#### Prognostic Clinical and Biological Markers for Amyotrophic Lateral Sclerosis Disease Progression: Validation and Implications for Clinical Trial Design and Analysis

<u>Authors</u>: Michael Benatar <sup>1</sup>, Eric A Macklin <sup>2</sup>, Andrea Malaspina <sup>3</sup>, Mary-Louise Rogers <sup>4</sup>, Eran Hornstein <sup>5</sup>, Vittoria Lombardi <sup>3</sup>, Danielle Renfrey <sup>4</sup>, Stephanie Shepheard <sup>4</sup>, Iddo Magen <sup>5</sup>, Yahel Cohen <sup>5</sup>, Volkan Granit <sup>1</sup>, Jeffrey M Statland <sup>6</sup>, Jeannine M Heckmann <sup>7</sup>, Rosa Rademakers <sup>8,9</sup>, Caroline A McHutchison <sup>10,11</sup>, Leonard Petrucelli <sup>9</sup>, Corey T McMillan <sup>12</sup>, and Joanne Wuu <sup>1</sup> on behalf of the CReATe Consortium PGB1 Study Investigators

## Affiliations:

- <sup>1</sup> Department of Neurology, University of Miami Miller School of Medicine, Miami, FL, USA
- <sup>2</sup> Departments of Neurology and Medicine, Massachusetts General Hospital, Harvard Medical School, Boston, MA USA
- <sup>3</sup> UCL Queen Square Motor Neuron Disease Center, UCL Queen Square Institute of Neurology, University College London, Queen Square, London, UK
- <sup>4</sup> Flinders Health and Medical Research Institute, College of Medicine and Public Health, Flinders University, Adelaide, South Australia, Australia
- <sup>5</sup> Department of Molecular Genetics and Molecular Neuroscience, Weizmann Institute of Science, Israel
- <sup>6</sup> Department of Neurology, University of Kansas Medical Center, Kansas City, KS USA
- <sup>7</sup> Division of Neurology, Department of Medicine, University of Cape Town, South Africa
- <sup>8</sup> VIB Center for Molecular Neurology, Department of Biomedical Sciences, University of Antwerp, Antwerp, Belgium
- <sup>9</sup> Department of Neuroscience, Mayo Clinic, Jacksonville, Florida, USA
- <sup>10</sup> School of Philosophy, Psychology, and Language Sciences, The University of Edinburgh, Edinburgh, UK.
- <sup>11</sup> Euan MacDonald Centre for Motor Neuron Disease Research, The University of Edinburgh, Edinburgh, UK
- <sup>12</sup> Department of Neurology, University of Pennsylvania Perelman School of Medicine, Philadelphia, PA, USA

<u>Correspondence to</u>: Michael Benatar MD, PhD, or Joanne Wuu, ScM, Department of Neurology, University of Miami Miller School of Medicine, 1120 NW 14 Street, CRB Suite 1300, Miami, FL, 33136, USA. Email: <u>mbenatar@miami.edu</u>; or <u>jwuu@miami.edu</u>.

# Structured Summary (max 300)

1

Background. With increasing recognition of the value of incorporating prognostic markers into amyotrophic lateral sclerosis (ALS) trial design and analysis plans, there is a pressing need to understand *which* among the prevailing clinical and biochemical markers have real value, and *how* they can be optimally used.

6

7 **Methods.** A subset of patients with ALS recruited through the multi-center Phenotype-

8 Genotype-Biomarker study (clinicaltrials.gov: NCT02327845) was identified as "trial-like" based

9 on meeting common trial eligibility criteria. Clinical phenotyping was performed by evaluators

10 trained in relevant assessments. Serum neurofilament light (NfL) and phosphorylated

11 neurofilament heavy (pNfH), urinary p75<sup>ECD</sup>, plasma microRNA-181, and an array of

12 biochemical and clinical measures were evaluated for their prognostic value. Associations with

13 functional progression were estimated by random-slopes mixed models of ALS functional rating

14 scale-revised (ALSFRS-R) score. Associations with survival were estimated by log-rank test and

15 Cox proportional hazards regression. Potential sample size savings from adjusting for given

- 16 biomarkers in a hypothetical trial were estimated.
- 17

Findings. Baseline serum NfL is a powerful prognostic biomarker, predicting survival and
ALSFRS-R rate of decline. Serum NfL <40pg/ml and >100pg/ml correspond to future ALSFRS-R
slopes of ~0.5 and 1.5 points/month, respectively. Serum NfL also adds value to the best
available clinical predictors, encapsulated by the European Network to Cure ALS (ENCALS)
predictor score. In models of functional decline, the addition of NfL yields ~25% sample size
saving above those achieved by inclusion of either clinical predictors or ENCALS score alone.
The prognostic value of serum pNfH, urinary p75<sup>ECD</sup>, and plasma miR-181ab is more limited.

25

Interpretation. Among the multitude of biomarkers considered, only blood NfL adds value to the
 ENCALS prediction model and should be incorporated into analysis plans for all ongoing and
 future ALS trials. Defined thresholds of NfL might also be used in trial design, for enrichment or
 stratified randomisation, to improve trial efficiency.

30

31 **Funding.** NIH (U01-NS107027, U54-NS092091). ALSA (16-TACL-242).

32

33 Key Words: Prognostic biomarkers, Context-of-use, ALS clinical trials, Neurofilament

## 35 Research in Context

36

# 37 Evidence Before This Study

38 The phenotypic heterogeneity of ALS poses a challenge for clinical trials, making it more difficult

- 39 to discern therapeutic effects of investigational agents amidst the noise of natural variability.
- 40 Prognostic markers are important tools to help mitigate this issue. A host of clinical markers and
- 41 putative biomarkers have been proposed to have prognostic value, but their relative utility,
- 42 especially when considered jointly, and the practical implications of their use, have not been well
- 43 defined.
- 44

## 45 Added Value of This Study

- 46 Using a trial-like population from a natural history study, in which clinical trial-grade phenotypic
- 47 data and multi-modal biomarker data were collected, we show that a subset of clinical factors,
- 48 encapsulated by the ENCALS predictive model score, and serum neurofilament light chain (NfL)
- 49 are the most powerful prognostic markers when considering either ALSFRS-R functional decline
- 50 or permanent assisted ventilation (PAV)/tracheostomy-free survival. Importantly, serum NfL adds
- 51 prognostic value even after adjusting for the ENCALS score, yielding an additional sample size
- 52 saving of ~27% in a hypothetical future clinical trial. While serum phosphorylated neurofilament
- 53 heavy chain (pNfH), urinary p75<sup>ECD</sup>, and plasma miR-181ab each holds some prognostic value,
- 54 when considered together with the ENCALS score and serum NfL, only p75<sup>ECD</sup> may yield
- 55 additional but modest sample size saving.
- 56

## 57 Implication of All Available Evidence

- 58 Blood NfL is a validated biomarker for multiple contexts-of-use. As a prognostic marker, it should
- 59 be used together with clinical predictors, such as the ENCALS predictive model score, in all
- 60 ongoing and future ALS clinical trials. The utility of urinary p75<sup>ECD</sup> and plasma miR-181ab is less
- 61 clear. Serum pNfH, as well as serum uric acid, albumin, creatinine, and C-reactive protein
- 62 (CRP), provide no additional prognostic information.
- 63

#### 64 Introduction

65 Clinical trials in the field of amyotrophic lateral sclerosis (ALS) must consider the phenotypic 66 heterogeneity of disease as well as the related challenge that clinically meaningful outcomes. 67 such as the rate of functional decline and survival, are typically insufficiently sensitive to detect 68 therapeutic effect in the early- and mid-phases of drug development. Biofluid biomarkers that 69 are fit-for-purpose, however, may help to meaningfully address this problem.<sup>1</sup> In patients with 70 clinically manifest ALS (as opposed to the pre-symptomatic population at elevated risk for ALS, 71 which is beyond the scope of this paper), prognostic biomarkers might be used in three broad 72 ways to improve the design and analysis of clinical trials. From a study design perspective, they 73 may be used as eligibility criteria to enrich for a population, in which a therapeutic effect might 74 be most apparent, or to stratify randomisation. They may also be used analytically to adjust for 75 phenotypic heterogeneity, thereby reducing the sample size needed to adequately power a trial using clinical outcome measures.<sup>2</sup> These approaches are not mutually exclusive, and indeed 76 77 could be combined, depending on the goals of a particular trial. In addition, response 78 biomarkers might be used to demonstrate target engagement or pharmacodynamic effect, and 79 perhaps even serve as surrogates that are reasonably likely to predict a future clinical benefit. 80

There remains, however, a significant gap between biomarker discovery, analytic validation, and preliminary reports of biomarker performance in samples of convenience on the one hand, and clinical validation on the other hand. The latter entails demonstrating the utility of a biomarker for a well-defined context-of-use in a large, carefully phenotyped clinical cohort.

85

Prior studies have identified clinical parameters predictive of disease progression<sup>2</sup> or survival.<sup>2,3</sup> 86 87 Moreover, among patients with ALS, neurofilament light chain (NfL) has emerged as the lead 88 prognostic and response biomarker <sup>4-8</sup> for a number of reasons: NfL can reliably be measured 89 in blood, there is a high correlation between blood and cerebrospinal fluid (CSF) concentrations, 90 and empiric data support these contexts-of-use based on results from serum or plasma. There 91 is, however, also persistent interest in the potential prognostic value of other biomarkers, including blood phosphorylated neurofilament heavy chain (pNfH);<sup>9</sup> urinary p75 neurotrophin 92 receptor extracellular domain (p75<sup>ECD</sup>);<sup>10</sup> microRNA-181 (miR-181);<sup>11</sup> and an array of analytes, 93 such as uric acid,<sup>12-20</sup> albumin,<sup>16</sup> creatinine,<sup>16,20,21</sup> and C-reactive protein (CRP),<sup>8,22,23</sup> that are 94 95 routinely quantified in the clinical arena.

97 In this study, we sought to clinically validate the utility of putative prognostic biofluid biomarkers 98 in the context of established clinical prognostic factors. The rationale is that a prognostic 99 biomarker would only be worth quantifying if it adds value to what can be learned from known 100 and readily available clinical parameters. Furthermore, a head-to-head comparison of clinical 101 markers and molecular biomarkers revealed their relative contributions to clinical trial design, 102 analysis, and result interpretation. Finally, we characterised the longitudinal trajectories of a 103 subset of biomarkers, to inform their potential future use as response biomarkers. While 104 prognostic clinical measures and biomarkers may have value in the clinical arena, such 105 individual use of these markers is beyond the scope of the current investigation which is 106 focused on the clinical trial utility of these markers,

107

#### 108 Methods

## 109 Study Population

- 110 Patients with ALS were enroled (between 2014 and 2019) at 12 centers in the United States and
- 111 1 center in South Africa through the prospective Phenotype-Genotype-Biomarker (PGB) study
- 112 (registered at clinicaltrials.gov NCT02327845) of the Clinical Research in ALS and Related
- 113 Disorders for Therapeutic Development (CReATe) Consortium. The PGB study enrolled 705
- 114 patients with ALS (n=472), primary lateral sclerosis (n=47), progressive muscular atrophy
- 115 (n=20), hereditary spastic paraplegia (n=162) and other related disorders (n=4). The goal was
- to evaluate participants serially over a period of 1.5-2 years to acquire longitudinal phenotypic
- data. Those with ALS, ALS-FTD and PMA were to be seen at Baseline, and Months 3, 6, 12 and
- 118 18; those with PLS, HSP and multisystem proteinopathy were to be seen at Baseline and
- 119 Months 6, 12, 18 and 24. Biological samples (blood and urine, as well as cerebrospinal fluid
- 120 when willing) were collected at all study visits. Periodic medical record reviews, in addition to
- direct communication with patients, were performed as needed to ascertain the timing of
- survival endpoint (permanent assisted ventilation [PAV; non-invasive ventilation > 23 hours/day],
- 123 tracheostomy, or death).
- 124
- 125 While PGB was designed to be broadly inclusive, the subset of patients with ALS who met
- 126 common trial eligibility criteria were designated as "trial-like" and served as the basis for this
- 127 report. Key inclusion/exclusion criteria included: diagnosis of ALS according to El Escorial
- 128 criteria, permitting those with cognitive or behavioural impairment (ALSci or ALSbi, respectively),
- 129 but excluding ALS-FTD; less than 3 years from onset of weakness; and an erect slow vital

130 capacity (SVC) of ≥50% predicted. All patients with ALS in PGB who met these criteria were131 included in the current report.

132

# 133 Clinical Assessments

134 The ALS Functional Rating Scale-Revised (ALSFRS-R), a 48-point scale that includes bulbar, 135 gross motor, fine motor, and respiratory domains,<sup>24</sup> was the principal measure of functional 136 status. Symptom onset was defined based on the first appearance of weakness or impaired 137 motor function. The estimated rate of change in ALSFRS-R between symptom onset and 138 baseline (ΔFRS), was defined as (48-baseline ALSFRS-R/months since symptom onset).<sup>25</sup> 139 Respiratory muscle function was quantified with slow vital capacity (SVC) in the erect position. 140 Alternate versions of the North American version of the Edinburgh Cognitive and Behavioural 141 ALS Screen (ECAS), including informant report, was used to evaluate cognitive and behavioural

142 function.<sup>26</sup> All evaluators were trained and certified for the performance of each of these

143 outcome measures. Biological sex, as well as race and ethnicity, were self-reported.

144

145 Ethics

146 The University of Miami institutional review board (IRB), which served as the central IRB for

147 CReATe, approved the study for all US sites study (protocol # 20160603) and the University of

148 Cape Town Health Sciences Human Research Ethics Committee approved the study in South

149 Africa (REF number 165/2017). All participants provided written informed consent.

150

151 Biological Samples

152 Biological specimens were collected, processed, and stored according to strict standard

153 operating procedures. Briefly, blood was collected in serum-separating BD vacutainers and

allowed to clot upright at room temperature for 1–2 hours. Following centrifugation (1,750 g for

155 10 min at 4°C), serum was aliquoted into cryogenic sterile freestanding conical microtubes

156 (Nalgene or Bio Plas Inc.) and stored at -80°C. Plasma was collected in K2 EDTA tubes,

157 centrifuged at 1,750g for 10 minutes at 4°C within 2 hours of collection, and aliquoted for

158 storage at -80°C. Urine was collected in a sterile collection cup, gently swirled, and transferred

to cryovials for immediate storage at -80°C.

160

161 Biomarker and Genetic Assays

162 Serum NfL and pNfH concentrations were quantified using the Simoa NfL and pNfH assays in

163 the laboratory of an author (AM). Established protocols for NfL (Simoa Nf-L Advantage Kit-

164 102258, Quanterix) and pNfH (Simoa pNF-Heavy Discovery Kit - 102669) analysis were used.
165 Each plate contained calibrators and quality controls. Samples were diluted to fall within the
166 range of the standard curve.

167

168 Urinary p75<sup>ECD</sup> was guantified by ELISA in the laboratory of an author (MLR) as previously described.<sup>10</sup> Briefly, urinary p75<sup>ECD</sup> was measured by a sandwich ELISA, that used a capture 169 monoclonal antibody (MLR1 at 8mg/ml) made to the extracellular region of p75<sup>27</sup> in Carbonate-170 171 Carbonate coating buffer (Ph 9.6). Another monoclonal antibody (NGFR5) to p75<sup>28</sup> was used as the detection antibody and biotinylated as per the manufacturer's instructions (Thermo Fisher 172 173 Scientific Australia, #UG283022) and used at 2.0mg/ml in the assay. Human p75<sup>ECD</sup> standard 174 was from R&D Systems (Lys29-Asn250; #367-NR). BlockAce (BioRad, BUF-029) was used as 175 blocking and sample buffer. The enzyme reaction was achieved using streptavidin horseradish 176 peroxidase (Jackson ImmunoResearch Laboratories, #JIO16030084) diluted to 1.0 mg /ml and 177 colour developed using tetramethylbenzidine (A:B; BioRad Australia, #1721067). The entire ELISA was accomplished as previously described <sup>29</sup> on a Hamilton Starlet Robot, integrated 178 179 with a Biotek 405 washer, and an MD reader (450nm); two calibrator human urine samples with 180 known p75<sup>ECD</sup> levels were included on each plate, and if the results from either had greater than 181 20% coefficient of variation, the results from the plate were rejected. The results were reported 182 as ng p75<sup>ECD</sup>/ ml urine and corrected by creatinine (mg/ml: measured by calorimetric method 183 using Enzo Life Sciences Creatinine Kits (ADI-907-030A) as per the manufacturer's 184 instructions). Samples with urinary creatinine below  $0.3 \pm 0.03$  mg/ml or above  $3.0 \pm 0.3$  mg ml 185 were rejected as per World Health Organization guidelines <sup>30</sup>. Final results are reported as ng p75<sup>ECD</sup>/mg creatinine. 186

187

188 Total RNA was extracted from plasma using the miRNeasy Micro Kit (Qiagen cat. 217084) and 189 quantified with a Qubit fluorometer using the RNA Broad Range Assay Kit (Thermo Fisher 190 Scientific cat. Q10210). For small RNA next-generation sequencing, libraries were prepared 191 from 7.5 ng of total RNA using the QIAseq miRNA Library Kit (cat. 331505) and QIAseq miRNA 192 NGS 48 Index IL (Qiagen cat. 331592) by an experimenter who was blinded to the identity of 193 samples. Precise linear quantification of miRNA gained by UMIs of random 12 nucleotides after 194 3' and 5' adapter ligation, within the reverse transcription primers. cDNA libraries were amplified 195 by PCR for 22 cycles, with a 3' primer that includes a six-nucleotide unique index, followed by 196 on-bead size selection and cleaning. Library concentration was determined with a Qubit 197 fluorometer (dsDNA High Sensitivity Assay Kit, Thermo Fisher Scientific, cat. Q32851) and

198 library size with TapeStation D1000 (Agilent, cat. Catalog number: Q32851). Libraries with 199 different indices were multiplexed and sequenced on a NovaSeg SP100 (Illumina), with 75-bp 200 single read and 6-bp index read. Human miRNA sequences were mapped using GeneGlobe 201 (Qiagen), normalized with the DESeq2 package and corrected for the library preparation batch. 202 Plasma miR-181a and miR-181b were quantified by small RNA next-generation sequencing in 203 the laboratory of an author (EH) as previously described,<sup>11</sup> and summarised as miR-181ab, the 204 combined expression of miR-181a and miR-181b. Serum uric acid, albumin, creatinine, and 205 CRP were assaved using the Roche Cobas C Analyzer in the Clinical Chemistry Laboratory at 206 the University of Miami. All biomarker studies were performed blind to clinical outcomes. 207

- 208 The presence of a *C9orf72* repeat expansion was determined in the laboratory of an author
- 209 (RR) using a two-step protocol, including a fluorescent PCR fragment-length analysis and a
- 210 repeat-primed PCR, with previously described oligos (ThermoFisher), as described elsewhere.<sup>31</sup>
- 211 The PCR reactions (Qiagen) for both assays included Betaine and DMSO additives
- 212 (MilliporeSigma). The FAM labeled products were run on a 3730xl DNA Analyzer (Applied
- 213 Biosystems) and sized with Genescan 400 using Genemapper software (ThermoFisher).
- 214

## 215 Statistics

Longitudinal change in ALSFRS-R total score, serum NfL, serum pNfH, and urinary p75<sup>ECD</sup> were 216 217 estimated in unadjusted mixed model repeated-measures analyses with visits windowed to the 218 closest planned assessment time (at 3, 6, 12, and 18 months) and in unadjusted mixed model 219 random-slopes analyses using the observed assessment times. The repeated-measures model 220 included a fixed effect of visit and assumed unstructured person-level variance-covariance 221 among repeated observations. The random-slopes model included a fixed effect of time and 222 assumed unstructured variance-covariance for the person-level random intercepts and slopes. 223 Biomarker concentrations were log-transformed prior to analysis and estimates were back-224 transformed. Back-transformation of visit-specific estimates yield values on the original scale of 225 measurement. Back-transformation of slopes or changes from baseline yield geometric mean 226 ratios which were further transformed by subtracting 1 and multiplying by 100 to express as 227 deviations in percentage change from 100%.

- 229 We examined an array of clinical measures (sex, onset age, bulbar onset, diagnostic delay,
- 230 ΔFRS [estimated rate of change in ALSFRS-R between symptom onset and baseline],<sup>25</sup>
- 231 baseline age, ALSFRS-R total score, slow vital capacity, ECAS-derived scores, and ENCALS

predictor score) and biofluid biomarkers (serum NfL, serum pNfH, urinary p75<sup>ECD</sup>, serum uric 232 233 acid, serum albumin, serum creatinine, serum CRP, plasma miR-181ab) as potential 234 prognostics of rate of disease progression as measured by ALSFRS-R total score and of 235 PAV/tracheostomy-free survival. We derived five scores from baseline ECAS assessments: total 236 score, ALS-specific score, ALS non-specific score, and dichotomous designations of cognitive 237 impairment (ALSci) and behavioural impairment (ALSbi) defined according to the revised Strong 238 criteria <sup>32</sup> and implemented in the PGB study.<sup>33</sup> ALSci and ALSbi designations were restricted to 239 English-speaking participants for whom robust normative data permitted reliable designation.<sup>26</sup> 240 The ENCALS linear predictive model score<sup>3</sup> (hereinafter "ENCALS predictor score" or 241 "ENCALS score") combines information from 8 clinical variables ( $\Delta$ FRS, bulbar onset, 242 diagnostic delay [months from symptom onset to diagnosis], age at onset, El Escorial definite 243 ALS, presence of FTD, presence of a C9orf72 repeat expansion, and percent predicted vital 244 capacity). Plasma miR-181ab was evaluated as a continuous measure, split at the median (24,590 UMI in the current study), and as defined by Magen et al <sup>11</sup> where miR181-ab was 245 246 defined as a poor prognostic when above the threshold of 39,300 UMI among those in the 247 middle tertile of NfL concentration (NfL 59-109.8pg/ml)<sup>11</sup>, and as defined by Magen et al but 248 using the median miR181ab value and the middle NfL tertile (44.8-80.8pg/ml) from the current 249 study.

250

251 Prognostic markers were assessed for their ability to predict the rate of progression in ALSFRS-252 R total score in random-slopes analyses and to predict PAV/tracheostomy-free survival by 253 Kaplan-Meier product-limit estimates and by Cox proportional hazards regression. In survival 254 analyses, time at risk began at the baseline visit (time zero) and continued to time last known 255 alive or time of PAV, tracheostomy, or death, if observed. Each model included one prognostic. 256 Continuous prognostics were evaluated both as continuous predictors after standardizing to unit 257 variance and when divided into quartiles. We focused on analyses after dividing prognostics into 258 quartiles (or fewer levels - e.g., for binary measures, where only 2 levels are possible) to avoid 259 the strong assumption of a linear association with rate of progression and survival across the full 260 range of a given prognostic and to permit comparison of all prognostics in a common 261 framework. Models were either unadjusted, adjusted for established core clinical predictors 262 (bulbar onset,  $\Delta$ FRS, and diagnostic delay for functional decline; plus baseline age for survival), 263 adjusted for ENCALS predictor score, or adjusted also for serum NfL. The adjusted models 264 sharpened estimates by accounting for known sources of variation and addressed whether a 265 given prognostic provided new information independent of known predictors of progression and

survival. Wald confidence intervals were used for estimates from random-slopes models.

267 Complementary log-log confidence intervals were used for estimates of median survival time.

- 268 Profile likelihood confidence intervals were used for estimates of hazard ratios.
- 269

270 In addition to estimating the clinical utility of each potential prognostic biomarker, we quantified 271 the proportional sample size saving that would result from adjusting for a given biomarker in a 272 hypothetical clinical trial. Reductions in sample size requirements based on a normal 273 approximation for a hypothetical clinical trial testing for slowing of ALSFRS-R progression, 274 analyzed in a random slopes model, were estimated based on reductions in standard error 275 estimates of the estimated slopes and resulting increases in the effect size after inclusion of a 276 given prognostic marker as a linear predictor. The proportional savings in sample size assume a 277 consistent but arbitrary allocation ratio, type 1 error control, and power between designs and an 278 assessment schedule similar to the present cohort. For any given choice of allocation ratio, type 279 1 error control, and power, the relative sample size required for two trials with equivalent 280 assessment schedule differs only as a function of the ratio of the respective effect sizes for tests 281 of the primary outcome, in the present case the estimated slope of ALSFRS-R. Note that effect 282 size ratios rather than variance ratios were used due to small variation in estimated slopes when 283 adding covariates.

284

A post-hoc analysis of the association between serum NfL and rate of progression in ALSFRS-R total score was performed using a cubic smoothing spline through the empirical Bayes ALSFRS-R slope estimates from an unadjusted random-slopes analysis and using a partial-linear spline in a longitudinal random-slopes analysis. Knots for the partial-linear spline were chosen posthoc, based on visual inspection, at 40 and 100 pg/mL to approximate the shape of the cubic smoothing spline.

- 291
- 292

Analyses were performed using SAS (version 9.4, SAS Institute, Cary NC) and R (version 4.0.3,
R Foundation for Statistical Computing, Vienna, Austria). Comparison-wise p-values are
reported with nominal significance at two-tailed p < 0.05. Results significant after correction by</li>
Holm-Bonferroni stepdown adjustment for multiple comparisons over 28 prognostic markers are
indicated.

- 298
- 299 Role of Funders

The funders of the study had no role in study design, data collection, statistical analysis, resultsinterpretation, or writing of the report.

302

#### 303 Results

## 304 Study Population

305 A total of 203 patients with ALS were included, with a mean (±SD) age of 57.1 (±12) years and a 306 slight male preponderance (55%). A genetic cause of ALS was identified in 24 (12%), most 307 commonly a C9orf72 hexanucleotide repeat expansion (n=20; 10%). Median disease duration 308 (time since symptom onset) at baseline was 14.4 months, with a mean SVC of 85% (±17) 309 predicted (Table 1a). ALSFRS-R declined by an average (±SE) of 0.89 (±0.05) points/month 310 (Figure 1a) with median (Q1-Q3) follow-up of 10.1 (5.8-16.3) months. SVC declined by an 311 average (±SE) of 1.8% (±0.15) predicted per month. 93 (46%) patients reached a survival 312 endpoint (PAV, tracheostomy, or death), with a median (Q1-Q3) survival time of 30.1 (17.4-47.7)

- 313 months observed from follow-up of 17.4 (10.6-29.9) months. 110 patients were censored (25
- study completion, 15 loss to follow-up, 11 withdrawal/dropout, and 59 administrative studyclosure).
- 316

## 317 Biomarker Profiles

318 Baseline serum NfL concentrations ranged from 9 to 214 pg/mL (Table 1b) and correlated with 319 subsequent rates of ALSFRS-R decline (Spearman r=-0.57, 95% CI [-0.66, -0.47], p<0.0001, 320 Figure 1b). Over the course of follow-up, serum NfL increased by an average (95% CI) of 0.98% 321 [0.57%, 1.38%] per month (Table 2, Figure 2a). Baseline serum pNfH concentrations ranged 322 from 3.4 to 4,177 pg/mL (Table 1b) and increased by an average of 0.45% [-0.12%, 1.03%] per month (Table 2, Figure 2b). Baseline urinary p75<sup>ECD</sup> levels ranged from 1.5 to 16.2 ng/mg 323 324 creatinine (Table 1b) and increased by an average of 2.59% [2.01%, 3.17%] per month (Table 2, 325 Figure 2c). Baseline plasma miR-181ab (product of miR-181a and miR-181b) concentration 326 ranged from 2,875 to 431,004 unique molecular identifiers (UMIs) (Table 1b). Baseline 327 concentrations of serum uric acid, albumin, creatinine, and CRP are summarised in Table 1b. 328 Baseline biomarker results, stratified by C9orf72 status and by sex are summarised in eTable 1 329 and eTable 2 respectively, with longitudinal changes in biomarkers stratified by sex in eTable 3.

- 330 Correlations among all prognostics at baseline are summarised in eTable 4.
- 331
- 332 Prognostic Markers for Survival

333 In univariate models, the strongest predictors of survival were the ENCALS score, baseline 334 serum NfL, and  $\Delta$ FRS. Median survival among those with ENCALS predictor scores in the 335 lowest vs. highest guartiles (i.e., lowest vs. highest predicted risk of PAV/tracheostomy-free 336 survival) were 48 vs. 17 months (Table 3, Figure 3a). Median survival among those with  $\Delta$ FRS 337 slopes in the lowest vs. highest quartiles (i.e., slowest vs. fastest pre-baseline slope) were 47 338 vs. 17 months (Table 3, Figure 3b). Median survival among those with baseline NfL 339 concentrations in the lowest vs. highest quartiles were 49 vs. 17 months (Table 3, Figure 3c). 340 Median survival among those with baseline miR181ab above vs. below 24.590 UMI were, 23 vs. 341 35 months (Table 3, Figure 3d). Bulbar onset, baseline ALSFRS-R, and baseline SVC

- 342 %predicted also predicted survival (Table 3).
- 343

344 In Cox proportional hazards models of time to death or equivalent, we evaluated the prognostic 345 utility of each clinical and biofluid marker when added as guartiles to multivariate models that 346 included either a core set of clinical predictors (bulbar,  $\Delta$ FRS, and diagnostic delay) or the 347 ENCALS predictor score (Table 3). Results from models with prognostic measures added as 348 linear terms are summarised in eTable 5. When a given prognostic is included both as quartiles 349 and as a linear term among the covariates, the results presented in Table 3 describe any non-350 linearity in the relationship with survival. Serum NfL remained the strongest predictor. For 351 example, in a model that already includes the ENCALS predictor score, the hazard ratio (HR) 352 for the fourth vs. first quartile values of NfL is 7.3 (Table 3). The addition of NfL as a linear term 353 to an ENCALS-adjusted Cox model, yields a HR of 1.83 for every 1 standard deviation increase 354 in NfL (eTable 5). To examine the prognostic value of plasma miR-181ab in these multivariate 355 models, we considered multiple analytic approaches. The previously published approach, in 356 which a higher miR-181ab is categorised as poor prognostic only for the middle tertile of NfL<sup>11</sup>, 357 reveals no prognostic value beyond that conferred by NfL alone, whether tertiles from a prior 358 study or the current cohort were used. By contrast, dichotomising at the median value in this 359 cohort (but not the threshold value identified in a prior study) added some prognostic value -360 with HRs of 1.65 and 1.73, respectively, when miR-181ab was added to the core set of clinical 361 predictors and the ENCALS predictor score (Table 3). None of the other biomarker candidates 362 considered - serum uric acid, albumin, creatinine, or CRP - added prognostic value in survival 363 analyses (Table 3). Similarly, none of our measures of cognitive/behavioural impairment 364 predicted survival (Table 3).

365

366 Prognostic Markers for Functional Decline

367 In univariate random-slope models of ALSFRS-R decline, ΔFRS, diagnostic delay, ENCALS 368 score, baseline NfL, and baseline pNfH were identified as prognostic markers (Table 4). 369 Although not developed for predicting functional decline, the ENCALS model predicted 370 differential rates of disease progression that ranged from -0.57 to -1.27 points/month among 371 those with the lowest vs. highest quartile ENCALS scores (Table 4, unadjusted). NfL is also a 372 powerful predictor of future functional decline, with slopes ranging from -0.41 to -1.49 373 points/month among those with the lowest vs. highest quartiles NfL values (Table 4). Results 374 from models with prognostic measures added as linear terms are summarised in eTable 6. 375 376 In multivariate models that already incorporate the ENCALS predictor score, quartiles of

baseline serum NfL added substantial prognostic value, with the rate of ALSFRS-R progression
ranging from -0.44 to -1.44 points/month among those with the lowest vs. highest quartile
values. ΔFRS and serum pNfH added much less prognostic value. Irrespective of the analytic
approach, plasma miR-181ab did not add prognostic value beyond that conferred by serum NfL
(Table 4). None of the other clinical markers (including measures of cognitive and behavioural
impairment), or biomarker candidates considered added prognostic value in random-slopes
models of ALSFRS-R functional decline (Table 4).

384

# 385 Impact of Prognostic Markers on Sample Size Savings for Future Clinical Trials

386 For the outcome measure of ALSFRS-R slope, the ENCALS model yields a 9% sample size 387 saving, compared to 30.9% for NfL alone (Table 5). The combination of ENCALS and NfL yields 388 a ~34% saving in sample size. In random slope models of ALSFRS-R that incorporate either the 389 core clinical predictors plus NfL, or ENCALS predictor score plus NfL, the addition of urinary 390 p75<sup>ECD</sup> yields an additional ~4% sample size saving, suggesting a modest additional utility of 391 this prognostic marker (with the caveat that this conclusion is based on incomplete baseline data for p75<sup>ECD</sup> in this sample). The addition of serum pNfH or plasma miR-181ab, however, 392 393 yielded no additional sample size saving, indicating that in multivariate models that incorporate 394 clinical predictors and NfL, these latter biomarkers add little prognostic value when the ALSFRS-395 R slope is the outcome measure (Table 5). None of the other clinical measures (including those 396 of cognitive or behavioural impairment) or biomarker candidates yielded sample size savings 397 when considered as prognostic markers.

400 The relationship between baseline NfL and future rate of functional decline, as measured by the 401 slope of the ALSFRS-R, is not linear (Figure 1b). In this dataset, the sigmoidal relationship 402 vields an estimate of thresholds that might be used either as eligibility criteria (for trial 403 enrichment) or to facilitate stratifying randomisation (to ensure equal balance of NfL-predicted 404 faster and slower disease progression rates across treatment groups). Baseline NfL levels <40 405 pg/mL corresponded to a future ALSFRS-R slope of ~0.5 points/month (i.e. slow progression), 406 whereas baseline levels >100 pg/mL corresponded to a future ALSFRS-R slope of ~1.5 407 points/month (i.e., fast progression). In the range from 40 to 100 pg/mL, ALSFRS-R slope 408 declines quickly for each incremental increase in baseline serum NfL concentration.

409

## 410 Discussion

411 This study comprehensively evaluated leading biochemical *prognostic* biomarker candidates. 412 alone and in combination, and examined their potential utility when combined with established 413 and emerging clinical predictors. This multivariate approach is essential to achieving a fuller 414 understanding of the practical value of candidate prognostic markers. Moreover, mindful that 415 observational studies typically enroll slower progressing patients, for greatest relevance to the 416 design and analysis of future trials we a priori focused our analysis on a trial-like population, the 417 subset of PGB participants who met clinical trial eligibility criteria. Absent a similar biomarker 418 study that utilizes the placebo group from clinical trial(s), our approach is the most robust to date 419 in providing clear answers about the utility of an array of prognostic biomarker candidates. 420

421 Serum NfL is a robust predictor of disease progression, whether the outcome is ASLFRS-R rate 422 of decline or survival time. While the overlap in survival curves for the second and third quartiles 423 of NfL (Figure 3c), for example, suggests limited prognostic value for NfL in the mid-range of 424 values when predicting survival, the relationship between NfL and future rate of functional 425 decline is steepest in the mid-range of values (Figure 1b). Moreover, not only does NfL provide 426 greater prognostic value than the ENCALS predictor score, the combination of NfL and the 427 ENCALS score yields more prognostic value than either NfL or ENCALS score alone. (Of note, 428 we have not fully explored potential transformations of NfL data to optimize its performance as a 429 prognostic marker beyond those displayed in Figure 1. Future research using fractional 430 polynomials or regression splines might further improve the value of NfL as a prognostic <sup>34</sup>. We 431 also acknowledge that some information is lost by dividing a continuous prognostic into 432 categories and that cut points for quartiles will vary from one dataset to another. The quartiles 433 provided here are intended to be descriptive of potential non-linearity in associations, not to be

434 prescriptive of future handling of such prognostics.) Serum pNfH, on the other hand, has some 435 prognostic value for functional decline, but not survival; and in models already adjusting for 436 clinical predictor(s) and NfL, it yielded no additional prognostic value. The prognostic utility of urinary p75<sup>ECD</sup> and plasma miR-181ab are more nuanced, with p75<sup>ECD</sup> yielding some sample 437 438 size saving when combined with clinical predictor(s) and NfL (recognizing that this conclusion is 439 based on incomplete baseline data for p75<sup>ECD</sup> in this sample). Serum uric acid, albumin, 440 creatinine, and CRP have no value as prognostic biomarkers irrespective of the outcome used. 441 Similarly, baseline cognitive and behavioural impairment, based on the ECAS, does not add

- 442 prognostic value.
- 443

444 While the greater prognostic value of blood NfL (than pNfH) may reflect a more critical role for

the NfL isoform in maintaining neuroaxonal structure and function under pathological conditions,

this may also reflect the better analytic performance of the blood NfL immunoassay. pNfH

447 assays in blood are still hampered by a matrix effect and lack of appropriate binding

448 reagents.<sup>35,36</sup> Analytic considerations may also be relevant to the performance of urinary p75<sup>ECD</sup>,

449 which has not yet achieved the same degree of analytic validation as NfL assays.<sup>37</sup>

450

451 The design of this study has both strengths and weaknesses. As an observational study rather 452 than a clinical trial, a limitation is that the intervals between study visits were wide (and 453 variable), requiring us to window study visits around defined time points for the repeated-454 measures analyses (see eTable 8). It is for this reason that we used observed times in a random 455 slopes analyses to estimate sample size savings from incorporation of various potential 456 prognostic biomarkers, despite the FDA's preference for a repeated-measures approach for 457 clinical trials where study visit windows are typically more rigidly controlled. Of note, many ALS 458 clinical trials have historically used this approach <sup>38-40</sup>. Moreover, the estimates themselves 459 depend on the duration of follow-up available at the time of analysis and would likely differ over 460 shorter or longer intervals. In addition, due to premature study closure (for administrative 461 reasons between funding cycles) and some attrition, follow up data at 3-, 6-, and 12-month were 462 available for only 85%, 80% and 52% of participants, respectively - resulting in less precise 463 estimates of ALSFRS-R values beyond 6 months. Vital status after a participant's last visit was 464 ascertained based on clinic notes at some sites, with potentially more complete data collection 465 on deaths; this leads to downward bias in estimates of absolute survival percentages but is 466 unlikely to bias estimates of prognostic value. Strengths of this study include the a priori 467 selection of a trial-like population, the rigorous attention to the guality of phenotypic data, and

the multimodal analysis of putative prognostic biomarkers. Of note, our claims of prognostic
utility do not imply any assumption of a causal relationship between a given prognostic and
progression rate or survival.

471

We also acknowledge the limitations of the ALSFRS-R as an outcome measure in clinical trials, notably the fact that it is not uni-dimensional (meaning that items on the scale measure domains other than functional status) <sup>41</sup>; that a one-point change can represent a variable amount of functional change depending on the question and the item <sup>42</sup>, providing a rationale for reporting the domain specific sub-scores of the ALSFRS-R <sup>43</sup>; and that the decline in ALSFRS-R is not linear across the entire course of disease <sup>44</sup>. Notwithstanding these considerations, the ALSFRS-R is typically linear during the follow-up period encompassed by clinical trials <sup>45</sup>, and

479 remains the principal functional outcome measure used in ALS clinical trials <sup>46</sup>.

480

While the longitudinal trajectory of a subset of the biomarkers was not the major focus of this investigation, we have observed subtle increases in NfL and pNfH over time (in contrast to the conventional wisdom that these are largely stable <sup>8,47-50</sup>). Also noteworthy is the marked increase and relatively consistent trajectory of urinary p75<sup>ECD</sup> (compared to NfL and pNfH).

485 suggesting that urinary p75<sup>ECD</sup> might have value as a response or monitoring biomarker.

486

It should be acknowledged that our evaluation of changes In biomarkers over time—and of the prognostic value of these biomarkers—has been conducted at a population (or group) level.
While statistically robust, conclusions from a population cannot necessarily be extrapolated to individual patients. NfL and other biomarkers considered in our analyses, therefore, remain largely research tools, with more limited (and speculative) value in the clinic setting when applied to individuals.

493

494 While confirmatory studies with larger sample sizes would add confidence to our conclusions, 495 the results of this study are nevertheless immediately relevant to all ongoing and future ALS 496 trials, even in the absence of formal qualification through regulatory agencies such as the 497 FDA.<sup>51</sup> Our findings are especially relevant to trials with 6-month treatment duration, the period 498 for which we have more complete data. First, baseline NfL should be incorporated into the 499 analysis plan for all clinical trials as a prognostic biomarker, whether functional decline or 500 survival is used as the primary outcome. Second, how one incorporates baseline NfL into trial 501 design – either as an eligibility criterion or as a stratification factor – depends on the purpose.

502 For example, if the goal is to enrich the trial population for either faster or slower progressing 503 patients, or to stratify randomisation based on anticipated rate of disease progression, then NfL 504 levels above or below a defined threshold might be used. Our data suggest serum NfL 505 thresholds of <40pg/mL for slow progressors and >100pg/mL for fast progressors. Between 40-506 100pg/ml, given the steep relationship between NfL increase and faster future rate of ALSFRS-507 R decline, multiple NfL strata may be required for randomisation (as permitted by study sample 508 size), in order to adequately control for heterogeneity of predicted disease progression rate. 509 (Importantly, the same threshold may not hold for predicting future survival.) Third, in a hypothetical clinical trial with ALSFRS-R slope as the outcome, except for urinary p75<sup>ECD</sup>, other 510 511 putative prognostic biomarkers yield very little in the way of sample size saving beyond those 512 conferred by the combination of established clinical predictor(s) and NfL. While incorporation of 513 plasma miR-181ab in such a model does not improve prediction of future rates of ALSFRS-R 514 decline or yield additional sample size savings, it may have some value in predicting survival. 515 516 This study exemplifies the critical importance of a multivariate approach to evaluating new 517 prognostic markers and highlights the necessity for novel markers to demonstrate value added 518 to existing predictors. Moreover, the implication of our finding that clinical predictors 519 (encapsulated, for example, by the ENCALS score) and blood-based measurement of NfL are 520 strong predictors of disease progression, is that both should be incorporated into all ongoing 521 and future Phase 2 and Phase 3 ALS trials. Moreover, the dual use of NfL as a prognostic and 522 response biomarker will aid interpretation of Phase 2 trial results and facilitate go/no-go 523 decisions about advancing experimental agents to Phase 3. Collectively, these modifications to 524 ALS trial design and analysis should accelerate the pace of ALS therapy development. 525

#### 527 Contributors

- 528 MB: Study concept/design, study oversight, major role in acquisition of data,
- analysis/interpretation of data, access to and verification of underlying data, anddrafting/revising the manuscript for content.
- 531 EAM: Analysis/interpretation of data, access to and verification of underlying data, and 532 drafting/revising the manuscript for content.
- 533 AM: Study concept/design, major role in acquisition of data, and drafting/revising the 534 manuscript for content.
- 535 MLR: Study concept/design, major role in acquisition of data, and drafting/revising the 536 manuscript for content.
- 537 EH: Major role in acquisition of data and drafting/revising the manuscript for content.
- 538 VL: Major role in acquisition of data.
- 539 DR: Major role in acquisition of data.
- 540 IM: Major role in acquisition of data and drafting/revising the manuscript for content.
- 541 YC: Major role in acquisition of data.
- 542 VG: Major role in acquisition of data.
- 543 JS: Major role in acquisition of data.
- 544 JH: Major role in acquisition of data.
- 545 RR: Major role in acquisition of data and editing the manuscript for content.
- 546 CAM: Major role in generation of summary cognitive/behavioural data.
- 547 LP: Study concept/design and editing the manuscript for content.
- 548 CM: Major role in acquisition of data and editing the manuscript for content.
- 549 JW: Study concept/design, study oversight, major role in acquisition of data,
- analysis/interpretation of data, access to and verification of underlying data, and
- 551 drafting/revising the manuscript for content.
- 552 All authors have read and approved the final version of the manuscript.
- 553 CReATe Consortium PGB Investigators contributed to data collection.
- 554

# 555 Data Sharing Statement

- 556 Following publication, de-identified participant data and a data dictionary defining each field in
- the dataset, will be made available following request to the corresponding authors and upon
- 558 execution of a data access agreement. Additional study related documents are not available.
- 559 **Declaration of Interests**

- 560 MB reports grants from the NIH (U01NS107027, U54NS092091) and the ALS Association (16-
- 561 TACL-242) in support of this work. He is also an unpaid member of the Board of Trustees
- for the ALS Association. He has served as a consultant to Alector, Alexion, Annexon,
- 563 Arrowhead, Biogen, Cartesian, Denali, Eli Lilly, Horizon, Immunovant, Novartis, Roche,
- 564 Sanofi, Takeda, UCB, and UniQure.
- 565 EAM reports grants from the NIH (U01NS107027, U54NS092091). He also serves as a
- 566 consultant to Annexon, Biogen, Bial Biotech, Cortexyme, Chase Therapeutics, Enterin, nQ
- 567 Medical, Partner Therapeutics, Stoparkinson Healthcare, and UCB. He has served on 568 DSMBs for NeuroSense Therapeutics, Novartis, and Sanofi.
- AM reports grants from the NIH (U01NS107027). He has also provided consulting services to
   Roche, Pfizer, and Accure Therapeutics.
- 571 MLR reports support from grants from the NIH (U01NS107027, U54NS092091) and FightMND.
- 572 EH has nothing to declare.
- 573 VL has nothing to declare.
- 574 DR has nothing to declare.
- 575 SS has nothing to declare.
- 576 IM has nothing to declare.
- 577 YC has nothing to declare.
- 578 VG currently is an employee of Biohaven Pharmaceutical Inc.
- 579 JS receives research funding from the NIH< MDA, FSHD Society, Friends of FSH Research,
- 580 and FSHD Canada. He also serves as a consultant or on scientific advisory boards for
- 581 Avidity, Fulcrum, Dyne, Armatus, Epic Bio, Roche, Lupin, and Entrada
- 582 JH has nothing to declare.
- 583 RR reports grants from the NIH (U54NS092091) in support of this work. She is also an unpaid
- 584 member of the Medical Advisory Board of the Association for Frontotemporal Dementias
- 585 (AFTD) and a paid member of the Scientific Advisory Board of the Kissick Family
- 586 Foundation FTD Grant Program
- 587 CAM reports funding from the ALS Association and the NIH (U54NS092091).
- 588 LP reports support from the Mayo Clinic Foundation and grants from the National Institute on
- 589 Aging (5P30AG0062677, U19AG063911) the National Institute of Neurological Disorders
- and Stroke (U54NS123743, R35NS097273, P01NS084974) and Target ALS Foundation.
- 591 CM reports grants from the NIH (AG066597, AG076411, AG066152, AG072979, NS109260,
- 592 NS092091), Department of Defense, and support from the Penn Institute on Aging,
- 593 Decrane Family PPA Fund, and Newhouse Fund.2

- 594 JW reports funding from the NIH (U01NS107027, U54NS092091) and the ALS Association (16-595 TACL-242) in support of this work.
- 596

597 The CReATe Consortium (U54NS092091) is part of the NIH Rare Diseases Clinical Research

- 598 Network (RDCRN), an initiative of the Office of Rare Diseases Research (ORDR), NCATS. This
- 599 Consortium is funded through a collaboration between NCATS and the NINDS. This work was
- also supported by a Clinical Trial Readiness grant (U01NS107027) from NINDS and by a grant
- from the ALS Association to support the CReATe Biorepository (grant ID 16-TACL-242).
- 602

# 603 Acknowledgements

604 The authors thank participants in the CReATe Consortium's Phenotype-Genotype-Biomarker (PGB1)

605 study; research staff at each clinical site; and the CReATe Consortium project management and data

- 606 management teams, genomics sub-core, and biorepository. The authors also thank members of the
- 607 University of Miami Laboratory for Clinical and Biological Studies for assistance with clinical chemistry
- 608 assays.
- 609
- 610
- 611

## 612 References

- 613
- 614 1. Benatar M, Boylan K, Jeromin A, et al. ALS biomarkers for therapy development: State of the field
  615 and future directions. *Muscle & nerve* 2016; 53(2): 169-82.
- 616 2. Witzel S, Frauhammer F, Steinacker P, et al. Neurofilament light and heterogeneity of disease
   617 progression in amyotrophic lateral sclerosis: development and validation of a prediction model to
   618 improve interventional trials. *Transl Neurodegener* 2021; **10**(1): 31.
- 619 3. Westeneng HJ, Debray TPA, Visser AE, et al. Prognosis for patients with amyotrophic lateral
  620 sclerosis: development and validation of a personalised prediction model. *Lancet Neurol* 2018;
  621 17(5): 423-33.
- Benatar M, Zhang L, Wang L, et al. Validation of serum neurofilaments as prognostic and potential
  pharmacodynamic biomarkers for ALS. *Neurology* 2020; **95**(1): e59-e69.
- 624 5. Miller TM, Cudkowicz ME, Genge A, et al. Trial of Antisense Oligonucleotide Tofersen for SOD1 ALS.
  625 The New England journal of medicine 2022; 387(12): 1099-110.
- 6. Benatar M, Wuu J, Turner MR. Neurofilament light chain in drug development for amyotrophic
  lateral sclerosis: a critical appraisal. *Brain : a journal of neurology* 2022.
- Huang F, Zhu Y, Hsiao-Nakamoto J, et al. Longitudinal biomarkers in amyotrophic lateral sclerosis.
  Annals of clinical and translational neurology 2020; 7(7): 1103-16.
- 630 8. Thompson AG, Gray E, Verber N, et al. Multicentre appraisal of amyotrophic lateral sclerosis
  631 biofluid biomarkers shows primacy of blood neurofilament light chain. *Brain Commun* 2022; 4(1):
  632 fcac029.
- Boylan KB, Glass JD, Crook JE, et al. Phosphorylated neurofilament heavy subunit (pNF-H) in
  peripheral blood and CSF as a potential prognostic biomarker in amyotrophic lateral sclerosis. J *Neurol Neurosurg Psychiatry* 2012.
- 636 10. Shepheard S, Wuu J, Cardoso M, et al. Urinary p75 extracellular domain; A biomarker for
   637 prognosis, progression and pharmacodynamic effect in ALS. *Neurology* 2017; 88(12): 1137-43.
- 63811.Magen I, Yacovzada NS, Yanowski E, et al. Circulating miR-181 is a prognostic biomarker for639amyotrophic lateral sclerosis. Nature neuroscience 2021; 24(11): 1534-41.
- Keizman D, Ish-Shalom M, Berliner S, et al. Low uric acid levels in serum of patients with ALS:
  further evidence for oxidative stress? *Journal of the neurological sciences* 2009; 285(1-2): 95-9.
- 642 13. Ikeda K, Hirayama T, Takazawa T, Kawabe K, Iwasaki Y. Relationships between disease progression
  643 and serum levels of lipid, urate, creatinine and ferritin in Japanese patients with amyotrophic
  644 lateral sclerosis: a cross-sectional study. *Intern Med* 2012; **51**(12): 1501-8.
- 645 14. Paganoni S, Zhang M, Quiroz Zarate A, et al. Uric acid levels predict survival in men with
  646 amyotrophic lateral sclerosis. *Journal of neurology* 2012; 259(9): 1923-8.
- 647 15. Zheng Z, Guo X, Wei Q, et al. Serum uric acid level is associated with the prevalence but not with
  648 survival of amyotrophic lateral sclerosis in a Chinese population. *Metab Brain Dis* 2014; 29(3): 771649 5.
- 650 16. Chio A, Calvo A, Bovio G, et al. Amyotrophic lateral sclerosis outcome measures and the role of
  albumin and creatinine: a population-based study. *JAMA neurology* 2014; **71**(9): 1134-42.
- Mandrioli J, Rosi E, Fini N, et al. Changes in routine laboratory tests and survival in amyotrophic
  lateral sclerosis. *Neurol Sci* 2017; **38**(12): 2177-82.
- 654 18. O'Reilly EJ, Liu D, Johns DR, et al. Serum urate at trial entry and ALS progression in EMPOWER.
  655 *Amyotroph Lateral Scler Frontotemporal Degener* 2017; **18**(1-2): 120-5.
- Paganoni S, Nicholson K, Chan J, et al. Urate levels predict survival in amyotrophic lateral sclerosis:
  Analysis of the expanded Pooled Resource Open-Access ALS clinical trials database. *Muscle & nerve* 2018; 57(3): 430-4.

- Kuffner R, Zach N, Norel R, et al. Crowdsourced analysis of clinical trial data to predict amyotrophic
  lateral sclerosis progression. *Nature biotechnology* 2015; **33**(1): 51-7.
- Paillisse C, Lacomblez L, Dib M, Bensimon G, Garcia-Acosta S, Meininger V. Prognostic factors for
  survival in amyotrophic lateral sclerosis patients treated with riluzole. *Amyotroph Lateral Scler Other Motor Neuron Disord* 2005; 6(1): 37-44.
- Lunetta C, Lizio A, Maestri E, et al. Serum C-Reactive Protein as a Prognostic Biomarker in
  Amyotrophic Lateral Sclerosis. *JAMA neurology* 2017; **74**(6): 660-7.
- Angel G, Peter RS, Rosenbohm A, et al. Adipokines, C-reactive protein and Amyotrophic Lateral
  Sclerosis results from a population- based ALS registry in Germany. *Sci Rep* 2017; 7(1): 4374.
- Cedarbaum JM, Stambler N, Malta E, et al. The ALSFRS-R: a revised ALS functional rating scale that
   incorporates assessments of respiratory function. BDNF ALS Study Group (Phase III). Journal of the
   *neurological sciences* 1999; **169**(1-2): 13-21.
- 671 25. Kimura F, Fujimura C, Ishida S, et al. Progression rate of ALSFRS-R at time of diagnosis predicts
  672 survival time in ALS. *Neurology* 2006; 66(2): 265-7.
- 673 26. McMillan CT, Wuu J, Rascovsky K, et al. Defining cognitive impairment in amyotrophic lateral
  674 sclerosis: an evaluation of empirical approaches. *Amyotroph Lateral Scler Frontotemporal Degener*675 2022; 23(7-8): 517-26.
- 676 27. Rogers ML, Atmosukarto I, Berhanu DA, Matusica D, Macardle P, Rush RA. Functional monoclonal antibodies to p75 neurotrophin receptor raised in knockout mice. *Journal of neuroscience methods* 2006; 158(1): 109-20.
- 679 28. Thompson SJ, Schatteman GC, Gown AM, Bothwell M. A monoclonal antibody against nerve
  680 growth factor receptor. Immunohistochemical analysis of normal and neoplastic human tissue. Am
  681 J Clin Pathol 1989; 92(4): 415-23.
- 682 29. Shepheard SR, Karnaros V, Benyamin B, et al. Urinary neopterin: A novel biomarker of disease
  683 progression in amyotrophic lateral sclerosis. *Eur J Neurol* 2022; **29**(4): 990-9.
- 68430.Mikheev MI, Lowry LK. WHO global project on biological monitoring of chemical exposure at the685workplace. Int Arch Occup Environ Health 1996; 68(6): 387-8.
- 686 31. DeJesus-Hernandez M, Mackenzie IR, Boeve BF, et al. Expanded GGGGCC Hexanucleotide Repeat in
   687 Noncoding Region of C9ORF72 Causes Chromosome 9p-Linked FTD and ALS. *Neuron* 2011.
- Strong MJ, Abrahams S, Goldstein LH, et al. Amyotrophic lateral sclerosis frontotemporal
   spectrum disorder (ALS-FTSD): Revised diagnostic criteria. *Amyotroph Lateral Scler Frontotemporal Degener* 2017; **18**(3-4): 153-74.
- 691 33. McHutchison CA, Wuu J, McMillan CT, et al. Temporal course of cognitive and behavioural changes
  692 in motor neuron diseases. *J Neurol Neurosurg Psychiatry* 2023.
- 693 34. Greenland S. Dose-response and trend analysis in epidemiology: alternatives to categorical analysis. *Epidemiology* 1995; 6(4): 356-65.
- Halbgebauer S, Steinacker P, Verde F, et al. Comparison of CSF and serum neurofilament light and
  heavy chain as differential diagnostic biomarkers for ALS. *J Neurol Neurosurg Psychiatry* 2022;
  93(1): 68-74.
- 698 36. Sturmey E, Malaspina A. Blood biomarkers in ALS: challenges, applications and novel frontiers.
  699 Acta Neurol Scand 2022; 146(4): 375-88.
- 700 37. Rogers ML, Schultz DW, Karnaros V, Shepheard SR. Urinary biomarkers for amyotrophic lateral
   701 sclerosis: candidates, opportunities and considerations. *Brain Commun* 2023; 5(6): fcad287.
- 702 38. Cudkowicz ME, Titus S, Kearney M, et al. Safety and efficacy of ceftriaxone for amyotrophic lateral sclerosis: a multi-stage, randomised, double-blind, placebo-controlled trial. *Lancet Neurol* 2014;
  704 13(11): 1083-91.

- Vucic S, Menon P, Huynh W, et al. Efficacy and safety of CNM-Au8 in amyotrophic lateral sclerosis
  (RESCUE-ALS study): a phase 2, randomised, double-blind, placebo-controlled trial and open label
  extension. *EClinicalMedicine* 2023; **60**: 102036.
- Schoenfeld DA, Finkelstein DM, Macklin E, et al. Design and analysis of a clinical trial using previous trials as historical control. *Clin Trials* 2019; 16(5): 531-8.
- van Eijk RPA, de Jongh AD, Nikolakopoulos S, et al. An old friend who has overstayed their
  welcome: the ALSFRS-R total score as primary endpoint for ALS clinical trials. *Amyotroph Lateral Scler Frontotemporal Degener* 2021; 22(3-4): 300-7.
- 713 42. Fournier CN. Considerations for Amyotrophic Lateral Sclerosis (ALS) Clinical Trial Design.
  714 *Neurotherapeutics* 2022; 19(4): 1180-92.
- 715 43. Rooney J, Burke T, Vajda A, Heverin M, Hardiman O. What does the ALSFRS-R really measure? A
  716 longitudinal and survival analysis of functional dimension subscores in amyotrophic lateral
  717 sclerosis. J Neurol Neurosurg Psychiatry 2017; 88(5): 381-5.
- 44. Gordon PH, Cheng B, Salachas F, et al. Progression in ALS is not linear but is curvilinear. *Journal of neurology* 2010; 257(10): 1713-7.
- Proudfoot M, Jones A, Talbot K, Al-Chalabi A, Turner MR. The ALSFRS as an outcome measure in
   therapeutic trials and its relationship to symptom onset. *Amyotroph Lateral Scler Frontotemporal Degener* 2016; **17**(5-6): 414-25.
- 46. Genge A, Cedarbaum JM, Shefner J, et al. The ALSFRS-R Summit: a global call to action on the use
  of the ALSFRS-R in ALS clinical trials. *Amyotroph Lateral Scler Frontotemporal Degener* 2024; 25(3-4): 382-7.
- 47. Lu CH, Macdonald-Wallis C, Gray E, et al. Neurofilament light chain: A prognostic biomarker in amyotrophic lateral sclerosis. *Neurology* 2015; 84(22): 2247-57.
- 48. Steinacker P, Huss A, Mayer B, et al. Diagnostic and prognostic significance of neurofilament light
  chain NF-L, but not progranulin and S100B, in the course of amyotrophic lateral sclerosis: Data
  from the German MND-net. *Amyotroph Lateral Scler Frontotemporal Degener* 2017; **18**(1-2): 1129.
- 49. Gille B, De Schaepdryver M, Goossens J, et al. Serum neurofilament light chain levels as a marker
  of upper motor neuron degeneration in patients with Amyotrophic Lateral Sclerosis. *Neuropathol*734 *Appl Neurobiol* 2019; 45(3): 291-304.
- 73550.Verde F, Steinacker P, Weishaupt JH, et al. Neurofilament light chain in serum for the diagnosis of<br/>amyotrophic lateral sclerosis. J Neurol Neurosurg Psychiatry 2019; **90**(2): 157-64.
- 737 51. Benatar M, Ostrow LW, Lewcock JW, et al. Biomarker Qualification for Neurofilament Light Chain in Amyotrophic Lateral Sclerosis: Theory and Practice. *Annals of neurology* 2024; **95**(2): 211-6.
- 739
- 740
- 741

# 742 Figure Legends

743

## 744 Figure 1. ALSFRS-R Slope and its Relationship to Baseline NfL

- 745 (a) Random slopes model of ALSFRS-R over time, with errors bars showing 95% confidence
- 746 intervals (CI). Faint grey dotted line illustrates the linear estimate of change in ALSFRS-R over
- time. (b) Relationship between baseline serum NfL (measured in duplicate) and future rate of
- progression of the ALSFRS-R (Spearman correlation coefficient = -0.57, 95% CI -0.66 to -0.47,
- p<0.0001) among n=203 study participants. The straight orange line shows the linear
- prediction. The bent blue line represents a partial-linear spline with knots chosen post-hoc at
- 40 and 100 pg/mL. The smooth green curve is a smoothing spline through the empirical Bayesslope estimates.
- 753

# 754 Figure 2. Longitudinal Biomarker Trajectories

- Longitudinal trajectories of (a) serum NfL; (b) serum pNfH; and (c) urinary p75<sup>ECD</sup>. Y-axis 755 756 shows percent change in each biomarker compared to baseline, plotted on a log scale. The 757 faint grey dotted line illustrates the linear estimate of biomarker change over time. Error bars 758 represent 95% confidence intervals (CI), widened at later time points due to participant attrition 759 over time and fewer biomarker data available. NfL and pNfH were measured in duplicate; 760 p75<sup>ECD</sup> quantified with a median of 3 replicates. Single NfL and pNfH values from the 18-month 761 visit of a single participant have been excluded (see footnote to Table 2 for detailed 762 explanation).
- 763

# 764 **Figure 3**. Kaplan-Meier Survival Curves

- Permanent assisted ventilation (PAV)- and tracheostomy-free survival for **(a)** the ENCALS predictor score, divided into quartiles; **(b)**  $\Delta$ FRS, divided into quartiles; **(c)** baseline serum NfL, divided into quartiles; and **(d)** baseline plasma miR-181ab dichotomised at the median value of
- 768 24,590 UMI. The range of values for each clinical or biological marker within a defined quartile,
- as well as the number of observations at each time point, are shown below each KM plot.
- 770 Shading represents pointwise log-log confidence intervals.
- 771 772
- 773

#### 774 Tables

#### 775

777

# 776 Table 1a. Baseline demographic and key clinical features

Characteristic Total N=203 Male 112 (55.2%) Sex Female 91 (44.8%) Race White 179 (88.2%) Black 15 (7.4%) Asian 4 (2.0%) Other 5 (2.5%) 36 (17.7%) Ethnicity Hispanic/Latino Non-Hispanic/Latino 167 (82.3%) C9orf72 20 (9.9%) Gene with a pathogenic variant 3 (1.5%) SOD1 TARDBP 1 (0.5%) [Unknown] 179 (88.2%) Age at onset, years Mean ± SD (Range) 57.1 ± 12.0 (15-82) Bulbar symptoms at onset No 147 (72.4%) Yes 56 (27.6%) Diagnostic delay, months Median (Q1-Q3) 8.2 (5.2-13.8) Time from symptom onset to baseline, Median (Q1-Q3) 14.4 (10.5-22.5) months Baseline  $\Delta$ FRS, points/month Median (Q1-Q3) 0.60 (0.4-0.9) Baseline age, years Mean ± SD (Range) 58.5 ± 12.1 (17-83) Baseline El Escorial category Clinically definite ALS 45 (22.2%) Clinically probable ALS 95 (46.8%) Other 63 (31.0%) Baseline ALSFRS-R total score Mean ± SD (Range)  $37.9 \pm 5.7 (15-48)$ Baseline SVC, %predicted Mean ± SD (Range) 85.3 ± 17.2 (52.0-135.5) Baseline ECAS total score Mean ± SD (Range)  $110.2 \pm 12.4 (44-130)$ 81.8 ± 10.3 (35-97) Baseline ECAS ALS-specific score<sup>1</sup> Mean ± SD (Range) Baseline ECAS ALS non-specific score Mean ± SD (Range)  $28.4 \pm 3.7 (9-35)$ Baseline cognitive impairment, by ECAS No 137 (87.8%) 19 (12.2%) Yes [n/a]<sup>3</sup> 47 (--) Baseline behavioural impairment, by ECAS No 95 (84.8%) Yes 17 (15.2%) [n/a] 91 (--) Survival duration from baseline, months Median (Q1-Q3) 30.1 (17.4-47.7) PAV, tracheostomy, or death occurrence 110 (54.2%) No Yes 93 (45.8%) Number of sample collections 48 (23.6%) 2 3 56 (27.6%)

Among English speakers (n=171)

<sup>2</sup> No pathogenic variant identified (by *C9orf72* testing and whole genome sequencing) in known diseasecausing genes.

47 (23.2%)

52 (25.6%)

<sup>3</sup> Not available, because a non-English version of ECAS was completed or if there was insufficient information to determine impairment status.

4

5

<sup>4</sup> Not available, because a non-English version of ECAS was completed, caregiver did not complete ECAS, or if there was insufficient information to determine impairment status.

- SD = Standard deviation.  $Q1 = 1^{st}$  quartile.  $Q3 = 3^{rd}$  quartile. ECAS = Edinburgh Cognitive and Behavioural ALS Screen.
- 788 SVC = Slow vital capacity. PAV = Permanent assisted ventilation.

# 792 Table 1b. Baseline biomarker data

Biomarker	N	Mean ±SD	Median (Q1, Q3)	Range
Serum NfL (pg/mL)	203	73.9 ± 47.0	67.9 (37.9, 92.5)	9.1 – 214
Serum pNfH (pg/mL)	203	598 ± 718	267 (110, 924)	3.4 – 4,177
Serum NfL/pNfH ratio	203	0.52 ± 1.56	0.18 (0.09, 0.50)	0.020 - 20.9
Urinary p75 <sup>ECD</sup> (ng/mg creatinine)	160 <sup>1</sup>	5.54 ± 2.42	5.05 (3.93, 6.44)	1.53 – 16.1
Serum uric acid (mg/dL)	203	5.19 ± 1.31	5.00 (4.20, 6.20)	2.60 - 8.90
Serum creatinine (mg/dL)	203	0.78 ± 0.20	0.77 (0.66, 0.90)	0.29 – 1.59
Serum albumin (g/dL)	203	4.55 ± 0.34	4.50 (4.30, 4.80)	3.60 - 5.80
Serum CRP (mg/dL)	203	0.27 ± 0.35	0.10 (0.10, 0.50)	0.10 – 2.30
Plasma miR-181a (UMI)	201	451 ± 208	418 (312, 552)	124 – 1,699
Plasma miR-181b (UMI)	201	66.2 ± 31.4	61.2 (44.8, 79.7)	13.8 – 263
Plasma miR-181ab (UMI/2)	201	34.663 ± 39.372	24,590 (13,638, 42,836)	3.148 - 447.480

UMI = unique molecular identifier

<sup>1</sup> Urinary p75<sup>ECD</sup> only available from a subset of study participants.

804805 Table 2. Longitudinal biomarker trajectories

Piemerker	Increase per month, relative to baseline						
Diomarker	Mean	(95% CI)	P-value				
Serum NfL (pg/mL) <sup>1</sup>	0.98%	(0.57%, 1.38%)	<0.0001				
Serum pNfH (pg/mL) <sup>1</sup>	0.45%	(-0.12%, 1.03%)	0.12				
Serum NfL/pNfH ratio	0.44%	(-0.09%, 0.97%)	0.10				
Urinary p75 <sup>ECD</sup> (ng/mg creatinine)	2.59%	(2.01%, 3.17%)	<0.0001				

Values are unadjusted for core clinical covariates

<sup>1</sup> One substantial outlier, from the 18-month visit (i.e. visit 5) of a participant, was excluded.

# Table 3. Prognostic markers of survival

	Unadjusted Analysis*					Adjusted Analysis*			
Prognostic Marker (in quartiles or binary)	Estimat	ed median by ı	survival (9 narker qua	95% CI) in n artile	Covariates included: Core clinical predictors <sup>2</sup> Covariate include ENCALS predictors <sup>3</sup>			ncluded: lictor score	
	Q1 / No	Q2	Q3	Q4 / Yes	p-value	HR (95% CI) [Q4 vs Q1] <sup>4</sup>	p-value	HR (95% CI) [Q4 vs Q1] <sup>5</sup>	p-value
Sex, male	25 (22-37)			31 (26-46)	0.62	1.13 (0.7-1.8)	0.61	0.98 (0.6-1.5)	0.91
Age at onset, years	47 (31-48)	25 (21-37)	24 (17-37)	22 (16-34)	0.023	1.33 (0.3-6.2)	0.28	1.34 (0.7-2.6)	0.59
Bulbar symptoms at onset	33 (30-46)			22 (17-27)	0.039	N/A		1.31 (0.8-2.0)	0.25
Diagnostic delay	30 (22-37)	27 (20-48)	27 (22-ne)	56 (21-ne)	0.096	0.65 (0.1 - 3.1)	0.73	1.66 (0.7-3.7)	0.43
Baseline ∆FRS	47 (34-ne)	35 (22-56)	30 (22-43)	17 (14-27)	<0.0001 <sup>13</sup>	3.64 (1.3-10.5)	0.11	1.62 (0.7-3.7)	0.39
Baseline age	46 (30-48)	23 (19-37)	25 (19-43)	30 (17-34)	0.073	0.46 (0.1-2.2)	0.10	1.21 (0. 6-2.3)	0.23
Baseline ALSFRS-R	22 (16-26)	24 (21-32)	48 (31-ne)	43 (31-48)	0.0010 <sup>13</sup>	0.55 (0.3-1.1)	0.03	0.61 (0.3-1.1)	0.04
Baseline SVC %predicted	20 (15-27)	31 (23-47)	34 (24-ne)	47 (27-ne)	0.0046	0.49 (0.3-0.9)	0.10	0.64 (0.3-1.2)	0.58
Baseline ECAS total <sup>6</sup>	31 (15-56)	30 (22-47)	31 (19-48)	48 (24-ne)	0.54	0.99 (0.5-2.1)	0.78	0.77 (0.4-1.6)	0.75
Baseline ECAS ALS-specific <sup>6</sup>	37 (18-ne)	30 (20-47)	23 (17-49)	48 (26-ne)	0.14	1.12 (0.5-2.6)	0.12	0.88 (0.4-2.0)	0.17
Baseline ECAS ALS non-specific <sup>6</sup>	25 (14-37)	47 (26-48)	30 (19-ne)	31 (23-48)	0.37	0.68 (0.3-1.4)	0.73	0.65 (0.3-1.3)	0.66
Baseline cognitive impairment <sup>6</sup>	30 (25-46)			37 (13-ne)	0.99	0.70 (0.3-1.5)	0.41	0.84 (0.3-1.7)	0.67
Baseline behavioural impairment <sup>6</sup>	34 (25-47)			18 (8-ne)	0.48	1.06 (0.4-2.4)	0.90	1.67 (0.7-3.6)	0.22
Baseline ENCALS predictor score	48 (39-ne)	35 (26-ne)	24 (20-32)	17 (13-25)	< 0.0001 13	4.55 (1.5 -14.7)	0.054	1.61 (0.3-7.4)	0.87
Baseline serum NfL	49 (46-ne)	30 (23-35)	26 (17-39)	17 (13-22)	<0.0001 13	7.71 (3.7 -17.1)	<0.0001 13	7.34 (3.7- 15.8)	<0.0001 13
Baseline serum pNfH	43 (30-ne)	23 (17-37)	25 (23-46)	26 (19-47)	0.18	1.74 (1.0-3.2)	0.28	1.68 (1.0-3.0)	0.36
Baseline urinary p75 <sup>ECD</sup>	34 (24-46)	31 (21-ne)	22 (14-ne)	30 (19-48)	0.77	0.92 (0.4-1.9)	0.94	0.65 (0.3-1.3)	0.57
Baseline serum uric acid	25 (21-37)	31 (15-46)	30 (23-34)	47 (25-56)	0.25	0.69 (0.4-1.3)	0.38	0.58 (0.3-1.1)	0.22
Baseline serum creatinine	31 (21-46)	24 (19-48)	31 (22-ne)	30 (23-47)	0.99	0.87 (0.5-1.6)	0.89	0.95 (0.5-1.7)	0.97
Baseline serum albumin	25 (20-35)	23 (19-52)	32 (22-47)	34 (27-47)	0.72	0.95 (0.5-1.8)	0.99	0.96 (0.5-1.8)	0.96
Baseline serum CRP	30 (2	4-39)	26 (9-ne)	30 (22-47)	0.85	1.07 (0.6-1.8)	0.96	1.08 (0.6-1.7)	0.95
Baseline plasma miR-181ab	35 (30-ne)	34 (24-52)	26 (20-47)	22 (17-30)	0.021	1.90 (1.0-3.6)	0.16	1.82 (1.0-3.3)	0.096
Baseline miR-181ab > 39,300 UMI	33 (26-48)			23 (17-32)	0.0062	1.55 (0.9-2.5)	0.075	1.55 (1.0-2.4)	0.050
Baseline miR-181ab > 24,590 UMI	35 (30-48)			23 (20-32)	0.016	1.65 (1.1-2.6)	0.030	1.73 (1.1-2.7)	0.014
Baseline NfL+miR181ab poor Px <sup>9</sup>	37 (30-48)			17 (15-21)	< 0.0001 13	2.69 (1.6-4.4)	0.0001 <sup>13</sup>	2.61 (1.7-4.1)	< 0.0001 <sup>13</sup>
Baseline NfL+miR181ab poor Px <sup>10</sup>	47 (34-48)			20 (16-22)	< 0.0001 13	3.14 (2.0-5.1)	< 0.0001 13	3.19 (2.0-5.0)	< 0.0001 13
Baseline NfL median split <sup>11</sup>	47 (31-48)			20 (16-25)	< 0.0001 13	2.24 (1.4-3.6)	0.0006 13	2.29 (1.5-3.6)	$0.0002^{13}$
Baseline NfL 4-level split <sup>12</sup>	47 (46-52)	24 (19-31)	32 (14-18)	17 (15-22)	<0.0001 <sup>13</sup>	4.28 (2.4-8.0)	0.0001 <sup>13</sup>	4.12 (2.3-7.5)	0.0001 <sup>13</sup>

\*Survival analysis. Q1-Q4 indicate quartiles of continuous predictors, with higher quartiles representing higher values. Yes/No in column headings captures the presence/absence of binary predictors. ne = not estimable.

<sup>1</sup> Without inclusion of covariate or prognostic marker in the model, median survival (95% Cl) = 30 (17-48) months.

<sup>2</sup> Core clinical predictors in survival analyses include bulbar onset, diagnostic delay,  $\Delta$ FRS, and baseline age.

- <sup>3</sup> ENCALS predictor score is derived from ΔFRS, bulbar onset, diagnostic delay, age at onset, SVC percent predicted, El Escorial definite ALS, presence of FTD, and presence of a *C9orf72* repeat expansion.
- <sup>4</sup> These HRs compare Q4 to Q1 of each prognostic marker in a model that also adjusts for the core clinical predictors of survival. While the adjusted analyses include diagnostic delay, ΔFRS, and baseline age as linear covariates, the potential additional prognostic value of each of these predictors (see respective rows) is evaluated by contrasting top and bottom quartiles to detect possible non-linear effects.

<sup>5</sup> These HRs compare Q4 to Q1 of each prognostic marker in a model that also adjusts for the ENCALS score as a linear covariate. The potential additional prognostic value of the ENCALS predictor score (see row) is evaluated by contrasting top and bottom quartiles to detect possible non-linear effects.

<sup>6</sup> Among English speakers (n=171)

- <sup>7</sup> Threshold of 39,300 UMI in plasma as defined by Magen et al <sup>11</sup>
- <sup>8</sup> Median of 24,590 UMI in plasma in the current dataset
- <sup>9</sup> Poor prognosis based on published optimal combination of NfL and miR-181ab, in which a poor prognostic factor is defined as either (NfL > 109.8pg/ml) or (NfL > 59.0pg/ml and miR-181ab > 39,300 UMI)<sup>11</sup>.
- <sup>10</sup> Poor prognosis based on recalculated combination of NfL and miR-181ab using thresholds obtained from the current dataset; a poor prognostic factor is defined as either (NfL > 80.8 pg/mL) or (NfL > 44.8 pg/mL and mIR-181ab > 24,590 UMI).
- <sup>11</sup> Median serum NfL = 67.9 pg/mL
- <sup>12</sup> Serum NfL 4-level split is at the 33<sup>rd</sup>, 50<sup>th</sup>, and 67<sup>th</sup> percentiles (44.8 pg/mL, 67.9 pg/mL, and 80.8 pg/mL, respectively), i.e., tertiles and median, rather than quartiles, to mimic construction of the NfL+miR18ab measure <sup>11</sup>.

<sup>13</sup> p-value remains statistically significant after adjustment for multiplicity. Holm-Bonferroni adjusted p-values are reported in eTable 7.

# Table 4. Prognostic markers of functional decline

	Unadjusted Analysis *					Adjusted Analysis *				
(in quartiles or binary)	ALSFRS-R slope (points/month): Estimate (SE) <sup>1</sup>					ALSFRS-R slope (points/month): Estimate (SE) <sup>1</sup>				· · · ·
	Q1 /No	Q2	Q3	Q4 / Yes	p-value	Q1 /No	Q2	Q3	Q4 / Yes	p-value
Sex, male	-0.90 (-1.1, -0.74)			-0.88 (-1.0, -0.73)	0.80	-0.90(-1.1, -0.75)			-0.88 (-1.0, -0.74)	0.83
Age at onset	-0.74 (95, -0.53)	-0.90 (-1.1, -0.70)	-1.00 (-1.2, -0.79)	-0.91 (-1.1, -0.67)	0.38	-0.87 (-1.1, -0.66)	-0.91 (-1.10, -0.72)	-0.95 (-1.1, -0.75)	-0.79 (-1.0, -0.56)	0.77
Bulbar symptoms at onset	-0.82 (94, -0.70)			-1.07 (-1.3, -0.87)	0.036	-0.84 (-0.96, -0.72)			-1.02 (-1.2, -0.82)	0.14
Diagnostic delay	-1.10 (-1.3,090)	-0.99 (-1.2, -0.79)	-0.70 (92, -0.48)	-0.73 (93, -0.52)	0.016	-0.99 (-1.2, -0.78)	-0.93 (-1.10, -0.73)	-0.73 (-0.94, -0.51)	-0.89 (-1.1, -0.66)	0.38
Baseline ΔFRS	-0.72 (-0.91, -0.54)	-0.83 (-1.0, -0.62)	-0.69 (89, -0.48)	-1.37 (-1.6, -1.1)	< 0.0001 10	-0.87 (-1.1, -0.66)	-0.83 (-1.00, -0.62)	-0.66 (-0.86, -0.45)	-1.22 (-1.5, -0.98)	0.0036
Baseline age	-0.76 (-0.97, -0.55)	-0.99 (-1.2, -0.79)	-0.91 (-1.1, -0.69)	-0.89 (-1.1, -0.66)	0.47	-0.88 (-1.1, -0.68)	-0.99 (-1.20, -0.79)	-0.87 (-1.1, -0.66)	-0.80 (-1.0, -0.58)	0.63
Baseline ALSFRS-R	-0.96 (-1.2, -0.75)	-0.92 (-1.1, -0.70)	-0.68 (92, -0.45)	-0.93 (-1.1, -0.73)	0.31	-0.88 (-1.1, -0.67)	-0.93 (-1.10, -0.72)	-0.70 (-0.92, -0.47)	-1.00 (-1.2, -0.80)	0.24
Baseline SVC %predicted	-1.08 (-1.3, -0.86)	-0.90 (-1.1, -0.70)	-0.85 (-1.1, -0.64)	-0.70 (92, -0.48)	0.12	-0.96 (-1.2, -0.74)	-0.92 (-1.10, -0.72)	-0.91 (-1.1, -0.71)	-0.73 (-0.94, -0.52)	0.44
Baseline ECAS total <sup>2</sup>	-0.99 (-1.2, -0.76)	-0.77 (-1.0, -0.53)	-0.79 (-1.0, -0.55)	-0.84 (-1.1, -0.60)	0.54	-0.98 (-1.2, -0.76)	-0.72 (-0.95, -0.49)	-0.80 (-1.0, -0.57)	-0.92 (-1.1, -0.68)	0.38
Baseline ECAS ALS-specific <sup>2</sup>	-0.83 (-1.1, -0.60)	-0.84 (-1.1, -0.62)	-0.92 (-1.2, -0.68)	-0.81 (-1.1, -0.55)	0.92	-0.84 (-1.1, -0.61)	-0.81 (-1.00, -0.59)	-0.93 (-1.2, -0.70)	-0.85 (-1.1, -0.60)	0.89
Baseline ECAS ALS non-specific <sup>2</sup>	-0.91 (-1.1, -0.67)	-0.70 (92, -0.48)	-0.98 (-1.2, -0.75)	-0.80 (-1.1, -0.54)	0.36	-0.87 (-1.1, -0.65)	-0.73 (-0.95, -0.52)	-0.99 (-1.2, -0.77)	-0.82 (-1.1, -0.56)	0.42
Baseline cognitive impairment <sup>2</sup>	-0.83 (-0.96, -0.70)			-0.89 (-1.2, -0.54)	0.73	-0.83 (-0.95, -0.70)			-0.89 (-1.2, -0.54)	0.75
Baseline behavioural impairment <sup>2</sup>	-0.78 (-0.93, -0.63)			-1.04 (-1.4, -0.66)	0.21	-0.76 (-0.90, -0.61)			-0.97 (-1.3, -0.61)	0.29
Baseline ENCALS predictor score	-0.57 (-0.76, -0.37)	-0.70 (89, -0.51)	-1.02 (-1.2, -0.81)	-1.27 (-1.5, -1.1)	< 0.0001 10	-0.62 (-0.90, -0.34)	-0.71 (-0.90, -0.51)	-1.00 (-1.2, -0.79)	-1.23 (-1.5, -0.96)	0.021
Baseline serum NfL	-0.41 (-0.58, -0.24)	-0.67 (84, -0.50)	-1.10 (-1.3, -0.92)	-1.49 (-1.7, -1.3)	<0.0001 10	-0.44 (-0.60, -0.27)	-0.71 (-0.88, -0.54)	-1.08 (-1.3, -0.90)	-1.44 (-1.6, -1.2)	<0.0001 10
Baseline serum pNfH	-0.68 (-0.88, -0.48)	-0.96 (-1.2, -0.74)	-0.82 (-1.0, -0.61)	-1.13 (-1.4, -0.91)	0.024	-0.70 (-0.89, -0.51)	-0.91 (-1.10, -0.71)	-0.86 (-1.1, -0.65)	-1.12 (-1.3, -0.91)	0.041
Baseline urinary p75 <sup>ECD</sup>	-0.93 (-1.2, -0.70)	-0.75 (-1.0, -0.50)	-1.03 (-1.3, -0.79)	-0.97 (-1.2, -0.73)	0.44	-0.99 (-1.2, -0.77)	-0.76 (-1.00, -0.52)	-0.96 (-1.2, -0.73)	-0.96 (-1.2, -0.73)	0.50
Baseline serum uric acid	-0.96 (-1.2, -0.75)	-0.85 (-1.1, -0.63)	-0.91 (-1.1, -0.70)	-0.82 (-1.0, -0.60)	0.82	-0.98 (-1.2, -0.78)	-0.82 (-1.00, -0.61)	-0.93 (-1.1, -0.73)	-0.81 (-1.0, -0.60)	0.61
Baseline serum creatinine	-0.90 (-1.1, -0.69)	-0.89 (-1.1, -0.67)	-0.80 (-1.0, -0.57)	-0.95 (-1.2, -0.74)	0.81	-0.91 (-1.1, -0.71)	-0.94 (-1.20, -0.73)	-0.76 (-0.98, -0.54)	-0.93 (-1.1, -0.74)	0.61
Baseline serum albumin	-0.82 (-1.0, -0.61)	-0.87 (-1.1, -0.64)	-1.03 (-1.2, -0.85)	-0.71 (98, -0.45)	0.23	-0.74 (-0.95, -0.54)	-0.83 (-1.00, -0.62)	-1.09 (-1.3, -0.92)	-0.76 (-1.0, -0.50)	0.039
Baseline serum CRP <sup>3</sup>	-0.85 (-0.9	97, -0.72)	-1.12 (-1.6, -0.66)	-0.97 (-1.2, -0.74)	0.38	-0.84 (-0.	96, -0.72)	-1.08 (-1.5, -0.64)	-0.98 (-1.2, -0.77)	0.37
Baseline plasma miR-181ab	-0.71 (-0.92, -0.50)	-0.82 (-1.0, -0.61)	-0.92 (-1.1, -0.70)	-1.11 (-1.3, -0.89)	0.061	-0.71 (-0.91, -0.51)	-0.83 (-1.00, -0.63)	-0.95 (-1.2, -0.74)	-1.07 (-1.3, -0.86)	0.085
Baseline miR-181ab > 39,300 UMI <sup>4</sup>	-0.80 (-0.93, -0.68)			-1.09 (-1.3, -0.89)	0.017	-0.82 (-0.94, -0.70)			-1.06 (-1.3, -0.87)	0.037
Baseline miR-181ab > 24,590 UMI <sup>5</sup>	-0.76 (-0.91, -0.61)			-1.01 (-1.2, -0.86)	0.023	-0.77 (-0.91, -0.63)			-1.01 (-1.2, -0.86)	0.022
Baseline NfL+miR181ab poor Px <sup>6</sup>	-0.69 (-0.80, -0.57)			-1.39 (-1.6, -1.2)	<0.0001 10	-0.71 (-0.82, -0.60)			-1.34 (-1.5, -1.1)	<0.0001 10
Baseline NfL+miR181ab poor Px <sup>7</sup>	-0.56 (-0.68, -0.43)			-1.28 (-1.4, -1.1)	< 0.0001 10	-0.59 (-0.72, -0.47)			-1.24 (-1.4, -1.1)	< 0.0001 10
Baseline NfL median split <sup>8</sup>	-0.54 (-0.66, -0.41)			-1.28 (-1.4, -1.1)	< 0.0001 10	-0.58 (-0.70, -0.45)			-1.24 (-1.4, -1.1)	< 0.0001 10
Baseline NfL 4-level split <sup>9</sup>	-0.44 (-0.59, -0.30)	-0.73 (94, -0.52)	-1.05 (0.11)	-1.41 (-1.6, -1.2)	< 0.0001 10	-0.48 (-0.63, -0.34)	-0.74 (-0.95, -0.53)	-1.05 (-1.3, -0.83)	-1.36 (-1.5, -1.2)	<0.0001 10

\* Random slopes model. ENCALS predictor score included in adjusted analysis. Q1-Q4 indicate quartiles of continuous predictors, with higher quartiles representing higher values. Yes/No in column headings captures the presence/absence of binary predictors.

<sup>1</sup> Without inclusion of covariate or prognostic marker in the model, ALSFRS-R slope (SE) = -0.89 (0.05) points/month.

<sup>2</sup> Among English speakers (n=171)

<sup>3</sup> More than 50% of observations were below the lower limit of quantification and were imputed at 0.1 mg/dL

- $^{\rm 4}$  Threshold of 39,300 UMI in plasma as defined by Magen et al  $^{\rm 11}$
- <sup>5</sup> Median of 24,590 UMI in plasma in the current dataset
- <sup>6</sup> Poor prognosis based on published optimal combination of NfL and miR-181ab, in which a poor prognostic factor is defined as either (NfL > 109.8 pg/mL) or (NfL > 59.0 pg/mL and miR-181ab > 39,300 UMI) <sup>11</sup>.
- <sup>7</sup> Poor prognosis based on recalculated combination of NfL and mIR-181ab using thresholds obtained from the current dataset; a poor prognostic factor is defined as either (NfL > 80.8 pg/mL) or (NfL > 44.8 pg/mL and mIR-181ab > 24,590 UMI).

<sup>8</sup> Median serum NfL = 67.9 pg/mL

<sup>9</sup> Serum NfL 4-level split is at the 33<sup>rd</sup>, 50<sup>th</sup>, and 67<sup>th</sup> percentiles (44.8 pg/mL, 67.9 pg/mL, and 80.8 pg/mL, respectively), i.e., tertiles and median, rather than quartiles, to mimic construction of the NfL+miR18ab measure <sup>11</sup>.

<sup>10</sup> p-value remains statistically significant after adjustment for multiplicity. Holm-Bonferroni adjusted p-values are reported in eTable 7.

# Table 5. Estimated total sample size savings in a random slopes model of ALSFRS-R progression that includes the prognostic marker and covariate(s)

		Covariate(s) included						
Prognostic Marker	Unadjusted	Core clinical predictors <sup>1</sup>	ENCALS predictor score <sup>2</sup>	Core clinical predictors <sup>1</sup> + NfL	ENCALS predictor score <sup>2</sup> + NfL			
ENCALS linear score	-9.0%	-10.4%		-33.6%				
Serum NfL	-30.9%	-33.4%	-33.6%					
Serum pNfH	-4.0%	-13.3%	-12.3%	-33.2%	-33.5%			
Urinary p75 <sup>ECD</sup>	-7.6%	-13.2%	-16.4%	-37.2%	-37.5%			
Serum uric acid	-0.8%	-8.2%	-8.2%	-33.2%	-33.3%			
Serum creatinine	1.2%	-8.3%	-7.8%	-33.0%	-33.2%			
Serum albumin	1.9%	-8.1%	-7.4%	-33.2%	-33.4%			
Serum CRP	-0.3%	-10.3%	-9.3%	-33.5%	-33.4%			
Plasma miR-181ab	-2.0%	-9.7%	-10.6%	-34.2%	-35.4%			
NfL+miR181ab poor Px <sup>3</sup>	-20.7%	-9.9%	-24.8%	-34.5%	-34.1%			
NfL+miR181ab poor Px $^4$	-25.1%	-9.3%	-28.9%	-33.8%	-34.9%			
NfL median split <sup>5</sup>	-26.5%	-29.2%	-29.4%	-34.2%	-34.4%			
NfL 4-level split <sup>6</sup>	-31.4%	-33.8%	-33.4%	-34.8%	-34.6%			

Values indicate the combined percent sample size reduction when the prognostic identified by a row heading is added to covariates described in column headings, in a hypothetical clinical trial with ALSFRS-R as the outcome measure, assuming the experimental therapeutic has a 30% treatment effect.

<sup>1</sup> Core clinical predictors of functional decline include bulbar onset, diagnostic delay, and  $\Delta$ FRS.

- <sup>2</sup> ENCALS predictor score is derived from ΔFRS, bulbar onset, diagnostic delay, age at onset, SVC percent predicted, EI Escorial definite ALS, presence of FTD, and presence of a *C9orf72* repeat expansion.
- <sup>3</sup> Poor prognosis based on published optimal combination of NfL and miR-181ab, in which a poor prognostic factor is defined as either (NfL > 109.8 pg/mL) or (NfL > 59.0 pg/mL and miR-181ab > 39,300 UMI) <sup>11</sup>.
- <sup>4</sup> Poor prognosis based on recalculated combination of NfL and mIR-181ab, using thresholds obtained from the current dataset; a poor prognostic factor is defined as either (NfL > 80.8 pg/mL) or (NfL > 44.8 pg/mL and mIR-181ab > 24,590 UMI<sup>2</sup>).
- <sup>5</sup> Median serum NfL = 67.9 pg/mL
- <sup>6</sup> Serum NfL 4-level split is at the 33<sup>rd</sup>, 50<sup>th</sup>, and 67<sup>th</sup> percentiles (44.8 pg/mL, 67.9 pg/mL, and 80.8 pg/mL, respectively), i.e., tertiles and median, rather than quartiles to mimic construction of the NfL+miR18ab measure <sup>11</sup>.



Baseline NfL (pg/mL)

а



