

Review

Open Access

6th International Symposium on Retroviral Nucleocapsid

Ben Berkhout¹, Robert Gorelick², Michael F Summers³, Yves Mély⁴ and Jean-Luc Darlix*⁵

Address: ¹Laboratory of Experimental Virology, Department of Medical Microbiology, Center for Infection and Immunity Amsterdam (CINIMA) Academic Medical Center of the University of Amsterdam K3-110, Meibergdreef 15, 1105 AZ Amsterdam, The Netherlands, ²AIDS Vaccine Program SAIC-Frederick, Inc. NCI-Frederick P.O. Box B Frederick, MD 21702-1201, USA, ³Department of Chemistry and Biochemistry and Howard Hughes Medical Institute, University of Maryland Baltimore County, 1000 Hilltop Circle, Baltimore, MD 21250, USA, ⁴Département Pharmacologie et Physico-chimie, UMR 7175 CNRS, Institut Gilbert Laustriat, Université Louis Pasteur, 74 route du Rhin, 67401 Illkirch, France and ⁵LaboRetro INSERM #758, Ecole Normale Supérieure de Lyon, IFR 128 Biosciences Lyon-Gerland, 69364 Lyon Cedex 07, France

Email: Ben Berkhout - b.berkhout@amc.uva.nl; Robert Gorelick - gorelick@ncifcrf.gov; Michael F Summers - summers@hhmi.umbc.edu; Yves Mély - yves.mely@pharma.u-strasbg.fr; Jean-Luc Darlix* - jldarlix@ens-lyon.fr

* Corresponding author

Published: 25 February 2008

Received: 18 December 2007

Retrovirology 2008, 5:21 doi:10.1186/1742-4690-5-21

Accepted: 25 February 2008

This article is available from: <http://www.retrovirology.com/content/5/1/21>

© 2008 Berkhout et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Retroviruses and LTR-retrotransposons are widespread in all living organisms and, in some instances such as for HIV, can be a serious threat to the human health. The retroviral nucleocapsid is the inner structure of the virus where several hundred nucleocapsid protein (NC) molecules coat the dimeric, genomic RNA. During the past twenty years, NC was found to play multiple roles in the viral life cycle (Fig. 1), notably during the copying of the genomic RNA into the proviral DNA by viral reverse transcriptase and integrase, and is therefore considered to be a prime target for anti-HIV therapy. The 6th NC symposium was held in the beautiful city of Amsterdam, the Netherlands, on the 20th and 21st of September 2007. All aspects of NC biology, from structure to function and to anti-HIV vaccination, were covered during this meeting.

Overview

In 1998, Larry O. Arthur and Louis E. Henderson of NCI-Frederick decided that the field of nucleocapsid (NC) research was at a stage in which an NC symposium would be very useful. There was a general realization that NC was central to many processes in retrovirus replication and the highly conserved NC Zn²⁺-fingers were necessary for genome packaging and early infection processes (Fig. 1). Drs Arthur and Henderson hosted the first symposium at NCI-Frederick, MD, USA, (June 1998) and termed it International Retroviral Nucleocapsid Symposium (IRNCS) to be sure to include all the significant work being conducted on NC throughout the world. A series of meetings have been held since the inception of the initial symposium, namely the 2nd IRNCS in September 1999 (J-L Darlix, ENS

Lyon, France), 3rd IRNCS in October 2001 (L. O. Arthur, Loews Annapolis, MD, USA), 4th IRNCS in September 2003 (Y. Mély, Faculté de Pharmacie, Strasbourg, France), and 5th IRNCS in September 2005 (L. Kleiman, McGill University, Montreal, Quebec, Canada). The 6th IRNCS was held in Amsterdam to discuss the most recent advances on the functions of the NC protein in the synthesis, maintenance and integration of the proviral DNA, and in virus particle assembly. All these topics have been covered at the meeting to gain a better understanding of the multifunctional nature of NC (Fig. 1). Moreover, recent findings on other viral and cellular proteins playing a role in the viral life cycle that are either associated with NC, or resemble it in structure or function, such as the antiviral

NC is Required for Many Replication Steps

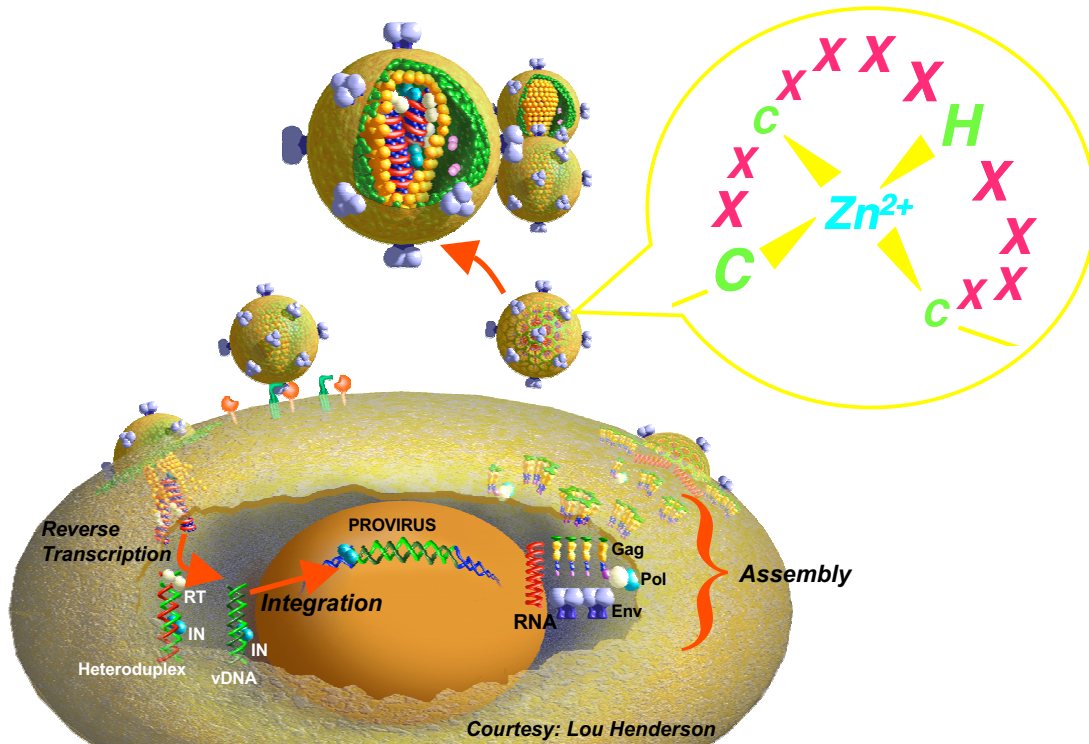


Figure 1

NC protein is required for many replication steps. Mature NC molecules coat the genomic RNA, in a dimeric form, in the viral particle and thus exert an essential function in virion structure. As illustrated in this cartoon – courtesy of Dr. Lou Henderson (NCI-Frederick, MD, USA) – NC protein chaperones conversion of the genomic RNA into the viral DNA by the RT enzyme. NC is also involved in the maintenance of the newly made viral DNA and in its integration into the host genome to form the provirus. In the course of virus assembly, the NC domain of Gag pilots genomic RNA selection which in turn acts as a scaffold for Gag oligomerization and assembly. As highlighted in the top circle, the CCHC zinc finger is central to all the NC-mediated functions. Thus, the retroviral zinc finger motif is viewed as a major target in anti-HIV drug development.

APOBEC3G protein, have been discussed at the conference.

NC structure and its relationship to function

This first session on NC structure-function relationships discussed the basis of NC recognition of the genomic RNA and its potent nucleic acid chaperoning activities, discovered in the late 80's [1-6] (for reviews see [7-9]). Using single DNA molecules that are stretched and melted by force, M. Williams (Northeastern University, Boston, MA, USA) and collaborators examined the influence of HIV and HTLV-1 NC on double stranded DNA destabilization and reannealing. It was found that HIV NC behaved as a pure chaperone while HTLV-1 NC was more akin a single stranded DNA binding protein [10-12] in agreement with the recent findings of K. Musier-Forsyth (OSU, Columbus,

OH, USA; see below). Based on standard assays to monitor the NC nucleic acid binding, aggregation and annealing activities, K. Musier-Forsyth and collaborators showed that the N-terminal 35 residues of HIV-1 NC were necessary and sufficient for chaperone function *in vitro*. Interestingly, the HIV-1 Gag polyprotein was also shown to be a nucleic acid chaperone protein, a property that is likely to facilitate genome dimerization and tRNA primer annealing during virion assembly. Results on different retroviral NC proteins showed that they exhibit significant differences in their overall chaperone activity, decreasing in the order HIV-1 ~RSV > MLV >> HTLV-1. Both K Musier-Forsyth and M Williams found that HTLV-1 NC's poor chaperone activity was caused by its acidic C-terminal domain. Using fluorescence-based techniques and the HIV-1 TAR stem-loop (SL) structure, Y Mély and his col-

leagues (Faculté de Pharmacie, Strasbourg, France) showed that the HIV-1 NC structural determinants, formed by the hydrophobic plateau at the top of the two zinc fingers and the flanking basic residues, are essential for its chaperoning function [13-15]. Similar structural determinants were also found to be essential for the chaperoning function of MoMLV NC that contains a unique zinc finger that is also flanked by basic residues [16].

Retroviral RNA structures and functions, and RNA-NC interactions

Several retroviral RNA structures, such as the 5' untranslated region (5'UTR) or leader, are the subject of intense interest because they are involved in the early and late phases of virus replication via multiple RNA-protein interactions, notably with NC. During retrovirus assembly NC, as the C-terminal domain of Gag, plays an essential role in specifically recruiting two copies of the full length genomic RNA and causing its dimerization. Deciphering the 3D structure of the Psi RNA packaging signal in the 5' UTR is key to our understanding of NC-genomic RNA interactions. To that end the group of M. Summers (UMBC, Baltimore, MD, USA) is investing much effort to establish the first high resolution model of the 100 nucleotide core encapsidation signal (Ψ^{CES}) of MoMuLV, which comprises stem loops (SL) B-D. Through a combination of nucleotide specific, segmental labeling, and sub-fragment analysis, they have obtained high quality and high resolution NMR (nuclear magnetic resonance) spectra indicative of dimer formation. In particular A289 and A293, which are part of the hairpin loop in SLB, give rise to signature peaks and nuclear Overhauser effect (NOE) patterns upon RNA dimer formation. Residues A330 and A364, which are part of SLC and SLD, respectively, give rise to downfield peaks diagnostic of kissing interactions between SLC and SLD. Additionally, using a novel transverse relaxation-optimized spectroscopy (TROSY) based heteronuclear single quantum correlation (HSQC) pulse sequence, residual dipolar couplings have been measured for isolated B-duplexes and this method is currently being applied to the 198-nucleotide dimeric (Ψ^{CES}) site. This work will lead to the first high resolution model of the dimeric Ψ^{CES} site. This group has also analysed interactions between the NC proteins and RNA packaging signals of other retroviruses, including MoMLV and Rous Sarcoma Virus (RSV). It was found that the native MoMLV Ψ allowed an average of 15 NCs to bind while mutant- Ψ , which can not form dimer, allowed only an average of two NC molecules to bind. These results show that exposure of NC binding sites by Ψ dimerization occurs in the entire Ψ -site, by which NC recognizes, selectively dimerizes, and packages its genomic RNA into the virion [17-19]. RSV is unusual in that its genome can be efficiently packaged by a relatively short, 82-nucleotide segment of the 5'-UTR called muPsi. Upon NC binding, muPsi adopts a stable

secondary structure that consists of three stem loops (SL-A, SL-B and SL-C) and an 8-base pair stem (O3), (see Figure 2) [20].

In the forefront of developing new technologies for high-throughput and accurate RNA secondary structure analysis, K Weeks (UNC Chapel Hill, NC, USA) reported the first generation of these technologies which involves Selective 2'-Hydroxyl Acylation analyzed by Primer Extension (called SHAPE) [21-24]. Because SHAPE makes it possible to obtain RNA secondary and tertiary structure information for hundreds or thousands of nucleotides over a few days or weeks, it is now becoming possible to tackle several ambitious problems involving the role of RNA structure in retroviral biology. Because of its completeness, high-throughput SHAPE analysis provides information sufficient to discriminate between otherwise contradictory models for HIV-1 genome structure. SHAPE analysis, performed *in virio*, allowed a series of previously unrecognized specific NC binding sites to be identified. *In virio* analysis also facilitated detailed characterizations of both the specific RNA binding and the contrasting duplex

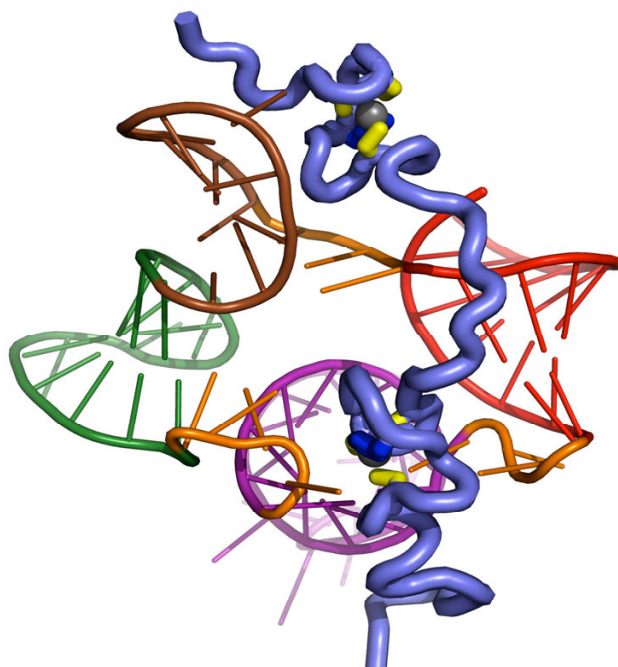


Figure 2
Structure of RSV NCp12 bound to the muPsi packaging signal. Structure of the Rous sarcoma virus nucleocapsid protein NCp12 bound to the cognate muPsi RNA packaging signal. The backbone of NC is shown as a ribbon (slate) and the zinc, cysteine and histidine groups are colored silver, yellow and blue, respectively. Coloring of secondary structural elements of the muPsi RNA are: SLA, purple; SLB, green; SLC, brown; O3, red; linker segments, orange (20).

destabilizing activity of NC protein. Further application of high-throughput SHAPE has significant potential to help establish the underlying connections between RNA structure and transcriptional and translational regulation and with other RNA-based processes in retroviral infectivity.

K. Purzycka and colleagues (Polish Academy of Sciences, Poznan, Poland) pursued their structural and dynamics analyses of the HIV-2 5' UTR RNA. They presented a new structure model for the DIS (dimer initiation site) of HIV-2 based on the high-throughput prediction of 3D RNA structures at low resolution [25,26] and molecular dynamic simulations [27].

Using functional assays A Das, M. Vrolijk and colleagues (B Berkhout, Amsterdam, the Netherlands) reinvestigated the role of the highly conserved TAR SL structure in HIV-1 replication [28]. They concluded that TAR has no essential function in HIV-1 biology other than to accommodate Tat-mediated activation of transcription [29]. But non-basepaired nucleotides at the 5' end of the 5' UTR can have adverse effects on its structure and functions in RNA dimerization and packaging mediated by the packaging signal (Ψ or Ψ). They concluded that HIV-1 requires a stable SL structure at the start of the viral transcripts to avoid misfolding of the leader RNA so that it can fulfill all its functions.

A. Lever and colleagues (University of Cambridge, UK) have long since been interested in the mechanism of HIV-1 genomic RNA selection and packaging during Gag assembly, that relies on the 5' Ψ packaging signal formed of SL structures [30]. The tip of SL1 is a dimer linkage site that has a purine-rich bulge proximal to it. This and a proximal bulge in SL3 appear to be metastable, probably facilitating RNA unwinding when Gag protein binds. They have identified another purine rich bulge in the SL1 stem loop which binds to the regulatory Rev protein. Disrupting Rev binding impairs virus replication. Some of the variant models in the structure of the Ψ RNA reflect the fact that the RNA undergoes significant conformational changes during its trafficking through the cell to the viral particle [31].

Role of NC in reverse transcription and recombination

NC is an obligatory constituent of the viral replication machinery whereby the genomic RNA is converted into the full-length double-stranded viral DNA by RT. This appears to be mediated by tight interactions between NC molecules, the genomic RNA that is in a dimeric form and RT. In a long standing effort to understand the role of NC in viral DNA synthesis, R Gorelick et al. (NCI-Frederick, MD, USA) [4] discussed their investigation of two HIV NC zinc-finger mutants, H23C and H44C, which exhibit

severe replication defects [32]. Analyses of early infection revealed that these mutations cause major defects in integration, but not reverse transcription: curiously, the rate of initiation and partial completion of reverse transcription was faster than wild-type [33]. Examination of virus particles prior to infection revealed that both mutant virions contain significantly more viral DNA than wild-type particles. Thus these NC mutations cause premature reverse transcription, and this may provide a partial explanation for their replication defects.

J. Mak and colleagues (Macfarlane Burnet Centre, Victoria, Australia) investigated the rate of HIV-1 recombination in cell cultures by monitoring sequence-tag redistribution in the *gag* and *pol* genes. They found that rates of recombination were high, corresponding to about 6–7 per cycle in T cells and up to 12–14 in monocyte-derived macrophages.

In addition to the genomic RNA, HIV-1 virions can package a substantial amount of spliced viral RNAs, but this seems to be independent of the NC zinc fingers, as reported by M. Mougel et al. (CNRS, Montpellier, France). Reverse transcription of these spliced RNAs takes place as efficiently as that of the genomic RNA, but poorly responds to the chain terminator antiviral drug AZT. Thus, AZT treatment might well increase the representativeness of spliced HIV-1 DNAs [34,35].

All viral RNA, the full length and the spliced forms, contain the TAR sequence at the 5' and 3' ends. NC can induce TAR dimerization as reported by E. Andersen et al (University of Aarhus, Denmark) [36]. According to these authors, TAR dimerization is important for the obligatory first strand transfer during cDNA synthesis and this TAR dimer has a parallel orientation depending, at least in part, on the dimerization initiation site (DIS). Completion of viral DNA synthesis necessitates the second strand transfer which takes place at the level of the PBS sequence. N Morellet et al (CNRS Paris, France) investigated this strand transfer in HIV-1 by means of NMR and fluorescence studies. They found that NC, notably the zinc fingers, chaperones this reaction by modifying the conformation of the loops of both PBS (+) and (-) sequences, promoting the formation of a kissing complex and the subsequent annealing of the PBS (+) and (-) sequences.

Gag assembly and the role of NC

The genomic RNA codes for the Gag and Gag-Pol polyprotein precursors which are synthesized by the host translation machinery. Several characteristics of the HIV-1 5' UTR, such as the 5' terminal TAR hairpin, its length and overall secondary structure, are likely to interfere with ribosomal scanning and suggest that translation is initiated by internal entry of ribosomes (IRES) [37,38]. To

gain further insight in the mechanism of translation initiation on the HIV-1 5'UTR, T. Abbink, K. Arts and B. Berkhout (Amsterdam, the Netherlands) introduced upstream AUGs (uAUG) at different positions in the 5'UTR and determined the effect on the expression of a downstream reporter gene under control of the gag AUG. If ribosomal scanning initiates upstream of the uAUG, translation will start on the uAUG, thereby inhibiting the expression of the downstream reporter gene. This allowed determination of the window of ribosomal scanning on the HIV-1 5' UTR. Their studies show that the inserted uAUG inhibited reporter gene expression at every position in the 5'UTR, thus indicating that the entire 5' UTR is scanned. In addition, they show that reinitiation of translation on the HIV-1 5'UTR is quite efficient if the upstream open reading frame is short (less than 20 bases). This finding is in agreement with previous findings [39] but different from others showing that HIV Gag translation operates by an IRES mechanism [40].

HIV-1 Gag polyprotein contains all of the molecular determinants required for its intracellular trafficking, and its assembly and budding in the form of virus-like particles (VLPs). The role of NC in Gag assembly has been studied for almost twenty years [1-5]. A. Rein and his colleagues (NCI-Frederick, MD, USA) have characterized the assembly properties of chimeric proteins in which the NC (and p1 and p6) domain of HIV-1 Gag is replaced by dimerizing or trimerizing leucine- or isoleucine-zipper motifs, respectively. These proteins assemble in mammalian cells into virus-like particles (VLPs) with morphology nearly, but not quite, identical to those formed by wild-type Gag. These VLPs contain at least 10-fold less RNA than wild-type VLPs. Their buoyant density is somewhat lower than that of wild-type VLPs; this difference is consistent with the hypothesis that the packing of the chimeric proteins in these VLPs is the same as that of wild-type Gag. The chimeric proteins co-assemble efficiently with wild-type Gag; thus the function of RNA in normal VLP assembly is not to act as a "string" upon which Gag protein molecules are aligned. The chimeric proteins were expressed in bacteria. They did not assemble spontaneously, but could be induced to assemble into VLPs by addition of either RNA or inositol hexakisphosphate.

D. Muriaux, J.L. Darlix and collaborators (INSERM Lyon, France) have been studying the trafficking and the assembly of HIV-1 in human cells. Viral and cellular factors are involved in these processes, in particular the NC domain of Gag, the genomic RNA, and cellular proteins of the endocytic pathways, and membrane microdomains [41]. They found that HIV-1 can bud into intracellular vesicles in addition to the plasma membrane of T cells [42]. The NC zinc fingers of HIV-1 Gag were found to be critical determinants of Gag assembly and localisation in endo-

somes [43]. Furthermore, they reported the involvement of a tetraspanin web in T-lymphoblastic cells that serves as a platform for HIV-1 assembly (Grigorov B., V. Attuil-Audenis, F. Perugi, M. Nedelec, S. Watson, J.-L. Darlix, C. Pique, H. Conjeaud & D. Muriaux: IMPLICATIONS OF TETRASPANINS IN HIV-1 FORMATION in infected T-lymphoblastic cells., submitted).

NC protein can exist in different forms in newly made mature HIV-1 virions, namely NCp15 (NC-SP2-p6), NCp9 (NCp7-SP2) and fully processed NCp7. J.A. Thomas (NCI-Frederick, MD, USA) presented work of collaborators where they investigated the requirement for proteolytic maturation of NC by mutating the protease cleavage sites necessary for the production of mature NCp9 (NCp7-SP2) or NCp7. Interestingly, viruses tailored to make either NCp9 + p6 or NCp7 + SP2 + p6 were fully infectious and could replicate in H9 cells. In contrast, viruses that made either NCp15 or NCp7 + p1-p6 were replication defective. Because the replication block was principally manifested by a severe reduction in integration, it is likely that NCp9 has a role in the integration process [44] and has also been shown to facilitate *in vitro* concerted integration even better than NCp7 [45].

Cellular and viral proteins associated with NC

Human APOBEC3G (hA3G) has been identified as an anti-HIV-1 host factor acting by deaminating the newly made cDNA. S. Cen and collaborators (Lady Davis Inst., Montreal, Canada) reported that hA3G inhibits HIV-1 reverse transcription independently of its editing activity. A reduction of 55% in early viral DNA synthesis by hA3G is correlated with a similar decrease in the tRNA^{Lys3} priming, which requires a hA3G/NCp7 interaction. A greater reduction of ~95% in late DNA synthesis results from the hA3G-induced inhibition of DNA strand transfer, which is correlated with its ability to prevent RNaseH degradation of the template RNA [46-48].

Y. Iwatani et al (NIH, Bethesda, MD, USA) have independently investigated possible effects of hA3G on RT and NC function *in vitro*. NC-mediated annealing and RNase H cleavage were not affected, but A3G inhibited all RT-catalyzed elongation reactions, independent of hA3G's catalytic activity. These data taken together with complementary biophysical analyses led to the conclusion that deaminase-independent inhibition of reverse transcription is determined by critical differences in the nucleic acid binding properties of NC, A3G, and RT [49,51].

In an attempt to understand the possible interactions between HIV-1 NC and hA3G, P Henry et al (NCI, Frederick, MD, USA) expressed hA3G in insect cells and purified it using metal affinity, ion exchange and size exclusion

chromatography. Immunoprecipitation of purified NC with purified A3G bound to magnetic beads in the presence and absence of RNA revealed that A3G pulls down NC more efficiently in the presence of RNA, with decreasing efficiency as excess RNA was added to dilute simultaneous binding. The effect of NC on A3G enzymatic activity was examined by adding purified NC to their scintillation proximity assay mixture at a ratio of 1 NC molecule per 2, 4 or 8 nucleotides of substrate DNA, either before or with the addition of A3G [50]. In all cases, NC was unable to reduce cytidine deamination of the substrate DNA. These data indicate that interactions between NC and A3G are indirect and mediated by an RNA bridge. In addition, the results suggest that concentrations of NC that are likely to be present in the reverse transcription complex do not interfere with A3G's cytidine deaminase activity.

J.C. Paillart (CNRS, Strasbourg, France) reported data on a putative new role of the viral factor Vif. Vif is a small basic protein essential to viral fitness and pathogenicity. Marquet, Paillart and coworkers recently showed that Vif preferentially binds to the 5' UTR of the HIV-1 genomic RNA [52] and to corresponding DNA sequences [53]. Now they report that Vif stimulates the early steps of reverse transcription (tRNA^{Lys3} annealing, strong-stop DNA synthesis, first strand transfer) and the formation of loose RNA dimers, suggesting that Vif is an RNA chaperone [54]. However, Vif is able to inhibit NC-induced reactions such as hybridization of tRNA^{Lys3} and formation of tight RNA dimers while it collaborates with NC to increase RT processivity. Thus, Vif might negatively regulate NC-assisted maturation of the RNA dimer and early steps of reverse transcription during assembly, but these inhibitory effects would be relieved after viral budding, due to the preferential exclusion of Vif from virions.

Antivirals and NC vaccines

In this session new anti-viral drugs targeting HIV-1 protease and NC were described. To combat the widespread development of HIV-1 protease resistance in AIDS patients, novel protease inhibitors (PI) with high potency against the known PI-resistant mutants have been developed. M. Nijhuis and collaborators (University of Utrecht, the Netherlands) investigated whether HIV-1 could yet again find a way to become less susceptible to these novel drugs [55]. *Ex vivo* selection experiments have shown that HIV-1 indeed can use an alternative mechanism by the selection of mutations in the NC/SP2 cleavage site instead of the viral protease. These changes cause resistance to all protease inhibitors by enhancing the processing efficiency of the altered substrate by wild type protease. Modulation of the cleavage efficiency of this site impacts both PI susceptibility and viral replication capacity. Further studies are required to determine to what extent NC/P2 cleavage

site mutations may explain virological failure during PI therapy.

Y. Mély (ULP, Strasbourg, France) in collaboration with J.L. Darlix (INSERM Lyon, France) and M. Gottikh (Moscow, Russia) developed a screening assay based on the ability of HIV-1 NCp7 to destabilize SL structures such as TAR (Fig. 3). Several lead molecules were identified that inhibit NC destabilization activity in the low μM range. These molecules bind to the NC zinc fingers but do not eject zinc. Small Trp-rich hexapeptides selected from phage display libraries can also inhibit the NCp7 destabilizing activity in the low μM range through competition with its target sequences [56-58]. Finally, peptide and oligonucleotide-based compounds targeting NCp7 with low nM activities have been found and they are currently being characterized *in vitro* and in cell culture assays.

P. Mahy from H-PHAR (Gosselies, Belgium) reported new results on HPH116, which is a zinc finger ejector targeting NCp7 protein. HPH116 is a modified synthetic form of azodicarbonamide produced under good manufacturing practices (GMP) conditions developed by H-PHAR. In a TZM-bl cell infection model, HPH116-treated viral particles were found to have lost their infectivity, with up to 100% inhibition at 1 μM HPH116. Viral inhibition was enhanced when the viral particles were pre-incubated with HPH116 at 0.25 μM and 0.1 μM Tenofovir (vs 5 μM Tenofovir alone), 0.3 μM AZT (vs 30 μM AZT alone), or

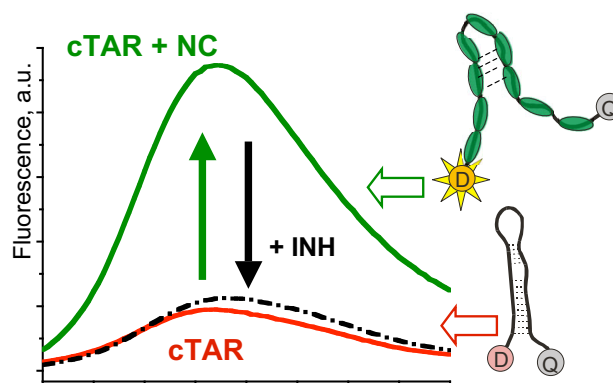


Figure 3
Principle of the NC inhibitor screening assay. This assay is based on the use of a HIV-1 TAR DNA (cTAR) sequence labeled at its 5' and 3' ends, by a fluorophore (D) and a quencher (Q), respectively. In the absence of NC, Q is close to D, leading to a small residual fluorescence of D. Due to its nucleic acid destabilizing properties, NC melts the cTAR structure up to the middle of the stem, leading Q to move away from D, thus causing a large fluorescence increase. A positive hit is detected through the restoration of the initial low fluorescence level.

with 0.125 μ M T20. Thus, when HPH116 is combined with other known antiretroviral drugs, it can induce a dramatic increase in virus inhibition.

S-acyl-2-mercaptobenzamide thioester lead compounds [59] were investigated as topical microbicides by Miller-Jenkins et al. Initial studies with C. Cheng-Mayer (Aaron Diamond AIDS Research Center, New York, USA) have shown that a lead compound protects macaques from SHIV infection in a dual-infection model of vaginal transmission. Three sites in NCp7 were found to be important for reactivity and specificity: position x+1 (where x is Cys36), x+2, and x+9 [60,61]. Furthermore, a secondary intramolecular S to N acyl transfer, which occurs after the primary intermolecular acyl transfer from the thioester to Cys36, was demonstrated. This secondary transfer is irreversible and leads to loss of NCp7 structure and, most probably, its function.

An update on work involving covalent modification of free S-H groups on internal viral proteins by electrophilic reagents to generate whole inactivated retroviral virions with functional envelope glycoproteins was presented by J. Lifson (NCI-Frederick, MD, USA). Studies with 4-vinyl pyridine showed that covalent modification of NC, without appreciable modification of other virion proteins is sufficient for inactivation. Inactivated viruses are being used for electron tomographic ultrastructural studies of whole virions and envelope glycoprotein spikes [62]. In additions tomographic studies were presented showing the interaction of Env with receptors on T cells [63]. *In vitro*, inactivated virions effectively primed both HIV-1 specific responses by MHC-I restricted responses by naïve CD8+ T cells and MHC-II restricted responses by naïve CD4+ T cells. Progress on the use of such particles as a vaccine immunogen was also presented.

Authors' contributions

BB organized the symposium. RG and JLD were co-organizers. MFS and YM provided figures 2 and 3, respectively. JLD wrote the report. BB, RG, MFS and YM corrected the report.

Acknowledgements

The 6th international Retroviral NC symposium was supported by the Office of AIDS RESEARCH (OAR, USA), SAIC FREDERICK (NCI, USA), the ROYAL DUTCH ACADEMY OF SCIENCES (The Netherlands), The Center for Infection and Immunity Amsterdam (CINIMA, the Netherlands), the city of Amsterdam (the Netherlands) the Dutch AIDS Fund (The Netherlands), ANRS (Paris, France), Clean Air Europe, Greiner bio-one Europe, SANYO Europe. Projects have been funded in whole or in part with federal funds from the National Cancer Institute, National Institutes of Health, under contract N01-CO-12400. The content of this publication does not necessarily reflect the views or policies of the Department of Health and Human Services, nor does mention of trade names, commercial products, or organizations imply endorsement by the U.S. Government.

References

- Méric C, Darlix JL, Spahr PF: **It is Rous sarcoma virus protein P12 and not P19 that binds tightly to Rous sarcoma virus RNA.** *J Mol Biol* 1984, **173(4)**:531-538.
- Méric C, Goff SP: **Characterization of Moloney murine leukemia virus mutants with single-amino-acid substitutions in the Cys-His box of the nucleocapsid protein.** *J Virol* 1989, **63(4)**:1558-1568.
- Méric C, Spahr PF: **Rous sarcoma virus nucleic acid-binding protein p12 is necessary for viral 70S RNA dimer formation and packaging.** *J Virol* 1986, **60(2)**:450-459.
- Gorelick RJ, Henderson LE, Hanser JP, Rein A: **Point mutants of Moloney murine leukemia virus that fail to package viral RNA: evidence for specific RNA recognition by a "zinc finger-like" protein sequence.** *Proc Natl Acad Sci USA* 1988, **85(22)**:8420-8424.
- Prats AC, Sarih L, Gabus C, Litvak S, Keith G, Darlix JL: **Small finger protein of avian and murine retroviruses has nucleic acid annealing activity and positions the replication primer tRNA onto genomic RNA.** *EMBO J* 1988, **7(6)**:1777-1783.
- Barat C, Lullien V, Schatz O, Keith G, Nugeyre MT, Grüninger-Leitch F, Barré-Sinoussi F, LeGrice SF, Darlix JL: **HIV-1 reverse transcriptase specifically interacts with the anticodon domain of its cognate primer tRNA.** *EMBO J* 1989, **8(11)**:3279-3285.
- Rein A, Henderson LE, Levin JG: **Nucleic-acid-chaperone activity of retroviral nucleocapsid proteins: significance for viral replication.** *Trends Biochem Sci* 1998, **23(8)**:297-301.
- Levin JG, Guo J, Rouzina I, Musier-Forsyth K: **Nucleic acid chaperone activity of HIV-1 nucleocapsid protein: critical role in reverse transcription and molecular mechanism.** *Prog Nucleic Acid Res Mol Biol* 2005, **80**:217-86.
- Darlix JL, Lapadat-Tapolksy M, de Rocquigny H, Roques BP: **First glimpses at structure-function relationships of the nucleocapsid protein of retroviruses.** *J Mol Biol* **254(4)**:523-537. 1995 Dec 8;
- McCauley MJ, Williams MC: **Mechanisms of DNA binding determined in optical tweezers experiments.** *Biopolymers* 2007, **85**:154-168.
- Cruceanu M, Gorelick RJ, Musier-Forsyth K, Rouzina I, Williams MC: **Rapid kinetics of protein-nucleic acid interaction is a major component of HIV-1 nucleocapsid protein's nucleic acid chaperone function.** *J Mol Biol* 2006, **363**:867-877.
- Pant K, Karpel RL, Rouzina I, Williams MC: **Salt dependent binding of T4 gene 32 protein to single and double-stranded DNA: single molecule force spectroscopy measurements.** *J Mol Biol* 2005, **349**:317-330.
- Morellet N, Jullian N, De Rocquigny H, Maigret B, Darlix JL, Roques BP: **Determination of the structure of the nucleocapsid protein NCp7 from the human immunodeficiency virus type 1 by IH NMR.** *Embo J* 1992, **11(8)**:3059-3065.
- Summers MF, Henderson LE, Chance MR, Bess JW Jr, South TL, Blake PR, Sagi I, Perez-Alvarado G, Sowder RC 3rd, Hare DR, et al.: **Nucleocapsid zinc fingers detected in retroviruses: EXAFS studies of intact viruses and the solution-state structure of the nucleocapsid protein from HIV-1.** *Protein Sci* 1992, **1(5)**:563-574.
- Beltz H, Clauss C, Piemont E, Ficheux D, Gorelick RJ, Roques B, Gabus C, Darlix JL, de Rocquigny H, Mely Y: **Structural determinants of HIV-1 nucleocapsid protein for cTAR DNA binding and destabilization, and correlation with inhibition of self-primed DNA synthesis.** *J Mol Biol* 2005, **348(5)**:1113-1126.
- Egelé C, Piémont E, Didier P, Ficheux D, Roques BP, Darlix JL, de Rocquigny H, Mély Y: **The single-finger nucleocapsid protein of Moloney Murine Leukaemia Virus binds and destabilizes the TAR sequences of HIV-1 but does not promote efficiently their annealing.** *Biochemistry* 2007, **46**:14650-14662.
- Dey A, York D, Smalls-Mantey A, Summers M: **Composition and Sequence-dependent Binding of RNA to the Nucleocapsid Protein of Moloney Murine Leukemia Virus.** *Biochemistry* 2005, **44**:3735-3744.
- D'Souza , Anwasha D, Habib D, Summers M: **NMR Structure of the 101-nucleotide Core Encapsulation Signal of the Moloney Murine Leukemia Virus.** *J Mol Biol* 2004, **337**:427-442.
- D'Souza V, Summers M: **Structural basis of packaging dimeric genome of Moloney murine leukemia virus.** *Nature* 2004, **431**:586-590.

20. Zhou J, Bean RL, Vogt VM, Summers M: **Solution structure of the Rous sarcoma virus nucleocapsid protein: muPsi RNA packaging signal complex.** *J Mol Biol* 2007, **365**(2):453-67.
21. Merino EJ, Wilkinson KA, Coughlan JL, Weeks KM: **RNA structure analysis at single nucleotide resolution by Selective 2'-Hydroxyl Acylation and Primer Extension (SHAPE).** *J Am Chem Soc* 2005, **127**:4223-4231.
22. Badorrek CS, Weeks KM: **Architecture of a gamma retroviral genomic RNA dimer.** *Biochemistry* 2006, **45**:12664-12672.
23. Wilkinson KA, Merino EJ, Weeks KM: **Selective 2'-hydroxyl acylation analyzed by primer extension (SHAPE): quantitative RNA structure analysis at single nucleotide resolution.** *Nature Protocols* 2006, **1**:1610-1616.
24. Mortimer SA, Weeks KM: **A fast acting reagent for accurate analysis of RNA secondary and tertiary structure by SHAPE chemistry.** *J Am Chem Soc* 2007, **129**:4144-4145.
25. Pachulska-Wieczorek K, Purzycka KJ, Adamiak RW: **New, extended hairpin form of the TAR-2 RNA domain points to the structural polymorphism at the 5'- end of the HIV-2 leader RNA.** *Nucleic Acids Res* 2006, **34**:2984-2997.
26. Popenda M, Bielecki , Adamiak RW: **High-throughput method for the prediction of low-resolution, three-dimensional RNA structures.** *Nucleic Acids Symposium Series* 2006, **50**:67-68.
27. Popenda M, Baewicz M, Szachniuk M, Adamiak RW: **RNA FRA-BASE version 1.0: an engine with a database to search for the three-dimensional fragments within RNA structures.** *Nucleic Acids Res* in press.
28. Berkhout B, Silverman RH, Jeang KT: **Tat trans-activates the human immunodeficiency virus through a nascent RNA target.** *Cell* 1989, **59**:273-282.
29. Das AT, Harwig A, Vrolijk MM, Berkhout B: **The TAR hairpin of human immunodeficiency virus type-I can be deleted when not required for Tat-mediated activation of transcription.** *Journal of Virology* 2007, **81**:7742-7748.
30. Lever AML: **HIV-1 RNA Packaging.** *Adv Pharmacol* 2007, **55**:1-32.
31. Greatorex JS, Palmer EA, Pomerantz RJ, Dangerfield J, Lever AML: **Mutation of the Rev binding loop in the HIV-1 leader causes a replication defect characterised by altered RNA trafficking and packaging.** *Journal of General Virology* 2006, **87**:3039-3044.
32. Gorelick RJ, Gagliardi TD, Bosche WJ, Wilttrout TA, Coren LV, Chabot DJ, Lifson JD, Henderson LE, Arthur LO: **Strict conservation of the retroviral nucleocapsid protein zinc finger is strongly influenced by its role in viral infection processes: characterization of HIV-1 particles containing mutant nucleocapsid zinc-coordinating sequences.** *Virology* 1999, **256**(1):92-104.
33. Thomas JA, Shulenin S, Coren LV, Bosche WJ, Gagliardi TD, Gorelick RJ, Oroszlan S: **Human immunodeficiency virus type I nucleocapsid zinc-finger mutations cause defects in reverse transcription and integration.** *Virology* 2006, **353**(1):41-51.
34. Houzet L, Smagulova F, Paillart JC, Maurel S, Morichaud Z, Marquet R, Mougél M: **HIV controls the selective packaging of genomic, spliced viral, and cellular RNAs into virions through different mechanisms.** *Nucleic Acids Res* 2007, **35**:2695-2704.
35. Houzet L, Morichaud Z, Mougél M, Morichaud Z: **Fully-spliced HIV-1 RNAs are reverse transcribed with similar efficiencies as the genomic RNA in virions and cells, but more efficiently in AZT-treated cells.** *Retrovirology* 2007, **4**:30.
36. Andersen ES, Contera SA, Knudsen B, Damgaard CK, Besenbacher F, Kjems J: **Role of the Trans-activation Response Element in Dimerization of HIV-1 RNA.** *J Biol Chem* 2004, **279**:22243-9.
37. Yilmaz A, Bolinger C, Boris-Lawrie K: **Retrovirus translation initiation: Issues and hypotheses derived from study of HIV-1.** *Current HIV Research* 2006, **4**:131-139.
38. Balvay L, Lopez Lastra M, Sargueil B, Darlix JL, Ohlmann T: **Translational control of retroviruses.** *Nat Rev Microbiol* 2007, **5**(2):128-140.
39. Luukkonen BG, Tan W, Schwartz S: **Efficiency of reinitiation of translation on human immunodeficiency virus type I mRNAs is determined by the length of the upstream open reading frame and by intercistronic distance.** *Journal Virology* 1995, **69**(7):4086-4094.
40. Brasey A, Lopez-Lastra M, Ohlmann T, Beerens N, Berkhout B, Darlix JL, Sonenberg N: **The leader of human immunodeficiency virus type I genomic RNA harbors an internal ribosome entry segment that is active during the G2/M phase of the cell cycle.** *J Virol* 2003, **77**(7):3939-49.
41. Muriaux D, Darlix JL, Cimarelli A: **Targeting HIV-1 budding.** *Curr Pharmacol Design* 2004, **10**:3725-39.
42. Grigorov B, Arcanger F, Roingard P, Darlix JL, Muriaux D: **Assembly of infectious HIV-1 in human epithelial and lymphoblastic cell lines.** *J Mol Biol* 2006, **359**:848-62.
43. Grigorov B, Décimo D, Smagulova F, Mougél M, Muriaux D, Darlix JL: **Intracellular HIV-1 Gag localization is impaired by mutations in the nucleocapsid zinc fingers.** *Retrovirology* 2007, **4**:54.
44. Coren LV, Thomas JA, Chertova E, Sowder RC 2nd, Gagliardi TD, Gorelick RJ, Ott DE: **Mutational analysis of the C-terminal gag cleavage sites in human immunodeficiency virus type I.** *J Virol* 2007, **81**(18):10047-54.
45. Gao K, Gorelick RJ, Johnson DG, Bushman F: **Cofactors for human immunodeficiency virus type I cDNA integration in vitro.** *J Virol* 2003, **77**(2):1598-1603.
46. Li X, Guo F, Zhang L, Kleiman L, Cen S: **Human APOBEC3G inhibits DNA strand transfer during reverse transcription of HIV-1.** *J Biol Chem* 2007, **282**(44):32065-32074.
47. Guo F, Cen S, Niu M, Yang Y, Gorelick RJ, Kleiman L: **The interaction of APOBEC3G with HIV-1 nucleocapsid inhibits tRNA^{Lys3} annealing to viral RNA.** *J Virol* 2007, **81**(20):11322-11331.
48. Guo F, Cen S, Niu M, Saadatmand J, Kleiman L: **The inhibition of tRNA^{Lys3}-primed reverse transcription by human APOBEC3G during HIV-1 replication.** *J Virol* 2006, **80**(23):11710-22.
49. Iwatani Y, Takeuchi H, Strebel K, Levin JG: *J Virol* 2006, **80**:5992-6002.
50. Svarovskaia ES, Xu H, Mbisa JL, Barr R, Gorelick RJ, Ono A, Freed EO, Hu WS, Pathak VK: **Human APOBEC3G is incorporated into HIV-1 virions through interactions with viral and nonviral RNAs.** *J Biol Chem* 2004, **279**(34):35822-35828.
51. Iwatani Y, Chan DSB, Wang F, Maynard KS, Sugiura W, Gronenborn AM, Rouzina I, Williams MC, Musier-Forsyth K, Levin JG: **Deaminase-independent inhibition of HIV-1 reverse transcription by APOBEC3G.** *Nucleic Acids Res* 2007, **35**(21):7096-7108.
52. Henriët S, Bernacchi S, Richer D, Decroly E, Vigne R, Ehresmann B, Ehresmann C, Paillart J-C, Marquet R: **Cooperative and specific binding of Vif to the 5' region of HIV-1 genomic RNA.** *J Mol Biol* 2005, **354**:55-72.
53. Bernacchi S, Henriët S, Dumas P, Paillart JC, Marquet R: **Characterization of the interaction between HIV-1 Vif protein and the viral genomic RNA: a fluorescence study.** *J Biol Chem* 2007, **282**:26361-26368.
54. Henriët S, Sinck L, Bec G, Gorelick RJ, Marquet R, Paillart J-C: **Vif is a RNA chaperone that could temporally regulate RNA dimerization and the early steps of HIV-1 reverse transcription.** *Nucleic Acids Res* 2007, **35**:5141-5153.
55. Nijhuis M, van Maarseveen NM, Lastere S, Schipper P, Coakley E, Glass B, Rovenska M, de Jong D, Chappay C, Goedegebuure IW, Heilek-Snyder G, Dulude D, Cammack N, Brakier-Gingras L, Konvalinka J, Parkin N, Kräusslich HG, Brun-Vezinet F, Boucher CA: **A novel substrate-based HIV-1 protease inhibitor drug resistance mechanism.** *PLoS Med* 2007, **4**(1):e36.
56. de Rocquigny H, Shvadchak V, Avilov S, Dong Chang Zhi, Dietrich U, Darlix J-L, Mély Y: **Targeting the viral nucleocapsid protein in anti-HIV-1 therapy.** *MiniRev Med Chem* 2008, **8**(1):24-35.
57. Raja C, Ferner J, Dietrich U, Avilov S, Ficheux D, Darlix JL, De Rocquigny H, Schwalbe H, Mely Y: **A Trp-rich hexapeptide inhibits nucleic acid destabilization chaperoned by the HIV-1 nucleocapsid protein.** *Biochemistry* 2006, **45**:9254-9265.
58. Dietz J, Koch J, Kaur A, Raja C, Stein S, Grez M, Pustowka A, Mensch S, Ferner J, Tempe R, Divita G, Mely Y, Schwalbe H, Dietrich U: **Antiviral activity of peptide ligands for the HIV-1 packaging signal Psiafter binding optimization on spot synthesis peptide membranes.** *Chem Med Chem* 2008 in press.
59. Srivastava P, Schito ML, Fattah RJ, Hara T, Hartman T, Buckheit RW, Turpin JA Jr, Inman JK, Appella E: **Optimization of unique, uncharged thioesters as inhibitors of HIV replication.** *Bioorg Med Chem* 2004, **12**(24):6437-50.
60. Miller Jenkins LM, Durell SR, Maynard AT, Stahl SJ, Inman JK, Appella E, Legault P, Omichinski JG: **Comparison of the Specificity of Interaction of Cellular and Viral Zinc-Binding Domains with 2-Mercaptobenzamide Thioesters.** *J Am Chem Soc* 2006, **128**:11964-76.

61. Miller Jenkins LM, Hara T, Durell SR, Hayashi R, Inman JK, Piquemal J-P, Gresh N, Appella E: **The Specificity of Acyl Transfer from 2-Mercaptobenzamide Thioesters to the HIV-1 Nucleocapsid Protein.** *J Am Chem Soc* 2007, **129**:11067-78.
62. Zhu P, Liu J, Bess J Jr, Chertova E, Lifson JD, Grise H, Ofek GA, Taylor KA, Roux KH: **Distribution and three-dimensional structure of AIDS virus envelope spikes.** *Nature* 2006, **441**:847-852.
63. Sougrat R, Bartesaghi A, Lifson JD, Bennett AE, Bess JW, et al.: **Electron tomography of the contact between T cells and SIV/HIV-1: implications for viral entry.** *PLoS pathogens* 2007, **3(5)**:e63.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

