

Biomaterials for Interbody Fusion in Bone Tissue Engineering

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In recent years, interbody fusion cages have played an important role in interbody fusion surgery for treating diseases like disc protrusion and spondylolisthesis. However, traditional cages cannot achieve satisfactory results due to their unreasonable design, poor material biocompatibility, and induced osteogenesis ability, limiting their application. There are currently 3 ways to improve the fusion effect, as follows. First, the interbody fusion cage is designed to facilitate bone ingrowth through the preliminary design. Second, choose interbody fusion cages made of different materials to meet the variable needs of interbody fusion. Finally, complete post-processing steps, such as coating the designed cage, to achieve a suitable osseointegration microstructure, and add other bioactive materials to achieve the most suitable biological microenvironment of bone tissue and improve the fusion effect. The focus of this review is on the design methods of interbody fusion cages, a comparison of the advantages and disadvantages of various materials, the influence of post-processing techniques and additional materials on interbody fusion, and the prospects for the future development of interbody fusion cages.

Keywords: 3D printing, interbody fusion cage, titanium, PEEK, absorbable material, growth factors, ceramic material, coating

1 INTRODUCTION

With population aging, spinal degenerative diseases have become a health problem that cannot be ignored, with a lifetime incidence of >50% (Chong et al., 2016; Lim et al., 2019; May et al., 2019; Warren et al., 2020). In the spinal orthopedics clinic, when nerve compression persists for a long time and conservative treatment fails, decompression surgery has remained the first choice for the treatment of intervertebral disc degeneration, but interbody fusion is crucial for the treatment of impaired vertebral stability, failure of intervertebral disc surgery, spinal deformity and spinal stenosis. (Jain et al., 2016; Mena et al., 2016; Phan and Mobbs 2016; Assem et al., 2017; Junes et al., 2020). The purpose of this operation is to decompress the nerve root, maintain the height of the intervertebral disc, and achieve bony fusion (Folman et al., 2003; Hoy et al., 2013; Hoy et al., 2019; Fan and Guo 2020). It means we need to convert fibrocartilage joints into hard and continuous bone segments (Smoljanovic et al., 2010). The autogenous iliac bone graft is the gold-standard method for spinal fusion (Vanek et al., 2012; Farrokhi et al., 2017). Harvested autologous bone slices have better osteoinductive properties compared to allogeneic bone grafts (Huang et al., 2014; Grgurevic et al., 2020). However, autologous bone grafts are limited by additional surgical trauma, donor site complications, and the inadequate autogenous bone supply (Sekerci et al., 2013; Fomekong et al., 2017). Fusion failure is often caused by cage loosening and related complications (5-35%), so

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TABLE 1 | Experimental studies published recently.

Study	Additive	Scaffold	Technique	Animal Model	Results	
(Zhang et al.)	SIM/poloxamer 407 Hydrogel	Ti	3D-printing	Rhesus Macaque	Promoted bone ingrowth and spinal fusion	
Liang et al. (2020)	-	HA/PA66	Infiltrating	Goat	Effectively improved intervertebral bony fusion	
Rodríguez-Vázquez and Ramos-Zúiga (2020)	_	Ch/HAp	Freeze-drying	Rat	Bone regeneration in a solid and well-structured fusion was induced	
Chu et al. (2019)	lliac crest bone graft	CS/PEEK (4:6)	Fabricated by compounding and injection-molding	Goat	Induced highly effective bone fusion	
Gunzburg et al. (2019)	Autologous bone	PEEK	Ti coating	Ovine	Nano-surfaced cages resulted in greater spinal stiffness changes	
Lu et al. (2019)	_	Tantalum	3D-knitted	Rabbit	Showed good biocompatibility and osteocompatibility	
Walsh et al. (2018)	-	PEEK	Coating	Sheep	Newly formed bone tracked along the thin Ti- coated surface	
McGilvray et al. (2018)	_	Titanium	3D-printing	Ovine	Reduced range of motion and increased bone ingrowth and construct stiffness	
(Kirk et al., 2017)	lliac crest bone graft	PEEK titanium composite (PTC)	Hybrid additive manufacturing approaches	Ovine	PTC constructs demonstrated significant reductions in range of motion and a significant increase in stiffness compared to PEEK and PSP	
Ren et al. (2017)	_	MAACP/HA/CS	_	Goat	Provided a good fusion effect, enough biomechanical stability, and close integration with the surrounding bone	
Bae et al. (2016)	RhBMP-2	PEEK	-	Sheep	RhBMP-2 dose-dependent osteoclastic resorption is a transient phenomenon	
Wheeler et al. (2016)	mesenchymal precursor cells (MPCs)	PEEK	_	Ovine	MPCs are an alternative to autologous for lumbar interbody fusion procedures	

choosing an appropriate interbody cage or not directly affects the success rate of surgery (Virk et al., 2017).

The ideal bone-substitute material should be characterized by favorable bone induction, good biocompatibility, suitable mechanical strength, and a high performance-to-price ratio (Warburton et al., 2019; Verma et al., 2020). At present, the commonly used materials for interbody fusion cages can be divided into metallic and non-metallic options (Park and Lehman, 2020; Verma et al., 2020). The most commonly used interbody cages are made of non-absorbent materials, such as titanium (Ti) and poly-ether-ether-ketone (PEEK). Ti alloy may cause implant subsidence and segmental instability due to its high elastic modulus, and the use of PEEK is limited due to its poor biocompatibility leading to chronic inflammatory responses (Guo et al., 2020). Absorbable materials include magnesium (Mg) and other polymer materials. With the mechanical strength weakens due to degradation, the materials may not be able to provide sufficient support and lead to the interbody fusion failure. Therefore, it is necessary to explore new materials that promote interbody fusion.

Three-dimensional (3D) printing is a new and widely popular technology (Egan et al., 2018; Burnard et al., 2020). Most importantly, 3D-printed cages can be customized for individual patients to achieve a high degree of implant-toanatomy match. An appropriate interbody cage is designed to maintain the height of the intervertebral body and keep the upper and lower vertebral bodies stable. In addition, the internal porous structure made by 3D-printing technology can induce new bone ingrowth, which realizes the fusion of the upper and lower vertebral bodies to achieve biological stability (Zhang et al., 2020a). Osseointegration can be further improved by making surface modifications and the addition of drugs and growth factors (Park and Lehman, 2020). Coating is a commonly used surface-modification technology, and better biological activity and osteogenic ability can be obtained with coating materials such as Ti and hyaluronic acid (HA). Commonly used drugs include simvastatin (SIM) and strontium ranelate (SRR), which can improve the local microenvironment and promote osteogenesis in patients with osteoporosis. Growth factors, such as bone morphogenetic protein (BMP) and epidermal growth factor, can promote interbody fusion by regulating the expression of osteogenic genes. And we list all Experimental studies and Clinical studies in Table 1 and Table 2 respectively.

Accordingly, we wrote this review to address above issues. Review was retrieved in PubMed databases and Web of Science by the keywords: "interbody fusion", "cage". A total of 658 papers were picked, we excluded some earlier published research and clinical articles, and 171 articles were finally selected for this review according to the criteria of inclusion and exclusion. This review introduces the application of interbody fusion cages in bone tissue engineering in detail and focuses on the design methods of interbody cages, including 3D printing, porosity design. The advantages and disadvantages of interbody cages made of different materials, such as Ti, tantalum, Mg, PEEK, and ceramics, are compared and improved methods are proposed. The effects of material post-processing techniques on interbody fusion are summarized, including coating techniques, and the effects of the internal addition of SIM, BMP, and SRR on

TABLE 2 | Clinical studies published recently.

Study	Additive	Scaffold	Technique	Case Number	Results
Mobbs et al. (2019)	Allograft	TI	3D-printing	1	Rapid recovery with significant fusion effects
Zippelius et al. (2018)	_	TI	3D-printing	50	Implant is safe and led to a very high fusion rate
Mokawem et al. (2019)	-	SiCaP- packed Tl	3D-printing	93	Provided excellent rates of solid fusion in TLIF and LLIF surgeries
Ohanisian and Dorsi (2019)	-	TI	3D-printing	1	Stimulates osteogenesis and enhances fusion with a remarkable improvement of symptoms
Arts et al. (2020)	_	TI	3D-printing	50	The fusion rates of porous titanium and PEEK with autograft were similar
Siu et al. (2018)	_	TI	3D-printing	30	The subsidence rate was lower compared to that of the PEEK implant
Krafft et al. (2019)	_	ТІ	3D-printing	1	The difficulties in deformity correction secondary to osteoporotic fractures were overcome

interbody fusion are discussed. In addition, the application challenges of different interbody cages and directions for further exploration are summarized.

STRATEGIES FOR DESIGNING INTERBODY FUSION CAGES

Traditional Interbody Fusion Cages

Before an interbody fusion cage design is chosen, the anatomical location, surgical approach, and implant size need to be considered. The anatomical model may be obtained through a computed tomography (CT) examination, and the matching interbody fusion cage can then be designed accordingly (Duan et al., 2016; Rieger et al., 2017; Sen et al., 2019; Gkt et al., 2020). Combined with the CT data of the intervertebral disc, the computer-aided design software (CAD) with computer-aided manufacturing (CAM) technology was used to design the 3D model of the interbody cage, and then the fused deposition modeling (FDM) and selective laser melting (SLM) technology were used to print interbody cage (Chen et al., 2019; Bassous et al., 2019). The anatomical data of the target implant area include the left-right diameter, anteroposterior diameter, intervertebral height, and the angle between the upper and lower surfaces. In addition, the increase in intervertebral height caused by the destruction of the upper and lower endplates during surgery should also be considered. Undersized implants may cause subsidence, while oversized implants can cause damage to neural structures due to excessive compression (Feng et al., 2014; Girod et al., 2017; Zhang et al., 2020b). The angle between the upper and lower surfaces mentioned above suggests the postoperative physiological curvature of the spine. A proper angle in this context will keep the posterior longitudinal ligament relatively tight after implantation, preventing the posterior longitudinal ligament from going slack and compressing the nerve. Moreover, an optimal angle can also avoid overstretching of the anterior longitudinal ligament, which may cause vertebral instability. Due to the different stages of fusion and the individual differences of the target, the designed interbody cages have various shapes, such as a kidney shape (Madan and Boeree 2003). Joffe et al. obtained previous data through CT imaging and designed a corresponding cervical fusion cage, then carried out 3D printing and implantation in a canine

cervical interbody fusion model. The implants were not ground or polished in this context because the rough surface structure increased the area of bone contact to facilitate bone growth (Joffe et al., 2019). A customized intervertebral device restores the intervertebral space and achieves a certain degree of interbody fusion (Joffe et al., 2019). In addition, by designing hollow cages, the implantation of autologous bone and other implants can be ensured to promote osseointegration. Walsh et al. designed an interbody fusion cage (measuring 4.5 mm high, 10 mm wide, and 20 mm long) with a central hole for transplant material injection, which was printed to fit the L4-L5 intervertebral space of sheep, providing support for clinical implantation (Walsh et al., 2018). The design of the fixed device cannot be ignored, and the nailboard system is the most commonly used method applied to fixation (Hah et al., 2020). Fixed device is generally used to maintain the stability of the intervertebral space for a better fusion effect (Caplan et al., 2020).

Porous Structures

The interconnected micropore structure of an interbody fusion cage can improve the bone-conduction ability, which also benefits cell proliferation and differentiation and bone regeneration. The pore size of human cancellous bone is 500-600 µm (Lim et al., 2019). Considering porous scaffolds applied in bone tissue engineering, the porosity should be >50%, especially in the range of 65-80%, where the structure and elasticity modulus are similar to those of human trabeculae (Zhang F et al., 2018; Bai H. T. et al., 2020). The stress between the cage and the endplates decreases with increasing porosity, while the range of motion of the vertebral bodies decreases with increasing fusion rates of the upper and lower vertebral bodies. Previous experiments have shown that a pore size of 50–500 μ m is beneficial for the adhesion, proliferation, and differentiation of osteoblasts (Zhang Z et al., 2018; Bai H. et al., 2020). In a comparison of goat interbody fusion models, McGilvray et al. found that the range of motion between the vertebral bodies of the animals using porous Ti interbody cages was significantly reduced and the stability of the interbody fusion was increased, indicating that it obtained a better fusion effect. The reason for this is that bone growth occurs in the cage's microporous structure, resulting in a greater overall mechanical structural stability and more effective fusion (McGilvray et al., 2018). The holes, in addition to providing space for new bone to grow in, can be used for the



injection of bioactive structures, such as BMPs, antibiotics, and bioceramics, as well as for post-processing for better fusion results (Schnitzer et al., 2020).

Other Special Structures

In designing the interbody fusion cage, there are some special options available to facilitate certain tasks, such as reducing the difficulty of surgery or promoting the fixation of adjacent vertebral bodies by integrating the cage and the fixator. Sasso et al. designed a hollow threaded cylinder with a removable end cover that provides strength while reducing the weight and increasing the space available for the bone autograft. Each cylinder component has multiple holes for implanting the head and tail as well as small transverse holes to enhance the vascularization of the graft in the device. The matching internal fixation device is then designed correspondingly. A good fusion rate was obtained in patients implanted with this device (Sasso et al., 2004) (Figure 1). In addition, the threaded structure on the surface of the fusion cage can enhance interbody fusion and reduce the occurrence of postoperative complications, causing autogenous bone transplantation to rarely be needed (Hacker et al., 2000; Burkus et al., 2002). The screw trajectory design, planned screw length, and device matching the patient's unique endplate anatomy should be considered when 3D-printing the threaded hole design to ensure uniform force on the endplate and device and to aid in fixation (Park et al., 2009; Caffrey et al., 2018; Easley et al., 2018). Mobbs et al. reported custom features with pre-angled threaded holes

designed to enhance post-implantation fixation and endplate interfaces that match patient anatomy to ensure even loading on the endplates and device. Their patient recovered quickly after surgery, and their clinical symptoms were significantly improved (Mobbs et al., 2019). Bionic structures can also be considered. Zippelius et al. designed a "banana-shaped" cage made of Ti with curved rails on the surface that can help embed the endplates of the vertebrae, securing the cage to the desired location on the ventral edge strip (such as on the rails) while extending the surface and facilitating better skeletal integration, and they obtained a good fusion rate in 60 patients by using this approach (Zippelius et al., 2018). In addition, Kim et al. designed a scalable interbody device (inserted in the form of contraction and expanded *in situ* after being correctly located in the intervertebral space), which is suitable for minimally invasive interbody fusion. Clinical validation found that this device resulted in improved patient-reported clinical outcomes, recovery of disc height, and better fusion rates together with reduced risks of implant displacement, subsidence, rupture, and collapse compared to static devices (Kim et al., 2016) (Figure 2).

3 INTERBODY FUSION CAGE MATERIALS

Ti and Its Alloy Material (Ti₆Al₄V)

 $\rm Ti_6Al_4V$ is typically selected as the raw material for the manufacture of interbody cages because of its strength,



excellent corrosion resistance, low density, biocompatibility, relatively low cost, and compatibility with magnetic resonance imaging (Dai and Jiang 2008; Knutsen et al., 2015; Burkus et al., 2017). The artifacts of Ti alloy generated on CT images are fewer than those associated with other metals, so the condition of the implant can be monitored after surgery (Joffe et al., 2019). 3Dprinted porous Ti₆Al₄V, which can be prepared by selective laser melting or electron beam melting, has a low elastic modulus similar to that of cortical bone, so it is a promising orthopedic scaffold material (Liu et al., 2016; Walsh et al., 2019). Advances in 3D-printing technology have enabled the manufacture of Ti cages with complex internal geometries. Postoperative follow-up of 93 patients who received transforaminal lumbar interbody fusion (TLIF) and lateral lumbar interbody fusion (LLIF) using silicatesubstituted calcium phosphate (SiCaP)-packaged 3D printing lamellar Ti cages was performed by Mokawem et al., and their results demonstrated that the 3D-printed lamellar Ti cages packaged with SiCaP ensured excellent solid fusion rates during TLIF and LLIF surgeries (Mokawem et al., 2019). 3D printing of Ti makes prostheses rougher. Modifying the surface of an implant, such as its surface roughness and morphology, promotes bone consolidation and prevents the implant from falling off of the vertebral body (Mobbs et al., 2019). Ti implants are beneficial for bone growth because of their metal

properties (i.e., valence charge), which attract neighboring proteins, cells, and body fluids (Cheng et al., 2020). In a previous study, however, Hauerberg *et al.* found that, for cervical radiculopathy caused by intervertebral disc herniation, there is no difference between simple discectomy and Ti cage fusion after discectomy (Hauerberg et al., 2008).

Ti has a high elastic modulus such that the porous Ti layer hardly deforms under compression during implantation or under physiological loading (Gunzburg et al., 2019). Arts et al. found that the fusion rate of porous Ti was similar to that of autologous bone-filled PEEK at 12 months during a prospective study of single-segment anterior cervical discectomy and fusion (ACDF) patients using 3D-printed porous Ti neck implants (Arts et al., 2020). Through a clinical comparison, Krafft et al. found that the sinking rate of 3D-printed porous Ti interbody fusion cages was significantly lower than that of PEEK interbody fusion cages (Krafft et al., 2020). 3D-printed lamellar Ti cages filled with SiCaP bone grafts can also achieve promising fusion rates in adult degenerative diseases by providing more consistent bone growth and biological fixation (McGilvray et al., 2018; Mokawem et al., 2019). However, the Ti₆Al₄V material has a high elastic modulus. Due to the different elastic moduli of cortical bone, the spinal fusion cage easily sinks, resulting in a decrease in intervertebral height, pedicle stenosis, and so on



(Seaman et al., 2017). At present, through the use of 3D-printing technology, Ti_6Al_4V materials have been successfully used in artificial hip replacements, semi-pelvic replacements, and artificial vertebral replacements in patients with spinal tumors. The advantages of 3D-printing technology in creating medical Ti alloys are gradually being highlighted due to the abundant processing methods in play (Niu et al., 2019).

Tantalum

Tantalum has been increasingly used in the field of orthopedics because of its good histocompatibility, strong corrosion resistance, and so on (Zardiackas et al., 2001; Veillette et al., 2006). Lu *et al.* demonstrated through *in vitro* and *in vivo* experiments that porous tantalum implants can achieve good fusion, and tantalum is expected to be an effective biomaterial for interbody fusion cages. Porous tantalum has a structure and mechanical properties to similar to those of human bone, allowing trabecular bone to grow into the pores of a cage implant to achieve better osseointegration. In addition, tantalum stimulates cell proliferation and enhances the osteogenic capacity of human osteoblasts, improving vertebral fusion efficiency (Lu et al., 2019) (**Figure 3**). Above all, compared to autogenous iliac bone graft, the adverse events in donor site can be avoided. On the other hand, tantalum has been reported in the treatment of infectious bone defects and bone tumors, which can be used in interbody fusion and spinal metastases after vertebrectomy (Hua et al., 2020; Wang J et al., 2020). At present, there are few reports about tantalum interbody fusion cages in the clinic and experiments, and its effect and characteristics need to be further verified (Iv, Beutler et al., 2010).

PEEK

PEEK is a kind of high-molecular-weight semi-crystal polyaromatic linear polymer and thermoplastic material characterized by biological inertia, radiation transmission, low density, strong corrosion resistance, and a bone-like hardness (Basgul et al., 2019). Compared to alloy, PEEK can minimize stress shielding and bone resorption around the implant to avoid implant loosening (McGilvray et al., 2018; Chu et al., 2019). However, the formation of a biofilm on the surface of PEEK inhibits its binding to the host bone, preventing solid fusion (McGilvray et al., 2017; Gunzburg et al., 2019). In addition, PEEK



observation; representative X-ray images with red arrows indicate the bone ingrowth. (C) 3D and longitudinal images of the interbody fusion at 4, 12, and 26 weeks based on the micro-CT scans. (D) Analysis of osseointegration at 4, 12, and 26 weeks based on histological assessment, including Van Gieson staining and fluorescence labeling by calcein (Chu et al., 2019).

cages can cause local inflammation, which may lead to bone nonunion and osteolysis (Mokawem et al., 2019; Cheng et al., 2020). In recent years, the technology for 3D-printing PEEK interbody cages has matured, and there have been reports of its successful clinical application. Because of PEEK high melting point, laser sintering is commonly used, and the mechanical properties, biocompatibility, and heat resistance of PEEK can be effectively improved by adding in other inorganic materials and polymers (Tan et al., 2005). It is reported that the fusion rate of PEEK interbody devices is similar to that of allogeneic bone grafts (Jain et al., 2020).

The calcium silicate (CS)/PEEK composite used by Chu *et al.* was prepared by infusing CS into PEEK through compounding and injection-molding techniques. According to the mechanical property test results, the best ratio of CS is 40% (wt%). Through the analysis of the results from a goat interbody fusion model, it was found that the CS/PEEK composite had good biomechanical stability, osteogenic effects, and osseointegration ability. This bioactive material has great potential to facilitate the

development of clinical interbody fusion cages (Chu et al., 2019) (Figure 4).

McGilvray et al. found that, compared to the standard PEEK stent, the range of motion of a PEEK-Ti composite stent made using a mixture of PEEK and Ti was significantly reduced, the hardness was significantly increased, and the fusion effect was better (McGilvray et al., 2017). Dai et al. confirmed in their study that a PEEK interbody fusion cage containing β -tricalcium phosphate was efficient for the treatment of cervical spondylosis (Dai and Jiang 2008). The elastic modulus of the PEEK interbody fusion cage is similar to that of vertebral cortical bone, which is beneficial for the dispersion of load and stress and greatly reduces the probability of the cage sinking (Lim et al., 2019). William et al. found that the introduction of HA into a scaffold made of PEEK matrix allowed for direct boneattachment growth and exhibited better fusion in a goat cervical fusion model. Although PEEK cages with HA can improve interbody fusion, there are no clinical studies to support their use (Walsh et al., 2016) (Figure 5). Also,



although PEEK interbody fusion cages are relatively mature

structures, their composite materials need to be further explored.

Absorbable Materials

Absorbable materials have relatively good biocompatibility and can be hydrolyzed or enzymatically hydrolyzed into non-toxic small molecules that can be absorbed by organisms, with some even participating in biological metabolism (Hojo et al., 2005). Polylactide (PLA), alkyl polyglycoside, and polydioxane are considered safe and sufficiently stable absorbable materials, while iron, Mg, and zinc are the main absorbable metal elements. Although the inconvenience of a second surgery can be avoided by harvesting biomaterials, the possible complications, such as soft-tissue reactions, unwanted osteolysis, and low primary mechanical loading capacity, also limit the use of absorbable interbody fusion cages (Ulum et al., 2019; Pisecky et al., 2021). The hardness of the bioabsorbable fusion cage is similar to that of bone, which may accelerate interbody fusion. Meanwhile, as the fusion cage degrades, the load is gradually transferred to the healed bone to aid with bone remodeling. At present, PLA is the most commonly used bioabsorbable scaffold material, but rapid degradation and osteolysis around the scaffold have been reported (Ren et al., 2017). In a previous study, Lu et al. compared a novel PLA/nanoscale \beta-tricalcium phosphate bioabsorbable self-retaining cervical cage (BCFC) with an autologous bone graft and PEEK cage for anterior cervical discectomy and interbody fusion and found that the BCFC device had better fusion than the autologous bone graft or PEEK cage. This device may be a potential alternative to current PEEK cages (Lu et al., 2017). Knutsen et al. designed a bioabsorbable polycaprolactone (PCL) cervical cage and tested its compression, compression-shear, and torsion, concluding that PCL bioabsorbable cages may require additional fixation to achieve the effect of fused vertebral bodies (Knutsen et al., 2015). Through a clinical comparison, Jiya et al. found that absorbable poly-L-lactide-co-D,L-lactide (PLDLLA) implants

are less effective in assisting with spinal fusion than PEEK implants, have a lower fusion rate, and are more prone to subsidence and osteolysis, so their benefits need to be further proven. The sterilization of PLDLLA cages by electron beam irradiation may lead to preliminary cage rupture, which is the main cause of mechanical failure (Jiya et al., 2009; Jiya et al., 2011). Ren et al. evaluated the fusion effect, biomechanical stability, and histological characteristics of a novel absorbable copolymer/nano-hydroxyapatite/calcium poly-amino acid sulfate (MAACP/HA/CS) interbody fusion cage in a goat cervical fusion model. Compared to a Ti cage and autogenous bone graft, the MAACP/HA/CS cage had a better fusion effect, combined closely with the surrounding bone, and maintained an appropriate intervertebral disc height (Ren et al., 2017). Although the absorbable material interbody fusion cage attained a good fusion rate because of its excellent biocompatibility and easy absorption, given its high failure rate, its clinical application needs to be further explored and studied.

Mg has a similar mechanical behavior to natural bone, good bone conduction activity, and radiation transparency, so it is widely regarded as a potential ideal absorbable orthopedic material superior to traditional metal and other biodegradable implants (Wang Y et al., 2020). The density of Mg and its alloy is close to that of human bone mineral, and its elastic modulus is about 45 GPa, which is lower than that of Ti₆Al₄V, tantalum, and other implantable metals (Staiger et al., 2006). Furthermore, it has been shown that implant-derived Mg induces local neuronal production of calcitonin gene-related polypeptide-a to improve bone-fracture healing (Zhang et al., 2016). The Mg²⁺ released from the degradation of Mg-based implants is considered an effective material to promote osteogenesis, but there are few studies on Mg-based implant fusion. Through degradation, interbody fusion, and biocompatibility testing of a goat cervical vertebra model, Guo et al. found that Mg-based interbody fusion cage had better histological fusion, but the total fusion area needed to be improved. This is the first report of successful histological fusion of a Mg-based interbody fusion cage (Guo et al., 2020). The main obstacle to limiting the clinical application of Mgbased cages is the adverse reactions caused by the rapid degradation of Mg. The degradation rate is faster in the first 3 weeks after implantation, then slows down gradually thereafter. However, excessive Mg accumulation caused by rapid implant corrosion will lead to a severe foreign body reaction, tissue stimulation, decreased mechanical strength of new bone, and abnormal precipitation of calcium, which will eventually hinder osteogenesis. Moreover, Mg generates a large amount of hydrogen during the corrosion process, resulting in tissue loosening or excessive pressure, which is not conducive to bone formation (Wessels et al., 2012). This may be because Mg scaffolds corrode faster than new bone formation, leading to local scaffold collapse, resulting in differences in intervertebral pressure and ultimately leading to fusion failure (Staiger et al., 2006). In an experiment of a silicon-coated Mg alloy (AZ31) fusion cage in a goat cervical vertebra model, Zhang et al. found that excessive Mg accumulation could inhibit the formation of new bone, the intervertebral Mg ion concentration increased

significantly after implantation, and the intervertebral Ca/P ratio decreased under the condition of high Mg accumulation, which may be the main reason for the failure of interbody fusion (Zhang F et al., 2018). However, there is only a single case of successful fusion in the literature; moreover, even after modification to balance the degradation rate, the fusion region is still too small to meet clinical needs, so more research is needed to improve it. It is a feasible method to slow down the degradation rate of Mg by reducing the purity of Mg (Chang et al., 2020). At present, Mg as an intraspinal plant is still in an exploratory stage, and its clinical application requires more experimental support.

Ceramic Materials

Bioceramics have inorganic components similar to natural bone which are widely used for bone tissue scaffolds fabricating. Due to its similar composition to natural bone, bioceramics present a significant osteogenic differentiating capability (Dxa et al., 2020). Bioceramic materials consist of calcium phosphate ceramics (CaP) such as hydroxyapatite (HA), tricalcium phosphate (TCP), and bioglass (Zhao et al., 2021). It is known that the biologically active CaO-SiO2-P2O5-B2O3 glass-ceramic can chemically combine with bone to form a carboxyapatite layer to promote the differentiation of human bone marrow mesenchymal stem cells into osteoblasts. Li et al. found that the fusion rate and improvement of clinical symptoms of CaO-SiO₂-P₂O₅-B₂O₃ glass-ceramic spacers were similar to those of Ti cages. In an in vivo model, cylindrical implants showed better osseointegration with adjacent bones than HA, and the surface coating was found to improve the osseointegration of the implant, while its compressive and flexural strength were ≥ 2 times that of HA alone. The compressive strength of CaO-SiO₂-P₂O₅-B₂O₃ glass-ceramics is 4 times that of PEEK and 1.3 times that of Ti, respectively. However, this increased mechanical strength correlates with a greater risk of sinking in patients with osteoporosis (Lee et al., 2016). Ceramic implants made of silicon nitride show better biocompatibility and bone conductivity and are expected to reduce complications like subsidence and displacement. Kersten et al. compared a silicon nitride interbody fusion cage and PEEK interbody fusion cage in patients with symptomatic degenerative lumbar disc disease for the first time, but their results have not yet been published (Kersten et al., 2014). HA is a widely used ceramic biomaterial because it is the basis of bone tissue. In addition, it also provides enough matrix for the endogenesis of tissue in the process of bone regeneration. It generally needs to be used in combination with other materials. When the scaffold is made of chitosan and HA in a ratio of 20-80, scaffold has both elasticity and an osteogenic ability. In a mouse spinal fusion model, a chitosan/HAp composite scaffold showed good biocompatibility and sufficient spinal stability and induced solid and well-structured bone regeneration (Rodriguez-Vazquez and Ramos-Zuniga, 2020). In addition, a hollow HA/polyamide 66 stent (HA/ PA66) has been used in anterior cervical reconstruction. Liang et al. verified in a goat cervical fusion model that porous HA/ PA66 with a 300-µm pore size is more conducive to bone growth and can more effectively promote interbody fusion, providing a



histology in the middle of the apertures at 4 and 8 weeks in the sagittal plane (Walsh et al., 2018). potential application prospect for intervertebral reconstruction Gunzburg *et al.* verified

4 POST-PROCESSING TECHNIQUES

after vertebrae resection (Liang et al., 2020).

Coating

Coating technology is also widely used in orthopedic implant modification. The coating of the cage can modify the surface of the material to amend and increase its surface roughness without changing its biomechanical properties. Coating an interbody cage reduces the vertebral body subsidence and pseudoarthrosis (Rao et al., 2014; MacBarb et al., 2017). Coating techniques, including plasma spraying, chemical vapor deposition, metal–organic chemical vapor deposition, electrochemical vapor deposition, molten coating, physical vapor deposition, thermal or diffusion conversion, and sol-gel, have been used to deposit micron and nanometer coatings on various substrates, including Ti and its alloys. Nano-coating technology refers to thin films with thicknesses of 1–1,000 nm or those less than micro-coatings (<1 μ m) (Choi et al., 2020). Here, we will introduce the coating techniques commonly used in interbody fusion.

Gunzburg et al. verified that a PEEK material coated with nano-Ti can achieve greater spinal stiffness than a pure PEEK material in spinal fusion using a goat model (Gunzburg et al., 2019). The Ti coating on PEEK retains the physical properties of the PEEK substrate and contains a surface that reacts better with the host. Ti-surfaced PEEK implants allow for earlier and better fusion of new bone formation and remodeling by increasing the anchorage of osteoblasts in the endplates and wells. The Ti coating aids in bone formation and remodeling. This is stimulating accomplished by the production of osteoprotegerin, transforming growth factor (TGF)-\u03b31, vascular endothelial growth factor (VEGF) A, fibroblast growth factor (FGF), and angiopoietin 1. Local levels of these factors were higher in Ti-coated implants than smooth Ti₆Al₄V or PEEK alloys (Bassous et al., 2019; Ohanisian and Dorsi 2019). In a prospective evaluation of 45 patients undergoing ACDF, Krayenbuhl et al. achieved good interbody fusion with a low complication rate by placing an empty plasma hole-covered Ti cage after anterior cervical microsurgery. Disc height and lordosis can be preserved with a low incidence of subsidence and good fusion rates (Krayenbühl et al., 2008). Walsh et al. (2018) found that newly formed bone grew along the surface of a nano-



Ti-coated PEEK fusion cage in a sheep model. On the other hand, the surface of PEEK cage shows non-reactive fiber tissue at the interface (**Figure 6**). The coated PEEK fusion cage had good adhesion and proliferation of bone marrow stromal cells (BMSCs) and can induce a higher cell growth rate and alkaline phosphatase level, which can promote bone growth (Liu et al., 2017).

The common method to enhance osteoblasts' response to a material is to apply HA to the surface of cage (MacBarb et al., 2017; Walsh et al., 2018). Sierra *et al.* 3D-printed a lumbar cage with HA coating and found that it had good biocompatibility through *in vitro* analysis, characterization, and testing (Serra et al., 2016). McBarb *et al.* designed and compared Ti plasma spray (TPS) coatings with and without nano-crystalline HA coating achieved better cell proliferation and osteogenic capacity compared to the uncoated TPS, but the difference was not statistically significant (MacBarb et al., 2017) (**Figure 7**).

The natural extracellular matrix protein is considered a candidate material for the development of biomaterials that can induce specific cell behaviors. As a surface coating of biomaterials, the purified protein can change the proliferation and differentiation of stem cells and the retention of growth factors, which provides a new idea for the coating technology of

printing materials. This could be explored in future experiments (Xing et al., 2020).

Internal Fillings

Obtaining good biocompatibility of fillers through internal filling of stents, promoting local proliferation, and obtaining a good osteogenic ability to promote interbody fusion are commonly used methods in basic experiments and clinical practice of spine surgery. Additives such as drugs, growth factors, and platelet-rich plasma (PRP) are added to the pre-designed porous interbody fusion cage by means of negative pressure suction, direct filling, and immersion (Bai H. T. et al., 2020; Leng et al., 2020).

4.2.1 Drugs

4.2.1.1 Simvastatin

Over the years, statins have been widely used to lower cholesterol and reduce the risk of heart attack (Song et al., 2003). SIM, an inhibitor of 3-hydroxy-3-methylglutaryl coenzyme A reductase (HMG-CoA), which has the advantages of a low price, good safety, and ability to promote bone growth in a prosthesis (Jin et al., 2021). SIM overcomes the shortcomings of the high cost and short half-life of biological factors like BMP-2 and VEGF used in previous studies (Zhang et al., 2020a). Topical application of SIM has a good effect on bone formation. Moreover, local



application of PLA has achieved good results in experiments of bone formation promotion and bone defect healing (Zhao et al., 2014; Tan et al., 2015). Using porous Ti cage filled with SIM/ poloxamer 407 hydrogel in a rhesus monkey interbody fusion model, Zhang *et al.* found that 3D-printed porous scaffolds containing SIM hydrogel promoted bone growth and spinal fusion. They also speculated that, in the early stage of osteogenesis, the continuous release of SIM from the hydrogel promotes the production of endogenous biological factors, such as BMP-2 and VEGF, which are key regulators of bone formation and angiogenesis during bone regeneration. However, with the passage of time, due to the body's metabolism, the osteogenic effect of SIM gradually decreases (Zhang et al., 2020b) (**Figure 8**).

4.2.1.2 Strontium Ranelate

SRR is an oral drug used for the prevention and treatment of osteoporosis. Different from other anti-osteoporotic drugs, SRR has unique dual effects on bone formation and bone resorption. At the same time, it reduces bone decomposition, stimulates bone reconstruction, prevents bone loss, improves bone strength, and reduces the risk of fracture (Rodrigues et al., 2018; Demirel and Kaya 2020). However, oral SSR may have no synthetic metabolic effect on human bone formation et al., 2018, Marx et al., 2020). Tai *et al.* found that SRR treatment had a relatively mild effect on bone strength and bone remodeling and confirmed its better interbody fusion through histological and mechanical tests in a mouse interbody fusion model, suggesting that SRR treatment can be used as a replenish for human interbody fusion, although an interbody fusion cage was not used in this mouse model. This experiment provides a new idea for interbody fusion under the umbrella of osteoporosis (Tsai et al., 2017).

4.2.2 Platelet-Rich Plasma

PRP obtained by centrifugation is a plasma product containing high concentrations of various growth factors, including plateletderived growth factor, insulin-like growth factor (IGF), and TGF- β . PRP is known to play an important role in fracture healing. A large number of studies have shown that PRP-modified scaffolds have a positive effect on bone regeneration and can significantly promote the repair of bone defects (Lowery et al., 1999; Carreon et al., 2005; Leng et al., 2020; Sochacki et al., 2020; Andrade et al., 2021). The application of PRP in a rat spinal fusion model significantly accelerated the speed of interbody fusion without any complications (Shiga et al., 2016). In addition, lyophilized PRP is also widely used due to its advantage of easy storage. Compared to traditional PRP, lyophilized PRP–coated Ti more strongly promoted the cell viability and osteogenic differentiation of BMSCs and triggered better bone formation. Interbody fusion provides a new idea (Qiao et al., 2020). It can be seen from previous literature that there are no animal experiments of interbody fusion cages combined with PRP, but PRP's role in promoting interbody fusion has been tested alone, which provides a direction for future experiments and research.

4.2.3 Growth Factors

Soluble growth factors obtained by protein separation and molecular cloning techniques include TGFs, BMPs, IGFs, FGFs, and epidermal growth factor. However, only some of these growth factors showed a significant bone-induction ability (Kandziora et al., 2003).

4.2.3.1 Bone Morphogenetic Protein

BMP has attracted wide attention because of its inherent potential to improve the process of ossification and induce pluripotent progenitor cells to differentiate along the direction of osteogenesis. RhBMP-2 is an osteoinductive protein that has been used to promote spinal fusion and can induce fusion when used as an implant with a suitable carrier, such as an absorbable sponge (Boden et al., 2000; Burkus et al., 2017). BMP acts as a signal to stimulate BMSCs to migrate to the implantation site and induces marrow stromal cells (MSCs) to proliferate and differentiate into osteoblasts (Grgurevic et al., Bae et al., 2016). Previous studies on BMP-2 engraftment have shown that it has many effects, including angiogenesis, cell recruitment, and even promotion of mesenchymal and hematopoietic progenitor cell proliferation. However, new research suggests that BMP behaves differently at different doses (Burkus et al., 2002; Meisel et al., 2008). By adding RhBMP-2 to the fusion cage, Pan et al. found that, although BMP accelerated the speed of interbody fusion, it also produced a significant inflammatory response. In clinical applications, BMP may lead to local adverse structural changes and eclectic bone biomechanical changes, osteolysis, and trabecular thinning (Pan et al., 2017). Sethi et al. found that the use of RhBMP-2 can increase the fusion rate, but the incidence of pre-vertebral soft tissue swelling also increases (Sethi et al., 2011). The most common adverse reactions include accidental swelling and inflammation as well as inflammatory cyst formation. Although there is no difference in clinical effect between interbody fusion and autograft placement, BMP can also lead to endplate absorption, sinking, increased incidence of cage migration, osteolysis, and ectopic bone formation (Sethi et al., 2011; Michielsen et al., 2013; Burkus et al., 2017). Use of RhBMP-2 is strictly limited in clinical applications. It is relatively effective in achieving bone fusion, and the fusion rate is similar to that of patients receiving

autografts. The incidence of complications was similar between autografts and BMP, but the rates of radiculitis and serum tumor were slightly higher in the BMP group (Khan et al., 2018). There was no significant difference in fusion rate between low and high doses (low: 33.3% and high: 46.7%), but a high dose could easily lead to an increased incidence of osteolysis, adverse events, and swelling (Okada et al., 2020). During a postoperative evaluation of 17 patients with lumbar degenerative disease after implantation of a PEEK interbody cage with 212 mg of a RhBMP collagen gel sponge, Meisel et al. found that there was no significant difference between the fusion rate and autologous bone graft, but there was temporary bone resorption around the fusion cage at 3 months without subsidence, pain, or complications (Meisel et al., 2008). In a sheep interbody fusion model, Bae et al. used an absorbable sponge with different doses of RhBMP-2 in a customized PEEK interbody fusion cage and found that the osteoclast activity and the corresponding peri-implant bone resorption rate were dosedependent and reached a peak 4 weeks after operation (Bae et al., 2016). Grgurevic et al. induced complete spinal fusion with recombinant human RhBMP-6 based on an autologous blood coagulant (ABC) in sheep model. The best dose was 100 µg of RhBMP-6/mLABC (Grgurevic et al.) (Figure 9). An absorbable collagen sponge soaked in recombinant BMP-2 and recombinant BMP-7 combined with bovine collagen have been approved as substitutes for biological bone grafts to close gaps and repair delayed and non-union fractures. Among them, the incidence of complications in different positions is varied. Anterior cervical approach always causes the most serious side effects including anterior cervical edema, osteolysis, graft sedimentation and wound infection. Followed by posterior cervical approach which can cause posterior neck pain. The side effects in anterior and posterior lumbar approaches are relatively mild. Moreover, the tumorigenic effect of BMP should also be noted (Joseph and Rampersaud, 2007). In summary, BMP can significantly promote spinal fusion in the short term, but its side effects and dose sensitivity limit its wide application. Currently, BMP-2 has been proved by Food and Drug Administration (FDA) and used as an INFUSE Bone Graft (Medtronic Sofamor Danek) clinically. The recommended dosage varied between studies, and we draw a comprehensive conclusion among references that 1.5 mg/cm3 is deemed as an ideal effective concentration of BMP-2 (Sethi et al., 2011). Moreover, the following researches should continuously focus on the appropriate BMP dosage weighing the biosafety and effectiveness.

4.2.3.2 Combination of IGF and TGF- β

When the combination of IGF and TGF- β is used for spinal fusion, it can achieve mutual promotion. By adding IGF and TGF- β to HA-coated cages in a goat cervical fusion model, Gu *et al.* found the cage with IGF and TGF- β had significantly higher tensile strengths in extension and scoliosis compared to the other groups, indicating that they achieved better spinal fusion (Gu et al., 2013). Kim *et al.* used a fusion cage with IGF combined with TGF- β in a cervical fusion sheep model and found through quantitative CT that the fusion cage group with IGF combined with TGF- β showed significantly higher values for bone mineral



density, bone mineral content, and bony callus volume at 12 weeks after implantation. This shows that these growth factors can improve the formation of the intervertebral bone matrix, but their long-term effects need to be further explored (Kim et al., 2015). By adding BMP and IGF/TGF- β into a Ti cage in a sheep cervical fusion model, Kandziora *et al.* found that the early fusion effects of BMP and IGF/TGF- β were similar. The callus in the BMP group and IGF/TGF- β group had higher bone mineral density and biomechanical stability during extension, rotation, and bending (Kandziora *et al.*, 2002). This fully illustrates the strong potential of the combined use of IGF/

 $\text{TGF-}\beta$ in spinal fusion and provides a strong basis for future clinical use.

4.2.4 Allogeneic Material

Allogeneic cancellous bone, tricalcium phosphate, decalcified bone matrix (DBM), mineralized collagen, biphasic calcium phosphate, and other alternative materials have been successfully applied to the additives of interbody fusion cage, and good results have been obtained (Cho et al., 2005; Dai and Jiang 2008; Liao et al., 2008; Topuz et al., 2009; Scholz et al., 2010). Among them, CS is a kind of bone-substitute material. CS



particles have been successfully used as implantable fillers in the treatment of periodontal bone defects and long bone defects (Thomas and Puleo 2010; Baranes and Kurtzman 2019). DBM has been proven to have the potential for bone conduction and bone induction because of the proteins and various growth factors present in the extracellular matrix. DBM alone can induce good bone bridging in lumbar interbody fusion, but the effect is still not as good as that of autogenous bone graft, so it is recommended that DBM be combined with other materials. Xie et al. found that, although the curative effect and fusion rate of a PEEK interbody fusion cage filled with CS/DBM and autogenous bone filler were similar, CS/DBM instead of autogenous bone had the advantages of less intraoperative blood loss and fewer complications at the donor site, so it is a good substitute for autotransplantation (Xie et al., 2015). Biomineralized collagen has a good osteogenic ability, and the osteogenic ability of adjacent bone surfaces has a profound clinical application value, showing a potential application prospect. In clinical cases, Barlocher et al. found that, compared to an autologous bone graft and threaded Ti cage, polymethyl methacrylate (PMMA) appears to be a good alternative to cage-assisted fusion. Although many complications caused by autologous transplantation are avoided, the inability to achieve bony fusion still limits its use (Bärlocher et al., 2002). Kong et al. created a new biomaterial mineralized collagen-PMMA bone cement by injecting mineralized collagen into PMMA. It was found that PMMA

bone cement showed a good osteogenic ability in a sheep model, but this finding needs to be verified in further clinical transformational experiments. Minimally invasive injectable lumbar interbody fusion with mineralized collagen–PMMA bone cement has far-reaching clinical value and shows potential application prospects (Kong et al., 2020). However, not all allogeneic fillers are conducive to fusion. Cho *et al.* found that the fusion effect of a fusion cage containing biphasic calcium phosphate ceramic was lower than that of autogenous iliac bone graft and delayed the fusion period (Cho et al., 2005).

5 INTERBODY FUSION CAGE PROSPECTS

At present, there are many kinds of cages used in clinical and basic experiments that need to be chosen according to their respective advantages and needs. In this review, we list the advantages of different cage materials, internal implants, design methods, and post-processing to inform our use and selection.

In orthopedics clinics, the design of microporous fusion cages is particularly suitable for bone ingrowth and osseointegration, and the research on microporous structures is relatively mature. There are many studies on bone ingrowth with different porosities and pore sizes. Generally, the porosity is 65-80%, and its structure and mechanical properties are similar to those of trabecular bone. A pore size of $50-500 \,\mu\text{m}$ is

conducive to the adhesion, proliferation, and differentiation of bone cells. With the innovation of additive manufacturing technology, production now offers unprecedented means of structuring and customization, and the complex anatomy of the spine is very suitable for patient-specific equipment. Improving implant designs through the 3D-printing process may improve osseointegration and reduce the sinking rate. Combined with the current, relatively popular approach of finite element analysis, the preliminary design can be established to analyze whether the elastic modulus under different porosities and pore sizes is similar to that of human bone so that better spinal fusion can be obtained. In addition, topoisomerism technology can render the contact area between the implant and bone larger, which will make bone ingrowth easier. At present, there is no interbody cage with a fixation device available in the implant itself, which is also a direction that can be explored. Combined with these technologies mentioned above, a relatively suitable interbody cage can be obtained before implantation, and then basic and clinical experiments are performed to verify the performance.

Nowadays, the 2 most commonly used materials of interbody fusion cages are Ti alloy and PEEK. While Ti alloy materials have good biocompatibility, their excessive strength is likely to cause fusion. Conversely, the strength of PEEK is similar to that of human bone, and it is often used because of the easy detection of transmitted rays, but its biological inertness limits its further use. However, by modifying the PEEK material or adding other compounds to print the cage, its biocompatibility has been improved. At present, the CS/PEEK cage has been proven to promote interbody fusion, but mixtures with other ceramic-like materials have not appeared, which provides a direction for future research. Tantalum, Mg, and their alloys and absorbable materials have relatively few applications, and their respective deficiencies limit their usefulness. In the future, a new Mg alloy will not fail due to rapid absorption and local collapse; instead, it will achieve a balance between the new bone and the Mg alloy scaffold to perfectly support the pressure between the vertebral bodies before eventually being perfectly replaced by the new bone. Other absorbable materials, such as PLA, can be modified to achieve a degree of compressive strength that can support intervertebral pressure, rendering this issue no longer a problem. In addition, copper alloys, cobalt alloys, and memory metals with antibacterial properties will provide new options for vertebral body fusion under different physiological conditions.

Transplantation of autologous bone and allogeneic bone is the most common method for interbody fusion, offering biocompatibility and no obverse adverse reactions. In addition to the above 2 materials, there are other internal plants and drugs that are conducive to osseointegration. Materials such as SIM, BMP, biomineralized collagen, and PRP all show good bone ingrowth induction function, but there are no precise data to guide their dosage, and further exploration and experiments are necessary. In the future, through research, there will be guidelines developed to standardize their use and dosage. In addition, it is known that, in the process of bone formation, different growth factors are required in different stages. In the future, different drug-release systems will be constructed, and different doses of drugs will be released at corresponding times to regulate intervertebral bone formation.

To sum up, interbody fusion is a process of bony fusion. The selection of materials and additives for general interbody cages serves to enhance their osteogenic properties, but, when designing an interbody cage, the developer must consider that the device needs to withstand the pressure from upper and lower vertebrae; otherwise, the collapse of the implant will lead to fusion failure.

6 CONCLUSION

Good selection and use of the interbody cage are essential in the prognosis of clinical spinal interbody fusion. However, there is still a shortage of systematic discussion of its usage to date. This review focused on the design, materials, and post-processing technologies of the interbody cage. 3D printing is commonly used in the design work, and the design of porous structures is conducive to the ingrowth of new bone, which is more effective in improving the fusion rate. Other special types of interbody cages have been designed to reduce the difficulty of surgery or to facilitate fixation. Ti alloy and PEEK are the more commonly used materials for interbody cages. Other materials are less frequently used; among them, Mg alloy is an absorbable material, and its rapid absorption limits its use. In addition, post-processing technology is also important, including coating technology and internal additives. Ti and HA coatings are more commonly used, and studies so far have found that coating technology can improve spinal fusion to a certain extent. Internal additives, including SIM, BMP, and PRP, can promote bone formation through their own metabolism; however, there are few related studies at present, which provides a direction for future research.

AUTHOR CONTRIBUTIONS

HZ: Writing-original draft, Funding acquisition. ZW: Supervision. YW: Conceptualization. ZL: Supervision. BC: Conceptualization. SL: Conceptualization. WL: Conceptualization. JJ: Conceptualization. MW: Investigation, Methodology.

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REFERENCES

- Andrade, A. L. L. D., Sardeli, A. V., Garcia, T. A., Livani, B., and Belangero, W. D. (2021). PRP Does Not Improve the Objective Outcomes of Anterior Cruciate Ligament Reconstruction: a Systematic Review and Meta-Analysis. *Knee Surg. Sports Traumatol. Arthrosc.* 29 (9), 3049–3058. doi:10.1007/s00167-020-06348-z
- Arts, M., Torensma, B., and Wolfs, J. (2020). Porous Titanium Cervical Interbody Fusion Device in the Treatment of Degenerative Cervical Radiculopathy; 1-year Results of a Prospective Controlled Trial. *Spine J.* 20 (7), 1065–1072. doi:10. 1016/j.spinee.2020.03.008
- Assem, Y., Mobbs, R. J., Pelletier, M. H., Phan, K., and Walsh, W. R. (2017). Radiological and Clinical Outcomes of Novel Ti/PEEK Combined Spinal Fusion Cages: a Systematic Review and Preclinical Evaluation. *Eur. Spine J.* 26 (3), 593–605. doi:10.1007/s00586-015-4353-8
- Bae, H. W., Patel, V. V., Sardar, Z. M., Badura, J. M., Pradhan, B. B., Seim, H. B., et al. (2016). Transient Local Bone Remodeling Effects of rhBMP-2 in an Ovine Interbody Spine Fusion Model. *J. Bone Jt. Surg.* 98 (24), 2061–2070. doi:10. 2106/jbjs.16.00345
- Bai, H. T., Cui, Y. T., Wang, C. Y., Wang, Z. H., Luo, W. B., Liu, Y. Z., et al. (2020a). 3D Printed Porous Biomimetic Composition Sustained Release Zoledronate to Promote Osteointegration of Osteoporotic Defects. *Mater. Des.* 189, 108513. doi:10.1016/j.matdes.2020.108513
- Bai, H., Zhao, Y., Wang, C., Wang, Z., Wang, J., Liu, H., et al. (2020b). Enhanced Osseointegration of Three-Dimensional Supramolecular Bioactive Interface through Osteoporotic Microenvironment Regulation. *Theranostics* 10 (11), 4779–4794. doi:10.7150/thno.43736
- Baranes, D., and Kurtzman, G. M. (2019). Biphasic Calcium Sulfate as an Alternative Grafting Material in Various Dental Applications. J. Oral Implantol. 45 (3), 247–255. doi:10.1563/aaid-joi-D-18-00306
- Bärlocher, C. B., Barth, A., Krauss, J. K., Binggeli, R., and Seiler, R. W. (2002). Comparative Evaluation of Microdiscectomy Only, Autograft Fusion, Polymethylmethacrylate Interposition, and Threaded Titanium Cage Fusion for Treatment of Single-Level Cervical Disc Disease: a Prospective Randomized Study in 125 Patients. *Neurosurg. Focus* 12 (1), E4. doi:10.3171/ foc.2002.12.1.5
- Basgul, C., Yu, T., Macdonald, D. W., Siskey, R., and Kurtz, M. S. M. (2019). Does Annealing Improve the Interlayer Adhesion and Structural Integrity of FFF 3D Printed PEEK Lumbar Spinal Cages? J. Mech. Behav. Biomed. Mater 102, 103455. doi:10.1016/j.jmbbm.2019.103455
- Bassous, N. J., Jones, C. L., and Webster, T. J. (2019). 3-D Printed Ti-6Al-4V Scaffolds for Supporting Osteoblast and Restricting Bacterial Functions without Using Drugs: Predictive Equations and Experiments. *Acta Biomater*. 96, 662–673. doi:10.1016/j.actbio.2019.06.055
- Boden, S. D., Zdeblick, T. A., Sandhu, H. S., and Heim, S. E. (2000). The Use of rhBMP-2 in Interbody Fusion Cages. Spine 25 (3), 376–381. doi:10.1097/ 00007632-200002010-00020
- Burkus, J. K., Dryer, R. F., Arnold, P. M., and Foley, K. T. (2017). Clinical and Radiographic Outcomes in Patients Undergoing Single-Level Anterior Cervical Arthrodesis. *Clin. Spine Surg.* 30 (9), E1321–E1332. doi:10.1097/bsd. 0000000000000409
- Burkus, J. K., Transfeldt, E. E., Kitchel, S. H., Watkins, R. G., and Balderston, R. A. (2002). Clinical and Radiographic Outcomes of Anterior Lumbar Interbody Fusion Using Recombinant Human Bone Morphogenetic Protein-2. *Spine* 27 (21), 2396–2408. doi:10.1097/00007632-200211010-00015
- Burnard, J. L., Parr, W. C. H., Choy, W. J., Walsh, W. R., and Mobbs, R. J. (2020). 3D-printed Spine Surgery Implants: a Systematic Review of the Efficacy and Clinical Safety Profile of Patient-specific and Off-The-Shelf Devices. *Eur. Spine* J. 29 (6), 1248–1260. doi:10.1007/s00586-019-06236-2
- Caffrey, J. P., Alonso, E., Masuda, K., Hunt, J. P., Carmody, C. N., Ganey, T. M., et al. (2018). Strains in Trussed Spine Interbody Fusion Implants Are Modulated by Load and Design. J. Mech. Behav. Biomed. Mater. 80, 203–208. doi:10.1016/j.jmbbm.2018.02.004
- Caplan, I. F., Sinha, S., Osiemo, B., Mcclintock, S. D., Schuster, J. M., Smith, H., et al. (2020). The Utility of Cervical Spine Bracing as a Postoperative Adjunct to Multilevel Anterior Cervical Spine Surgery. *Int. J. Spine Surg.* 14 (2), 151–157. doi:10.14444/7022

- Carreon, L. Y., Glassman, S. D., Anekstein, Y., and Puno, R. M. (2005). Platelet Gel (AGF) Fails to Increase Fusion Rates in Instrumented Posterolateral Fusions. *Spine* 30 (9), E243–E246. doi:10.1097/01.brs.0000160846.85397.44
- Chang, C.-W., Chung, Y.-H., Chang, C.-J., Chen, Y.-N., Li, C.-T., Chang, C.-H., et al. (2020). Computational Comparison of Bone Cement and Poly Aryl-Ether-Ether-Ketone Spacer in Single-Segment Posterior Lumbar Interbody Fusion: a Pilot Study. *Phys. Eng. Sci. Med.* 43 (1), 163–173. doi:10.1007/s13246-019-00832-8
- Chen, C., Hao, Y., Bai, X., Ni, J. J., Chung, S. M., Liu, F., et al. (2019). 3D Printed Porous Ti6Al4V Cage: Effects of Additive Angle on Surface Properties and Biocompatibility; Bone Ingrowth in Beagle Tibia Model. *Mater Des.* 2019, 175. doi:10.1016/j.matdes.2019.107824
- Cheng, B. C., Jaffee, S., Averick, S., Swink, I., Horvath, S., and Zhukauskas, R. (2020). A Comparative Study of Three Biomaterials in an Ovine Bone Defect Model. *Spine J.* 20 (3), 457–464. doi:10.1016/j.spinee.2019.10.003
- Cho, D.-Y., Lee, W.-Y., Sheu, P.-C., and Chen, C.-C. (2005). Cage Containing a Biphasic Calcium Phosphate Ceramic (Triosite) for the Treatment of Cervical Spondylosis. Surg. Neurol. 63 (6), 497–503. doi:10.1016/j.surneu.2004.10.016
- Choi, A. H., Karacan, I., and Ben-Nissan, B. (2020). Surface Modifications of Titanium Alloy Using Nanobioceramic-Based Coatings to Improve Osseointegration: a Review. *Mater. Technol.* 35 (11-12), 742–751. doi:10. 1080/10667857.2018.1490848
- Chong, E., Mobbs, R. J., Pelletier, M. H., and Walsh, W. R. (2016). Titanium/ Polyetheretherketone Cages for Cervical Arthrodesis with Degenerative and Traumatic Pathologies: Early Clinical Outcomes and Fusion Rates. Orthop. Surg. 8 (1), 19–26. doi:10.1111/os.12221
- Chu, L., Li, R., Liao, Z., Yang, Y., Dai, J., Zhang, K., et al. (2019). Highly Effective Bone Fusion Induced by the Interbody Cage Made of Calcium Silicate/PEEK in a Goat Model. Acs Biomaterials Sci. Eng. 5 (5), 2409–2416. doi:10.1021/ acsbiomaterials.8b01193
- Dai, L.-Y., and Jiang, L.-S. (2008). Anterior Cervical Fusion with Interbody Cage Containing β -tricalcium Phosphate Augmented with Plate Fixation: a Prospective Randomized Study with 2-year Follow-Up. *Eur. Spine J.* 17 (5), 698–705. doi:10.1007/s00586-008-0643-8
- Demirel, M., and Kaya, A. I. (2020). Effect of Strontium-Containing Compounds on Bone Grafts. J. Mater. ence 55 (3). doi:10.1007/s10853-020-04451-7
- Duan, Y., Yang, Y., Wang, Y., Liu, H., Hong, Y., Gong, Q., et al. (2016). Comparison of Anterior Cervical Discectomy and Fusion with the Zero-Profile Device versus Plate and Cage in Treating Cervical Degenerative Disc Disease: A Meta-Analysis. J. Clin. Neurosci. 33, 11–18. doi:10.1016/j.jocn.2016.01.046
- Dxa, B., Jz, B., Cz, A., Db, C., Hy, B., Lm, B., et al. (2020). The Role of Calcium Phosphate Surface Structure in Osteogenesis and the Mechanisms Involved. *Acta Biomater.* 106, 22–33.
- Easley, J., Puttlitz, C. M., Seim, H., Ramo, N., Abjornson, C., Cammisa, F. P., et al. (2018). Biomechanical and Histologic Assessment of a Novel Screw Retention Technology in an Ovine Lumbar Fusion Model. *Spine J.* 18 (12), 2302–2315. doi:10.1016/j.spinee.2018.07.021
- Egan, P. F., Bauer, I., Shea, K., and Ferguson, S. J. (2018). Mechanics of 3D Printed Lattices for Biomedical Devices. *J. Mech. Des.* 141 (3).
- Fan, W., and Guo, L. X. (2020). The Effect of Non-fusion Dynamic Stabilization on Biomechanical Responses of the Implanted Lumbar Spine during Whole-Body Vibration. *Comput. Methods Programs Biomed.* 192, 105441. doi:10.1016/j. cmpb.2020.105441
- Farrokhi, M. R., Nikoo, Z., Gholami, M., and Hosseini, K. (2017). Comparison between Acrylic Cage and Polyetheretherketone (PEEK) Cage in Single-Level Anterior Cervical Discectomy and Fusion. *Clin. Spine Surg.* 30 (1), 38–46. doi:10.1097/bsd.00000000000251
- Feng, Y., Chen, L., Gu, Y., Zhang, Z.-M., Yang, H.-L., and Tang, T.-S. (2014). Influence of the Posterior Lumbar Interbody Fusion on the Sagittal Spino-Pelvic Parameters in Isthmic L5-S1 Spondylolisthesis. J. Spinal Disord. Tech. 27 (1), E20–E25. doi:10.1097/bsd.0b013e31828af6f0
- Folman, Y., Lee, S.-H., Silvera, J. R., and Gepstein, R. (2003). Posterior Lumbar Interbody Fusion for Degenerative Disc Disease Using a Minimally Invasive B-Twin Expandable Spinal Spacer. J. Spinal Disord. Tech. 16 (5), 455–460. doi:10.1097/00024720-200310000-00004
- Fomekong, E., Dufrane, D., Berg, B. V., André, W., Aouassar, N., Veriter, S., et al. (2017). Application of a Three-Dimensional Graft of Autologous Osteodifferentiated Adipose Stem Cells in Patients Undergoing Minimally

Invasive Transforaminal Lumbar Interbody Fusion: Clinical Proof of Concept. Acta Neurochir. 159 (3), 527–536. doi:10.1007/s00701-016-3051-6

- Girod, P.-P., Hartmann, S., Kavakebi, P., Obernauer, J., Verius, M., and Thomé, C.
 (2017). Asymmetric Pedicle Subtractionosteotomy (aPSO) Guided by a 3D-Printed Model to Correct a Combined Fixed Sagittal and Coronal Imbalance. *Neurosurg. Rev.* 40 (4), 689–693. doi:10.1007/s10143-017-0882-4
- Gkt, A., Mgo, B., Ml, B., Rgt, B., and Pmlb, C. (2020). Measuring the Performance of Patient-specific Solutions for Minimally Invasive Transforaminal Lumbar Interbody Fusion Surgery. J. Clin. Neurosci. 71, 43–50.
- Grgurevic, L., Erjavec, I., Gupta, M., Pecin, M., Bordukalo-Niksic, T., Stokovic, N., et al. (2020). Autologous Blood Coagulum Containing rhBMP6 Induces New Bone Formation to Promote Anterior Lumbar Interbody Fusion (ALIF) and Posterolateral Lumbar Fusion (PLF) of Spine in Sheep. *Bone* 138, 115448. doi:10.1016/j.bone.2020.115448
- Gu, Y., Zhang, F., Lineaweaver, W. C., Zhang, J., Qi, L. J., Wang, J., et al. (2013). In Vivo Study of Hydroxyapatite-Coated Hat Type Cervical Intervertebral Fusion Cage Combined with IGF-I and TGF-B1 in the Goat Model. Clin. Spine Surg. 29 (5), E267–E275. doi:10.1097/BSD.0b013e3182781d52
- Gunzburg, R., Colloca, C. J., Jones, C. F., Hall, D. J., Callary, J. S., Hegazy, M. A., et al. (2019). Does Nanoscale Porous Titanium Coating Increase Lumbar Spinal Stiffness of an Interbody Fusion Cage? an *In Vivo* Biomechanical Analysis in an Ovine Model. *Clin. Biomech. (Bristol, Avon)* 67, 187–196. doi:10.1016/j. clinbiomech.2019.04.024
- Guo, X., Xu, H., Zhang, F., and Lu, F. (2020). Bioabsorbable High-Purity Magnesium Interbody Cage: Degradation, Interbody Fusion, and Biocompatibility from a Goat Cervical Spine Model. Ann. Transl. Med. 8 (17), 1054. doi:10.21037/atm-20-225
- Hacker, R. J., Cauthen, J. C., Gilbert, T. J., and Griffith, S. L. (2000). A Prospective Randomized Multicenter Clinical Evaluation of an Anterior Cervical Fusion Cage. Spine 25 (20), 2646–2655. doi:10.1097/00007632-200010150-00017
- Hah, R. J., Alluri, R., and Anderson, P. A. (2020). Biomechanical Analysis of an Anterior Cervical Discectomy and Fusion Pseudarthrosis Model Revised with Machined Interfacet Allograft Spacers. *Glob. Spine J.* 10 (8), 973–981. doi:10. 1177/2192568219884265
- Hauerberg, J., Kosteljanetz, M., Bøge-Rasmussen, T., Dons, K., Springborg, P. J. B., and Wagner, A. (2008). Anterior Cervical Discectomy with or without Fusion with Ray Titanium Cage. Spine 33 (5), 458–464. doi:10.1097/brs. 0b013e3181657dac
- Hojo, Y., Kotani, Y., Ito, M., Abumi, K., Kadosawa, T., Shikinami, Y., et al. (2005).
 A Biomechanical and Histological Evaluation of a Bioresorbable Lumbar Interbody Fusion Cage. *Biomaterials* 26 (15), 2643–2651. doi:10.1016/j. biomaterials.2004.07.020
- Høy, K., Bünger, C., Niederman, B., Helmig, P., Hansen, E. S., Li, H., et al. (2013). Transforaminal Lumbar Interbody Fusion (TLIF) versus Posterolateral Instrumented Fusion (PLF) in Degenerative Lumbar Disorders: a Randomized Clinical Trial with 2-year Follow-Up. *Eur. Spine J.* 22 (9), 2022–2029. doi:10.1007/s00586-013-2760-2
- Høy, K., Grycel, B., Andersen, T., and Bünger, C. (2019). Does Transforaminal Lumbar Interbody Fusion Produce Leg Pain?-Results from a RCT. J. Orthop. Surg. Hong. Kong) 27 (3), 2309499019869469. doi:10.1177/2309499019869469
- Hua, L., Lei, T., Qian, H., Zhang, Y., and Lei, Y. P. (2020). 3D-printed Porous Tantalum: Recent Application in Various Drug Delivery Systems to Repair Hard Tissue Defects. *Expert Opin. Drug Deliv.* 18 (122), 625–634. doi:10.1080/ 17425247.2021.1860015
- Huang, H., Jiang, C., Feng, Z., and Jiang, X. (2014). Comparing the Process of Creeping Substitution between Allograft Bone and Local Bone Grafting in Lumbar Interbody Fusion. *Eur. Spine J.* 23 (10), 2068–2074. doi:10.1007/ s00586-014-3388-6
- Iunes, E. A., Barletta, E. A., Barba Belsuzarri, T. A., Onishi, F. J., Cavalheiro, S., and Joaquim, A. F. (2020). Correlation between Different Interbody Grafts and Pseudarthrosis after Anterior Cervical Discectomy and Fusion Compared with Control Group: Systematic Review. *World Neurosurg.* 134, 272–279. doi:10. 1016/j.wneu.2019.10.100
- Iv, J. P. M., Beutler, W., Peppelman, W., Harris, R., Slotkin, E., and Gillette, J. (2010). Clinical Outcomes with Porous Tantalum in Lumbar Interbody Fusion. *Spine J.* 10 (9-supp-S), S147–S148.
- Jain, A., Marrache, M., Harris, A., Puvanesarajah, V., Neuman, B. J., Buser, Z., et al. (2020). Structural Allograft versus PEEK Implants in Anterior Cervical

Discectomy and Fusion: A Systematic Review. Glob. Spine J. 10 (6), 775–783. doi:10.1177/2192568219883256

- Jain, S., Eltorai, A. E. M., Ruttiman, R., and Daniels, A. H. (2016). Advances in Spinal Interbody Cages. Orthop. Surg. 8 (3), 278–284. doi:10.1111/os.12264
- Jin, H., Ji, Y., Cui, Y., Xu, L., and Wang, H. J. (2021). Simvastatin-Incorporated Drug Delivery Systems for Bone Regeneration. ACS Biomater. Sci. Eng. 7 (6), 2177–2191. doi:10.1021/acsbiomaterials.1c00462
- Jiya, T., Smit, T., Deddens, J., and Mullender, M. (2009). Posterior Lumbar Interbody Fusion Using Nonresorbable Poly-Ether-Ether-Ketone versus Resorb Able Poly-L-Lactide-Co-D,l-Lactide Fusion Devices A Prospective, Randomized Study to Assess Fusion and Clinical Outcome. Spine 34 (3), 233–237. doi:10.1097/BRS.0b013e318194ed00
- Jiya, T. U., Smit, T., van Royen, B. J., and Mullender, M. (2011). Posterior Lumbar Interbody Fusion Using Non Resorbable Poly-Ether-Ether-Ketone versus Resorbable Poly-L-Lactide-Co-D,l-Lactide Fusion Devices. Clinical Outcome at a Minimum of 2-year Follow-Up. *Eur. Spine J.* 20 (4), 618–622. doi:10.1007/ s00586-010-1568-6
- Joffe, M. R., Parr, W. C. H., Tan, C., Walsh, W. R., and Brunel, L. (2019). Development of a Customized Interbody Fusion Device for Treatment of Canine Disc-Associated Cervical Spondylomyelopathy. *Vet. Comp. Orthop. Traumatol.* 32 (1), 79–86. doi:10.1055/s-0038-1676075
- Joseph, V., and Rampersaud, Y. R. (2007). Heterotopic Bone Formation with the Use of rhBMP2 in Posterior Minimal Access Interbody Fusion. Spine 32, 2885–2890. doi:10.1097/brs.0b013e31815b7596
- Kandziora, F., Pflugmacher, R., Scholz, M., Knispel, C., Hiller, T., Schollmeier, G., et al. (2002). Comparison of BMP-2 and Combined IGF-I/TGF-ss1 Application in a Sheep Cervical Spine Fusion Model. *Eur. Spine J.* 11 (5), 482–493. doi:10. 1007/s00586-001-0384-4
- Kandziora, F., Pflugmacher, R., Scholz, M., Schafer, J., Schollmeier, G., Schmidmaier, G., et al. (2003). Dose-dependent Effects of Combined IGF-I and TGF-?1 Application in a Sheep Cervical Spine Fusion Model. *Eur. spine J.* 12 (5), 464–473. doi:10.1007/s00586-002-0483-x
- Kerolus, M., Turel, M. K., Tan, L., and Deutsch, H. (2016). Stand-Alone Anterior Lumbar Interbody Fusion: Indications, Techniques, Surgical Outcomes and Complications: Expert Review of Medical Devices: Vol 0, No Ja. Expert Rev. Med. Devices 13 (12), 1127–1136. doi:10.1080/17434440. 2016.1254039
- Kersten, R. F., van Gaalen, S. M., Arts, M. P., Roes, K. C., de Gast, A., Corbin, T. P., et al. (2014). The SNAP Trial: a Double Blind Multi-Center Randomized Controlled Trial of a Silicon Nitride versus a PEEK Cage in Transforaminal Lumbar Interbody Fusion in Patients with Symptomatic Degenerative Lumbar Disc Disorders: Study Protocol. *BMC Musculoskelet. Disord.* 15, 57. doi:10. 1186/1471-2474-15-57
- Khan, T. R., Pearce, K. R., McAnany, S. J., Peters, C. M., Gupta, M. C., and Zebala, L. P. (2018). Comparison of Transforaminal Lumbar Interbody Fusion Outcomes in Patients Receiving rhBMP-2 versus Autograft. *Spine J.* 18 (3), 439–446. doi:10.1016/j.spinee.2017.08.230
- Kim, C. W., Doerr, T. M., Luna, I. Y., Joshua, G., Shen, S. R., Fu, X., et al. (2016). Minimally Invasive Transforaminal Lumbar Interbody Fusion Using Expandable Technology: A Clinical and Radiographic Analysis of 50 Patients. World Neurosurg. 90, 228–235. doi:10.1016/j.wneu.2016.02.075
- Kim, J., Lin, B., Kim, S., Choi, B., Evseenko, D., and Lee, M. (2015). TGF-β1 Conjugated Chitosan Collagen Hydrogels Induce Chondrogenic Differentiation of Human Synovium-Derived Stem Cells. J. Biol. Eng. 9 (1), 1. doi:10.1186/1754-1611-9-1
- Knutsen, A. R., Borkowski, S. L., Ebramzadeh, E., Flanagan, C. L., Hollister, S. J., and Sangiorgio, S. N. (2015). Static and Dynamic Fatigue Behavior of Topology Designed and Conventional 3D Printed Bioresorbable PCL Cervical Interbody Fusion Devices. J. Mech. Behav. Biomed. Mater. 49, 332–342. doi:10.1016/j. jmbbm.2015.05.015
- Kong, J., Ma, J., Wu, Z., Wang, H., Peng, X., Wang, S., et al. (2020). Minimally Invasive Injectable Lumbar Interbody Fusion with Mineralized Collagen-Modified PMMA Bone Cement: A New Animal Model. J. Appl. Biomater. Funct. Mater 18, 2280800020903630. doi:10.1177/2280800020903630
- Krafft, P. R., Osburn, B., Vivas, A. C., Rao, G., and Alikhani, P. (2019). Novel Titanium Cages for Minimally Invasive Lateral Lumbar Interbody Fusion: First Assessment of Subsidence. *Spine Surg. Relat. Res.* 4 (2), 171–177. doi:10.22603/ ssrr.2019-0089

- Krafft, P. R., Osburn, B., Vivas, A. C., Rao, G., and Alikhani, P. (2020). Novel Titanium Cages for Minimally Invasive Lateral Lumbar Interbody Fusion: First Assessment of Subsidence. *Spine Surg. Relat. Res.* 4 (2), 171–177. doi:10.22603/ ssrr.2019-0089
- Krayenbühl, N., Schneider, C., Landolt, H., and Fandino, J. (2008). Use of an Empty, Plasmapore-Covered Titanium Cage for Interbody Fusion after Anterior Cervical Microdiscectomy. J. Clin. Neurosci. 15 (1), 11–17. doi:10. 1016/j.jocn.2006.12.011
- Lee, J. H., Kong, C.-B., Yang, J. J., Shim, H.-J., Koo, K.-H., Kim, J., et al. (2016). Comparison of Fusion Rate and Clinical Results between CaO-SiO2-P205-B203 Bioactive Glass Ceramics Spacer with Titanium Cages in Posterior Lumbar Interbody Fusion. *Spine J.* 16 (11), 1367–1376. doi:10.1016/j.spinee. 2016.07.531
- Leng, Y., Ren, G., Cui, Y., Peng, C., Wang, J., Wu, D., et al. (2020). Platelet-rich Plasma-Enhanced Osseointegration of Decellularized Bone Matrix in Critical-Size Radial Defects in Rabbits. *Ann. Transl. Med.* 8 (5), 198. doi:10.21037/atm. 2020.01.53
- Liang, X., Li, F., Gong, X., Li, J., Li, S. Q., Liu, Z., et al. (2020). In Vivo evaluation of Porous Nanohydroxyapatite/polyamide 66 Struts in a Goat Cervical Fusion Model. Sci. Rep. 10 (1), 10495. doi:10.1038/s41598-020-65341-1
- Liao, J. C., Niu, C. C., Chen, W. J., and Chen, L. H. (2008). Polyetheretherketone (PEEK) Cage Filled with Cancellous Allograft in Anterior Cervical Discectomy and Fusion. *Int. Orthop.* 32 (5), 643–648. doi:10.1007/s00264-007-0378-x
- Lim, K.-M., Park, T.-H., Lee, S.-J., and Park, S.-J. (2019). Design and Biomechanical Verification of Additive Manufactured Composite Spinal Cage Composed of Porous Titanium Cover and PEEK Body. *Appl. Sci.* 9 (20), 4258. doi:10.3390/ app9204258
- Liu, H., Li, W., Liu, C., Tan, J., Wang, H., Hai, B., et al. (2016). Incorporating Simvastatin/poloxamer 407 Hydrogel into 3D-Printed Porous Ti6Al4V Scaffolds for the Promotion of Angiogenesis, Osseointegration and Bone Ingrowth. *Biofabrication* 8 (4), 045012. doi:10.1088/1758-5090/8/4/045012
- Liu, X., Han, F., Zhao, P., Lin, C., Wen, X., and Ye, X. (2017). Layer-by-layer Self-Assembled Multilayers on PEEK Implants Improve Osseointegration in an Osteoporosis Rabbit Model. *Nanomedicine Nanotechnol. Biol. Med.* 13 (4), 1423–1433. doi:10.1016/j.nano.2017.01.011
- Lowery, G. L., Kulkarni, S., and Pennisi, A. E. (1999). Use of Autologous Growth Factors in Lumbar Spinal Fusion. *Bone* 25 (2), 47s–50s. doi:10.1016/s8756-3282(99)00132-5
- Lu, C., Qian, C., Li-Bo, J., Xiao-Fan, Y., Chong, B., Hui-Ren, W., et al. (2017). Bioabsorbable Self-Retaining PLA/nano-sized β-TCP Cervical Spine Interbody Fusion Cage in Goat Models: An *In Vivo* Study. *Int. J. Nanomedicine* 12, 7197–7205.
- Lu, M., Xu, S., Lei, Z.-X., Lu, D., Cao, W., Huttula, M., et al. (2019). Application of a Novel Porous Tantalum Implant in Rabbit Anterior Lumbar Spine Fusion Model. *Chin. Med. J.* 132 (1), 51–62. doi:10.1097/cm9.000000000000000
- MacBarb, R. F., Lindsey, D. P., Bahney, C. S., Woods, S. A., Wolfe, M. L., and Yerby, S. A. (2017). Fortifying the Bone-Implant Interface Part 1: An *In Vitro* Evaluation of 3D-Printed and TPS Porous Surfaces. *Int. J. Spine Surg.* 11, 15. doi:10.14444/4015
- Madan, S., and Boeree, N. R. (2003). Outcome of the Graf Ligamentoplasty Procedure Compared with Anterior Lumbar Interbody Fusion with the Hartshill Horseshoe Cage. *Eur. Spine J.* 12 (4), 361–368. doi:10.1007/s00586-002-0488-5
- Marx, D., Rahimnejad Yazdi, A., Papini, M., and Towler, M. (2020). A Review of the Latest Insights into the Mechanism of Action of Strontium in Bone. *Bone Rep.* 12, 100273. doi:10.1016/j.bonr.2020.100273
- May, R. D., Frauchiger, D. A., Albers, C. E., Tekari, A., Benneker, L. M., Klenke, F. M., et al. (2019). Application of Cytokines of the Bone Morphogenetic Protein (BMP) Family in Spinal Fusion Effects on the Bone, Intervertebral Disc and Mesenchymal Stromal Cells. Cscr 14 (8), 618–643. doi:10.2174/1574888x14666190628103528
- McGilvray, K. C., Easley, J., Seim, H. B., Regan, D., Berven, S. H., Hsu, W. K., et al. (2018). Bony Ingrowth Potential of 3D-Printed Porous Titanium Alloy: a Direct Comparison of Interbody Cage Materials in an *In Vivo* Ovine Lumbar Fusion Model. *Spine J.* 18 (7), 1250–1260. doi:10.1016/j.spinee.2018.02.018
- McGilvray, K. C., Waldorff, E. I., Easley, J., Seim, H. B., Zhang, N., Linovitz, R. J., et al. (2017). Evaluation of a Polyetheretherketone (PEEK) Titanium Composite Interbody Spacer in an Ovine Lumbar Interbody Fusion Model: Biomechanical,

Microcomputed Tomographic, and Histologic Analyses. Spine J. 17 (12), 1907–1916. doi:10.1016/j.spinee.2017.06.034

- Meisel, H. J., Schnöring, M., Hohaus, C., Minkus, Y., Beier, A., Ganey, T., et al. (2008). Posterior Lumbar Interbody Fusion Using rhBMP-2. *Eur. Spine J.* 17 (12), 1735–1744. doi:10.1007/s00586-008-0799-2
- Michielsen, J., Sys, J., Rigaux, A., and Bertrand, C. (2013). The Effect of Recombinant Human Bone Morphogenetic Protein-2 in Single-Level Posterior Lumbar Interbody Arthrodesis. J. Bone Jt. Surgery-American 95 (10), 873–880. doi:10.2106/jbjs.1.00137
- Mobbs, R. J., Parr, W. C. H., Choy, W. J., McEvoy, A., Walsh, W. R., and Phan, K. (2019). Anterior Lumbar Interbody Fusion Using a Personalized Approach: Is Custom the Future of Implants for Anterior Lumbar Interbody Fusion Surgery? *World Neurosurg*. 124, 452–458. doi:10.1016/j.wneu.2018.12.144
- Mokawem, M., Katzouraki, G., Harman, C. L., and Lee, R. (2019). Lumbar Interbody Fusion Rates with 3D-Printed Lamellar Titanium Cages Using a Silicate-Substituted Calcium Phosphate Bone Graft. J. Clin. Neurosci. 68, 134–139. doi:10.1016/j.jocn.2019.07.011
- Niu, J., Sun, Z., Chang, H., and Zhou, L. (2019). Review on 3D Printing of Biomedical Titanium Alloy. *Rare Metal Materials and Engineering* 48 (5), 1697–1706.
- Ohanisian, L., and Dorsi, M. J. (2019). A Novel 3D Printed Titanium Implant for Anterior Cervical Discectomy and Fusion. *Cureus* 11 (1), e3952. doi:10.7759/ cureus.3952
- Okada, R., Kaito, T., Ishiguro, H., Kushioka, J., Otsuru, S., Kanayama, S., et al. (2020). Assessment of Effects of rhBMP-2 on Interbody Fusion with a Novel Rat Model. *Spine J.* 20 (5), 821–829. doi:10.1016/j.spinee.2019.12.014
- Pan, H. C., Lee, S., Ting, K., Shen, J., Wang, C., Nguyen, A., et al. (2017). Cyst-Like Osteolytic Formations in Recombinant Human Bone Morphogenetic Protein-2 (rhBMP-2) Augmented Sheep Spinal Fusion. *Am. J. Pathology* 187 (7), 1485–1495. doi:10.1016/j.ajpath.2017.03.010
- Park, H.-W., Lee, J.-K., Moon, S.-J., Seo, S.-K., Lee, J.-H., and Kim, S.-H. (2009). The Efficacy of the Synthetic Interbody Cage and Grafton for Anterior Cervical Fusion. Spine 34 (17), E591–E595. doi:10.1097/brs.0b013e3181ab8b9a
- Park, P. J., and Lehman, R. A. (2020). Optimizing the Spinal Interbody Implant: Current Advances in Material Modification and Surface Treatment Technologies. *Curr. Rev. Musculoskelet. Med.* 13 (6), 688–695. doi:10.1007/ s12178-020-09673-5
- Phan, K., and Mobbs, R. J. (2016). Evolution of Design of Interbody Cages for Anterior Lumbar Interbody Fusion. Orthop. Surg. 8 (3), 270–277. doi:10.1111/ os.12259
- Pisecky, L., Luger, M., Klasan, A., Gotterbarm, T., Klotz, M. C., and Hochgatterer, R. (2021). Bioabsorbable Implants in Forefoot Surgery: a Review of Materials, Possibilities and Disadvantages. *Efort Open Rev.* 6 (12), 1132–1139. doi:10. 1302/2058-5241.6.200157
- Qiao, S. C., Sheng, Q. B., Li, Z. H., Wu, D. L., Zhu, Y., Lai, H. C., et al. (2020). 3Dprinted Ti6Al4V Scaffolds Coated with Freeze-Dried Platelet-Rich Plasma as Bioactive Interface for Enhancing Osseointegration in Osteoporosis. *Mater. Des.* 194. doi:10.1016/j.matdes.2020.108825
- Rao, P. J., Pelletier, M. H., Walsh, W. R., and Mobbs, R. J. (2014). Spine Interbody Implants: Material Selection and Modification, Functionalization and Bioactivation of Surfaces to Improve Osseointegration. *Orthop. Surg.* 6 (2), 81–89. doi:10.1111/os.12098
- Ren, C., Song, Y., Xue, Y., Yang, X., and Zhou, C. (2017). Evaluation of Bioabsorbable Multiamino Acid Copolymer/Nanohydroxyapatite/Calcium Sulfate Cage in a Goat Spine Model. *World Neurosurg.* 103, 341–347. doi:10.1016/j.wneu.2017.04.005
- Rieger, B., Jiang, H., Reinshagen, C., Molcanyi, M., Zivcak, J., Grönemeyer, D., et al. (2017). Effects of Preoperative Simulation on Minimally Invasive Hybrid Lumbar Interbody Fusion. *World Neurosurg*. 106, 578–588. doi:10.1016/j. wneu.2017.06.184
- Rodrigues, T. A., Freire, A. O., Bonfim, B. F., Cartágenes, M. S. S. S., and Garcia, J. B. S. (2018). Strontium Ranelate as a Possible Disease-Modifying Osteoarthritis Drug: a Systematic Review. *Braz J. Med. Biol. Res.* 51 (8), e7440. doi:10.1590/ 1414-431x20187440
- Rodríguez-Vázquez, M., and Ramos-Zúñiga, R. (2020). Chitosan-Hydroxyapatite Scaffold for Tissue Engineering in Experimental Lumbar Laminectomy and Posterolateral Spinal Fusion in Wistar Rats. Asian Spine J. 14 (2), 139–147. doi:10.31616/asj.2019.0091

- Sasso, R. C., Kitchel, S. H., and Dawson, E. G. (2004). A Prospective, Randomized Controlled Clinical Trial of Anterior Lumbar Interbody Fusion Using a Titanium Cylindrical Threaded Fusion Device. *Spine* 29 (2), 113–121. doi:10.1097/01.brs.0000107007.31714.77
- Scardueli, C. R., Bizelli-Silveira, C., Marcantonio, R. A. C., Marcantonio, E., Stavropoulos, A., and Spin-Neto, R. (2018). Systemic Administration of Strontium Ranelate to Enhance the Osseointegration of Implants: Systematic Review of Animal Studies. *Int. J. Implant Dent.* 4 (1), 21. doi:10.1186/s40729-018-0132-8
- Schnitzer, M., Hudák, R., Sedlačko, P., Rajtúková, V., Findrik Balogová, A., Živčák, J., et al. (2020). A Comparison of Experimental Compressive Axial Loading Testing with a Numerical Simulation of Topologically Optimized Cervical Implants Made by Selective Laser Melting. J. Biotechnol. 322, 33–42. doi:10. 1016/j.jbiotec.2020.07.010
- Scholz, M., Schleicher, P., Eindorf, T., Friedersdorff, F., Gelinsky, M., König, U., et al. (2010). Cages Augmented with Mineralized Collagen and Platelet-Rich Plasma as an Osteoconductive/inductive Combination for Interbody Fusion. *Spine* 35 (7), 740–746. doi:10.1097/brs.0b013e3181bdc6cc
- Seaman, S., Kerezoudis, P., Bydon, M., Torner, J. C., and Hitchon, P. W. (2017). Titanium vs. Polyetheretherketone (PEEK) Interbody Fusion: Meta-Analysis and Review of the Literature. *J. Clin. Neurosci.* 44, 23–29. doi:10.1016/j.jocn. 2017.06.062
- Sekerci, Z., Uğur, A., Ergün, R., and Sanli, M. (2013). Early Changes in the Cervical Foraminal Area after Anterior Interbody Fusion with Polyetheretherketone (PEEK) Cage Containing Synthetic Bone Particulate: a Prospective Study of 20 Cases. *Neurol. Res.* 28 (5), 568–571. doi:10.1179/ 016164106X110382
- Serra, T., Capelli, C., Toumpaniari, R., Orriss, I. R., Leong, J. J., Dalgarno, K., et al. (2016). Design and Fabrication of 3D-Printed Anatomically Shaped Lumbar Cage for Intervertebral Disc (IVD) Degeneration Treatment. *Biofabrication* 8 (3), 035001. doi:10.1088/1758-5090/8/3/035001
- Sethi, A., Craig, J., Bartol, S., Chen, W., Jacobson, M., Coe, C., et al. (2011). Radiographic and CT Evaluation of Recombinant Human Bone Morphogenetic Protein-2-Assisted Spinal Interbody Fusion. Am. J. Roentgenol. 197 (1), W128–W133. doi:10.2214/ajr.10.5484
- Shiga, Y., Orita, S., Kubota, G., Kamoda, H., Yamashita, M., Matsuura, Y., et al. (2016). Freeze-Dried Platelet-Rich Plasma Accelerates Bone Union with Adequate Rigidity in Posterolateral Lumbar Fusion Surgery Model in Rats. *Sci. Rep.* 6, 36715. doi:10.1038/srep36715
- Siu, T. L., Rogers, J. M., Lin, K., Thompson, R., and Owbridge, M. (2018). Custom-Made Titanium 3-Dimensional Printed Interbody Cages for Treatment of Osteoporotic Fracture-Related Spinal Deformity. *World Neurosurg.* 111, 1–5. doi:10.1016/j.wneu.2017.11.160
- Smoljanovic, T., Bojanic, I., and Vlahovic, Z. (2010). Safety of Posterior Interbody Fusions of the Lumbar Spine Using rhBMP-2. J. Spinal Disord. Tech. 23 (1), 78. doi:10.1097/bsd.0b013e3181c1474d
- Sochacki, K. R., Safran, M. R., Abrams, G. D., Donahue, J., Chu, C., and Sherman, S. L. (2020). Platelet-Rich Plasma Augmentation for Isolated Arthroscopic Meniscal Repairs Leads to Significantly Lower Failure Rates: A Systematic Review of Comparative Studies. Orthop. J. Sports Med. 8 (11), 2325967120964534. doi:10.1177/2325967120964534
- Song, C., Guo, Z., Ma, Q., Chen, Z., Liu, Z., Jia, H., et al. (2003). Simvastatin Induces Osteoblastic Differentiation and Inhibits Adipocytic Differentiation in Mouse Bone Marrow Stromal Cells. *Biochem. Biophysical Res. Commun.* 308 (3), 458–462. doi:10.1016/s0006-291x(03)01408-6
- Staiger, M. P., Pietak, A. M., Huadmai, J., and Dias, G. (2006). Magnesium and its Alloys as Orthopedic Biomaterials: A Review. *Biomaterials* 27 (9), 1728–1734. doi:10.1016/j.biomaterials.2005.10.003
- Tan, J., Yang, N., Fu, X., Cui, Y., and Song, C. (2015). Single-Dose Local Simvastatin Injection Improves Implant Fixation via Increased Angiogenesis and Bone Formation in an Ovariectomized Rat Model. *Med. Sci. Monit.* 21, 1428–1439. doi:10.12659/MSM.892247
- Tan, K. H., Chua, C. K., Leong, K. F., Naing, M. W., and Cheah, C. M. (2005).
 Fabrication and Characterization of Three-Dimensional Poly(ether-Ether-Ketone)/-Hydroxyapatite Biocomposite Scaffolds Using Laser Sintering. Proc. Inst. Mech. Eng. H. 219 (3), 183–194. doi:10.1243/ 095441105x9345

- Thomas, M. V., and Puleo, D. A. (2010). Calcium Sulfate: Properties and Clinical Applications. J. Biomed. Mater Res. B Appl. Biomater. 88 (2), 597–610. doi:10. 1002/jbm.b.31269
- Topuz, K., Çolak, A., Kaya, S., Şimşek, H., Kutlay, M., Demircan, M. N., et al. (2009). Two-level Contiguous Cervical Disc Disease Treated with Peek Cages Packed with Demineralized Bone Matrix: Results of 3-year Follow-Up. *Eur. Spine J.* 18 (2), 238–243. doi:10.1007/s00586-008-0869-5
- Tsai, T. T., Tai, C. L., Ho, N. Y., Lai, P. L., Fu, T. S., Niu, C. C., et al. (2017). Effects of Strontium Ranelate on Spinal Interbody Fusion Surgery in an Osteoporotic Rat Model. *Plos One* 12 (1), e0167296. doi:10.1371/journal.pone.0167296
- Ulum, M. F., Caesarendra, W., Alavi, R., and Hermawan, H. (2019). *In-Vivo* Corrosion Characterization and Assessment of Absorbable Metal Implants. *Coatings* 9 (5), 282. doi:10.3390/coatings9050282
- Vanek, P., Bradac, O., Delacy, P., Saur, K., Belsan, T., and Benes, V. (2012). Comparison of 3 Fusion Techniques in the Treatment of the Degenerative Cervical Spine Disease. Is Stand-Alone Autograft Really the "Gold Standard?". *Spine* 37 (19), 1645–1651. doi:10.1097/brs.0b013e31825413fe
- Veillette, C. J. H., Mehdian, H., Schemitsch, E. H., and McKee, M. D. (2006). Survivorship Analysis and Radiographic Outcome Following Tantalum Rod Insertion for Osteonecrosis of the Femoral Head. J. Bone Jt. Surgery-American Volume 88, 48–55. doi:10.2106/00004623-200611001-00008
- Verma, R., Virk, S., and Qureshi, S. (2020). Interbody Fusions in the Lumbar Spine: A Review. HSS Jrnl 16 (2), 162–167. doi:10.1007/s11420-019-09737-4
- Virk, S. S., Coble, D., Bertone, A. L., Hussein, H. H., and Khan, S. N. (2017). Experimental Design and Surgical Approach to Create a Spinal Fusion Model in a New Zealand White Rabbit (*Oryctolagus cuniculus*). *J. Invest. Surg.* 30 (4), 226–234. doi:10.1080/08941939.2016.1235748
- Walsh, W. R., Pelletier, M. H., Bertollo, N., Christou, C., and Tan, C. (2016). Does PEEK/HA Enhance Bone Formation Compared with PEEK in a Sheep Cervical Fusion Model? *Clin. Orthop. Relat. Res.* 474 (11), 2364–2372. doi:10.1007/ s11999-016-4994-x
- Walsh, W. R., Pelletier, M. H., Christou, C., He, J., Vizesi, F., and Boden, S. D. (2018). The *In Vivo* Response to a Novel Ti Coating Compared with Polyether Ether Ketone: Evaluation of the Periphery and Inner Surfaces of an Implant. *Spine J.* 18 (7), 1231–1240. doi:10.1016/j.spinee.2018.02.017
- Walsh, W. R., Pelletier, M. H., Wang, T., Lovric, V., Morberg, P., and Mobbs, R. J. (2019). Does Implantation Site Influence Bone Ingrowth into 3D-Printed Porous Implants? *Spine J.* 19 (11), 1885–1898. doi:10.1016/j.spinee.2019.06.020
- Wang, J. L., Xu, J. K., Hopkins, C., Chow, D. H., and Qin, L. (2020). Biodegradable Magnesium-Based Implants in Orthopedics-A General Review and Perspectives. Adv. Sci. (Weinh) 7 (8), 1902443. doi:10.1002/advs.201902443
- Wang, Y., Wei, R., Subedi, D., Jiang, H., Yan, J., and Li, J. (2020). Tantalum Fusion Device in Anterior Cervical Discectomy and Fusion for Treatment of Cervical Degeneration Disease. *Clin. Spine Surg.* 33 (3), 111–119. doi:10.1097/bsd. 000000000000875
- Warburton, A., Girdler, S. J., Mikhail, C. M., Ahn, A., and Cho, S. K. (2019). Biomaterials in Spinal Implants: A Review. *Neurospine* 17 (1), 101–110. doi:10. 14245/ns.1938296.148
- Warren, J. M., Mazzoleni, A. P., and Hey, L. A. (2020). Development and Validation of a Computationally Efficient Finite Element Model of the Human Lumbar Spine: Application to Disc Degeneration. *Int. J. Spine Surg.* 14 (4), 502–510. doi:10.14444/7066
- Wessels, V., Mené, G. L., Fischerauer, S. F., Kraus, T., Weinberg, A. M., Uggowitzer,
 P. J., et al. (2012). *In Vivo* Performance and Structural Relaxation of Biodegradable Bone Implants Made from Mg Zn Ca Bulk Metallic Glasses. *Adv. Eng. Mater.* 14 (6), B357–B364. doi:10.1002/adem.201180200
- Wheeler, D. L., Fredericks, D. C., Dryer, R. F., and Bae, H. W. (2016). Allogeneic Mesenchymal Precursor Cells (MPCs) Combined with an Osteoconductive Scaffold to Promote Lumbar Interbody Spine Fusion in an Ovine Model. *Spine J.* 16 (3), 389–399. doi:10.1016/j.spinee.2015.08.019
- Xie, Y., Li, H., Yuan, J., Fu, L., Yang, J., and Zhang, P. (2015). A Prospective Randomized Comparison of PEEK Cage Containing Calcium Sulphate or Demineralized Bone Matrix with Autograft in Anterior Cervical Interbody Fusion. *Int. Orthop. (SICOT)* 39 (6), 1129–1136. doi:10.1007/s00264-014-2610-9
- Xing, H., Lee, H., Luo, L., and Kyriakides, T. R. (2020). Extracellular Matrix-Derived Biomaterials in Engineering Cell Function. *Biotechnol. Adv.* 42, 107421. doi:10.1016/j.biotechadv.2019.107421

- Yang, S., Yu, Y., Liu, X., Hou, Z. T., Xu, J., Wu, W., et al. (2019). Clinical and Radiological Results Comparison of Allograft and Polyetheretherketone Cage for One to Two-Level Anterior Cervical Discectomy and Fusion: A CONSORT-Compliant Article. *Med. Baltim.* 98 (45), e17935. doi:10.1097/MD. 000000000017935
- Zardiackas, L. D., Parsell, D. E., Dillon, L. D., Mitchell, D. W., Nunnery, L. A., and Poggie, R. (2001). Structure, Metallurgy, and Mechanical Properties of a Porous Tantalum Foam. *J. Biomed. Mat. Res.* 58 (2), 180–187. doi:10.1002/1097-4636(2001)58:2<180::aid-jbm1005>3.0.co;2-5
- Zhang, F., Xu, H., Wang, H., Geng, F., Ma, X., Shao, M., et al. (2018). Quantitative Analysis of Near-Implant Magnesium Accumulation for a Si-Containing Coated AZ31 Cage from a Goat Cervical Spine Fusion Model. BMC Musculoskelet. Disord. 19, 105. doi:10.1186/s12891-018-2027-5
- Zhang, W., Sun, C., Zhu, J., Zhang, W., Leng, H., and Song, C. (2020a). 3D Printed Porous Titanium Cages Filled with Simvastatin Hydrogel Promotes Bone Ingrowth and Spinal Fusion in Rhesus Macaques. *Biomater. Sci.* 8 (15), 4147–4156. doi:10.1039/d0bm00361a
- Zhang, W., Sun, C., Zhu, J., Zhang, W., Leng, H., and Song, C. (2020b). 3D Printed Porous Titanium Cages Filled with Simvastatin Hydrogel Promotes Bone Ingrowth and Spinal Fusion in Rhesus Macaques. *Biomaterials Sci.* 8 (15), 4147–4156. doi:10.1039/d0bm00361a
- Zhang, Y., Xu, J., Ruan, Y. C., Yu, M. K., O'Laughlin, M., Wise, H., et al. (2016). Implant-derived Magnesium Induces Local Neuronal Production of CGRP to Improve Bone-Fracture Healing in Rats. *Nat. Med.* 22 (10), 1160–1169. doi:10. 1038/nm.4162
- Zhang, Z., Li, H., Fogel, G. R., Xiang, D., Liao, Z., and Liu, W. (2018). Finite Element Model Predicts the Biomechanical Performance of Transforaminal Lumbar Interbody Fusion with Various Porous Additive Manufactured Cages. *Comput. Biol. Med.* 95, 167–174. doi:10.1016/j.compbiomed.2018. 02.016

- Zhao, R., Yang, R., Cooper, P. R., Khurshid, Z., Shavandi, A., and Ratnayake, J. (2021). Bone Grafts and Substitutes in Dentistry: A Review of Current Trends and Developments. *Molecules* 26 (10), 3007. doi:10.3390/molecules26103007
- Zhao, S., Wen, F., He, F., Liu, L., and Yang, G. (2014). In Vitro and In Vivo Evaluation of the Osteogenic Ability of Implant Surfaces with a Local Delivery of Simvastatin. Int. J. Oral Maxillofac. Implants 29 (1), 211–220. doi:10.11607/ jomi.3147
- Zippelius, T., Strube, P., Suleymanov, F., Putzier, M., and Hölzl, A. (2018). Sicherheit und Effektivität eines in Electron-Beam-Melting-Technik hergestellten Titan-Mesh-Cages zur intersomatischen lumbalen Fusion; Safety and efficacy of an electron beam melting technique-manufactured titanium mesh cage for lumbar interbody fusion. Der Orthopäde.

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