



Complete Genome Sequence of *Klebsiella pneumoniae* Myophage Muenster

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ABSTRACT *Klebsiella pneumoniae* is associated with antibiotic-resistant nosocomial infections. Here, we present the annotated genome sequence of the *Klebsiella* jumbo phage Muenster. The Muenster genome sequence (346,937 bp) encodes 6 tRNAs and 561 putative protein-coding genes, including 9 tail fibers, suggesting a genetic mechanism to broaden the host range.

Klebsiella pneumoniae is a Gram-negative opportunistic pathogen that causes severe nosocomial infections and is likely responsible for the spread of carbapenem resistance (1). Increasing drug resistance has led to the need for alternative treatment options such as phage and phage-derived treatments. We isolated and annotated the genome sequence of the *Klebsiella* phage Muenster to investigate a unique group of phages, known as jumbo phages (genomes larger than 200 kbp) (2), and assess their potential for therapy.

Muenster was isolated from wastewater collected from College Station, TX (GPS coordinates, 30.616035, −96.279732), by the soft-agar overlay method (3) and grown on carbapenem-resistant *K. pneumoniae* clinical isolate 44819 (GenBank accession no. [NZ_LDDU000000000](https://www.ncbi.nlm.nih.gov/nuclot/NZ_LDDU000000000)) aerobically at 37°C using tryptic soy medium. Transmission electron microscopy imaging performed at the Texas A&M Microscopy and Imaging Center by negative staining with 2% (wt/vol) uranyl acetate showed a myophage morphology for Muenster (4). Phage genomic DNA was prepared using a modified Promega Wizard DNA cleanup kit protocol as described previously (5). A DNA library was prepared using a TruSeq Nano kit with 300-bp inserts and then sequenced with the Illumina iSeq100 platform with paired-end 2 × 150-bp reads. The reads obtained (1,000,282 in total) were analyzed by FastQC v0.11.19 (www.bioinformatics.babraham.ac.uk/projects/fastqc) for quality control and then assembled into a single contig with 152-fold coverage using SPAdes v3.5.0 (6). The genome was closed by PCR and Sanger sequencing off the ends of the contig using a primer set (forward, 5′-GCACTATGGGTGTTGGTGGG-3′; reverse, 5′-TCAGCGTCGTCTGCATCAAA-3′). Putative protein-coding genes were annotated using GLIMMER v3 (7) and MetaGeneAnnotator v1.0 (8), and tRNAs were detected with ARAGORN v2.36 (9). Sequence homology analysis from BLASTp v2.9.0 (10) against the NCBI nonredundant (nr), Swiss-Prot, and TrEMBL databases (11), conserved folding analysis using HHpred v3.2.0 (12), and conserved domain analyses using InterProScan v5.33 (13) and TMHMM v2.0 (14) were used to assign the putative gene functions. TransTermHP (15) was used to predict the rho-independent terminators. Minus HHpred, all tools were accessed in the CPT Galaxy-Apollo interface (<https://cpt.tamu.edu/galaxy-pub>) (16–18). Default settings were used for all analyses.

Muenster has a 346,937-bp genome sequence with 561 putative protein-coding genes and 6 tRNAs for a 91% coding density. The genome has a 31.9% GC content, which is significantly lower than that of its host at ~57.7% (19). The genome sequence was reopened to reflect a 20,030-bp direct terminal repeat predicted by PhageTerm (20). Genome-wide sequence similarity analysis using progressiveMauve v2.4 (21)

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showed that Muenster is most closely related to other *Klebsiella* jumbo phages, K64-1 (GenBank accession no. [AB897757](#)) and vB_KleM-RaK2 ([JQ513383](#)), with ~90% nucleotide sequence similarity. This suggests that Muenster belongs to the genus *Alcyoneusvirus*. Four hundred forty-six genes encoded hypothetical proteins. Thirty-six coding DNA sequences (CDSs) had significant similarity to T4 proteins with annotated functions, and five more had high-probability HHpred scores for T4 proteins. The Muenster genome sequence has a 20,030-bp direct terminal repeat, but its large terminase is closely related to T4 gp17, despite the T4 packaging having a headful-packaging mechanism. Muenster encodes nine putative tail fibers, most of which are followed by transcriptional terminators. This suggests that the phage uses a controlled genetic mechanism to broaden the host range by modifying the tail fibers (22).

Data availability. The genome sequence of phage Muenster was deposited under GenBank accession no. [MT708547](#) and BioSample accession no. [SAMN14646297](#). The BioProject accession no. is [PRJNA222858](#), and the SRA accession no. is [SRR11575707](#).

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REFERENCES

- Lee C-R, Lee JH, Park KS, Kim YB, Jeong BC, Lee SH. 2016. Global dissemination of carbapenemase-producing *Klebsiella pneumoniae*: epidemiology, genetic context, treatment options, and detection methods. *Front Microbiol* 7:895. <https://doi.org/10.3389/fmicb.2016.00895>.
- Yuan Y, Gao M. 2017. Jumbo bacteriophages: an overview. *Front Microbiol* 8:403. <https://doi.org/10.3389/fmicb.2017.00403>.
- Adams MH. 1959. Bacteriophages. Interscience Publishers, Inc., New York, NY.
- Valentine RC, Shapiro BM, Stadtman ER. 1968. Regulation of glutamine synthetase. XII. Electron microscopy of the enzyme from *Escherichia coli*. *Biochemistry* 7:2143–2152. <https://doi.org/10.1021/bi00846a017>.
- Summer EJ. 2009. Preparation of a phage DNA fragment library for whole genome shotgun sequencing. *Methods Mol Biol* 502:27–46. https://doi.org/10.1007/978-1-60327-565-1_4.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27:4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
- Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. *DNA Res* 15:387–396. <https://doi.org/10.1093/dnares/dsn027>.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
- The UniProt Consortium. 2018. UniProt: the universal protein knowledge-base. *Nucleic Acids Res* 46:2699. <https://doi.org/10.1093/nar/gky092>.
- Zimmermann L, Stephens A, Nam S-Z, Rau D, Kubler J, Lozajic M, Gabler F, Soding J, Lupas AN, Alva V. 2018. A completely reimplemented MPI bioinformatics toolkit with a new HHpred server at its core. *J Mol Biol* 430:2237–2243. <https://doi.org/10.1016/j.jmb.2017.12.007>.
- Jones P, Binns D, Chang H-Y, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong S-Y, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30:1236–1240. <https://doi.org/10.1093/bioinformatics/btu031>.
- Krogh A, Larsson B, von Heijne G, Sonnhammer ELL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 305:567–580. <https://doi.org/10.1006/jmbi.2000.4315>.
- Kingsford CL, Ayanbule K, Salzberg SL. 2007. Rapid, accurate, computational discovery of Rho-independent transcription terminators illuminates their relationship to DNA uptake. *Genome Biol* 8:R22. <https://doi.org/10.1186/gb-2007-8-2-r22>.
- Afgan E, Baker D, Batut B, van den Beek M, Bouvier D, Cech M, Chilton J, Clements D, Coraor N, Gruning BA, Guerler A, Hillman-Jackson J, Hiltmann S, Jalili V, Rasche H, Soranzo N, Goeds J, Taylor J, Nekrutenko A, Blankenberg D. 2018. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2018 update. *Nucleic Acids Res* 46:W537–W544. <https://doi.org/10.1093/nar/gky379>.
- Ramsey J, Rasche H, Maughmer C, Criscione A, Mijalis E, Liu M, Hu JC, Young R, Gill JJ. 2020. Galaxy and Apollo as a biologist-friendly interface for high-quality cooperative phage genome annotation. *PLoS Comput Biol* 16:e1008214. <https://doi.org/10.1371/journal.pcbi.1008214>.
- Lee E, Helt GA, Reese JT, Munoz-Torres MC, Childers CP, Buels RM, Stein L, Holmes IH, Elisk CG, Lewis SE. 2013. Web Apollo: a Web-based genomic annotation editing platform. *Genome Biol* 14:R93. <https://doi.org/10.1186/gb-2013-14-8-r93>.
- Wu K-M, Li L-H, Yan J-J, Tsao N, Liao T-L, Tsai H-C, Fung C-P, Chen H-J, Liu Y-M, Wang J-T, Fang C-T, Chang S-C, Shu H-Y, Liu T-T, Chen Y-T, Shiau Y-R, Lauderdale T-L, Su I-J, Kirby R, Tsai S-F. 2009. Genome sequencing and comparative analysis of *Klebsiella pneumoniae* NTUH-K2044, a strain

- causing liver abscess and meningitis. *J Bacteriol* 191:4492–4501. <https://doi.org/10.1128/JB.00315-09>.
20. Garneau JR, Depardieu F, Fortier L-C, Bikard D, Monot M. 2017. Phage-Term: a tool for fast and accurate determination of phage termini and packaging mechanism using next-generation sequencing data. *Sci Rep* 7:8292. <https://doi.org/10.1038/s41598-017-07910-5>.
21. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 5:e11147. <https://doi.org/10.1371/journal.pone.0011147>.
22. Sandmeyer H. 1994. Acquisition and rearrangement of sequence motifs in the evolution of bacteriophage tail fibres. *Mol Microbiol* 12:343–350. <https://doi.org/10.1111/j.1365-2958.1994.tb01023.x>.