

Original article:

BROMELAIN CAPPED GOLD NANOPARTICLES AS THE NOVEL DRUG DELIVERY CARRIERS TO AGGRANDIZE EFFECT OF THE ANTIBIOTIC LEVOFLOXACIN

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ABSTRACT

To develop bromelain capped gold nanoparticles (BRN capped Au-NPs) as the effective drug delivery carriers of the antibiotic levofloxacin (LvN) and evaluate antibacterial potential of its bioconjugated form compared to pure LvN. BRN capped Au-NPs were synthesized by *in vitro* method and bioconjugated to LvN using 1-ethyl-3-(3-dimethylamino-propyl)-carbodiimide as activator to form Au-BRN-LvN-NPs. These were characterized for mean particle size by dynamic light scattering analysis, zeta potential by Zetasizer nanosystem analysis and transmission electron microscopy (TEM) on carbon coated TEM copper grids by TEM respectively. Drug loading efficiency of LvN was calculated using UV-visible spectroscopy by standard curve of pure LvN. Antibacterial efficacy of Au-BRN-LvN-NPs and pure LvN was determined by evaluating minimum inhibitory concentration (MIC) against *Staphylococcus aureus* and *Eschereschia coli*. Two peaks were observed in Au-BRN-LvN-NPs spectrum one at 307 nm and other at 526 nm while one peak in BRN capped Au-NPs at 522 nm during UV spectroscopy suggesting red shift. The drug loading efficiency of LvN was found to be $84.8 \pm 2.41\%$. The diameter of Au-BRN-LvN-NPs and BRN capped Au-NPs were found to be $(58.65 \pm 2 \text{ nm}, 38.11 \pm 2 \text{ nm})$, zeta potential $(-9.01 \text{ mV}, -13.8 \text{ mV})$ and surface morphology $(\sim 13.2 \text{ nm}, 11.4 \text{ nm})$ respectively. The MICs against *S. aureus* and *E. coli* were found to be $(0.128 \mu\text{g/mL}, 1.10 \mu\text{g/mL})$ for Au-BRN-LvN-NPs and $(0.547 \mu\text{g/mL}, 1.96 \mu\text{g/mL})$ for pure LvN. The results suggested that BRN capped Au-NPs can be used as effective drug delivery carriers of the antibiotic LvN. The Au-BRN-LvN-NPs exhibited enhanced antibacterial activity compared to pure LvN alone. (Graphical abstract see Figure 1)

Keywords: Antibacterial activity, bromelain, gold nanoparticles, levofloxacin, novel drug delivery carriers

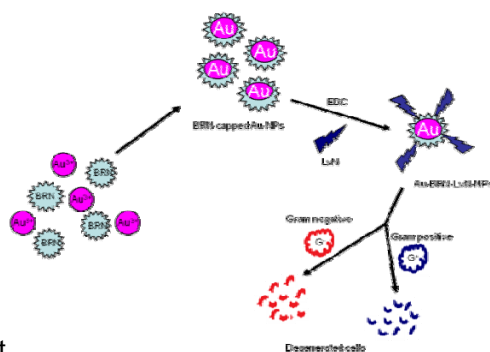


Figure 1: Graphical abstract

INTRODUCTION

The metallic nanoparticles derived from noble metals such as gold has attracted the researchers all over the world regarding their escalating use in delivery of drugs (Emerich and Thanos, 2006). These particles build up new physico-chemical properties which are absent in individual molecules. Specially, the gold nanoparticles are being targeted nowadays by the researchers since it not only encompasses the advantages of “nano-sizing” but it also includes surface functionalization necessary to augment stability and enable conjugation with biomolecules (Couto et al., 2016). Surface functionalization of the gold nanoparticles has opened new frontiers for delivery of drugs such as anticancer and the antimicrobial drugs (Dhar et al., 2008, Selvaraj and Alagar, 2007). Bovine serum albumin capped gold nanoparticles of aminoglycosidic antibiotics and the acridine derivatives bioconjugated on citrate stabilized gold nanoparticles have been used as the effective drug delivery vehicles (Rastogi et al., 2012, Mitra et al, 2014). Thus, these exciting approaches have motivated the search for novel molecules for the synthesis of gold nanoparticles.

Levofloxacin (LvN) which is chemically (S)-9-fluoro-2,3-dihydro-3-methyl-10-(4-methylpiperazin-1-yl)-7-oxo-7H-pyrido[1,2,3-de]-1,4-benzoxazine-6-carboxylic acid, is a broad spectrum fluoroquinolone antibiotic active against both Gram positive and Gram negative bacteria functioning by inhibiting the type II topoisomerase enzymes namely DNA gyrase and topoisomerase IV (Drlica and Zhao, 1997). It is recommended as a first-line treatment option by ‘Infectious Disease Society of America’ for catheter associated urinary tract infections in adults and in prosthetic joint infection in combination with rifampicin (Hooton et al., 2010; Osmon et al., 2013).

Bromelain (BRN) is a proteolytic enzyme derived from the plant *Ananas comosus* belonging to family Bromeliaceae (Taussig and Batkin, 1988). It has shown analgesic, anti-inflammatory, anti-arthritis, an-

titumour, anthelmintic, antimicrobial and several other pharmacological activities (Leipner et al, 2001; Beez et al, 2007; Rowan et al., 1990; Praveen et al., 2014). It was found to be less toxic having mutagenic potential (Castell et al., 1997; Chobotova et al., 2010). Hence, the present study was aimed to develop bromelain capped gold nanoparticles as the effective drug delivery carrier of the antibiotic levofloxacin and to evaluate the antibacterial potential of its bioconjugated form as compared to pure LvN alone.

MATERIALS AND METHODS

Chemicals and reagents

All the chemicals and solvents used in the present investigation were of analytical grade. Levofloxacin was procured from Yarrow Chem Products, Mumbai (India) and bromelain from Merck Chemicals Darmstadt, Germany.

Synthesis of gold nanoparticles

Gold nanoparticles (Au-NPs) were synthesized by *in vitro* method. The 3 μ l of 1.0 mM H[AuCl₄] prepared in 50 mM phosphate buffer was taken in 3 mL of freshly prepared bromelain (0.33 mg/mL) and incubated at the temperature of 40 °C for 48 h in an incubator. The reaction mixture without bromelain was used as a control. To ensure the formation of nanoparticles, the samples were removed from the reaction mixture at regular intervals and analyzed by UV-visible spectroscopy. On completion of the reaction, Au-NPs were collected by centrifugation at 30000 g for 30 min, washed twice with MilliQ water and the excess bromelain was removed by treating with 50 % ethanol (v/v) (Khan et al., 2015a).

Bioconjugation of gold nanoparticles to levofloxacin

Synthesized Au-NPs were bioconjugated to LvN by using 1-ethyl-3-(3-dimethylamino-propyl)-carbodiimide (EDC) as the activator (Timkovich, 1977). The 5 mM EDC was added to the 5 ml reaction mixture containing 250 μ g of LvN, 250 μ g of Au-

NPs and 50 mM MES/HEPES buffer (HiMedia laboratories, India) in aliquots within 3 h at 30 °C for the coupling process. The bioconjugates so formed were separated from unconjugated Au-NPs by passing the reaction mixture through Biogel P-30 gel filtration column pre-equilibrated with 20 mM HEPES buffer (*pH* 6.0) containing 150 mM sodium chloride. The fractions were scanned between 200-900 nm and subsequently pooled. The pooled samples were then dialyzed against distilled water and used for further characterization (Khan et al., 2015b). The samples were removed at regular intervals and analyzed in UV-visible spectroscopy (Shimadzu dual-beam spectrophotometer, model UV-1601 PC, Japan) at a resolution of 1 nm to confirm the formation of 'BRN capped Au-NPs' and 'levofloxacin conjugated with BRN capped Au-NPs' (Au-BRN-LvN-NPs).

Characterization of synthesized gold nanoparticles and bioconjugated gold nanoparticles

Measurement of mean particle size and zeta potential of 'BRN capped Au-NPs' and Au-BRN-LvN-NPs

The samples of 'BRN capped Au-NPs' and Au-BRN-LvN-NPs were sonicated separately for 1 min and then, taken in a DTS0112-low volume disposable sizing cuvette of 1.5 mL for the measurement of mean particle size with a dynamic light scattering (DLS) particle size analyzer (Zetasizer Nano-ZS, model ZEN3600, Malvern Instrument Ltd, Malvern, UK) (Akhtar et al., 2014). Zeta potential values were also assessed to check the stability of the gold nanoparticles with a Malvern Zetasizer Nanosystem (Worcestershire, UK).

Transmission electron microscopy of 'BRN capped Au-NPs' and Au-BRN-LvN-NPs

Transmission electron microscopy (TEM) was analyzed by drop drying the solution of biosynthesized 'BRN capped Au-NPs' and Au-BRN-LvN-NPs on carbon coated TEM copper grids followed by meas-

urements on TEM (FEI Company, Tecnaia G2 Spirit BioTWIN) at an accelerating voltage of 80 kV.

Fourier transform infrared spectroscopy of 'BRN capped Au-NPs' and Au-BRN-LvN-NPs

The Fourier transform infrared (FTIR) spectra of 'BRN capped Au-NPs' and Au-BRN-LvN-NPs were recorded using Perkin-Elmer Spectrum Two FT-IR (Perkin Elmer Inc., Tres Cantos, Madrid) equipped with a Universal attenuated total reflectance sampling device and scanned at room temperature in transmission mode over the wave number range of 4000-650 cm^{-1} at a resolution of 4 cm^{-1} .

Drug loading efficiency of levofloxacin

Drug loading efficiency of LvN was calculated using UV-visible spectroscopy at the wavelength of 307 nm/526 nm where the standard curve of the pure drug LvN was established and the amount of unbound LvN was calculated from it. The actual amount of bioconjugated LvN was calculated by subtracting unbound LvN from the total amount of LvN added. The amount of bioconjugated LvN was calculated using the equation [% bioconjugation = (amount of drug conjugated / total drug added) \times 100].

Evaluation of antibacterial efficacy of bioconjugated levofloxacin, Au-BRN-LvN-NPs over pure levofloxacin alone

The antibacterial efficacy of bioconjugated LvN (Au-BRN-LvN-NPs) was determined by evaluating the minimum inhibitory concentration (MIC) of Au-BRN-LvN-NPs and pure LvN against Gram positive bacteria *Staphylococcus aureus* (NCIM No. 2079) and Gram negative bacteria *Escherichia coli* (NCIM No. 2065) (Amsterdam, 1996). The bacterial strains in mid logarithmic phase were harvested by centrifugation, washed with 10 mM sodium phosphate buffer (PB) of *pH* 7.4, and diluted to 2×10^5 colony forming units per ml (CFU/mL) in PB containing 0.03 % Luria-Bertani (LB) broth. Au-BRN-LvN-NPs were serially diluted in

50 μ L of LB medium in 96-well microtitre plates so as to achieve the desired concentrations with bacterial inoculums (5×10^4 CFU/well) and were incubated at 37 °C overnight. The MIC was taken as the lowest Au-BRN-LvN-NPs concentration at which growth was inhibited (Khan et al., 2008). For agar plate count method (Stenger et al., 1998), 25 μ L aliquot of bacteria (1×10^5 CFU/mL) was incubated with 25 μ L of samples at 37 °C for 2 h (Hamamoto et al., 2002). The mixtures were 10-fold serially diluted in PB, plated on LB agar and then incubated overnight at 37 °C. Bacterial colonies were enumerated following the day. After the determination of the MIC from the microtitre plate wells with no visible growth, the samples were removed for serial subcultivation (2 μ L) into microtitre plates containing 100 μ L of broth/well and further incubated for 24 h in order to determine the bactericidal activity measurement (MBC), the lowest concentration at which there is no visible bacterial growth indicating 99.5 % killing of the original inoculums. The absorbance of each well was measured at a wavelength of 620 nm by microtitre plate reader (Bio-Rad laboratories Inc., India) and compared with a control. Autoclaved water and 'BRN capped Au-NPs' were used as a negative control for each experiment. The proce-

cedure was repeated for the pure levofloxacin alone too.

RESULTS

Bioconjugation of gold nanoparticles to levofloxacin and drug loading efficiency of levofloxacin

The plasmon band was observed for the wine red colloidal gold nanoparticles at 522 nm in the UV-visible spectrum. The colloidal solution containing 'BRN capped Au-NPs' had shown very intense and characteristic pink red color. Two peaks were observed in Au-BRN-LvN-NPs spectrum one at 307 nm and other one at 526 nm during ultraviolet (UV) spectroscopy (Figure 2). The drug loading efficiency of LvN was found to be 84.8 ± 2.41 %.

Measurement of mean particle size and zeta potential of 'BRN capped Au-NPs' and Au-BRN-LvN-NPs

The diameter of Au-BRN-LvN-NPs was found to be 58.65 ± 2 nm while that of its unconjugated form 'BRN capped Au-NPs' as 38.11 ± 2 nm (Figure 3). The zeta potential of Au-BRN-LvN-NPs was found to be -9.01 mV and that of its unconjugated form 'BRN capped Au-NPs' -13.8 mV (Figure 4).

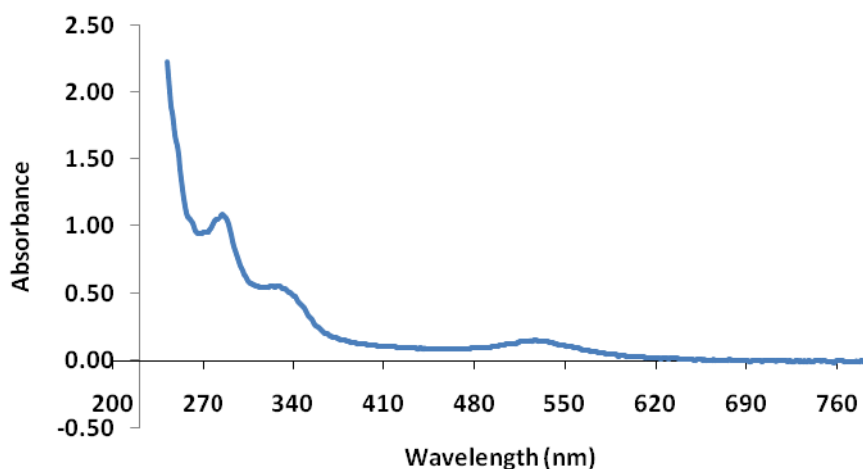


Figure 2: UV-visible absorption spectra of BRN capped gold nanoparticles conjugated with levofloxacin (Au-BRN-LvN-NPs)

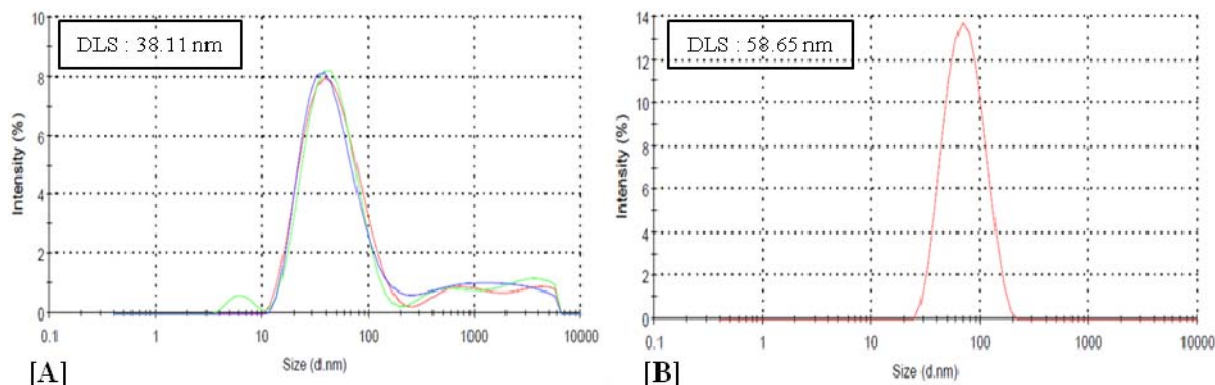


Figure 3: Dynamic light scattering (DLS) analysis of [A] BRN capped gold nanoparticles (BRN capped Au-NPs), and [B] BRN capped gold nanoparticles conjugated with levofloxacin (Au-BRN-LvN-NPs)

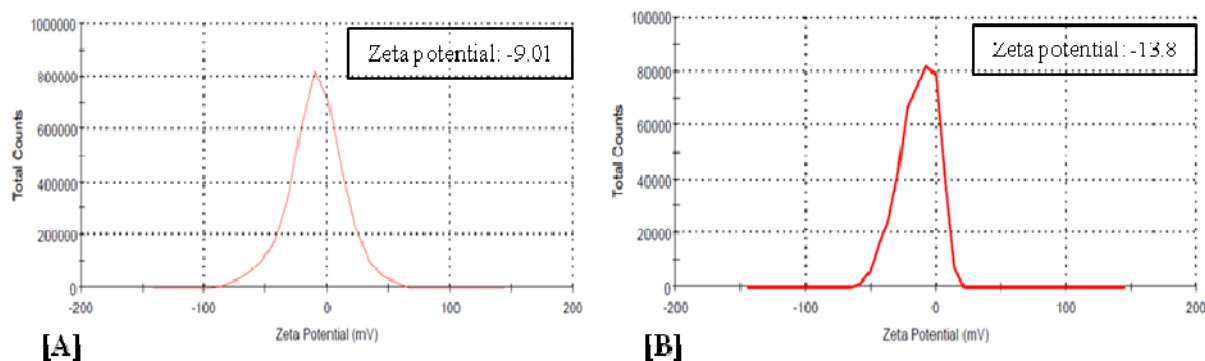


Figure 4: Zetasizer analysis of [A] BRN capped gold nanoparticles conjugated with levofloxacin (Au-BRN-LvN-NPs), and [B] BRN capped gold nanoparticles (BRN capped Au-NPs).

Transmission electron microscopy of ‘BRN capped Au-NPs’ and Au-BRN-LvN-NPs

Surface morphological study of ‘BRN capped Au-NPs’ and Au-BRN-LvN-NPs showed the formation of spherical particles with the particle size of 11.4 nm of ‘BRN capped Au-NPs’. The particle size of Au-BRN-LvN-NPs was found to be ~13.2 nm and it retained the spherical shape (Figure 5).

Fourier transform infrared spectroscopy of ‘BRN capped Au-NPs’ and Au-BRN-LvN-NPs

FTIR spectrum of ‘BRN capped Au-NPs’ is shown in Figure 6(A) which has showed the characteristic C–N stretch vibration frequencies of monoalkyl guanidinium assigned to the observed IR bands at 1641, 1425-1256 and 1100 cm^{-1} . The band at 1760-1670 cm^{-1} (s) showed the presence of C = O groups (amides at ~1640 cm^{-1}). FTIR spectrum of

Au-BRN-LvN-NPs is shown in Figure 6(B) which has shown the presence of peak at 1634.24 cm^{-1} due to the presence of C = O (str) of the amide I linkage and the presence of peak at 3338 cm^{-1} due OH stretching vibrations.

Evaluation of antibacterial efficacy of bio-conjugated levofloxacin over free levofloxacin

Percent inhibition of the bacteria was increasing with increasing doses of the pure levofloxacin and Au-BRN-LvN-NPs. The MICs of Au-BRN-LvN-NPs against *S. aureus* (NCIM No. 2079) and *E. coli* (NCIM No. 2065) were found to be 0.128 $\mu\text{g/ml}$ and 1.10 $\mu\text{g/ml}$ respectively, whereas MICs of the pure levofloxacin against *S. aureus* and *E. coli* were found to be 0.547 $\mu\text{g/ml}$ and 1.96 $\mu\text{g/ml}$ respectively (Figure 7).

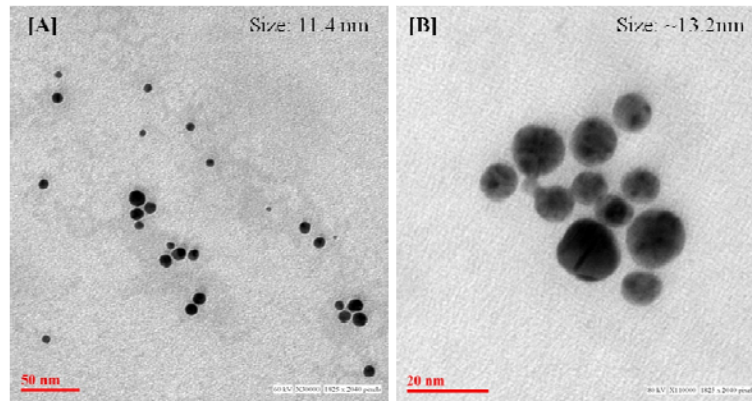


Figure 5: Transmission electron microscopic (TEM) images of **[A]** BRN capped gold nanoparticles (BRN capped Au-NPs), and **[B]** BRN capped gold nanoparticles conjugated with levofloxacin (Au-BRN-LvN-NPs)

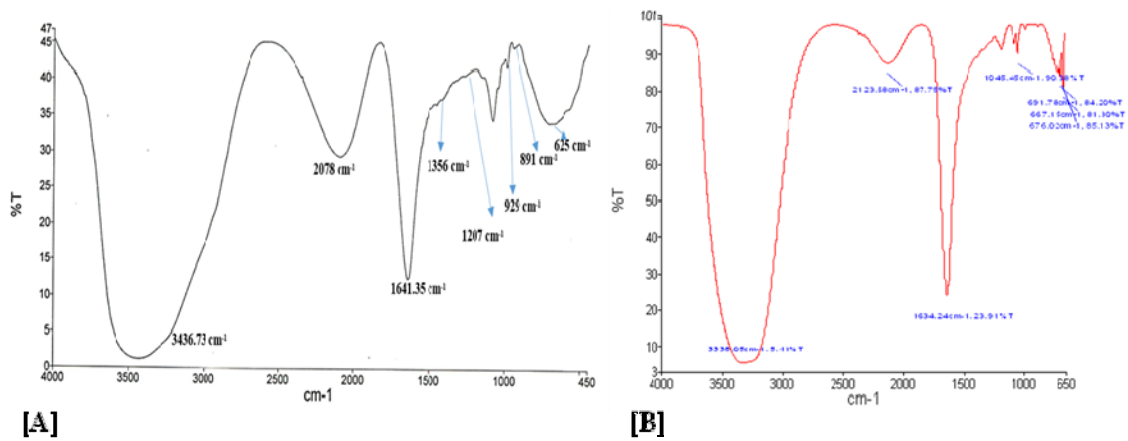


Figure 6: Fourier transform infrared (FTIR) spectra of **[A]** Pure levofloxacin, and **[B]** BRN capped gold nanoparticles conjugated with levofloxacin (Au-BRN-LvN-NPs)

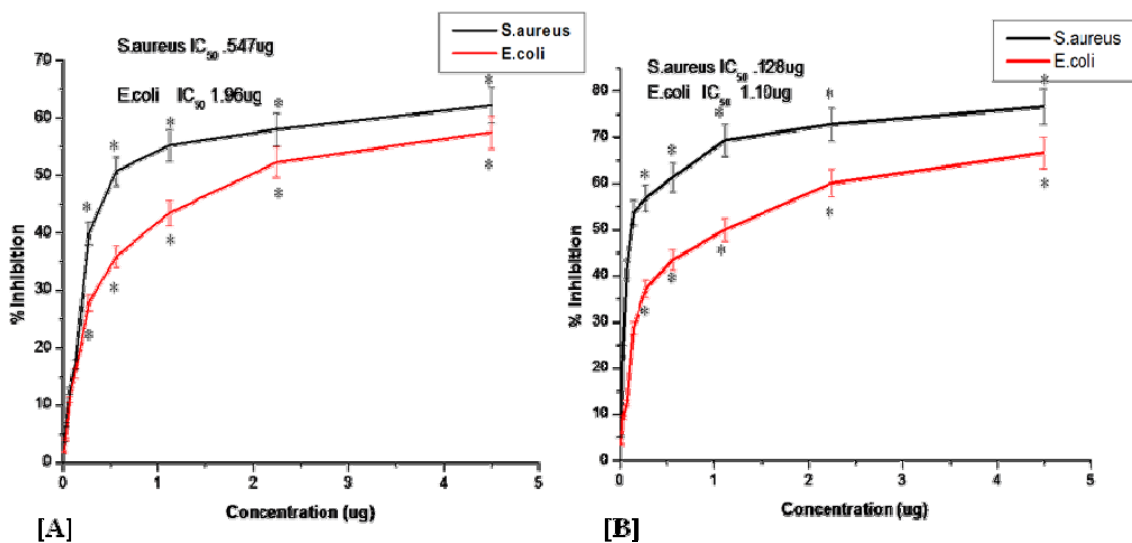


Figure 7: Plot of percent inhibition of bacterial strains versus different concentrations of **[A]** pure levofloxacin, and **[B]** BRN capped gold nanoparticles conjugated with levofloxacin (Au-BRN-LvN-NPs)

DISCUSSION

The plasmon band was observed for the wine red colloidal Au-NPs at 522 nm in the UV-visible spectrum which is characteristic of Au-NPs (Liao et al., 2006). The colloidal solution containing 'BRN capped Au-NPs' had shown very intense and characteristic pink red color confirming the formation of Au-NPs. However, two peaks were observed during UV-Visible spectral analysis of Au-BRN-LvN-NPs one at 307 nm due to aromatic acid transitions of LvN and other one at 526 nm due to red shift. This red shift in the plasmon band suggests different dielectric environment which confers conjugation of LvN with 'BRN capped Au-NPs'. The amount of bioconjugated LvN (Au-BRN-LvN-NPs) was found to be 84.8 ± 2.41 % indicating that the LvN was efficiently loaded on synthesized BRN capped Au-NPs by using EDC as the activator.

DLS provides hydrodynamic diameter which includes inorganic core and the thin electric dipole layer of the solvent that adheres to the surface of the nanoparticles (Akhtar et al., 2014). The diameter of Au-BRN-LvN-NPs was found to be 58.65 ± 2 nm while that of its unconjugated form 'BRN capped Au-NPs' 38.11 ± 2 nm proving that bioconjugation of LvN has taken place to form Au-BRN-LvN-NPs. Zeta potential gives an indication of the stability of a colloidal system against agglomeration. The zeta potential of Au-BRN-LvN-NPs was found to be -9.01 mV which is well in range to prevent agglomeration as compared to its unconjugated form 'BRN capped Au-NPs' which was found to be -13.8 mV supporting the absence of nanoparticle aggregation after conjugation. This change in zeta potential may be ascribed to the decrease in number of carboxylic groups which are involved in conjugation of LvN.

During surface morphological study by TEM, the particle size of Au-BRN-LvN-NPs was found to be ~ 13.2 nm while that of 'BRN capped Au-NPs' as 11.4 nm. It is in coherence with the DLS and further corroborates bioconjugation of LvN with 'BRN

capped Au-NPs' and supports the absence of aggregation of nanoparticles after bioconjugation.

From the results of the study, it was found that the percent inhibition of the bacteria *S. aureus* and *E. coli* were increasing with increasing doses of pure levofloxacin and Au-BRN-LvN-NPs suggesting their antibacterial activity against *S. aureus* and *E. coli* in dose dependent manner. The functionalized nanoparticles showed superior antibacterial activity compared to pure LvN at the similar concentration. From the results of the study, it is also clear that there was reduction in the IC₅₀ values from 0.547 µg/ml and 1.96 µg/ml of pure LvN to 0.128 µg/ml and 1.10 µg/ml of Au-BRN-LvN-NPs against *S. aureus* and *E. coli* respectively which may be attributed to disruptions in the cell wall and membrane structures of bacteria by it leading to loss of cellular integrity and finally cell death. The IC₅₀ value for Au-BRN-LvN-NPs was less against *S. aureus* as compared to that of *E. coli* suggesting that Au-BRN-LvN-NPs are more effective against *S. aureus* bacterium. On the other hand, Au-BRN-LvN-NPs are also able to generate reactive oxygen species which may play a synergistic role in killing bacteria (Nadeau et al., 2008). The probable reasons for superior antibacterial activity of Au-BRN-LvN-NPs may be its superior stability and transport of a huge number of LvN molecules into a highly localized area at the site of particle-bacterium contact (Burygin et al., 2009).

CONCLUSION

The results suggest that bromelain capped gold nanoparticles can be used as effective carriers for levofloxacin molecules. The bioconjugated bromelain capped gold nanoparticles exhibited superior antibacterial activity against both Gram negative and Gram positive bacteria compared to pure levofloxacin which may be due to its superior stability and transport of a huge number of LvN molecules into a highly localized area at the site of particle-bacterium contact.

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Conflict of interest

The authors declare no conflict of interest.

REFERENCES

- Akhtar J, Siddiqui HH, Badruddeen, Fareed S, Aqil M. Nanomulsion as a carrier for efficient delivery of metformin. *Curr Drug Deliv*. 2014;11:243-52.
- Amsterdam D. Susceptibility testing of antimicrobials in liquid media. *Antibiotics Lab Med*. 1996;4:61-143.
- Beez R, Lopes MTP, Salas CE, Hernandez M. *In vivo* antitumoral activity of stem pineapple (*Ananas comosus*) bromelain. *Planta Med*. 2007;73:1377-83.
- Burygin G, Khlebtsov B, Shantrokha A, Dykman L, Bogatyrev V, Khlebtsov N. On the enhanced antibacterial activity of antibiotics mixed with gold nanoparticles. *Nanoscale Res Lett*. 2009;4:794-801.
- Castell JV, Friedrich G, Kuhn CS, Poppe GE. Intestinal absorption of undegraded proteins in men: presence of bromelain in plasma after oral intake. *Am J Physiol Gastrointest Liver Physiol*. 1997;273:G139-46.
- Chobotova K, Vernallis AB, Majid FAA. Bromelain's activity and potential as an anti-cancer agent: current evidence and perspectives. *Cancer Lett*. 2010;290:148-56.
- Couto C, Vitorino R, Daniel-da-Silva AL. Gold nanoparticles and bioconjugation: a pathway for proteomic applications. *Crit Rev Biotechnol*. 2016;10:1-13.
- Dhar S, Reddy EM, Shiras A, Pokharkar V, Prasad BEE. Natural gum reduced/stabilized gold nanoparticles for drug delivery formulations. *Chem Eur J*. 2008;14:10244-50.
- Drlica K, Zhao X. DNA gyrase, topoisomerase IV, and the 4-quinolones. *Microbiol Mol Biol Rev*. 1997;61:377-92.
- Emerich DF, Thanos CG. The pinpoint promise of nanoparticle-based drug delivery and molecular diagnosis. *Biomol Eng*. 2006;23:171.
- Hamamoto K, Kida Y, Zhang Y, Shimizu T, Kuwano K. Antimicrobial activity and stability to proteolysis of small linear cationic peptides with D-amino acid substitutions. *Microbiol Immunol*. 2002;46:741-9.
- Hooton TM, Bradley SF, Cardenas DD, Colgan R, Geerlings SE, Rice JC, et al. Diagnosis, prevention, and treatment of catheter-associated urinary tract infection in adults, 2009: International Clinical Practice Guidelines from the Infectious Diseases Society of America. *Clin Infect Dis*. 2010;50:625-63.
- Khan MS, Siddiqui SA, Siddiqui MSRA, Goswami U, Srinivasan KV, Khan MI. Antibacterial activity of synthesized 2, 4, 5-trisubstituted imidazole derivatives. *Chem Biol Drug Des*. 2008;72:197-204.
- Khan S, Rizvi SMD, Avaish M, Arshad M, Bagga P, Khan MS. A novel process for size controlled biosynthesis of gold nanoparticles using bromelain. *Materials Lett*. 2015a;159:373-6.
- Khan S, Haseeb M, Baig MH, Bagga PS, Siddiqui HH, Kamal MA, et al. Improved efficiency and stability of secnidazole- An ideal delivery system. *Saudi J Biol Sci*. 2015b;22:42-9.
- Leipner J, Iten F, Saller R. Therapy with proteolytic enzymes in rheumatic disorders. *BioDrugs*. 2001;15:779-89.
- Liao H, Nehl CL, Hafner JH. Biomedical applications of plasmon resonant metal nanoparticles. *Nanomedicine*. 2006;1:201-8.
- Mitra P, Chakraborty PK, Saha P, Ray P, Basu S. Antibacterial efficacy of acridine derivatives conjugated with gold nanoparticles. *Int J Pharm*. 2014;473:636-43.
- Nadeau JL, Clarke SJ, Suresh AK, Khatchadourian RA, Dumas EM. The relationship of QD composition and conjugate to cellular uptake and toxicity. In: Osiński M, Jovin TM, Yamamoto K (eds.): *Colloidal quantum dots for biomedical applications III* (paper no. 68660J). Bellingham, WA: SPIE, 2008. (Proceedings of SPIE, Vol. 6866); (Progress in biomedical optics and imaging, Vol. 9, No. 25).
- Osmon DR, Berbari EF, Berendt AR, Lew D, Zimmerli W, Steckelberg JM, et al. Executive summary: diagnosis and management of prosthetic joint infection: clinical practice guidelines by the Infectious Diseases Society of America. *Clin Infect Dis*. 2013;56:1-10.

- Praveen NC, Rajesh A, Madan M, Chaurasia VR, Hiremath NV, Sharma AM. *In vitro* evaluation of antibacterial efficacy of pineapple extract (bromelain) on periodontal pathogens. J Int Oral Health. 2014;6: 96.
- Rastogi L, Kora AJ, Arunachalam J. Highly stable, protein capped gold nanoparticles as effective drug delivery vehicles for amino-glycosidic antibiotics. Mater Sci Eng C. 2012;32:1571-7.
- Rowan AD, Buttle DJ, Barrett AJ. The cysteine proteinases of the pineapple plant. Biochem J. 1990;266: 869-75.
- Selvaraj V, Alagar M. Analytical detection and biological assay of antileukemic drug 5-fluorouracil using gold nanoparticles as probe. Int J Pharm. 2007; 337:275-81.
- Stenger S, Hanson DA, Teitelbaum R, Dewan P, Niazi KR, Froelich CJ, et al. An antimicrobial activity of cytolytic T cells mediated by granulysin. Science. 1998;282:121-5.
- Taussig SJ, Batkin S. Bromelain, the enzyme complex of pineapple (*Ananas comosus*) and its clinical application - an update. J Ethnopharmacol. 1988;22:191-203.
- Timkovich R. Detection of the stable addition of carbodiimide to proteins. Anal Biochem. 1977;79:135-43.