

Herpes simplex virus-based nerve targeting gene therapy in pain management

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Abstract: Chronic pain represents a major medical burden not only in terms of suffering but also in terms of economic costs. Traditional medical approaches have so far proven insufficient in treating chronic pain and new approaches are necessary. Gene therapy with herpes simplex virus (HSV)-based vectors offers the ability to directly target specific regions of the neuraxis involved in pain transmission including the primary afferent nociceptor. This opens up new targets to interact with that are either not available to traditional systemic drugs or cannot be adequately acted upon without substantial adverse off-target effects. Having access to the entire neuron, which HSV-based vector gene therapy enables, expands treatment options beyond merely treating symptoms and allows for altering the basic biology of the nerve. In this paper, we discuss several HSV-based gene therapy vectors that our group and others have used to target specific neuronal functions involved in the processing of nociception in order to develop new therapies for the treatment of chronic pain.

Keywords: pain, gene therapy, herpes simplex virus, opioids, voltage-gated sodium channels, inhibitory neurotransmitters

Introduction

Pain as an unpleasant experience to a noxious stimulus is a useful evolutionary adaptation. It is usually associated with two phenomena, ie, hyperalgesia, or increased pain sensitivity, and allodynia, or a painful response to a normally innocuous stimulus. Typically these two phenomena fade as the initiating noxious stimulus is removed and/or the resultant tissue injury resolves. In some instances this pain persists in the absence of stimuli or tissue injury. At this point, the pain is no longer adaptive and becomes a disease itself; this is generally termed “chronic pain”. Chronic pain can result from a myriad of causes, including arthritis and rheumatism, back or neck problems, frequent headaches, painful neuropathies, trigeminal neuralgia or mandibular joint disorder, pelvic and abdominal disorders, and fibromyalgia or complex regional pain syndrome. The current estimate of the number of adult Americans suffering from chronic pain is approximately 100 million.^{1,2} One recent Internet-based survey showed that 30.7% of respondents reported some form of chronic pain lasting 6 months or longer.³ Of these subjects, 32% reported their chronic pain as severe (≥ 7 on a scale from 0 to 10). The financial cost of chronic pain in the US is over \$600 billion, of which half are direct medical expenditures and half lost productivity.¹ Clearly, chronic pain represents a major cause of morbidity, significantly impairing individual quality of life and imposing a substantial burden on society.

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Nociceptive versus neuropathic pain

Pain is often described as being either nociceptive or neuropathic. Nociceptive pain results from activation of neuronal pain pathways from damage to non-neuronal tissues, such as muscles, joints, viscera, and skin. This type of damage excites local primary afferent nociceptors (PANs) that are activated by specific receptors or ion channels sensitive to heat, mechanical stimuli, protons, or cold.⁴ The central projections of PANs, which have their cell bodies in the dorsal root ganglia, synapse with nociceptive-specific and wide dynamic range projection neurons in the dorsal horn of the spinal cord, which, in turn, project centrally to transmit the affective and discriminatory components of pain. These components of pain processing are further influenced by excitatory and inhibitory interneurons within the dorsal horn.⁵ Neuropathic pain results from a lesion, disease, or dysfunction of the nociceptive pathway itself, either in the presence or absence of other tissue damage, although not all lesions of this pathway result in pain.^{6,7}

Injury to tissues leading to nociceptive pain cause the release of several inflammatory molecules capable of stimulating PANs, including hydrogen and potassium ions, serotonin, substance P, glutamate, histamine, bradykinins/tachykinins, prostaglandins, and leukotrienes.^{8,9} In a process known as peripheral sensitization, these compounds alter the response characteristics of the primary nociceptors, made up of unmyelinated C-fibers and thinly myelinated A δ fibers, by activating second messenger systems such as protein kinase A and protein kinase C (PKC) in primary nociceptors, causing increased sensitivity of endogenous receptors and altered sodium channel distribution, resulting in a lowered activation threshold and increased sensitivity to pain.^{4,10} This may also have the added effect of activating previously silent nociceptors. The end result of all these changes is an increase in nociceptive input into the spinal cord.

Prolonged nociceptive input results in similar changes in the spinal cord, resulting in "central sensitization". Activation of intracellular kinases within projection neurons causes phosphorylation of ion channels and receptors, resulting in immediate central sensitization.⁴ In addition, the same second messenger systems activate transcription factors, causing altered gene expression and long-lasting changes in the physiology of projection neurons, resulting in delayed central sensitization. Central sensitization is characterized by an increase in receptive field size, a reduction in threshold, and an increase in responsiveness in individual projection neurons which can cause nociceptive-specific neurons to behave like wide dynamic range neurons and respond to

normally nonpainful stimuli, resulting in allodynia.^{4,11-13} The phenomenon of windup in wide dynamic range neurons is also involved in central sensitization, and is known to involve activation of the N-methyl-D-aspartate subtype glutamate receptor.¹⁴

Peripheral nerve injury can lead to neuropathic pain through mechanisms that are not completely understood.⁷ Injured afferent neurons develop spontaneous neural activity and ectopic discharges¹⁵ while the uninjured afferent neurons can undergo peripheral sensitization as described above.¹⁶ Changes in trophic factor support affect both injured and uninjured afferents, and can alter gene expression in dorsal root ganglia neurons, resulting in increased pain sensitivity.^{17,18} Central sensitization also occurs in response to nerve injury, but unlike that involved with inflammation, which is dependent on ongoing nociceptive input, the central sensitization associated with neuropathic pain can involve different sets of afferents.⁷ For example, low-threshold A β fibers responsive to touch shift to a nociceptor-like phenotype or synapse on sensitized nociceptive-specific and wide dynamic range neurons, resulting in a painful sensation to a light touch stimulus (allodynia),¹⁹ and there is a change in the subtypes of voltage-gated sodium channels that are expressed in injured PANs, resulting in altered signal propagation.²⁰ Peripheral nerve damage is also associated with apoptosis of dorsal horn interneurons.²¹ These largely inhibitory neurons synapse with both the PANs and projection neurons and express gamma aminobutyric acid (GABA), glycine, and endogenous opioid peptides such as enkephalin and endomorphin in order to modulate pain signaling from the periphery. A downregulation in the receptors for these inhibitory neurotransmitters also occurs in response to nerve injury.²¹

In addition to the above mechanisms, substance P and glutamate released by the PANs and prostaglandins and nitrous oxide released by the nociceptive-specific and wide dynamic range neurons activate glia within the dorsal horn, which in turn release proinflammatory cytokines and growth factors that have hyperalgesic effects, resulting in a positive feedback mechanism of increased and prolonged pain.²²

Difficulty in treating chronic pain

Chronic pain is a debilitating disease because, for many patients, currently available analgesics either do not offer adequate pain relief or produce intolerable side effects. In regards to nociceptive pain, the best treatment is repair of the injured tissue; however, for many chronic conditions, such as arthritis or cancer, this may not be possible. While nonsteroidal anti-inflammatory drugs are used to treat

mild inflammatory pain, opioids or narcotic analgesics are an essential component of chronic nociceptive pain management. These drugs, eg, morphine, act at opioid μ , δ , and κ receptors in the dorsal horn to inhibit nociceptive transmission.²³ Unfortunately, these receptors are found throughout the neuraxis and body tissues, so the long-term use of opioid drugs is limited by adverse systemic effects, including nausea, constipation, dizziness, drowsiness, and respiratory depression, the development of tolerance, and ultimately dependence on such drugs.

The diverse mechanisms underlying neuropathic pain make it difficult to establish a unified clinical management approach because patients who present with the same symptoms may respond differently to the same medication. Generally, patients with neuropathic pain respond poorly to nonsteroidal anti-inflammatory drugs or opioids. The classes of medications that have proven most beneficial to date are anticonvulsants and antidepressants, no doubt reflecting the role that abnormal neuronal activity plays in the pathophysiology of neuropathic pain.²⁴ While some patients respond well to these types of drugs, it is estimated that half of all patients suffering from neuropathic pain receive little, if any, pain relief from currently available drugs.²⁵

Gene therapy approaches for treating chronic pain

The medical and economic burden of chronic pain coupled with insufficient treatment options make the development of novel approaches necessary, and gene therapy is one such approach. Gene therapy (or gene transfer) refers to the introduction and expression of DNA or RNA sequences in tissues and/or cells. It can be used to express proteins, or other gene products, that directly target the neurologic components underlying nociception, such as receptors, neurotransmitters, ion channels, second messenger systems, and biochemical mediators of inflammation.^{26,27} Gene therapy has several benefits compared with traditional pharmaceuticals in the treatment of chronic pain. Therapeutic genes can be delivered to specific regions and/or cells in the neuraxis, resulting in a localized concentration of the gene product. This significantly increases effectiveness while simultaneously reducing systemic adverse effects compared with traditional drugs. Small molecule drugs typically require large systemic doses to ensure adequate local concentrations of drug products, often resulting in side effects. Through targeted delivery and/or the use of specialized gene products (eg, transcription factors, dominant/negative genes, small interfering (si)RNA), a far greater number of potential therapeutic targets become available to act upon and inhibit

pain signaling. Loss of efficacy due to tolerance is very unlikely to occur with gene therapy due to the relatively small amount of therapeutic protein expressed and the tendency to utilize endogenous protein products, which are not normally subject to tolerance. Gene therapy offers the possibility of long-term expression of therapeutic proteins in specific cells, an especially attractive benefit for treating chronic conditions. Lastly, there is almost no possibility for abuse or addiction because the effect remains local and does not interact with higher brain centers.

Any location along the pain pathway can be targeted by gene therapy, but the PANs and their synapse with second order neurons in the dorsal horn are ideal because they are integral in the pathophysiology of chronic pain. Three gene therapy strategies have been employed to deliver analgesic genes to the nervous system: cell-based therapies, usually transplantation of transformed cells into the subarachnoid space;^{28–30} use of plasmids or oligonucleotides, sometimes encapsulated in liposomes to facilitate entry into cells;^{31–33} and use of viral-based vectors. Viral vector gene therapy takes advantage of the natural ability of viruses to infect and have their genes expressed by host cells and has been the most utilized system for gene delivery into the nervous system.^{34,35}

Herpes simplex virus as a vector system for treating chronic pain

Among the several viral-based vector systems capable of gene delivery, herpes simplex virus-1 (HSV)-based vectors are especially useful for delivery to the nervous system and PANs. HSV is a large human virus containing 152 kb of linear double-stranded DNA encoding at least 75 gene products. The virus particle consists of a nucleocapsid surrounded by an envelope containing glycoproteins essential for virus attachment and penetration into cells. The virus genome consists of two segments, the unique long (U_L) and unique short (U_S), each flanked by inverted repeats. Approximately half of the gene products are not essential for growth in cell culture. Once these nonessential genes have been deleted, 40–50 kb of foreign DNA can be accommodated within the virus.

In its natural life cycle, wild-type HSV is spread by direct contact. It infects and replicates in skin or mucous membrane and released viral particles are taken up by sensory nerve terminals.³⁶ These particles travel via retrograde transport along axons to the neuronal perikaryon in the dorsal root ganglia, where wild-type HSV may either re-enter the lytic cycle or alternatively establish a latent state that is characterized by the persistence of viral genomes as nonreplicating intranuclear episomal elements. In the lytic life cycle, viral

immediate early genes transactivate expression of viral early genes, the products of which principally code for proteins required for viral DNA replication.³⁷ Replication-defective HSV vectors can be engineered by removing one or more genes that code for immediate early gene products. These vectors can only replicate in specialized cell lines that provide the missing gene product but are still capable of infecting sensory neurons and expressing an inserted transgene product.^{38–40} Transgene expression is mediated by the choice of promoter. Short-term (weeks) expression is achieved using a strong promoter such as the immediate early gene promoter derived from human cytomegalovirus. HSV possesses a natural promoter system that is uniquely active during latency when all the other viral promoters are repressed.⁴¹ This latency active promoter has been used for the long-term (months) expression of transgenes in sensory neurons of the peripheral and central nervous systems.^{42,43}

Due to the natural biology of the parent virus, HSV-based vectors injected into the dermal layer of the skin will infect the sensory nerves innervating that area. Thus, by selecting the correct dermatome for direct vector inoculation, one can specifically target the sensory neurons in which one wants to express the therapeutic product. This restricts the “therapy” to the most appropriate region of the body, both minimizing the possibility of side effects while maximizing therapeutic efficacy.

Strategies for using HSV vectors to treat pain

Because HSV-based vectors can easily be delivered to sensory neurons, several potential therapeutic targets are available that could interfere with the processing of noxious stimuli from the periphery to the central nervous system (Figure 1). One such target is the synapse between the PAN and projection neuron within the dorsal horn. Projections from the brainstem can alter pain perception through interaction with interneurons located in the dorsal horn.⁴⁴ These interneurons use inhibitory neurotransmitters to modulate the activity of both primary nociceptors and projection neurons, and as discussed above, pathologic changes in these neurons have been observed with chronic pain. Our group and others have used replication-defective HSV-based vectors to overexpress and/or increase the activity of these inhibitory neurotransmitters in PANs, and thus increase the overall inhibitory tone at this synapse.

The first of these transmitters to be expressed was the endogenous opioid peptide enkephalin (ENK), which is the natural ligand for the delta opioid receptor.⁴⁵ HSV vector-mediated transfer of the ENK gene to lumbar dorsal root ganglia of rodents via footpad inoculation has been demonstrated to reduce C-fiber-evoked pain behaviors in response to acute inflammatory stimuli, such as injection of capsaicin⁴⁶ or formalin,⁴⁷ and decrease hyperalgesia associated with chronic

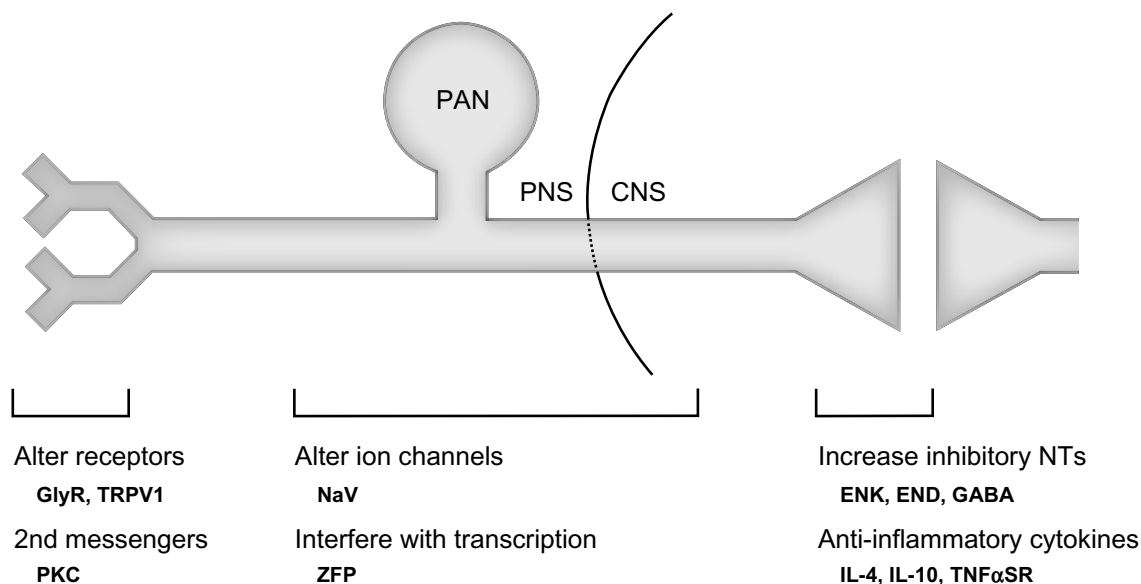


Figure 1 Potential therapeutic target areas to treat pain that are accessible via HSV-based gene transfer. The illustration represents a primary afferent nociceptor with its receptive field in the periphery and its synapse with a projection neuron in the dorsal horn of the spinal cord. HSV-based gene transfer can be used to interfere with pain processing at many locations along this route. Specific targets (shown in bold) that have been demonstrated to decrease nociceptive responses in animal models of pain are shown for each location.

Abbreviations: GlyR, glycine receptor; TRPV1, transient receptor potential vanilloid type 1; PKC, protein kinase C; NaV, voltage-gated sodium channel; ZFP, zinc finger protein transcription factor; ENK, enkephalin; END, endomorphin; GABA, gamma aminobutyric acid; IL-4, interleukin-4; IL-10, interleukin-10; TNF α SR, tumor necrosis factor alpha soluble receptor; HSV, herpes simplex virus; PAN, primary afferent nociceptor; PNS, peripheral nervous system; CNS, central nervous system; NTs, neurotransmitters.

inflammation^{48,49} and bone cancer.⁵⁰ Similarly, footpad inoculation has been shown to reduce nociceptive behavior in rodent models of chronic neuropathic pain, including spinal nerve ligation⁵¹ and painful diabetic neuropathy,⁵² while application to the rat vibrissal pad has reduced neuropathic pain associated with trigeminal neuralgia.⁵³ Others have since shown that ENK-expressing HSV vectors directly applied to the bladder or pancreas produce analgesia in visceral pain models of cystitis and pancreatitis⁵⁴⁻⁵⁶ and extended findings to primates.⁵⁷ All of these studies led to the first human gene therapy trial for the treatment of pain. In a ten-person, open-label, dose-escalation trial, NP2, an ENK-expressing HSV vector, demonstrated a positive safety profile with no serious adverse events in patients with severe focal pain due to cancer. Analysis of daily numeric pain scores over the course of the study (28 days) suggested a dose-related analgesic response to the drug.⁵⁸

Two other inhibitory neurotransmitters that play a role in nociceptive transmission at the dorsal horn synapse have been utilized as HSV vector transgenes, ie, GABA and endomorphin. GABA is the dominant inhibitory neurotransmitter in the spinal cord and exerts tonic inhibitory control over nociceptive transmission primarily via GABA_A and GABA_B receptors found presynaptically and postsynaptically on dorsal horn neurons. The evidence suggests that prolonged nociceptor excitation reduces the GABA-mediated tonic inhibition within the dorsal horn by reducing presynaptic GABA production and/or release.⁵⁹ GABA synthesis in PANs can be increased using HSV vectors expressing the rate-limiting enzyme in the biosynthesis of GABA from glutamate, ie, glutamic acid decarboxylase. HSV vectors expressing glutamic acid decarboxylase are more effective in reducing neuropathic pain in rodent models of spinal nerve ligation, spinal cord injury, or painful diabetic neuropathy⁶⁰⁻⁶³ compared with ENK-expressing vectors, but less effective in reducing pain-related behaviors in animal models of inflammatory pain (formalin test, bone cancer; JRG, personal observation). Endomorphin is a natural ligand of the mu opioid receptor, which is the major site of action for narcotic drugs such as morphine. While no natural endomorphin gene has been identified, our group engineered an HSV vector containing a synthetic endomorphin gene construct based on the known amino acid sequence. This vector demonstrated antinociceptive properties in the spinal nerve ligation model of neuropathic pain⁶⁴ and the complete Freund's adjuvant model of inflammatory pain.⁶⁵ Interestingly, this vector demonstrated an analgesic effect in rats with opiate tolerance induced by daily treatments with morphine, suggesting that there is

no cross-tolerance between morphine and vector-mediated endomorphin.

Aside from increasing inhibitory tone at the dorsal horn synapse, HSV vectors can be used to release anti-inflammatory molecules at this site to decrease the effects of glia-mediated neuroimmune activation that occur in response to chronic pain. Anti-inflammatory cytokines such as interleukin-4 (IL-4), IL-10, and IL-13 have been shown to have analgesic properties in several different animal models of acute and chronic pain.⁶⁶⁻⁷⁰ Along the lines of those experiments, HSV-mediated expression of IL-4 or tumor necrosis factor alpha soluble receptor in lumbar dorsal root ganglia neurons following footpad inoculation significantly decreases pain-related behavior in the spinal nerve ligation model of neuropathic pain^{71,72} and an HSV vector expressing IL-10 decreases spontaneous paw flinches in the formalin model of acute inflammatory pain.⁷³ Other studies have used direct intrathecal administration to the lumbar spinal cord of adenovirus or adenoassociated virus-based vectors expressing IL-10 to reduce neuropathic pain in animals.^{74,75}

Another set of potential therapeutic targets accessible to HSV vectors are the receptors that transduce noxious stimuli to electrical potentials within the PANs. These targets are only now being examined, but could provide a more fine-tuned approach to pain control, ie, decrease pain from a specific stimuli such as heat. For example, the transient receptor potential vanilloid type 1 (TRPV1) receptor is an ion channel activated by high temperature and involved in painful heat sensation. Transduction of rodent dorsal root ganglia by footpad inoculation with an HSV vector expressing a mutant TRPV1 gene that does not form the channel results in a significantly attenuated thermal response,⁷⁶ but does not affect an animal's response to cold temperature (JRG, personal observation). A similar effect has been achieved by using an HSV vector that expresses a dominant negative protein kinase C epsilon. TRPV1 activity is regulated by phosphorylation via protein kinase C epsilon and introduction of a dominant negative form of the enzyme alters the response of rats to capsaicin.⁷⁷ Another approach involves the insertion of novel receptors into sensory neurons. Our group has used an HSV vector expressing the glycine receptor as a "regulatable target" in this regard. Glycine is an inhibitory neurotransmitter used throughout the nervous system but primary sensory neurons do not express glycine receptors. Inoculation into the footpad in rats with a glycine receptor vector resulted in a reduction in inflammatory pain-related behavior following application of glycine to the paw.⁷⁸ This method could be

refined by using vectors that express channels or receptors that respond to other ligands or drugs.

Other attractive therapeutic targets within the PANs are the ion channels responsible for propagating the transduced stimuli at the periphery to the terminals located in the dorsal horn. Voltage-gated sodium channels are especially interesting candidates because these proteins lie at points of convergence and are necessary for normal nociceptive processing. It is known that the voltage-gated sodium channels NaV1.7, NaV1.8, and NaV1.9 play roles in nociception, and modifying their function has become a natural focus in recent studies.⁷⁹ The sodium channel subunit NaV1.7 plays a significant role in action spike initiation, and natural gain-of-function and loss-of-function mutations are associated with clinical pain syndromes of paroxysmal extreme pain disorder and congenital insensitivity to pain, respectively.^{80,81} Further, the reductions in diabetic neuropathy pain-related behavior associated with the HSV-glutamic acid decarboxylase and HSV-ENK gene therapy studies discussed above appear to be partly attributable to decreased expression of NaV1.7 in dorsal root ganglia neurons.^{52,63} Not surprisingly, knockdown of NaV1.7 via HSV-mediated expression of a microRNA or an antisense oligodeoxynucleotide has been shown to reduce hyperalgesia in models of painful diabetic neuropathy and inflammation.^{82,83} NaV1.8 and NaV1.9 are tetrodotoxin-resistant voltage-gated sodium channels expressed in nociceptors. NaV1.8-null mice have decreased behavioral responses to thermal and mechanical stimuli, indicating a role for nociception for this particular channel,⁸⁴ but a clearer role for NaV1.9 has been more difficult to discern.⁷⁹ In pilot data collected by our group, we demonstrated that HSV vectors expressing zinc finger protein transcription repressors targeted against NaV1.8 can reduce transcription of this protein *in vitro* and reduce nociceptive behaviors in a rat model of neuropathic pain.⁸⁵

Future outlook

As chronic pain continues to be a major health issue that is underserved with currently available drugs, new treatment approaches will be developed and tested. Gene therapy with HSV-based vectors is one such approach and offers the ability to directly act upon the specific neurons and area of the nervous system that are involved in the processing of nociception. However, there remain dosing, safety, and cost issues that will need to be addressed before gene therapy becomes a fully accepted treatment option for the management of pain. Treatment for chronic pain will depend on long-term expression of the therapeutic gene product, which

could be achieved by using a long-term promoter but will more likely involve repeat dosing of the vector product. Our group has demonstrated that readministration of HSV-based vectors is both safe and effective in re-establishing analgesia in rodent models of nociceptive and neuropathic pain.^{47,51,60,71} Long-term expression of a transgene carries its own risks. Once applied, many viral vector-based gene transfer vectors, HSV included, could persist in cells for the life of the patient. This could be quite problematic if the patient had an adverse reaction to the expressed transgene. For this reason, it may be preferential to use short-duration promoters in vector constructs. Alternatively, a regulatable system capable of turning gene expression “on” and “off” could be used.⁸⁶ While such a system has been used in HSV vectors to control the expression of erythropoietin in the treatment of diabetic neuropathy, it has not yet been utilized to treat pain.⁸⁷ A practical concern is one of cost. The process and subsequent costs associated with growing, isolating, and purifying gene transfer vectors are substantially different from those associated with traditional small molecule therapeutics. The ultimate cost per treatment can only be known once indications and target populations have been identified. We believe that the potential increased efficacy of this approach or the ability to treat otherwise intractable patient populations will drive the acceptance of gene therapy to treat pain, and enhanced manufacturing processes and market forces will stabilize the costs to a point acceptable to both patients and payers. Drug products based on HSV vectors that utilize a short-duration promoter such as the human cytomegalovirus promoter would be expected to be readministered every 2–3 months. The choice of transgene would depend on the indication and preclinical animal model studies; as stated above, some transgenes are more efficacious than others in specific pain models.

Several targets, especially within the PAN, have been examined to date, resulting in a substantial amount of preclinical data supporting an HSV-based gene therapy approach in pain management. HSV vectors enable retrograde transduction of PANs via intradermal application but direct injection to nervous system structures is also possible. Zou et al demonstrated that direct injection of an ENK-expressing HSV amplicon vector into the rat ventral periaqueductal gray region resulted in an increased pain threshold in chronic constriction injury-induced neuropathic pain.⁸⁸ Intradermal inoculation of HSV vectors may be sufficient for well localized focal pain, but application to higher nervous system structures such as the periaqueductal gray may be needed to treat more global pain disorders, such as complex regional pain syndrome.

Future studies will be needed to address this issue. To date, preclinical animal studies have focused on targets aiming to treat pain symptoms. Increasing the inhibitory tone at the primary synapse using ENK, endomorphin, or periaqueductal gray-expressing vectors are intended to interfere with pain transmission, but do not alter the underlying pathology. The use of anti-inflammatory vectors may affect central sensitization, but do not necessarily treat the causative problem. As our understanding of the pathophysiology of chronic pain increases, new targets will reveal themselves and vectors designed to treat the cause and not just the symptoms will be developed, tested, and moved into the clinic.

Disclosure

The authors report no conflicts of interest in this work.

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