

Lactobacillus paracasei HP7 with *Portulaca oleracea* Linn. Alleviates Scopolamine-Induced Cognitive Decline via Regulation of Neurotrophic Factor and Inflammation Signals in Mice

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ABSTRACT: People often experience cognitive deterioration of various degrees, from early-stage mild cognitive impairment to severe cognitive decline. Cognitive deterioration is related to many diseases and studied to alleviated inflammation reaction or oxidative stress. In the present study, the levels of various memory-related proteins: brain-derived neurotrophic factor (BDNF), amyloid beta (A β) 42, A β 40, interleukin-6 and tumor necrosis factor-alpha were measured. Among *Lactobacillus paracasei* HP7 (HP7), *Portulaca oleracea* Linn. (PO) and HP7 together with PO (HP7A), the HP7A group had the best effect on increasing BDNF expression and suppressing A β 40 expression. Also, we measured the protective effect on scopolamine-induced cognitive decline in mice. In the acquisition test, the HP7A group most reliably relieved cognitive decline from days 2 to 5 of scopolamine injection. When the probe test was performed on the day 6 of scopolamine injection, the HP7A group had the shortest escape latency. Based on the results of the Morris water maze tasks, we suggest that HP7A is most useful for ameliorating cognitive decline. It is suggested that the HP7A ameliorating scopolamine-induced cognitive decline via the increase of BDNF expression and the suppression of A β 40 expression.

Keywords: cognitive decline, inflammation, *Lactobacillus*, neurotrophic factor, *Portulaca oleracea* Linn.

INTRODUCTION

Cognitive deterioration with human aging is related to many diseases characterized by chronic processes, such as inflammation or oxidative stress (Lu et al., 2014; Paniz et al., 2017). There has been showed a critical role of inflammation in cognitive ability or memory deficits, since cognitive deteriorations like Alzheimer disorders have relation with up-regulated inflammatory cytokine levels (Barrientos et al., 2009). Moreover, according to Lu et al. (2014), deficits of brain-derived neurotrophic factor (BDNF) lead to the deterioration of brain disorders, such as cognitive decline and up-regulated BDNF level can inhibit the amyloid beta (A β) accumulation, which lead to memory loss and learning disabilities (Eckert et al., 2003; Arancibia et al., 2008; Lu et al., 2014). Thus, the development of drugs or supplements for cognitive decline is becoming ever more urgent.

Recent evidence has revealed that probiotics can improve cognitive function by alleviating increased inflammatory cytokine level. Namely, *Lactobacillus pentosus* has been found to improve age-related memory impairment

and scopolamine-induced cognitive impairment *in vivo* (Jeong et al., 2015). *Lactobacillus paracasei* has been reported to prevent age-related cognitive decline in senescent accelerated prone 8 mice (Corpuz et al., 2018; Huang et al., 2018). According to Yun et al. (2020), probiotics including *Lactobacillus* have alleviative effect against cognitive decline by regulating inflammatory cytokine such as interleukin (IL)-1 β , IL-6, or tumor necrosis factor (TNF) expression.

Furthermore, *Portulaca oleracea* Linn. (PO) is well known for its cognition-enhancing properties (Sumathi and Christinal, 2016; Noorbakhshnia and Karimi-Zandi, 2017; Wang et al., 2017). This PO has been termed the 'Global Panacea' and listed as one of the most-used medicinal plants by the World Health Organization (Lim and Quah, 2017). As the nickname 'longevity vegetable' suggests, pharmacological studies have demonstrated that this plant has anti-aging and cognition-improving properties (Hongxing et al., 2007). PO, like prebiotics, also has the effect of increasing the amount of *Lactobacillus* in the broiler caecum (Zhao et al., 2013). However, the synergistic effects of probiotics with PO on cognitive impair-

Received 10 August 2022; Revised 28 September 2022; Accepted 1 October 2022; Published online 31 December 2022

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ment or anti-inflammatory effects have not been studied. Also, when they mixed or administered together, the BDNF or A β level which have critical relationship with cognitive decline was not reported yet.

Therefore, this study aims to investigate the synergistic effect of alleviative effect of inflammation as well as regulatory effects of neurotrophic factor of HP7A [*L. paracasei* HP7 (HP7) together with PO], and to confirm the cognitive improvement effect in mice using Morris water maze tasks.

MATERIALS AND METHODS

Reagents and equipment

Chemicals and assay kits used in this study include scopolamine, corticosterone (Sigma-Aldrich Co., St. Louis, MO, USA), mouse A β 40, A β 42, BDNF, TNF- α and IL-6 enzyme-linked immunosorbent assay (ELISA) kit (R&D Systems, Minneapolis, MN, USA), human IL-6 and TNF ELISA kit (BD Biosciences, Franklin Lakes, NJ, USA). Cell viability assay kit (Precaregene, Anyang, Korea) reagent was used for assay cell proliferation.

The water extract preparation of *L. paracasei* HP7, PO, and HP7A

HP7 was cultured for 24 h at 35°C in Man-Rogosa-Sharpe broth (Difco Laboratories, Detroit, MI, USA). After drying, the powdered HP7 was stored at -20°C until further use. For *in vivo* assays, HP7 was resuspended at the concentration of 1×10^9 colony forming unit (CFU)/mL in sterile phosphate-buffered saline. To prepare *P. oleracea* extract, dried aerial part of PO was purchased from Humanherb (Daegu, Korea). To prepare the water extract for the animal experiments, the dried sliced aerial part of PO was refluxed by ten to twenty times the weight (w/w) of water for 6 to 16 h. The extracts were concentrated and dried to yield PO water extracts. For *in vivo* assays, PO extract was resuspended at the concentration of 20 mg/mL in sterile phosphate-buffered saline. In order to prepare HP7A, HP7 was resuspended at the concentration of 1×10^9 CFU/mL in sterile phosphate-buffered saline and PO extract was resuspended altogether at the concentration of 20 mg/mL in sterile phosphate-buffered saline.

Cell cultures

The human neuroblastoma cells (SK-N-SH) were cultured in Eagle's minimum essential medium containing 1% antibiotic solution (Gibco, Carlsbad, CA, USA) and 10% heat-inactivated fetal bovine serum in 5% CO $_2$ at 37°C. For *in vitro* assay, cells (1×10^5 cells/mL) were incubated in 6 well plates with corticosterone (250 μ M) in the presence of samples, HP7 (1×10^6 CFU/well), PO (50 μ g/mL), or HP7A (1×10^6 CFU HP7 with 100 μ g/mL PO/well) (Table 1). The culture supernatants were recovered for analysis by ELISA.

Animals and experiment design

Five weeks old male-C57BL/6 mice were purchased from Orient Bio (Seongnam, Korea). Animals were maintained under controlled climatic conditions (21.2 ~ 23.6°C, 43 ~ 65% relative humidity, 12-h light cycle with lights on during 07:00 ~ 19:00). The animal protocol in this study was reviewed and approved based on ethical procedures and scientific care by the Ethics Committee at Chaon Corp. (Yongin, Korea; IACUC no. CE21047). The mice were divided into control, scopolamine, donepezil, HP7, PO, and HP7A groups, with six mice in each group. Donepezil is a medicine that treats some types of dementia. It alleviates mental dysfunctions such as memory-deficit. Mice in the donepezil, HP7, PO, and HP7A group orally received 5 mg/kg of donepezil (Su et al., 2011), 1×10^8 CFU/mice *L. paracasei* HP7, 100 mg/kg PO, and 1×10^8 CFU/mice *L. paracasei* HP7 with 100 mg/kg PO, respectively, for 13 days (Table 2). On the day 8, all mice except those in the control group started to be injected intraperitoneally with scopolamine (1 mg/kg) 1 h before behavioral tests. After the final behavioral test, mice were sacrificed and hippocampal tissues were collected. The schedule of animal experiments is shown in Fig. 1.

Morris water maze task

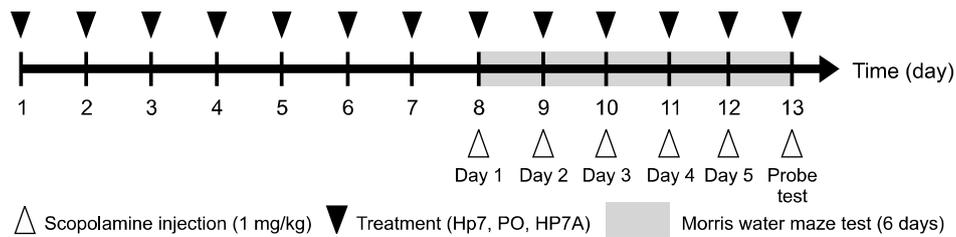
The Morris water maze is commonly used to investigate spatial memory and learning. A circular swimming pool which is 90 cm in diameter and 45 cm in height is filled with opaque water mixed with 500 mL of milk to a depth of 30 cm. The water temperature was maintained at $20 \pm 1^\circ$ C. There were four sectors marked by white platforms which are 6 cm in diameter and 29 cm in height in the pool. The white platforms were invisible as they were lo-

Table 1. The experimental groups in SK-N-SH cells for *in vitro*

Group	Treatment	Corticosterone (250 μ M)
Control	Vehicle	(-)
Corticosterone	Corticosterone (250 μ M)	(+)
HP7	<i>Lactobacillus paracasei</i> HP7 (1×10^6 CFU/well)	(+)
PO	<i>Portulaca oleracea</i> Linn. (50 μ g/mL)	(+)
HP7A	<i>L. paracasei</i> HP7 (1×10^6 CFU/well) added to <i>P. oleracea</i> Linn. (100 μ g/mL)	(+)

Table 2. The animal experimental groups for *in vivo*

Group	Treatment	No. of mice	Scopolamine (1 mg/kg)
Control	Vehicle	6	(-)
Scopolamine	Scopolamine (1 mg/kg)	6	(+)
Donepezil	Donepezil (5 mg/kg)	6	(+)
HP7	<i>Lactobacillus paracasei</i> HP7 (1×10^8 CFU/mice)	6	(+)
PO	<i>Portulaca oleracea</i> Linn. (100 mg/kg)	6	(+)
HP7A	<i>L. paracasei</i> HP7 (1×10^8 CFU/mice) added to <i>P. oleracea</i> Linn. (100 mg/kg)	6	(+)

**Fig. 1.** Experimental design of HP7, PO, and HP7A treatment in scopolamine injected mice. HP7, PO, and HP7A were treated for 7 days before Morris water maze test. Probe test was done after scopolamine was treated for 5 days. HP7, *Lactobacillus paracasei* HP7; PO, *Portulaca oleracea* Linn.; HP7A, HP7 together with PO.

cated 1 cm below the surface of the water. Mice were trained for 60 s on the first day of the experiment. During next 4 days after the training, the learning trials were conducted 4 times per a day; the mouse was allowed to stay on the platform for 10 s either when it found a platform or when it did not find within 60 s. Each trial was performed with inter-trial interval of 30 s. Mice were dried in their cages under an infrared lamp between trials. The latency time was recorded while mice reached to the invisible platform using a Smart 3.0 video tracking system (Panlab, Barcelona, Spain). During 4 trials in a day, mice faced each quadrant of the pool wall. The probe trial session was done after the last training trial period. Mice were placed in the pool without the platform to swim and find a platform for 60 s. Moreover, target crossing time, the motion trail of the mouse and the swimming time in the quadrant were recorded.

Tissue analysis

Hippocampus samples were washed with cold phosphate-buffered saline and homogenized on ice using a bead homogenizer. The samples were then subjected to two freeze-thaw cycles to further degrade cell membranes and the supernatants were collected by centrifugation at 16,000 g for min. After all sample concentrations had been equalized, these samples were assayed immediately or stored at -20°C until further use.

Statistical analysis

Datasets are presented as the mean \pm standard error of the mean. Between-group differences were evaluated using Dunnett's multiple comparison test with one-way or two-way ANOVAs, and were deemed statistically significant at $P < 0.05$.

RESULTS

The effect of HP7A on levels of inflammatory cytokines and neuroblastoma cell proliferation in *in vitro* assays

We investigated the inhibitory action of HP7, PO, and HP7A on the levels of IL-6 and TNF production in SK-N-SH cells (Fig. 2). HP7A treatment demonstrated inhibition of IL-6 and TNF production in SK-N-SH cell induced by corticosterone ($P < 0.05$). HP7A treatment also showed protective effects on neuroblast cell proliferation in SK-N-SH cells induced by corticosterone ($P < 0.05$). Furthermore, the synergistic effect of HP7A on recovery of IL-6 and TNF was observed.

The effect of HP7, PO, and HP7A on the hippocampus

We measured the effects of donepezil, HP7, PO, and HP7A on BDNF, A β 42, and A β 40 protein expression in the hippocampus (Fig. 3). The BDNF protein expression level in the hippocampus was reduced from 509.3 pg/mL in the control group to 284.8 pg/mL in the scopolamine group ($P < 0.001$). These levels increased to 403.0 pg/mL, 404.7 pg/mL, and 423.3 pg/mL in the donepezil ($P < 0.05$), HP7 ($P < 0.05$), and HP7A ($P < 0.01$; Fig. 3A) group, respectively. We also observed that A β 42 and A β 40 protein expression levels in the hippocampus increased from 19.01 pg/mL and 37.72 pg/mL in the control group, respectively, to 35.78 pg/mL and 72.35 pg/mL, respectively, in the scopolamine treated group ($P < 0.05$). In terms of the A β 42 expression level, only the HP7 group exhibited a significant decrease to 22.60 pg/mL ($P < 0.05$; Fig. 3B). The A β 40 expression levels decreased to 39.23 pg/mL, 40.95 pg/mL, and 38.23 pg/mL in the HP7 ($P < 0.05$), PO ($P < 0.05$), and HP7A ($P < 0.01$; Fig. 3C) groups, respectively. We measured the effects of donepezil, HP7,

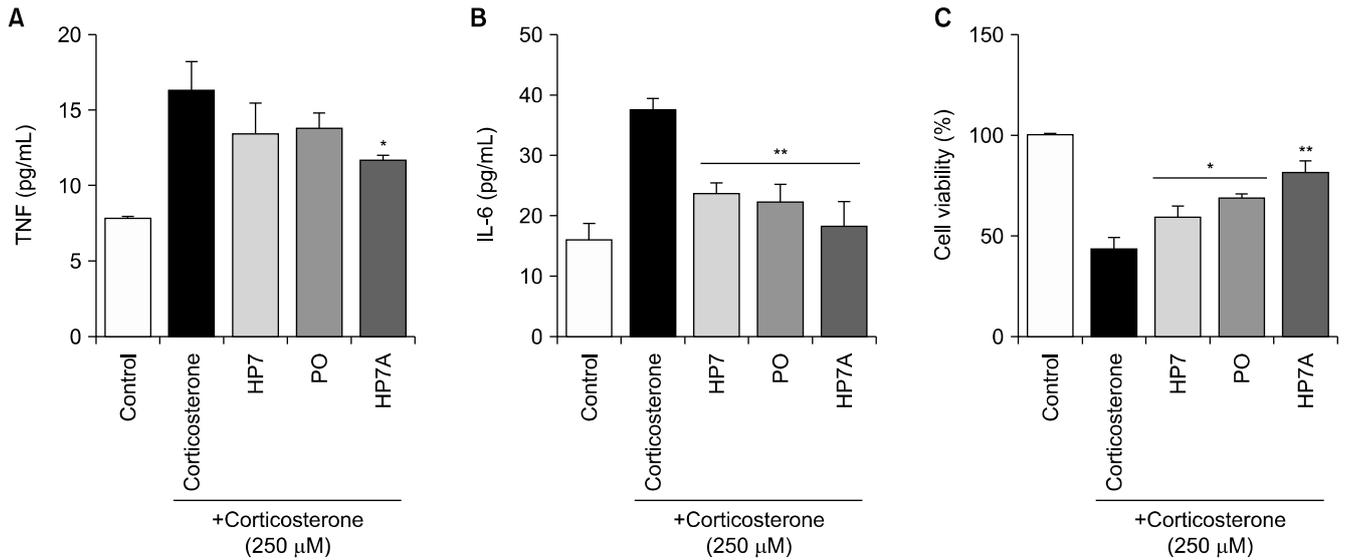


Fig. 2. The effects of HP7, PO, and HP7A on corticosterone-stimulated SK-N-SH assay. Tumor necrosis factor (TNF) (A) and interleukin (IL)-6 (B) concentrations were assessed using ELISA and neuroblastoma cell proliferation (C) was measured by using cell viability assay kit. Neuroblastoma cells stimulated with corticosterone except the control group. * $P < 0.05$ and ** $P < 0.01$ vs. corticosterone-treated group. Control, vehicle treated cell; Corticosterone, corticosterone 250 μM ; HP7, corticosterone-treated cell with *Lactobacillus paracasei* HP7 (1×10^6 CFU/well); PO, corticosterone-treated cell with *Portulaca oleracea* Linn. (50 $\mu\text{g}/\text{mL}$); HP7A, HP7 (1×10^6 CFU/well) added to PO (100 $\mu\text{g}/\text{mL}$); CFU, colony forming unit.

PO, and HP7A on IL-6 and TNF- α protein expression in the hippocampus (Fig. 4). The levels of inflammatory markers IL-6 and TNF- α in the hippocampus increased from 5.89 pg/mL and 27.79 pg/mL in the control group to 14.23 pg/mL and 340.0 pg/mL, respectively, in the scopolamine group ($P < 0.05$ and $P < 0.01$, respectively). HP7 was effective in reducing IL-6 and TNF- α levels to 7.06 pg/mL and 96.57 pg/mL, which were similar to

those of the control group ($P < 0.05$).

The effect of HP7, PO, and HP7A on acquisition test for cognitive performance

As shown in Fig. 5A, the escape latency of the five groups tended to decrease from days 1 to day 4, which indicates that training improved learning capacity. On days 2, 3, and 4, escape latencies in the scopolamine group in-

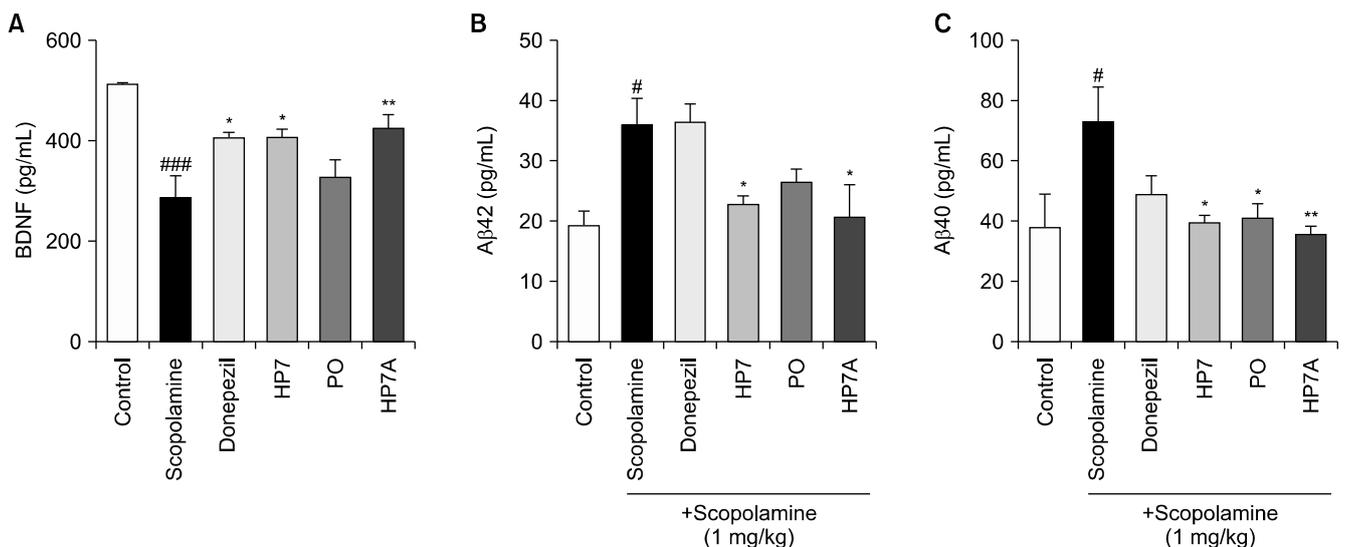


Fig. 3. The effects of HP7, PO, and HP7A on hippocampal brain-derived neurotrophic factor (BDNF), amyloid beta (A β) 42, and A β 40 levels. Mouse hippocampus samples were collected after induced by scopolamine and BDNF (A), A β 42 (B), and A β 40 (C) concentrations were assessed using ELISA. Mice were orally administered test agents or an equal volume of vehicle once daily for 13 days. # $P < 0.05$ and ### $P < 0.001$ vs. control group; * $P < 0.05$ and ** $P < 0.01$ vs. scopolamine-treated group. Control, mice treated with vehicle; Scopolamine, scopolamine-injected mice treated with vehicle; Donepezil, scopolamine-injected mice treated with donepezil (5 mg/kg); HP7, scopolamine-injected mice treated with *Lactobacillus paracasei* HP7 (1×10^8 CFU/mice); PO, scopolamine-injected mice treated with *Portulaca oleracea* Linn. (100 mg/kg); HP7A, HP7 (1×10^8 CFU/mice) added to PO (100 mg/kg); CFU, colony forming unit.

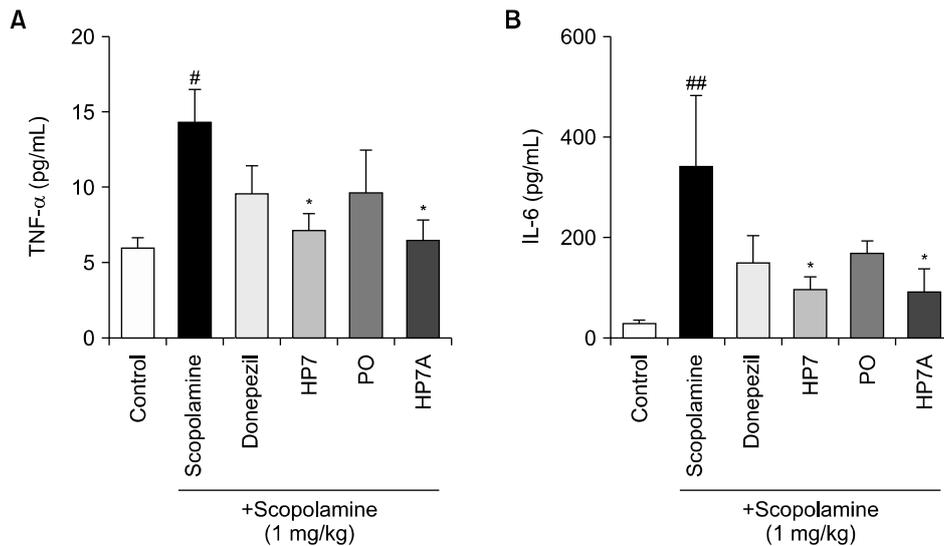


Fig. 4. The effects of HP7, PO, and HP7A on hippocampal interleukin (IL)-6 and tumor necrosis factor (TNF)- α levels. Mouse hippocampus samples were collected after induced by scopolamine, and TNF- α (A) and IL-6 (B) were assessed by ELISA. Mice were orally administered test agents or an equal volume of vehicle once daily for 13 days. [#] $P < 0.05$ and ^{##} $P < 0.01$ vs. control group; * $P < 0.05$ vs. scopolamine-treated group. Control, mice treated with vehicle; Scopolamine, scopolamine-injected mice treated with vehicle; Donepezil, scopolamine-injected mice treated with donepezil (5 mg/kg); HP7, scopolamine-injected mice treated with *Lactobacillus paracasei* HP7 (1×10^8 CFU/mice); PO, scopolamine-injected mice treated with *Portulaca oleracea* Linn. (100 mg/kg); HP7A, HP7 (1×10^8 CFU/mice) added to PO (100 mg/kg); CFU, colony forming unit.

creased to 1.95, 2.09, and 4.44 times of those of the control group ($P < 0.01$, $P < 0.001$, and $P < 0.001$, respectively), which indicated that chronic treatment of scopolamine impaired the learning capacity of mice. The average escape latency of the donepezil, HP7, and HP7A groups on day 2 reduced to 86%, 87%, and 72% of that of the scopolamine group; however, only the PO group reached significance, at 70% ($P < 0.05$). Compared to the scopolamine group, the average escape latency of the donepezil and PO groups on day 3 reduced to 70% and 69%; however, only the HP7 and HP7A groups reached the significance levels of 57% and 60% ($P < 0.01$). Compared to the scopolamine group, the average escape latency of the PO group on day 4 reduced to 73%; however, only the donepezil, HP7, and HP7A groups reached the significance levels of 52%, 51%, and 50% ($P < 0.01$), which indicates that HP7 and HP7A can enhance the learning capacity of mice.

The effect of HP7, PO, and HP7A on probe test for cognitive performance

We measured the effects of donepezil, HP7, PO, and HP7A on the escape latency in the probe test of the Morris water maze task (Fig. 5B). Scopolamine increased the escape latencies to 41.09 s, which indicates that repetitive treatment of scopolamine impairs the spatial memory capacity of mice. Donepezil, HP7, PO, and HP7A reduced the escape latency to 14.69 s, 18.40 s, 13.80 s, and 10.99 s on the probe test, respectively, which indicates that HP7, PO, and HP7A alleviate spatial memory impairment, similar to donepezil ($P < 0.01$, $P < 0.05$, $P < 0.01$,

and $P < 0.01$, respectively). Scopolamine decreased target crossings to 23% in the probe test. Donepezil, HP7, PO, and HP7A restored the target crossings to 91%, 73%, 77%, and 82%, but these were not significant (Fig. 5C). As shown in Fig. 5D and 5E, the time spent in the target platform position decreased non-significantly to 31.01% in scopolamine-treated group, compared to 52.88% in the control group. Compared to the scopolamine-treated mice, the time donepezil, HP7, PO, and HP7A groups spent in the target platform position quadrant non-significantly increased to 48.06%, 36.4%, 39.65%, and 39.57%.

DISCUSSION

Cholinergic disorder is an important feature of Alzheimer's disease pathology (Sarter and Bruno, 2004; Ishrat et al., 2009), since it can lead to memory loss and cognitive impairment over the years (Terry and Buccafusco, 2003). Previous reported that scopolamine is an anti-cholinergic drug, but it can cause memory loss in healthy young subjects mimicking the memory impairment observed in elderly people not under treatment with anti-dementia drugs (Tariot et al., 1996). Thus, we investigated the anti-inflammatory effects of HP7, PO, and HP7A on scopolamine-induced cognitive decline in mice as well as the protective effects of neurotrophic factor, such as BDNF.

Numerous studies have explored that BDNF plays a critical role in memory and learning (Hall et al., 2000; Mizuno et al., 2000; Alonso et al., 2002; Broad et al.,

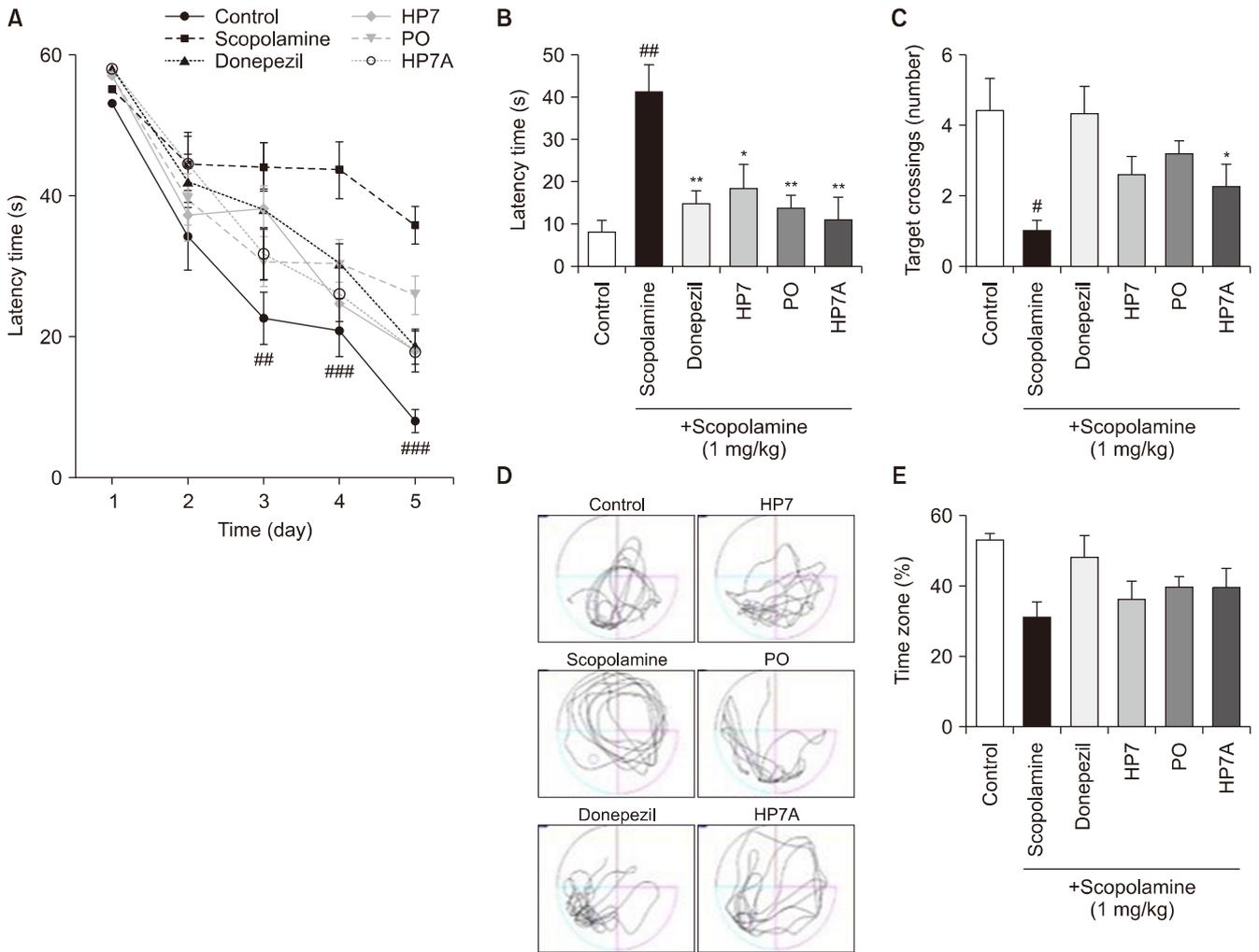


Fig. 5. The effect of HP7, PO, and HP7A on cognitive performance in Morris water maze tests. (A) Latency time (s) in the acquisition test. (B) Latency time (s), (C) target crossing, (D) motion trails of mice, and (E) time in zone (%) in the probe test. All tests were carried out 1 h after the administration of test agents or vehicle. $^{\#}P < 0.05$, $^{\#\#}P < 0.01$, $^{\#\#\#}P < 0.001$ vs. control group; $^*P < 0.05$ and $^{**}P < 0.01$ vs. scopolamine-treated group. Control (●), mice treated with vehicle; Scopolamine (■), scopolamine-injected mice treated with vehicle; Donepezil (▲), scopolamine-injected mice treated with donepezil (5 mg/kg); HP7 (◆), scopolamine-injected mice treated with *Lactobacillus paracasei* HP7 (1×10^8 CFU/mice); PO (▼), scopolamine-injected mice treated with *Portulaca oleracea* Linn. (100 mg/kg); HP7A (○), HP7 (1×10^8 CFU/mice) added to PO (100 mg/kg); CFU, colony forming unit.

2002; Ma et al., 2011). However, this is the first study to examine BDNF expression levels in the hippocampus after HP7 or PO administration in mice. BDNF promotes synaptic transmission and plasticity, which play a key role in memory formation and storage (Park et al., 2012). BDNF participates in the formation of long-term potentiation by increasing N-methyl-D-aspartate receptor sensitivity (Figurov et al., 1996; Madara and Levine, 2008). Our results showed that the HP7A group almost restored hippocampal BDNF levels in scopolamine-injected mice, which suggests that HP7A ameliorates scopolamine-induced cognitive decline by the increase of BDNF expression.

According to a recent study, chronic administration of scopolamine results in an increase of A β levels in brain (Hernández-Rodríguez et al., 2020). Amyloid plaques are one of the main remarks of Alzheimer's disease. The sol-

uble A β molecules consisted of 40- or 42-residue peptide transforms into plaque-associated fibers (Walsh et al., 1997). The A β 42 variant is more hydrophobic and tends to aggregate easily to develop fibrillogenesis than A β 40. A β 40 takes more various conformational structures than A β 42, which attribute to the oligomerization pathway and detection time during aggregation (Vestergaard et al., 2005). A β accumulation results in cell death, which in turn leads to memory loss and learning disabilities (McGlenon et al., 1999; Eckert et al., 2003). However, it has been reported that increased BDNF signaling protects cells from A β deposition and improves cognitive decline (Arancibia et al., 2008; Moghbelinejad et al., 2014). We measured A β concentrations in hippocampal tissue. The HP7A group most markedly reduced A β 40 levels, compared to the scopolamine group. These results demonstrate that HP7A can suppress A β accumulation by in-

creasing BDNF expression, as previously reported (Koh et al., 2018).

In the acquisition test, the HP7A group most reliably relieved cognitive decline from day 2 to 5. Compared to the scopolamine group on the day 6 for probe test, the HP7A group had the shortest escape latency. Although the results of target crossing and time in zone were not significant, our results suggest that HP7A can most alleviate spatial memory impairments and excellently increase the learning capacity of mice. Next, we measured the levels of various memory-related proteins: BDNF, A β 42, A β 40, TNF- α , and IL-6.

We confirmed the recovery effects of HP7, PO and HP7A on inflammatory cytokine such as TNF and IL-6 in corticosterone stimulated neuroblastoma cell, SK-N-SH (Kim et al., 2018). HP7 and PO also showed protective effects of neuroblastoma cell proliferation that stimulated by corticosterone, respectively. Interestingly, HP7A showed synergic effect on recovery of inflammatory cytokine as well as neuroblastoma cell protective effects on corticosterone stimulation. There is growing evidence that neuritis can interfere with cognition (Li et al., 2012; Allison and Ditor, 2014; Moon et al., 2014). Neuritis also plays an important role in neurodegenerative diseases such as Alzheimer's disease and ischemic stroke (DeLegge and Smoke, 2008; Hovens et al., 2014). TNF- α is a key pro-inflammatory cytokine that induces the secretion of other cytokines (Qin et al., 2008). Previous studies have revealed that inflammatory mediators stimulate apoptosis pathways and neuronal cell death (Semmler et al., 2007; Belarbi et al., 2012). It has been found that cytokines such as TNF- α interfere with long-term potentiation and neurogenesis in the hippocampus (Kim and Diamond, 2002; Monje et al., 2003). It has also been demonstrated that increased production of IL-6 leads to the dysfunction of neuronal stem cells and the inhibition of neurogenesis, resulting in impaired cognition and learning abilities (Monje et al., 2003).

Collectively, our findings suggest HP7A, a mixture of *L. paracasei* HP7 and PO, showed synergies of alleviative effects on inflammatory cytokines or protective effect in neuroblastoma cell. In addition, HP7A administration improved cognitive decline in scopolamine-induced mice via regulation of neurotrophic factor, BDNF and suppression of inflammatory signals, such as TNF, IL-6 and A β 40 or 42 level in hippocampus. Also, HP7A alleviated cognitive performance on Morris water maze test.

These health-promoting effect of the administration of HP7A can be related not only to the viability of probiotics but also to gut microbiota. According to the previous study, the combination of probiotics and plant extracts can improve the viability of probiotics during storage (Michael et al., 2015). It is possible that PO enhanced the viability of HP7, resulting in HP7A showed greater effica-

cy than either PO or HP7. The combination of probiotics and prebiotics is called synbiotics when there is a proven health-benefit in co-administration (Bilal et al., 2022). The other possible mechanism of the synbiotics of HP7A may be relevant to gut microbiota. Supplementation with synbiotics can affect gut microbiota and regulate microbial metabolites including short chain fatty acids and secondary metabolites of plant extracts which are small organic molecules beneficial to gut health (Russell and Duthie, 2011; Bilal et al., 2022). However, further study for underlying mechanism of synergistic effect of HP7A remains the subject of investigation.

FUNDING

This work was supported by the Korea Institute of Planning and Evaluation for Technology in Food, Agriculture, Forestry (IPET) through the High Value-added Food Technology Development Program, funded by the Ministry of Agriculture, Food and Rural Affairs (MAFRA) (grant no. 318027-04).

AUTHOR DISCLOSURE STATEMENT

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

Conceptualization: JHK, JHR, SDP, JJS, JLL. Methodology: JHK, JHR. Software: JHK, JHR. Validation: JHK, HK. Formal analysis: JHK. Investigation: JHK, JHR. Writing – original draft preparation: JHK. Writing review and editing: SDP. Visualization: JHK. All authors have read and agreed to the published version of the manuscript.

REFERENCES

- Allison DJ, Ditor DS. The common inflammatory etiology of depression and cognitive impairment: a therapeutic target. *J Neuroinflammation*. 2014. 11:151. <https://doi.org/10.1186/s12974-014-0151-1>
- Alonso M, Vianna MR, Depino AM, Mello e Souza T, Pereira P, Szapiro G, et al. BDNF-triggered events in the rat hippocampus are required for both short- and long-term memory formation. *Hippocampus*. 2002. 12:551-560.
- Arancibia S, Silhol M, Moulière F, Meffre J, Höllinger I, Maurice T, et al. Protective effect of BDNF against beta-amyloid induced neurotoxicity *in vitro* and *in vivo* in rats. *Neurobiol Dis*. 2008. 31:316-326.
- Barrientos RM, Frank MG, Hein AM, Higgins EA, Watkins LR, Rudy JW, et al. Time course of hippocampal IL-1 β and memory consolidation impairments in aging rats following peripheral infection. *Brain Behav Immun*. 2009. 23:46-54.

- Belarbi K, Jopson T, Tweedie D, Arellano C, Luo W, Greig NH, et al. TNF- α protein synthesis inhibitor restores neuronal function and reverses cognitive deficits induced by chronic neuroinflammation. *J Neuroinflammation*. 2012. 9:23. <https://doi.org/10.1186/1742-2094-9-23>
- Bilal M, Ashraf S, Zhao X. Dietary component-induced inflammation and its amelioration by prebiotics, probiotics, and synbiotics. *Front Nutr*. 2022. 9:931458. <https://doi.org/10.3389/fnut.2022.931458>
- Broad KD, Mimmack ML, Keverne EB, Kendrick KM. Increased BDNF and trk-B mRNA expression in cortical and limbic regions following formation of a social recognition memory. *Eur J Neurosci*. 2002. 16:2166-2174.
- Corpuz HM, Ichikawa S, Arimura M, Mihara T, Kumagai T, Mitani T, et al. Long-term diet supplementation with *Lactobacillus paracasei* K71 prevents age-related cognitive decline in senescence-accelerated mouse prone 8. *Nutrients*. 2018. 10:762. <https://doi.org/10.3390/nu10060762>
- DeLegge MH, Smoke A. Neurodegeneration and inflammation. *Nutr Clin Pract*. 2008. 23:35-41.
- Eckert A, Keil U, Marques CA, Bonert A, Frey C, Schüssel K, et al. Mitochondrial dysfunction, apoptotic cell death, and Alzheimer's disease. *Biochem Pharmacol*. 2003. 66:1627-1634.
- Figurov A, Pozzo-Miller LD, Olafsson P, Wang T, Lu B. Regulation of synaptic responses to high-frequency stimulation and LTP by neurotrophins in the hippocampus. *Nature*. 1996. 381:706-709.
- Hall J, Thomas KL, Everitt BJ. Rapid and selective induction of BDNF expression in the hippocampus during contextual learning. *Nat Neurosci*. 2000. 3:533-535.
- Hernández-Rodríguez M, Arciniega-Martínez IM, García-Marín ID, Correa-Basurto J, Rosales-Hernández MC. Chronic administration of scopolamine increased GSK3 β P9, beta secretase, amyloid beta, and oxidative stress in the hippocampus of Wistar rats. *Mol Neurobiol*. 2020. 57:3979-3988.
- Hongxing Z, Nancai Y, Guofu H, Jianbo S, Yanxia W, Hanju H, et al. Neuroprotective effects of purslane herb aqueous extracts against D-galactose induced neurotoxicity. *Chem Biol Interact*. 2007. 170:145-152.
- Hovens IB, Schoemaker RG, van der Zee EA, Absalom AR, Heine-man E, van Leeuwen BL. Postoperative cognitive dysfunction: Involvement of neuroinflammation and neuronal functioning. *Brain Behav Immun*. 2014. 38:202-210.
- Huang SY, Chen LH, Wang MF, Hsu CC, Chan CH, Li JX, et al. *Lactobacillus paracasei* PS23 delays progression of age-related cognitive decline in senescence accelerated mouse prone 8 (SAMP8) mice. *Nutrients*. 2018. 10:894. <https://doi.org/10.3390/nu10070894>
- Ishrat T, Hoda MN, Khan MB, Yousuf S, Ahmad M, Khan MM, et al. Amelioration of cognitive deficits and neurodegeneration by curcumin in rat model of sporadic dementia of Alzheimer's type (SDAT). *Eur Neuropsychopharmacol*. 2009. 19:636-647.
- Jeong JJ, Woo JY, Kim KA, Han MJ, Kim DH. *Lactobacillus pentosus* var. *plantarum* C29 ameliorates age-dependent memory impairment in Fischer 344 rats. *Lett Appl Microbiol*. 2015. 60:307-314.
- Kim DH, Jeon SJ, Son KH, Jung JW, Lee S, Yoon BH, et al. The ameliorating effect of oroxylin A on scopolamine-induced memory impairment in mice. *Neurobiol Learn Mem*. 2007. 87:536-546.
- Kim JJ, Diamond DM. The stressed hippocampus, synaptic plasticity and lost memories. *Nat Rev Neurosci*. 2002. 3:453-462.
- Kim YH, Im AR, Park BK, Paek SH, Choi G, Kim YR, et al. Anti-depressant-like and neuroprotective effects of ethanol extract from the root bark of *Hibiscus syriacus* L. *Biomed Res Int*. 2018. 2018:7383869. <https://doi.org/10.1155/2018/7383869>
- Koh EJ, Kim KJ, Choi J, Kang DH, Lee BY. *Spirulina maxima* extract prevents cell death through BDNF activation against amyloid beta 1-42 (A β ₁₋₄₂) induced neurotoxicity in PC12 cells. *Neurosci Lett*. 2018. 673:33-38.
- Lim YY, Quah EPL. Antioxidant properties of different cultivars of *Portulaca oleracea*. *Food Chem*. 2007. 103:734-740.
- Lu B, Nagappan G, Lu Y. BDNF and synaptic plasticity, cognitive function, and dysfunction. In: Lewin G, Carter B, editors. *Neurotrophic Factors. Handbook of Experimental Pharmacology*. Springer, Berlin, Germany. 2014. p 223-250.
- Ma L, Wang DD, Zhang TY, Yu H, Wang Y, Huang SH, et al. Region-specific involvement of BDNF secretion and synthesis in conditioned taste aversion memory formation. *J Neurosci*. 2011. 31:2079-2090.
- Madara JC, Levine ES. Presynaptic and postsynaptic NMDA receptors mediate distinct effects of brain-derived neurotrophic factor on synaptic transmission. *J Neurophysiol*. 2008. 100:3175-3184.
- McGleenon BM, Dynan KB, Passmore AP. Acetylcholinesterase inhibitors in Alzheimer's disease. *Br J Clin Pharmacol*. 1999. 48:471-480.
- Michael M, Phebus RK, Schmidt KA. Plant extract enhances the viability of *Lactobacillus delbrueckii* subsp. *bulgaricus* and *Lactobacillus acidophilus* in probiotic nonfat yogurt. *Food Sci Nutr*. 2015. 3:48-55.
- Mizuno M, Yamada K, Olariu A, Nawa H, Nabeshima T. Involvement of brain-derived neurotrophic factor in spatial memory formation and maintenance in a radial arm maze test in rats. *J Neurosci*. 2000. 20:7116-7121.
- Moghbelinejad S, Nassiri-Asl M, Farivar TN, Abbasi E, Sheikhi M, Taghiloo M, et al. Rutin activates the MAPK pathway and BDNF gene expression on beta-amyloid induced neurotoxicity in rats. *Toxicol Lett*. 2014. 224:108-113.
- Monje ML, Toda H, Palmer TD. Inflammatory blockade restores adult hippocampal neurogenesis. *Science*. 2003. 302:1760-1765.
- Moon M, Kim HG, Choi JG, Oh H, Lee PK, Ha SK, et al. 6-Shogaol, an active constituent of ginger, attenuates neuroinflammation and cognitive deficits in animal models of dementia. *Biochem Biophys Res Commun*. 2014. 449:8-13.
- Noorbakhshnia M, Karimi-Zandi L. *Portulaca oleracea* L. prevents lipopolysaccharide-induced passive avoidance learning and memory and TNF- α impairments in hippocampus of rat. *Physiol Behav*. 2017. 169:69-73.
- Paniz C, Bairros A, Valentini J, Charão M, Bulcão R, Moro A, et al. The influence of the serum vitamin C levels on oxidative stress biomarkers in elderly women. *Clin Biochem*. 2007. 40:1367-1372.
- Park SJ, Kim DH, Jung JM, Kim JM, Cai M, Liu X, et al. The ameliorating effects of stigmasterol on scopolamine-induced memory impairments in mice. *Eur J Pharmacol*. 2012. 676:64-70.
- Qin L, He J, Hanes RN, Pluzarev O, Hong JS, Crews FT. Increased systemic and brain cytokine production and neuroinflammation by endotoxin following ethanol treatment. *J Neuroinflammation*. 2008. 5:10. <https://doi.org/10.1186/1742-2094-5-10>
- Russell W, Duthie G. Plant secondary metabolites and gut health: the case for phenolic acids. *Proc Nutr Soc*. 2011. 70:389-396.
- Sarter M, Bruno JP. Developmental origins of the age-related decline in cortical cholinergic function and associated cognitive abilities. *Neurobiol Aging*. 2004. 25:1127-1139.
- Semmler A, Frisch C, Debeir T, Ramanathan M, Okulla T, Klockgether T, et al. Long-term cognitive impairment, neuronal loss and reduced cortical cholinergic innervation after recovery from sepsis in a rodent model. *Exp Neurol*. 2007. 204:733-740.
- Su D, Zhao Y, Wang B, Xu H, Li W, Chen J, et al. Isoflurane-induced spatial memory impairment in mice is prevented by the acetylcholinesterase inhibitor donepezil. *PLoS One*. 2011. 6:e27632. <https://doi.org/10.1371/journal.pone.0027632>
- Sumathi T, Christinal J. Neuroprotective effect of *Portulaca oleracea*

- ethanolic extract ameliorates methylmercury induced cognitive dysfunction and oxidative stress in cerebellum and cortex of rat brain. *Biol Trace Elem Res*. 2016. 172:155-165.
- Tariot PN, Patel SV, Cox C, Henderson RE. Age-related decline in central cholinergic function demonstrated with scopolamine. *Psychopharmacology*. 1996. 125:50-56.
- Terry AV Jr, Buccafusco JJ. The cholinergic hypothesis of age and Alzheimer's disease-related cognitive deficits: recent challenges and their implications for novel drug development. *J Pharmacol Exp Ther*. 2003. 306:821-827.
- Vestergaard M, Kerman K, Saito M, Nagatani N, Takamura Y, Tamiya E. A rapid label-free electrochemical detection and kinetic study of Alzheimer's amyloid beta aggregation. *J Am Chem Soc*. 2005. 127:11892-11893.
- Walsh DM, Lomakin A, Benedek GB, Condron MM, Teplow DB. Amyloid β -protein fibrillogenesis. Detection of a protofibrillar intermediate. *J Biol Chem*. 1997. 272:22364-22372.
- Wang P, Sun H, Liu D, Jiao Z, Yue S, He X, et al. Protective effect of a phenolic extract containing indoline amides from *Portulaca oleracea* against cognitive impairment in senescent mice induced by large dose of D-galactose/NaNO₂. *J Ethnopharmacol*. 2017. 203:252-259.
- Yun SW, Kim JK, Lee KE, Oh YJ, Choi HJ, Han MJ, et al. A probiotic *Lactobacillus gasseri* alleviates *Escherichia coli*-induced cognitive impairment and depression in mice by regulating IL-1 β expression and gut microbiota. *Nutrients*. 2020. 12:3441. <https://doi.org/10.3390/nu12113441>
- Zhao XH, He X, Yang XF, Zhong XH. Effect of *Portulaca oleracea* extracts on growth performance and microbial populations in ceca of broilers. *Poult Sci*. 2013. 92:1343-1347.