

Lipid accumulation product and visceral adiposity index are associated with dietary patterns in adult Americans

Mohsen Mazidi, PhD^{a,b,*}, Hong-kai Gao, PhD^{a,b}, Andre Pascal Kengne, MD^c

Abstract

In the present study, we aimed to examine the association between lipid accumulation product (LAP) and visceral adiposity index (VAI) with dietary pattern (DP) in the US adults. Participants of the National Health and Nutrition Examination Survey (NHANES) with data available on dietary intake from 2005 to 2010 were included. DPs were derived by principal component analysis. We applied analysis of covariance and multivariable-adjusted linear regressions accounting for the masked variance and utilizing the proposed weighting methodology. The analytical sample comprised 18,318 participants (mean age = 45.8 years), of whom 48.3% (n = 8607) were men with no age difference by gender (P = .126). The first DP was representative of a diet rich in carbohydrate and sugar, total fat and saturated fatty acid (SFA), high-caloric dieatry pattern; the second DP was highly loaded with vitamins, minerals and fiber (nutrient-dense dietary pattern), and the third DP was mainly representative of high dietary polyunsaturated fatty acids (PUFAs) and monounsaturated fatty acids (MUFAs) (healthy fat DP). The adjusted (age, sex, race, physical activity, smoking, C-reactive protein) mean of LAP, VAI and glucose homeostasis indices increased across increasing quarters of the first DP score (all P < .001), while across increasing score of the second DP, the adjusted mean of LAP, VAI, glucose homeostasis indices decreased (all P < .001). Findings were similar in adjusted linear regressions models. Our findings support that affordable measurements, such as VAI and LAP, could be good alternative surrogate markers of visceral fat. They are also significantly related to DPs in same line as with glucose/insulin homeostasis and anthropometric indices.

Abbreviations: AMPM = agriculture automated multiple-pass method, ANCOVA = analysis of covariance, apVAT = anthropometrically predicted visceral adipose tissue, BMI = body mass index, DP = dietary pattern, HbA1c = glycated hemoglobin, HDL = high-density lipoprotein, HOMA-IR = homeostatic model assessment of insulin resistance, LAP = lipid accumulation product, NHANES = National Health and Nutrition Examination Survey, MUFA = monounsaturated fatty acids, PUFA = polyunsaturated fatty acids, SFA = saturated fatty acid, TG = triglyceride, VAI = visceral adiposity index, VAT = visceral adipose tissue, WC = circumferences of the waist.

Keywords: Glucose, insulin, lipid accumulation product, visceral adiposity index

Editor: Kei Nakajima.

Mohsen Mazidi and Hong-kai Gao contributed equally to this work.

MM was supported by a TWAS studentship of the Chinese Academy of Sciences.

For data collection and physical examination of the NHANES, informed consent was obtained from all adult participants, and the National Centre for Health Statistics Research Ethics Review Board approved the protocol.

All the data are from public access database.

The authors have no funding and conflicts of interest to disclose.

^a Key State Laboratory of Molecular Developmental Biology, Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, ^bDepartment of General Surgery, The General Hospital of Chinese People's Armed Police Forces, Beijing, China, ^cNon-Communicable Disease Research Unit, South African Medical Research Council and University of Cape Town, Cape Town, South Africa.

* Correspondence: Mohsen Mazidi, Key State Laboratory of Molecular Developmental Biology, Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, Beijing 100101, China (e-mail: moshen@genetics.ac.cn).

Copyright © 2018 the Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Medicine (2018) 97:19(e0322)

Received: 27 June 2017 / Received in final form: 8 December 2017 / Accepted: 14 March 2018

http://dx.doi.org/10.1097/MD.000000000010322

1. Introduction

Excess visceral adipose tissue (VAT) is one of the most deleterious fat depots in the body, with strong links with cardiovascular disease, and certain types of cancer.^[1,2] Lipid accumulation product (LAP) index, a recently developed biomarker of central fat accumulation, has been recommended as a precise indicator of the risk of insulin resistance, metabolic syndrome, type 2 diabetes, and cardiovascular disease.^[3–5] Higher LAP has been associated with abnormal glucose homeostasis and insulin resistance, as well as elevated alanine aminotransferase in healthy individuals.^[6] A Chinese study has shown that both LAP and visceral adiposity index (VAI) were effective markers for stratifying adults for obesity phenotypes.^[7] In addition, another study reported that LAP was a helpful indicator for the screening for metabolic syndrome.^[8]

The VAT seems to be affected by diet and lifestyle modifications.^[9,10] Furthermore, it has been suggested that VAT is mainly influenced by the non-caloric qualitative aspects of diet, although evidence on the association between macronutrient composition of diet and VAT, is still limited. A recent investigation indicated that consuming energy mainly as carbohydrate or fat for 3 months did not affect visceral fat and metabolic syndrome in a low-processed, lower-glycemic dietary context.^[11] There are contradictory findings regarding the

association between different dietary patterns (DPs), LAP, and VAI. Significant association between carbohydrate intake,^[12,13] dietary fatty acids^[14] including saturated fatty acids (SFAs), monounsaturated fatty acids (MUFAs), polyunsaturated fatty acids (PUFAs) with VAT has not been reported in all investigations.^[8] However, it is important to note that foods and nutrients are consumed in combination, and complex combinations of nutrients are likely to be interactive or to have a synergistic effect.^[15] The approach of evaluating single nutrients or foods might therefore be limited in terms of potential interactions and high inter-correlations between several food components, which might make it challenging to estimate the general, or independent impacts of different nutrients or foods, perhaps slight and thus untraceable impacts of a single nutrient may be concealed, and the concern of multiple comparison is also crucial in this area.^[16] Therefore, in an efforts to overcome these issues, the analysis of DPs has gained importance.^[15,17] A DP is a comprehensive variable that incorporates the intake of numerous nutrients or nutrient groups and that has a more impact on disease risk than does any single nutrient.^[16,17]

The mechanisms by which nutrient patterns affect the risk of chronic conditions are not fully understood and there is good evidence that it is a combination of nutrients, rather than an individual one, that will affect the risk. Therefore, a pattern of nutrients may provide more information about probable underlying mechanisms.^[18,19]

The aim of present study is to investigate the association between LAP, VAI with DPs, alongside markers of glucose/ insulin homeostasis (which are well-characterized correlates of VAT) in randomly selected nationally representative samples of the US adults.

2. Methods

2.1. Population

The National Health and Nutrition Examination Surveys (NHANESs) conducted between 2005 and 2010 were used for this study. NHANESs are repeated cross-sectional surveys conducted by the US National Center for Health Statistics, applying protocols and procedures described in details previously.^[20,21] NHANES uses a complex, multistage, and stratified sampling design to select a sample representative of the civilian and non-institutionalized resident population of the United States. Data on demographic information and interviews are collected using questionnaires administered during home visits, while anthropometrical, inflammation, and biochemistry data are collected by trained personnel using mobile examination units. Methods for biochemical analyses are described in the NHANES Laboratory/Medical Technologists Procedures Manual.^[20,22-24] NHANES is open access database and all the information on the data access and analysis can be found at https://www.cdc.gov/nchs/nhanes/index.htm.

Dietary intake was assessed via 24 hours recall obtained by a trained interviewer during the mobile examination center visit, with the use of a computer-assisted dietary interview system with standardized probes, that is, the United States Department of Agriculture Automated Multiple-Pass Method (AMPM).^[25,26] Briefly, the type and quantity of all foods and beverages consumed in a single 24-hour period before the dietary interview (from midnight to midnight) were collected with the use of AMPM. AMPM is designed to enhance complete and accurate data collection while reducing respondent burden.^[26,27]

A blood specimen was drawn from the participant's antecubital vein by a trained phlebotomist. Glycated hemoglobin (HbA1c) was measured using a Tosoh A1C 2.2 plus glycohemoglobin analyzer (San Francisco). Fasting plasma glucose was measured by a hexokinase method using a Roche/Hitachi 911 analyzer (New Jersey) and Roche Modular P chemistry analyzer (New Jersey). Insulin was measured using an enzymelinked immunosorbent assay (Merocodia, Uppsala, Sweden).^[20] Homeostatic model assessment of insulin resistance (HOMA-IR) was calculated as follows: (FBG [nmol/L] × Insulin [mU/mL]/22.5) using fasting values.^[28] Other laboratory-test details are available in the NHANES Laboratory/Medical Technologists Procedures Manual.^[29] Details on information on high-sensitivity C-reactive protein (hsCRP) concentration measurements are available elsewhere.^[24]

The triglyceride (TG)-glucose (TyG) index was calculated as the ln(Fasting TG [mg/dL] × Glucose [mg/dL]/2).^[30] The anthropometrically predicted VAT (apVAT) was estimated with sexspecific validated equations that included age, body mass index (BMI), and circumferences of the waist (WC) and thigh.^[31] The equation for men was: $6 \times WC - 4.41 \times Proximal thigh$ circumference + 1.19 × Age – 213.65; and the equation for women was: $2.15 \times WC - 3.63 \times Proximal thigh + 1.46 \times Age + 6:22 \times BMI - 92.713.^[31] VAI was calculated using sex-specific$ formulas: males (WC/39.68 + [1.88 × BMI]) × (TGs/1.03) ×(1.31/high-density lipoprotein [HDL]); females: (WC/36.58 +[1.89 × BMI]) × (TGs/0.81) × (1.52/HDL), where both TGs andHDL levels are expressed in mmol/L.^[32] LPA was calculated as(WC/65) × TG in men, and (WC/58) × TG in women.^[33]

2.2. Statistical analysis

We analyzed the data in compliance with the prescribed guidelines for analysis of complex NHANES data set, taking into account the masked variance and utilizing the proposed weighting methodology.^[20,34] Factor analysis was applied with orthogonal transformation (varimax procedure) to derive DPs based on the nutrients and bioactive compounds. Factors were retained for further analysis based on their natural interpretation and eigenvalues on the Screen test.^[35] We computed the factor score for each nutrient pattern by summing up intakes of nutrients weighted by their factor loadings.^[35] Each participant received a factor score for the identified pattern. We categorized the subjects based on quarters of nutrient pattern scores. We computed age, sex, race, physical activity, smoking, CRP, and history of diabetese adjusted means of our outcomes across quarter of DPs by using analysis of covariance. Adjusted multivariate linear regressions (age, sex, race, physical activity, smoking, CRP) were used to examine the association of score of food pattern with adiposity. All tests were two-sided, and P < .05was the level of significance.

3. Results

The analytical sample comprised 18,318 participants, of whom 48.3% (n=8607) were men. The mean age was 45.8 years in the overall sample and did not vary significantly in men and women (P=.126). White (non-Hispanic) participants formed the majority (69.4%) of the population. Furthermore, 56.1% (n=8759) of the participants were married, and 19.8% were current smokers (23.9% of men and 16.7% of women). The PCA method uncovered 3 DPs altogether explaining 55.9% of the variance in dietary nutrient consumption. The first DP was representative of

Table 1

Demographic and clinical characters of subjects.

Characteristics		Overall	Р
Sex	Men (%)	48.3%	<.001
	Women (%)	51.7%	
Age (y)	47.1±1.1		
Race/ethnicity	White (non-Hispanic) (%)	69.4%	<.001
	Non-Hispanic Black (%)	11.5%	
	Mexican-American (%)	8.4%	
	Other Hispanic (%)	4.5%	
	Other (%)	6.2%	
Marital status	Married (%)	56.1%	<.001
	Widowed (%)	61.1%	
	Divorced (%)	10.1%	
	Never married (%)	17.9%	
Education status	Less than high school (%)	19.1%	<.001
	Completed high school (%)	24.4%	
	More than high school (%)	56.4%	
Body mass index (kg/m ²)		28.7±0.05	
Waist circumference (cm)		98.2 <u>+</u> 0.12	
TyG index		8.78±0.002	
Serum Hs-CRP (mg/dL)		0.43±0.001	
Serum Apolipoprotein (B) (mg/dL)		94.2 <u>+</u> 0.25	
Fasting blood glucose (mg/dL)		100.2±0.021	
Plasma insulin (µU/mL)		2.31 ± 0.008	
HOMA-IR		3.66±0.051	
HbA1c (%)		5.66 ± 0.004	
apVAT		179.2 <u>+</u> 1.18	
Lipid accumulation product		68.5±0.56	
Visceral adiposity index		2.56 ± 0.02	

Value expressed as a mean and SEM or percent.

apVAT = anthropometrically predicted visceral adipose tissue, HbA1c = glycated hemoglobin, HOMA-B = homeostatic model assessment of β -cell function, HOMA-IR = homeostatic model assessment of insulin resistance, Hs-CRP = high-senility C-reactive protein, TyG index = triglyceride-glucose index.

a diet high in carbohydrate and sugar, total fat and saturated fatty acid (SFA), high-caloric dieatry pattern; the second DP was highly loaded with vitamins, minerals, and fiber (nutrient-dense dietary patten); and the third DP was mainly representative of high dietary PUFAs and MUFAs (healthy fat DP) (Table 1).

The adjusted (age, sex, race, physical activity, smoking, CRP) means of adposity factors (apVAT, LAP, VAI) and glucose/ insulin homeostasis (FBG, insulin, HOMA-IR) increased across increasing quarters of the first DP score (all P < .001, Table 2), while across increasing score of the second DP, the adjusted mean of apVAT, LAP, VAI, FBG, insulin, and HOMA-IR decreased (all P < .001, Table 2). Across increasing quarters of the third DP, just LAP and FBG showed significantly decreasing trend (P < .001, Table 2). Adjusted linear regressions revealed a significant and positive association between first DP and adiposity and glucose/ insulin homeostasis factors (all P < .001, Table 2), whereas there was a significant and negative association between second DP and same factors (all P < .001, Table 2).

4. Discussion

Findings from this study revealed that adiposity factors and markers of glucose/insulin homeostasis were positively associated with the diet which highly consisting of the carbohydrate and sugar, total fat and SFA, and inversly associated with diet comprising vitamins, minerals, and fiber. Moreover, we found a negative association between LAP and diet highly loaded with PUFA and MUFA.

In contrast to our findings, in a prospective study, no relation was detected between SFAs, MUFAs, PUFAs, and 5-year percent change in VAT^[14]; however, one cross-sectional study revealed a positive association between fat intake and VAT in overweight young adults aged 17 to 35 years.^[14] An Iranian investigation reported that increasing MUFA by decreasing total protein or PUFA in isoenergetic diets was positively associated with visceral adiposity index changes.^[36,37] The hypothesis that MUFAs are healthy fatty acids comes from studies investigating the impacts of olive oil, whereas further studies suggest MUFA intakes from animal sources to have different effects.^[36,37]

Contrary to our results, some observational studies did not find a significant association between carbohydrate intake and VAT^[12,13]; however, it has been proposed that replacing carbohydrate with total protein was positively associated with VAI in women only.^[37] A recent Iranian investigation reported that higher dietary proportions of protein and animal-derived MUFA could be positively associated with VAI; in addition, in isoenergetic diet, replacing carbohydrate, MUFAs, and PUFAs with protein was positively associated with 3-year changes in VAI.^[37] However, no significant association was reported between 2-year changes in total protein intake and change in VAT in 85 overweight adolescents aged 11 to 17 years,^[38] as well total protein intake was also not associated with 5-year percent change in VAT in 1114 black and hispanic overweight adults in another prospective study.^[14]

An investigation reported that LAP and VAI were markers of insulin resistance and metabolic-related disturbances in young women with polycystic ovary syndrome.^[39] Recent meta-analysis investigated the effects of saturated fat, polyunsaturated fat,

Table 2

Adjusted mean of adiposity factors across quarters of dietary patterns and adjusted linear regression between adiposity factors and score of dietary patterns.

	Dietary pattern quarters (first)					Dietary pattern quarters (second)						Dietary pattern quarters (third)						
Variables	Q1	Q2	Q3	Q4	Р	β	Q1	Q2	Q3	Q4	Р	β	Q1	Q2	Q3	Q4	Р	β
apVAT	176 ± 2.6	179±2.3	184±2.2	187 ± 1.9	<.00	1 0.424	189.2±1.5	186.5 ± 2.0	184.2±2.2	176.8±2.9	<.001	-0.073	180.1±2.8	181.1±3.1	180.2±2.8	180.3 ± 2.5	.182	-0.001
Lipid accumulation Product	64.3±1.2	65.8±1.1	68.4 ± 0.9	73.1±1.1	<.00	1 0.638	72.3±1.1	70.4±1.3	68.2±1.7	66.9±1.2	<.001	-1.240	74.2±1.1	73.2±1.2	71.6±1.5	69.3±1.4	<.001	-0.236
Visceral adiposity index	2.1±0.08	2.2±0.05	2.3 ± 0.03	2.7±0.01	<.00	1 4.632	2.9±0.08	2.6±0.01	2.5±0.03	2.3±0.05	5 <.001	-9.361	2.7±0.08	2.4±0.05	2.4±0.04	2.5±0.03	.096	0.002
Fasting blood glucose (mg/dL)	102.3±1.1	105.3±1.0	109.8 ± 0.9	115.9±1.2	<.00	1 0.652	115.2±1.0	111.9±1.1	110.5±1.0	104.9±1.9	<.001	-0.536	119.2±1.2	115.3±2.0	114.7±1.4	110.2±2.0	<.001	-0.293
Plasma insulin (µU/mL)	2.01 ± 0.01	2.09 ± 0.02	2.19±0.01	2.43±0.01	<.00	1 0.439	2.69 ± 0.02	2.43±0.01	2.12±0.03	2.10 ± 0.0^{-1}	1 <.001	-0.432	2.13±0.01	2.52±0.02	2.04±0.03	2.33±0.01	.0235	0.023
HOMA-IR	3.42 ± 0.11	3.51 ± 0.12	3.64 ± 0.36	3.92 ± 0.15	<.00	1 0.492	3.86 ± 0.12	3.79 ± 0.23	3.56±0.19	3.35 ± 0.14	4 <.001	-0.346	3.82	3.79	3.25	3.12	<.001	-0.183

Both analysis of covariance and linear regression corrected for age, sex, race, physical activity, smoking, C-reactive protein, and history of diabetese.

Values in bold indicated for the significant β value for the linear regression.

apVAT = anthropometrically predicted visceral adipose tissue, HOMA-IR = homeostatic model assessment of insulin resistance.

monounsaturated fat, and carbohydrate on glucose-insulin homeostasis.^[40] It reported that only energy intake substitution with PUFA was associated with lower fasting glucose, lower HbA1c, improved HOMA-IR, and improved insulin secretion capacity. Furthermore, insulin secretion capacity similarly improved when PUFA replaced MUFA. Experimental studies showed that PUFA suppresses oxidative stress, hepatic lipogenesis and steatosis, pancreatic lipotoxicity, and insulin resistance.^[41] In addition, MUFA consumption did not appear to significantly influence fasting glucose, compared to others macronutrients, however, was reported to reduce HbA1c and improve HOMA-IR in comparison to either carbohydrate or SFA.^[40]

There are several limitations to this study. First, the results based on this cross-sectional study, although it is nationally representative, cannot demonstrate a causal relationship between DPs and VAT. Second, although our analysis included known potential confounding variables that can affect adiposity in terms of environmental and genetic factors, residual confounding variables may still exist. Moreover, we did not have data on the direct measurement of the VAT for validation. This study has several strengths. We had a large sample, selected randomly from general population; therefore, the results obtained from nationally representative samples can be extrapolated to the general population.

5. Conclusion

In conclusion, our findings suggest that LAP and VAI could potentially be used as indirect measures of VAT in routine setting and for research purpose, considering that they are likely more affordable than other advocated indirect measures such as markers of insulin resistance. Also they are significantly related to the DPs in same line with glucose/insulin homeostasis and anthropometrics indices.

Author contributions

Conceptualization: Hong- Gao, Andre Pascal Pascal.

Data curation: Mohsen Mazidi.

Formal analysis: Andre Pascal Kengne, Mohsen Mazidi.

Methodology: Andre Pascal Pascal.

Supervision: Hong- Gao, Andre Pascal Pascal.

Writing - original draft: Mohsen Mazidi, Hong- Gao.

Writing – review & editing: Andre Pascal Pascal.

References

- Brown JC, Harhay MO, Harhay MN. Anthropometrically-predicted visceral adipose tissue and mortality among men and women in the third national health and nutrition examination survey (NHANES III). Am J Hum Biol 2017;29:
- [2] Després J-P. Body fat distribution and risk of cardiovascular disease: an update. Circulation 2012;126:1301–13.
- [3] Mirmiran P, Bahadoran Z, Azizi F. Lipid accumulation product is associated with insulin resistance, lipid peroxidation, and systemic inflammation in type 2 diabetic patients. Endocrinol Metab 2014; 29: 443–9.
- [4] Wakabayashi I, Daimon T. A strong association between lipid accumulation product and diabetes mellitus in Japanese women and men. J Atheroscler Thromb 2014;21:282–8.
- [5] Nascimento-Ferreira MV, Rendo-Urteaga T, Vilanova-Campelo RC, et al. The lipid accumulation product is a powerful tool to predict metabolic syndrome in undiagnosed Brazilian adults. Clin Nutr 2017; 36:1693–700.
- [6] Oh JY, Sung YA, Lee HJ. The lipid accumulation product as a useful index for identifying abnormal glucose regulation in young Korean women. Diabet Med 2013;30:436–42.

- [7] Du T, Yu X, Zhang J, et al. Lipid accumulation product and visceral adiposity index are effective markers for identifying the metabolically obese normal-weight phenotype. Acta Diabetol 2015;52:855–63.
- [8] Guo S-X, Zhang X-H, Zhang J-Y, et al. Visceral adiposity and anthropometric indicators as screening tools of metabolic syndrome among low income rural adults in Xinjiang. Sci Rep 2016;6:36091.
- [9] Fischer K, Moewes D, Koch M, et al. MRI-determined total volumes of visceral and subcutaneous abdominal and trunk adipose tissue are differentially and sex-dependently associated with patterns of estimated usual nutrient intake in a northern German population. Am J Clin Nutr 2015;101:794–807.
- [10] Fischer K, Pick JA, Moewes D, et al. Qualitative aspects of diet affecting visceral and subcutaneous abdominal adipose tissue: a systematic review of observational and controlled intervention studies. Nutr Rev 2015; 73:191–215.
- [11] Veum VL, Laupsa-Borge J, Eng Ø, et al. Visceral adiposity and metabolic syndrome after very high-fat and low-fat isocaloric diets: a randomized controlled trial. Am J Clin Nutr 2017;105:85–99.
- [12] Bailey BW, Sullivan DK, Kirk EP, et al. Dietary predictors of visceral adiposity in overweight young adults. Br J Nutr 2010;103:1702–5.
- [13] Kondoh T, Takase H, Yamaguchi TF, et al. Association of dietary factors with abdominal subcutaneous and visceral adiposity in Japanese men. Obes Res Clin Pract 2014;8:e16–25.
- [14] Hairston KG, Vitolins MZ, Norris JM, et al. Lifestyle factors and 5-year abdominal fat accumulation in a minority cohort: the IRAS Family Study. Obesity (Silver Spring, MD) 2012;20:421–7.
- [15] Jacobs DRJr, Steffen LM. Nutrients, foods, and dietary patterns as exposures in research: a framework for food synergy. Am J Clin Nutr 2003;78:5085–135.
- [16] Hu FB. Dietary pattern analysis: a new direction in nutritional epidemiology. Curr Opin Lipidol 2002;13:3–9.
- [17] Jacques PF, Tucker KL. Are dietary patterns useful for understanding the role of diet in chronic disease? Am J Clin Nutr 2001;73:1–2.
- [18] Salehi-Abargouei A, Esmaillzadeh A, Azadbakht L, et al. Nutrient patterns and their relation to general and abdominal obesity in Iranian adults: findings from the SEPAHAN Study. Eur J Nutr 2016;55:505–18.
- [19] Samieri C, Ginder Coupez V, Lorrain S, et al. Nutrient patterns and risk of fracture in older subjects: results from the Three-City Study. Osteoporos Int 2013;24:1295–305.
- [20] Mohsen Mazidi EDM, Maciej , Banach . The association of telomere length and serum 25-hydroxyvitamin D levels in US adults: the National Health and Nutrition Examination Survey. Arch Med Sci 2017;13:61–5.
- [21] Kalk WJ, Joffe BI. The metabolic syndrome, insulin resistance, and its surrogates in African and white subjects with type 2 diabetes in South Africa. Metab Syndr Relat Disord 2008;6:247–55.
- [22] Needham BL, Adler N, Gregorich S, et al. Socioeconomic status, health behavior, and leukocyte telomere length in the National Health and Nutrition Examination Survey, 1999–2002. Soc Sci Med 2013;85:1–8.
- [23] Remer T. Influence of nutrition on acid-base balance metabolic aspects. Eur J Nutr 2001;40:214–20.
- [24] Available at: http://www.cdc.gov/NCHS/data/nhanes/nhanes_09_10/ CRP_F_met.pdf. Accessed August 19, 2013.
- [25] Ahluwalia N, Andreeva VA, Kesse-Guyot E, et al. Dietary patterns, inflammation and the metabolic syndrome. Diabetes Metab 2013;39: 99–110.
- [26] Ahluwalia N, Dwyer J, Terry A, et al. Update on NHANES dietary data: focus on collection, release, analytical considerations, and uses to inform public policy. Adv Nutr 2016;7:121–34.
- [27] Moshfegh AJ, Rhodes DG, Baer DJ, et al. The US Department of Agriculture Automated Multiple-Pass Method reduces bias in the collection of energy intakes. Am J Clin Nutr 2008;88:324–32.
- [28] Musso G, Gambino R, Bo S, et al. Should nonalcoholic fatty liver disease be included in the definition of metabolic syndrome? A cross-sectional comparison with Adult Treatment Panel III criteria in nonobese nondiabetic subjects. Diabetes Care 2008;31:562–8.
- [29] National Center for Health Statistics CfDCaPNHaNESA. Available at: http://www.cdc.gov/nchs/nhanes.htm. Accessed April 1, 2017.
- [30] Simental-Mendia LE, Rodriguez-Moran M, Guerrero-Romero F. The product of fasting glucose and triglycerides as surrogate for identifying insulin resistance in apparently healthy subjects. Metab Syndr Relat Disord 2008;6:299–304.
- [31] Samouda H, Dutour A, Chaumoitre K, et al. VAT=TAAT-SAAT: innovative anthropometric model to predict visceral adipose tissue without resort to CT-Scan or DXA. Obesity (Silver Spring) 2013;21: E41–50.

- [32] Amato MC, Giordano C, Galia M, et al. Visceral Adiposity Index: a reliable indicator of visceral fat function associated with cardiometabolic risk. Diabetes Care 2010;33:920–2.
- [33] Onat A, Avci GS, Barlan MM, et al. Measures of abdominal obesity assessed for visceral adiposity and relation to coronary risk. Int J Obes Relat Metab Disord 2004;28:1018–25.
- [34] Mazidi M, Mikhailidis DP, Banach M. Higher dietary acid load is associated with higher likelihood of peripheral arterial disease among American adults. J Diabetes Complications 2018;pii: S1056-8727(17) 31704-X. doi: 10.1016/j.jdiacomp.2018.03.001. [Epub ahead of print].
- [35] Stanhope KL. Role of fructose-containing sugars in the epidemics of obesity and metabolic syndrome. Annu Rev Med 2012;63:329–43.
- [36] Hoffman R, Gerber M. Evaluating and adapting the Mediterranean diet for non-Mediterranean populations: a critical appraisal. Nutr Rev 2013;71:573–84.
- [37] Moslehi N, Ehsani B, Mirmiran P, et al. Association of dietary proportions of macronutrients with visceral adiposity index: non-

substitution and iso-energetic substitution models in a prospective study. Nutrients 2015;7:8859–70.

- [38] Davis JN, Alexander KE, Ventura EE, et al. Inverse relation between dietary fiber intake and visceral adiposity in overweight Latino youth. Am J Clin Nutr 2009;90:1160–6.
- [39] Abruzzese GA, Cerrrone GE, Gamez JM, et al. Lipid accumulation product (LAP) and visceral adiposity index (VAI) as markers of insulin resistance and metabolic associated disturbances in young argentine women with polycystic ovary syndrome. Horm Metab Res 2017;49: 23–9.
- [40] Imamura F, Micha R, Wu JH, et al. Effects of saturated fat, polyunsaturated fat, monounsaturated fat, and carbohydrate on glucose-insulin homeostasis: a systematic review and meta-analysis of randomised controlled feeding trials. PLoS Med 2016;13:e1002087.
- [41] Rosqvist F, Iggman D, Kullberg J, et al. Overfeeding polyunsaturated and saturated fat causes distinct effects on liver and visceral fat accumulation in humans. Diabetes 2014;63:2356–68.