

# Preventive Effects of *Lactobacillus* Mixture on Experimental *E. coli* Urinary Tract Infection in Infant Rats

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**Purpose:** Urinary tract infection (UTI) is an ascending infection of fecal uropathogens, urogenital lactobacilli are suggested to play a role in the prevention of UTI. This study was to investigate whether *lactobacillus* mixture (LM) could prevent the experimental infantile UTI. **Materials and Methods:** The LM were composed of three *lactobacillus* strains (*L. gasseri*, *L. rhamnosus*, and *L. reuteri*). Mother rats were grouped as *lactobacillus* (LB) group I (LB I, n=22), II (LB II, n=24) and control (n=20). LB I and LB II were fed with LM (1 mL/day) and control with phosphate-buffered saline (PBS) from late pregnancy through lactation. All newborn rats were breast-fed and their urine and stool were collected at the end of the 3rd week to compare *lactobacillus* colony. Then, infant rats from LB II were treated with intravesical instillation of LM. Infant rats from LB I and control were instilled with PBS. Twenty-four hours later, experimental UTI was introduced by intravesical instillation of standard *E. coli* strain. After 72 hours later, the infant rats were sacrificed for histologic examination. **Results:** Lactobacilli colonies in urine and stool were not statistically different among the three groups. The incidence of pyelonephritis in the LB II was 16.7% (4/24), LB I 72.7% (16/22) and control 75.0% (15/20) ( $p=0.015$ ). The incidence of cystitis was not significantly different among the three groups. **Conclusion:** The intravesically instilled LM significantly prevented experimental pyelonephritis in infant rats, however, LM administered orally to the pregnant and lactating mother rats did not.

**Key Words:** Lactobacillus, intravesical instillation, cystitis, pyelonephritis

## INTRODUCTION

Urinary tract infection (UTI) is the most common bacterial infection in infants and has been documented as an ascending infection of the patient's own fecal uropathogens, evidenced by genomic profiling study.<sup>1</sup> Therefore, the role of *lactobacillus*, the most dominant urogenital microflora, has been a focus in preventing UTI.<sup>2,3</sup>

Intraurethraly instilled *lactobacillus* strains were proven to prevent experimental UTI in adult animal models,<sup>4,6</sup> and clinical application of *lactobacillus* suppositories showed beneficial effects in reducing the recurrence rate of UTI in adult women.<sup>7-9</sup> However, the preventive effect of oral *lactobacillus* probiotics against

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UTI has not sufficiently been investigated in adults.<sup>10,11</sup>

In early infancy when the UTI incidence is extremely high, postnatal development of genitourinary *lactobacilli* is considered very important in preventing UTI. It was confirmed that some maternal *lactobacilli* are transferred to their infants during birth and through breast milk.<sup>12-14</sup> Probiotic *lactobacilli*, administered to lactating mother rats, were also proven to be transferred to their infant guts<sup>15,16</sup> and then to genitourinary tract.<sup>17</sup> However, a question of whether this mother to infant transmission prevents infantile UTI has not been clarified.

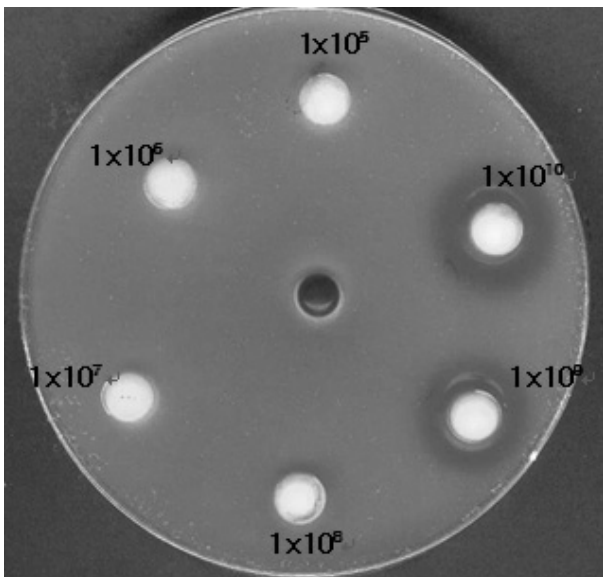
Therefore, we evaluated whether *lactobacilli* orally administered to pregnant and lactating mother rats could prevent experimental UTI of their infant rats, and compared the effects with those of intravesically instilled *lactobacilli* to infant rats.

## MATERIALS AND METHODS

### Materials

#### *Lactobacillus* mixture

For this study, we selected three *lactobacillus* strains (*L. gasseri*, *L. rhamnosus*, *L. reuteri*), which were isolated from the healthy infant feces and mixed. For optimal dosage, the *lactobacillus* mixture (LM) was incubated in the DeMan-Rogosa-Sharpe (MRS) agar medium to make six different dosages [ $10^6$ - $10^{11}$  colony forming unit (CFU)/mL], which



**Fig. 1.** Inhibitory zone (diameter, mm) at different dosage of *lactobacillus* mixture against standard *E. coli* (ATCC 25922). The optimal dosage (CFU/mL) for maximal inhibition was determined as  $1 \times 10^{10}$  CFU/mL.

were tested for the antimicrobial activities against standard *E. coli* strain (ATCC No. 25922, Seattle, WA, USA). We selected the dosage  $10^{10}$  CFU/mL, whose antimicrobial activity was maximal (Fig. 1).

#### *The experimental infant rats*

Forty-eight breast-fed Sprague Dawley infant rats (Seoul, Korea), whose mother rats were fed the LM (1 mL/day) via gavage tube from late pregnancy through lactation, were evenly allocated to the *lactobacillus* (LB) I (n=24) or II group (n=24). The control group included 20 infant rats, whose mother rats were given phosphate-buffer solution (PBS, 1 mL). All infant rats were breast-fed for 3 weeks, when the same LM (1 mL) was intravesically instilled to infant rats of the LB II group and PBS to those of the LB I and the control group. The study protocol was approved by the ethical committee of the hospital.

#### *Urine and stool culture for lactobacillus*

At age 3 weeks of infant rats, urines and stools were collected. Fecal specimens were placed to a 50 mL test tube together with sterile saline and shaken for 1 minute. For *lactobacillus* culture, urines and diluted fecal supernatants (200  $\mu$ L) were inoculated into *lactobacillus*-selective MRS agar (Oxoid, Basingstroke, UK) and incubated anaerobically at 37°C for 48 hours. *Lactobacillus* was confirmed by Gram-positive white, smooth bacillus, and colonies (CFU/mL) were counted.

#### *Intravesical instillation of the lactobacillus mixture*

After completion of stool and urine collection, the same LM (1 mL), given to the mother rats, was instilled into the bladder of infant rats of the LB II group using a 16-gauge silicone catheter after ketamine anesthesia. PBS (1 mL) was instilled into LB I and the control group.

#### *Induction of experimental E. coli UTI*

Twenty-four hours after intravesical instillation of the LM or PBS, 1 mL of standard *E. coli* strain ( $10^7$  CFU/mL) was instilled into the bladder of infant rats to induce experimental *E. coli* UTI through a 16-gauge silicone catheter after ketamine anesthesia.

#### *Histopathological examination*

Seventy-two hours after intravesical instillation of *E. coli*, the infant rats were sacrificed for histopathologic examination. Both kidneys and bladder were extracted and fixed for

24 hours in 10% buffered formalin solution. The kidney was embedded in paraffin, sectioned 3  $\mu\text{m}$  in thickness using a rotatory microtome, and Hematoxylin-Eosin staining and Masson-Trichrome staining were performed. According to the histopathological changes, cystitis and pyelonephritis were diagnosed and the incidence was compared. For semiquantitative evaluation, the severity of inflammation in cystitis and pyelonephritis was scored. The cystitis score ranged from 0 to 3 (0 point without inflammatory cells, 1 point with a few inflammatory cells in the submucosa, 2 points with more than 5 focal inflammatory cells in the submucosa, and 3 points with diffuse inflammatory cells in the submucosa). The pyelonephritis score ranged from 0 to 4 (0 point without inflammation, 1 point with occasional inflammation in the pelvic mucosa, 2 points with continuous inflammation along the pelvic mucosa, 3 points with focal inflammation from the pelvic mucosa to the renal medulla, and 4 points with diffuse inflammation to the renal medulla).

#### Statistical methods

For statistical analysis, SPSS version 11.5 (SPSS Inc., Chicago, IL, USA) was used. ANOVA test was performed for comparison of the number of *lactobacillus* or *E. coli*, histopathological findings, the incidence and the score of cystitis

and pyelonephritis between LB I, II and control groups. For the comparison of the number of *lactobacillus* and *E. coli* CFU/mL after experimental *E. coli* UTI, Wilcoxon signed rank test was used. *p* value less than 0.05 was determined to be statistically significant.

## RESULTS

#### *Lactobacillus* colonization status in stools and urines of infant rats

The numbers of *lactobacillus* CFUs in the stools of infant rats were  $3.5 \times 10^8 \pm 6.6 \times 10^7$  CFU/mL in the LB I group,  $4.1 \times 10^8 \pm 2.5 \times 10^8$  CFU/mL in the LB II group, and  $1.7 \times 10^7 \pm 7.3 \times 10^6$  CFU/mL in the control group ( $p > 0.05$ ). The numbers of *lactobacillus* CFUs in the urines of infant rats were  $2930 \pm 314.4$  CFU/mL in the LB I group,  $5892 \pm 370.5$  CFU/mL in the LB II group, and  $1658 \pm 361.1$  CFU/mL in the control group ( $p > 0.05$ ) (Fig. 2).

#### The incidence of experimental UTI

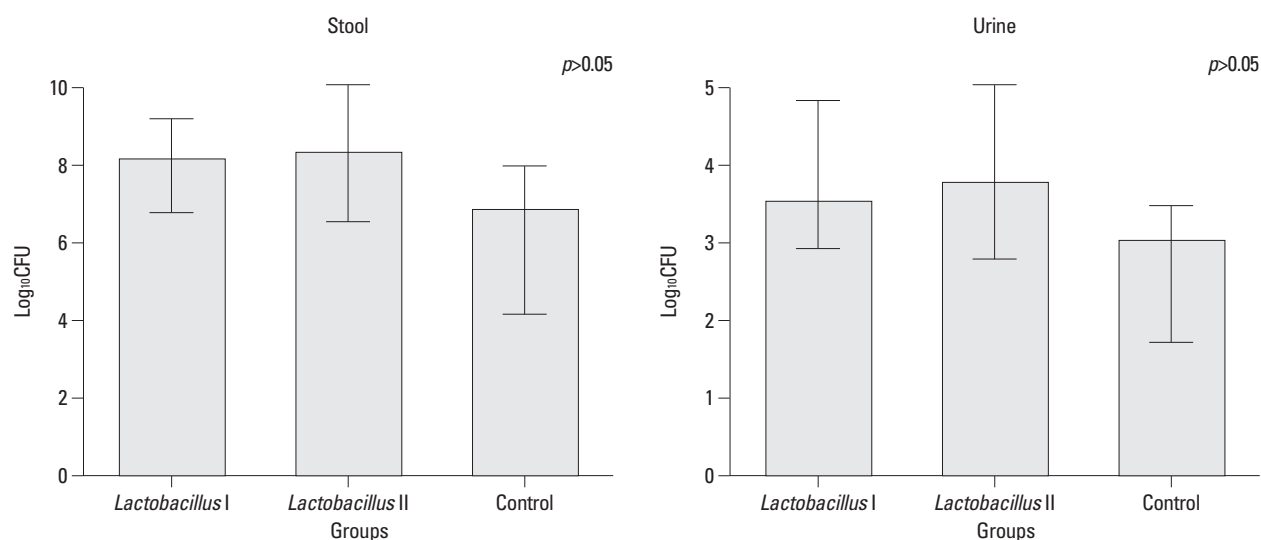
The incidence of cystitis was 95.5% (21/22) in the LB I group, 87.5% (21/24) in the LB II group, and 100% (20/20) in the control group ( $p > 0.05$ ). The incidence of pyelone-

**Table 1.** Incidence of Experimental *E. coli* Urinary Tract Infection in Infant Rats

UTI	<i>Lactobacillus</i> I (n=22)	<i>Lactobacillus</i> II (n=24)	Control (n=20)
Cystitis	21 (95.5%)	21 (87.5%)	20 (100%)
Pyelonephritis	16 (72.7%)	4 (16.7%)*	15 (75.0%)
Total	21 (95.5%)	21 (87.5%)	20 (100%)

UTI, urinary tract infection.

\* $p = 0.015$  vs. control.



**Fig. 2.** Stool and urine *lactobacillus* colonization in *lactobacillus* I, II and control group.

phritis was 16.7% (4/24) in the LB II group, which was significantly lower than 75.0% (15/20) of the control group ( $p=0.015$ ) (Table 1).

### The semiquantitative inflammatory score of cystitis and pyelonephritis

The cystitis score of the LB II group ( $1.25\pm 0.98$ ) was lower than those of the LB I group ( $1.5\pm 0.78$ ) and control group ( $1.8\pm 1.06$ ) ( $p>0.05$ ) (Fig. 3). The pyelonephritis score of the LB II group ( $0.33\pm 0.68$ ) was significantly lower than those of the LB I group ( $1.31\pm 1.21$ ) and control group ( $1.95\pm 1.72$ ) ( $p=0.006$ ) (Fig. 3).

## DISCUSSION

In preventing infantile UTI, postnatal development of genitourinary *lactobacilli* is considered very important. Indeed, *lactobacilli* in the maternal vagina are the first source of *lactobacilli* of newborn infants. While passing through the birth canal, maternal vaginal *lactobacilli* are transferred to the sterile neonate's gut for the first time.<sup>12</sup> *Lactobacilli* in breast milk are the second important source of infant's gut *lactobacilli*.<sup>13</sup> Approximately 6 days after birth, the number of CFU/mL of *lactobacillus* in the feces of breast-fed infants was 1000 times more than that of enterobacteriae, but 10 times less in the feces of bottle-fed infants.<sup>14</sup> Probiotic *lactobacilli*, supplied during pregnant and lactating period, were proved to colonize the infant's gut.<sup>15,16</sup> These vertically transmitted *lactobacilli* are transferred from gut to geni-

tourinary tract.<sup>17</sup>

Many earlier studies demonstrated that the number of urogenital *lactobacilli* is significantly decreased in infants with UTI<sup>18</sup> as well as in woman with urethritis and recurrent UTI.<sup>7-9,19-22</sup> Antimicrobial activities of *lactobacillus* strains against uropathogens have been studied in many *in vitro* tests.<sup>23-26</sup> *Lactobacillus* strains impeded the adherence of uropathogens by secreting biosurfactants,<sup>23</sup> compete with uropathogens in the binding site on vaginal epithelial cells,<sup>24</sup> and inhibit the growth of uropathogens by secreting hydrogen peroxide, lactate, bacteriocin, and other antimicrobial molecules.<sup>25</sup> They also enhance the local immunity of the intestinal mucosa, and improve the innate immunity and cell-mediated immunity by activating monocytes.<sup>26</sup>

The preventive effects of intravesically instilled *lactobacilli* against UTI have been proven in adult animal models with different strains and different dosages.<sup>4,6</sup> When *L. casei* GR1 ( $5\times 10^9$  CFU/mL), isolated from healthy adult women, was intravesically instilled to adult rats and was then swabbed twice weekly for 21 days onto the introitus before challenge with an uropathogen suspension (*E. coli*, *K. pneumoniae*, *P. aeruginosa*), experimental UTI was prevented in 84% of the animals.<sup>4</sup> *L. casei* shirota strain ( $1\times 10^8$  CFU/day), when administered intraurethrally to a mouse 24 hours prior to the induction of experimental *E. coli*, dramatically inhibited *E. coli* growth and inflammatory responses in the urinary tract.<sup>5</sup> Furthermore, intraurethral instillation of the indigenous *L. murinus* strain ( $1\times 10^8$  CFU/mL) also significantly prevented *Proteus mirabilis* ascending UTI in a mouse model.<sup>6</sup>

In the present study, intravesically instilled *lactobacilli* to infant rats showed the significant preventive effect against experimental pyelonephritis, whereas orally administered *lactobacilli* to pregnant and lactating mother rats did not prevent the infection of their infant rats. This might be due to insufficient increase of *lactobacillus* colonization in their stools and urines. Further studies are necessary to find ideal *lactobacillus* strains and an optimal oral dosage that are compatible with preventive effect of intravesically instilled *lactobacillus*.

□ Cystitis score    ■ Pyelonephritis score

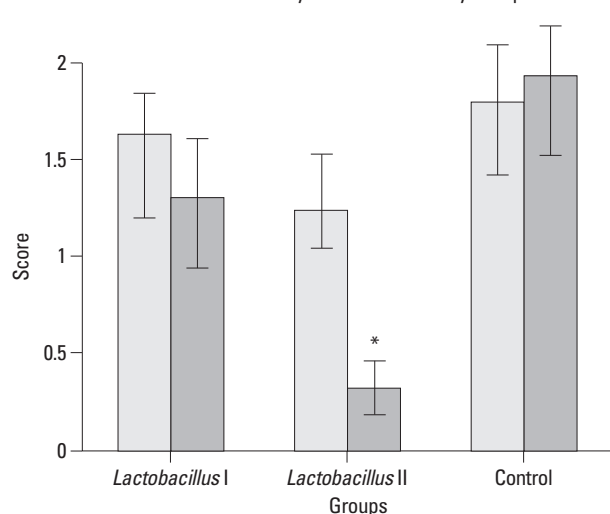


Fig. 3. Cystitis and pyelonephritis score in experimental *E. coli* urinary tract infection (cystitis score  $p>0.05$ , pyelonephritis score  $*p=0.006$  vs. control).

## REFERENCES

- Usein CR, Damian M, Tatu-Chițoiu D, Căpușă C, Făgăraș R, Mircescu G. Comparison of genomic profiles of Escherichia coli isolates from urinary tract infections. Roum Arch Microbiol Immunol 2003;62:137-54.
- Redondo-Lopez V, Cook RL, Sobel JD. Emerging role of lactoba-

- cilli in the control and maintenance of the vaginal bacterial microflora. *Rev Infect Dis* 1990;12:856-72.
3. Reid G, Bruce AW. Probiotics to prevent urinary tract infections: the rationale and evidence. *World J Urol* 2006;24:28-32.
  4. Reid G, Chan RC, Bruce AW, Costerton JW. Prevention of urinary tract infection in rats with an indigenous *Lactobacillus casei* strain. *Infect Immun* 1985;49:320-4.
  5. Asahara T, Nomoto K, Watanuki M, Yokokura T. Antimicrobial activity of intraurethrally administered probiotic *Lactobacillus casei* in a murine model of *Escherichia coli* urinary tract infection. *Antimicrob Agents Chemother* 2001;45:1751-60.
  6. Fraga M, Scavone P, Zunino P. Preventive and therapeutic administration of an indigenous *Lactobacillus* sp. strain against *Proteus mirabilis* ascending urinary tract infection in a mouse model. *Antonie Van Leeuwenhoek* 2005;88:25-34.
  7. Reid G, Bruce AW, Taylor M. Instillation of *Lactobacillus* and stimulation of indigenous organisms to prevent recurrence of urinary tract infection. *Microecol Ther* 1995;23:32-45.
  8. Uehara S, Monden K, Nomoto K, Seno Y, Kariyama R, Kumon H. A pilot study evaluating the safety and effectiveness of *Lactobacillus* vaginal suppositories in patients with recurrent urinary tract infection. *Int J Antimicrob Agents* 2006;28 Suppl 1:S30-4.
  9. Stapleton AE, Au-Yeung M, Hooton TM, Fredricks DN, Roberts PL, Czaja CA, et al. Randomized, placebo-controlled phase 2 trial of a *Lactobacillus crispatus* probiotic given intravaginally for prevention of recurrent urinary tract infection. *Clin Infect Dis* 2011; 52:1212-7.
  10. Lee SJ, Shim YH, Cho SJ, Lee JW. Probiotics prophylaxis in children with persistent primary vesicoureteral reflux. *Pediatr Nephrol* 2007;22:1315-20.
  11. Abad CL, Safdar N. The role of *Lactobacillus* probiotics in the treatment or prevention of urogenital infections--a systematic review. *J Chemother* 2009;21:243-52.
  12. Matsumiya Y, Kato N, Watanabe K, Kato H. Molecular epidemiological study of vertical transmission of vaginal *Lactobacillus* species from mothers to newborn infants in Japanese, by arbitrarily primed polymerase chain reaction. *J Infect Chemother* 2002;8:43-9.
  13. Martín R, Langa S, Reviriego C, Jiménez E, Marín ML, Xaus J, et al. Human milk is a source of lactic acid bacteria for the infant gut. *J Pediatr* 2003;143:754-8.
  14. Rinne M, Kalliomaki M, Arvilommi H, Salminen S, Isolauri E. Effect of probiotics and breastfeeding on the bifidobacterium and *Lactobacillus/enterococcus* microbiota and humoral immune responses. *J Pediatr* 2005;147:186-91.
  15. Hall MA, Cole CB, Smith SL, Fuller R, Rolles CJ. Factors influencing the presence of faecal *Lactobacilli* in early infancy. *Arch Dis Child* 1990;65:185-8.
  16. Sanz Y. Gut microbiota and probiotics in maternal and infant health. *Am J Clin Nutr* 2011;94(6 Suppl):2000S-5S.
  17. Morelli L, Zonenenschain D, Del Piano M, Cognein P. Utilization of the intestinal tract as a delivery system for urogenital probiotics. *J Clin Gastroenterol* 2004;38(6 Suppl):S107-10.
  18. Lee JW, Shim YH, Lee SJ. *Lactobacillus* colonization status in infants with urinary tract infection. *Pediatr Nephrol* 2009;24:135-9.
  19. Bruce AW, Chadwick P, Hassan A, VanCott GF. Recurrent urethritis in women. *Can Med Assoc J* 1973;108:973-6.
  20. Marrie TJ, Swantee CA, Hartlen M. Aerobic and anaerobic urethral flora of healthy females in various physiological age groups and of females with urinary tract infections. *J Clin Microbiol* 1980;11:654-9.
  21. Gupta K, Stapleton AE, Hooton TM, Roberts PL, Fennell CL, Stamm WE. Inverse association of H<sub>2</sub>O<sub>2</sub>-producing *Lactobacilli* and vaginal *Escherichia coli* colonization in women with recurrent urinary tract infections. *J Infect Dis* 1998;178:446-50.
  22. Kirjavainen PV, Pautler S, Baroja ML, Anukam K, Crowley K, Carter K, et al. Abnormal immunological profile and vaginal microbiota in women prone to urinary tract infections. *Clin Vaccine Immunol* 2009;16:29-36.
  23. Velraeds MM, van der Mei HC, Reid G, Busscher HJ. Inhibition of initial adhesion of uropathogenic *Enterococcus faecalis* by biosurfactants from *Lactobacillus* isolates. *Appl Environ Microbiol* 1996;62:1958-63.
  24. Zárate G, Nader-Macias ME. Influence of probiotic vaginal *Lactobacilli* on in vitro adhesion of urogenital pathogens to vaginal epithelial cells. *Lett Appl Microbiol* 2006;43:174-80.
  25. Rodríguez JM, Martínez MI, Horn N, Dodd HM. Heterologous production of bacteriocins by lactic acid bacteria. *Int J Food Microbiol* 2003;80:101-16.
  26. Matsuzaki T, Chin J. Modulating immune responses with probiotic bacteria. *Immunol Cell Biol* 2000;78:67-73.