


CASE REPORT

Transformation from adenocarcinoma to squamous cell lung carcinoma with MET amplification after lorlatinib resistance: A case report

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Abstract

To date, several studies have described the mechanism of resistance to first- or second-generation anaplastic lymphoma kinase (ALK) inhibitors. Secondary ALK mutations, ALK gene amplification, and other bypass signal activations (i.e., KRAS mutation, EGFR mutation, amplification of KIT, and increased autophosphorylation of EGFR) are known as resistance mechanisms. However, little has been previously reported on acquired resistance mechanisms to lorlatinib. Here, we report a case of a patient with ALK-positive lung adenocarcinoma that acquired resistance to lorlatinib during treatment for brain metastasis and showed histological transformation to squamous cell carcinoma with MET amplification. We also review the previous literature on the resistance mechanism to ALK inhibitors.

KEYWORDS

Anaplastic lymphoma kinase, lorlatinib, lung cancer, squamous cell lung cancer transformation

INTRODUCTION

Anaplastic lymphoma kinase (ALK) rearrangements are found in 3%–5% of patients with non-small cell lung cancer (NSCLC).^{1,2} Currently, there are four ALK inhibitors approved in Japan by the Pharmaceuticals and Medical Devices Agency, five in Europe approved by the European Medical Agency, and five in the USA approved by the Food and Drug Administration. Based on two phase 3 trials that compared first-line alectinib with crizotinib in ALK-rearranged NSCLC, the standard first treatment for ALK-positive lung adenocarcinoma is alectinib.^{3,4} Additionally, the third-generation ALK inhibitor lorlatinib has been shown to be effective in patients with acquired resistance to first-generation or second-generation ALK inhibitors.⁵

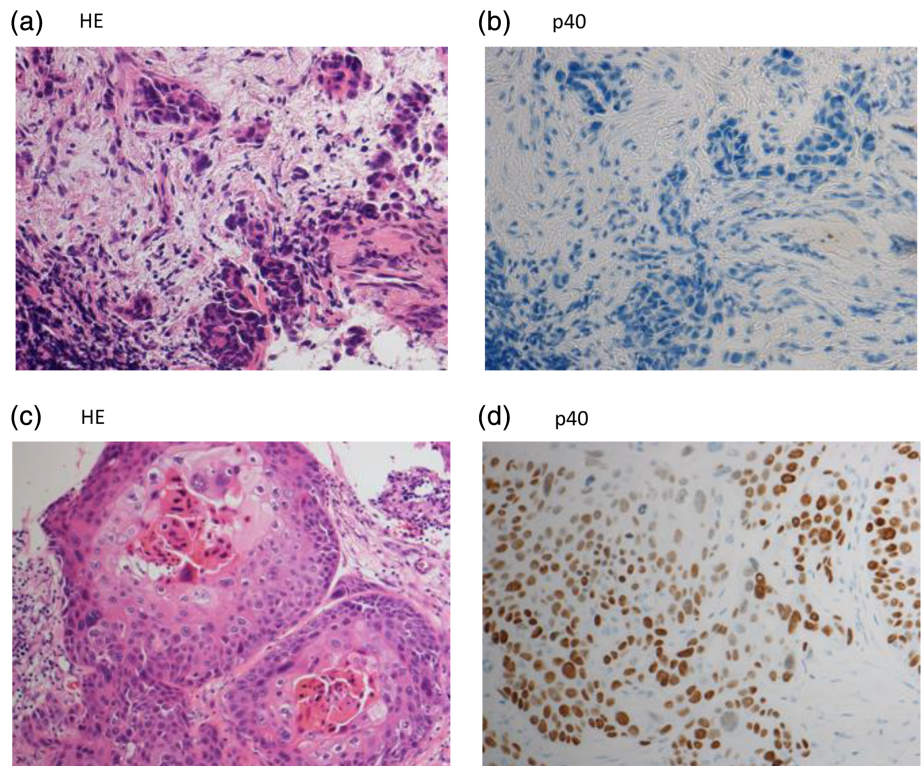
However, there have been few reports on acquired resistance mechanisms to lorlatinib, except for compound mutations in ALK. Here, we report a case of a patient with ALK-positive lung adenocarcinoma that acquired resistance to lorlatinib during treatment for brain metastasis and

showed histological transformation to squamous cell carcinoma with MET amplification.

CASE REPORT

A 58-year-old female non-smoker was diagnosed with clinical T1aN3M0 stage IIIB adenocarcinoma in 2011 (Figure 1 (a), (b)). She subsequently received concurrent chemoradiotherapy (cisplatin plus vinorelbine with thoracic radiotherapy of 60 Gy in 30 fractions). Computed tomography (CT) showed a good partial response after three cycles of this regimen. However, grade 1 radiation pneumonitis was identified on CT, and further consolidation chemotherapy was eventually discontinued. Her lung cancer lesion and radiation pneumonitis were monitored with CT without any treatment. Four months later, radiation pneumonitis had improved on CT, but progressive disease was identified. She received 13 cycles of pemetrexed, five cycles of docetaxel, and six cycles of gemcitabine. Multiple brain metastases were

FIGURE 1 Pathological findings of the patient. (a, b) Hematoxylin and eosin staining showing adenocarcinoma histology with p40 negative expression at diagnosis. (c, d) Brain tumor samples after resistance to lorlatinib showing transformation to squamous cell carcinoma with p40-positive expression



identified on magnetic resonance imaging (MRI), and the lesions were treated with Gamma knife radiosurgery. Additionally, the test for *ALK* rearrangement, approved in Japan in 2012, was performed and her lung cancer specimen was found to have *ALK* rearrangement with fluorescence in situ hybridization (FISH); she subsequently received crizotinib as fifth-line therapy for 21 months. Due to recurrence of thoracic lesions and brain metastases, she was treated with alectinib for 27 months, ceritinib for three months, three cycles of pemetrexed plus bevacizumab, and seven more courses of Gamma knife treatment. However, owing to the recurrence of brain metastases (Figure 2(a)), she received lorlatinib as ninth-line treatment. Four months after initiation of lorlatinib, the brain metastases were under control (Figure 2(b)). During lorlatinib treatment, dose reduction and temporary drug discontinuation were required because of grade 3 edema and grade 2 peripheral sensory neuropathy. Six months after initiation of lorlatinib, she presented at our hospital with hemiparesis. Brain MRI showed an enlargement of the metastatic lesion in the right temporal lobe with severe parenchymal edema (Figure 2(c)). In order to relieve the patient's symptoms, we removed the tumor and the hemiparesis improved. A summary of the treatment course is shown in Figure 3.

Histopathological review showed the cancer cells in the metastatic brain specimen had changed from adenocarcinoma to squamous cell lung carcinoma (Figure 1(c), (d)). Next-generation sequencing (NGS) of 46 oncogenes was performed with Oncomine Dx Target Test (ThermoFisher Scientific), and it showed no secondary mutations of *ALK* or

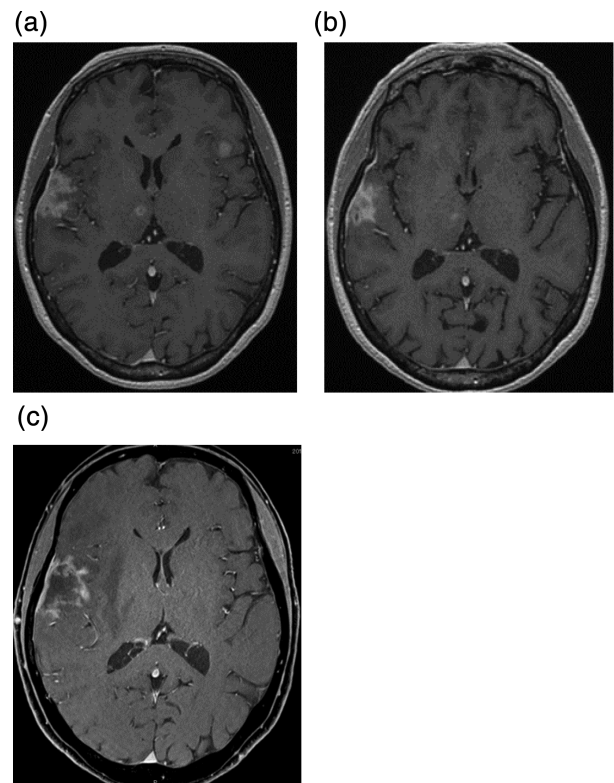


FIGURE 2 Brain magnetic resonance imaging (MRI). T1 weighted images with contrast enhancement. (a) Multiple brain metastases were found at the time after focal radiation. (b) The brain metastases lesions were controlled for four months after lorlatinib treatment. (c) The lesion in the right temporal lobe had enlarged with severe parenchymal edema six months after lorlatinib treatment

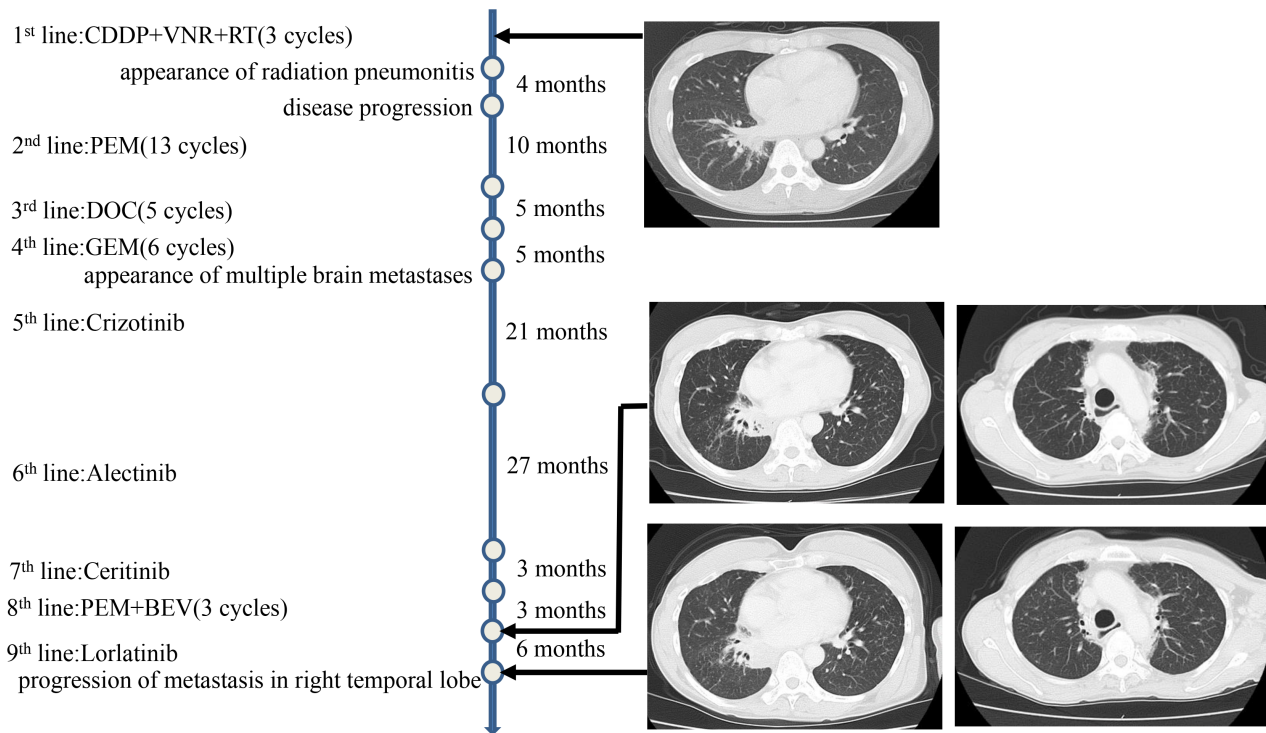


FIGURE 3 Schematic summary of the treatment course. Six months after initiation of lorlatinib, brain-magnetic resonance imaging (MRI) showed progression of metastasis in the right temporal lobe but computed tomography (CT) of the thoracoabdominal region showed stable disease

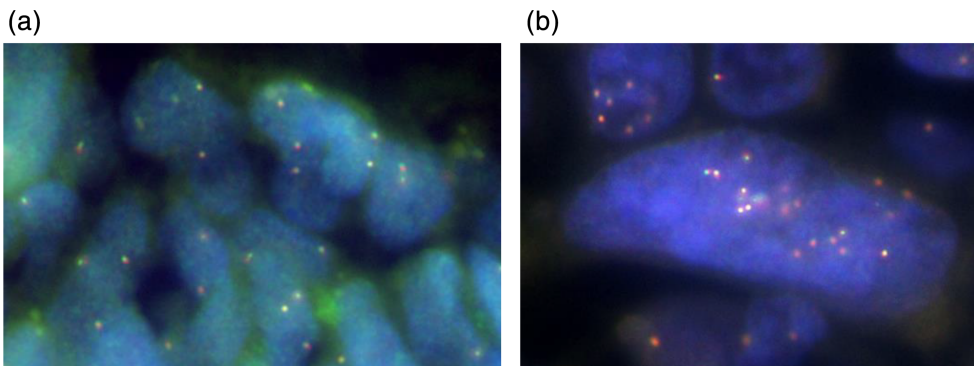


FIGURE 4 (a) Amplification of *MET* was evaluated with fluorescence in situ hybridization in the lung biopsy sample at the time of diagnosis; and (b) in the brain tumor sample after lorlatinib resistance. *MET* amplification was observed in some cells in the sample at the time of diagnosis, but *MET* amplification obviously increased in the sample after lorlatinib resistance

other driver oncogenes. Amplification of *MET* was evaluated with FISH in the samples at the time of diagnosis and after lorlatinib resistance (Figure 4(a), (b)). *MET* amplification was observed in some cells in the sample at the time of diagnosis, but *MET* amplification obviously increased in the sample after lorlatinib resistance.

Lorlatinib was resumed after brain tumor resection and continued until CNS progression was confirmed on brain MRI.

DISCUSSION

We report a case of adenocarcinoma that transformed to squamous cell lung carcinoma with *MET* amplification after resistance to lorlatinib.

There are three generations of ALK inhibitors. Crizotinib is a first-generation ALK inhibitor; ceritinib, alectinib, and brigatinib are second generation inhibitors and lorlatinib is the third generation. Second- and third-generation ALK inhibitors have been developed to overcome resistance to previous generation inhibitors. Secondary *ALK* mutations are major causes of resistance to inhibitors and were found in 20%–35% of tumors resistant to crizotinib.^{6–8} *ALK* gene amplification was also found in 8.3% of crizotinib-resistant tumors, and other bypass signal activations such as *KRAS* mutation, *EGFR* mutation, amplification of *KIT*, and increased autophosphorylation of *EGFR* were found in tumors resistant to crizotinib.^{6–8} Secondary *ALK* mutations were found more frequently in tumors resistant to second-generation ALK inhibitors than in those resistant to first-generation inhibitors.⁸ Two studies recently reported

the mechanism of lorlatinib resistance.^{9,10} One study investigated the mechanism of lorlatinib resistance in longitudinal tumor samples from five patients with *ALK*-positive lung cancer. Similar epithelial-mesenchymal-transition (EMT)-mediated resistance was found in two patients, *ALK* kinase domain compound mutation-mediated resistance was found in two patients, and *NF2* biallelic loss of function mutations were found in one patient.⁹ Another study showed *MET* amplification in six (12%) of 52 biopsies following administration of a second-generation *ALK* inhibitor and in five (22%) of 23 post lorlatinib biopsies. In addition, two tumor specimens harbored an identical *ST7-MET* rearrangement, one of which had concurrent *MET* amplification. Dual *ALK/MET* inhibition resensitized a patient-derived cell line harboring both *ST7-MET* and *MET* amplification to *ALK* inhibitors.¹⁰

Histological transformation after first- or second-generation *ALK* inhibitor treatment has been reported in several studies, wherein most reported on transformation from adenocarcinoma to small-cell lung carcinoma.¹¹ Transformation from adenocarcinoma to squamous cell lung carcinoma was reported after resistance to crizotinib in one case and after resistance to alectinib in another case.^{12,13}

Our study has some limitations. The tumor specimen after lorlatinib treatment was compared with that taken at the diagnosis of adenocarcinoma. Transformation to squamous cell lung carcinoma with *MET* amplification may have been acquired as a result of previous treatments before lorlatinib. However, the metastatic lesion in the brain had initially responded to lorlatinib and progressed during lorlatinib treatment. These two changes could be related to lorlatinib resistance. The results of a previous report also suggest that *MET* amplification could be related to lorlatinib resistance in our case.¹⁰ Additionally, lorlatinib is used as a second or further line chemotherapy in clinical practice. It would be difficult to compare tumor samples taken after lorlatinib with those taken just before lorlatinib treatment.

A pooled analysis was conducted to investigate the characteristics and outcomes of 17 patients with *EGFR*-mutated adenocarcinoma who developed a transformation to squamous cell histology after treatment with *EGFR* tyrosine kinase inhibitors (*EGFR*-TKIs).¹⁴ Most patients were women (82%), 41% were former smokers, and no current smokers were identified. The median time to squamous cell transformation was 11.5 months. In all cases, basal *EGFR* mutation was maintained, 11 patients (65%) developed an acquired mutation in exon 20, and a T790M mutation appeared in eight patients (47%). The median survival after squamous cell carcinoma diagnosis was 3.5 months. In the case reported here, there was no smoking history, basal *ALK* translocation was maintained, and *MET* amplification was found in addition to squamous cell transformation. Progression-free survival of patients treated with crizotinib has been previously reported to be significantly shorter in squamous cell lung carcinoma with *ALK* rearrangement

than in adenocarcinoma with *ALK* rearrangement.¹⁵ This suggests that squamous cell histology can be related to resistance to *ALK* inhibitors. The mechanism of resistance to *ALK* inhibitors, especially lorlatinib, might be multiple and complex. In our case, squamous cell transformation and *MET* amplification were found in the same patient after treatment with lorlatinib. In this case, it may be difficult to overcome resistance with a molecularly targeted agent aiming to inhibit just one molecule. As reported in the phase 3 CROWN trial which compared lorlatinib with crizotinib, and supports lorlatinib as a future first-line standard treatment in *ALK*-positive non-small cell lung cancer, knowing the mechanism of resistance to lorlatinib is becoming more important.¹⁶ Further investigation is needed to overcome resistance to *ALK* inhibitors in patients with *ALK*-positive NSCLC.

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