Research Paper

HYAL2 methylation in peripheral blood as a potential marker for the detection of pancreatic cancer: a case control study

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ABSTRACT

Pancreatic ductal adenocarcinoma (PDAC) is a highly lethal malignancy which is mostly diagnosed in advanced and inoperable stages though surgery remains the only curable therapeutic approach. Early detection markers are urgently needed to improve diagnosis. Altered hyaluronoglucosaminidase 2 gene (HYAL2) DNA methylation in peripheral blood is known to be associated with malignancy at early stage but has not been evaluated in PDAC patients. This study evaluates the association between bloodbased HYAL2 methylation and PDAC by a case-control study with 191 controls and 82 PDAC patients. Decreased methylation of all four investigated HYAL2 methylation sites showed highly significant association with PDAC (odds ratio (ORs) per -10% methylation ranging from 2.03 to 12.74, depending on the specific CpG site, p < 0.0001for all). HYAL2 methylation sites were also distinguishable between stage I&II PDAC (61 subjects) and controls (ORs per-10% methylation from 3.17 - 23.04, p < 0.0001for all). Thus, HYAL2 methylation level enabled a very good discrimination of PDAC cases from healthy controls (area under curve (AUC) = 0.92, 95% Confidence interval (C.I.): 0.88 - 0.96), and was also powerful for the detection of PDAC at stage I&II (AUC = 0.93, 95% C.I.: 0.89 - 0.98). Moreover, the blood-based HYAL2 methylation pattern was similar among PDAC patients with differential clinical characteristics, and showed no correlation with the overall survival of PDAC patients. Our study reveals a strong association between decreased HYAL2 methylation in peripheral blood and PDAC, and provides a promising blood-based marker for the detection of PDAC.

INTRODUCTION

Pancreatic ductal adenocarcinoma (PDAC) is a highly malignant cancer and the fourth leading course of cancer-related mortality with 40,000 deaths in Europe each year [1]. The therapeutic options are devastating and surgery remains the main curative treatment approach for only 20% of patients, whereas the other patients are not operable. Even in the 20% patients who could be treated by radical pancreatico-duonodectomy, the 5-year survival is also just about 30% and chemotherapy had only limited improvement in survival [2–3]. Overall, 10-year survival of PDAC patients is only 1.1% [4]. Since early detection strategies are missing, most patients present clinically with a progressed and incurable disease which has very limited curative therapeutic approaches.

Major risk factors for PDAC are known to be chronic pancreatitis and environmental risk factors such as tobacco use, high caloric diet and alcohol as well as inherited cancer susceptibility syndromes in up to 10% of the cases [5]. Multiple genes that are associated with wellknown hereditary cancer syndromes such as *BRCA1* and *BRCA2*, *PALB2*, *CDKN2A*, *ATM* and the DNA mismatch repair genes were found to be associated with pancreatic ductal adenocarcinoma as well [6]. Recently, inherited mutations in correlation with PDAC have gained focused attention, and a pathogenic *BRCA1* and *BRCA2* mutation was identified in a large cohort in 4.6% among pancreatic ductal adenocarcinoma patients [6]. Nonetheless, those risk stratifications attempts have not yet been implemented in daily clinic to improve diagnostic tools.

There are several approaches to identify new biomarkers and to classify pancreatic cancer carriers from healthy persons. DNA methylation is described as one of the earliest and most common events in the process of cancer development, affecting control of gene transcription and the architecture of the cell nucleus [7, 8]. Recent literature has reported the correlation between aberrant methylation of DNA from peripheral blood and multiple cancers, such as breast, ovarian, head and neck, as well as bladder cancer [9–20].

Hyaluronidases (HYALs) are key regulators of hyaluronan (HA) metabolism, and HYAL2 is known to be expressed in somatic tissue and blood cells and to initiate HA degradation [21], whereas HYAL1 expression in tumor tissue was recently found to be associated with endometrial carcinoma aggressiveness and described as an independent prognostic factor for early disease recurrence [22]. At the somatic level, overexpression of HYAL2 is correlated with a higher occurrence of metastasis and shorter survival of triple negative breast cancer [18]. Whole exome sequencing discovered *HYAL2* mutations in the recurrent B-cell lymphoma [23]. Furthermore, the methylation level of *HYAL2* in tumor tissue has been shown to predict overall and progression-free survival in colorectal cancer [24]. Decreased methylation of *HYAL2* has been reported in the peripheral blood DNA of breast cancer patients and head and neck cancer patients [11, 25].

To investigate the association between blood-based *HYAL2* methylation and PDAC, we hereby conducted a case-control study and analysed the *HYAL2* methylation in PDAC patients with various clinical characteristics (Table 1) and healthy control individuals.

RESULTS

Decreased *HYAL2* methylation is associated with PDAC

HYAL2 methylation in peripheral blood DNA associated to PDAC was evaluated using the HYAL2-A amplicon [25] harboring four CpG sites (CpG 1, CpG 2, CpG 3, CpG 4) for analyses with Sequenom MALDI-TOF (matrix-assisted laser desorption ionization time-of-flight) mass spectrometry. The HYAL2_CpG_3 site showed the most significantly lower methylation levels in PDAC cases than in controls (PDAC cases: median = 0.27 (inter quartile range (IQR) = 0.22-0.33); controls: median = 0.43 (IQR = 0.38-0.46; odds ratio (OR) per -10% methylation = 12.74, 95% C.I. = 6.75-24.04, $p = 4.03 \times 10^{-15}$ by logistic regression adjusted for age and gender, Figure 1A and Table 2). Methylation levels of the other three CpG loci also showed significantly lower levels among the PDAC cases than the controls (all OR per -10% methylation > 2.00, $p < 1.00 \times 10^{-5}$ by logistic regression adjusted for age and gender, Figure 1A and Table 2). The methylation levels of the other three CpG loci in HYAL2-A amplicon [25] were also strongly correlated with the methylation level of HYAL2 CpG 3 (all Spearman rho > 0.65, $p < 4.60 \times 10^{-34}$, Table 2).

Decreased *HYAL2* methylation is associated with PDAC at stage I&II

Since the PDAC patients recruited in this study are part of a surgical collective, more early and resectable cases (Stage I&II) were included than typical for the general PDAC population. 60 out of the 82 PDAC are at the stage I and stage II. Compared to the healthy controls, the status of *HYAL2* methylation was also significantly decreased in PDAC cases at relatively early stage. The methylation status of all *HYAL2* CpG sites was also correlated with stage I&II PDAC as shown by the OR > $3.17 (p < 1.40 \times 10^{-7}$ by logistic regression adjusted for age and gender, Figure 1B and Table 3). Same as the PDAC in general, the most altered methylation for stage I&II PDAC compared to controls was detected in the HYAL2_CpG_3 site (OR = 23.04, 95% C.I. = 9.74 - 54.53, $p = 9.49 \times 10^{-13}$) (Figure 1B and Table 3).

Clinical characteristics	Group	N (%)	Median of age
Tumour size	T1	3 (3.7%)	64.2
	T2	1 (1.2%)	63.1
	Т3	62 (75.6%)	63.2
	T4	4 (4.9%)	50.7
	Unknown	12 (14.6%)	68.5
Lymph node involvement	pN0	16 (19.5%)	61.5
	pN1	54 (65.9%)	63.2
	Unknown	12 (14.6%)	68.5
Status of distant metastasis	M0	66 (80.5%)	62.8
	M1	12 (14.6%)	69.1
	Unknown	4 (4.9%)	67.6
Tumour stage	Stage I	4 (4.9%)	63.7
	Stage II	57 (69.5%)	62.9
	Stage III	4 (4.9%)	50.7
	Stage IV	12 (14.6%)	69.1
	Unknown	5 (6.1%)	68.8
Grading	Grade 1	2 (2.4%)	68.5
	Grade 2	33 (40.2%)	64.4
	Grade 3	22 (26.8%)	60.5
	Unknown	25 (30.5%)	63.6
Gender	Male	49 (59.8%)	63.1
	Female	33 (40.2%)	64.4

Table 1: The clinical characteristics of PDAC patients

HYAL2 methylation has no correlation with the clinical characteristics of PDAC

With the aim to further identify subgroups at risk for PDAC, the relation between blood-based *HYAL2* methylation status and clinical characteristics of PDAC (Table 1) was analysed. The methylation levels of all four CpG sites in HYAL2-A amplicon showed no correlation with tumor size, lymph node involvement, status of distant metastasis, tumor stage, grading as shown in Table 4. This gives further proof that *HYAL2* methylation could detect PDAC regardless of stage, and thus, is suitable for the early detection. We further evaluated the influence of age and gender to *HYAL2* methylation. Interestingly, *HYAL2* methylation showed a significant inverse correlation with older age and a positive correlation with male gender in the control group but not in the case group (Table 5).

HYAL2 methylation has no correlation with the overall survival of PDAC

Among the 82 PDAC patients, 61 had complete follow up for their overall survival record for four years (stage I = 3, stage II = 47, stage III = 2, stage IV = 4). In the four years-time period, 8 PDAC patients were still alive (all 3 Stage I PDAC patients, 3 out of all 9 Stage IIA patients, and 2 out of all 38 Stage IIB patients). 53 PDAC patients died with a median overall survival time of 423 days, range from 32 days to 1475 days. The cox regression analysis was performed to evaluate the influence of individual factors to the overall survival of PDAC patients (tumor size was not analysed in the cox regression, because no statistical power can be reached when only 4 patients had T1 and T2 tumors). In the univariate analysis, positive lymph nodes, status of distant metastasis, and higher grading showed significant

CpG sites	Differences in methylation levels					tions to _CpG_3
	Controls median (IQR)	PDAC cases median (IQR)	OR (95 % CI) * per -10% methylation	<i>p</i> -value *	Spearman rho	<i>p</i> -value
HYAL2_CpG_1	0.35 (0.31-0.40)	0.25 (0.19-0.34)	2.03 (1.49-2.78)	9.20E-06	0.65	4.55E-34
HYAL2_CpG_2	0.27 (0.23-0.31)	0.14 (0.09-0.18)	8.50 (4.96-14.57)	6.61E-15	0.82	1.09E-67
HYAL2_CpG_3	0.43 (0.38-0.46)	0.27 (0.22-0.33)	12.74 (6.75-24.04)	4.03E-15	1.00	-
HYAL2_CpG_4	0.61 (0.56-0.66)	0.47 (0.43-0.55)	5.10 (3.32-7.85)	1.27E-13	0.82	1.79E-66

* Logistic regression, adjusted for age and gender





CpG sites	Controls median (IQR)	PDAC stage I&II median (IQR)	OR (95 % CI) * per -10% methylation	<i>p</i> -value *
HYAL2_CpG_1	0.35 (0.31-0.40)	0.25 (0.20-0.33)	3.17 (2.06-4.86)	1.36E-07
HYAL2_CpG_2	0.27 (0.23-0.31)	0.14 (0.09-0.19)	10.98 (5.76-20.92)	3.19E-13
HYAL2_CpG_3	0.43 (0.38-0.46)	0.28 (0.23-0.34)	23.04 (9.74-54.53)	9.49E-13
HYAL2_CpG_4	0.61 (0.56-0.66)	0.48 (0.43-0.55)	5.12 (3.15-8.27)	3.55E-11

* Logistic regression, adjusted for age and gender

Table 4: The methylation of HYAL2 in PDAC	patients with different clinical characteristics

Clinical	Group (N)	Median of	Median of methylation levels			
characteristics (N)		age	HYAL2_ CpG_1	HYAL2_ CpG_2	HYAL2_ CpG_3	HYAL2_ CpG_4
Tumour size (70)	< pT3 (4)	64.20	0.27	0.16	0.24	0.46
	pT3 & pT4 (66)	62.64	0.25	0.14	0.26	0.48
	<i>p</i> -value *	0.544	0.839	0.693	0.770	0.444
Lymph node	pN0 (16)	61.51	0.27	0.12	0.26	0.47
involvement (70)	pN1 (54)	63.56	0.25	0.14	0.28	0.48
	<i>p</i> -value *	0.611	0.279	0.249	0.788	0.725
Status of distant	M0 (66)	62.73	0.25	0.14	0.26	0.48
metastasis (78)	M1 (12)	69.09	0.30	0.13	0.26	0.45
	<i>p</i> -value *	0.144	0.267	0.811	0.822	0.779
Tumour stage (77)	Stage I&II (61)	63.00	0.25	0.14	0.28	0.48
	Stage III&IV (16)	65.50	0.27	0.16	0.24	0.45
	<i>p</i> -value *	0.899	0.407	0.939	0.403	0.888
Grading (57)	Grade 1&2 (35)	64.42	0.26	0.14	0.28	0.49
	Grade 3 (22)	60.50	0.23	0.15	0.24	0.46
	<i>p</i> -value *	0.422	0.133	0.693	0.111	0.083

influence on the survival time (p < 0.05), whereas gender, age and *HYAL2* methylation levels had no significant correlation with the overall survival of PDAC patients (Table 6). In Figure 2, Kaplan Meier curve also shows that the overall survival time is significantly correlated with the status of lymph nodes, status of distant metastasis and grading. In the following multivariate analysis using the significant factors (p < 0.1) selected from the univariate model, status of distant metastasis and grading correlated significantly with the overall survival of PDAC patients (status of distant metastasis, hazard ratio = 5.97, p = 0.006; higher grading, hazard ratio = 3.52, p = 0.0003, Table 6).

HYAL2 methylation as marker for the detection of PDAC

The possibility of using *HYAL2* methylation as detection marker of PDAC was evaluated with the receiver operating characteristic [26] curve analyses adjusted for age and gender by logistic regression. The methylation levels of all *HYAL2* methylation sites pooled together revealed strong discriminatory power between PDAC cases and controls (area under curve (AUC = 0.92, 95% C.I. = 0.88-0.96, sensitivity 75.6%, specificity 93.7%), and between stage I&II PDAC cases and controls (AUC = 0.93, 95% C.I. = 0.89-0.98, sensitivity 66.7%,

 Table 5: Correlation between HYAL2 methylation and age and gender

CpG sites	Contr	Controls		es
	Spearman rho	<i>p</i> -value	Spearman rho	<i>p</i> -value
HYAL2_CpG_1	-0.159	0.028	0.042	0.711
HYAL2_CpG_2	-0.161	0.026	0.124	0.266
HYAL2_CpG_3	-0.150	0.039	0.088	0.431
HYAL2_CpG_4	-0.107	0.142	0.060	0.593

CpG sites	Controls		Case	28
	Spearman rho	<i>p</i> -value	Spearman rho	<i>p</i> -value
HYAL2_CpG_1	0.142	0.049	0.149	0.181
HYAL2_CpG_2	0.143	0.048	0.036	0.750
HYAL2_CpG_3	0.179	0.013	-0.051	0.652
HYAL2_CpG_4	0.160	0.027	-0.066	0.554

Table 6: Prognostic factors of overall surviva	l in PDAC nationts, r	asult from cox regression model
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Factors	Total event (N)	Univariate		Total event (N)UnivariateM		Multivariat	te
		HR (95 % CI)	<i>p</i> -value	HR (95 % CI)	<i>p</i> -value		
Positive lymph node	48	3.22 (1.36-7.65)	0.008	1.77 (0.66-4.72)	0.253		
Distant metastasis	51	2.31 (1.01-5.28)	0.046	5.97 (1.66-21.47)	0.006		
Higher grading	42	3.52 (1.84-6.75)	0.0001	3.52 (1.79-6.93)	0.0003		
Male gender	53	1.20 (0.69-2.09)	0.514	-	-		
Older age *	53	1.00 (0.58-1.71)	0.985	-	-		
Lower HYAL2_CpG_1 §	53	1.05 (0.61-1.80)	0.866	-	-		
Lower HYAL2_CpG_2 §	53	0.95 (0.55-1.64)	0.840	-	-		
Lower HYAL2_CpG_3 §	53	1.16 (0.67-2.00)	0.590	-	-		
Lower HYAL2_CpG_4 §	53	0.91 (0.53-1.58)	0.745	-	-		

* Age older than or equal to the medium age of the PDAC patients

[§] Methylation level lower than the medium methylation levels of the PDAC patients

specificity 95.3%) (Table 7). Among the four HYAL2 CpG sites, HYAL2_CpG_2 and HYAL2_CpG_3 CpG sites contributed the most distinguish power, and each of the CpG sites could distinguish all the PDAC cases and the I&II PDAC cases from controls with AUC > 0.90 as shown in Table 7.

DISCUSSION

This is the first case-control study that describes a significant correlation between decreased methylation of HYAL2 in peripheral blood among PDAC patients compared to healthy controls. PDAC remains a highly

lethal cancer, with about 80% of the patients diagnosed at advanced and unresectable stage (20% are at stage III, median survival 8-14 months; 60% are at stage IV, median survival 4-6 months). The other resectable patients (stage 0, stage I, stage II and even part stage III) have a median survival of about 20 months after the surgery [27]. In our study, we also observed that the early detection of PDAC could dramatically increase the survival time of patients (in our study, 100% of Stage I, 33.3% of Stage IIA and 5% of Stage IIB patients survived for more than four years). The detection of PDAC at resectable stage (before and including stage II) can improve survival time of patients for more than 10 months; whereas clinicians are devoted to improve 2-3 months' survival of advanced PDAC patients in many Phase III clinical studies [28-29]. Early detection markers for resectable pancreatic neoplasia are urgently needed. However, the carbohydrate antigen (CA) 19-9, the tumor marker of pancreatic cancer, is related to tumor size and has a limited sensitivity for small cancers [30]. The positive predictive value of CA 19-9 is especially low in asymptomatic individuals and is therefore not recommended as screening tool [31]. Genetic analysis found several PDAC specific genes and identifies individuals with a germline mutation associated with predispositions for familial cancers. Among pancreatic cancer patients 3-16% are either syndromic or familial [32]. Therefore, diagnostic tools are urgently needed to provide surveillance strategies for predisposed carriers [32]. Additionally, a sub-classification of gene mutation carriers with an increased risk of developing pancreatic cancer is desirable to identify especially those people at higher risk, in order to develop more individualized surveillance programs.

HYAL2 is known to be increased in proliferative processes and was described as a tumor suppressor gene involved in cell adhesion, cell mobility, chemokinesis, cancer progression, angiogenesis and metastasis [21, 33–37]. Recently, we have shown lower *HYAL2* methylation level and higher HYAL2 gene expression in the peripheral blood of breast cancer patients compared to controls [25]. This is in agreement with previous studies publishing decreased *HYAL2* methylation in the peripheral blood in the patients with head and neck squamous cell carcinoma compared to controls [11]. Here, we report *HYAL2* hypomethylation in the blood of PDAC patients comparing to the healthy controls. It is possible that hypomethylation of *HYAL2* may reflect a broader change in DNA methylation across many CpG sites in the genome

that occurs in patients with certain cancer syndromes. But it is interesting that CpG 3 of HYAL2 showed the most significant correlation to PDAC, whereas CpG 4 of HYAL2 was the most significant site correlated to breast cancer. Studies with larger samples size and multiple cancers will be helpful to understand whether there are cancer type specific methylation patterns. It has been known that the proportion of DNA from cancer cells in blood has a ratio of about 1:1000 comparing to the DNA from blood cells [38], and thus, very likely the change of DNA methylation mainly originates from white blood cells. In our previous study in breast cancer, we suggested that the change of leucocytes subpopulation proportion and the breast cancer associated differential HYAL2 methylation in leucocytes subpopulations (T cells and probably other cell types except B cells) are the main reasons for the origin of breast cancer associated differential methylation in blood [25]. Unfortunately, in our study we only have DNA materials from the whole blood of the PDAC patients and controls, and no fresh blood samples available; therefore we could not tell whether this altered DNA methylation is because of the change in proportion of cells and methylation of this gene is cell type specific. Another way to estimate the cell proportion is via statistical calculation using epigenomewide Array data, which is also not available for us. This is a limitation of our study, which should be further explored in the future. In addition, as no questionnaires to the life style were given to the patients and the blood donors, there was in lack of details to the life style factors of the patients and healthy controls. Except age and gender, other potential methylation related factors, such as tobacco smoking and alcohol drinking are unfortunately not available in our study. In future studies these clinical and life style data of study subjects should be collected.



Figure 2: The association of clinical characteristic with overall survival of PDAC by Kaplan–Meier curves. (A) Association of lymph node with overall survival of PDAC. (B) Association of distant metastatic status with overall survival of PDAC. (C) Association of grading with overall survival of PDAC. Low grading: grading 1 and 2; High grading: grading 3.

All 82 PDAC cases vs. all 191 con	trols		
CpG sites	AUC, 95% CI	Sensitivity (positive predictive value)	Specificity (negative predictive rate)
HYAL2_CpG_1	0.74 (0.66-0.81)	37.80%	98.40%
HYAL2_CpG_2	0.90 (0.85-0.95)	73.20%	94.20%
HYAL2_CpG_3	0.92 (0.88-0.96)	74.40%	94.80%
HYAL2_CpG_4	0.85 (0.80-0.91)	59.80%	92.70%
All HYAL2 CpG sites	0.92 (0.88-0.96)	75.60%	93.70%
60 PDAC cases of Stage I&II vs.	all 191 controls		
CpG sites	AUC, 95% CI	Sensitivity (positive predictive value)	Specificity (negative predictive rate)
HYAL2_CpG_1	0.77 (0.68-0.85)	41.70%	98.40%
HYAL2_CpG_2	0.90 (0.84-0.95)	68.30%	94.20%
HYAL2_CpG_3	0.93 (0.89-0.97)	68.30%	94.80%
HYAL2_CpG_4	0.85 (0.79-0.91)	53.30%	94.80%
All HYAL2 CpG sites	0.93 (0.89-0.98)	66.70%	95.30%

Table 7: The discriminatory power of methylation in HYAL2 to distinguish PDAC cases from healthy controls

* Logistic regression, adjusted for age and gender

The mechanisms of cancer-related aberrant HYAL2 methylation in blood remain unknown. So far, there is also no report about the function of HYAL2 in blood cells. Very recently, copy number variation in the gene region of *HYAL2* was reported to be associated with diffuse large B-cell lymphoma [23], indicating that HYAL2 is functional in the malignancy of B cells. In our study, we disclosed the highly significant difference of HYAL2 methylation between PDAC cases and controls, and suggested its potential in the clinical usage as a marker for the PDAC. We could not find a correlation between the altered HYAL2 methylation in blood cells and the clinical pathological characteristics. The similar HYAL2 methylation levels in the patients with early and advance PDAC, as well as the comparable distinguish power for HYAL2 methylation for the PDAC in general and stage I&II PDAC, give strong indication that blood-based HYAL2 methylation might be sufficient for the detection of resectable PDAC. Our finding of HYLA2 hypomethylation in the blood of PDAC patients by case-control study cannot answer the question of whether methylation marker presents cancer risk or cancer progress. In a prospective cass-cohort analysis of DNA methylation in blood and breast cancer, Xu and colleagues found the change of methylation in women whose blood sample was collected less than 1 year before diagnosis is more pronounced than in women who provided blood in the year before their diagnosis. Although Xu's study indicates that blood-based methylation might be marker for preclinical disease via showing the correlation between marker strength and the time to diagnosis, the hypothesis should be taken with caution due to limited sample size. More prospective casecohort studies with larger sample size will be needed to explore whether methylation is a marker of predisposition to disease or a marker of preclinical disease. In the future studies, it will be interesting to analyse other potential markers. The combination of HYAL2 methylation and other markers, especially the markers representing different pathway or mechanism, might provide a even better panel for the detection of early PDAC, and might even be useful for sub-classification and prognosis [22].

To highlight, this study disclosed the correlation between altered DNA methylation in whole blood (mainly from blood cells) and PDAC, and suggested outstanding power for the detection of PDAC, and has great potential for the detection of early PDAC. In contrast, so far circulating free DNA methylation markers from serum or plasma has shown limited evidence for the clinical usage of PDAC detection due to limited sensitivity to the early stage tumors [39]. Although blood-based HYAL2 methylation has shown significant difference between PDAC patients and controls in our study, the power of this study is limited by the small sample size. An additional remark also for our study is that the methylation difference has been compared to healthy controls but not to nonmalignant pancreatic diseases or other cancers. Validation studies in a second cohort with larger sample sizes and even prospective studies among high risk persons are needed to address the effect of predictive potential in healthy subjects.

MATERIALS AND METHODS

Study population

The present study was approved by the local Ethics Committee of the University of Heidelberg, Germany. Each participant provided written informed consent. All participants were Caucasian. Blood sample was drawn by venipuncture, collected into Li-Heparin-Gel Monovette (S-Monovette® 7.5ml LH; cat# 01.1604.001; Sarstedt AG; Nümbrecht, Germany) and stored at -80oC until DNA isolation. The DNA is extracted by the QIAamp DNA Blood Mini Kit (Cat No./ID: 51104) according to manufacturer's instructions.

Peripheral blood samples were obtained from 82 PDAC patients (stage I = 4, stage II = 57, stage III = 4, stage IV = 12, stage unknown = 5) at the University of Heidelberg University Hospital of Surgery in Heidelberg from September of 2009 to August of 2011. The PDAC patients have a median age of 63.7 years old (range from 39 to 79 years old), and have 49 males (59.8%) and 33 females (40.2%). Since the PDAC patients recruited in this study are part of a surgical collective, more early and resectable cases (Stage I&II) were obtained than typical for the general PDAC population. In PDAC surgery, if metastases are found, the operation is finished as exploration without resection but with histologic confirmation. Thus, several Stage III and Stage IV cases are also included in the study. The clinical characteristics of PDAC were confirmed by pathology in all cases. All blood samples from the enrolled PDAC patients were taken before the surgery and pancreatic cancer related treatment. Detailed clinical data of the PDAC patients are shown in Table 3. In the follow up period, 21 PDAC patients had no or not complete follow up data, whereas the remaining 61 PDAC patients were followed for four years with 53 death event in this period. No progression free survival data is available for the PDCA cases.

Peripheral blood samples from controls were consecutively collected from healthy blood donors (N = 191) by the German Red Cross Blood Service of Baden-Württemberg-Hessen (Mannheim, Germany) from 2004 to 2012 as described elsewhere [25, 40]. The controls have a median age of 61.0 years old (range from 21 to 68 years old), and have 115 males (60.2%) and 76 females (39.8%). No further exclusion or inclusion criteria were applied for the controls.

MALDI-TOF mass spectrometry

MALDI-TOF mass spectrometry (Sequenom) described by Breitling *et al.* [9, 41] was used in all the validation and further exploring rounds. In brief, DNA was bisulfite converted by EZ-96 DNA Methylation Gold Kit (Zymo Research) and amplified by bisulfite-specific primers (no SNPs in the primers) and PCR amplicons HYAL2-A were described as previously used [25]. The

PCR products were used according to the standard protocol of Sequenom EpiTyper Assay, and further cleaned by Resin and dispensed to a 384 SpectroCHIP by a Nanodispenser as described by us before. The chips were read by a Sequenom Mass Spectrometer system. Data were collected by SpectroACQUIRE v3.3.1.3 software and visualized with MassArray EpiTyper v1.2 software. For each batch of MassARRAY analysis, same amount of cases and controls were randomly selected from the cohort [25].

Statistical analyses

All the statistical analyses were conducted by SPSS Statistics 21. Spearman's rank correlation coefficient was used to assess the correlations. The comparisons between two and multiple groups were performed by logistic regression models and non-parametric tests. The logistic regression results were adjusted for possible and available confounding effects such as age and gender by including additional co-variables into the logistic regression models. Receiver operating characteristic [26] curve analysis assessed the discriminatory power of methylation levels. Cox-regression analyses were used to estimate the influence of factors to the survival of PDAC patients. All statistical tests were two-sided, and *p* values < 0.05 were defined as statistically significant.

Abbreviations

AUC: area under curve; CA: carbohydrate antigen: CI: confidence interval; HYAL2: hyaluronoglucosaminidase 2 gene; HR: hazard ratio; IQR: inter quartile range; MALDI-TOF: matrix-assisted laser desorption ionization time-of-flight; PDAC: pancreatic ductal adenocarcinoma; OR: odds ratio

Author contributions

SSc: Drafting the article, Critical revision of the article, Final approval of the version to be published

RY: Data collection, Data analysis and interpretation, Drafting the article, Critical revision of the article, Final approval of the version to be published

SSt: Data collection, Conception or design of the work, Final approval of the version to be published

FC: Data collection, Final approval of the version to be published

NG: Data collection, Final approval of the version to be published

PB: Data collection, Final approval of the version to be published

FB: Data collection, Final approval of the version to be published

OS: Data collection, Data Interpretation, Final approval of the version to be published

TH: Data collection, Final approval of the version to be published

CS: Data collection, Final approval of the version to be published

BB: Conception or design of the work, Supervision of the work, Data interpretation, Critical revision of the article, Final approval of the version to be published.

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CONFLICTS OF INTEREST

Rongxi Yang and Barbara Burwinkel are inventors of a provisional patent application relating to the subject matter of this manuscript and therefore declare a potential conflict of interests.

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