Glutamate receptor GluAl subunit is implicated in capsaicin induced modulation of amygdala LTP but not LTD

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Capsaicin has been shown to modulate synaptic plasticity in various brain regions including the amygdala. Whereas in the lateral amygdala the modulatory effect of capsaicin on long-term potentiation (LA-LTP) is mediated by TRPVI channels, we have recently shown that capsaicin-induced enhancement of long term depression (LA-LTD) is mediated by TRPMI receptors. However, the underlying mechanism by which capsaicin modulates synaptic plasticity is poorly understood. In the present study, we investigate the modulatory effect of capsaicin on synaptic plasticity in mice lacking the AMPAR subunit GluA1. Capsaicin reduced the magnitude of LA-LTP in slices derived from wild-type mice as previously described, whereas this capsaicin-induced suppression was absent in GluA1-deficient mice. In contrast, neither LA-LTD nor the capsaicin-mediated enhancement of LA-LTD was changed in GluA1 knockout mice. Our data indicate that capsaicin-induced modulation of LA-LTP via TRPV1 involves GluA1-containing AMPARs whereas capsaicin-induced modulation of LA-LTD via TRPM1 is independent of the expression of the AMPAR GluA1 subunit.

The lateral amygdala (LA) receives sensory and cortical information and has been identified as a key structure for fear memory. Fear conditioning and extinction depend on intact long-term increases (LTP) or decreases (long-term depression, LTD) in synaptic efficacy in the LA.

In horizontal slices, LA-LTP and LA-LTD can be elicited by stimulation of external capsule fibers (EC) or intranuclear afferents (intranuclear stimulation, IN) with different induction and expression mechanisms (Drephal et al. 2006; Schubert and Albrecht 2008; Müller et al. 2009). In contrast to coronal slices, stimulation of EC fibers in horizontal slices includes afferents from the lateral entorhinal and perirhinal cortex that also synapse in the LA (von Bohlen und Halbach and Albrecht 2002). Stimulation within the LA also activates local connections within the LA and afferents from other amygdaloid nuclei (von Bohlen und Halbach and Albrecht 2002).

Induction of LA-LTP requires a high increase in intracellular calcium which is, depending on the induction protocols and the stimulated afferents, mainly mediated by NMDA receptors, voltage-gated Ca²⁺ channels (Bauer et al. 2002; Drephal et al. 2006; Fourcaudot et al. 2009) or group I metabotropic GluRs (Lee et al. 2002). LTD induction requires a moderate increase in intracellular calcium due to activation of postsynaptic NMDARs, group II mGluRs or L-type calcium channels (Kaschel et al. 2004; Tchekalarova and Albrecht 2007; Lucas et al. 2013). Previous studies in horizontal slices of the LA have shown that the application of LFS to presynaptic fibers within the LA reliably elicits LTD (intranuclear; IN) (Tchekalarova and Albrecht 2007; Müller et al. 2009), whereas stimulating fibers in the EC does not (Müller et al. 2009).

Excitatory responses in the LA evoked by electrical stimulation of afferents, are predominantly mediated by AMPA receptors (AMPARs). AMPARs are homo- and hetero-tetramers assembled from the subunits GluA1–GluA4 (Traynelis et al. 2010). In the LA, the majority of AMPARs in projection neurons contain the

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GluA2 subunit composed by GLUA1/GluA2 or GluA2/GluA3 hetero-tetramers whereas on interneurons AMPARs lacking GluA2 have been described (Sah and Lopez De Armentia 2003). Insertion or removal of synaptic AMPARs are known to underlie various forms of LTP and LTD in the LA. The GluA1 subunit is critical for the expression of LA-LTP, fear conditioning, anxiety, and aggression (Vekovischeva et al. 2004; Rumpel et al. 2005; Feyder et al. 2007; Humeau et al. 2007; Nedelescu et al. 2010; Tran and Keele 2016).

Members of the transient receptor potential (TRP) channel family, a family of nonselective cation channels, have been shown to modify synaptic plasticity in various brain regions including the hippocampus (Gibson et al. 2008; Li et al. 2008; Chávez et al. 2010; Menigoz et al. 2016), entorhinal cortex (Banke 2016), nucleus accumbens (Grueter et al. 2010), and superior colliculus (Maione et al. 2009). In the LA, we have previously demonstrated that LTP is attenuated by capsaicin via activation of TRPV1 (Zschenderlein et al. 2011) whereas LTD is enhanced by capsaicin via activation of TRPM1 (Gebhardt et al. 2016).

In the present study, we investigate the effect of capsaicin on LA-LTP and LA-LTD in horizontal brain slices from genetically modified mice lacking AMPAR subunit GluA1. Here, we find a significant attenuation of high frequency stimulation (HFS)-induced LTP in slices from homozygous (-/-) and heterozygous (+/-) mice by stimulation of EC or IN fibers as well as an absence of the capsaicin-induced alteration of LA-LTP which was observed in wild-type (wt) mice. In contrast, LA-LTD induced by low frequency stimulation (LFS) was unaltered in slices from homozygous (-/-) and heterozygous (-/-) and heterozygous (+/-) mice and the capsaicin-induced effect on LA-LTD was preserved.

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Results

LA-LTP is significantly reduced in homozygous and heterozygous GluAl knockout mice

GluA1-dependent mechanisms of glutamatergic synaptic plasticity have been shown to be pathway specific in coronal slices. Here, we study LA-LTP in horizontal slices, because LA projection neurons receive different afferent inputs than those in coronal slices: whereas in coronal slices the cortico- and the thalamic LA pathway can be used for stimulation, in horizontal slices LA-LTP also can be induced by stimulation of intranuclear fibers within the LA (IN-LA). We initially tested the I/O curves in slices derived from male and female homozygous GluA1^{-/-}, heterozygous GluA1^{+/-}, and wt GluA1^{+/+} mice to examine the effect of GluA1 on synaptic function in the amygdala using the IN-LA stimulation site. No significant differences in I/ O-curves between the groups or sexes were observed (Fig. 1A,B), which shows that basal transmission was not impaired in the examined LA-afferents in GluA1 mutant mice. Next, we examined whether HFS could induce a stable LA-LTP in horizontal slices of homozygous, heterozygous GluA1 mice and wt GluA1+/+ mice. Consistent with earlier findings from horizontal slices derived from rats or mice (Drephal et al. 2006; Müller et al. 2009), our HFS protocol applied to afferents fibers running within the LA (IN-LA) and to afferents running through the external capsule (EC-LA) induced a significant increase in field potential amplitude in male wt GluA1^{+/+} mice (IN-LA: $146.4 \pm 4.5\%$ [*n* = 9 slices], Fig. 1C; EC-LA: $146.4 \pm 3.8\%$ [*n*=11 slices], Fig. 2A). However, the magnitude of both IN-LA LTP (GluA1^{+/-}: 124.8 ± 4.0% [n=8 slices], Fig. 1E; GluA1^{-/-}: 124.1 ± 5.8% [n=8 slices], Fig. 1G) and EC-LA LTP (GluA1^{+/-}: $129.8 \pm 6.6\%$ [*n*=11 slices], Fig. 2B; GluA1^{-/-}: 120.6±3.9% [n = 10 slices], Fig. 2C) was significantly reduced in male heterozygous and male homozygous GluA1 deficient mice compared with LTP magnitude in wt GluA1^{+/+} mice as illustrated in the bar graph (Figs. 1I, 2D).

The HFS-induced potentiation of the field potential amplitudes (IN-LA) was also observed in female wt GluA1^{+/+} mice (150.7 \pm 7.2% [*n*=8 slices]; Fig. 1D), female GluA1^{+/-} mice (117.9 \pm 6.0% [*n*=12 slices], Fig. 1F) and female GluA1^{-/-} mice (127.3 \pm 6.5% [*n*=8 slices]; Fig. 1H). We did not find any significant difference in IN-LA LTP between males and females.

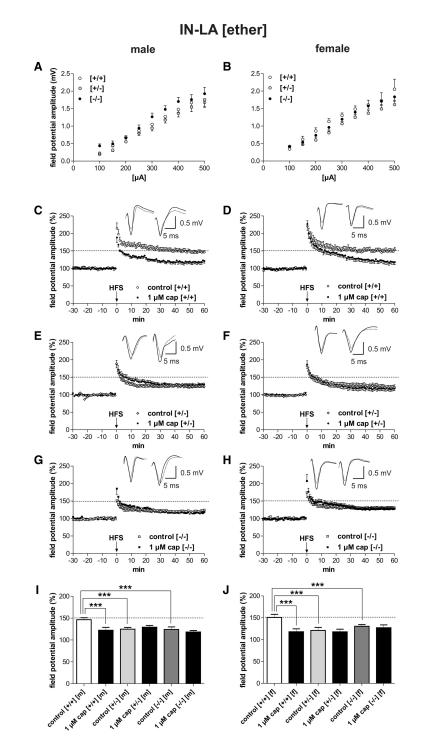


Figure 1. Capsaicin-mediated reduction of LA-LTP induced by intranuclear stimulation (IN) in slices derived from male and female wt mice is absent in GluA1 knockout mice. (*A*) Input/output curves recorded in horizontal brain slices derived from male wild-type (wt) mice (n = 39 slices), GluA1^{+/-} (n = 28 slices), and GluA1^{-/-} knockout mice (n = 41 slices). (*B*) Input/output curves recorded in horizontal brain slices derived from female wt mice (n = 35 slices), GluA1^{+/-} (n = 40 slices). and GluA1^{-/-} knockout mice (n = 35 slices), GluA1^{+/-} (n = 40 slices), and GluA1^{-/-} knockout mice (n = 35 slices). Basal transmission was not significantly different in mutant mice compared with wt. Capsaicin significantly reduced LA-LTP after HFS of fibers running through the LA in male (m) (*C*) and female (f) (*D*) wt mice. (*E*, *F*) The capsaicin-induced suppression of LA-LTP is missing in heterozygous GluA1 knockout mice. (*G*, *H*) The capsaicin-induced suppression of LA-LTP is also missing in homozygous GluA1 knockout mice. Represent averaged amplitudes (mean ± SEM) normalized with respect to baseline values. Representative traces were recorded 5 min prior to tetanus (dashed lines) and 60 min after tetanic stimulation (solid lines) in the LA. (*I*,*I*) Bar histograms of data points averaged 59 to 60 min after HFS, and normalized with respect to baseline (mean ± SEM). Significant differences are indicated. (***) *P*<0.001.

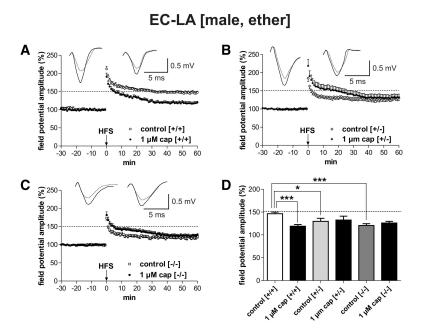


Figure 2. Capsaicin-induced reduction of EC-induced LA-LTP in slices derived from wt mice is absent in GluA1 knockout mice. (*A*) Capsaicin significantly reduced LA-LTP after high-frequency stimulation (HFS) of external capsule fibers (EC) in male wt mice in comparison with controls. (*B*,*C*) The capsaicin-induced suppression of LA-LTP is missing in GluA1^{+/-} and GluA1^{-/-} mice. Data points represent averaged amplitudes (mean ± SEM) normalized with respect to baseline values. Representative traces were recorded 5 min prior to tetanus (dashed lines) and 60 min after tetanic stimulation (solid lines) in the LA. (*D*) Bar histograms of data points averaged 59–60 min after HFS, and normalized with respect to baseline (mean ± SEM). Significant differences are indicated. (*) *P*<0.05; (***) *P*<0.001.

There is no capsaicin-dependent modulation of LA-LTP in homozygous and heterozygous GluAl knockout mice

Our previous data have demonstrated that capsaicin modulates the magnitude of LA-LTP induced by HFS of both EC-fibers and IN-afferents in horizontal slices of C57Bl/6 mice. This capsaicinmediated modulation depends on the isoflurane concentration during anesthesia. We have observed a similar HFS-induced increase in the magnitude of LA-LTP in mice anesthetized with ether or 4% isoflurane before decapitation whereas after anesthesia with high isoflurane concentrations (8%) the magnitude of LA-LTP was significantly smaller (Kulisch et al. 2011). When ether or low isoflurane (4%) concentration were used for anesthesia capsaicin reduced the magnitude of LA-LTP. In contrast, high isoflurane concentrations (8%) cause a sensitization of TRPV1 receptor and its activation by capsaicin led to an enhancement of LA-LTP (Zschenderlein et al. 2011). Please note that all experiments were performed using ether anesthesia if not otherwise stated.

As expected, the magnitude of LA-LTP was significantly reduced by 1 μ M capsaicin in wt littermates (IN-LA: 122.2±6.7% [n=7 slices]; P<0.05; Fig. 1C; EC-LA: 119.0±3.7% [n=10 slices]; P<0.05; Fig. 2A). In contrast, in homozygous and heterozygous GluA1 knockout mice, capsaicin did not show a significant effect on IN-LA LTP (GluA1^{+/-} males: 129.2±4.0% [n=9 slices]; female 117.9±6.0% [n=12 slices]; GluA1^{-/-} males: 118.2±3.4% [n=10 slices]; females: 127.3±6.5% [n=8 slices]; Fig. 1). These results indicate that GluA1 is involved in the mediation of capsaicin-induced effects on synaptic plasticity in the amygdala in both males and females.

Similarly, in slices of homozygous and heterozygous GluA1 knockout mice capsaicin had no significant effect on EC-LA LTP (GluA1^{+/-} control: $129.8 \pm 6.6\%$ [n = 11 slices] versus cap: $132.6 \pm 8.4\%$ [n = 15 slices]; GluA1^{-/-} control: $120.6 \pm 3.9\%$ [n = 10 slices]

versus cap: $126.1 \pm 3.7\%$ [n = 17 slices]). Thus, we could not find differences in the involvement of GluA1 subunit in the mediation of TRPV1 activation depending on the input being stimulated.

After high isoflurane anesthesia EC-LA LTP was facilitated by pretreatment with capsaicin in male wild-type mice (drug-free control: $127.6 \pm 3.6\%$ [n = 9 slices] versus cap: $154.0 \pm 7.3\%$ [n = 9 slices]; P<0.05; Fig. 3A,C). The capsaicin-induced enhancement of LA-LTP in the high-isoflurane group could be blocked by the TRPV1 antagonist AMG 9810 (130.1 \pm 5.3% [*n* = 9 slices]; Fig. 3A) and was not observed in brain slices derived from male GluA1^{-/-} mice (drug-free control: $120.5 \pm 7.3\%$ [*n* = 8 slices]; Fig. 3B,C). The pretreatment of slices with AMG 9810 and capsaicin did not cause a significant change in the magnitude of LA-LTP in GluA1 knockout mice (AMG $9819 + \text{cap:} 116.8 \pm 7.5\% [n = 9 \text{ slices}]).$ These results, which are summarized in Figure 3C, also support the suggestion that the GluA1 subunit is involved in the mediation of capsaicin-induced effects on LA-LTP.

Genetic deletion of GluAl had no effect on LTD or capsaicin-induced modulation of LTD

We have recently shown that the application of low-frequency stimulation (LFS, 900 pulses at 1 Hz) elicits a strong LTD when stimulating presynaptic fibers within the LA (intranuclear; IN) and that capsaicin significantly enhanced this form of LFS-induced LTD via activation of TRPM1 receptors. Here, we examined whether this LFS-induced IN-LA-LTD is impaired in GluA1 knockout mice. As shown in Figure 4, we did not find any significant differences in IN-LA-LTD between the wt-littermates, heterozygous and homozygous mice or between sexes. In all animal groups, capsaicin caused a significant enhancement of LA-LTD (male +/+: $82.2 \pm 5.1\%$ [n = 9 slices] versus cap: $69.6 \pm 3.7\%$ [*n* = 10 slices], *P* = 0.007, Fig. 4A; +/-: 87.2 \pm 4.9\% [*n* = 8 slices] versus cap: $75.5 \pm 4.3\%$ [n = 9 slices], P = 0.02, Fig. 4C; $-/-: 84.5 \pm 5.0\%$ [n = 7 slices] versus cap: 73.5 ± 5.1% [n = 8 slices], P = 0.04, Fig. 4E; females +/+: 86.7 ± 3.2% [n = 8 slices] versus cap: $74.3 \pm 6.7\%$ [*n* = 7 slices], *P* < 0.0001, Fig. 4B; +/-: 85.6 ± 3.7\% [*n* = 10 slices] versus cap: $72.6 \pm 1.7\%$ [*n* = 7 slices], *P* = 0.0007, Fig. 4D; $-/-: 89.6 \pm 7.7\%$ [n = 7 slices] versus cap: 72.2 ± 2.2% [n = 9 slices], P = 0.002, Fig. 4F). Our results suggest that, in contrast to LA-LTP, LA-LTD and capsaicin-induced effects on LA-LTD are independent of GluA1-containing AMPARs.

Discussion

The present study shows that LA-LTP was impaired in horizontal brain slices of homo- and heterozygous GluA1 knockout mice. This impairment was independent of stimulation site and activation of TRPV1 receptor did not modulate LA-LTP in these slices.

Furthermore, we did not observe significant gender differences in the magnitude of LA-LTP which is in line with previous data from rat (Drephal et al. 2006) or mice slices (Staschewski et al. 2011) using high frequency stimulation as LTP induction protocol.

EC-LA [male, high lsoflurane]

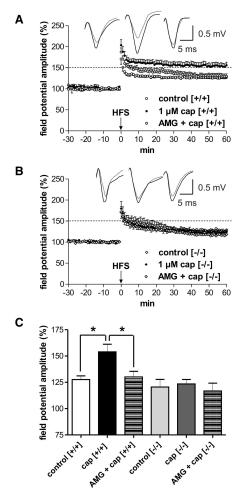


Figure 3. In slices derived from male mice anesthetized with 8% isoflurane before decapitation LTP-enhancing effect of capsaicin after EC stimulation was absent in GluA1 knockout mice. (A) Deep isoflurane anesthesia before decapitation caused a reduction of LA-LTP in slices derived from male wt mice. In these slices capsaicin evoked a strong increase in the magnitude of LA-LTP. The capsaicin-induced enhancement of LA-LTP could be blocked by the specific TRPV1 antagonist AMG9810. (B) In male GluA1 knockout mice the capsaicin-induced effect on LA-LTP was absent. The co-application of the TRPV1 antagonist together with capsaicin did not change the magnitude of LA-LTP. Data points represent averaged amplitudes (mean \pm SEM) normalized with respect to baseline values. (C) Bar histograms of data points averaged 57 to 60 min after HFS, and normalized with respect to baseline (mean \pm SEM). Significant differences are indicated. (*) P < 0.05.

It is known that hippocampal synaptic structure and function exhibit marked variations during the estrus cycle of female rats (Warren et al. 1995; Prange-Kiel et al. 2009; Sabaliauskas et al. 2015). We have shown that rat amygdala slices prepared from females in proestrus or estrus exhibited greater LA-LTP than slices from females in diestrus (Schubert et al. 2008). The results of our present investigation suggest that LA-LTP obtained during different states of the estrus cycle in female mice are responsible for the missing gender difference.

Conversely, we found that LA-LTD and the modulatory effect of capsaicin on LA-LTD were unchanged in GluA1 knockout mice. Our data also indicate that basal synaptic transmission was intact in GluA1-deficient mice because we did not find significant differences in I/O curves in slices derived from GluA1-deficient and wild-type mice. Presumably, removal of GluA1 containing AMPA receptors mainly affects extrasynaptic receptors in LA neurons whereas remaining and compensatory GluA2/GluA3-containing receptors are transported and targeted to synapses, potentially acting primarily to maintain basal transmission as suggested previously by for hippocampal neurons (Zamanillo et al. 1999).

Impairment of LTP in GluA1 deficient mice was previously reported for CA3 to CA1 synapses (Zamanillo et al. 1999) as well as for thalamic and cortical inputs to LA in coronal slices (Humeau et al. 2007), whereas LTP was unaltered in cortical pyramidal neurons of adult mice (Frey et al. 2009). In our experiments performed in mature adult mice slices LA-LTP was reduced but still inducible in heterozygous and homozygous GluA1-deficient mice using both IN- and EC-stimulation. Recently it has been reported that a strong induction protocol is able to induce LTP in CA1 neurons independent of AMPAR subunit type as long as a sufficient number of extrasynaptic surface-expressed receptors can be recruited to the postsynaptic membrane (Granger et al. 2013). Thus, in GluA1 knockout mice the decreased number of extrasynaptic surfaceexpressed AMPA receptors might account for the LTP impairment in CA1 neurons. However, it is unclear whether trafficking and surface expression of synaptic AMPA receptors in LA neurons are similarly regulated as in hippocampal synapses. For instance, it has been recently hypothesized that synaptic recruitment of AMPA receptors from extracellular sites is regulated differently in hippocampal and visual cortical synapses, which indicates that synaptic AMPA receptor regulation might depend on brain region specific mechanisms (He et al. 2011).

Here, we show for the first time that the modulatory effect of capsaicin on the magnitude of LA-LTP is dependent on AMPAR. In a previous study, we found that capsaicin-induced attenuation of LA-LTP is absent in nNOS^{-/-} mice and that it can be blocked by application of the unspecific NOS antagonist L-Name (Zschenderlein et al. 2011). However, NOS inhibitors and genetic deletion of neuronal NOS decrease binding of GluA1 to stargazin, a member of the family of transmembrane AMPAR regulatory proteins (TARPs), which mediates surface expression of AMPARs (Selvakumar et al. 2009). Our data indicate that the capsaicin-induced changes in LA-LTP are selective for GluA1-containing AMPARs, suggesting that capsaicin might inhibit surface expression of GluA1 containing AMPARs via the NOS pathway.

Sensitization of TRPV1 by a high dosage of isoflurane before decapitation of mice causes capsaicin to increase LA-LTP instead of decreasing it (Zschenderlein et al. 2011). This increase of LA-LTP could be blocked by a specific TRPV1 receptor antagonist and was not present in GluA1 knockout mice. A similar effect of capsaicin on regulation of synaptic AMPARs was observed in nonpeptidergic C-fibers. In a capsaicin model of acute inflammatory hyperalgesia, synapses exhibited potentiation due to a significantly higher density of GluA1 containing AMPARs after capsaicin application (Larsson and Broman 2008). In accordance with these observations, GluA1 knockout mice show reduced acute inflammatory hyperalgesia (Hartmann et al. 2004).

In contrast to our results obtained in LTP experiments, LA-LTD induced by LFS of fibers running through the LA deriving not only from the cortical areas but also from the basolateral nucleus of the amygdala was unaffected in GluA1 deficient mice. As for LTP the role of AMPARs in LTD is best investigated in hippocampal neurons. Although previous studies suggested that dephosphorylation of GluA1 subunit may have a role in hippocampal LTD (Lee et al. 2003, 2010), LTD was still inducible in CA1 pyramidal neurons where GluA1 subunit was constitutively knocked out (Granger and Nicoll 2014) as well as in hippocampal slices with genetic deletion of GluA1 (Selcher et al. 2012).

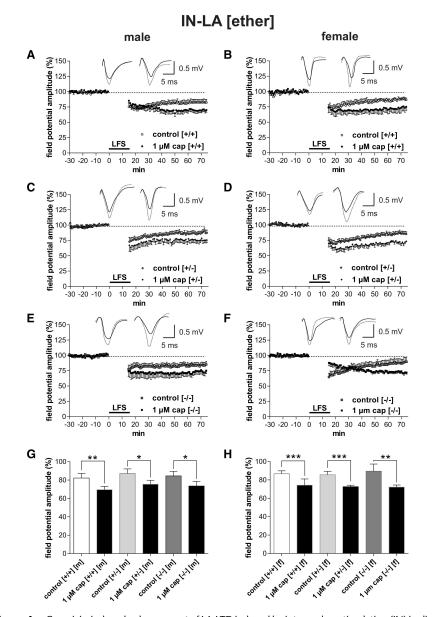


Figure 4. Capsaicin-induced enhancement of LA-LTD induced by intranuclear stimulation (IN) in slices derived from male and female wt mice is preserved in GluA1 knockout mice. Capsaicin significantly enhanced LA-LTD after LFS of fibers running through the LA in male (m) (*A*) and female (f) (*B*) wt mice. (*C*, *D*) The capsaicin-induced enhancement of LA-LTD is preserved in heterozygous GluA1 knockout mice. (*E*,*F*) The capsaicin-induced enhancement of LA-LTD is also preserved in homozygous GluA1 knockout mice. U*(E*,*F)* The capsaicin-induced enhancement of LA-LTD is also preserved in homozygous GluA1 knockout mice. Data points represent averaged amplitudes (mean \pm SEM) normalized with respect to baseline values. Representative traces were recorded 5 min prior to tetanus (dashed lines) and 60 min after tetanic stimulation (solid lines) in the LA. (*G*,*H*) Bar histograms of data points averaged 59 to 60 min after LFS, and normalized with respect to baseline (mean \pm SEM). Significant differences are indicated. (*) *P*<0.05; (**) *P*<0.01; (***) *P*<0.001.

In a previous study, we have identified several physiological mechanisms, which contribute to the capsaicin-induced enhancement of LA-LTD (i.e., antagonists of Group I mGluR, NMDAR, and TRPC channels block capsaicin-induced LA-LTD). Furthermore, we showed an involvement of GABAergic interneurons (Gebhardt et al. 2016). Our finding that neither LA-LTD nor the capsaicin-induced enhancement of LA-LTD depend on GluA1 AMPAR subunit in LA neurons supports the hypothesis that reduction in synaptic strength is independent of glutamate receptor subtype in the LA.

Materials and Methods

Animals and housing

The gene-targeted mouse lines for the GluA1 knockout were generated in the laboratory of R. Sprengel and have been described in detail (Zamanillo et al. 1999). The GluA1 knockout mice are kept as a heterozygous line. For electrophysiological experiments, the colonies of wild-type and knockout mice were simultaneously produced by pairing heterozygous mice from this line. We monitored breeding continuously by assessing the genetic status of the animals by performing PCR on the tail-tip of each mouse. At the time of the extracellular electrophysiological recordings, wt and GluA1 knockout mice were 3 to 5 mo old. To preclude bias, the experimenters measuring LTP or LTD were blinded to the genotype of the mice analyzed. Animals were housed in standardized conditions with an artificial 12-h darklight cycle, room temperature of 22°C and approximately 80% humidity. Mice had free access to food and water. In the past, the extent of sex-biased effects in mammals was obscured by the fact that most investigators examined animals of only one sex (typically males) or did not specifically mention the sex of the tested animals (Jazin and Cahill 2010). Moreover, it has been shown that an interaction between gonadal steroids and glutamate involves hormone regulation of glutamate receptors (Diano et al. 1997; Palomero-Gallagher et al. 2003). To consider this issue, we performed our electrophysiological experiments partly both in males and females. The estrus cvcle was not monitored in intact, cycling mice. All of the experimental protocols were approved by government authorities (Landesamt für Gesundheit und Soziales Berlin ID: T0344/05 and T0381/ 11) and performed according to the German Animal Welfare Act of May 25, 1998, and the European Communities Council Directive of November 24, 1986 (86/609/EEC). All efforts were made to minimize suffering. Between three and eight animals were used for each extracellular experiment (*n* = number of slices).

Electrophysiology

The mice were anesthetized with ether and decapitated (Kulisch et al. 2011). Because isoflurane can cause TRPV1 sensitization (Harrison and Nau 2008), mice were anesthetized with 8% isoflur-

ane for sensitization (Kulisch et al. 2011) and decapitated in an additional series. The brains were quickly removed and placed in ice-cold artificial cerebrospinal fluid (ACSF) (in mM: 129 NaCl, 3 KCl, 1.6 CaCl₂, 1.8 MgSO₄, 1.25 NaH₂PO₄, 10 glucose, and 21 NaHCO₃). For extracellular recordings, hemisected horizontal slices (400 µm) were prepared using a vibroslicer (Campden Instruments) and placed in an interface chamber, where they were allowed to equilibrate for 120 min at 35°C. The slices were superfused continuously with ACSF (1.2 mL/min). The pH was maintained at 7.4 (95% O₂ and 5% CO₂). Glass microelectrodes (Science Products) were filled with ACSF (tip resistance 3 MΩ)

and placed in the LA to record field potentials. Bipolar stimulation electrodes were used to stimulate fibers within the LA (IN) or in the external capsule (EC). An input/output response curve was constructed by varying the intensity of the single-pulse stimulation and averaging six responses per intensity. The stimulus intensity that evoked a field potential with amplitude equal to 50% of the maximal response was then used for all subsequent stimulations. Once a stable baseline of responses had been obtained for at least 20 min, either HFS (two trains at 100 Hz, 1 sec duration, 30 sec apart) was delivered as to induce LTP or low frequency stimulation (LFS; 1 Hz, 15 min) was applied to induce LTD (Müller et al. 2009). Subsequent responses to single stimuli were recorded for at least 60 min, and their amplitude quantified as a percent change with respect to baseline. The HFS paradigm was chosen because theta burst stimulation did not produce consistent and reliable induction of LA-LTP.

Drug application

All drugs were bath-applied at the indicated concentrations starting at least 30 min before HFS or LFS. Capsaicin and AMG 9810 was obtained from Tocris Bioscience.

Data analysis

Extracellular data were collected and averaged using the custommade software Signal 2 (Cambridge Electronic Design). We defined the field potential amplitude as the absolute DC voltage of a vertical line running from the minimal point of the field potential to its intersection with a line running tangential to the field potential onset and offset points. The significance of differences between groups (n=number of slices) was calculated with a two-way ANOVA (GraphPad Prism 5; GraphPad software). Significance was set at P < 0.05. To express and compare changes in the amplitudes of the field potentials between the animal groups, we averaged responses from the period of 59 to 60 min after HFS or LFS.

Acknowledgments

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