Submit a Manuscript: https://www.f6publishing.com

World J Methodol 2021 November 20; 11(6): 278-293

ISSN 2222-0682 (online) DOI: 10.5662/wim.v11.i6.278

MINIREVIEWS

Preterm nutrition and neurodevelopmental outcomes

Alyson Margaret Skinner, Hassib Narchi

ORCID number: Alyson Margaret Skinner 0000-0001-7723-8015; Hassib Narchi 0000-0001-6258-2657.

Author contributions: Both authors contributed to the writing of the manuscript; Narchi H designed the figure.

Conflict-of-interest statement: No conflict of interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Specialty type: Medical laboratory technology

Country/Territory of origin: United Kingdom

Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0

Alyson Margaret Skinner, Department of Paediatrics, Manor Hospital, Walsall Healthcare NHS Trust, Walsall WS2 9PS, West Midlands, United Kingdom

Alyson Margaret Skinner, Hassib Narchi, Department of Paediatrics, College of Medicine and Health Sciences, United Arab Emirates University, Al Ain PO Box 17666, United Arab Emirates

Corresponding author: Alyson Margaret Skinner, BPharm, BSc, FRCP, MBChB, MD, MSc, Doctor, Department of Paediatrics, Manor Hospital, Walsall Healthcare NHS Trust, Moat Road, Walsall WS2 9PS, West Midlands, United Kingdom. alysonskinner8@gmail.com

Abstract

Survival of preterm infants has been steadily improving in recent years because of many recent advances in perinatal and neonatal medicine. Despite these advances, the growth of survivors does not reach the ideal target level of the normal fetus of the same gestational age. Postnatal weight gain is often not achieved because extrauterine growth has higher energy requirements than intrauterine growth, due to the intensive care environment, illness and inadequate nutrition. Although many other factors influence infant brain development, including family socioeconomic and educational background, the role of nutrition is considerable and fortunately, amenable to intervention. In the preterm neonate, the brain is the most metabolically demanding organ, consuming the largest proportions of energy and nutrient intake for its function and programmed growth and maturation. Weight gain, linear and head circumference growth are all markers of nutritional status and are independently associated with long-term neurodevelopment. Brain development is not only the result of nutrients intake, but in addition, of the interaction with growth factors which depend on adequate nutrient supply and overall health status. This explains why conditions such as sepsis, necrotizing enterocolitis and chronic lung disease alter the distribution and accretion of nutrients thereby suppressing growth factor synthesis. In this review, we will focus on the direct role of nutrition on neurodevelopment, emphasizing why it should be started without delay. The nutritional requirements of the preterm infant will be discussed, followed by the effects of general nutritional interventions and specific nutrients, as well as the role of nutritional supplements on neurodevelopment. The primordial role of human breast milk, breast milk fortifiers and human milk oligosaccharides will be discussed in detail. We will also examine the role of nutrition in preventing neonatal complications which can affect neurodevelopment in their own right.

Key Words: Brain; Nutrition; Preterm infants; Neurodevelopmental outcomes; Newborn;



Grade E (Poor): 0

Received: March 28, 2021

Peer-review started: March 28, 2021 First decision: June 17, 2021

Revised: July 13, 2021 Accepted: September 27, 2021 Article in press: September 27, 2021

Published online: November 20.

P-Reviewer: Zeng H S-Editor: Wang LL L-Editor: A

P-Editor: Wang LL



Breast milk

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Survival of preterm infants has been steadily improving because of the many recent advances in perinatal and neonatal medicine. However, neither the growth of survivors reaches the ideal target level of the normal foetus of the same gestational age, nor is the optimum postnatal weight gain often achieved. In the preterm neonate, the brain is the most metabolically demanding organ. Growth is a marker of nutritional status and is also independently associated with long-term neurodevelopment. In this review, we will discuss the direct role of nutrition and the effect of general and specific nutritional interventions on neurodevelopment.

Citation: Skinner AM, Narchi H. Preterm nutrition and neurodevelopmental outcomes. World J

Methodol 2021; 11(6): 278-293

URL: https://www.wjgnet.com/2222-0682/full/v11/i6/278.htm

DOI: https://dx.doi.org/10.5662/wjm.v11.i6.278

INTRODUCTION

Survival of preterm infants has been steadily improving in recent years because of the many advances in perinatal and neonatal medicine, such as antenatal corticosteroids administration, surfactant therapy and novel mechanical ventilation modalities. Despite these advances, the growth of preterm infants, as indicated by anthropometric indices and body composition, does not reach the target level of the normal foetus of the same gestational age. Often optimal postnatal weight gain is not achieved because extrauterine growth has higher energy requirements than intrauterine growth, due to the intensive care environment, illnesses and inadequate provision of nutrition. The persisting suboptimal nutrition in survivors remains an issue and is associated with adverse outcomes, such as chronic lung disease (CLD), necrotising enterocolitis (NEC), sepsis and, more worrying, neurodevelopmental impairment, especially in the cognitive domains, which is increasing in prevalence[1]. These adverse developmental outcomes can be secondary to abnormal brain development, postnatal brain injury and suboptimal nutrition occurring during hospital admission. Therefore, preterm birth can be considered a nutritional emergency.

Improved neurodevelopmental outcomes in extremely low birth weight (ELBW) infants are associated with a reduction in the incidence of CLD, apnea, intraventricular hemorrhage (IVH) and any associated nutritional deficiency[2]. Although many other factors influence infant brain development, not least family socioeconomic and educational background, the role of nutrition is considerable and fortunately, amenable to intervention[3].

In the preterm neonate, the brain is the most metabolically demanding organ and consumes the largest amount of energy and nutrients for its function and programmed growth and maturation[4]. Starting in the prenatal period and continuing until the second year of life, the development of the cerebral white and gray matter structures of the central nervous system (CNS) involves many processes, especially between 22 and 42 wk post-conception. Cell replication, neurogenesis, neuronal differentiation, cell migration, myelination, synaptogenesis, progression and regression of specific structures in different periods of development and neuroplasticity, are some of these processes[4]. The number of cell replication cycles in the CNS is decreased in malnutrition, thereby reducing total brain DNA and leading to a restriction in dendritic arborization and a reduction in the connections between neurons[5]. These cellular mechanisms affect the pre-oligodendrocyte, the widely disseminated activated microglia and the reactive astrocytes. As they occur over a relatively long time period, neurorestorative interventions, such as better nutrition can improve brain development[6]. Also, alterations in dietary precursors may affect neurotransmitter levels, for example, serotonin, norepinephrine, dopamine and acetylcholine in specific brain regions. In addition, essential and nonessential lipid components play an important role in the structural composition of the brain and of myelin sheaths. The resulting CNS impairment, caused by these nutrient deficiencies, involves motor and cognitive development and social abilities. They are also associated with alterations in the sleep-wake cycle organization, neurovegetative activities during sleep, waking electroencephalographic (EEG) activity and visual and auditory evoked responses.

The neurodevelopment of premature babies is very sensitive to nutrition in the first few weeks of life, and these effects may be long lasting until the age of eight years[7]. Cerebellar development is most affected by nutritional deprivation around the time of birth, and also, synaptic connectivity occurring before the third year of life. Nutrition is positively associated with weight gain, increased brain volumes and white matter maturation on magnetic resonance imaging at term equivalent age and neurodevelopment in infancy[7-9].

Adequate early nutrition may also attenuate the adverse effects of neonatal illness. Studies of EEG maturational age (EMA) during periods of quiet sleep which analysed spectral power, continuity and interhemispheric synchrony, have shown that a lower intake of calories and carbohydrate is associated with a greater reduction of spectral amplitude in the delta band. A lower protein intake was associated with higher discontinuity. In contrast, both higher proteins and lipid intake were associated with better developmental increase in interhemispheric synchrony and with brain activity maturation [10]. Increased caloric and protein intakes in preterm infants are associated with a higher fat free mass (a key marker for organ growth) and better neurodevelopment outcomes[11]. Protein and energy deficits in preterm nutrition are commonly associated with other nutritional deficiencies and also with psychosocial deprivation. Such interactions impact on the brain's structural development, disrupting neurodevelopment and adversely affecting cognitive performance[12].

Weight gain, linear and head circumference growth are all markers of nutritional status and are independently associated with long-term neurodevelopment. Brain development is not only a result of nutrients intake, but also of the interaction with growth factors, which depend on adequate nutrient status and overall health status. Sepsis, inflammation (occurring in NEC, CLD) and corticosteroids administration, alter the distribution and accretion of nutrients and suppress growth factor synthesis. Therefore, in addition to provision of adequate nutrients, strategies to reduce the incidence of these conditions are also required to optimise brain growth and develo-

In this review, we will focus on the direct role of nutrition on neurodevelopment. First, we will discuss the nutritional requirements of the preterm infant, followed by the effects of general nutritional interventions, specific nutrients and nutritional supplements on neurodevelopment[9]. We will then examine the role of nutrition in preventing neonatal complications, such as sepsis, NEC and CLD, which can also affect neurodevelopment in their own right (Figure 1)[4,13].

NUTRITIONAL REQUIREMENTS

Macronutrients refer to carbohydrates, fats and proteins while micronutrients include trace elements, electrolytes, and vitamins. Nutritional requirements, based on the estimated average requirement of a specific population group, are defined as the type and amount of nutrients needed to support normal health, growth and development. However, as preterm infants do not constitute a homogeneous population, their requirements must be individualised based on their clinical condition and developmental stage[14].

The high growth rate of premature infants requires a proportionally very high intake of all nutrients. With the general assumption that adequate postnatal growth should approximate the in-utero growth of a normal foetus, nutrient requirements were initially derived from postmortem measurement of the size and body composition of fetuses at varying gestational ages. The resulting nutritional intakes were estimated as the amounts required to replicate intrauterine rates of growth and nutrient accretion. The traditional method for estimating protein requirement is the factorial method, based on the body composition of the foetus with an estimate of the inevitable urinary nitrogen losses (occurring even in the absence of nitrogen intake) and of the amount deposited in-utero for accretion of tissue, corrected for efficiency of absorption and deposition[15]. Improvements to this model now take into account, not only the estimation of intrauterine accretion rate, organ development and factorial estimates of requirements, but also nutrient interactions, supplemental feeding and long-term developmental outcomes[16].

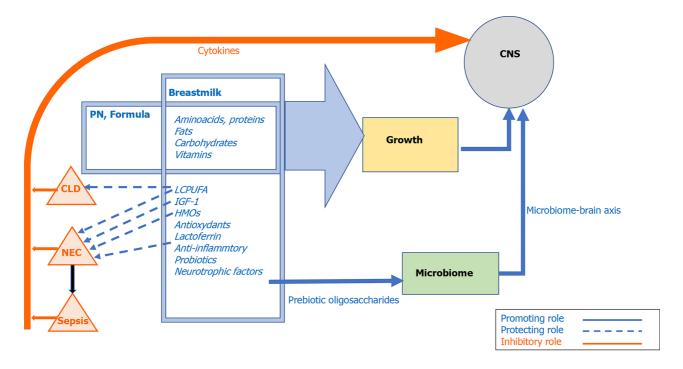


Figure 1 Direct effect of nutrients on brain development of the preterm infant and indirect impacts in preventing some neonatal complications, which can affect neurodevelopment in their own right. PN: Parenteral nutrition; LCPUFA: Long chain polyunsaturated fatty acids; IGF-1: Insulin-like growth factor 1; HMOs: Human milk oligosaccharides; CLD: Chronic lung disease; NEC: Necrotising enterocolitis.

Energy, stored in the body in the form of adenosine triphosphate (ATP), is required for several cellular and organ functions. Preterm infants have low energy reserves because they are born before adequate accumulation of fat and glycogen has occurred. In healthy premature infants, a recommended daily energy intake of 110-130 kcal/kg/day allows a growth rate similar to that of the intrauterine growth rate. For the neonate, carbohydrates are the main energy source (4 kcal/gram), in the form of glucose. Glucose synthetic rates in preterm infants are much higher at 6-8 mg/kg/min compared to 3-5 mg/kg/min in term infants. Because of their high energy-density (9 kcal/gram), fats or lipids are a very good source of energy. In addition, they also provide essential fatty acids indispensable for brain and visual development. Proteins are essential for normal growth and development (see below). After the initial free water diuresis, 2-3 mmol/kg/day of sodium and potassium (as a minimum) will be required to maintain serum levels in the normal range. Preterm infants often develop low sodium levels in the second week of life due to renal immaturity resulting in sodium loss. A normal serum sodium level is a requirement for adequate growth. Calcium, magnesium and phosphorus are essential for tissue structure and function. Vitamins are important for growth and development facilitating many reactions in intermediary metabolism. Preterm infants have low body stores of fat-soluble vitamins at birth due to limited transfer of lipid-soluble substrate across the placenta and, therefore, provision of the latter should be started as soon as lipid containing parenteral nutrition is initiated. A summary of the high nutrient requirements of the preterm infant is shown in Table 1. Human milk alone cannot provide the nutritional needs (especially protein) of preterm infants and requires fortification.

Providing preterm infants with adequate nutritional support presents unique challenges. Failure to provide adequate nutrient intakes at all stages of development places them at risk of impaired neurodevelopment. Therefore, every effort must thus be made to meet their complete nutritional needs from birth[15]. The main and most important reasons for growth failure in premature infants are the physiological limitations preventing the provision of nutrients by enteral feeding. This, in turn is compounded by the very high nutritional needs. Provision of nutrients has to take into account the immaturity of the intestinal tract, a major risk factor for the development of NEC. To overcome this, parenteral nutrition is used during the early days and often weeks of life. Although there are problems associated with parenteral nutrition, especially that of infection, failure to provide nutrients parenterally would place these infants at high risk of impaired neurodevelopment and host defenses. Providing early nutrient intakes by the parenteral route or with breast milk when possible, are therefore crucial to optimise long-term health outcomes[3].

Table 1 Nutritional requirements ¹				
Body weight	500-1000 g	1000-1500 g	1500-2200 g	2200-3000g
Foetal weight gain (g/kg/d)	19.0	17.4	16.4	13.4
Protein(g/kg/d)	4.0	3.9	3.7	3.4
Energy (kcal/kg/d)	106	115	123	130
Protein/energy (g/100 kcal)	3.8	3.4	3.0	2.6
Glucose	6-8 mg/kg/min			
Lipids	2.5-3.5 g/kg/d			
Calcium	60-90 mg/kg/d			
Phosphorus	40-70 mg/kg/d			

¹Estimated by factorial and empirical methods, derived from several references in this article[14-16].

INDIVIDUAL NUTRIENTS DIRECTLY AFFECTING NEURODEVELOPMENT IN PRETERM INFANTS

Caloric intake

Caloric intake in a large population of extremely premature infants (born < 28 wk' gestation) was associated with the development of retinopathy of prematurity (ROP) [17]. In preterm infants, caloric deprivation is associated with a significantly low composite score on Bayley-III scales performance at 18-24 mo corrected age [13]. In the same study, late onset sepsis, NEC, and CLD were also found to be risk factors for impaired neurodevelopment. Improving nutritional support, specifically total caloric intake is therefore necessary to prevent neurodevelopment impairment.

Protein

Adequate protein intake is crucial for satisfactory growth and neurodevelopment in preterm infants. It has been recommended that a high protein intake starts on day 1 of life to limit extrauterine growth restriction, which in itself negatively affects neurodevelopment. Brain magnetic resonance imaging (MRI) studies in the first month of life in extremely preterm infants, who had received early nutrition, demonstrated larger brain volumes in those who had received a higher protein and caloric intake than controls[18]. In very low birth weight (VLBW) newborns, serial cranial ultrasound measurements of the size of several brain structures in the first month of life showed a positive correlation between enteral protein intake and brain volume[19]. Supplemental enteral proteins were also shown to reduce postnatal growth restriction and improve neurological outcomes in preterm infants < 1000 g and in small for gestational age (SGA) infants (birth weight <10th percentile)[20]. Similarly, parenteral amino acid intake within the first two weeks of life was correlated with neurodevelopmental outcomes at 2 years, with higher language and motor scores on the 2-year Bayley Scales of Infant and Toddler Development (Bayley III)[21].

However, not all studies have reported a consistently positive relationship between early protein intake and neurocognitive outcomes. Protein supplementation in very low birth weight infants (VLBW) (birth weight < 1500 g) showed a significant positive correlation with neurological development in communication, auditory, verbal language, cognitive function, social connection and gross motor development, but not with fine motor, problem solving or personal-social relationships in the third year of life[22]. However, this study involved small population numbers and protein supplementation was stopped when the infants weighed 1500 g. In preterm infants born before 32 wk' gestation or with birth weight < 1500 g, addition of fortifier to expressed human milk to increase protein intake, was associated with better head growth and weight gain at 40 wk' postmenstrual age. However, this intervention had no benefits on long-term growth and neurodevelopment at 12 to 18 mo corrected age [23]. Likewise, early aggressive parenteral nutrition with high-dose amino acids and mixed fat emulsions did not improve neurodevelopmental scores of VLBW infants, nor decrease the incidence of major disabilities[24]. Again this study included a small study population divided into 5 groups with different parenteral feeding regimes.

Fats and long-chain polyunsaturated fatty acids

Long-chain polyunsaturated fatty acids (LCPUFA), including docosahexaenoic acid (DHA) and arachidonic acid (ARA), are essential for neurodevelopment and normal vision. In-utero DHA accretion occurs primarily in the last trimester of pregnancy supporting rapid growth and brain development. Premature infants, born before the completion of this process are therefore relatively deficient in these essential fatty acids. This deficiency persists for a long period after birth because of ineffective conversion from precursor fatty acids, lower fat stores, and limited nutritional provision of DHA and ARA. Besides long-term visual and neurodevelopmental risks, very preterm infants have significant morbidity and mortality from diseases specific to premature birth, including CLD, NEC and ROP. There is increasing evidence that DHA has protective benefits against these complications[25]. The ratio of n-3 to n-6 LCPUFAs, which generates eicosanoids, is important in moderating the effects of hypoxia, inflammation, infection, thrombosis and oxidative damage of key organs such as the lungs, brain and retina [26]. These actions improve the prognosis for ELBW and VLBW infants. By providing LCPUFAs, including DHA, breastfeeding improves cognitive development, especially in infants with a proven lower activity of DHA synthesis[27].

The outcomes of supplementation with PUFAs are still contested. Although early PUFAs supplementation in preterm infants had a positive impact on early childhood psychomotor development and visual acuity, they had no significant effects on global intelligence quotient (IQ) in childhood[28]. Similarly, a metanalysis of LCPUFAs supplementation in preterm infants showed a significant improvement in their neurodevelopment (assessed by the Mental Development Index of the Bayley Scales at one to three years of age). Supplementation during lactation accelerated neurodevelopment but their final developmental outcome remained unchanged[29]. Other studies have shown no beneficial effect of early high-dose mixed fat emulsions on neurodevelopment of VLBW infants[24,30]. Studies of formula milk supplemented with LCPUFAs showed mixed results, some reporting better psychomotor outcomes [31] while others showed no long-term benefits on visual or psychomotor development[32]. Infants enrolled in the supplemented formula study were mature in general and the quality of evidence was low for both visual and neurodevelopmental outcomes. Current recommendations for follow-on formula suggest that they should contain omega-3 DHA equal to the mean content of human milk, and also AHA[33].

lodine supplementation

Iodide supplementation in babies born before 31 wk' gestational age provided no benefit to neurodevelopment measured at 2 years of age[34]. Similarly, a large highquality study showed no beneficial effects of iodine supplementation for preterm infants[35].

Iron supplementation

In infants born ≥ 34 wk gestational age, latent in-utero iron deficiency (serum ferritin < 75 ng/mL at birth) was associated with abnormal auditory neural maturation [36]. The age at which to begin iron supplementation remains controversial. It is often given from day 28 of life onwards, assuming until then, the infant has sufficient iron stores from the breakdown of fetal hemoglobin. As breast milk does not provide sufficient iron to sustain a premature infant's growth and normal haemopoietic function, breastfed infants require iron supplementation. Formula-fed babies do not need iron supplements as milk formulas are iron fortified.

BREAST MILK

Fresh maternal milk contains factors involved in antioxidant and anti-inflammatory defense[35], gut microbiome establishment and the maturation of immune defenses [37,38], feed tolerability and metabolism[39] and also protects infants against NEC [16]. For these and other reasons, human milk is the preferred choice for feeding premature infants. It reduces the risk of morbidities and promotes growth through elevated serum levels of insulin-like growth factor 1 (IGF-1), an essential intrauterine hormonal mediator of growth[40].

Human milk contains critical nutrients and other neurotrophic factors that benefit the brain of preterm infants[41]. A systematic review of neurodevelopment outcomes of ex-preterm children found better test scores in those who had been fed breast milk compared with formula feeds, when evaluated in the first three years of life and also, between the age of five to eleven years [42]. In a prospective study of 316 VLBW infants, correcting for gestational age (GA), being born SGA, and for complications at birth and during hospital admission, better neurodevelopment at 24 mo corrected age was associated with breast milk feeds[43]. VLBW infants fed fortified human milk had increased head circumference velocity and decreased regional white matter diffusivity when assessed by magnetic resonance diffusion tensor imaging suggesting improved maturation of the cerebral connective tracts[44]. They also had improved visual perception as judged by their response to global motion[45]. Brain MRI studies at term equivalent age, using network-based statistics, tract-based spatial statistics and volumetric analysis of white matter water diffusion parameters, reported that feeding with breast milk in the weeks after preterm birth was associated with improved structural connectivity of developing networks and greater fractional anisotropy (FA)weighted connectome in major white matter fasciculi [46]. There were also better outcomes observed in infants fed breast milk for more than 90% of their hospital admission duration compared with those who received breast milk for 75% of the same period.

When a mother expressed breast milk is unavailable, donor breast milk is used widely to feed premature babies. The methods used to process donor milk reduce the levels of protective and growth promoting factors found in fresh breast milk. However, donor milk still offers protection against NEC but remains inferior to fresh human milk in promoting the growth and development of very preterm infants[47].

Breast milk fortification

Human milk, although the preferred feeding for premature infants because of its protective effects, does not provide adequate amounts of nutrients to support the rapid growth and development of the premature baby and therefore, must be fortified. Failure to fortify mother's milk places the baby at risk of neurodevelopmental impairment. As mentioned earlier, an enhanced nutrient supply with breast milk fortification to VLBW infants was associated with increased head circumference growth and decreased regional white matter mean diffusivity seen on magnetic resonance diffusion tensor imaging, suggesting improved maturation of cerebral connective tracts[44]. A recent study of long-term outcomes of preterm infants fed an exclusive human milk-based diet using a donor human milk-based fortifier reported an absence of severe cognitive developmental delay using a cutoff score of 70 (Bayley Scales of Infant and Toddler Development III) measured at 18-22 mo corrected gestational age (GA)[48].

Some controversies still exist. An earlier study showed no significant differences at 18 mo corrected age in mean Bayley-III cognitive composite score, language composite score, or motor composite score in VLBW infants fed fortified donor milk when compared with formula fed babies [49]. Similarly, a study of the balance of risks and benefits of feeding formula vs donor breast milk for preterm or LBW infants found no evidence of an effect on long-term neurodevelopment, although the level of evidence was moderate [50]. This may be explained by the effects of the pasteurisation process on donor milk causing reduced levels of the non-nutritive content. Even fewer data are available for growth and developmental outcomes assessed beyond infancy and many do not show consistent effects. Feeding with a nutrient enriched formula had no effect on Bayley Mental Development Index scores at 18 mo post term or on cerebral palsy, and there were no effects on long-term development[51].

Human milk oligosaccharides

Human breast milk has two types of carbohydrates, lactose and oligosaccharides. HMOs are indigestible unconjugated complex carbohydrates forming the third most abundant component of breast milk, after lactose and lipids. They are a complex mixture of over 200 non-digestible and non-nutritional carbohydrates. There are three main HMO categories based on their building blocks. The first category comprises non-fucosylated neutral (core) HMOs, which are the foundations upon which other HMOs are built, lacto-N-tetraose isomer (LNT) being the most abundant representative in this category. The fucosylated neutral HMOs form the second category, with 2'FL the most abundant fucosylated HMO and DFL (LDFT) among the 10 most abundant representatives in this group. The third category is composed of the sialyated acidic HMOs, 6'SL is the leading representative of this group followed by 3'SL.

In animal studies, using a genome-wide profiling of the intestinal epithelial transcriptome in response to HMOs, the latter were found to protect against NEC in part by altering the differentiation of the crypt-villus axis host transcriptome in 225 target genes involved in cell proliferation and differentiation, including upregulation

of the stem cell differentiation marker HMGCS2. HMOs also directly induced a series of biological processes to protect the intestine [52]. In another experiment, milk supplementation with 2'-FL and 6'-SL, but not lactose, prevented NEC in mice and piglet models and attenuated NEC inflammation in the ileum, in part through TLR4 inhibition with reduced apoptosis and inflammation[53].

In neonates, HMOs have been shown to decrease the incidence of late-onset sepsis and NEC, both conditions being associated with neurodevelopment impairment. Reducing the risk of these complications leads to less disruption of feeds which is also beneficial for normal neurodevelopment. In the premature baby's gut, HMOs and the microbiome of breast milk act as immunomodulatory agents that provide intestinal homeostasis through regulation of the microbiome and protection of the intestinal barrier, thereby protecting the premature baby at risk of NEC[54]. The concentrations of different HMOs found in neonates vary according to the lactation period and maternal secretory phenotype status[55]. Fucosyl--N-hexose (FDSLNH) was associated with a reduced risk of late-onset sepsis in VLBW infants[55].

An animal study examined the effect of HMOs on cognition, novel object recognition task, brain development, and hippocampal gene expression using magnetic resonance imaging procedures to assess structural brain development and hippocampal tissue analysis for mRNA expression. The results showed improved recognition memory and better absolute and relative volumes of the cortical and subcortical brain regions. Hippocampal mRNA expression of GABRB2, SLC1A7, CHRM3, and GLRA4 were also affected by HMOs[56].

Relative abundances of several individual HMOs were associated with normal growth and neurodevelopment in infants, mainly motor development at 12 mo, including ability to stand or walk alone. They were also associated with language skills, socioemotional development, executive function and working memory at 18 mo, mostly among secretors[57].

Microbiota and probiotics

While the newborn term microbiome is normally dominated by Bifidobacterium species, the microbial composition and functions in preterm babies has a higher initial percentage of Lactobacillaceae [58]. It has been established that intestinal dysbiosis, (abnormal microbial colonisation), can occur as a result of exogenous factors such as mode of delivery, formula feeding, and exposure to antibiotics. These factors predispose preterm infants to sepsis and NEC in association with an already existing impaired intestinal barrier and immature immunity. The prolonged use of antibiotics and parenteral nutrition were reported to have significant adverse effects on the Lactobacillus and Bifidobacterium levels in the gut of preterm infants[59].

Inflammation and perinatal infection play a crucial role in the pathogenesis of white matter injury in preterm infants. Therefore, nutritional components with immunomodulatory and/or anti-inflammatory effects may serve as neuroprotective agents. There is growing evidence of the existence of a microbiome-gut-brain axis in which the microbiome interacts with the brain through immunological, endocrine, and neural pathways. Consequently, nutritional components that influence gut microbiota may also exert beneficial effects on the developing brain. Based on these properties, probiotics, prebiotic oligosaccharides, and certain amino acids may offer neuroprotection. Certain nutritional components may also have a neuroprotective role against white matter injury through modulation of inflammation and infection and may influence the microbiome-gut-brain axis[60].

As stated above, the normal term newborn microbiome is dominated by Bifidobacterium species but the establishment of abnormal microbial colonisation (intestinal dysbiosis) can be influenced by several factors, thereby increasing the risk of developing sepsis and NEC. Probiotics have been shown to reduce rates of necrotising enterocolitis (NEC), sepsis, and mortality[61]. However, research in this area has been complicated by studying different bacterial strains, administration of different doses of probiotics and for different time periods of administration. Provided all safety issues are met, it is currently recommended that either Lactobacillus rhamnosus GG ATCC53103 or the combination of Bifidobacterium infantis Bb-02, Bifidobacterium lactis Bb-12, and Streptococcus thermophilus TH-4 would help reduce the rates of NEC[62]. It is essential that probiotic strains should be devoid of transferable antibiotic resistance genes.

MEDIUM AND LONG-TERM EFFECTS OF CONDITIONS INFLUENCED BY NUTRITION WHICH HAVE AN IMPACT ON NEURODEVELOPMENT

Growth

The long-term effect of inadequate early postnatal growth (EPG) is a growing concern. Poor head growth during hospital admission and post discharge is associated with delayed motor and cognitive development between 16 and 36 mo of age[63]. A systematic review of the relationship between EPG before 3 years of age and neurodevelopmental outcome, showed that EPG was positively associated with good neurodevelopmental outcome in a non-linear association, with a plateau attained with higher weight gain, suggesting a possible ceiling effect [64]. Similarly, post-discharge failure to thrive was found to be significantly correlated with poor neurodevelopmental outcomes. There was an association between lower body weight (< 3rd percentile) at corrected ages of 6, 12, and 24 mo and poor neurodevelopmental outcomes among 30% of VLBW premature infants[65]. Extra uterine growth retardation (EUGR) of VLBW infants is significantly associated with low mental development index (MDI) at a corrected age of 24 mo[66].

Studies examining the relationship between body composition and neurodevelopment in VLBW infants reported higher growth, fat mass and fat-free mass were associated with larger cerebellar volumes at term[67]. Also, greater fat-free mass (measured by air displacement plethysmography) gained by VLBW premature infants as inpatients, was associated with improved cognitive and motor scores at 12 months corrected age (CA)[68]. Increased provision of protein and calories during the first week of life is positively associated with fat-free mass gains, but not fat mass gains. Frondas-Chauty et al[69] reported a deficit of fat-free mass in VLBW infants at discharge was associated with neurological impairment at 24 mo (corrected age), independent of sex, GA and BW.

Necrotising enterocolitis, intestinal failure, and sepsis

NEC often occurs in premature infants at a time of rapid brain growth and development and is associated with poor neurodevelopmental outcomes. The pathogenesis of neurodevelopmental impairment following NEC is likely multi-factorial, involving both nutritional and non-nutritional factors[70]. A variety of circulating cytokines are known to be associated with white matter injury. There is also increased risk of secondary blood stream infections and subsequent nutritional compromise[71]. Breast milk, probiotics and lactoferrin have all been shown to reduce the incidence of NEC and late-onset sepsis[72].

NEC is also a major cause of intestinal failure (IF), which in turn is associated with developmental delay in multiple domains. While gross motor skills appear to be most significantly affected, cognitive deficits become more apparent at 26 months[73]. In children with IF assessed before 15 mo corrected age, 88% had abnormal general movements and over 50% had suspect or abnormal gross motor scores. Factors significantly associated with poorer outcomes at 12-15 mo included prematurity, low birth weight, CNS co-morbidity, longer neonatal intensive care admission, NEC, number of operations and conjugated hyperbilirubinemia. Thus, multiple risk factors contribute to early developmental delay in children with IF and close developmental follow-up is essential in these patients[73].

GENERAL DIETARY INTERVENTION

Early feeding

As mentioned earlier, the preterm brain is very vulnerable to undernutrition in the first few weeks of life, even over a period as short as 4 wk, and these effects persist up to the age of 8 years[74]. Suboptimal nutrient intakes permanently affect later cognitive attainment and therefore, early nutrient intakes and breast milk provision are essential[3]. A positive association between several components of early nutrition, growth, brain volumes, white matter maturation, and neurodevelopment in infancy has been shown[7,75]. Immediate breast milk feeding results in elevated insulin-like growth factor 1 (IGF-1), an essential intrauterine hormonal mediator of growth, measured at term, in the serum of preterm infants[40]. Early administration of optimal postnatal parenteral and enteral nutrients helps to prevent neurodevelopmental impairment caused by extrauterine growth restriction, NEC, sepsis, CLD dysplasia, and ROP[72].

Early introduction of enteral feedings to stimulate or prime the gut even in the acutely ill preterm infant, is well tolerated [76,77]. Several metanalyses have shown the safety of early trophic feeding with breast milk within 48 h after birth and progressing the volume of enteral feeding before four days of life at a rate of 24 mL/kg/d in clinically stable very preterm and VLBW infants. These measures did not lead to higher mortality or an increased incidence of NEC[77,78].

However, despite the introduction of more aggressive early feeding guidelines and improved energy and nutrient intakes, many preterm infants do not achieve their recommended nutrition intake in the first week of life, especially those with morbidities[79].

FEEDING IN PRACTICE

During the immediate postnatal period, the aim of nutritional support is twofold: i) to provide an early and uninterrupted flow of nutrients to ensure the anabolic state that existed in-utero continues with minimal or no interruption and ii) to stimulate the immature gastrointestinal tract to undergo maturation. However, as described above, early provision of the preterm infant with the recommended nutritional intakes is fraught with difficulties. The latter include comorbidities which preclude normal enteral feeding such as respiratory distress, haemodynamic instability and sepsis. Comorbidities also indirectly affect the delivery of adequate nutrition because a significant proportion of the total daily fluids may be used to deliver several medications, thereby limiting the amount of actual nutrition given to the baby. This problem can be overcome to some extent by concentrating the constituents of TPN in a smaller volume. In addition, gut immaturity limits the amount of enteral feed the baby will tolerate and is also a major risk factor for NEC, which will further impair enteral feeding. Furthermore, the high nutritional requirements cannot be fully provided immediately after birth and thus need progressive and cautious escalation. All these factors contribute to the risk of undernutrition with its resulting outcomes.

Strategy for nutritional support

Nutritional support of preterm infants occurs in four distinct phases, each with its own risks and challenges. During the early phase, nutrients are almost exclusively provided via the parenteral route, while small enteral feedings (gastrointestinal priming or trophic feeds) are used to stimulate the intestinal tract towards maturation. During the subsequent transition phase, enteral feeding is slowly advanced as the intestinal tract shows evidence of maturation, and parenteral nutrition is gradually phased out. During the late phase, infants are exclusively enteral fed and are expected to grow normally. If provided with the necessary nutrients, preterm infants may also show catch-up growth, making up for lost time during the early phase. Preterm infants continue to have special nutritional needs after discharge from hospital.

The importance of providing early nutrition has led to the use of parenteral nutrition immediately after birth with the knowledge that it differs from enteral nutrition in nutrient composition and carries the risk of infection. Nevertheless, the use of total parenteral nutrition (TPN) has contributed to improving survival rates and at its most fundamental level, it helps the newborn premature infant maintain a neutral nitrogen balance.

While nutrients are provided parenterally immediately after birth, starting trophic feeds with breast milk from day 1 of life, with volumes as low as 2 mL every 4 to 6 h, is the safest and most effective method of stimulating the intestinal tract and accelerating its maturation.[18]

When breast milk is not available, pasteurised donor milk (free of viruses such as HIV and cytomegalovirus) can be used for gastrointestinal priming. Although pasteurisation diminishes some of the protective and trophic factors of human milk, donor milk still remains protective against NEC and sepsis while maintaining trophic effects. Using gastrointestinal priming has been shown to lead to earlier establishment of full feedings and to earlier hospital discharge without an increase in morbidities. Moreover, earlier achievement of full feedings has been shown to decrease the risk of

Increasing enteral feed volumes

Once intestinal tract maturation occurs, enteral nutrients can be progressively increased while parenteral nutrition is proportionally reduced before being discontinued. Gastrointestinal motility is a marker of gut maturation and is monitored by the

assessment of gastric residuals. Improving gastric emptying (facilitated by breast milk) reflects the improving ability to digest and absorb nutrients. Feeding volumes can be gradually increased by 20-25 mL/kg each day as gastric residuals permit. The absence of significant gastric residuals should be ascertained before feeding volume is further increased. Although more rapid increases in feed volumes have been achieved safely, intestinal maturation requires time and therefore more rapid increases are not necessary. Parenteral nutrition is usually maintained until enteral feedings reach 120 mL/kg/day after which it may be discontinued, provided the baby is tolerating enteral feeds.

Breast milk fortification

When feeding volumes of 80-100 mL/kg/day are tolerated, fortification of breast milk is usually initiated, although in some neonatal units, fortification is started at an earlier stage. Half-strength fortification may be tried before proceeding to full strength.

Once full feed volumes (≥ 150 mL/kg/day) are established, the aim is to facilitate a growth rate similar to intrauterine growth. If the growth rate is not satisfactory, enteral intake may be increased by 10%-20%. The best nutrition is fortified human mother's milk or, when not available, fortified donor milk or special formulas with higher protein content (3.3-3.6 g/100 kcal). Babies should also be given multivitamins and, if being exclusively fed with expressed breast milk, iron supplements from day 28 onwards. It is important to measure serum sodium and phosphate levels regularly (minimum once weekly). Serum sodium levels should be kept in the normal range to facilitate the action of IGF1 at cellular level. Serum phosphate levels should be at least 2.0 mmol/L as the premature baby's requirement for phosphate is high, not only for bone development but also for intermediary metabolism.

After discharge from hospital, preterm infants continue to have high nutrient requirements in addition to potential accrued deficits in bone mineral content. There is therefore a need for continued fortification of human milk and mineral and vitamin supplementation or to feed with an enriched post-discharge formula.

DIRECTIONS FOR FUTURE RESEARCH

Despite the major improvements already made in preterm infant nutrition, several unknowns remain and should direct future research efforts. Many studies report historical data in small populations.

There is a lack of a standardised approach to report nutritional intake data and measures of outcomes. Also, there is substantial variation in methods used to estimate and calculate nutritional intakes, making comparison amongst studies difficult and meta-analysis unreliable. Future research should focus on the development of minimal reporting sets of standardised nutritional interventions, and an agreed checklist for standardised reporting of neonatal nutrition and core outcomes.

More research is needed to determine the optimal nutrition, growth rates and body composition in preterm infants that are associated with the best neurocognitive benefits while minimising the long-term risk of chronic diseases.

Adequately powered randomised controlled trials on the late commencement of parenteral nutrition in term and late preterm infants who have IF as well as trials of intensified initial parenteral nutrition are required. Also, large-scale adequately powered randomised controlled trials should determine the optimal intake of amino acids and the effects of caloric balance in parenteral nutrition on the brain and neurodevelopment. The comparison of different types and amounts of protein in multi-component fortifiers on long-term neurodevelopment merits further investigation.

Further studies investigating the interactions between maternal HMOs and the intestinal microbiome, and the identification of specific pathways by which individual HMO structures exert protective actions are needed. More information on the development of a simple, high-throughput method to allow full characterization of HMOs is required and also, to determine if a causal relation exists between HMOs and neurodevelopmental outcome.

That LCPUFAs, particularly DHA, provide beneficial effects on preterm infants needs further exploration. These potential effects include better neurological outcomes at 2 years and reduction in the incidence and severity of neonatal morbidities such as CLD, NEC, ROP and possibly sepsis. Knowing how LCPUFAs affect different steps of the immune and anti-inflammatory response would contribute to determining the optimal LCPUFA requirements for good neurodevelopmental outcomes.

CONCLUSION

Despite the many difficulties encountered in nutrition research, several studies to date have demonstrated a positive relationship between aspects of nutrition when optimised and growth and neurocognitive outcomes for premature babies. Survival rates of premature infants, including those born at the extremes of viability, have improved enormously but future research needs to address how better growth and neurodevelopmental outcomes may be achieved. As more knowledge becomes available, neonatologists and all those involved in neonatal care, can significantly improve nutritional management, particularly in terms of quality, thereby making a major contribution to the lives of individuals, families and society.

REFERENCES

- Nakanishi H, Suenaga H, Uchiyama A, Kono Y, Kusuda S; Neonatal Research Network, Japan. Trends in the neurodevelopmental outcomes among preterm infants from 2003-2012: a retrospective cohort study in Japan. *J Perinatol* 2018; **38**: 917-928 [PMID: 29679045 DOI: 10.1038/s41372-018-0061-7]
- Asami M, Kamei A, Nakakarumai M, Shirasawa S, Akasaka M, Araya N, Tanifuji S, Chida S. Intellectual outcomes of extremely preterm infants at school age. Pediatr Int 2017; 59: 570-577 [PMID: 27935152 DOI: 10.1111/ped.13215]
- Embleton ND. Early nutrition and later outcomes in preterm infants. World Rev Nutr Diet 2013; 106: 26-32 [PMID: 23428677 DOI: 10.1159/000342553]
- Ramel SE, Georgieff MK. Preterm nutrition and the brain. World Rev Nutr Diet 2014; 110: 190-200 [PMID: 24751630 DOI: 10.1159/000358467]
- Dobbing J, Hopewell JW, Lynch A. Vulnerability of developing brain. VII. Permanent deficit of neurons in cerebral and cerebellar cortex following early mild undernutrition. Exp Neurol 1971; 32: 439-447 [PMID: 5110226 DOI: 10.1016/0014-4886(71)90010-0]
- Volpe JJ. Dysmaturation of Premature Brain: Importance, Cellular Mechanisms, and Potential Interventions. Pediatr Neurol 2019; 95: 42-66 [PMID: 30975474 DOI: 10.1016/j.pediatrneurol.2019.02.016]
- Coviello C, Keunen K, Kersbergen KJ, Groenendaal F, Leemans A, Peels B, Isgum I, Viergever MA, de Vries LS, Buonocore G, Carnielli VP, Benders MJNL. Effects of early nutrition and growth on brain volumes, white matter microstructure, and neurodevelopmental outcome in preterm newborns. Pediatr Res 2018; 83: 102-110 [PMID: 28915232 DOI: 10.1038/pr.2017.227]
- Sammallahti S, Kajantie E, Matinolli HM, Pyhälä R, Lahti J, Heinonen K, Lahti M, Pesonen AK, Eriksson JG, Hovi P, Järvenpää AL, Andersson S, Raikkonen K. Nutrition after preterm birth and adult neurocognitive outcomes. PLoS One 2017; 12: e0185632 [PMID: 28957424 DOI: 10.1371/journal.pone.0185632]
- Schneider J, Fischer Fumeaux CJ, Duerden EG, Guo T, Foong J, Graz MB, Hagmann P, Chakravarty MM, Hüppi PS, Beauport L, Truttmann AC, Miller SP. Nutrient Intake in the First Two Weeks of Life and Brain Growth in Preterm Neonates. *Pediatrics* 2018; **141** [PMID: 29440285 DOI: 10.1542/peds.2017-2169]
- Marchi V, Stevenson N, Koolen N, Mazziotti R, Moscuzza F, Salvadori S, Pieri R, Ghirri P, Guzzetta A, Vanhatalo S. Measuring Cot-Side the Effects of Parenteral Nutrition on Preterm Cortical Function. Front Hum Neurosci 2020; 14: 69 [PMID: 32256325 DOI: 10.3389/fnhum.2020.00069]
- Ramel SE, Haapala J, Super J, Boys C, Demerath EW. Nutrition, Illness and Body Composition in Very Low Birth Weight Preterm Infants: Implications for Nutritional Management and Neurocognitive Outcomes. Nutrients 2020; 12 [PMID: 31947964 DOI: 10.3390/nu12010145]
- Ehrenkranz RA, Dusick AM, Vohr BR, Wright LL, Wrage LA, Poole WK. Growth in the neonatal intensive care unit influences neurodevelopmental and growth outcomes of extremely low birth weight infants. Pediatrics 2006; 117: 1253-1261 [PMID: 16585322 DOI: 10.1542/peds.2005-1368]
- Lithoxopoulou M, Rallis D, Christou H, Goutsiou E, Varaklioti A, Karagianni P, Tsakalidis C, 13 Domeyer P, Kuriakeli G, Soubasi V. Early caloric deprivation in preterm infants affects Bayley-III scales performance at 18-24 mo of corrected age. Res Dev Disabil 2019; 91: 103429 [PMID: 31272067 DOI: 10.1016/j.ridd.2019.103429]
- Thureen P, Heird WC. Protein and energy requirements of the preterm/Low birthweight (LBW) infant. Pediatr Res 2005; 57: 95R-98R [PMID: 15817496 DOI: 10.1203/01.PDR.0000160434.69916.34]
- Ziegler EE. Meeting the nutritional needs of the low-birth-weight infant. Ann Nutr Metab 2011; 58 Suppl 1: 8-18 [PMID: 21701163 DOI: 10.1159/000323381]
- Klein CJ. Nutrient requirements for preterm infant formulas. J Nutr 2002; 132: 1395S-1577S [PMID: 12042465 DOI: 10.1093/jn/132.6.1395S]
- Vander Veen DK, Martin CR, Mehendale R, Allred EN, Dammann O, Leviton A; ELGAN Study Investigators. Early nutrition and weight gain in preterm newborns and the risk of retinopathy of prematurity. PLoS One 2013; 8: e64325 [PMID: 23734194 DOI: 10.1371/journal.pone.0064325]
- van Beek PE, Claessens NHP, Makropoulos A, Groenendaal F, de Vries LS, Counsell SJ, Benders

289



- MJNL. Increase in Brain Volumes after Implementation of a Nutrition Regimen in Infants Born Extremely Preterm. J Pediatr 2020; 223: 57-63.e5 [PMID: 32389719 DOI: 10.1016/j.jpeds.2020.04.063]
- Terrin G, De Nardo MC, Boscarino G, Di Chiara M, Cellitti R, Ciccarelli S, Gasparini C, Parisi P, Urna M, Ronchi B, Russo A, Sabatini G, De Curtis M. Early Protein Intake Influences Neonatal Brain Measurements in Preterms: An Observational Study. Front Neurol 2020; 11: 885 [PMID: 32982918 DOI: 10.3389/fneur.2020.00885]
- Biasini A, Monti F, Laguardia MC, Stella M, Marvulli L, Neri E. High protein intake in human/maternal milk fortification for ≤ 1250 gr infants: intrahospital growth and neurodevelopmental outcome at two years. Acta Biomed 2018; 88: 470-476 [PMID: 29350662 DOI: 10.23750/abm.v88i4.5316]
- Yang J, Chang SS, Poon WB. Relationship Between Amino Acid and Energy Intake and Long-Term Growth and Neurodevelopmental Outcomes in Very Low Birth Weight Infants. JPEN J Parenter Enteral Nutr 2016; 40: 820-826 [PMID: 25690346 DOI: 10.1177/0148607115572833]
- Kashaki M, Samghabadi FM, Bordbar A. Effect of Fortification of Breast Milk in Conjugation with Protein Supplement on Neurodevelopment of Preterm Low Birth Weight Infants at 3 Years. Med Arch 2019; 73: 344-350 [PMID: 31819309 DOI: 10.5455/medarh.2019.73.344-350]
- Dogra S, Thakur A, Garg P, Kler N. Effect of Differential Enteral Protein on Growth and Neurodevelopment in Infants < 1500 g: A Randomized Controlled Trial. J Pediatr Gastroenterol Nutr 2017; 64: e126-e132 [PMID: 27801753 DOI: 10.1097/mpg.000000000001451]
- Roelants JA, Vlaardingerbroek H, van den Akker CHP, de Jonge RCJ, van Goudoever JB, Vermeulen MJ. Two-Year Follow-up of a Randomized Controlled Nutrition Intervention Trial in Very Low-Birth-Weight Infants. JPEN J Parenter Enteral Nutr 2018; 42: 122-131 [PMID: 27875287 DOI: 10.1177/01486071166781961
- Harris WS, Baack ML. Beyond building better brains: bridging the docosahexaenoic acid (DHA) gap of prematurity. J Perinatol 2015; **35**: 1-7 [PMID: 25357095 DOI: 10.1038/jp.2014.195]
- 26 Uauy R, Mena P. Long-chain polyunsaturated fatty acids supplementation in preterm infants. Curr
- Koletzko B. Human Milk Lipids. Ann Nutr Metab 2016; 69 Suppl 2: 28-40 [PMID: 28103608 DOI: 2.7 10.1159/000452819]
- Shulkin M, Pimpin L, Bellinger D, Kranz S, Fawzi W, Duggan C, Mozaffarian D. n-3 Fatty Acid Supplementation in Mothers, Preterm Infants, and Term Infants and Childhood Psychomotor and Visual Development: A Systematic Review and Meta-Analysis. J Nutr 2018; 148: 409-418 [PMID: 29546296 DOI: 10.1093/jn/nxx031]
- Wang Q, Cui Q, Yan C. The Effect of Supplementation of Long-Chain Polyunsaturated Fatty Acids During Lactation on Neurodevelopmental Outcomes of Preterm Infant From Infancy to School Age: A Systematic Review and Meta-analysis. Pediatr Neurol 2016; 59: 54-61.e1 [PMID: 27318249 DOI: 10.1016/j.pediatrneurol.2016.02.017]
- Thanhaeuser M, Fuiko R, Oberleitner-Leeb C, Brandstaetter S, Binder C, Thajer A, Huber-Dangl M, Haiden N, Pablik E, Berger A, Repa A. A Randomized Trial of Parenteral Nutrition Using a Mixed Lipid Emulsion Containing Fish Oil in Infants of Extremely Low Birth Weight: Neurodevelopmental Outcome at 12 and 24 Months Corrected Age, A Secondary Outcome Analysis. J Pediatr 2020 [PMID: 32590001 DOI: 10.1016/j.jpeds.2020.06.056]
- 31 Alshweki A, Muñuzuri AP, Baña AM, de Castro MJ, Andrade F, Aldamiz-Echevarría L, de Pipaón MS, Fraga JM, Couce ML. Effects of different arachidonic acid supplementation on psychomotor development in very preterm infants; a randomized controlled trial. Nutr J 2015; 14: 101 [PMID: 26424477 DOI: 10.1186/s12937-015-0091-3]
- 32 Moon K, Rao SC, Schulzke SM, Patole SK, Simmer K. Longchain polyunsaturated fatty acid supplementation in preterm infants. Cochrane Database Syst Rev 2016; 12: CD000375 [PMID: 27995607 DOI: 10.1002/14651858.CD000375.pub5]
- Koletzko B, Bergmann K, Brenna JT, Calder PC, Campoy C, Clandinin MT, Colombo J, Daly M, Decsi T, Demmelmair H, Domellöf M, FidlerMis N, Gonzalez-Casanova I, van Goudoever JB, Hadjipanayis A, Hernell O, Lapillonne A, Mader S, Martin CR, Matthäus V, Ramakrishan U, Smuts CM, Strain SJJ, Tanjung C, Tounian P, Carlson SE. Should formula for infants provide arachidonic acid along with DHA? Am J Clin Nutr 2020; 111: 10-16 [PMID: 31665201 DOI: 10.1093/ajcn/nqz252]
- 34 Walsh V, Brown JVE, McGuire W. Iodine supplementation for the prevention of mortality and adverse neurodevelopmental outcomes in preterm infants. Cochrane Database Syst Rev 2019; 2: CD005253 [PMID: 30802939 DOI: 10.1002/14651858.CD005253.pub3]
- Cacho NT, Lawrence RM. Innate Immunity and Breast Milk. Front Immunol 2017; 8: 584 [PMID: 28611768 DOI: 10.3389/fimmu.2017.00584]
- Choudhury V, Amin SB, Agarwal A, Srivastava LM, Soni A, Saluja S. Latent iron deficiency at birth influences auditory neural maturation in late preterm and term infants. Am J Clin Nutr 2015; 102: 1030-1034 [PMID: 26310540 DOI: 10.3945/ajcn.115.113084]
- Andreas NJ, Kampmann B, Mehring Le-Doare K. Human breast milk: A review on its composition and bioactivity. Early Hum Dev 2015; 91: 629-635 [PMID: 26375355 DOI: 10.1016/j.earlhumdev.2015.08.013]
- Paramasivam K, Michie C, Opara E, Jewell AP. Human breast milk immunology: a review. Int J Fertil Womens Med 2006; 51: 208-217 [PMID: 17269587]



- Donovan S. Role of human milk components in gastrointestinal development: Current knowledge and future NEEDS. J Pediatr 2006: 149: S49-S61
- 40 Alzaree FA, AbuShady MM, Atti MA, Fathy GA, Galal EM, Ali A, Elias TR. Effect of Early Breast Milk Nutrition on Serum Insulin-Like Growth Factor-1 in Preterm Infants. Open Access Maced J Med Sci 2019; 7: 77-81 [PMID: 30740165 DOI: 10.3889/oamjms.2019.035]
- Li R, Xia W, Zhang Z, Wu K. S100B protein, brain-derived neurotrophic factor, and glial cell linederived neurotrophic factor in human milk. PLoS One 2011; 6: e21663 [PMID: 21738758 DOI: 10.1371/journal.pone.0021663]
- Koo W, Tank S, Martin S, Shi R. Human milk and neurodevelopment in children with very low birth weight: a systematic review. Nutr J 2014; 13: 94 [PMID: 25231364 DOI: 10.1186/1475-2891-13-94]
- Gibertoni D, Corvaglia L, Vandini S, Rucci P, Savini S, Alessandroni R, Sansavini A, Fantini MP, Faldella G. Positive effect of human milk feeding during NICU hospitalization on 24 mo neurodevelopment of very low birth weight infants: an Italian cohort study. PLoS One 2015; 10: e0116552 [PMID: 25590630 DOI: 10.1371/journal.pone.0116552]
- Strømmen K, Blakstad EW, Moltu SJ, Almaas AN, Westerberg AC, Amlien IK, Rønnestad AE, Nakstad B, Drevon CA, Bjørnerud A, Courivaud F, Hol PK, Veierød MB, Fjell AM, Walhovd KB, Iversen PO. Enhanced nutrient supply to very low birth weight infants is associated with improved white matter maturation and head growth. Neonatology 2015; 107: 68-75 [PMID: 25401387 DOI: 10.1159/0003681811
- Blakstad EW, Strømmen K, Moltu SJ, Wattam-Bell J, Nordheim T, Almaas AN, Grønn M, Rønnestad AE, Brække K, Iversen PO, von Hofsten C, Veierød MB, Westerberg AC, Drevon CA, Nakstad B. Improved Visual Perception in Very Low Birth Weight Infants on Enhanced Nutrient Supply. Neonatology 2015; **108**: 30-37 [PMID: 25967892 DOI: 10.1159/000381660]
- Blesa M, Sullivan G, Anblagan D, Telford EJ, Quigley AJ, Sparrow SA, Serag A, Semple SI, Bastin ME, Boardman JP. Early breast milk exposure modifies brain connectivity in preterm infants. Neuroimage 2019; **184**: 431-439 [PMID: 30240903 DOI: 10.1016/j.neuroimage.2018.09.045]
- Hård AL, Nilsson AK, Lund AM, Hansen-Pupp I, Smith LEH, Hellström A. Review shows that donor milk does not promote the growth and development of preterm infants as well as maternal milk. Acta Paediatr 2019; 108: 998-1007 [PMID: 30565323 DOI: 10.1111/apa.14702]
- Bergner EM, Shypailo R, Visuthranukul C, Hagan J, O'Donnell AR, Hawthorne KM, Abrams SA, Hair AB. Growth, Body Composition, and Neurodevelopmental Outcomes at 2 Years Among Preterm Infants Fed an Exclusive Human Milk Diet in the Neonatal Intensive Care Unit: A Pilot Study. Breastfeed Med 2020; 15: 304-311 [PMID: 32298596 DOI: 10.1089/bfm.2019.0210]
- O'Connor DL, Gibbins S, Kiss A, Bando N, Brennan-Donnan J, Ng E, Campbell DM, Vaz S, Fusch C, Asztalos E, Church P, Kelly E, Ly L, Daneman A, Unger S; GTA DoMINO Feeding Group. Effect of Supplemental Donor Human Milk Compared With Preterm Formula on Neurodevelopment of Very Low-Birth-Weight Infants at 18 Months: A Randomized Clinical Trial. JAMA 2016; 316: 1897-1905 [PMID: 27825008 DOI: 10.1001/jama.2016.16144]
- Quigley M, Embleton ND, McGuire W. Formula vs donor breast milk for feeding preterm or low birth weight infants. Cochrane Database Syst Rev 2019; 7: CD002971 [PMID: 31322731 DOI: 10.1002/14651858.CD002971.pub5]
- Walsh V, Brown JVE, Askie LM, Embleton ND, McGuire W. Nutrient-enriched formula vs standard formula for preterm infants. Cochrane Database Syst Rev 2019; 7: CD004204 [PMID: 31314903 DOI: 10.1002/14651858.CD004204.pub3]
- Li B, Wu RY, Horne RG, Ahmed A, Lee D, Robinson SC, Zhu H, Lee C, Cadete M, Johnson-Henry KC, Landberg E, Alganabi M, Abrahamsson T, Delgado-Olguin P, Pierro A, Sherman PM. Human Milk Oligosaccharides Protect against Necrotizing Enterocolitis by Activating Intestinal Cell Differentiation. Mol Nutr Food Res 2020; 64: e2000519 [PMID: 32926533 DOI: 10.1002/mnfr.202000519]
- Sodhi CP, Wipf P, Yamaguchi Y, Fulton WB, Kovler M, Niño DF, Zhou Q, Banfield E, Werts AD, Ladd MR, Buck RH, Goehring KC, Prindle T Jr, Wang S, Jia H, Lu P, Hackam DJ. The human milk oligosaccharides 2'-fucosyllactose and 6'-sialyllactose protect against the development of necrotizing enterocolitis by inhibiting toll-like receptor 4 signaling. Pediatr Res 2021; 89: 91-101 [PMID: 32221473 DOI: 10.1038/s41390-020-0852-31
- Nolan LS, Rimer JM, Good M. The Role of Human Milk Oligosaccharides and Probiotics on the Neonatal Microbiome and Risk of Necrotizing Enterocolitis: A Narrative Review. Nutrients 2020; 12 [PMID: 33036184 DOI: 10.3390/nu12103052]
- Torres Roldan VD, Urtecho S M, Gupta J, Yonemitsu C, Cárcamo CP, Bode L, Ochoa TJ. Human milk oligosaccharides and their association with late-onset neonatal sepsis in Peruvian very-low-birthweight infants. Am J Clin Nutr 2020; 112: 106-112 [PMID: 32401307 DOI: 10.1093/ajcn/nqaa102]
- Fleming SA, Mudd AT, Hauser J, Yan J, Metairon S, Steiner P, Donovan SM, Dilger RN. Human and Bovine Milk Oligosaccharides Elicit Improved Recognition Memory Concurrent With Alterations in Regional Brain Volumes and Hippocampal mRNA Expression. Front Neurosci 2020; 14: 770 [PMID: 32903658 DOI: 10.3389/fnins.2020.00770]
- Jorgensen JM, Young R, Ashorn P, Ashorn U, Chaima D, Davis JCC, Goonatilleke E, Kumwenda C, Lebrilla CB, Maleta K, Prado EL, Sadalaki J, Totten SM, Wu LD, Zivkovic AM, Dewey KG. Associations of human milk oligosaccharides and bioactive proteins with infant growth and development among Malawian mother-infant dyads. Am J Clin Nutr 2020 [PMID: 33096556 DOI: 10.1093/ajcn/nqaa272]

291

- Arboleya S, Sánchez B, Milani C, Duranti S, Solís G, Fernández N, de los Reyes-Gavilán CG, Ventura M, Margolles A, Gueimonde M. Intestinal microbiota development in preterm neonates and effect of perinatal antibiotics. J Pediatr 2015; 166: 538-544 [PMID: 25444008 DOI: 10.1016/j.jpeds.2014.09.041]
- Jia J, Xun P, Wang X, He K, Tang Q, Zhang T, Wang Y, Tang W, Lu L, Yan W, Wang W, Hu T, Cai 59 W. Impact of Postnatal Antibiotics and Parenteral Nutrition on the Gut Microbiota in Preterm Infants During Early Life. JPEN J Parenter Enteral Nutr 2020; 44: 639-654 [PMID: 31452218 DOI: 10.1002/jpen.1695]
- Keunen K, van Elburg RM, van Bel F, Benders MJ. Impact of nutrition on brain development and its neuroprotective implications following preterm birth. Pediatr Res 2015; 77: 148-155 [PMID: 25314585 DOI: 10.1038/pr.2014.171]
- Robertson C, Savva GM, Clapuci R, Jones J, Maimouni H, Brown E, Minocha A, Hall LJ, Clarke P. Incidence of necrotising enterocolitis before and after introducing routine prophylactic Lactobacillus and Bifidobacterium probiotics. Arch Dis Child Fetal Neonatal Ed 2020; 105: 380-386 [PMID: 31666311 DOI: 10.1136/archdischild-2019-317346]
- van den Akker CHP, van Goudoever JB, Shamir R, Domellöf M, Embleton ND, Hojsak I, Lapillonne A, Mihatsch WA, Berni Canani R, Bronsky J, Campoy C, Fewtrell MS, Fidler Mis N, Guarino A, Hulst JM, Indrio F, Kolaček S, Orel R, Vandenplas Y, Weizman Z, Szajewska H. Probiotics and Preterm Infants: A Position Paper by the European Society for Paediatric Gastroenterology Hepatology and Nutrition Committee on Nutrition and the European Society for Paediatric Gastroenterology Hepatology and Nutrition Working Group for Probiotics and Prebiotics. J Pediatr Gastroenterol Nutr 2020; 70: 664-680 [PMID: 32332478 DOI: 10.1097/MPG.00000000000002655]
- Raghuram K, Yang J, Church PT, Cieslak Z, Synnes A, Mukerji A, Shah PS; Canadian Neonatal Network; Canadian Neonatal Follow-Up Network Investigators. Head Growth Trajectory and Neurodevelopmental Outcomes in Preterm Neonates. Pediatrics 2017; 140 [PMID: 28759409 DOI: 10.1542/peds.2017-0216]
- Taine M, Charles MA, Beltrand J, Rozé JC, Léger J, Botton J, Heude B. Early postnatal growth and neurodevelopment in children born moderately preterm or small for gestational age at term: A systematic review. Paediatr Perinat Epidemiol 2018; 32: 268-280 [PMID: 29691880 DOI: 10.1111/ppe.12468]
- Hsu CT, Chen CH, Lin MC, Wang TM, Hsu YC. Post-discharge body weight and neurodevelopmental outcomes among very low birth weight infants in Taiwan: A nationwide cohort study. PLoS One 2018; 13: e0192574 [PMID: 29444139 DOI: 10.1371/journal.pone.0192574]
- Chien HC, Chen CH, Wang TM, Hsu YC, Lin MC. Neurodevelopmental outcomes of infants with very low birth weights are associated with the severity of their extra-uterine growth retardation. Pediatr Neonatol 2018; 59: 168-175 [PMID: 28866004 DOI: 10.1016/j.pedneo.2017.08.003]
- Paviotti G, De Cunto A, Zennaro F, Boz G, Travan L, Cont G, Bua J, Demarini S. Higher growth, fat and fat-free masses correlate with larger cerebellar volumes in preterm infants at term. Acta Paediatr 2017; 106: 918-925 [PMID: 28295577 DOI: 10.1111/apa.13829]
- Ramel SE, Gray HL, Christiansen E, Boys C, Georgieff MK, Demerath EW. Greater Early Gains in Fat-Free Mass, but Not Fat Mass, Are Associated with Improved Neurodevelopment at 1 Year Corrected Age for Prematurity in Very Low Birth Weight Preterm Infants. J Pediatr 2016; 173: 108-115 [PMID: 27056450 DOI: 10.1016/j.jpeds.2016.03.003]
- Frondas-Chauty A, Simon L, Flamant C, Hanf M, Darmaun D, Rozé JC. Deficit of Fat Free Mass in Very Preterm Infants at Discharge is Associated with Neurological Impairment at Age 2 Years. J Pediatr 2018; 196: 301-304 [PMID: 29336797 DOI: 10.1016/j.jpeds.2017.12.017]
- Hickey M, Georgieff M, Ramel S. Neurodevelopmental outcomes following necrotizing enterocolitis. Semin Fetal Neonatal Med 2018; 23: 426-432 [PMID: 30145060 DOI: 10.1016/j.siny.2018.08.005]
- 71 Adams-Chapman I. Necrotizing Enterocolitis and Neurodevelopmental Outcome. Clin Perinatol 2018; **45**: 453-466 [PMID: 30144849 DOI: 10.1016/j.clp.2018.05.014]
- Hsiao CC, Tsai ML, Chen CC, Lin HC. Early optimal nutrition improves neurodevelopmental 72 outcomes for very preterm infants. Nutr Rev 2014; 72: 532-540 [PMID: 24938866 DOI: 10.1111/nure.12110]
- So S, Patterson C, Gold A, Rogers A, Belza C, de Silva N, Avitzur Y, Wales PW. Neurodevelopmental outcomes of infants with intestinal failure at 12 and 26 mo corrected age. Early Hum Dev 2019; 130: 38-43 [PMID: 30660017 DOI: 10.1016/j.earlhumdev.2018.12.020]
- Lucas A, Morley R, Cole TJ. Randomised trial of early diet in preterm babies and later intelligence quotient. BMJ 1998; 317: 1481-1487 [PMID: 9831573 DOI: 10.1136/bmj.317.7171.1481]
- Hiltunen H, Löyttyniemi E, Isolauri E, Rautava S. Early Nutrition and Growth until the Corrected Age of 2 Years in Extremely Preterm Infants. Neonatology 2018; 113: 100-107 [PMID: 29131014 DOI: 10.1159/0004806331
- Strodtbeck F. The role of early enteral nutrition in protecting premature infants from sepsis. Crit Care Nurs Clin North Am 2003; 15: 79-87 [PMID: 12597043 DOI: 10.1016/s0899-5885(02)00043-6]
- Hay WW. Optimizing nutrition of the preterm infant. Zhongguo Dangdai Erke Zazhi 2017; 19: 1-21
- Kwok TC, Dorling J, Gale C. Early enteral feeding in preterm infants. Semin Perinatol 2019; 43: 151159 [PMID: 31443906 DOI: 10.1053/j.semperi.2019.06.007]
- Ng DV, Brennan-Donnan J, Unger S, Bando N, Gibbins S, Nash A, Kiss A, O'Connor DL. How



Close Are We to Achieving Energy and Nutrient Goals for Very Low Birth Weight Infants in the First Week? JPEN J Parenter Enteral Nutr 2017; 41: 500-506 [PMID: 26160253 DOI: 10.1177/0148607115594674]





Published by Baishideng Publishing Group Inc

7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: https://www.f6publishing.com/helpdesk

https://www.wjgnet.com

