



Platelet-Rich Plasma as a **Potential New Strategy in the Endometrium Treatment in** Assisted Reproductive Technology

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Lin Y. Qi J and Sun Y (2021) Platelet-Rich Plasma as a Potential New Strategy in the Endometrium Treatment in Assisted Reproductive Technology. Front. Endocrinol. 12:707584. doi: 10.3389/fendo.2021.707584 The success rate of assisted reproduction techniques (ART) has long been less than satisfactory albeit the great progress made in recent years, demonstrating the need for alternative options in the ART cycles. Growing evidence correlates the effect of intrauterine platelet-rich plasma (PRP) infusion on the endometrium with reassuring reproductive results. Thus, in this review, we focus on the current clinical and mechanical evidence on PRP and its effect on endometrial receptivity, and assess the features, benefits and limitations of the current studies and potential risks of PRP in ART.

Keywords: platelet-rich plasma, endometrial receptivity, mechanisms, assisted reproduction techniques, risk

INTRODUCTION

Successful implantation requires a receptive endometrium, a functional embryo and a synchronized interaction between blastocyst and endometrium (1). Though advances in assisted reproduction techniques (ART) have led to remarkable improvements in pregnancy rates, implantation failure has long been an unsolved problem. One-third of implantation failures can be attributed to embryo reasons, while unsatisfactory endometrial receptivity and poor embryo-endometrium communication account for the remaining two-thirds of such failures (1). Despite the great progress in embryo election and quality improvement, many patients are still undergoing repeated implantation failure (RIF). Poor endometrial receptivity has become a bottleneck issue in the ART field, and the lack of evidence-based treatments signifies the issue. Clinicians and scientists have struggled to find an effective therapeutic solution.

Recently, increasing evidence has shown the positive role played by autologous platelet-rich plasma (PRP) in treating endometrium (2, 3). PRP is a volume of plasma, obtained by centrifugation of the patient's whole blood, that has a platelet count above baseline (2, 4). With the intrauterine infusion of PRP, numerous proteins, several growth factors (GFs), and cytokines stored in the platelet act on the endometrium through the promotion of cell proliferation and neoangiogenesis, and the anti-inflammatory properties, resulting in successful implantation (2).

PRP treatment is gaining researchers' attention due to its unique advantages. As an autologous biologic material, PRP minimizes the risk of immune reactions and contagious diseases (5). In addition, the application of PRP is considered less invasive since it is produced from peripheral blood (6).

In this article, we review PRP and its application to clinical medicine, with an emphasis on reproduction. We then address the current clinical and mechanical evidence for the influence of PRP on endometrial receptivity. Last but not least, we analyze the limitations of the current studies and potential risks of PRP in co-treatment in ART.

PRP AND ITS CLINICAL APPLICATION

PRP can be defined as a volume of plasma that has a platelet count above baseline prepared by centrifugation of the patient's peripheral blood (2, 4). Platelets are non-nucleated cell fragments evolved from bone marrow-derived megakaryocytes. The cytoplasm of platelets is comprised of two parts: the granules accumulated chromomere and the agranular hyalomere rich in cytoskeletal proteins. Platelet granules contain multiple proteins including antimicrobial peptides (7), fibronectin and vitronectin (8), GFs [including platelet-derived growth factor (PDGF), the epidermal growth factor (EGF), the transforming growth factor β -I (TGF β -I), the vascular endothelial growth factor (VEGF), the hepatocyte growth factor (HGF), the basic fibroblast growth factor (bFGF) (9)], and cytokines [comprising anti-inflammatory and pro-inflammatory interleukins, interleukin-4 (IL-4), IL-8, IL-13, IL-17, tumor necrosis factor- α (TNF- α), and interferon (IFN)- α (4)]. These proteins are secreted in the event of injuries so that platelets are activated and then delivered to the site of injury or deficiency (4, 8), allowing PRP to participate in cell proliferation, migration, tissue growth and healing, neoangiogenesis (2), inflammatory processes (7, 10), chemotaxis and immune responses (3).

Platelet concentrates can be sorted into four categories based on their leucocyte and fibrin content: pure platelet-rich plasma (P-PRP), leucocyte- and platelet-rich plasma (L-PRP), pure platelet-rich fibrin (P-PRF), and leucocyte and platelet-rich fibrin (L-PRF) (11). Each one has a particular bioaction and has been applied in a specific clinical field. However, the overall goal is to achieve a concentrating platelet, as well as factors of two to three times in whole blood, in order to make PRP a more effective instrument than peripheral blood in the processes described above (2). Factors in platelet granules continue exocytosing after their first-round release at activation, maintaining their levels three- to five-fold higher as compared to baseline values (12).

Applications of PRP are commonly disciplined in various conditions related to regenerative medicine without known effective treatments, such as osteoarthritis (13) and ligament injuries (14) in orthopedics, skin rejuvenation and hair loss in dermatology (15) and breast augmentation and wound healing in aesthetic surgery (16). Latest reports also record the application of PRP in reproduction: PRP has been found to facilitate growth rate and cell viability of primordial or primary to pre-antral stage follicles, and improve ovarian function, pregnancy and birth rate (17–20). In this article, we will review and summarize the preparation procedure of PRP, the function of PRP on endometrium and its contribution to the improvement of reproductive results.

PRP PREPARATION

Most studies share similar PRP preparation procedures drawing patients' own venous blood aiming at obtaining a three-layer separation from the whole blood first, which contains the cellular plasma in supernatant, the intermediate buffy coat containing concentrated platelets, and the bottom red blood cells, and then mixing the pellet of platelets with 0.5-1ml of supernatant to get the final PRP sample. However, multiple inconsistent details in various studies may potentially lead to contradictory outcomes. For example, in the relevant articles, the amount of venous blood drawn from the patient varied from 8ml to 18ml, and the relative centrifugal force differed from 300*g for 10min to 3300*g for 7min. Therefore, different concentrations of platelets and leukocytes could be obtained, which may in turn impact the amount of various types of growth factors playing a key part in the PRP samples. Still, researchers haven't reached a consensus on whether to use the activated or inactivated platelets, as well as the type of the platelet agonists for activation, including calcium chloride, thrombin, etc. (21, 22). Only a minority is centrifuged again in order to remove cellular debris through a filter (21). Besides, few paid attention to the temperature when processing the PRP. Since the standard PRP preparation protocol is the prerequisite of the evaluation of different results in various trials, it is crucial to make every attempt to develop the optimal protocol for PRP preparation.

CLINICAL EVIDENCE OF PRP ON ENDOMETRIAL RECEPTIVITY

Research in the effect of PRP on the endometrium has surged in recent five years. Most studies apply PRP in clinical cases of patients with thin endometrium, repeated implantation failure (RIF), chronic endometritis (CE) and Asherman Syndrome (AS), since unfortunately, there has not yet been any effective remedy for these distressing problems. It has been demonstrated that intrauterine PRP infusion positively influences the reproductive results, including endometrial thickness, clinical pregnancy, live birth, etc., thus can be potentially included in the different protocols for endometrial preparation. Tables 1-3 outline the main characteristics of almost all related and eligible studies, among which five studies are randomized clinical trials (RCTs), and one being a single arm RCT. The studies cover different populations, including patients with thin endometrium (<7mm) in 8 studies, patients with RIF in 7 studies, and patients with refractory endometrium and suboptimal endometrial growth. No consensus has been reached on the optimal timing of PRP injection. For the patients with thin endometrium, the timing of PRP infusion varied from the 8th day to the 17th day of HRT cycle, and for those who underwent RIF, PRP therapy was usually applied 2 days before the embryo transfer (ET). Seventeen out of eighteen studies administered the PRP with the dose of 0.5-1.0 ml, the remaining one with a dose <0.5ml. We will now analyze the related studies of intrauterine PRP infusion published in various clinical ART settings.

TABLE 1 | Studies assessing the effect of platelet-rich plasma (PRP) treatment in patients with thin endometrium in vitro fertilization.

Year of Publication	Country	Year of	RCT	Objects	Sam	ple Size	Intervention	Control	Time of PRP Infusion	Transfer	Outcome	Measures		Effects		Referer
		Study			Case	Control				Туре			Case (After PRP treatment)	Control (Before PRP treatment)	P Value*	
2015	China	March 2014 to June 2014	No	Women aged 31-39 with thin endometrium (<7 mm) after standard HRT	5	/	HRT+Intrauterine infusion of 0.5-1 ml of PRP	/	On the 10th day of HRT cycle (another dose of infusion of PRP was performed 72h later if the	FET	Endometrial thickness	Reached 7mm with one or two doses	5/5	0/5	/	(23)
									endometrial thickness was		Clinical pregr	lancy	5/5	0/5	/	
									not satisfied)		Live birth		4/5	0/5	/	
2017	Iran	September 2015 to May 2016	No	Women with a history of inadequate endometrial growth (<7	10	/	HRT+Intrauterine infusion of 0.5ml PRP	/	On the 11-12th day and repeated on 13-14th day of HRT cycle	FET	Endometrial thickness	Reached 7mm with one dose	0/10	0/10	/	(24)
				mm) in FET								Reached 7mm with two infusions	10/10	0/10	/	
											Chemical pre	gnancy	5/10	0/10	/	
											Clinical pregr	-	4/10	0/10	/	
2018	Iran	September	Yes	Women aged 18-42	40	43	HRT+Intrauterine		On the 13th day of HRT	FET		hickness (mm)	8.67 ± 0.64	8.04 ± 0.27	0.001	↑ (25)
		2016 to		with thin endometrium			infusion of 0.5-	without	cycle		Clinical pregr	ancy (per-	13/40	6/43 (14.0%)	0.044	1
		January		(<7 mm)			1.0ml of PRP	intrauterine			cycle)		(32.5%)			
		2017						administration			Implantation		21%	9.37%	0.002	1
											Ongoing pre	gnancy (per-	11/40	6/43 (14.0%)	0.127	()
											cycle)		(27.0%)			
2019	Iran	2016 to	Yes	Women who had a	30	30	HRT+Intrauterine	Sham	On day 11-12 and was	FET	Endometrial	One dose	$5.993 \pm$	5.453 ±	0.63	(-) (26)
		2017		history of canceled			infusion of 0.5ml of	catheter	repeated after 48 h		thickness		0.701	0.823		
				frozen-thawed embryo			PRP					Two doses	7.213 ±	5.767 ±	< 0.001	1
				transfer cycle due to a									0.188	0.973		
				thin endometrium (<7							Chemical pre		12/30(40%)	2/30(6.7%)	0.031	1
2019	01.	11.0015		mm)	0.4	00				CCT	Clinical pregr	-	10/30(33.3%)	1/30 (3.3%)	0.048	1
2019	China	July 2015	No	Women aged under 40 with thin endometrium	34	30	HRT+Intrauterine infusion of 0.5-	Underwent ET without		FET		hickness (mm)	7.65 ± 0.22 15/34	6.52 ± 0.31	0.013	↑ (22) ↑
		to July 2016		(<7 mm)			1.0ml of PRP	intrauterine	cycle		Clinical pregr	lancy	(44.12%)	6/30(20%)	0.036	I
		2010		(<7 11111)				administration			Implantation	rate	19/34	7/30	0.018	†
								administration			πραπαιοπ	ato	(27.94%)	(11.67%)	0.010	I
2019	South	December	No	Women age of 20-45	20	/	HRT+Intrauterine	/	On day 10 and was	FET	Endometrial t	hickness (mm)	6.0 ± 1.1	(11.0770) 5.4 ± 0.8	0.070	(-) (27)
1010	Korea	2015 to		years who had a	20	,	infusion of 0.7-	,	repeated at 3 day intervals		Clinical pregr		6/20	0/20	0.020	() (=·) ↑
		June 2017		history of two or more			1.0ml of PRP		until the EMT reached 7		Live birth		4/20	0/20	0.106	(-)
				failed IVF cycles and					mm, 3 days before ET		Implantation	rate	7/55	0/52	0.015	1
				refractory thin					,,.							
				endometrium (<7mm)												
2020	India	July 2018	No	Women aged 27 to 39	32	/	HRT+ Instillation of	/	7–10 days after the	FET	Endometrial	≥7 mm	24/32(75%)	0/32	/	(28)
		to		years, suffering from			4ml PRP(1ml in		injection of leuprolide (on		thickness	(embryo	. /			
		September		primary or secondary			each wall of the		day 16 of OCP), 22-			transfer				
		2019		infertility with thin			uterine cavity) into		27days before ET			done)				
				endometrium (<7 mm)			the					6–7 mm	4/32(12.5%)	0/32	/	
												(embryo				

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Year of Co Publication	Country Year of Shudv	T OT RCI	Objects	sample size	Intervention	Control		Iranster Outcome Measures Tyne	ő	Effects		Keterence
		6		Case Control			-		Case	Control	P *orlow	
									Anter PRP treatment	(Atter (Before PRP PRP treatment) treatment)		
					endomyometrial			transfer not	not			
					junction			done)				
								<6 mm no	10 4/32(12.5%)	0/32	/	
								improvement	nent			
								(embryo				
								transfer not	Tot			
								done)				
								Serum _b -hCG positive	12/24(50%)	0/24	/	
								Clinical pregnancy	10/24	0/24	/	
									(41.66%)			
								Live birth	5/24(20.83%)	0/24	/	
2020 Bra	Brazil Januar	January to No	Patients with	21 /	HRT+Intrauterine /		PRP was administered on FET	- Clinical pregnancy	0.667	0/21	/	(29)
	December	hber	endometrium thickness		infusion of 0.5ml of		the 14th to 17th day of	Ongoing pregnancy or live	ive 0.54	0/21	/	
	2018		below 5 mm during		РПР		HRT every second day,	birth				
			preparation for FET				for a total of three					
							infusions.					

Thin Endometrium

Endometrium is one of the most prominent factors in implantation and pregnancy. Multiple studies suggest that pregnancy rate may increase with growing endometrial thickness within a certain range (40-43). It has been suggested that the minimal endometrial thickness required for embryo transfer is 7 mm at the end of follicular phase (44, 45). Thus, endometrial thickness less than 7 mm is considered to be thin, which occurs in 2.4% of the *in vitro* fertilization cycles and is associated with a lower possibility of pregnancy (44, 46).

Six studies on PRP intrauterine infusion treatment of infertile women with thin endometrium were conducted in frozen embryo transfer (FET) cycles on the 10th to 13th day of hormone replacement therapy (HRT) cycle, reaching a consensus on the significant improvement in the pregnancy rate and the endometrial thickness with the PRP treatment (Table 1). Three articles displayed 100% otherwise infertile cases reaching endometrial thickness of 7mm with one or two times of PRP infusion (23, 24, 26), and another 2 revealed PRP infusion leading to significantly increased endometrial thickness compared with the controls (22, 25). The study in Brazil also showed reassuring clinical pregnancy, ongoing pregnancy or live birth rates after the PRP treatment, but did not mention the endometrial thickness improvement (29). A study in South Korea found no statistically significant difference in the endometrium thickness and the live birth rates between the PRP and control groups but there was a significant statistical difference in implantation and clinical pregnancy rates between the PRP group and the controls (12.7 and 30%, respectively) (27). The largest RCT study by Eftekhar et al. (25) was conducted on 83 infertile women, including 40 patients with 0.5-1.0ml intrauterine PRP infusion on the 13th day of HRT cycle and 43 controls who underwent embryo transfer without intrauterine administration. The endometrium of the patients with PRP treatment expanded significantly to 8.67 \pm 0.64 (from 6.09 \pm 0.47) while the endometrium thickness of the controls merely increased to 8.04 ± 0.27 (from 6.15 ± 0.37) (p=0.001). The percycle clinical pregnancy rate raised from 14% in controls to 32.5% in the PRP group (p=0.044). However, there is no statistically significant difference in ongoing pregnancy rate between the two groups.

A recent research on 32 women suffering from primary or secondary infertility with thin endometrium (<7 mm) concluded that the injection of PRP (1ml in each wall of the uterine cavity) guided by hysteroscopy into the endomyometrial junction. In this study, PRP injection improved endometrial thickness and pregnancy rates in cases of previously canceled embryo transfer due to a thin endometrium (28). All studies have led to the conclusion that infertile women with thin endometrium benefit from PRP treatment, whereas the experimental conditions should be noticed and controlled, including PRP preparation process, PRP infusion dose, inclusion criteria etc.

Repeated Implantation Failure

Infertile women with RIF represent another large potential group that could benefit from the PRP administration. The definition of RIF slightly differs based on various sets of standards. Most TABLE 2 | Studies assessing the effect of platelet-rich plasma (PRP) treatment in repeated implantation failure (RIF) patients in vitro fertilization.

Year of	Country	Year of	RCT	Objects	San	ple Size	Intervention	Control	Time of	Transfer	Outcome		Effects		Referen
Publication		Study			Case	Control			PRP Infusion	Туре	Measures	Cases	Control	P Value*	
2016	Iran	March to June	A single arm	Women aged under 40 who failed to conceive after 3 or more ET with high-quality	20	/	HRT+Intrauterine infusion of 0.5ml of PRP	/	48h before	FET	Clinical pregnancy	18/20 (90%)	0/20	/	(30)
		2016	preliminary study of an	embryos					ET		Ongoing pregnancy	16/20 (80%)	0/20	/	
			RCT								Molar pregnancy	1/20 (5%)	0/20	/	
2019	Turkey	January 2014 to January	No	Patients aged between 21 and 39 with a history of at least three consecutive failed IVF	34	36	HRT+Intrauterine infusion of 1ml of PRP	Underwent ET without intrauterine	48h before ET	FET	Endometrial thickness (mm)	10/(8– 14)	6.25(4.3– 6.9)	<0.001	↑ (31)
		2017						administration	2.		Clinical pregnancy	17/34 (50%)	12/36 (33.3%)	0.042	↑
											Live birth	14/34 (41.2%)	6/36 (16.7%)	0.045	†
2019	Iran	2016- 2017	No	Patients with history of more than 2 repeated failed embryo transfer cycles	67	56	HRT+Intrauterine infusion of 1ml of PRP	Underwent ET with systemic administration of	48h before ET	FET	Chemical pregnancy Clinical	29/67 (43.3%) 27/67	15/56 (26.8%) 12/56	0.057	(-) (32) ↑
2019	Brazil	February	No	Patients with ≥2embryo transfers, and at	33	33	HRT+Intrauterine infusion of 0.7ml	GCSF Patients in their	48h	ICSI	pregnancy Clinical	(40.3%) 12/33	(21.4%) 10/33	0.025	T (-) (33)
		2017 to October		least 5 good-morphological embryos were transferred			of PRP+subcutaneous G-CSF injection (300mg/0.5ml started	first IVF/ICSI cycle attempt	before ET		pregnancy Implantation	(36.4%) 14/77	(30.3%) 12/68	0.90	(-)
		2017					simultaneously to PRP and was administered subcutaneously every	without PRP or G-CSF			rate Miscarriage	(18.2%) 3/12	(17.6%) 1/10	0.43	(-)
							week)	treatment			rate Ongoing pregnancy	(25.0%) 9/33 (27.3%)	(9.0%) 9/33 (27.3%)	0.99	(-)
											Live birth	9/33 (27.3%)	9/33 (27.3%)	0.99	(-)
2020	Iran	2016- 2017	Yes	Patients aged below 40 years with history of 3 or more embryo transfer failures with high-	49	48	HRT+Intrauterine infusion of 0.5 ml of PRP	Underwent ET without	48h before	FET	Chemical pregnancy	26/49 (53.06)	13/48 (27.08)	0.009	↑ (34)
2020	Iran	2016-	No	quality embryos and candidates for FET Women aged under 35 with a failure to	42	43	HRT+Intrauterine infusion of 1ml of	intrauterine administration Underwent ET	ET 48h	FET	Clinical pregnancy Biochemical	22/49 (44.89%) 15/42	8/48 (16.66%) 16/43	0.003	↑ (-) (35)
		2018		achieve a clinical pregnancy after the transfer of at least four good-quality embryos in at			PRP	without intrauterine	before ET		pregnancy Clinical	(35.7%) 13/42	(37.2%) 16/43	0.54	(-)
				least three fresh or frozen cycles and normal endometrial thickness (≥7 mm)				administration			pregnancy Ongoing	(31.0%) 11/42	(37.2%) 11/43 (25.6%)	0.90	(-)
2020	Iran	2016- 2019	Yes	Women aged between 20–40years who failed to be pregnant after three or more	55	43	HRT+Intrauterine infusion of 0.5ml of PRP	Underwent ET without	48h before	FET	pregnancy Clinical pregnancy	(26.8%) 29/55 (52.7%)	(23.8%) 10/43 (23.3%)	0.001	↑ (36)
				embryo transfer of embryos with good quality				intrauterine administration	ET		Ongoing pregnancy	28/55 (50.9%)	7/43 (16.3%)	0.001	Ť
											Implantation rate	35/55 (63.6%)	15/43 (34.9%)	0.001	î

*: ↑, increase; (-), no significant change.

TABLE 3 | Studies assessing the effect of platelet-rich plasma (PRP) treatment in other patients in vitro fertilization.

Year of	Country	Year of	RCT	Objects	Sam	ple Size	Intervention	Control	Time of PRP	Transfer Type			Effects			Referenc
Publication		Study			Case	Control			Infusion	Туре		Cases/ After PRP treatment	Control/ Before PRP treatment	P Value*		
2017	India	January to August 2016	No	Women aged 20-40 with suboptimal endometrial growth (endometrium thickness < 7mm despite standard dose of estradiol valerate (up to 16mg/day), or suboptimal endometrial vascularity (<5 vascular signals reaching the central zone (zones 3 and 4 as per Applebaum grading) of the endometrium), and repeated cycle cancellations	64	/	HRT +Intrauterine infusion of 0.5-0.8ml of PRP	/	On the 15th or 16th day of HRT cycle, and another dose 72h later if necessary	FET	Endometrial thickness (mm) Vascularity Serumβ-hCG	7.22 sparse, moder vascularity pat sparse → moo pattern (52) persistant spa pattern (4) 39/64	tern (17) dest vascularity	< 0.00001 /	Ţ	(37)
											positive Clinical pregnancy	(60.93%) 29/64 (45.31%)	0/64	/		
2018	China	January to October 2017	No	Women aged 27-43 who underwent RIF with thin endometrium (<7 mm) or suboptimal endometrial vascularity with <5 vascular signals reaching the central zone (zones 3 and 4 as per Applebaum grading)	20	/	HRT +Intrauterine infusion of 0.5-0.8ml of PRP	1	Not mentioned	FET	Endometrial thickness Pregnancy rate	7.82 ± 1.04 12/20(60%)	5.55 ± 0.71 0/20	<0.0001	ſ	(21)
2018	Venezuela	February 2016 to February	No	Women aged 33-45 with a history of refractory endometrium (characterized by atrophy with endometrial interface measurements below 6 mm by ultrasound; and/or	19	/	HRT +Intrauterine infusion of	/	On the 10th day of HRT cycle, and	IVF-ET	Serumβ-hCG positive Live birth	14/19 (73.7%) 5/19(26.3%)	0/19 0/19	/		(38)
		2017		the Asherman syndrome) and and at least one failed IVF attempt			1ml of PRP		then 72 hours after the first		Ongoing pregnancy	5/19(26.3%)	0/19	/		
									administration.		Chemical pregnancy Anembryonic pregnancies	2/19(10.5%) 1/19(5.3%)	0/19 0/19	/		
											Endometrial thickness	<7mm (with one dose); <9mm(with two doses)	Not mentioned	/		
2018	India	January 2018 to November 2018	No	Women aged 25-40 who had at least one previous FET failure with endometrium 7 mm or more in thickness	42	56	HRT +Intrauterine infusion of 0.3-0.4ml of PRP	Underwent ET without intrauterine administration	On the 8th or 9th day of HRT cycle	FET	Clinical pregnancy	47.6%	42.8%	0.09	(-)	(39)

*: ↑, increase; (-), no significant change.

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studies defined RIF as the failure of achieving a pregnancy after at least three embryo transfer cycles in which one or two highgrade embryos are transferred. Since the etiology of RIF has not been clinically confirmed, it is rather difficult to implement targeted therapy, leading to RIF patients being among the most difficult patients to treat, with no proven standard treatments.

Seven related studies, including 3 RCTs and 4 cohorts, were conducted, of which 6 studies detected encouraging results (Table 2). Two RCTs and a cohort study all implied significant improvements of clinical pregnancy rate with the use of intrauterine PRP infusion compared with controls, as well as other studied endpoints, including endometrial thickness, chemical pregnancy, implantation rate, ongoing pregnancy, and live birth rate. The RCT carried out in Iran with a population of 98 women with RIF indicated that intrauterine infusion of PRP increased the clinical pregnancy (intrauterine PRP infusion group vs control group: 48.3% vs 23.3%, p=0.001), ongoing pregnancy (intrauterine PRP infusion group vs control group: 46.7% vs 11.7%, p=0.001) and implantation rate (intrauterine PRP infusion group vs control group: 58.3% vs 25%; p=0.001) in PRP group compared with control group (36). Nazari et al. demonstrated similar results in their RCT, where increased clinical (intrauterine PRP infusion group vs control group: 44.89% vs 16.66%, p= 0.003) and chemical pregnancy rate (intrauterine PRP infusion group vs control group: 53.06% vs 27.08%) were observed in PRP group compared with control group (34). Mehrafza et al. compared the effect of intrauterine infusion of PRP with systemic administration of granulocyte colony stimulating factor(G-CSF) in a sample size of 123, and they demonstrated that intrauterine infusion of PRP could significantly enhance clinical pregnancy rate in RIF patients in comparison with systemic administration of G-CSF (32). One study demonstrated the efficacy of the combined use of intrauterine PRP and subcutaneous G-CSF (33). When compared with control patients in their first in vitro fertilization (IVF)/intracytoplasmic sperm injection (ICSI) cycle attempt without PRP or G-CSF treatment, no statistically significant difference regarding implantation, clinical pregnancy and miscarriage rates was found between the two groups, suggesting the combined use of intrauterine PRP and subcutaneous G-CSF a potential new strategy for endometrial receptivity improvement (33). However, another study holds opposing opinions. Tehraninejad et al. nonrandomly recruited 85 patients with RIF and normal endometrial thickness (≥ 7 mm), out of which 42 received 1ml PRP, and 43 were included in the control group. It is worth noting that there was no significant difference in the pregnancy outcomes between the two groups, including biochemical, clinical and ongoing pregnancy (≥ 20 weeks of gestation) rate (35). Till now, most studies on PRP and RIF confirm the positive role of PRP, but the efficacy of PRP on RIF patients remains controversial. The study design and the inclusion criteria should be controlled in order to investigate the effect of PRP.

Chronic Endometritis

Chronic endometritis is a persistent inflammatory condition of the endometrial mucosa with plasmacyte infiltration in the endometrial stromal compartment caused by various bacterial pathogens (47, 48). In the endometrium of CE, the local expression of multiple genes, including chemokines, cytokines and apoptotic proteins, was dysregulated (49). Various reports indicate that the incidence of CE in infertile women who underwent IVF cycles is approximately 15%, and over 30% of the patients with RIF are diagnosed with CE, the presence of which is associated with overall lower success rates in ART (50-52). Since CE could modify the endometrial decidualization, it may affect the endometrial receptivity to accommodate pregnancy (53). Oral antimicrobial regimes are considered to be a traditional remedy for CE. Nevertheless, certain CE cases do not respond satisfactorily to these widely applied treatments such as doxycycline, ciprofloxacin and metronidazole (54), driving research to address the need for alternative options when managing these cases. Recent research suggested that PRP may be employed as an ideal CE treatment (55). The case report presents a woman diagnosed with CE with premature ovarian insufficiency and a history of six failed donated embryo transfers. Twin pregnancy and birth were finally achieved with 1.5ml intrauterine PRP infusion curing the CE, after the woman underwent another new failed ET with antibiotic treatment. Another case showed 5 infertile women with CE achieved endometrial histologically recovery and 100% positive serum β-HCG results with the application of intrauterine infusion of 0.5-0.8ml of PRP every 2 days for 3 cycles (56). Clearly more studies on larger patient population, as well as more RCTs, are needed for clinically meaningful conclusions.

Asherman Syndrome

Asherman Syndrome (AS) consists of the development of intrauterine adhesions as a consequence of trauma, radiation, or infection in the endometrium. AS remains one of the most challenging pathologies fertility specialists encounter in daily practice. Prevalence of this syndrome varies from 2.8 to 45.5% in infertile women (57, 58).

There were only several cases of AS treated with intrauterine PRP infusion. Relevantly, it was reported that a 31-year-old women was diagnosed as AS with a postpartum curettage history (59). The patient received 1 ml autologous PRP on the 12th day of HRT, and the procedure was repeated 3 days after, allowing endometrial tissue regeneration, with increased vascularity, endometrium thickness, and restoration of endometrial function that eventually led to a successful pregnancy. In a separate study, two women with AS experienced recurrent pregnancy loss after various treatments were reported (60). Both finally succeeded in pregnancy with PRP therapy, despite the persistent thin lining, indicating that PRP may not only help endometrial growth, but also enhance its functional properties. It can be interpreted that the removal of scar tissue, leading to the exposure of the normal endometrial cells to the GFs and cytokines in PRP, helps boost the existing cellular functions involved in tissue regeneration.

Others

There are also some studies which contain different inclusion criteria described in detail in **Table 3**. All 4 studies take the

patients with at least one instance of failed embryo transfer previously as the research object, among which 3 studies point out that the use of autologous PRP holds promise in all the studied results, including endometrium thickness and diverse pregnancy tests (21, 37, 38). Kaur et al. (39) collected data of 98 patients who had at least one previous FET failure with endometrium 7 mm or more in thickness and divided them into the intervention group with the treatment of intrauterine infusion of 0.3-0.4ml of PRP and the control group. The intervention failed to increase the clinical pregnancy rate in this study. A related meta-analysis pointed out that the administered PRP at a dose of 0.5–1 ml was more effective than those administered PRP at doses of ≤ 0.5 ml and ≥ 1 ml (3). Therefore, the dose of the PRP used may have a biased impact on the results.

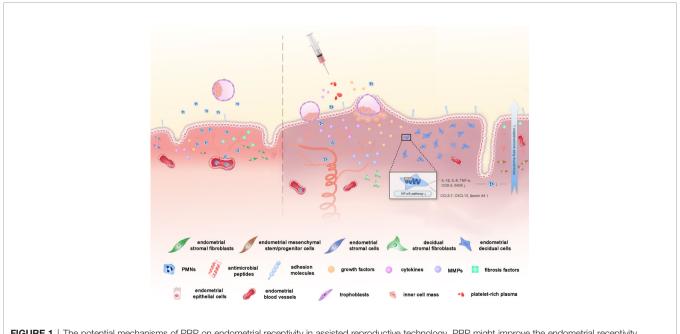
POTENTIAL MECHANISMS OF PRP ON ENDOMETRIAL RECEPTIVITY

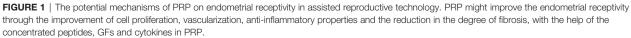
As growing evidence shows the correlation between satisfactory reproductive results and intrauterine PRP infusion treatment, researchers embark on the investigation of the underlying mechanisms of PRP in enhancing endometrial receptivity. Below are the potential mechanisms that account for the above clinical phenomena in the current literature (**Figure 1**).

First and foremost, PRP is known to play a critical role in the proliferation, regeneration and differentiation of cells due to its housing of a number of GFs, such as VEGF, PDGF, EGF, TGF, IGF1 and other cytokines, released upon platelet activation at the site of injury or inflammation (3, 61). Among the GFs, VEGF is responsible for vascularization as well. Evidence shows that PRP

stimulates various endometrial cell proliferation including epithelial cells, endometrial stromal fibroblasts, and endometrial mesenchymal stem/progenitor cells (21, 61, 62). Addition of PRP significantly increased cell proliferation in human endometrial stromal cell line (ICE7), and the replication of the effects was observed when primary stromal cells were used. Murine model also showed that a significantly increased Ki67 expression in endometrium after PRP treatment (63). Gaining expression of adhesion molecules and attracted stem cells and increased endometrial cell migration with PRP infusion may in turn stimulate endometrial growth and receptivity (8, 61). Research showed that a higher level of Hoxa10, one of the primary endometrial markers of receptivity, was detected in the endometrium after the application of PRP (63). Therefore, PRP rich in GFs can be the potential solution to the infertility patients with thin endometrium, which has been pathophysiologically characterized by the state of high blood flow resistance and VEGF down-regulation with inadequate epithelial growth and vascularization (64-68).

Considering that tissue growth and cell proliferation depend not only on single GFs, but on an overall profitable environment, another potential explanation for the action of PRP on endometrial is *via* its anti-inflammatory and anti-microbial property (2). Therapeutic PRP works by inhibiting nuclear factor kappa-B (NF- κ B) (69), a key valve of the inflammatory process, modulating prostaglandin-endoperoxide synthase 2 (COX-2) expression and other crucial inflammatory and proinflammatory cytokines, which have an important role in the implantation process (3). On one hand, PRP increases chemokines such as chemokine ligand 5 (CCL5), CCL7, CXCmotif chemokine ligand 13 (CXCL13) (61) and lipoxin A4 (69)





that reduces the attraction of polymorphonuclear neutrophils (PMNs) to the inflamed tissue, through blood vessel walls into the uterine lumen (69), leading to decreased intrauterine fluid accumulation, thus alleviating the inflammation (69, 70). On the other hand, PRP poses a down regulation effect of cytokines, such as IL-1 β , IL-8, TNF- α , COX-2 and inducible nitric oxide synthase (iNOS) expression *in vitro* and in animal models (62, 71). Additionally, the recorded presence of antimicrobial peptides in platelet secretory granules can also interpret the anti-inflammatory function of PRP (7). Since the endometrial environment should be switched to an anti-inflammatory state in the mid-secretory phase to prevent fetal rejection (72, 73), the mechanism of PRP treatment illustrated above may account for its reassuring clinical effects.

Last but not least, PRP treatment may have succeeded in reducing the degree of fibrosis *via* down-regulating expression of fibrosis-related factors including collagen type 1A (COL1A1), transforming growth factor β 1 (TGF β 1), tissue inhibitor of metalloproteinase-1 (TIMP1), and finally enhanced implantation sites and live birth rate ensued (6, 71). Matrix metalloproteinase (MMP)1, MMP3, MMP7, and MMP26, members of MMP gene family, involved in tissue regeneration and wound healing *via* degradation of extracellular matrix (ECM) and wound remodeling (74), are activated by a wide range of cytokines and GFs present in PRP and then are elevated (62). Hence, in terms of intrauterine adhesions, where factors stimulating the formation of fibrotic tissue into the intrauterine environment are released, PRP may be a potential ideal cure due to the mechanism above (6).

Apart from the cases above, studies show that PRP synergizes with other treatments to enhance the endometrial regeneration. In a rat model of intrauterine adhesion (75), PRP amplifies the therapeutic effect of menstrual blood-derived stromal cell (MenSC), optimizing the pregnancy outcomes through the improvement of the epithelial thickness, vascularization, the decrease of inflammation and fibrosis with augmented soluble paracrine factors released by MenSCs including insulin-like Growth Factor-1 (IGF-1), stromal cell-derived factor-1 (SDF-1) and thrombospondin-1 (TSP-1), and suppression in inflammation by increasing IL-1 β , IL-4, IL-10 and decreasing IL-6. The repairment derives from the stimulation of the Hippo signaling pathway with the regulation of the downstream factors connective tissue growth factor (CTGF), WNT5a and growth differentiation factor 5 (GDF-5). In another rat model of uterine horn damage (76), PRP therapy improves endometrial regeneration after BMSC transplant therapy, likely mediated through the NF-KB signaling pathway subunit p50 to directly induce the expression IL-10, and through the increase in endometrial thickness and angiogenesis.

LIMITATIONS OF THE CURRENT RESEARCH AND POTENTIAL RISKS OF PRP TREATMENT IN ART

Admittedly, both laboratory and clinical data make intrauterine PRP infusion a promising therapeutic approach in treating uterine endometrium. However, several limitations should be considered regarding the interpretation of the present studies and outcomes.

First, little is known about the best dosage and time or duration for the PRP treatment in endometrium. The importance of standard techniques and a highly repeatable protocol with sufficient description to prepare platelets for clinical use should be highlighted. Second, patients should be conferred upon separate treatment according to their putative cause of implantation failure in a more precise manner, so that the conclusion of the best beneficiary population of PRP treatment would be more convincing. Third, more rigid clinical trials, especially larger scaled RCTs, are warranted, even though they might be quite challenging to perform. Since intrauterine PRP infusion may result in local mechanical injuries which may affect endometrium receptivity, it is better to include sham catheter performance acting as a placebo, which is absent in most of the current studies, in the control group. However, due to its invasiveness and uncertain efficacy, it may well be that few patients are willing to make the attempt. Fourth, the indication of the PRP treatment should be considered more rigorous. As an autologous biologic material, PRP does appear to be considered relatively safe. However, some doubts remain on whether it is suitable for every infertile woman with thin endometrium, RIF and CE. Till now, there was only one study indicated that plasma from AS/endometrial atrophy patients would be comparable to plasma from healthy subjects (63). PRP primarily contains platelets, and some contains leucocytes as well. It is still not clear whether PRP extracted from the blood of patients with blood disorders like inflammation (CE, upper respiratory tract infection, etc.), leukemia and thrombocytopenia are different from that from healthy people, and for those patients, whether PRP therapy is still appliable and effective needs further discussion. Fifth, as PRP is a potential clinical application, more detailed mechanism is necessary. Last but not least, long-term health and complications of the resulting child, such as the high risk of endometrial cancer, should be considered. Due to PRP therapy being a novel application in reproductive medicine, it has been only 7 years since PRP infusion was first applied to endometrium. As reproductive results and endometrial thickness being the primary outcomes, follow-ups are basically terminated as the end of the pregnancy. Hence, as a treatment to be promoted for clinical use, a higher level of safety assessment with longer follow-up duration is necessary, for PRP introduces much greater concentrations of hematopoietic cells into the implantation environment that may have unknown detrimental effects on the embryo. In the circumstances, further studies are encouraged to assess the existing findings and clarify the medium- and long-term outcomes.

CONCLUSION

Clinical studies for the efficacy of PRP in improving uterine endometrial function remain controversial. Most studies suggested that PRP treatment resulted in positive reproductive results in patients with thin endometrium, RIF, CE and AS. However, the best dosage and timing of PRP application remains unknown. Few studies assess the underlying mechanisms of the PRP treatment. PRP might improve the endometrial receptivity through the improvement of cell proliferation, vascularization, anti-inflammatory properties and the reduction in the degree of fibrosis, with the help of the concentrated peptides, GFs and cytokines in PRP. But for its potential clinical application, further explication of the mechanism is necessary. Though increasing evidence suggests the possible value of PRP treatment on the endometrium of infertile women, more carefully designed studies, especially RCTs, on larger scales are required. Further, the standard PRP preparation procedure and the strict indication of the PRP treatment should be published. Studies and followups are needed on the long-term health and complications of the resulting child(ren).

AUTHOR CONTRIBUTIONS

YL drafted the manuscript. JQ and YS performed the critical revisions. All authors listed did contribute to the composing and

REFERENCES

- Craciunas L, Gallos I, Chu J, Bourne T, Quenby S, Brosens JJ, et al. Conventional and Modern Markers of Endometrial Receptivity: A Systematic Review and Meta-Analysis. *Hum Reprod Update* (2019) 25 (2):202–23. doi: 10.1093/humupd/dmy044
- Bos-Mikich A, de Oliveira R, Frantz N. Platelet-Rich Plasma Therapy and Reproductive Medicine. J Assist Reprod Genet (2018) 35(5):753-6. doi: 10.1007/s10815-018-1159-8
- Maleki-Hajiagha A, Razavi M, Rouholamin S, Rezaeinejad M, Maroufizadeh S, Sepidarkish M. Intrauterine Infusion of Autologous Platelet-Rich Plasma in Women Undergoing Assisted Reproduction: A Systematic Review and Meta-Analysis. J Reprod Immunol (2020) 137:103078. doi: 10.1016/j.jri.2019.103078
- Bos-Mikich A, Ferreira MO, de Oliveira R, Frantz N. Platelet-Rich Plasma or Blood-Derived Products to Improve Endometrial Receptivity? J Assist Reprod Genet (2019) 36(4):613–20. doi: 10.1007/s10815-018-1386-z
- Marques LF, Stessuk T, Camargo IC, Sabeh Junior N, dos Santos L, Ribeiro-Paes JT. Platelet-Rich Plasma (PRP): Methodological Aspects and Clinical Applications. *Platelets* (2015) 26(2):101–13. doi: 10.3109/09537104. 2014.881991
- Kim JH, Park M, Paek JY, Lee WS, Song H, Lyu SW. Intrauterine Infusion of Human Platelet-Rich Plasma Improves Endometrial Regeneration and Pregnancy Outcomes in a Murine Model of Asherman's Syndrome. *Front Physiol* (2020) 11:105. doi: 10.3389/fphys.2020.00105
- Drago L, Bortolin M, Vassena C, Taschieri S, Del Fabbro M. Antimicrobial Activity of Pure Platelet-Rich Plasma Against Microorganisms Isolated From Oral Cavity. *BMC Microbiol* (2013) 13:47. doi: 10.1186/1471-2180-13-47
- Urman B, Boza A, Balaban B. Platelet-Rich Plasma Another Add-on Treatment Getting Out of Hand? How can Clinicians Preserve the Best Interest of Their Patients? *Hum Reprod* (2019) 34(11):2099–103. doi: 10.1093/ humrep/dez190
- Pietrzak WS, Eppley BL. Platelet Rich Plasma: Biology and New Technology. J Craniofac Surg (2005) 16(6):1043–54. doi: 10.1097/01.scs.0000186454.07097.bf
- Bendinelli P, Matteucci E, Dogliotti G, Corsi MM, Banfi G, Maroni P, et al. Molecular Basis of Anti-Inflammatory Action of Platelet-Rich Plasma on Human Chondrocytes: Mechanisms of NF-κb Inhibition via HGF. J Cell Physiol (2010) 225(3):757–66. doi: 10.1002/jcp.22274
- Dohan Ehrenfest DM, Rasmusson L, Albrektsson T. Classification of Platelet Concentrates: From Pure Platelet-Rich Plasma (P-PRP) to Leucocyte- and Platelet-Rich Fibrin (L-PRF). *Trends Biotechnol* (2009) 27(3):158–67. doi: 10.1016/j.tibtech.2008.11.009

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- Zhu Y, Yuan M, Meng HY, Wang AY, Guo QY, Wang Y, et al. Basic Science and Clinical Application of Platelet-Rich Plasma for Cartilage Defects and Osteoarthritis: A Review. Osteoarthritis Cartilage (2013) 21(11):1627–37. doi: 10.1016/j.joca.2013.07.017
- Whitney KE, Liebowitz A, Bolia IK, Chahla J, Ravuri S, Evans TA, et al. Current Perspectives on Biological Approaches for Osteoarthritis. Ann N Y Acad Sci (2017) 1410(1):26–43. doi: 10.1111/nyas.13554
- Wasterlain AS, Braun HJ, Harris AH, Kim HJ, Dragoo JL. The Systemic Effects of Platelet-Rich Plasma Injection. Am J Sports Med (2013) 41(1):186– 93. doi: 10.1177/0363546512466383
- Zhang M, Park G, Zhou B, Luo D. Applications and Efficacy of Platelet-Rich Plasma in Dermatology: A Clinical Review. J Cosmet Dermatol (2018) 17 (5):660–5. doi: 10.1111/jocd.12673
- Samadi P, Sheykhhasan M, Khoshinani HM. The Use of Platelet-Rich Plasma in Aesthetic and Regenerative Medicine: A Comprehensive Review. *Aesthetic Plast Surg* (2019) 43(3):803–14. doi: 10.1007/s00266-018-1293-9
- Callejo J, Salvador C, González-Nuñez S, Almeida L, Rodriguez L, Marqués L, et al. Live Birth in a Woman Without Ovaries After Autograft of Frozen-Thawed Ovarian Tissue Combined With Growth Factors. J Ovarian Res (2013) 6(1):33. doi: 10.1186/1757-2215-6-33
- Sills ES, Wood SH. Autologous Activated Platelet-Rich Plasma Injection Into Adult Human Ovary Tissue: Molecular Mechanism, Analysis, and Discussion of Reproductive Response. *Biosci Rep* (2019) 39(6). doi: 10.1042/ BSR20190805
- Hosseini L, Shirazi A, Naderi MM, Shams-Esfandabadi N, Borjian Boroujeni S, Sarvari A, et al. Platelet-Rich Plasma Promotes the Development of Isolated Human Primordial and Primary Follicles to the Preantral Stage. *Reprod BioMed Online* (2017) 35(4):343–50. doi: 10.1016/j.rbmo.2017.04.007
- Sfakianoudis K, Simopoulou M, Nitsos N, Rapani A, Pantou A, Vaxevanoglou T, et al. A Case Series on Platelet-Rich Plasma Revolutionary Management of Poor Responder Patients. *Gynecol Obstet Invest* (2019) 84(1):99–106. doi: 10.1159/000491697
- 21. Wang X, Liu L, Mou S, Zhao H, Fang J, Xiang Y, et al. Investigation of Platelet-Rich Plasma in Increasing Proliferation and Migration of Endometrial Mesenchymal Stem Cells and Improving Pregnancy Outcome of Patients With Thin Endometrium. *J Cell Biochem* (2018) 120(5):7403–11. doi: 10.1002/ jcb.28014
- Chang Y, Li J, Wei LN, Pang J, Chen J, Liang X. Autologous Platelet-Rich Plasma Infusion Improves Clinical Pregnancy Rate in Frozen Embryo Transfer Cycles for Women With Thin Endometrium. *Medicine* (2019) 98 (3):e14062. doi: 10.1097/MD.000000000014062

- Chang Y, Li J, Chen Y, Wei L, Yang X. Autologous Platelet-Rich Plasma Promotes Endometrial Growth and Improves Pregnancy Outcome During *In Vitro* Fertilization. *Int J Clin Exp Med* (2015) 8(1):1286–90.
- Zadehmodarres S, Salehpour S, Saharkhiz N, Nazari L. Treatment of Thin Endometrium With Autologous Platelet-Rich Plasma: A Pilot Study. *JBRA* Assist Reprod (2017) 21(1):54–6. doi: 10.5935/1518-0557.20170013
- Eftekhar M, Neghab N, Naghshineh E, Khani P. Can Autologous Platelet Rich Plasma Expand Endometrial Thickness and Improve Pregnancy Rate During Frozen-Thawed Embryo Transfer Cycle? A Randomized Clinical Trial. *Taiwan J Obstet Gynecol* (2018) 57(6):810–3. doi: 10.1016/j.tjog.2018.10.007
- Nazari L, Salehpour S, Hoseini S, Zadehmodarres S, Azargashb E. Effects of Autologous Platelet-Rich Plasma on Endometrial Expansion in Patients Undergoing Frozen-Thawed Embryo Transfer: A Double-Blind RCT. Int J Reprod BioMed (2019) 17(6):443–8. doi: 10.18502/ijrm.v17i6.4816
- Kim H, Shin JE, Koo HS, Kwon H, Choi DH, Kim JH. Effect of Autologous Platelet-Rich Plasma Treatment on Refractory Thin Endometrium During the Frozen Embryo Transfer Cycle: A Pilot Study. *Front Endocrinol (Lausanne)* (2019) 10:61. doi: 10.3389/fendo.2019.00061
- Agarwal M, Mettler L, Jain S, Meshram S, Gunther V, Alkatout I. Management of a Thin Endometrium by Hysteroscopic Instillation of Platelet-Rich Plasma Into The Endomyometrial Junction: A Pilot Study. *J Clin Med* (2020) 9(9):2795. doi: 10.3390/jcm9092795
- Frantz N, Ferreira M, Kulmann MI, Frantz G, Bos-Mikich A, Oliveira R. Platelet-Rich Plasma as an Effective Alternative Approach for Improving Endometrial Receptivity - A Clinical Retrospective Study. *JBRA Assist Reprod* (2020) 24(4):442–6. doi: 10.5935/1518-0557.20200026
- Nazari L, Salehpour S, Hoseini S, Zadehmodarres S, Ajori L. Effects of Autologous Platelet-Rich Plasma on Implantation and Pregnancy in Repeated Implantation Failure: A Pilot Study. *Int J Reprod Biomed* (2016) 14(10):625–8. doi: 10.29252/ijrm.14.10.625
- Coksuer H, Akdemir Y, Ulas Barut M. Improved *In Vitro* Fertilization Success and Pregnancy Outcome With Autologous Platelet-Rich Plasma Treatment in Unexplained Infertility Patients That had Repeated Implantation Failure History. *Gynecol Endocrinol* (2019) 35(9):815–8. doi: 10.1080/09513590.2019.1597344
- 32. Mehrafza M, Kabodmehri R, Nikpouri Z, Pourseify G, Raoufi A, Eftekhari A, et al. Comparing the Impact of Autologous Platelet-Rich Plasma and Granulocyte Colony Stimulating Factor on Pregnancy Outcome in Patients With Repeated Implantation Failure. J Reprod Infertil (2019) 20(1):35–41.
- 33. Dieamant F, Vagnini LD, Petersen CG, Mauri AL, Renzi A, Petersen B, et al. New Therapeutic Protocol for Improvement of Endometrial Receptivity (PRIMER) for Patients With Recurrent Implantation Failure (RIF) - A Pilot Study. *JBRA Assist Reprod* (2019) 23(3):250–4. doi: 10.5935/1518-0557.20190035
- 34. Nazari L, Salehpour S, Hosseini MS, Hashemi Moghanjoughi P. The Effects of Autologous Platelet-Rich Plasma in Repeated Implantation Failure: A Randomized Controlled Trial. *Hum Fertil (Camb)* (2020) 23(3):209–13. doi: 10.1080/14647273.2019.1569268
- 35. Tehraninejad ES, Kashani NG, Hosseini A, Tarafdari A. Autologous Platelet-Rich Plasma Infusion Does Not Improve Pregnancy Outcomes in Frozen Embryo Transfer Cycles in Women With History of Repeated Implantation Failure Without Thin Endometrium. J Obstet Gynaecol Res (2021) 41(1):147– 51. doi: 10.1111/jog.14445
- 36. Zamaniyan M, Peyvandi S, Gorji HH, Moradi S, Jamal J, Yahya Poor Aghmashhadi F, et al. Effect of Platelet-Rich Plasma on Pregnancy Outcomes in Infertile Women With Recurrent Implantation Failure: A Randomized Controlled Trial. *Gynecol Endocrinol* (2020) 2:1-5. doi: 10.1080/09513590.2020.1756247
- Tandulwadkar SR, Naralkar MV, Surana AD, Selvakarthick M, Kharat AH. Autologous Intrauterine Platelet-Rich Plasma Instillation for Suboptimal Endometrium in Frozen Embryo Transfer Cycles: A Pilot Study. J Hum Reprod Sci (2017) 10(3):208–12. doi: 10.4103/jhrs.JHRS_28_17
- Molina A, Sanchez J, Sanchez W, Vielma V. Platelet-Rich Plasma as an Adjuvant in the Endometrial Preparation of Patients With Refractory Endometrium. JBRA Assist Reprod (2018) 22(1):42–8. doi: 10.5935/1518-0557.20180009
- Kaur J, Madhavan A, Naidu P, Rani K, Mahajan N. Intrauterine Autologous Platelet-Rich Plasma Therapy to Improve Implantation Rates in Patients Undergoing Frozen Embryo Transfer: A Pilot Study. Onco Fertil J (2018) 1 (2):81–5. doi: 10.4103/tofj.tofj_11_18

- Abdalla HI, Brooks AA, Johnson MR, Kirkland A, Thomas A, Studd JW. Endometrial Thickness: A Predictor of Implantation in Ovum Recipients? *Hum Reprod* (1994) 9(2):363–5. doi: 10.1093/oxfordjournals.humrep.a138509
- El-Toukhy T, Coomarasamy A, Khairy M, Sunkara K, Seed P, Khalaf Y, et al. The Relationship Between Endometrial Thickness and Outcome of Medicated Frozen Embryo Replacement Cycles. *Fertil Steril* (2008) 89(4):832–9. doi: 10.1016/j.fertnstert.2007.04.031
- 42. Gallos ID, Khairy M, Chu J, Rajkhowa M, Tobias A, Campbell A, et al. Optimal Endometrial Thickness to Maximize Live Births and Minimize Pregnancy Losses: Analysis of 25,767 Fresh Embryo Transfers. *Reprod BioMed Online* (2018) 37(5):542–8. doi: 10.1016/j.rbmo.2018.08.025
- Gonen Y, Casper RF, Jacobson W, Blankier J. Endometrial Thickness and Growth During Ovarian Stimulation: A Possible Predictor of Implantation in *In Vitro* Fertilization. *Fertil Steril* (1989) 52(3):446–50. doi: 10.1016/s0015-0282(16)60916-0
- 44. Kasius A, Smit JG, Torrance HL, Eijkemans MJ, Mol BW, Opmeer BC, et al. Endometrial Thickness and Pregnancy Rates After IVF: A Systematic Review and Meta-Analysis. *Hum Reprod Update* (2014) 20(4):530–41. doi: 10.1093/ humupd/dmu011
- Tomic V, Kasum M, Vucic K. Impact of Embryo Quality and Endometrial Thickness on Implantation in Natural Cycle Ivf. Arch Gynecol Obstetr (2020) 301(5):1325–30. doi: 10.1007/s00404-020-05507-4
- Kovacs P, Matyas S, Boda K, Kaali SG. The Effect of Endometrial Thickness on IVF/ICSI Outcome. *Hum Reprod* (2003) 18(11):2337–41. doi: 10.1093/ humrep/deg461
- Kitaya K, Yasuo T. Immunohistochemistrical and Clinicopathological Characterization of Chronic Endometritis. Am J Reprod Immunol (2011) 66 (5):410–5. doi: 10.1111/j.1600-0897.2011.01051.x
- Moreno I, Cicinelli E, Garcia-Grau I, Gonzalez-Monfort M, Bau D, Vilella F, et al. The Diagnosis of Chronic Endometritis in Infertile Asymptomatic Women: A Comparative Study of Histology, Microbial Cultures, Hysteroscopy, and Molecular Microbiology. *Am J Obstet Gynecol* (2018) 218(6):602.e1-602.e16. doi: 10.1016/j.ajog.2018.02.012
- 49. Pietro CD, Cicinelli E, Guglielmino MR, Ragusa M, Farina M, Palumbo MA, et al. Altered Transcriptional Regulation of Cytokines, Growth Factors, and Apoptotic Proteins in the Endometrium of Infertile Women With Chronic Endometritis. *Am J Reprod Immunol* (2013) 69(5):509–17. doi: 10.1111/aji.12076
- Cicinelli E, Matteo M, Tinelli R, Lepera A, Alfonso R, Indraccolo U, et al. Prevalence of Chronic Endometritis in Repeated Unexplained Implantation Failure and the IVF Success Rate After Antibiotic Therapy. *Hum Reprod* (2015) 30(2):323–30. doi: 10.1093/humrep/deu292
- Johnston-MacAnanny EB, Hartnett J, Engmann LL, Nulsen JC, Sanders MM, Benadiva CA. Chronic Endometritis is a Frequent Finding in Women With Recurrent Implantation Failure After *In Vitro* Fertilization. *Fertil Steril* (2010) 93(2):437–41. doi: 10.1016/j.fertnstert.2008.12.131
- Romero R, Espinoza J, Mazor M. Can Endometrial Infection/Inflammation Explain Implantation Failure, Spontaneous Abortion, and Preterm Birth After In Vitro Fertilization? Fertil Steril (2004) 82(4):799–804. doi: 10.1016/ j.fertnstert.2004.05.076
- Wu D, Kimura F, Zheng L, Ishida M, Niwa Y, Hirata K, et al. Chronic Endometritis Modifies Decidualization in Human Endometrial Stromal Cells. *Reprod Biol Endocrinol* (2017) 15(1):16. doi: 10.1186/s12958-017-0233-x
- 54. Kitaya K, Matsubayashi H, Takaya Y, Nishiyama R, Yamaguchi K, Takeuchi T, et al. Live Birth Rate Following Oral Antibiotic Treatment for Chronic Endometritis in Infertile Women With Repeated Implantation Failure. Am J Reprod Immunol (2017) 78(5):e12719. doi: 10.1111/aji.12719
- 55. Sfakianoudis K, Simopoulou M, Nitsos N, Lazaros L, Rapani A, Pantou A, et al. Successful Implantation and Live Birth Following Autologous Platelet-Rich Plasma Treatment for a Patient With Recurrent Implantation Failure and Chronic Endometritis. *In Vivo* (2019) 33(2):515–21. doi: 10.21873/ invivo.11504
- Li F, Cui Y, Zhao D, Bao H, Hao C. Outcome Study of Five Cases Receiving *In Vitro* Fertilization After Treatment of Intrauterine Platelet-Rich Plasma (PRP) for Chronic Endometritis. *Panminerva Med* (2021). doi: 10.23736/S0031-0808.20.04247-0
- 57. Al-Inany H. Intrauterine Adhesions. An Update. Acta Obstet Gynecol Scand (2001) 80(11):986–93. doi: 10.1034/j.1600-0412.2001.801103.x

- March CM, Israel R, March AD. Hysteroscopic Management of Intrauterine Adhesions. Am J Obstet Gynecol (1978) 130(6):653–7. doi: 10.1016/0002-9378 (78)90322-8
- Puente Gonzalo E, Alonso Pacheco L, Vega Jiménez A, Vitale SG, Raffone A, Laganà AS. Intrauterine Infusion of Platelet-Rich Plasma for Severe Asherman Syndrome: A Cutting-Edge Approach. Updates Surg (2020). doi: 10.1007/ s13304-020-00828-0
- Aghajanova L, Cedars MI, Huddleston HG. Platelet-Rich Plasma in the Management of Asherman Syndrome: Case Report. J Assist Reprod Genet (2018) 35(5):771–5. doi: 10.1007/s10815-018-1135-3
- Aghajanova L, Houshdaran S, Balayan S, Manvelyan E, Irwin JC, Huddleston HG, et al. *In Vitro* Evidence That Platelet-Rich Plasma Stimulates Cellular Processes Involved in Endometrial Regeneration. *J Assist Reprod Genet* (2018) 35(5):757–70. doi: 10.1007/s10815-018-1130-8
- 62. Marini MG, Perrini C, Esposti P, Corradetti B, Bizzaro D, Riccaboni P, et al. Effects of Platelet-Rich Plasma in a Model of Bovine Endometrial Inflammation *In Vitro*. *Reprod Biol Endocrinol* (2016) 14(1):58. doi: 10.1186/s12958-016-0195-4
- 63. de Miguel-Gómez L, López-Martínez S, Campo H, Francés-Herrero E, Faus A, Díaz A, et al. Comparison of Different Sources of Platelet-Rich Plasma as Treatment Option for Infertility-Causing Endometrial Pathologies. *Fertil Steril* (2020) 115(2):490–500. doi: 10.1016/j.fertnstert.2020.07.053
- 64. Chien LW, Au HK, Chen PL, Xiao J, Tzeng CR. Assessment of Uterine Receptivity by the Endometrial-Subendometrial Blood Flow Distribution Pattern in Women Undergoing *In Vitro* Fertilization-Embryo Transfer. *Fertil Steril* (2002) 78(2):245–51. doi: 10.1016/s0015-0282(02)03223-5
- Friedler S, Schenker JG, Herman A, Lewin A. The Role of Ultrasonography in the Evaluation of Endometrial Receptivity Following Assisted Reproductive Treatments: A Critical Review. *Hum Reprod Update* (1996) 2(4):323–35. doi: 10.1093/humupd/2.4.323
- Miwa I, Tamura H, Takasaki A, Yamagata Y, Shimamura K, Sugino N. Pathophysiologic Features of "Thin" Endometrium. *Fertil Steril* (2009) 91 (4):998–1004. doi: 10.1016/j.fertnstert.2008.01.029
- Smith SK. Angiogenesis, Vascular Endothelial Growth Factor and the Endometrium. *Hum Reprod Update* (1998) 4(5):509–19. doi: 10.1093/ humupd/4.5.509
- Smith SK. Angiogenesis and Implantation. *Hum Reprod* (2000) 15 Suppl 6:59– 66. doi: 10.1007/s10815-007-9152-7
- 69. Segabinazzi LG, Friso AM, Correal SB, Crespilho AM, Dell'Aqua JA, Miro J, et al. Uterine Clinical Findings, Fertility Rate, Leucocyte Migration, and COX-2 Protein Levels in the Endometrial Tissue of Susceptible Mares Treated With Platelet-Rich Plasma Before and After AI. *Theriogenology* (2017) 104:120–6. doi: 10.1016/j.theriogenology.2017.08.007

- Reghini MF, Ramires Neto C, Segabinazzi LG, Castro Chaves MM, Dell'Aqua Cde P, Bussiere MC, et al. Inflammatory Response in Chronic Degenerative Endometritis Mares Treated With Platelet-Rich Plasma. *Theriogenology* (2016) 86(2):516–22. doi: 10.1016/j.theriogenology.2016.01.029
- 71. Jang HY, Myoung SM, Choe JM, Kim T, Cheon YP, Kim YM, et al. Effects of Autologous Platelet-Rich Plasma on Regeneration of Damaged Endometrium in Female Rats. *Yonsei Med J* (2017) 58(6):1195–203. doi: 10.3349/ ymj.2017.58.6.1195
- Griffith OW, Chavan AR, Protopapas S, Maziarz J, Romero R, Wagner GP. Embryo Implantation Evolved From an Ancestral Inflammatory Attachment Reaction. *Proc Natl Acad Sci U S A* (2017) 114(32):E6566–75. doi: 10.1073/ pnas.1701129114
- Mor G, Cardenas I, Abrahams V, Guller S. Inflammation and Pregnancy: The Role of the Immune System at the Implantation Site. *Ann N Y Acad Sci* (2011) 1221:80–7. doi: 10.1111/j.1749-6632.2010.05938.x
- Caley MP, Martins VL, O'Toole EA. Metalloproteinases and Wound Healing. *Adv Wound Care (New Rochelle)* (2015) 4(4):225–34. doi: 10.1089/ wound.2014.0581
- Zhang S, Li P, Yuan Z, Tan J. Platelet-Rich Plasma Improves Therapeutic Effects of Menstrual Blood-Derived Stromal Cells in Rat Model of Intrauterine Adhesion. Stem Cell Res Ther (2019) 10(1):61. doi: 10.1186/s13287-019-1155-7
- 76. Zhou Y, Shen H, Wu Y, Zhao X, Pei J, Mou Z, et al. Platelet-Rich Plasma Therapy Enhances the Beneficial Effect of Bone Marrow Stem Cell Transplant on Endometrial Regeneration. *Front Cell Dev Biol* (2020). 8:52. doi: 10.3389/ fcell.2020.00052

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