



Complete Genome Sequence of the Red-Pigmented Strain Serratia marcescens SCQ1 and Its Four Spontaneous Pigment Mutants

Microbiology[®]

Resource Announcements

Tingting Xiang, a Rui Liu, a Jing Xu, Cailing Xu, Nuo Wang, Wei Zhou, Yu Zhao, Minhui Luo, Congji Wana

a Laboratory of Invertebrate Pathology and Applied Microbiology, College of Sericulture, Textile and Biomass Sciences, Southwest University, Chongqing, China

ABSTRACT Serratia marcescens SCQ1 is a red-pigmented bacterium isolated from silkworm larva with septicemia. Pigment-deficient spontaneous mutants arise when *S. marcescens* SCQ1 is incubated under relatively stable laboratory conditions for a long time. Here, we present the complete genome sequence of SCQ1 and the resequenced genomes of four spontaneous pigment mutants.

erratia marcescens, a kind of Gram-negative bacterium, belongs to the family Enterobacteriaceae, which is widely distributed in diverse environments, including water, soil, plants, and animals (1). Some strains of S. marcescens may produce a red pigment, prodigiosin, which is a secondary metabolite with multiple bioactivities that has a linear tripyrrole skeleton (2-methyl-3-pentyl-6-methoxyprodiginine) (2-4). S. marcescens SCQ1 was isolated from the hemolymph of diseased silkworm larvae in a sericultural area of Chongqing, China. The hemolymph of silkworm larvae was streaked onto an LB plate and incubated at 28°C for 24 h. Red colonies were picked and restreaked onto LB medium 3 times to obtain the axenic strain S. marcescens SCQ1, which was stored at -20° C. Strain SCQ1 was then passaged 20 times in liquid LB medium (pH 7), incubated at 28°C with shaking (200 rpm). Every day, 500 μ l of culture was transferred into 50 ml of fresh LB. After 20 transfers, 50 μ l of the diluted cultures was plated onto LB medium. Colonies with different pigment phenotypes were picked, and the mutant strains SCQ1-1M (pink), SCQ1-2M (white), SCQ1-3M (white), and SCQ1-4M (white sides and red center) were characterized by different colony colors (5). The genetic changes underlying the pigment deficiencies in the S. marcescens mutants have not yet been identified.

S. marcescens SCQ1 and the other four spontaneous mutants were cultured in LB broth at 28°C. Genomic DNA was extracted using the E.Z.N.A. bacterial DNA kit (Omega Bio-tek, Norcross, GA). DNA concentration and quality were assessed using a NanoDrop 2000 UV-visible (UV-vis) spectrophotometer (Implen, CA, USA) and 1% agarose gel electrophoresis. Genomic DNA of SCQ1 was sheared into \sim 10-kb fragments using a q-TUBE device (Covaris). A library was constructed for SCQ1 using the SMRTbell Express template preparation kit v2.0 (Pacific Biosciences, USA) and sequenced using single-molecule real-time (SMRT) sequencing technology on the PacBio RS II sequencing platform (6). A total of 3,152,565,048 bases and 320,553 subreads were generated. The mean subread length and N_{50} value were 9,834 bp and 11,305 bp, respectively. The reads were assembled using Falcon v2.0.0. Genomic DNA from SCQ1-1M, SCQ1-2M, SCQ1-3M, and SCQ1-4M was sheared into 400- to 500-bp fragments using the M220 sonicator (Covaris, Woburn, MA, USA). Paired-end sequencing libraries were prepared for SCQ1-1M, SCQ1-2M, SCQ1-3M, and SCQ1-4M using the NEBNext Ultra DNA library prep kit (NEB, USA). Whole-genome sequencing of SCQ1-1M, SCQ1-2M, and SCQ1-3M was performed using the HiSeq X Ten platform (Illumina, San Diego, CA, USA), and whole-genome sequencing of SCQ1-4M was performed using the Illumina HiSeq 4000 platform to generate 2×150 -bp paired-end reads. Quality control on the Illumina data was assessed with FastQC v0.11.5 (7). The adapter sequences, low-quality reads, and reads

Citation Xiang T, Liu R, Xu J, Xu C, Wang N, Zhou W, Zhao Y, Luo M, Wan Y. 2021. Complete genome sequence of the redpigmented strain *Serratia marcescens* SCQ1 and its four spontaneous pigment mutants. Microbiol Resour Announc 10:e01456-20. https://doi.org/10.1128/MRA.01456-20.

Editor Julia A. Maresca, University of Delaware

Copyright © 2021 Xiang et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Yongji Wan, canbl3312@126.com.

Received 18 December 2020 Accepted 13 March 2021 Published 15 April 2021

AMERICAN SOCIETY FOR

MICROBIOLOGY

containing more than 10% of N bases were removed. Default parameters were used except where otherwise noted.

The assembled SCQ1 genome sequence was 5,063,944 bp with an average coverage of \sim 622 \times and a GC content of 61.33%, and it consists of a 4,994,354-bp circular chromosome and a 69,590-bp plasmid. The genome was annotated using the National Center for Biotechnology Information (NCBI) Prokaryotic Genome Annotation Pipeline (PGAP) v4.13 (8). It contains 4,692 protein-encoding genes, 22 rRNA genes, 91 tRNA genes, 14 noncoding RNA (ncRNA) genes, 25 pseudogenes, and 1 CRISPR array. We predicted 7 genomic islands using IslandPath-DIOMB v0.2 software (9) and 4 prophage sequences using PhiSpy v2.3 software (10). For whole-genome sequencing of SCQ1-1M, SCQ1-2M, SCQ1-3M, and SCQ1-4M, we obtained 20,976,838, 23,788,882, 18,999,970, and 8,876,328 clean reads, respectively. Using the Burrows-Wheeler Aligner (BWA) v0.7.8 (11), the reads were aligned to the SCQ1 assembled genome sequence. We used SAMtools v0.1.18 (12) for converting the SAM and BAM formats and sorting, indexing, and calling variants. Identified variants were annotated using the ANNOVAR v2019-09-01 tool (13). The resequencing depth was at least 184 \times with coverage of more than 99.99%. In total, 26 single nucleotide polymorphisms (SNPs) were detected, including 8 in SCQ1-1M, 8 in SCQ1-2M, 8 in SCQ1-3M, and 2 in SCQ1-4M. All of them were substitutions of base, and no indels were detected in the genomes of the mutants. No SNPs were found in the prodigiosin biosynthetic gene cluster in strains SCQ1-2M, SCQ1-3M, and SCQ1-4M. In strain SCQ1-1M (pink), one SNP was found in the pigJ gene. This SNP is a nonsynonymous substitution (A to G), which caused the change of an amino acid from Asp to Gly and may impact the activity of PigJ (betaketoacyl synthase).

Data availability. The complete genome sequence of *S. marcescens* SCQ1 has been deposited in GenBank under accession numbers CP063354 (chromosome) and CP063355 (plasmid). The raw sequencing data for SCQ1, SCQ1-1M, SCQ1-2M, SCQ1-3M, and SCQ1-4M have been deposited in the NCBI SRA database under BioProject accession number PRJNA386063 and SRA accession numbers SRR12825186, SRR12737688, SRR12825185, SRR12825184, and SRR12825183, respectively.

ACKNOWLEDGMENTS

This work was supported by the China Qujing Academician and Experts Workstation fund (grant number WYJ20170413) and the China Agriculture Research System (number CARS-22).

REFERENCES

- Petersen LM, Tisa LS. 2013. Friend or foe? A review of the mechanisms that drive *Serratia* towards diverse lifestyles. Can J Microbiol 59:627–640. https://doi.org/10.1139/cjm-2013-0343.
- Harris AKP, Williamson NR, Slater H, Cox A, Abbasi S, Foulds I, Simonsen HT, Leeper FJ, Salmond GPC. 2004. The *Serratia* gene cluster encoding biosynthesis of the red antibiotic, prodigiosin, shows species- and strain-dependent genome context variation. Microbiology (Reading) 150:3547–3560. https:// doi.org/10.1099/mic.0.27222-0.
- Williamson NR, Fineran PC, Leeper FJ, Salmond GPC. 2006. The biosynthesis and regulation of bacterial prodiginines. Nat Rev Microbiol 4:887–899. https://doi.org/10.1038/nrmicro1531.
- Stankovic N, Senerovic L, Ilic-Tomic T, Vasiljevic B, Nikodinovic-Runic J. 2014. Properties and applications of undecylprodigiosin and other bacterial prodigiosins. Appl Microbiol Biotechnol 98:3841–3858. https://doi.org/10.1007/ s00253-014-5590-1.
- Zhou W, Li J, Chen J, Liu X, Xiang T, Zhang L, Wan Y. 2016. The red pigment prodigiosin is not an essential virulence factor in entomopathogenic Serratia marcescens. J Invertebr Pathol 136:92–94. https://doi.org/ 10.1016/j.jip.2016.03.011.
- Korlach J, Bjornson KP, Chaudhuri BP, Cicero RL, Flusberg BA, Gray JJ, Holden D, Saxena R, Wegener J, Turner SW. 2010. Real-time DNA sequencing from single polymerase molecules. Methods Enzymol 472:431–455. https://doi.org/10.1016/S0076-6879(10)72001-2.

- 7. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. Babraham Bioinformatics, Cambridge, United Kingdom.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/ 10.1093/nar/gkw569.
- Hsiao W, Wan I, Jones SJ, Brinkman FSL. 2003. IslandPath: aiding detection of genomic islands in prokaryotes. Bioinformatics 19:418–420. https://doi .org/10.1093/bioinformatics/btg004.
- Akhter S, Aziz RK, Edwards RA. 2012. PhiSpy: a novel algorithm for finding prophages in bacterial genomes that combines similarity- and composition-based strategies. Nucleic Acids Res 40:e126. https://doi.org/10.1093/ nar/gks406.
- 11. Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. Bioinformatics 25:1754–1760. https://doi.org/10.1093/bioinformatics/btp324.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. Bioinformatics 25:2078–2079. https://doi.org/10.1093/bioinformatics/btp352.
- Wang K, Li M, Hakonarson H. 2010. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. Nucleic Acids Res 38:e164. https://doi.org/10.1093/nar/gkq603.