

Review





2018; 9(17): 3084-3092. doi: 10.7150/jca.26422

# Exosome-related tumor microenvironment

Cheng Hu<sup>1,2</sup>, Meijuan Chen<sup>1,2</sup>, Rilei Jiang<sup>1,2</sup>, Yuanyuan Guo<sup>1,2</sup>, Mianhua Wu<sup>2</sup>, Xu Zhang<sup>1,2</sup>

1. School of Medicine and Life Sciences , Nanjing University of Chinese Medicine, Nanjing 210023, Jiangsu, P.R. China

 Jiangsu Collaborative Innovation Center of Traditional Chinese Medicine (TCM) Prevention and Treatment of Tumor, Nanjing University of Chinese Medicine, Nanjing 210023, Jiangsu, P.R. China

🖂 Corresponding author: Dr Xu Zhang, The School of Medicine and Life Science and Jiangsu Collaborative Innovation Center of Traditional Chinese Medicine (TCM) Prevention and Treatment of Tumor, Nanjing University of Chinese Medicine, Nanjing 210023, P.R. China. Email: zhangxu@njucm.edu.cn

© Ivyspring International Publisher. This is an open access article distributed under the terms of the Creative Commons Attribution (CC BY-NC) license (https://creativecommons.org/licenses/by-nc/4.0/). See http://ivyspring.com/terms for full terms and conditions.

Received: 2018.04.01; Accepted: 2018.07.05; Published: 2018.08.06

#### Abstract

The tumor microenvironment (tumor cells are located in the internal and external environment) is vital for the occurrence, growth and metastasis of tumors. An increasing number of studies have shown that exosomes are closely related to the tumor microenvironment. The mechanisms involved, however, are unclear. The focus of this review is on the exosome-related tumor microenvironment and other relevant factors, such as hypoxia, inflammation and angiogenesis. Many studies have suggested that exosomes are important mediators of metastasis, angiogenesis, and immune modulation in the tumor microenvironment. Additionally, exosomes can be isolated from bodily fluids of cancer patients, including urine, blood, saliva, milk, tumor effusion, cerebrospinal fluid, amniotic fluid and so on. Consequently, exosomes are potential biomarkers for clinical predictions and are also good drug carriers because they can cross the biofilm without triggering an immune response. Collectively, these findings illustrate that exosomes are crucial for developing potential targets for a new generation of pharmaceutical therapies that would improve the tumor microenvironment.

Key words: exosome; tumor microenvironment; angiogenesis; hypoxia; inflammation

# Introduction

It is acknowledged that cancer is the deadliest disease, responsible for disrupting human health worldwide. Cancer has long been widely studied by the scientific community, and more researchers are performing cancer research. Currently, there are many studies on cancer, but studies show that the effects of chemotherapy and radiation therapy on tumor cells themselves not only demonstrate adverse reactions but also unsatisfactory treatment effects. The main problem in current research is ignoring tumor microenvironment, the environment which tumor cells live in, which plays an important role in the evolution of cancer. The tumor microenvironment is a complex internal environment that is important for tumor survival and includes various stromal cells, extracellular matrix components and biological molecules. The important cellular components of the tumor microenvironment include fibroblasts, macrophages, inflammatory cells and important

biomolecules, including various growth factors, inflammatory factors, and proteases [1-3]. Factors related to the tumor microenvironment, such as hypoxia [4], inflammation, angiogenesis [5] and exosomes, play pivotal roles in tumor development, invasion and metastasis [6]. Matrix cells in the tumor microenvironment communicate with cancer cells through non-cell material, such as exosomes, and play crucial roles in tumor evolution and progression [7-9]. Exosomes can function as transporters in the tumor microenvironment and as biomarkers in clinical diagnoses [10]. Exosomes can help miRNA to be released into the tumor microenvironment [11], and miRNA plays vital roles in many biological functions in the tumor microenvironment, such as mediation of proliferation, apoptosis, migration and invasiveness of tumor cells [12]. The mRNA distribution in exosomes of tumor patients can be used to identify tumor biomarkers and provide a more specific target

for the treatment and prediction of tumors.

### 1. Tumor microenvironment

During tumor development, the tumor microenvironment is a pivotal pathological environment and can even serve as a bridge. Tumor development, recurrence and metastases are not only determined by cancer cells but are also associated with non-tumor cells of the tumor microenvironment. It is increasingly recognized that the tumor microenvironment, which includes cells such as macrophages, dendritic cells, T cells, endothelial cells and fibroblasts, as well as extracellular matrix (ECM) components, proteases, and cytokines, plays a role in tumor evolution and metastasis [2, 8, 13]. The tumor microenvironment plays a vital role in the transition from invasion growth before tumor cells invade the matrix [14]. In most cases, the late-stage matrix supports the development of the tumor more than the early stage matrix [15]. Stephen Paget in 1889 proposal that metastasis depends on cross-talk between selected cancer cells (the seeds) and specific tumor microenvironments (the soil) shows how cancer cells (the seeds) adapt to their environment (the soil). Therefore, this hypothesis is a good illustration of the relationships between cancer cells and the tumor microenvironment [16]. Traditional tumor studies focus on the tumor cells themselves, the seeds in the "seeds and soil" hypothesis, but neglect soil research, while the microenvironment, the soil of tumor cell growth and metastasis, is quite vital [17]. There is a large difference between the tumor microenvironment and normal tissue, and although the target hallmarks of the tumor microenvironment improve cancer therapies, the may tumor microenvironment is characterized by various pathologic responses, such as hypoxia, inflammation and angiogenesis [18-20].

# 2. Exosome-related tumor microenvironment

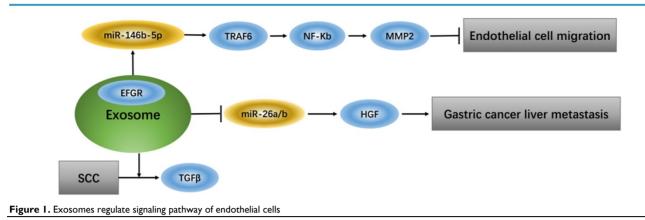
We know that the tumor microenvironment is very important in cancer development and that exosomes are a significant component of the tumor microenvironment [10]. Exosomes are thought to be between 30 and 100 nanometers in size with a classical "cup" or "dish" morphology and are released by almost all cell types, non-tumor cells or tumor cells. Exosomes carrying a broad range of cargoes, including nucleic acids, heat shock proteins and various enzymes [7, 10, 21-25]. Exosomes can promote cancer metastasis and influence other cells as follows: (1) the exosomes that come from tumor cells induce epithelial mesenchymal transition and degrade the matrix; (2) tumor-secreted exosomes directly or indirectly disturb endothelial cells via activating macrophages; (3) circulating tumor cells (CSTs) and the tumor activate platelets to release exosomes, influencing immune cells; (4) exosomes that are attached to the tumor up-regulate adhesive molecules on endothelial cells; and (5) exosomes can disseminate tumor cells in a suitable niche to proliferate, forming a micro-metastasis [26].

#### 2.1 Exosome and cell-to-cell communication

Exosomes are important components of extracellular vehicles that can mediate intercellular communication between tumor microenvironment components and play a role in the formation and development of tumors [27]. Presently, there are four main forms of communication between exosomes and cells: first, exosomes act as signaling complexes to stimulate target cells; second, exosomes transmit receptors between cells; third, exosomes transport functional proteins to receptor cells; and fourth, exosomes deliver genetic information to receptor cells via mRNA and miRNA [28]. Tumor cells can not only release more exosomes than normal cells but the contents of exosomes from tumors are also significantly different than those from normal cells. A large difference between exosomes from cancer cells and those from normal cells is that exosomes from cancer cells contain more miRNA than those from normal cells [29]. The two-way communication between tumor cells and the stroma or tumor can significantly affect the progression of disease and sensitivity of the tumor to treatment, and extracellular vesicles mediate the communication between cancer cells and the neighboring microenvironment [30].

#### 2.2 Exosomes and pathways

Exosomes that deliver epidermal growth factor receptor (EGFR) play a key role in regulating signaling pathways of endothelial cells because EGFR, which changes position, can effectively activate hepatocyte growth factor (HGF) by suppressing expression of its upstream miRNA (miR-26a/b). Additionally, up-regulation of liver HGF promotes gastric cancer liver metastasis, while down-regulation of liver HGF suppresses metastasis [31]. TGF $\beta$  type II receptor (T $\beta$ RII) is a common component in exosomes from squamous cell carcinoma (SCC) tumor cells and can stimulate TGF $\beta$  signaling in the tumor microenvironment [32]. Tumor-associated macrophage (TAM)-derived exosomes from ovarian cancer can target the miR-146b-5p/TRAF6/NF-кB/MMP2 pathway to suppress endothelial cell migration [33] (Figure 1).



# 3. Other factors related to the tumor microenvironment

# 3.1 Hypoxia and angiogenesis (VEGF)

Hypoxia-inducible factor 1 (HIF-1) is thought to be the key effector of the cell response to hypoxia, which is involved in promoting angiogenesis-related transcription genes [34]. When breast cancer cells and cancer-associated fibroblasts are subjected to hypoxia, HIF-1a/GPER signaling participates in the regulation of VEGF expression [35]. An in vivo study showed that the tumor ascorbate levels are negatively associated with tumor growth and that HIF-1 and its target proteins can control angiogenesis [36]. Expression of vascular endothelial growth factor (VEGF) is found at high levels in some hypoxic tumors. In vitro experiments have shown that soluble vascular endothelial growth factor receptor-2 (sVEGFR2) specifically binds to human vascular endothelial growth factor-A (VEGF-A), which inhibits tumor growth, and the high expression level of sVEGFR2 under hypoxic conditions plays a role in inhibition of the tumor angiogenesis [37]. It has been shown that the PTEN/PI3K/AKT signaling axis, which proceeds through the proteasome, controls hypoxia-induced HIF1a to avoid hypoxic degradation in macrophages and that this signaling pathway controls tumor-induced angiogenesis [38].

# 3.1.1 Angiogenesis

The tumor vasculature is a key component of the microenvironment and not only influences tumor behavior but is also related to the treatment effect [39]. Angiogenesis is a process that is associated with trauma, embryonic formation and solid tumors. The influence of proangiogenic factor overproduction in tumors leads to deformed vessel structures. When tumor blood vessels are abnormal, blood perfusion is low after hypoxia perfusion, leading to a microenvironment that promotes metastasis and invasion [40]. In more than 80% of pancreatic ductal

adenocarcinoma, the tumor-related transmembrane glycoprotein MUC1 is overexpressed, and increasing the expression levels of neuropilin-1 (NRP1, a co-receptor of VEGF) and its ligand VEGF lead to a proangiogenic tumor microenvironment [41]. PI3K modulates the tumor vasculature, either directly (by inhibiting endothelial cells) or indirectly (by inhibiting angiogenesis-promoting, tumor-associated myeloid cells and VEGF production by tumor cells) [42]. Identification of specific and non-redundant roles of class I PI3K isoforms in the tumor microenvironment has revealed the way that this hub regulates cancer progression [43]. Pro-inflammatory cytokines in the tumor microenvironment are known for their ability to either inhibit or promote cancer progression. Interleukin-31 (IL-31), which belongs to the pro-inflammatory IL-6 cytokine family, has been characterized in autoimmune disease and tumorigenesis, and IL-31 inhibits tumor growth partly via an anti-angiogenic effect [44]. In the tumor microenvironment, high expression of the extracellular matrix component tenascin-C (TNC) is related to decreased patient survival. TNC also has direct antiangiogenic effects on endothelial cells, and via tumor cells and cancer-associated fibroblasts, TNC regulates paracrine proangiogenic signal transduction [45]. TAp73, a member of the p53 family, regulates tumor angiogenesis by inhibiting proangiogenic and pro-inflammatory cytokines [46].

# 3.1.2 Hypoxia

Oxygen homeostasis is required by the body to generate energy. A decrease in oxygen or excess oxygen can be deleterious for cellular adaptation and growth. Hypoxia has a role not only in pathological conditions but also in physiological conditions [47]. A hypoxic solid tumor microenvironment can promote tumor metastasis, epithelial mesenchymal transition and angiogenesis [48]. In various tumors, hypoxia is associated with a poor response to treatment, such that hypoxia-inducible factor 1 regulates many genes that have critical cellular functions, leading cancer cells to adapt to the hypoxic environment [49]. It has been reported that the expression levels of HIF-1a and tumor-infiltrating lymphocytes (TILs) are positively correlated with the tumor microenvironment of esophageal squamous cell carcinoma and that patients with high levels of HIF-1a and TILs have the worst survival rates [50]. Thus, in gastric cancer cells, the hypoxic environment independently induces angiopoietin-like protein 4 and HIF-1a, which have adverse effects on the progression of the tumor [51]. Recent research has shown that in the tumor microenvironment, hypoxia and HIF-1a impact heat shock protein 70 (Hsp70) and the major histocompatibility class I chain-related proteins A and B (MICA/B), which are located on the cell membrane and, as ligands, affect the sensitivity of natural killer cell-mediated cytotoxicity [52]. Hypoxia-dependent and -independent alterations in immunological surveillance lead to different immune avoidance tactics [53]. There is some evidence that, in tumors, hypoxic induction factor (HIF) plays an inhibitory role in the regulation of the immune response [54, 55]. In hepatocellular carcinoma (HCC), transcatheter arterial chemoembolization (TACE) surgery increases the hypoxia status, and in HCC tissue, HIF-1a protein increases. HIF-1a stimulates expression the expression of COX-2 protein and positively regulates EMT, which promotes HCC invasion and metastasis, resulting in a poor prognosis [56]. Regarding pancreatic cancer, in a hypoxic tumor microenvironment, expression of receptor for advanced glycation end products (RAGE) is mainly controlled by NF-kB, but not HIF-1a, and RAGE is positively regulates KRAS-driven HIF-1a signaling and hypoxia-induced autophagy via the RAGE-KRAS-HIF1a pathway [57]. Pancreatic cancer cells exist in the hypoxic microenvironment and contain numerous factors that impact tumor survival, proliferation and metastasis, such as HIF-1a, which up-regulates and then induces miR-21 overexpression, allowing cells to avoid apoptosis [58]. In some late glioblastoma hypoxia microenvironments, IL-1b has been found to inhibit the transactivation activity of HIF-1, inducing expression of adrenomedullin (AM), which helps glioblastoma cells fight apoptosis induced by hypoxia [59]. In the tumor microenvironment, either chronic or cycling hypoxia enhances the expression or function of Livin, which promotes anti-apoptosis and resistance to ionizing radiation and temozolomide in glioblastoma [60]. Cycling hypoxia contributes to the resistance to cytotoxic therapies through anti-apoptotic effects, cycling hypoxia-induced Bcl-xL expression via ROS-mediated HIF-1a and NF-кВ activation, which plays an important role in the tumor microenvironment [61].

### 3.2 Inflammation

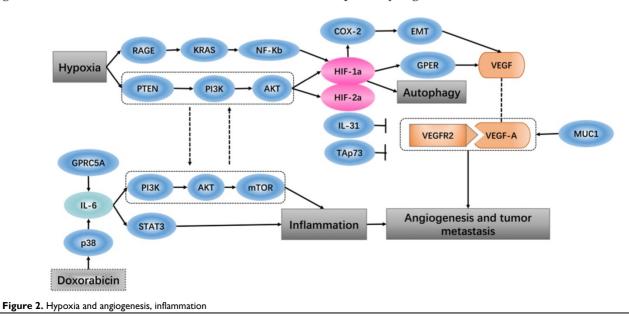
In many tumors, inflammatory infiltration is a marker that results in the continued release of anti-inflammatory, pro-inflammatory, and immunosuppressive cytokines the in tumor microenvironment [62]. The effect of the pro-inflammatory environment on cancer is to promote carcinogenesis by providing cytokines, growth factors and chemokines to maintain the cell proliferation rate, stimulate angiogenesis and inhibit cell apoptosis [63]. Inflammation acts as a double-edged sword; on the one hand, the protective response of inflammation can eliminate harmful stimuli and restore tissue homeostasis. On the other hand, uncontrolled inflammation leads to the development of malignancies [64, 65]. In many solid tumors, due to the inflammatory microenvironment, cells express a high concentration of macrophageassociated markers, which indicate a poor clinical consequence [66]. A study showed that, in the tumor microenvironment, the pro-inflammatory cytokine IL-6, which is produced by endothelial cells, promotes chemical resistance, indicating that the chemotherapy drug doxorubicin induces the release of acute IL-6 in reactive oxygen-mediated p38 vitro through activation. Doxorubicin leads to endothelial senescence mainly through the senescence-related inflammatory factor IL-6, which functions in the PI3K/AKT/mTOR pathway [67]. Prostate-specific IL-6 can autonomously induce prostate oncogenesis by expanding local inflammation, activating the STAT3 pathway and increasing paracrine insulin-like growth factor (IGF) signaling to reprogram gene expression[68]. We know that G protein-coupled receptor family C group 5 member A (GPRC5A) is a lung tumor suppressor that can be induced by retinoic acid, and overexpression of GPRC5A in head and neck squamous cells can suppress the activation of IL-6-induced STAT3 and inhibit tumor growth [69] (Figure 2).

# 4. Exosomes in cancer

It has been reported that glioma-derived exosomes that contain linc-POU3F3 can regulate the tumor microenvironment and influence angiogenesis. HBMECs treated with exosomes that have increased amounts of linc-POU3F3 can upregulate Angio, bFGFR, VEGF and bFGF expression, and these gene expression levels protein are mainly and pro-angiogenesis factors in angiogenesis regulation [70]. Exosomes contain high levels of miR-210 in the hypoxic tumor environment, and exosome-released miR-210 from hypoxic cells can inhibit Ephrin-A3 and PTP1B, which are the target genes of miR-210 and have an intimate connection with VEGF expression

and endothelial cell recruitment [71]. Under hypoxic conditions, lung cancer cells produce more exosomes than under normoxic conditions, and the expression levels of miR-23a are up-regulated in these exosomes. Hypoxia lung cancer-derived exosomal miR-23a induces prolyl hydroxylase 1/2 and tight junction protein ZO-1 to activate vascular endothelial cells; in simple terms, hypoxia tumor cells increase angiogenesis and permeability by regulating the exosomal miR-23a  $\rightarrow$  PHD1/2  $\rightarrow$  HIF-1a and  $\rightarrow$  ZO-1 regulatory pathway [72]. Studies have shown that small extracellular vesicles (sEVs) can be used as important media by senescent cells to promote tumor progression. The sEVs released by senescent cells carry EphA2, which can combine with ephrin-A1 and EphA2/ephrin-A1 to reverse signaling, eventually promoting cancer cell proliferation [73]. A special extracellular vesicle subgroup, microvesicles (MVs), from breast cancer cells activates VEGF receptors and promotes tumor angiogenesis, mainly via 90-kDa VEGF (VEGF90k), which interacts with Hsp90 and then combines with the vesicles. MV-associated VEGF90k is weakly expressed following exposure to bevacizumab, which is ineffective at blocking the activation of related VEGF receptors in MVs, and using an Hsp90 inhibitor that affects the release of VEGF90K from MVs can restore the sensitivity of VEGF90K to bevacizumab [74].

In in vivo and in vitro experiments of hepatocellular carcinoma (HCC), exosomes from HCC can induce sorafenib resistance, mainly via the HGF/c-Met/Akt signaling pathway, and inhibit sorafenib-induced apoptosis [75]. Compared with un-activated macrophages, exosomes from M2 macrophages express a higher level of miR-21. In gastric cancer cells, exosomal transfer of miR-21 from M2 macrophages mediates the resistance to DDP as well as the down-regulation of PTEN and up-regulation AKT. The apoptosis-associated gene Bcl-2 is also increased along with miR-21 overexpression [76]. miR-21 delivered by exosomes, comes from neighboring stromal cells in the omental tumor microenvironment, is transferred to ovarian cancer cells can conferring chemo-resistance. In ovarian cancer, APAF1 is the downstream target of miR-21, and their mRNA expression is inversely Additionally, APAF1 correlated. can mediate miR-21-induced paclitaxel resistance in ovarian cancer [77]. It has been reported that exosomes from breast cancer cells treated with paclitaxel (PTX) promote cell survival and chemo-resistance. PTX-treated cells generate exosomes enriched with survivin, and survivin from exosomes promotes cell survival [78]. It was discovered that shikonin inhibits the proliferation of MCF-7 cells by reducing tumor-derived exosomal miR-128, while miR-128 in exosomes negatively regulates the gene level of Bax in MCF-7 cells and promotes cell proliferation [79]. Exosomes involved in miR-7 can mediate apoptosis and inhibit tumor growth because miR-7 is a key sensitizer for TRAIL (TNF-related apoptosis inducing ligand)-induced apoptosis, and XIAP is the direct downstream of miR-7 [80]. It has been reported that miRNAs from exosomes can be used as intercommunication media between tumor cells and macrophages. TWEAK stimulation increases the level of miR-7 in both macrophages and macrophagesecreted exosomes. The level of miR-7 in recipient epithelial ovarian cancer (EOC) cells is increased due to EOC cell ingestion of the corresponding exosomes, and miR-7 eventually weakens EGFR/AKT/ERK1/2 pathway signal conduction related to EOC transfer



[81]. Gemcitabine increases cancer-associated fibroblast (CAF)-derived exosome release, and these exosomes play a role in the up-regulation of Snail and miR-146a expression. Snail is а chemo-resistance-inducing factor in recipient epithelial cells that promotes proliferation and drug resistance [82]. Tumor cell-derived extracellular vesicles (EVs) can cause apoptosis of human mesenchymal cells, which leads to the destruction of the peritoneal cortex and promotion of metastasis of tumor cells. EVs from ascites of ovarian cancer patients carry MMP1 mRNA, which can induce apoptosis of interstitial cells [83]. In cancer, exosomes act as communicators between tumors and their microenvironment, and exosome secretion due to lung cancer cell regulation causes cancer cell migration via TGF- $\beta$  and IL-10 [84]. Pyruvate kinase type M2 (PKM2) is an important enzyme of anaerobic glycolysis in tumor cells that plays a pivotal role in exosome release from tumor cells. Phosphorylated synaptosome-associated protein 23 (SNAP-23) is the phosphorylated substrate of PKM2 in tumor cells, which regulates the retention and release of exosomes, PKM2 phosphorylation and dimers not only transforms tumor cell metabolism from oxidative phosphorylation to anaerobic glycolysis but also promotes the secretion of exosomes in tumor cells by direct phosphorylation of SNAP23 [85] (Figure 3).

# 5. Tumor-derived exosomal miRNA as biomarkers

An early diagnosis is important for the prognosis of cancer. miRNAs in tumor-derived exosomes are potential diagnostic biomarkers for early-stage, 3089

non-small cell lung cancer (NSCLC) for next-generation sequencing compared with normal healthy cells. In NSCLC, miR-181-5p, miR-30a-3p, miR-30e-3p and miR-361-5p in exosomes are specific biomarkers for adenocarcinoma, and miR-10b-5p, miR-15b-5p and miR-320b in exosomes are specific biomarkers for squamous cell carcinoma [86]. miR-96 in exosomes is a serum biomarker for lung cancer, and its expression is positively correlated with high-grade, metastatic lung cancer. Expression of LIM-domainonly protein 7 (LMO7) is inversely related to the grade in lung cancer, and LMO7 overexpression has an inhibitory effect on miR-96 [87]. In the tumor microenvironment, miR-25-3p and miR-92a-3p are released into the serum by liposarcoma (LPS) cells through extracellular vehicles, and these miRNAs can be used as potential diagnostic biomarkers. miR-25-3p and miR-92a-3p, in a TLR7/8-dependent manner, activate tumor-associated macrophages to induce IL-6 secretion. In turn, IL-6 promotes LPS cell invasion proliferation, and transfer through interactions with the surrounding environment [88].

# Conclusions

Studies on tumors have mainly focused on the tumor cells, ignoring the microenvironment of tumor survival. The tumor microenvironment is a good soil for tumor growth, proliferation, invasion and metastasis [89]. Exosome are widely distributed in different body fluids and have a long half-life. Exosomes can influence the occurrence and development of tumors by interacting with various factors in the tumor microenvironment, such as hypoxia-, inflammation- and angiogenesis-associated

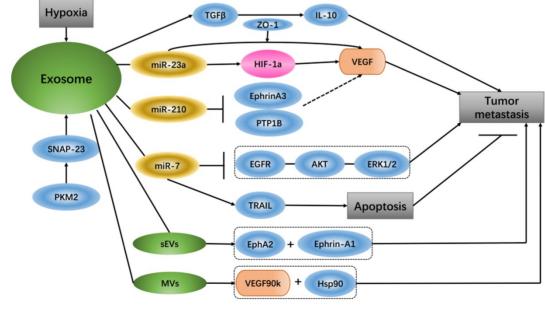


Figure 3. Exosomes in cancer

factors. Exosomes are nano-scale vesicles that are secreted by cells. As carriers, exosomes from tumor cells carry genetic material and proteins to other cells, leading to important information transmission between cells [90]. Intercellular communication is an indispensable cornerstone for the maintenance and development of multicellular tissue. The tumor microenvironment is a local environment composed of tumor cells with local infiltrating immune cells and stromal cells as well as the active media secreted by them.

Additionally, tumor-derived exosomes, through with other cells of interactions the tumor microenvironment, can regulate angiogenesis, tumor progression, metastasis and immune escape. The pathological processes mediated by exosomes give exosomes large potential as biomarkers [91]. Understanding the mechanism of exosomes is of great value for early screening, accurate diagnosis and prognosis assessment. Exosomes play an important role in tumor metastasis. They locate the position of transfer and activate specific cells to secrete specific cytokines to recruit cells to form the microenvironment before the tumor metastasizes. Exosomes induce transformation of epithelial mesenchymal cells and have the ability to migrate as well as promote the migration of tumor cells. Thus, miRNA can be transported to tumor cells to regulate the expression of corresponding genes that affect tumor cell growth. Through different proteins, miRNA can promote angiogenesis and induce immune escape, affecting the occurrence of metastatic tumor development.

Exosomes that are secreted can be used as the medium of signal transduction in tumor sensitivity to chemotherapy drugs and participate in the tumor process. Studying the mechanism of drug resistance by exosomes is helpful to better understand and treat neoplasms. Exosomes can be stable in urine, blood, saliva, milk, tumor effusion, cerebrospinal fluid, amniotic fluid and other fluids, especially in the tumor microenvironment. Exosomes can move back and forth in fluid, allowing them to cross the biofilm and deliver drugs without triggering an immune response [92]. However, it is difficult to purify exosomes, which is a difficult problem to overcome for current research and application.

#### Acknowledgements

This work was supported by grants from the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD), the Research Foundation of Education Bureau of Jiangsu Province (16KJA360001) (to X.Z.), the National Natural Science Foundation of China (81503374) (to M.C.) and the People Programme (Marie Curie Actions) of the European Union's Seventh Framework Programme FP7/2007-2013/ under REA grant agreement No. PIRSES-GA-2013-612589: CHETCH (China and Europe taking care of healthcare solutions). The authors thank the Editor and Reviewers for their significant contributions during the revision period.

#### **Author Contributions**

Dr Xu Zhang designed the review. Cheng Hu, Meijuan Chen, Rilei Jiang and Yuanyuan Guo collected the data from publications. Cheng Hu developed the database, wrote the manuscript, and edited the final text. Dr Xu Zhang and Dr Mianhua Wu provided financial support. All authors read and approved the final manuscript.

#### **Competing Interests**

The authors have declared that no competing interest exists.

#### References

- Chen F, Zhuang X, Lin L, Yu P, Wang Y, Shi Y, et al. New horizons in tumor microenvironment biology: challenges and opportunities. BMC medicine. 2015; 13: 45.
- Chou J, Shahi P, Werb Z. microRNA-mediated regulation of the tumor microenvironment. Cell cycle. 2013; 12: 3262-71.
- Cao Q, Yan X, Chen K, Huang Q, Melancon MP, Lopez G, et al. Macrophages as a potential tumor-microenvironment target for noninvasive imaging of early response to anticancer therapy. Biomaterials. 2018; 152: 63-76.
- Semenza GL. The hypoxic tumor microenvironment: A driving force for breast cancer progression. Biochimica et biophysica acta. 2016; 1863: 382-91.
- Lee E, Pandey NB, Popel AS. Crosstalk between cancer cells and blood endothelial and lymphatic endothelial cells in tumour and organ microenvironment. Expert reviews in molecular medicine. 2015; 17: e3.
- Wang M, Zhao J, Zhang L, Wei F, Lian Y, Wu Y, et al. Role of tumor microenvironment in tumorigenesis. Journal of Cancer. 2017; 8: 761-73.
- Zhang X, Yuan X, Shi H, Wu L, Qian H, Xu W. Exosomes in cancer: small particle, big player. Journal of hematology & oncology. 2015; 8: 83.
- Whiteside TL. Tumor-Derived Exosomes and Their Role in Cancer Progression. Advances in clinical chemistry. 2016; 74: 103-41.
- Guo W, Gao Y, Li N, Shao F, Wang C, Wang P, et al. Exosomes: New players in cancer (Review). Oncology reports. 2017; 38: 665-75.
- Wang Z, Chen JQ, Liu JL, Tian L. Exosomes in tumor microenvironment: novel transporters and biomarkers. Journal of translational medicine. 2016; 14: 297.
- Kahlert C, Kalluri R. Exosomes in tumor microenvironment influence cancer progression and metastasis. Journal of molecular medicine. 2013; 91: 431-7.
- Jiang X, Hu S, Liu Q, Qian C, Liu Z, Luo D. Exosomal microRNA remodels the tumor microenvironment. PeerJ. 2017; 5: e4196.
- 13. Hui L, Chen Y. Tumor microenvironment: Sanctuary of the devil. Cancer letters. 2015; 368: 7-13.
- Ma XJ, Dahiya S, Richardson E, Erlander M, Sgroi DC. Gene expression profiling of the tumor microenvironment during breast cancer progression. Breast cancer research : BCR. 2009; 11: R7.
- 15. Egeblad M, Nakasone ES, Werb Z. Tumors as organs: complex tissues that interface with the entire organism. Developmental cell. 2010; 18: 884-901.
- 16. Fidler IJ. The pathogenesis of cancer metastasis: the 'seed and soil' hypothesis revisited. Nature reviews Cancer. 2003; 3: 453-8.
- Ribelles N, Santonja A, Pajares B, Llacer C, Alba E. The seed and soil hypothesis revisited: current state of knowledge of inherited genes on prognosis in breast cancer. Cancer treatment reviews. 2014; 40: 293-9.
- Justus CR, Sanderlin EJ, Yang LV. Molecular Connections between Cancer Cell Metabolism and the Tumor Microenvironment. International journal of molecular sciences. 2015; 16: 11055-86.
- LeBleu VS. Imaging the Tumor Microenvironment. Cancer journal. 2015; 21: 174-8.
- Ivey JW, Bonakdar M, Kanitkar A, Davalos RV, Verbridge SS. Improving cancer therapies by targeting the physical and chemical hallmarks of the tumor microenvironment. Cancer letters. 2016; 380: 330-9.
- 21. Jia Y, Chen Y, Wang Q, Jayasinghe U, Luo X, Wei Q, et al. Exosome: emerging biomarker in breast cancer. Oncotarget. 2017.
- Vader P, Breakefield XO, Wood MJ. Extracellular vesicles: emerging targets for cancer therapy. Trends in molecular medicine. 2014; 20: 385-93.

- H. Rashed M, Bayraktar E, K. Helal G, Abd-Ellah M, Amero P, Chavez-Reyes A, et al. Exosomes: From Garbage Bins to Promising Therapeutic Targets. International Journal of Molecular Sciences. 2017; 18: 538.
- Roma-Rodrigues C, Fernandes AR, Baptista PV. Exosome in Tumour Microenvironment: Overview of the Crosstalk between Normal and Cancer Cells. BioMed Research International. 2014; 2014: 1-10.
- Roma-Rodrigues C, Raposo LR, Cabral R, Paradinha F, Baptista PV, Fernandes AR. Tumor Microenvironment Modulation via Gold Nanoparticles Targeting Malicious Exosomes: Implications for Cancer Diagnostics and Therapy. International journal of molecular sciences. 2017; 18.
- An T, Qin S, Xu Y, Tang Y, Huang Y, Situ B, et al. Exosomes serve as tumour markers for personalized diagnostics owing to their important role in cancer metastasis. Journal of extracellular vesicles. 2015; 4: 27522.
- Achreja A, Zhao H, Yang L, Yun TH, Marini J, Nagrath D. Exo-MFA A 13C metabolic flux analysis framework to dissect tumor microenvironment-secreted exosome contributions towards cancer cell metabolism. Metabolic engineering. 2017.
- Camussi G, Deregibus MC, Bruno S, Cantaluppi V, Biancone L. Exosomes/microvesicles as a mechanism of cell-to-cell communication. Kidney international. 2010; 78: 838-48.
- Milane L, Singh A, Mattheolabakis G, Suresh M, Amiji MM. Exosome mediated communication within the tumor microenvironment. Journal of controlled release : official journal of the Controlled Release Society. 2015; 219: 278-94.
- Han L, Xu J, Xu Q, Zhang B, Lam EW, Sun Y. Extracellular vesicles in the tumor microenvironment: Therapeutic resistance, clinical biomarkers, and targeting strategies. Medicinal research reviews. 2017; 37: 1318-49.
- Zhang H, Deng T, Liu R, Bai M, Zhou L, Wang X, et al. Exosome-delivered EGFR regulates liver microenvironment to promote gastric cancer liver metastasis. Nature Communications. 2017; 8: 15016.
- Languino LR, Singh A, Prisco M, Inman GJ, Luginbuhl A, Curry JM, et al. Exosome-mediated transfer from the tumor microenvironment increases TGFbeta signaling in squamous cell carcinoma. American journal of translational research. 2016; 8: 2432-7.
- Wu Q, Wu X, Ying X, Zhu Q, Wang X, Jiang L, et al. Suppression of endothelial cell migration by tumor associated macrophage-derived exosomes is reversed by epithelial ovarian cancer exosomal lncRNA. Cancer cell international. 2017; 17: 62.
- Toth RK, Warfel NA. Strange Bedfellows: Nuclear Factor, Erythroid 2-Like 2 (Nrf2) and Hypoxia-Inducible Factor 1 (HIF-1) in Tumor Hypoxia. Antioxidants. 2017; 6.
- De Francesco EM, Lappano R, Santolla MF, Marsico S, Caruso A, Maggiolini M. HIF-1alpha/GPER signaling mediates the expression of VEGF induced by hypoxia in breast cancer associated fibroblasts (CAFs). Breast cancer research : BCR. 2013; 15: R64.
- Campbell EJ, Vissers MC, Bozonet S, Dyer A, Robinson BA, Dachs GU. Restoring physiological levels of ascorbate slows tumor growth and moderates HIF-1 pathway activity in Gulo(-/-) mice. Cancer medicine. 2015; 4: 303-14.
- Collet G, Lamerant-Fayel N, Tertil M, El Hafny-Rahbi B, Stepniewski J, Guichard A, et al. Hypoxia-regulated overexpression of soluble VEGFR2 controls angiogenesis and inhibits tumor growth. Molecular cancer therapeutics. 2014; 13: 165-78.
- Joshi S, Singh AR, Zulcic M, Durden DL. A macrophage-dominant PI3K isoform controls hypoxia-induced HIF1alpha and HIF2alpha stability and tumor growth, angiogenesis, and metastasis. Molecular cancer research : MCR. 2014; 12: 1520-31.
- Hendry SA, Farnsworth RH, Solomon B, Achen MG, Stacker SA, Fox SB. The Role of the Tumor Vasculature in the Host Immune Response: Implications for Therapeutic Strategies Targeting the Tumor Microenvironment. Frontiers in immunology. 2016; 7: 621.
- Li W, Li X, Liu S, Yang W, Pan F, Yang XY, et al. Gold nanoparticles attenuate metastasis by tumor vasculature normalization and epithelial-mesenchymal transition inhibition. International journal of nanomedicine. 2017; 12: 3509-20.
- Zhou R, Curry JM, Roy LD, Grover P, Haider J, Moore LJ, et al. A novel association of neuropilin-1 and MUC1 in pancreatic ductal adenocarcinoma: role in induction of VEGF signaling and angiogenesis. Oncogene. 2016; 35: 5608-18.
- Okkenhaug K, Graupera M, Vanhaesebroeck B. Targeting PI3K in Cancer: Impact on Tumor Cells, Their Protective Stroma, Angiogenesis, and Immunotherapy. Cancer discovery. 2016; 6: 1090-105.
- Soler A, Angulo-Urarte A, Graupera M. PI3K at the crossroads of tumor angiogenesis signaling pathways. Molecular & cellular oncology. 2015; 2: e975624.
- Davidi S, Fremder E, Kan T, Raviv Z, Timaner M, Karin N, et al. The antiangiogenic role of the pro-inflammatory cytokine interleukin-31. Oncotarget. 2017; 8: 16430-44.
- Rupp T, Langlois B, Koczorowska MM, Radwanska A, Sun Z, Hussenet T, et al. Tenascin-C Orchestrates Glioblastoma Angiogenesis by Modulation of Proand Anti-angiogenic Signaling. Cell reports. 2016; 17: 2607-19.
- 46. Stantic M, Sakil HA, Zirath H, Fang T, Sanz G, Fernandez-Woodbridge A, et al. TAp73 suppresses tumor angiogenesis through repression of proangiogenic cytokines and HIF-1alpha activity. Proceedings of the National Academy of Sciences of the United States of America. 2015; 112: 220-5.

- Kumar H, Choi DK. Hypoxia Inducible Factor Pathway and Physiological Adaptation: A Cell Survival Pathway? Mediators of inflammation. 2015; 2015: 584758.
- Wang M, Zhao X, Zhu D, Liu T, Liang X, Liu F, et al. HIF-1alpha promoted vasculogenic mimicry formation in hepatocellular carcinoma through LOXL2 up-regulation in hypoxic tumor microenvironment. Journal of experimental & clinical cancer research : CR. 2017; 36: 60.
- Li S, Wei Q, Li Q, Zhang B, Xiao Q. Down-regulating HIF-1alpha by lentivirus-mediated shRNA for therapy of triple negative breast cancer. Cancer biology & therapy. 2015; 16: 866-75.
- Zhang L, Ye SB, Li ZL, Ma G, Chen SP, He J, et al. Increased HIF-1alpha expression in tumor cells and lymphocytes of tumor microenvironments predicts unfavorable survival in esophageal squamous cell carcinoma patients. International journal of clinical and experimental pathology. 2014; 7: 3887-97.
- Kubo H, Kitajima Y, Kai K, Nakamura J, Miyake S, Yanagihara K, et al. Regulation and clinical significance of the hypoxia-induced expression of ANGPTL4 in gastric cancer. Oncology letters. 2016; 11: 1026-34.
- Schilling D, Tetzlaff F, Konrad S, Li W, Multhoff G. A hypoxia-induced decrease of either MICA/B or Hsp70 on the membrane of tumor cells mediates immune escape from NK cells. Cell stress & chaperones. 2015; 20: 139-47.
- Eckert AW, Wickenhauser C, Salins PC, Kappler M, Bukur J, Seliger B. Clinical relevance of the tumor microenvironment and immune escape of oral squamous cell carcinoma. Journal of translational medicine. 2016; 14: 85.
- Kumar V, Gabrilovich DI. Hypoxia-inducible factors in regulation of immune responses in tumour microenvironment. Immunology. 2014; 143: 512-9.
- 55. Palazon A, Aragones J, Morales-Kastresana A, de Landazuri MO, Melero I. Molecular pathways: hypoxia response in immune cells fighting or promoting cancer. Clinical cancer research : an official journal of the American Association for Cancer Research. 2012; 18: 1207-13.
- Huang M, Wang L, Chen J, Bai M, Zhou C, Liu S, et al. Regulation of COX-2 expression and epithelial-to-mesenchymal transition by hypoxia-inducible factor-1alpha is associated with poor prognosis in hepatocellular carcinoma patients post TACE surgery. International journal of oncology. 2016; 48: 2144-54.
- Kang R, Hou W, Zhang Q, Chen R, Lee YJ, Bartlett DL, et al. RAGE is essential for oncogenic KRAS-mediated hypoxic signaling in pancreatic cancer. Cell death & disease. 2014; 5: e1480.
- Mace TA, Collins AL, Wojcik SE, Croce CM, Lesinski GB, Bloomston M. Hypoxia induces the overexpression of microRNA-21 in pancreatic cancer cells. The Journal of surgical research. 2013; 184: 855-60.
- Sun W, Depping R, Jelkmann W. Interleukin-1beta promotes hypoxia-induced apoptosis of glioblastoma cells by inhibiting hypoxia-inducible factor-1 mediated adrenomedullin production. Cell death & disease. 2014; 5: e1020.
- Hsieh CH, Lin YJ, Wu CP, Lee HT, Shyu WC, Wang CC. Livin contributes to tumor hypoxia-induced resistance to cytotoxic therapies in glioblastoma multiforme. Clinical cancer research : an official journal of the American Association for Cancer Research. 2015; 21: 460-70.
- Chen WL, Wang CC, Lin YJ, Wu CP, Hsieh CH. Cycling hypoxia induces chemoresistance through the activation of reactive oxygen species-mediated B-cell lymphoma extra-long pathway in glioblastoma multiforme. Journal of translational medicine. 2015; 13: 389.
- Rodriguez YI, Campos LE, Castro MG, Aladhami A, Oskeritzian CA, Alvarez SE. Sphingosine-1 Phosphate: A New Modulator of Immune Plasticity in the Tumor Microenvironment. Frontiers in oncology. 2016; 6: 218.
- Inacio Pinto N, Carnier J, Oyama LM, Otoch JP, Alcantara PS, Tokeshi F, et al. Cancer as a Proinflammatory Environment: Metastasis and Cachexia. Mediators of inflammation. 2015; 2015: 791060.
- Zhang Q, Zhu B, Li Y. Resolution of Cancer-Promoting Inflammation: A New Approach for Anticancer Therapy. Frontiers in immunology. 2017; 8: 71.
- Serhan CN. Pro-resolving lipid mediators are leads for resolution physiology. Nature. 2014; 510: 92-101.
- Serkova NJ. Nanoparticle-Based Magnetic Resonance Imaging on Tumor-Associated Macrophages and Inflammation. Frontiers in immunology. 2017; 8: 590.
- Bent EH, Gilbert LA, Hemann MT. A senescence secretory switch mediated by PI3K/AKT/mTOR activation controls chemoprotective endothelial secretory responses. Genes & development. 2016; 30: 1811-21.
- Liu G, Zhang J, Frey L, Gang X, Wu K, Liu Q, et al. Prostate-specific IL-6 transgene autonomously induce prostate neoplasm through amplifying inflammation in the prostate and peri-prostatic adipose tissue. Journal of hematology & oncology. 2017; 10: 14.
- Liu S, Ye D, Wang T, Guo W, Song H, Liao Y, et al. Repression of GPRC5A is associated with activated STAT3, which contributes to tumor progression of head and neck squamous cell carcinoma. Cancer cell international. 2017; 17: 34.
- Lang HL, Hu GW, Chen Y, Liu Y, Tu W, Lu YM, et al. Glioma cells promote angiogenesis through the release of exosomes containing long non-coding RNA POU3F3. European review for medical and pharmacological sciences. 2017; 21: 959-72.
- Jung KO, Youn H, Lee CH, Kang KW, Chung JK. Visualization of exosome-mediated miR-210 transfer from hypoxic tumor cells. Oncotarget. 2017; 8: 9899-910.
- 72. Hsu YL, Hung JY, Chang WA, Lin YS, Pan YC, Tsai PH, et al. Hypoxic lung cancer-secreted exosomal miR-23a increased angiogenesis and vascular

permeability by targeting prolyl hydroxylase and tight junction protein ZO-1. Oncogene. 2017; 36: 4929-42.

- Takasugi M, Okada R, Takahashi A, Virya Chen D, Watanabe S, Hara E. Small extracellular vesicles secreted from senescent cells promote cancer cell proliferation through EphA2. Nature communications. 2017; 8: 15729.
- Feng Q, Zhang C, Lum D, Druso JE, Blank B, Wilson KF, et al. A class of extracellular vesicles from breast cancer cells activates VEGF receptors and tumour angiogenesis. Nature communications. 2017; 8: 14450.
- Qu Z, Wu J, Wu J, Luo D, Jiang C, Ding Y. Exosomes derived from HCC cells induce sorafenib resistance in hepatocellular carcinoma both in vivo and in vitro. Journal of experimental & clinical cancer research : CR. 2016; 35: 159.
- Zheng P, Chen L, Yuan X, Luo Q, Liu Y, Xie G, et al. Exosomal transfer of tumor-associated macrophage-derived miR-21 confers cisplatin resistance in gastric cancer cells. Journal of experimental & clinical cancer research : CR. 2017; 36: 53.
- Au Yeung CL, Co NN, Tsuruga T, Yeung TL, Kwan SY, Leung CS, et al. Exosomal transfer of stroma-derived miR21 confers paclitaxel resistance in ovarian cancer cells through targeting APAF1. Nature communications. 2016; 7: 11150.
- Kreger BT, Johansen ER, Cerione RA, Antonyak MA. The Enrichment of Survivin in Exosomes from Breast Cancer Cells Treated with Paclitaxel Promotes Cell Survival and Chemoresistance. Cancers. 2016; 8.
- Wei Y, Li M, Cui S, Wang D, Zhang CY, Zen K, et al. Shikonin Inhibits the Proliferation of Human Breast Cancer Cells by Reducing Tumor-Derived Exosomes. Molecules. 2016; 21.
- Zhang X, Zhang X, Hu S, Zheng M, Zhang J, Zhao J, et al. Identification of miRNA-7 by genome-wide analysis as a critical sensitizer for TRAIL-induced apoptosis in glioblastoma cells. Nucleic acids research. 2017; 45: 5930-44.
- Hu Y, Li D, Wu A, Qiu X, Di W, Huang L, et al. TWEAK-stimulated macrophages inhibit metastasis of epithelial ovarian cancer via exosomal shuttling of microRNA. Cancer letters. 2017; 393: 60-7.
- Richards KE, Zeleniak AE, Fishel ML, Wu J, Littlepage LE, Hill R. Cancer-associated fibroblast exosomes regulate survival and proliferation of pancreatic cancer cells. Oncogene. 2017; 36: 1770-8.
- Yokoi A, Yoshioka Y, Yamamoto Y, Ishikawa M, Ikeda SI, Kato T, et al. Malignant extracellular vesicles carrying MMP1 mRNA facilitate peritoneal dissemination in ovarian cancer. Nature communications. 2017; 8: 14470.
- Wang Y, Yi J, Chen X, Zhang Y, Xu M, Yang Z. The regulation of cancer cell migration by lung cancer cell-derived exosomes through TGF-beta and IL-10. Oncology letters. 2016; 11: 1527-30.
- Wei Y, Wang D, Jin F, Bian Z, Li L, Liang H, et al. Pyruvate kinase type M2 promotes tumour cell exosome release via phosphorylating synaptosome-associated protein 23. Nature communications. 2017; 8: 14041.
- 86. Jin X, Chen Y, Chen H, Fei S, Chen D, Cai X, et al. Evaluation of tumor-derived exosomal miRNA as potential diagnostic biomarkers for early stage non-small-cell lung cancer using next-generation sequencing. Clinical cancer research : an official journal of the American Association for Cancer Research. 2017.
- Wu H, Zhou J, Mei S, Wu D, Mu Z, Chen B, et al. Circulating exosomal microRNA-96 promotes cell proliferation, migration and drug resistance by targeting LMO7. Journal of cellular and molecular medicine. 2017; 21: 1228-36.
- Casadei L, Calore F, Creighton CJ, Guescini M, Batte K, Iwenofu OH, et al. Exosome-Derived miR-25-3p and miR-92a-3p Stimulate Liposarcoma Progression. Cancer research. 2017; 77: 3846-56.
- Hede K. Environmental protection: studies highlight importance of tumor microenvironment. Journal of the National Cancer Institute. 2004; 96: 1120-1.
- Soung YH, Nguyen T, Cao H, Lee J, Chung J. Emerging roles of exosomes in cancer invasion and metastasis. BMB reports. 2016; 49: 18-25.
- Li W, Li C, Zhou T, Liu X, Liu X, Li X, et al. Role of exosomal proteins in cancer diagnosis. Molecular cancer. 2017; 16: 145.
- Batrakova EV, Kim MS. Using exosomes, naturally-equipped nanocarriers, for drug delivery. Journal of controlled release : official journal of the Controlled Release Society. 2015; 219: 396-405.