Relation of oxidant-antioxidant imbalance with disease progression in patients with asthma

Asrar Ahmad^{1,2}, Mohammad Shameem², Qayyum Husain¹

Biochemistry, Faculty of Life Sciences, Aligarh Muslim University, Aligarh, ²Department of T.B. and Respiratory Diseases. Jawaharlal Nehru Medical College, Aligarh Muslim University, Aligarh, India

¹Department of

Address for

correspondence: Prof. Qayyum Husain, Department of Biochemistry, Faculty of Life Sciences, Aligarh Muslim University, Aligarh-202002, India. E-mail: qayyumbiochem@ gmail.com

> Submission: 16-03-12 Accepted: 02-06-12



Access this article online

Website: www.thoracicmedicine.org

DOI: 10.4103/1817-1737.102182

Abstract

CONTEXT: Asthma is a chronic airway disorder which is associated to the inflammatory cells. Inflammatory and immune cells generate more reactive oxygen species in patients suffering from asthma which leads to tissue injury.

AIMS: To investigate the role of oxidant-antioxidant imbalance in disease progression of asthmatic patients.

SETTINGS AND DESIGN: In this study, 130 asthmatic patients and 70 healthy controls were documented.

METHODS: For this malondialdehyde level, total protein carbonyls, sulfhydryls, activity of superoxide dismutase (SOD), catalase, glutathione peroxidase (GPx), total blood glutathione, and total antioxidant capacity (FRAP) were measured.

STATISTICAL ANALYSIS USED: Analysis of the data was done using unpaired student t test and one-way ANOVA analysis. P < 0.05 was considered significant.

RESULTS: The present work showed that the systemic levels of MDA (4.19 ± 0.10 nmol/ml, P < 0.001) and protein carbonyls (1.13 ± 0.02 nmol/mg, P < 0.001) were found to be remarkably higher in asthmatic patients while protein sulfhydryls ($0.55 \pm 0.01 \text{ mmol/l}$, P < 0.05) decreased as compared to controls ($2.84 \pm 0.12 \text{ nmol/m}$], $0.79 \pm 0.02 \text{ nmol/}$ mg and 0.60 ± 0.02 mmol/l, respectively). We also observed decrease in activities of SOD (2047 ± 50.34 U/g Hb, P < 0.05), catalase (4374 ± 67.98 U/g Hb, P < 0.01), and GPx (40.97 ± 1.05 U/g Hb, P < 0.01) in erythrocytes compared to control (2217 ± 60.11 U/g Hb, 4746 ± 89.94 U/g Hb, and 48.37 ± 2.47 U/g Hb, respectively). FRAP level (750.90 \pm 21.22 μ mol/l, P < 0.05) in plasma was decreased, whereas total blood glutathione increased $(0.94 \pm 0.02 \text{ mmol/l}, P < 0.05)$ as seen in control (840.40 ± 28.39 µmol/l and 0.84 ± 0.04 mmol/l).

CONCLUSIONS: This work supports and describes the hypothesis that an imbalance between oxidant-antioxidant is associated to the oxidative stress which plays a significant role in severity of the disease.

Key words:

Antioxidants, asthma, oxidants, oxidative stress, reactive oxygen species

sthma is the most frequent chronic airway ${
m A}$ disease and is one of the major global health problems^[1] which is characterized by chronic inflammation of the airways involving variable and recurrent airflow obstruction and increased airway responsiveness to a variety of stimuli.^[2,3] Recent research suggests that airway inflammation is the most proximate cause of the recurrent episodes of airflow limitation in asthma and is associated with increased oxidative stress.^[4] Thus, oxidative stress (oxidantantioxidant imbalance) has been increasingly recognized as a major factor contributing to the chronic inflammation. When oxidative stress overwhelms antioxidant mechanisms, reactive oxygen species (ROS) interact with proteins, lipids, and DNA leading to pathological consequences.^[5] Oxidative stress is thought to initiate airway inflammation; therefore, it is an important component of asthma pathogenesis.[6] Many evidences suggest that ROS produced both intracellularly by lung parenchymal cells and extracellularly by lung macrophages. It is

present in all forms of asthma including mild and asymptomatic cases and involves central as well as peripheral airways.[7,8]

Increased generation of oxidants have been reported in asthma patients than in healthy individuals,^[9,10] which aggravated airway inflammation by inducing diverse pro-inflammatory mediators including macrophages, neutrophils and eosinophils.[11,12] Numerous studies have suggested that oxidative stress is caused by overproduction of various free radicals or by an insufficient antioxidant defense system in asthma and thus it contributes to the tissue damage which is induced by inflammatory cells.^[9,13-16] These include a range of toxic ROS such as superoxide radical (O_2^{\bullet}) , hydrogen peroxide (H_2O_2) , hypochlorous acid (HOCl), and hydroxyl radical (OH[•]).^[11,14] In asthmatic patients, ROS play a vital role in pathophysiologic changes such as increased lipid peroxidation, airway responsiveness, production of chemoattractants, and vascular permeability.^[17] To counter the oxidant-mediated toxicity, lung

and blood cells are endowed with several antioxidants including glutathione peroxidase (GPx), superoxide dismutase (SOD), catalase, glutathione, α -tocopherol, and ascorbic acid.^[18,19] The increased oxidative stress in asthma is reflected by plasma level of malondialdehyde (MDA) and protein carbonyls in both bronchoalveolar lavage (BAL) fluid and peripheral blood samples;^[12,20] together with increased oxidized glutathione in BAL fluid.^[21] It has also been reported that markers of oxidative stress such as H₂O₂, 8-isoprostane, nitric oxide, and carbon monoxide were found in higher concentration in exhaled air of asthmatic patients.^[10,16,22] Changes in antioxidant defenses have noticed due to oxidative stress in asthmatic patients as decreased in GPx activity in whole blood, plasma, and platelets.^[23,24] Furthermore, protein sulfhydryls and total antioxidant capacity were also found to be decreased in plasma concomitant with increased SOD activity in BAL cells.^[25]

In the present study, the imbalance between oxidant and antioxidant level was determined in asthmatic patients which plays a key role in severity of the disease. Moreover, we have also correlated it with the degree of airflow obstruction (FEV1 % predicted) in patients with asthma.

Methods

Study population

This study was conducted in the Department of TB and Respiratory Diseases, Jawaharlal Nehru Medical College Hospital, Aligarh Muslim University, Aligarh, India, on patients attending outpatient department (OPD) and inpatient departments (IPD). We documented 130 (12-83 years of age) patients with bronchial asthma. Clinical severity of asthma was determined using criteria (appropriate clinical and respiratory function tests) defined in global initiative for asthma guidelines (GINA).^[26] The diagnosis was established on the basis of recurrent symptoms of breathlessness, chest tightness, and wheezing with an improvement of greater than 12% in forced expiratory volume in 1 second (FEV1) after inhalation of 200 µg of salbutamol from a nebulizer.

None of the patients were taking any antioxidant supplements and did not have any symptoms of lower or upper respiratory tract infection. Those patients who had pulmonary or systemic disease or an acute exacerbation of asthma within the past 4 weeks were in exclusion criterion. No drug was allowed on the day of testing. Seventy control subjects (10-75 years of age) were selected without respiratory symptoms and normal lung functions. All recruited subjects were nonsmokers and majority of the patients included in the present study were on treatment with inhaled corticosteroids. Written informed consent was obtained from each recruited subjects and the Institutional Ethics Committee of the Medical College approved the study.

Each patient's demographic profile, clinical, radiological findings, associated disease, pulmonary function measurement, history of allergy, and treatment details were recorded.

Evaluation of lung function

Pulmonary function tests were performed in each patient as well as in controls. Lung function parameters were measured by Easy on PC spirometer (ndd Medizintechnik AG, Zurich, Switzerland) and the best value from 3 manoeuvres was recorded as an absolute value. FEV1 and FVC were expressed as a percentage of the predicted values for age, sex, and height.

Anthropometry

All anthropometric measurements were taken according to a standardized protocol. Body weight was measured with participants wearing light clothes without shoes by using a balance beam scale. Height was measured at the same time. Body mass index (BMI) was calculated as body weight divided by height squared (kg/m²).

Collection of blood samples and storage

After a detailed history and pulmonary function test, 5.0 ml fasting venous blood was drawn into heparinized vials and tubes without anticoagulants. Tubes were centrifuged at 1500 g for 15 minutes to obtain plasma. Samples were divided into 0.5 ml aliquots. Erythrocytes and plasma were separated and stored at -80°C until biochemical measurements were taken.

Study measurements

Lipid peroxide assay

Plasma level of MDA was measured by measuring thiobarbituric acid reactive substances (TBARS) according to the method described by Buege and Aust.^[27] For assay, 0.5 ml of plasma sample was mixed with equal volume of 0.67% TBA and 30% of TCA in tubes. The mixture was heated in a boiling water bath for 20 minutes. The tubes were centrifuged at 4320 *g* for 15 minutes. The absorbance of the pink supernatant was taken at 530 nm on the UV-visible spectrophotometer. Results were expressed in nmol of MDA/ml of plasma.

Plasma protein carbonyl content assay

The measurement of protein oxidation was carried out by the estimation of total carbonyl content in the sample using 2,4-dinitrophenylhdrazine^[28] and its absorbance was recorded at 360 and 280 nm on the UV-visible spectrophotometer. The carbonyl formation was expressed as the ratio of absorbance at 360 nm to 280 nm and the units were expressed in nmol/mg protein of plasma.

Plasma protein sulfhydryl assay

The estimation of plasma protein sulfhydryl level was performed at 412 nm on the UV-visible spectrophotometer according to the method described by Sedalk and Lindsay.^[29] Results were expressed in mmol/l of plasma.

Superoxide dismutase assay in erythrocytes

SOD activity was assayed according to the method of Marklund and Marklund on the basis of autoxidation of pyrogallol.^[30] The change in absorbance was recorded at 412 nm on the UV-visible spectrophotometer for 3 minutes. Results were expressed in U/g Hb.

Catalase activity in erythrocytes

The quantitation of catalase activity in erythrocyte hemolysates was based on the oxidation of H_2O_2 . The activity of catalase was measured by taking 1.95 ml of potassium phosphate buffer (50 mM, pH 7.0), 50 µl of the sample to which 1 ml of H_2O_2 (30 mM) was pipetted in dark conditions.^[31] The solution was immediately read at 240 nm for 3 minutes by UV-visible spectrophotometer. The results were expressed in U/g Hb.

GPx activity in erythrocytes

GPx activity was determined spectrophotometrically in erythrocyte hemolysates using the technique adapted by Paglia and Valentine.^[32] GPx catalyses the oxidation of glutathione (GSH) by cumene hydroperoxide. In the presence of GSH reductase and NADPH, the oxidized GSH is reduced and NADPH is oxidized to NADP. GPx activity was measured by the decrease in NADPH absorbance at 340 nm on UV-visible spectrophotometer. Its activity was expressed in U/g Hb.

Total blood glutathione assay

The assay was carried out according to the method of Griffith,^[33] in which the reaction was initiated by addition of DTNB and the rate of reduction was measured at 412 nm on UV-visible spectrophotometer. Results were expressed in mmol/l.

Total antioxidant capacity (Ferric reducing antioxidant power of plasma assay)

The total antioxidant power in the plasma was estimated by measuring the ferric reducing ability by using the method of Benzie and Strain.^[34] The absorbance was taken at 593 nm by UV-visible spectrophotometer and total antioxidant status was calculated using ferrous sulfate as standard. The results were expressed in µmol/l.

Statistical analysis of data

Statistical analyses were performed with statistical package for the social sciences for Windows (Version 16.0; SPSS Inc) and Graph Pad Prism 5.01. Data were expressed as Mean ± SEM. The unpaired *t* test was used to compare the healthy controls with patients. The biochemical parameters were compared among patients with different asthma severity group by using one-way ANOVA. The relationship between different study parameters and the degree of airways obstruction was evaluated by computing the Pearson's correlation coefficient. P < 0.05 was considered significant.

Results

Table 1 shows the clinical and demographic characteristics of the study subjects. Patients with asthma had a significantly lower FEV1 (% predicted) and other spirometry indices than controls. The post changes in FEV1, FVC, and FEV1/FVC (% predicted) after nebulization with 200 µg of salbutamol were found remarkably at higher level than controls. Relative percent of the measured anthropometric and spirometric parameters have also been demonstrated in Figure 1.

It was shown that oxidative stress indicator measured as TBARS in plasma (4.19 \pm 0.10 nmol/ml, *P* < 0.001; Figure 2a) and total protein carbonyls (1.13 \pm 0.02 nmol/mg, *P* < 0.001; Figure 2b) were markedly enhanced in asthmatic patients as compared to control (2.84 \pm 0.12 nmol/ml and 0.79 \pm 0.02 nmol/mg, respectively). It was also observed that the level of total protein sulfhydryls (0.55 \pm 0.01 mmol/l, *P* < 0.05; Figure 2c) was decreased in patients with asthma as compared to healthy controls (0.60 \pm 0.02 mmol/l).

Figure 3a-c illustrates that the activity of SOD (2047 \pm 50.34 U/g Hb, *P* < 0.05), catalase (4374 \pm 67.98 U/g Hb, *P* < 0.01), and GPx (40.97 \pm 1.05 U/g Hb, *P* < 0.01) in erythrocytes were remarkably lower in asthmatic patients than the healthy controls (2217 \pm 60.11 U/g Hb; 4746 \pm 89.94 U/g Hb, and 48.37 \pm 2.47 U/g Hb, respectively) while the levels of total blood glutathione (0.94 \pm 0.02 mmol/l, *P* < 0.05; Figure 4a) were significantly increased in asthmatic patients as seen in controls (0.84 \pm 0.04 mmol/l). It was observed that total antioxidant status in plasma, which is measured as FRAP (750.90 \pm 21.22 µmol/l, *P* < 0.05) was remarkably decreased

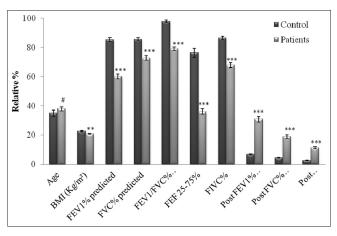


Figure 1: Relative percent of the measured anthropometric and spirometric parameters of asthmatic patients comparative to controls ${}^{\#}P > 0.05$, Non-significant; "P < 0.01; "P < 0.001

Characteristic	Asthmatic patients (N: 130)	Healthy controls (N: 70)	P value	
Age [#]	37.91 ± 1.38	34.97 ± 2.05	0.224	
Sex (Male/Female)#	90/40	52/18	0.454	
BMI (kg/m ²) ^{**}	20.85 ± 0.31	22.56 ± 0.53	0.003	
FEV1 (% predicted)***	60.14 ± 1.65	85.39 ± 1.31	0.000	
FVC (% predicted)***	72.84 ± 1.49	85.60 ± 1.12	0.000	
FEV1/FVC (% predicted)***	79.18 ± 1.33	97.86 ± 0.84	0.000	
FEF 25-75%***	36.17 ± 1.92	76.43 ± 3.07	0.000	
FIVC% ***	67.88 ± 1.72	86.57 ± 1.17	0.000	
Post FEV1% change***	30.80 ± 1.77	6.80 ± 0.35	0.000	
Post FVC% change***	18.92 ± 1.26	4.37 ± 0.26	0.000	
Post FEV1/FVC% change***	11.23 ± 0.70	2.43 ± 0.20	0.000	

BMI = body mass index, FEV1 = forced expiratory volume in 1 second, FVC = forced vital capacity, FEF = forced expiratory flow, FIVC = forced inspiratory vital capacity The probability unpaired t test is statistically significant 'P < 0.05; '''P < 0.001; *P > 0.05, Non-significant.

Parameters	Intermittent (N: 25)	Mild Persistent (N: 30)	Moderate persistent (N: 38)	Severe persistent (N: 37)
Age#	34.84 ± 3.22	39.43 ± 3.44	35.79 ± 2.09	40.92 ± 2.53
Sex (Male/Female)#	15/10	24/6	23/15	28/9
BMI (Kg/m ²) [*]	21.78 ± 0.56	21.64 ± 0.69	21.13 ± 0.64	19.31 ± 0.49
TBARS [•] (nmol of MDA/ml of plasma)	3.69 ± 0.20	4.04 ± 0.23	4.29 ± 0.22	4.55 ± 0.15
Total protein carbonyls [*] (nmol/mg protein of plasma)	1.02 ± 0.05	1.09 ± 0.04	1.16 ± 0.04	1.19 ± 0.03
Total protein sulfhydryls (mmol/l of plasma)	0.58 ± 0.02	0.58 ± 0.01	0.55 ± 0.01	0.52 ± 0.01
Erythrocytic SOD [#] (U/g Hb)	2076 ± 110.30	2042 ± 122.10	2015 ± 103.70	2063 ± 72.67
Erythrocytic Catalase [*] (U/g Hb)	4114 ± 129.40	4169 ± 136.40	4479 ± 146.00	4608 ± 109.5
Erythrocytic GPx [*] (U/g Hb)	43.91 ± 2.96	42.06 ± 2.11	42.78 ± 1.99	36.25 ± 1.40
Total blood glutathione [*] (mmol/l)	0.85 ± 0.04	0.89 ± 0.04	0.95 ± 0.04	1.02 ± 0.03
Total antioxidant status of plasma (FRAP) µmol/l	829.0 ± 50.24	813.3 ± 44.87	736.8 ± 42.44	661.9 ± 29.79

Table 2: Comparison of indices of oxidant-antioxidant status and other demographic variables among patients with asthma severity

TBARS = thiobarbituric acid reactive substances, MDA = Malondialdehyde, SOD = superoxide dismutase, GPx = glutathione peroxidase, FRAP = ferric reducing antioxidant power Data are expressed in Mean \pm SEM and One-way ANOVA analysis for comparison of group; *P > 0.05, Non-significant; P < 0.05

Table 3: Coefficients of correlation between oxidation variables and FEV1 (% predicted)

Study parameter	Correlation coefficients (r)		
	FEV1 of controls	FEV1 of asthmatics	
Plasma TBARS	-0.33**	-0.39**	
Total protein carbonyls of plasma	-0.18#	-0.35**	
Total protein sulfhydryls of plasma	0.37**	0.42**	
Erythrocytic SOD	0.33**	-0.07#	
Erythrocytic Catalase	0.43**	0.01#	
Erythrocytic GPx	0.42**	0.39**	
Total blood glutathione	-0.41**	-0.43**	
Total antioxidant status of plasma	0.49**	0.42**	

r denotes Pearson correlation coefficient; *P > 0.05, Non-significant; *P < 0.01

in asthmatic patients as compared to controls (840.40 \pm 28.39 μ mol/l; Figure 4b).

There were marked differences in biochemical parameters and other demographic variables among patients with different severities of asthma except age, sex, and SOD activity [Table 2]. Pearson's correlation coefficients of different studied oxidative variables with FEV1 (% predicted) showed remarkable negative correlation between plasma TBARS and total blood glutathione with FEV1 (% predicted) in patients with asthma as well as in controls, but there was no statistically significant difference found between total protein carbonyls and FEV1 (% predicted) in controls (r = -0.18, P > 0.05) [Table 3]. A remarkable positive correlation was found between total protein sulfhydryls, erythrocyte GPx, and total antioxidant status with FEV1 (% predicted) in asthmatic patients as well as in controls. Conversely, erythrocyte SOD (r = -0.07, P > 0.05) exhibited a slight negative correlation, whereas catalase showed an insignificant positive correlation in asthmatic patients (r =0.01, P > 0.05).

Discussion

Several studies have revealed that oxidants-antioxidants balance is an essential factor for the normal physiological function of the lungs. An increased oxidant and/or decreased antioxidant may reverse the physiologic oxidant-antioxidant balance in favor of oxidants. Thus, oxidative stress plays an important role in the pathogenesis of asthma and oxidants caused lipid peroxidation by oxidizing fatty acids and polyunsaturated lipids of cell membranes.

The oxidant–antioxidant status was investigated in blood (red blood cell and plasma) because it is an easily available source and also considered as an important pool of antioxidant defenses in the body rather than BAL fluid which is obtained through an invasive bronchoscope technique.^[35] Furthermore, asthma is an inflammatory disease in which cells are recruited from the peripheral blood.

In this work, it has been observed that the asthmatic patients have increased oxidative stress. It was shown by remarkable increase in lipid peroxidation product (MDA) and protein carbonyls as well as decrease in protein sulfhydryls in plasma [Figure 2a-c]. These are accompanied by alterations in several endogenous enzymatic antioxidants in blood including decreased GPx and SOD activities. The distinguished changes were noticed in catalase activity of erythrocytes and total antioxidant power of plasma of asthmatic patients. Rahman et al. reported that the plasma MDA level was higher in asthmatic patients than controls as well as in patients with asthma exacerbation as compared to stable asthma.^[12] Similarly, another study entails that MDA level in BAL fluid was higher in mild to moderate asthmatic patients.^[20] Moreover, protein carbonyl content was also significantly higher in asthmatic patients because most of the amino acids can be oxidized by ROS. Peroxynitrite anion is a potent oxidant that mediates not only the oxidation of both non-protein and protein sulfhydryls but also induces lipid peroxidation.[36]

Several studies have demonstrated alterations in different endogenous antioxidants in patients with asthma. The alterations in antioxidant defenses may involve either an increase or a decrease depending on the changes occurring due to a defense response. SOD is an intracellular antioxidant enzyme, helping in removing superoxide anion. In some studies, it has been reported that SOD activity in airway epithelium and erythrocytes of asthmatic patients was lower

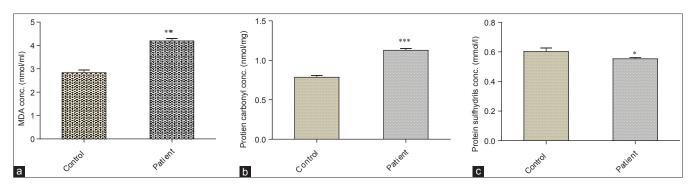


Figure 2: (a-c) Levels of lipid peroxidation products measured as TBARS expressed as nmol of MDA/ml of plasma, ""P < 0.001; total protein carbonyls in plasma expressed as nmol/mg of protein, ""P < 0.001; and total protein sulfhydryls in plasma expressed as mmol/l, 'P < 0.05. Data are expressed in Mean ± SEM

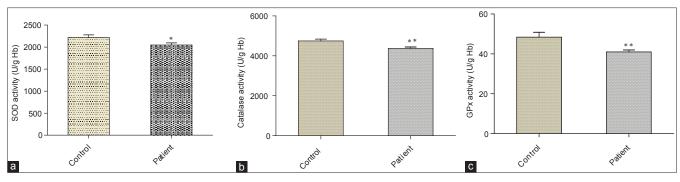


Figure 3: (a-c) Superoxide dismutase, catalase, and glutathione peroxidase activity in erythrocytes expressed as U/g Hb; 'P < 0.05 and "P < 0.01. Data are expressed in Mean ± SEM

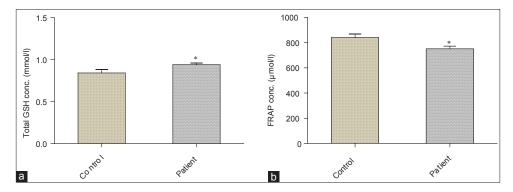


Figure 4: (a and b) Total blood glutathione and ferric reducing antioxidant power in plasma expressed as mmol/l and µmol/l, respectively, 'P < 0.05.

than healthy controls.^[17,37] We have also noticed significantly lower activities of SOD and catalase in red blood cells [Figure 3a,b] and these findings were very much similar to the earlier reported by Rai and Phadke (2006).^[38]

In a previous study, it has been reported that GPx play a significant role in peroxyl scavenging mechanism and maintaining functional integration of the cell membrane. Here, we observed a significant decrease in GPx activity in asthmatic patients as compared to controls [Figure 3c], which led to the high elevation of lipid peroxidation products as reported in the same investigated samples. Lower GPx level in asthmatic patients can be related to the clinical presentation of the disease and it indicates the presence of H_2O_2 in breath condensate of exhaled air which is elevated in asthmatics patients.^[37] Decreased activity of GPx has been well documented in literature^[39] and it is in favor of our findings. Selenium is an essential component of GPx and it indirectly helps in protecting cells against damage caused by free radicals. This might arise as a result of deficiency of selenium or inactivation caused by OH* and O₂[•].^[25,40,41] A number of studies have been done to measure the selenium deficiency in asthma through the antioxidant effects of GPx and it has been found that plasma levels of selenium was significantly lower in asthmatic patients.^[42-44] GPx is essential for removing toxic lipid oxidation products and H₂O₂, which are continuously generated as a result of sequestration and infiltration of inflammatory leukocytes in the lung. Low SOD activity together with low GPx activity in asthmatic patients proved the contribution of oxidative stress in the etiology of asthma. It was reported that the glutathione system is altered in lung inflammatory conditions such as asthma and many reports have shown that alterations in glutathione level have been found in asthmatic airways.[45]

imbalance.

Acknowledgments

The authors wish to thank all the subjects who participated in this study and Dr. Mohammad Faisal Siddiqui for work in his Interdisciplinary Brain Research Centre lab and critically reading manuscript.

References

- Masoli M, Fabian D, Holt S, Beasley R. The global burden of asthma: Executive summary of the GINA Dissemination Committee report. Allergy 2004;59:469-78.
- 2. Chung KF. Role of inflammation in the hyper reactivity of the airways in asthma. Thorax 1986;41:657-62.
- 3. Barnes PJ. Reactive oxygen species and airway inflammation. Free Radic Biol Med 1990;9:235-43.
- Vignola AM, Chanez P, Campbell AM, Souques F, Lebel B, Enander I, *et al.* Airway inflammation in mild intermittent and in persistent asthma. Am J Respir Crit Care Med 1998;157:403-9.
- Ciencewicki J, Trivedi S, Kleeberger SR. Oxidants and the pathogenesis of lung diseases. J Allergy Clin Immunol 2008;122:456-68.
- Riedl MA, Nel AE. Importance of oxidative stress in the pathogenesis and treatment of asthma. Curr Opin Allergy Clin Immunol 2008;8:49-56.
- Bousquet J, Jeffery PK, Busse WW, Johnson M, Vignola AM. Asthma. From bronchoconstriction to airways inflammation and remodeling. Am J Respir Crit Care Med 2000;161:1720-45.
- Hamid Q, Song Y, Kotsimbos TC, Minshall E, Bai TR, Hegele RG, et al. Inflammation of small airways in asthma. J Allergy Clin Immunol 1997;100:44-51.
- 9. Henricks PA, Nijkamp FP. Reactive oxygen species as mediators in asthma. Pulm Pharmacol Ther 2001;14:409-20.
- 10. Rahman I, Biswas SK, Kode A. Oxidant and antioxidant balance in the airways and airway diseases. Eur J Pharmacol 2006;533:222-39.
- 11. Terada LS. Specificity in reactive oxidant signaling: Think globally, act locally. J Cell Biol 2006;174:615-23.
- Rahman I, Morrison D, Donaldson K, MacNee W. Systemic oxidative stress in asthma, COPD and smokers. Am J Respir Crit Care Med 1996;154:1055-60.
- Fujisawa T. Role of oxygen radicals on bronchial asthma. Current Drug Targets Inflamm Allergy 2005;4:505-9.
- 14. Caramori G, Papi A. Oxidants and asthma. Thorax 2004;59:170-3.
- Kirkham P, Rahman I. Oxidative stress in asthma and COPD: Antioxidants as a therapeutic strategy. Pharmacol Ther 2006;111:476-94.
- Comhair SA, Erzurum SC. Antioxidant responses to oxidantmediated lung diseases. Am J Physiol Lung Cell Mol Physiol 2002;283:L246-L55.
- De Raeve HR, Thunnissen FB, Kaneko FT, Guo FH, Lewis M, Kavuru MS, *et al.* Decreased Cu, Zn-SOD activity in asthmatic airway epithelium: Correction by inhaled corticosteroid *in vivo*. Am J Physiol 1997;272:L148-54.
- Psarras S, Caramori G, Contoli M, Papadopoulos N, Papi A. Oxidants in asthma and in chronic obstructive pulmonary disease (COPD). Curr Pharm Des 2005;11:2053-62.
- 19. Bowler RP. Oxidative stress in the pathogenesis of asthma. Curr Allergy Asthma Rep 2004;4:116-22.
- 20. Ozaras R, Tahan V, Turkmen S, Talay F, Besirli K, Aydin S, *et al.* Changes in malondialdehyde levels in bronchoalveolar fluid and serum by the treatment of asthma with inhaled steroid and beta2-agonist. Respirology 2000;5:289-92.
- 21. Wood LJ, Fitzgerald DA, Gibson PG, Cooper DM, Garg ML. Lipid peroxidation as determined by plasma isoprostanes is related to

The present finding of our study also shows that the asthmatic patients have significantly higher level of total blood glutathione as compared to control subjects [Figure 4a]. Mak *et al.* have also reported that the total glutathione level increased in erythrocytes of asthmatic individuals.^[46] There have been several reports on the decreased antioxidant capacity in asthmatic patients with intense oxidative load. In this study, we found very low total antioxidant status [Figure 4b] which is correlated to the findings reported by Rahman *et al.*^[12]

The present study also demonstrates that the measures of oxidative stress and antioxidants differ significantly between patients grouped according to severity of asthma except SOD [Table 2]. It revealed that severe persistent asthmatics have highest levels of plasma lipid peroxidation products and simultaneously lowest level of plasma antioxidants. An increasing trend was observed in the level of MDA, total protein carbonyls, and total blood glutathione among patients with intermittent to severe asthma while total protein sulfhydryls, enzymatic antioxidants, and total antioxidant capacity shows a decreasing pattern in these respect (P < 0.05). We have also observed a dramatic enhancement in catalase activity among asthmatic patients (intermittent to severe). Al-Abdulla et al. also reported that the mean serum level of MDA was significantly raised with increasing severity of asthmatic attack among patients grouped according to degree of severity.^[47] Previously, Kanazawa et al. have also found differences related to severity in patients with acute exacerbations.[48] The association between asthma severity and anthropometric measurement demonstrates that age and gender was unrelated to disease severity similar to our findings.[49] Our results are also of interest in the context of increased oxidative burden remarkably associated to the pulmonary airways obstruction [Table 3]. Markers of lipid peroxidation (TBARS), total protein carbonyls, and total blood glutathione level showed negative associations with FEV1 (% predicted). Furthermore, it was also noticed that total antioxidant status, erythrocyte GPx, and total protein sulfhydryls were positively associated with FEV1 (% predicted). Schunemann et al. found a remarkable negative correlation between plasma TBARS and red cell GSH, but they did not find any significant correlation with GPx and total antioxidant status.^[50] A negative association in erythrocyte SOD activity was also observed, but this was not significant. Wood and co-workers also reported that SOD activity was negatively associated with the severity of asthma.[30] Some workers also demonstrated that the superoxide anion release was greater in patients with exacerbation of their disease and it was also inversely correlated with FEV1.[48,51] It suggested that the lower antioxidant defense cause greater degree of airway obstruction (decreased FEV1 % predicted).

In conclusion, this work describes the role of oxidative stress and antioxidants in severity of disease which provides a strong evidence for oxidant-antioxidant imbalance in disease progression. The findings of this study further strengthen the evidence that the altered oxidant-antioxidant balance is associated with airways obstruction. In further studies, we realized the need to elucidate pathophysiological mechanism involved in lung injury related to oxidant-antioxidant disease severity in mild asthma. Lipids 2000;35:967-74.

- Chanez P, Dent G, Yukawa T, Barnes PJ, Chang KF. Generation of oxygen free radicals from blood eosinophils from asthma patients after stimulation with PAF or phorbol ester. Eur Respir J 1990;3:1002-7.
- 23. Kelly FJ, Mudway L, Blomberg A, Frew A, Sandstrom T. Altered lung antioxidant status in patients with mild asthma. Lancet 1999;354:482-3.
- Kharitonov SA, Yates D, Robbins RA, Logan-Sinclair R, Shinebourne EA, Barnes PJ. Increased nitric oxide in exhaled air of asthmatic patients. Lancet 1994;343:133-5.
- Pearson DJ, Saurez-Mendez VJ, Day JP, Miller PF. Selenium status in relation to reduced glutathione peroxidase activity in aspirinsensitive asthma. Clin Exp Allergy 1991;21:203-8.
- Global Initiative for Asthma (GINA) Report [Internet]. Global Strategy for Asthma Management and Prevention. Updated 2007.Available at: http://www.ginasthma.com. Accessed November 10, 2008.
- Buege JA, Aust SD. Microsomal lipid peroxidation. Methods Enzymol 1978;52:302-10.
- Levine RL, Williams JA, Stadtman ER, Schacter E. Carbonyl assays for determination of oxidatively modified proteins. Methods Enzymol 1994;233:346-57.
- 29. Sedalk J, Lindsay RH. Estimation of total, protein-bound and non-protein sulfhydryl groups in tissues with Ellman's reagent. Anal Biochem 1968;25:192-205.
- 30. Marklund S, Marklund G. Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. Eur J Biochem 1974;47:469-74.
- 31. Aebi H. Catalase in vitro. Methods Enzymol 1984;105:121-6.
- Paglia DE, Valentine WN. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. J Lab Clin Med 1967;70:158-69.
- Griffith OW. Determination of glutathione and glutathione disulfide using glutathione reductase and 2-vinylpyridine. Anal Biochem 1980;106:207-12.
- Benzie IFF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of "Antioxidant Power": The FRAP assay. Anal Biochem 1996;239:70-6.
- Loft S, Poulsen HE. Markers of oxidative damage to DNA: Antioxidants and molecular damage. Methods Enzymol 1999;300:166-84.
- Radi R, Beckman JS, Bush KM, Freeman BA. Peroxynitrite oxidation of sulfhydrils. The cytotoxic potential of superoxide and nitric oxide. J Biol Chem 1991;266:4244-50.
- Fenech AG, Ellul-Micallef R. Selenium, glutathione peroxidase and superoxide dismutase in maltese asthmatic patients: Effect of glucocorticoid administration. Pulm Pharmacol Ther 1998;11:301-8.

- 38. Rai RR, Phadke MS. Plasma oxidant-antioxidant status in different respiratory disorders. Ind J Clin Biochem 2006;21:161-4.
- Antczak A, Nowak D, Shariati B, Krol M, Piasecka G, Kurmanowska Z. Increased hydrogen peroxide and thiobarbituric acid-reactive products in expired breath condensate of asthmatic patients. Eur Respir J 1997;10:1235-41.
- 40. Blum J, Fridovich I. Inactivation of glutathione peroxidase by superoxide radical. Arch Biochem Biophys 1985;240:500-8.
- Searle AJ, Wilson RL. Glutathione peroxidase: Effect of superoxide, hydroxyl and bromine radicals on enzyme activity. Int J Radiat Biol Relat Stud Phys Chem Med 1980;37: 213-7.
- 42. Jahnova E, Horvathova M, Gazdik F, Weissova S. Effects of selenium supplementation on expression of adhesion molecules in corticoid-dependent asthmatics. Bratisl Lek Listy 2002;103:12-6.
- Stone J, Hinks LJ, Beasley R, Holgate ST, Clayton BA. Reduced selenium status of patients with asthma. Clin Sci (Lond) 1989;77:495-500.
- Kadrabova J, Mad'aric A, Kovaciková Z, Podivínsky F, Ginter E, Gazdík F. Selenium status is decreased in patients with intrinsic asthma. Biol Trace Elem Res 1996;52:241-8.
- Smith LJ, Houston M, Anderson J. Increased levels of glutathione in bronchoalveolar lavage fluid from patients with asthma. Am Rev Respir Dis 1993;147(6 Pt 1):1461-4.
- Mak JC, Leung HC, Ho SP, Law BK, Lam WK, Tsang KW, et al. Systemic oxidative and antioxidative status in Chinese patients with asthma. J Allergy Clin Immunol 2004;114:260-4.
- Al-Abdulla NO, Al Naama LM, Hassan MK. Antioxidant status in acute asthmatic attack in children. J Pak Med Assoc 2010;60:1023-7.
- Kanazawa H, Kurihara N, Hirata K, Takeda T. The role of free radicals in airway obstruction in asthmatic patients. Chest 1991;100:1319-22.
- 49. Varraso R, Siroux V, Maccario J, Pin I, Kauffmann F. Asthma severity is associated with body mass index and early menarche in women. Am J Respir Crit Care Med 2005;171:334-9.
- Schunemann HJ, Muti P, Freudenheim JL, Armstrong D, Browne R, Klocke RA, *et al.* Oxidative stress and lung function. Am J Epidemiol 1997;146:939-48.
- Comhair SA, Ricci KS, Arroliga M, Lara AR, Dweik RA, Song W, et al. Correlation of systemic superoxide dismutase deficiency to airflow obstruction in asthma. Am J Respir Crit Care Med 2005;72:306-13.

How to cite this article: Ahmad A, Shameem M, Husain Q. Relation of oxidant-antioxidant imbalance with disease progression in patients with asthma. Ann Thorac Med 2012;7:226-32.

Source of Support: Nil, Conflict of Interest: None declared.

Announcement

Android App



A free application to browse and search the journal's content is now available for Android based mobiles and devices. The application provides "Table of Contents" of the latest issues, which are stored on the device for future offline browsing. Internet connection is required to access the back issues and search facility. The application is compatible with all the versions of Android. The application can be downloaded from https://market.android.com/details?id=comm.app.medknow. For suggestions and comments do write back to us.