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Breisgau 79106, Germany Tel: +49-170-5613312 Fax: +49-766-44037772

E-mail: gkirste@icloud.com

Corresponding author: Guenter Kirste Department of Surgery, University Hospital of Freiburg, Albert Ludwig University of

Freiburg, Hugstetter St., 55, Freiburg im

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Cold but not too cold: advances in hypothermic and normothermic organ perfusion

Guenter Kirste

Department of Surgery, University Hospital of Freiburg, Albert Ludwig University of Freiburg, Freiburg im Breisgau, Germany

Transplantation is the method of choice and, in many cases, the only method of treatment for patients with end-stage organ disease. Excellent results have been achieved, and the main focus today is to extend the number of available donors. The use of extended-criteria donors or donors after circulatory death is standard, but is accompanied by an increased risk of ischemia reperfusion injury. This review presents newly developed machine perfusion techniques using hypothermic, subnormothermic, or normothermic conditions, with or without oxygenation. Possibilities for treatment and quality assessment in decision-making about organ acceptability are also discussed.

Keywords: Organ donation; Organ protection; Cold storage; Normothermic machine perfusion; Ischemic reperfusion injury

INTRODUCTION

With the very first transplantations in clinical medicine [1], discussions started about how best to protect organs from damage, how to keep them alive and functioning, and how to preserve them over an extended period of time [2]. Knowing that a decrease in temperature slows all biological processes and reactions, simple cold storage was the basis of organ preservation over many decades. Cold storage and perfusion with cold solutions to wash out the blood from small vessels in order to avoid clotting were used with quite good success over many years [3].

In this context, scientific discussions focused on the composition of the perfusion fluids to establish an equilibrium of electrolytes similar to that in a physiologic environment, with a high concentration of intracellular potassium and a normal concentration of extracellular sodium. To establish such an equilibrium, cell membranes need to be intact and to work at least on a low level. This type of work in the cell needs energy and produces substances such as lactate, the accumulation of which changes the pH, causing further cell damage.

Several perfusion solutions are available, and some of them are effective in the wash-out of blood and cells, such as Ringer solution and EuroCollins [4]. In particular, Euro-Collins and its modifications have been used over many years as a cheap and effective perfusion solution making it possible to keep kidneys viable over up to 24 hours.

However, with the increasing number of liver transplantations and transplantations of other organs, several experimental trials were conducted to find better solutions for organ protection. These efforts led to the development of two solutions, which are widely accepted and are still in use all over the world. Both have different underlying principles. Histidine-tryptophan-ketoglutarate (HTK) solution, developed by Brettschneider in Germany [5,6], is solely intended to keep the pH value stable through a very potent buffer system [7]. In contrast, the University of Wisconsin (UW) solution supplies cells with a number of substances delivering energy to the cell [8].

HIGHLIGHTS

- To establish good graft function Ischemia reperfusion injury (IRI) must be reduced as much as possible.
- Normothermic machine perfusion (NMP) is currently the best method to avoid IRI.
- NMP is specifically important in heart transplantation and is the standard procedure in DCD.
- More sophisticated perfusion technique allows evaluation and even treatment of organs for transplantation.

UW solution was developed at the University of Wisconsin by Southard and Belzer with the intention of keeping cells alive and working [9]. HTK was already in use as a cardioplegic solution in heart surgery and was well known to clinicians, but it had not been tested for the extended storage and protection of organs. HTK was used as a whole-body perfusion solution for all abdominal organs, including the pancreas and small bowel, and for the heart and lungs. UW solution was only used for abdominal organs. The combined use of both solutions made it necessary to encircle and clamp the abdominal aorta directly below the diaphragm when both retrieval teams (thoracic and abdominal) were ready to start perfusion.

While UW solution can be used in small volumes, sometimes even after precooling and wash-out of blood with Ringer solution, HTK solution is used with a high volume (e.g., 20 L or more) depending on the body weight to establish a stable buffer system. Some publications reported inferior outcomes with the use of HTK solution [10]; however, this was often only due to the use of inadequate volume for perfusion.

Several trials compared both solutions [11-14]. Not all of them reached statistical significance. However, when using a reduced incidence of delayed graft function (DGF) as a surrogate for good organ protection, there are data showing superiority of both HTK and UW solutions in comparison to others such as EuroCollins, Marshall, and Celsior [15].

The extended analysis done by Opelz and Döhler [16] with data from the Collaborative Transplant Study (CTS) registry demonstrated that in kidney transplantation, all solutions were safe and effective when the ischemia time was restricted to 18 hours. Only with the extension of ischemia time to more than 36 hours was there reduced graft survival at 3 years, specifically in the very small group of

HTK-perfused kidneys.

One could ask, is it necessary to further extend the ischemia time? There are several reasons for doing so. First, with the introduction of human leukocyte antigen (HLA) typing and the allocation of organs on the basis of a good HLA match, the time for organ transportation has increased due to longer distances between the donor hospital and recipient center. Second, there are increasingly many donors not only for kidney donation, but also for liver and pancreas donation. The liver and pancreas are less tolerant of ischemia than the kidney. With the limited number of trained transplant surgeons and organs allocated to donor centers, liver transplantation is performed as the first procedure, followed by kidney transplantation, making it necessary to extend the storage time of the kidneys. Third, time is necessary for an advanced evaluation of donors to prevent the transmission of infectious diseases from the donor to recipient [17].

With the intention to allow longer storage time, some centers have continued to use the technique of machine perfusion. Belzer [3] was one of the pioneers in the field. As early as the 1960s, he published a 17-hour preservation time on a machine followed by successful transplantation. However, simple cold storage with EuroCollins solution or other solutions had the advantage of being very simple and cost-effective. The disadvantage of this technique was the accumulation of metabolites and the change in pH, leading to ischemia reperfusion injury (IRI), DGF, and reduced graft survival of the transplanted organ [18]. These effects were more evident with the use of organs with extended donor criteria (ECD) or donors after circulatory death (DCD) [19]. In the 1970s and even 1980s, trauma was the main cause of brain death. This is no longer the case. The donor age has increased enormously, and the majority of cases now involve brain death as a consequence of stroke and cerebral bleeding. Moreover, many donors have a long history of hypertension, diabetes, and other diseases affecting organ quality [20,21]. ECD and organs from DCD are widely used. In many countries, these are the only categories of donors with increasing numbers, as the numbers of standard criteria donors (SCD) and living donors (LD) are stabilizing or even decreasing. Of course, when using organs from ECD and DCD, the risk of developing IRI is elevated [22]. This issue has prompted a renewed discussion on how to avoid IRI by using pump machines or supplying nutrients or oxygen.

Machine perfusion also makes it possible to evaluate the quality of an organ and to decide whether it is accept-

able. It has even become feasible to supply organs with substances that improve organ quality [21]. Thus, discussions about whether to keep an organ on the machine and use the time to achieve better organ function or to perform transplantation as quickly as possible to avoid damage have started again.

The current state of the discussion is reviewed below, with a main focus on abdominal organ perfusion, preservation, and protection.

KIDNEY PRESERVATION

Simple cold perfusion and cold storage remains the method of choice for SCD kidneys if the ischemia time is not extended to more than 18 to 24 hours. In practice, this means that transplantation should be performed as soon as possible, even in the middle of the night if necessary [23]. However, this is no longer feasible in several countries, including the Netherlands, as restrictions in working time and hours of personnel have made it necessary to postpone kidney transplantation to the regular schedule of the next day. These regulations have resulted in longer preservation times. The question of whether machine perfusion is superior to cold storage and allows a longer preservation time was first addressed in a randomized trial published by Moers et al. [24,25].

This trial included donors from the Netherlands, Belgium, and the federal state North Rhine-Westphalia in Germany with one kidney of the same donor transplanted after cold storage and the other perfused on a kidney pump provided by Organ Recovery Systems. This landmark publication showed that machine perfusion was associated with a significant decrease in graft loss at 1 year. The number of cases with DGF was reduced in the machine perfusion group, without reaching statistical significance. However, a subgroup analysis of ECD kidneys in the same trial showed a significantly reduced risk of DGF, implying that this methodology at least has advantages in the protection of non-SCD kidneys [26].

With the background of this publication, several perfusion systems and machines were either developed or modified. It is difficult to compare these machines and the published results because they differ in terms of the temperature used, oxygen and nutrient supply, duration of perfusion, and technical details such as pulsatile or non-pulsatile perfusion, automation and monitoring. No scientific trials have directly compared the use and efficacy of one apparatus to that of another [27-31].

Moreover, there is growing interest in using different techniques sequentially, such as starting with cold perfusion and storage followed by rewarming before implantation [32-34]. Depending on the availability of techniques in the donor hospital, the most frequently used models involve starting with simple cold storage followed by cold perfusion at a dedicated (usually university-based) procurement center, followed by rewarming either with or without oxygen shortly before transplantation. A number of different methods for organ perfusion have been recurrently discussed (Table 1) [35]. The idea of pulsatile normothermic perfusion is not new at all. The very first attempts of organ preservation tried to mimic physiologic conditions as much as possible [36]. In the 1960s, Belzer et al. [3] introduced the method of hypothermic perfusion of the kidneys, which was simpler than normothermic machine perfusion (NMP). However, the simpler the better. The breakthrough in simple cold storage came when Collins et al. [4] introduced specific solutions for cold organ preservation.

Nonetheless, even in conditions of reduced metabolism under cold conditions, there is an accumulation of metabolites leading to IRI. The increasing number of ECD and even the use of high numbers of DCD in several countries led to growing interest in the revival of a more physiologic environment during perfusion and storage [37,38]. This requires advanced technology, ideally with NMP from the donor hospital to the transplant center [39]. Other perfusion solutions can be used, adding different nutrients. In general, normothermic perfusion enables normal cellular metabolism, specifically when adding oxygen [40]. It could even allow repair of injuries and more sophisticated testing of functionality and viability [21,41]. The need to add oxygen for organ perfusion was already discussed in the 1970s. Several experimental and clinical studies were published

Method

- · Cold (hypothermic) perfusion and storage
- · Hypothermic machine perfusion
- Hypothermic oxygenated machine perfusion
- Subnormothermic machine perfusion
- Normothermic machine perfusion
- · Normothermic oxygenated machine perfusion
- Normothermic regional perfusion in donors after circulatory death

using oxygen and measuring oxygen pressure on the surface of transplanted kidneys [42]. However adding oxygen requires an oxygen carrier, which ideally would be blood. Other carriers or the simple use of oxygen as an additive to the solution did not show meaningful effects [43-46].

In general, the results of kidney transplantation are very good when using sophisticated protocols of immunosuppression and patient monitoring. These protocols and HLA matching as the basis for the allocation of kidneys have helped to achieve these good results. However, it was demonstrated years ago by the CTS that the difference in graft function at 3 years between cases with zero mismatches and those with a full-house match was not as large as the difference based on the simple and subjective impression of the transplant surgeon regarding whether a kidney was good or suboptimal [47].

Thus, to further improve the results of kidney transplantation, the focus is on organs that are suboptimal or originate from ECD or DCD. The Kidney Donor Risk Index, preimplantation biopsy, analysis of pump and perfusate parameters, and other parameters might help to distinguish between high-quality and suboptimal kidneys [48-52]. As a very sophisticated method, the use of nuclear magnetic resonance imaging to measure the adenosine triphosphate content of cells in a kidney ready for implantation showed close correlations with outcomes [53,54]; however, this technique has never been introduced into clinical practice. These parameters are not sufficiently exact or reproducible to determine whether to discard a kidney. Furthermore, research is needed to evaluate whether the improvement of kidney function using pump techniques is detectable. The study by Moers et al. [24] introduced hypothermic machine perfusion (HMP) without oxygenation and excluded the effect of all donor parameters by allocating one kidney from the same donor to HMP and the other to static cold storage (SCS). The donors were consecutive cases from the Netherlands, Belgium, and North Rhine-Westphalia in Germany, all above the age of 16. In total, 359 donors were enrolled, and 336 in the HMP group and 336 in the SCS group could be assessed. In the analysis of the whole study population, there was a significant difference concerning DGF, but only in the subgroup analysis of ECD, and in further studies with DCD the advantage of the HMP could be demonstrated very clearly [26,55]. Further analysis revealed that "traditional" risk factors such as cold ischemia time (CIT), time on dialysis, and the origin of kidney disease had an even higher impact on graft survival. It is well known that CIT is an important risk

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factor. A reason for the good results in LD is the extremely short ischemia time of only a few hours, or even shorter at some centers. To reduce CIT, NMP should be used, not HMP as in the Moers et al.'s study [24]. This technique was introduced by a group from Cambridge and evaluated in different settings, most often starting with a period of SCS followed by NMP, either as pre-warming before transplantation or over the whole time of storage and transportation [35,56]. Other studies with the same approach have been conducted in DCD. The evaluation is done using a specific newly developed scoring system. However, the authors warned not to use this scoring system alone as the basis for deciding whether to discard kidneys [57]. The parameters used include macroscopic appearance, blood flow, und urine output.

Whether prolonged NMP can help restore or even improve kidney function is not yet clear. The promising results from Kaths et al. [58] in an experimental setting using porcine kidneys from donation after brain death (DBD) or DCD showed that NMP was at least equivalent to SCS in terms of tubular injury and kidney function. Studies with long-term preservation involving 24-hour NMP showed that the kidneys were well preserved and functional. This proves that NMP for over 24 hours is possible. However, these results are not that very surprising, since several cases have been published wherein SCS alone and CIT for over 36 to 48 hours were followed by successful transplantation. Nevertheless, there is certainly a place for NMP in kidney preservation when it comes to ECD and DCD. A further study (the HOPE study) focused on the question of whether NMP with additional oxygen could improve kidney function [59]. The results are promising, with a clear advantage of NMP over SCS. Other studies, such as the COPE, POMP, COMPARE have not yet been finished [60].

HMP opens the possibility of evaluating kidney function with the intention to increase the number of available organs from ECD and DCD. NMP could be effective for the repair of kidneys either alone or by administration of various drugs [61]. Even immune modulation and other methods to reduce organ damage, such as the use of anti-inflammatory drugs, can be introduced. Early and specific immunosuppression becomes feasible. These new developments are excellently summarized in a review published by Hamelink et al. [62].

REGIONAL NORMOTHERMIC PERFUSION

With the introduction of extracorporeal membrane oxygenation in intensive care, a method became available that also seemed to be suitable for regional perfusion of organs after death declaration. A number of questions have been answered concerning maintenance of the permanence principle for death during in situ regional perfusion [63]. However, there is no international agreement about the use of DCD.

In the overwhelming majority of scientific publications, death in DCD is defined as the permanent cessation of brain circulation [64]. In countries where DCD is legally not prohibited, methods have been developed to start regional perfusion of abdominal organs and even restart heart beating without reperfusion of the brain. These techniques have been extensively discussed by a number of expert groups in the UK, Canada, and other countries [63]. After confirmation of death in DCD and before starting normothermic regional perfusion (NRP), it is considered appropriate to perform surgery to ligate or divide the aortic arch vessels for cases with thoracic NRP or to occlude the descending aorta for abdominal NRP [65]. Opening the vessels of the supra-aortic arch to the atmosphere ensures that there is no reperfusion to the brain. The details of the underlying principles and the methodology have been described in detail by Manara et al. [63] as a result of an international working group defining death [64,66].

With strict compliance with these regulations not to restore perfusion of the brain, NMP offers a unique opportunity to maintain physiological conditions through oxygen and nutrient delivery. Moreover, it provides enough time to evaluate the suitability of organs for transplantation, and the possibility of treating an organ and restoring complete function is even apparent.

The first results of NRP in DCD were published in 2014 from a group in the UK [67]. Thirty-two kidney transplants, 11 liver transplants, two combined pancreas-kidney transplants, and three double lung transplants were successfully performed. In a second trial, 43 livers were transplanted after NRP from DCD and compared to a historical group of livers after cold storage [68]. The IRI decreased significantly. Another group from Spain obtained similar results.

LIVER PERFUSION

The first randomized trial comparing SCS with NRP in liver procurement was published in 2018 [69]. Most recently, in January 2022, a randomized multicenter trial (PROTECT) compared 293 patients that received livers, of whom 151 were in the Organ Care System (TransMedics, Andover, MA, USA), while 142 were transplanted after cold storage [70]. The primary endpoint was a significant decrease in early allograft dysfunction, which occurred in 27 out of 151 (18%) livers in the Organ Care System arm and in 44 out of 142 (31%) livers in the cold storage arm. The livers in the Organ Care System group showed a significant reduction of IRI. The use of livers from DCD increased significantly. There was a significant reduction of ischemic-type biliary lesions in the Organ Care System group. These results indicate that the use of machine perfusion, at least for livers from ECD or DCD, will be the standard in the future. However, a number of guestions remain open: (1) The distinction between SCD and ECD, (2) Assessment of the liver on the machine, (3) time on the machine, (4) cold perfusion in the donor followed by NMP or NRP, (5) possibilities of treating the liver (e.g., a steatotic liver), (6) Treatment of hepatitis C-positive donor livers, (7) Changes in the immune status of the liver cells to avoid rejection. These questions need to be addressed in randomized trials.

The results of the trials published to date have made it possible to increase the number of livers for transplantation, specifically from ECD [71-73]. Assessment of the liver and its function is critical for further increasing the number of available livers. Researchers from both Birmingham and Cambridge have been working on this guestion [74]. Lactate clearance, glucose metabolism, level of transaminases, bile production, und pH stability during NMP are suitable parameters for predicting outcomes after transplantation. However, a further analysis of bile composition with bile pH, glucose, and bicarbonate is needed to predict ischemic-type biliary lesions. The work of this group proved a number of parameters to be of predictive value (Table 2) [35]. The validity of these parameters was even challenged by a study by the Birmingham group [75]. Livers that were declined by seven transplant centers were perfused and, if the above criteria were fulfilled, transplanted with good 90-day patient survival.

In many cases, the reason to classify a liver as an ECD organ is steatosis. Some centers have reported that 13% to 28% of livers were steatotic, while others have reported far more, with rates exceeding 60% depending on the

Table 2. Cambridge criteria of variables associated with successful transplantation of NMP livers [35]

Criteria

- Maximum bile pH >7.5
- Bile glucose concentration <3 mmol/L or >10 mmol/L less than perfusate glucose
- Ability to maintain perfusate pH >7.2 without >30 mol bicarbonate supplementation
- Falling glucose beyond 2 hours or perfusate glucose under 10 mmol/L, which on challenge with 2.5 g of glucose, does fall subsequently
- Peak lactate fall >4.4 mmol/L/kg/hr
- Alanine aminotransferase <6,000 U/L at 2 hours

NMP, normothermic machine perfusion.

grade of steatosis and whether the fat is accumulated in the liver cells in micro- or macro-bubbles. Several experimental attempts have been made to reduce fat in the liver. While the approach of using NMP seems to be quite successful in reducing the fat in liver cells in animal experiments, the same approach has not been that successful in clinical settings [76-78]. Even the addition of defatting agents showed only a minimal reduction [79,80]. It is not clear whether an even more extended perfusion time is necessary to reduce fat, or whether other approaches such as lipid aphaeresis are needed. There is certainly an advantage of NMP versus HMP when looking at approaches to reduce fat in the liver [81].

Other approaches are possible and subject to clinical trials. Adding antiviral substances such as miravirsen to the perfusion could reduce reinfection with hepatitis C virus. The downregulation of pro-inflammatory cells, gene therapy, and immune modulation using CTLA4-Ig are other approaches. Most of these have only been tested in experimental studies in animals.

Few studies have directly compared NMP and HMP. Instead, the addition of oxygen to either method is a promising approach. Dutkowski et al. [82] showed good results in DCD livers concerning graft survival and reduction of ischemic-type biliary lesions when adding oxygen in comparison to cold storage [83-85]. This was also confirmed by a number of other investigators. However, few cases were studied and some of the studies only compared the results with historical groups.

There is a need to systematically evaluate HMP versus NMP, both with and without the addition of oxygen. Subnormothermic machine perfusion (SMP) has also been studied, but not directly compared with cold storage [86].

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To date, there is no scientifically proven way to decide which method of perfusion should be used in different types of donors. Machine perfusion is certainly more challenging from a technical standpoint. Even with the Organ Care System liver perfusion system from TransMedics, which is the simplest device on the market, there is a need for training personnel to avoid mistakes and casualties in the beginning. In many cases, it might be suitable in the management of the donor to simply perfuse a liver with cold solution, transport it to a procurement center, and decide which method of NMP should be used, with the addition of whichever substances are deemed to be necessary in a specific case [87]. Organ perfusion and protection have become more difficult and challenging. The use of these techniques may even be possible in split liver procurement [88]. This possibility would involve the creation of dedicated and well-equipped procurement centers dealing with the optimal perfusion of different organs, assessment of function, evaluation of outflow parameters, allocation of each organ to a specific recipient, and subsequent transfer to the transplant center.

The improvement in perfusion and preservation techniques is very important. However, in most recently published studies, the simply calculated parameters of donor hepatectomy time and implantation time have also shown significant impacts on early graft dysfunction [89,90]. Therefore a skilled and well-trained surgeon is still the best way to improve transplant results.

PANCREAS AND ISLET CELLS

Major problems in pancreas transplantation are the development of posttransplant pancreatitis, steatosis in the organ, and non-function or insufficient function of insulin producing β -cells. Only a few studies have directly compared SCS with either HMP or NMP [91]. Insulin secretion was detectable during machine perfusion, but with very reduced or limited exocrine function. It is not clear whether the use of machine perfusion would increase the number of available organs from ECD or reduce IRI. A study used discarded organs with oxygenated HMP over 6 hours [92]. There were no signs of cell oedema, and the islet cells could be isolated with good viability and function. However, no data on the success rate of whole-organ transplantation have been published. A prolonged CIT seems to be detrimental for pancreatic transplantation. The only option

to reduce the CIT, of course, is NMP. Barlow et al. [93] has implemented this approach, showing good function and results, but with a small number of cases so far. Before introducing this technique as standard in the organ donation process, further evaluation and studies are needed. Monitoring exocrine function during perfusion is certainly a good way to evaluate the organ. Which type of perfusion solution and which treatment should be used to restore good function must also be evaluated [94].

HEART AND LUNG PERFUSION

The situation of heart transplantation is completely different from that of abdominal organ transplantation. CIT must be limited to 4 hours, or at most 6 hours. With longer CIT, heart transplantations fail in most circumstances. With such a short CIT compared to kidneys, the risk of IRI is far lower. Therefore, the approach of cardiac surgeons to optimize perfusion systems is driven by purposes other than reducing the CIT. First, there is the need to increase the number of available hearts for transplantation by using ECD and even DCD, and second, perfusion systems prolong the time available for transportation and storage. This gives more time for the transplant team to prepare the recipient for implantation despite often having a history of several operations involving the implantation of multiple ventricular assist systems, valve replacements, and so on. Third, a perfusion system allows well-monitored reconditioning of the heart, evaluation of biological parameters (especially lactate concentration), and evaluation of functional pump parameters (e.g., by changing the preload and afterload of the heart) [95,96]. Even an assessment of the heart on the machine by coronary angiography is possible. Treatment of the heart is possible, and gene therapy could be tested in the future.

Evidently, several factors make life easier for the heart surgeon, the most important of which is prolonging the time between the first cold ischemia and the second cold ischemia, when the heart is off the machine and sewed into the recipient.

The development of these machines was not driven primarily by a scientific approach, but by the idea of making certain options technically possible. The first such machine ready for clinical use is the Organ Care System [97]. The use of this machine and its attachment for the heart needs experience and training. Most trials so far have set the primary endpoint as non-inferiority compared to simple cold storage [98]. Some trials failed and were stopped mostly because centers did not have enough experience and training to run the system. Other single-center trials, however, showed significant improvements in early and late outcomes [99,100]. Many more ECD and even DCD organs were used and transplanted successfully [101-104]. The first PROTECT trial in Europe reported a 30-day graft survival rate of 100% using SCD organs [105]. The first trial in the United States, named PROCEED [97] reported a 30day patient survival rate of 93% [106]. In both trials, the combined primary and secondary CIT was significantly reduced to 60 to 80 minutes, but with a prolonged machine preservation time.

However, the management of the apparatus is complex. A substantial amount (1 to 1.5 L) of donor blood is needed and rapidly taken from the donor via aortic cannulation within 40 seconds prior to aortic clamping. Abdominal retrieval teams do not favor this procedure because of the rapid fall in blood pressure. Moreover, the requirement for blood to be used in different machines is even higher when NMP for the lung, liver, and kidneys is considered as well.

At Harefield Hospital in London, García Sáez et al. [98] has reported a high number of Organ Care System cases. Thirty donors were evaluated, and the hearts were attached to the machine with the target of a coronary artery flow of 900 to 1,000 mL/min. If function deteriorates on the machine or the heart does not return to sinus rhythm, manual massage in the apparatus is possible as well as defibrillation shock. Every 30 minutes, lactate is measured together with electrolytes and other parameters. Lactate should stay stable at a level of <5 mmol/L and always be less on the venous side than on the arterial side. With increasing lactate levels, the risk of non-function after implantation is high. With more experience with the system, even hearts with a runtime on the machine of 10 to 11 hours were used [107]. Nonetheless, there was still a reduction in combined CIT. Heart transplantation using DCD hearts has become feasible. Despite the high costs of the machine and the one-time usable material, the use of Organ Care System has become the standard at many centers.

In lung transplantation, *ex vivo* lung perfusion (EVLP) was developed to evaluate lungs specifically from DCD before implantation. The Ontario group demonstrated that the treatment of ECD lungs is possible to bring them to a status good enough for transplantation [108,109].

However, the greatest number of additional donors can be reached by consequently using every single case of DCD [110]. Different apparatuses have been developed, not all of which are portable. The Organ Care System produced by TransMedics is the system used most widely. The flow in the machine is 2-2.5 L/min, and air is ventilated into the lung from outside. The device allows monitoring of various parameters, such as pulmonary artery pressure, pulmonary vascular resistance, and peak airway pressure. The introduction of EVLP was followed by an increase in the number of available organs for transplantation. A study has shown that the results were more or less the same when comparing DBD lungs with DCD lungs [111]. Some very preliminary data show that standard cold perfusion and storage can be used initially, followed by attachment to an EVLP system. This could make it easier for a procurement team in a small rural hospital to start with cold perfusion, followed by quick transport to a specialized perfusion center and transplantation after a period of treatment and evaluation on the machine.

CONCLUSIONS

Organ transplantation is a very successful treatment modality, and in many cases, it is the only therapy for endstage organ diseases. In recent decades, the main focus of scientific investigations and research has been on ways to improve immunosuppression. Numerous protocols have been compared and numerous substances are licensed and available. The protocols for studies to compare or to prove superiority have become extremely difficult. Specifically, a large number of cases is required to reach statistical significance. Donor criteria have changed enormously, as well as recipient factors.

The main focus now is to increase the number of donors, which is only possible by using more ECD or even organs from DCD. Therefore, there is increasing interest in methods of evaluating and improving the function and quality of organs. To improve function means first to exclude all damaging factors and second to use methods for treatment.

CIT is certainly the most important factor for IRI. Several approaches to reduce CIT have been evaluated, such as regional machine perfusion, NMP, SMP, and others. The results of ongoing studies might provide insights on how to decide, how to proceed, and which method to choose.

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The limiting factor for the use of NMP systems in several organs (e.g., the heart, lungs, liver, and kidneys) is the amount of blood available from a single donor. Other oxygen carriers need to be evaluated. Further improvement is feasible using better oxygen delivery to the organ, drugs for the repair of cells, or gene therapy to make the organ less susceptible to rejection. Other methods, such as supercooling and storage at temperatures below 0°C, remain on the horizon of scientific work, but need further evaluation [112].

The costs for organ perfusion increase with the use of machines. However, there are clear advantages at least for normothermic and subnormothermic techniques for regional or single organ perfusion of the heart, lung, and liver is concerned.

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Conflict of Interest

Guenter Kirste is an editorial board member of the journal but was not involved in the peer reviewer selection, evaluation, or decision process of this article. No other potential conflict of interest relevant to this article was reported.

ORCID

Guenter Kirste

https://orcid.org/0000-0002-7184-5751

REFERENCES

- 1. Leeson S, Desai SP. Medical and ethical challenges during the first successful human kidney transplantation in 1954 at Peter Bent Brigham Hospital, Boston. Anesth Analg 2015;120:239-45.
- 2. Belzer FO, Ashby BS, Dunphy JE. 24-Hour and 72-hour preservation of canine kidneys. Lancet 1967;2:536-8.
- Belzer FO, Ashby BS, Gulyassy PF, Powell M. Successful seventeen-hour preservation and transplantation of human-cadaver kidney. N Engl J Med 1968;278:608-10.
- 4. Collins GM, Bravo-Shugarman M, Terasaki PI. Kidney preservation for transportation: initial perfusion and 30 hours' ice storage. Lancet 1969;2:1219-22.
- Groenewoud AF, Buchholz B, Gubernatis F, Hölscher M, Hoyer J, Isemer F, et al. First results of the multi-

center study of HTK protection for kidney transplants. Transplant Proc 1990;22:2212.

- Isemer FE, Ludwig A, Schunck O, Bretschneider HJ, Peiper HJ. Kidney procurement with the HTK solution of Bretschneider. Transplant Proc 1988;20:885-6.
- Eble FJ, Kirste G, eds. Single centre results of organ preservation with Eurocollins, HTK and UW solution. Proceedings of Custodiol-Symposium; 1992 Jun 29; Freiburg im Breisgau, Germany.
- 8. Ploeg RJ. Kidney preservation with the UW and Euro-Collins solutions: a preliminary report of a clinical comparison. Transplantation 1990;49:281-4.
- Southard JH. Biochemistry and cell physiology of organ preservation. In: Collins GM, Dubernard JM, Land W, Persijn GG, eds. Procurement, preservation and allocation of vascularized organs. Dordrecht, NL: Springer; 1997. p. 103-13.
- Stewart ZA, Lonze BE, Warren DS, Dagher NN, Singer AL, Montgomery RA, et al. Histidine-tryptophan-ketoglutarate (HTK) is associated with reduced graft survival of deceased donor kidney transplants. Am J Transplant 2009;9:1048-54.
- 11. Pedotti P, Cardillo M, Rigotti P, Gerunda G, Merenda R, Cillo U, et al. A comparative prospective study of two available solutions for kidney and liver preservation. Transplantation 2004;77:1540-5.
- Lynch RJ, Kubus J, Chenault RH, Pelletier SJ, Campbell DA, Englesbe MJ. Comparison of histidine-tryptophan-ketoglutarate and University of Wisconsin preservation in renal transplantation. Am J Transplant 2008;8:567-73.
- de Boer JD, Strelniece A, van Rosmalen M, de Vries E, Ysebaert D, Guba M, et al. The effect of histidine-tryptophan-ketoglutarate solution and University of Wisconsin solution: an analysis of the Eurotransplant registry. Transplantation 2018;102:1870-7.
- 14. Groenewoud AF, Thorogood J. Current status of the Eurotransplant randomized multicenter study comparing kidney graft preservation with histidine-tryptophan-ketogluterate, University of Wisconsin, and Euro-Collins solutions. The HTK Study Group. Transplant Proc 1993;25(1 Pt 2):1582-5.
- 15. O'Callaghan JM, Knight SR, Morgan RD, Morris PJ. Preservation solutions for static cold storage of kidney allografts: a systematic review and meta-analysis. Am J Transplant 2012;12:896-906.
- 16. Opelz G, Döhler B. Multicenter analysis of kidney preservation. Transplantation 2007;83:247-53.

- 17. European Directorate for the Quality of Medicines and HealthCare. Guide to the quality and safety of organs for transplantation, 8th ed [Internet]. Strasbourg: European Directorate for the Quality of Medicines and Healthcare; 2022 [cited 2022 Mar 8]. Available from: https://www.edqm.eu.
- Kayler LK, Srinivas TR, Schold JD. Influence of CIT-induced DGF on kidney transplant outcomes. Am J Transplant 2011;11:2657-64.
- Wind J, Snoeijs MG, van der Vliet JA, Winkens B, Christiaans MH, Hoitsma AJ, et al. Preservation of kidneys from controlled donors after cardiac death. Br J Surg 2011;98:1260-6.
- Echterdiek F, Latus J, Döhler B, Schwenger V, Süsal C. Influence of cold ischemia time on the outcome of kidney transplants from donors aged 70 years and above: a collaborative transplant study report. Transplantation 2021;105:2461-9.
- 21. Weissenbacher A, Stone JP, Lo Faro ML, Hunter JP, Ploeg RJ, Coussios CC, et al. Hemodynamics and metabolic parameters in normothermic kidney preservation are linked with donor factors, perfusate cells, and cytokines. Front Med (Lausanne) 2022;8:801098.
- Fischer-Fröhlich CL, Kutschmann M, Feindt J, Schmidtmann I, Kirste G, Frühauf NR, et al. Influence of deceased donor and pretransplant recipient parameters on early overall kidney graft-survival in Germany. J Transplant 2015;2015:307230.
- 23. Terasaki PI. Cold ischemia time: time to rethink the risk for kidneys? Am J Transplant 2011;11:2551-2.
- 24. Moers C, Smits JM, Maathuis MH, Treckmann J, van Gelder F, Napieralski BP, et al. Machine perfusion or cold storage in deceased-donor kidney transplantation. N Engl J Med 2009;360:7-19.
- 25. Moers C, Pirenne J, Paul A, Ploeg RJ; Machine Preservation Trial Study Group. Machine perfusion or cold storage in deceased-donor kidney transplantation. N Engl J Med 2012;366:770-1.
- 26. Treckmann J, Moers C, Smits JM, Gallinat A, Maathuis MH, van Kasterop-Kutz M, et al. Machine perfusion versus cold storage for preservation of kidneys from expanded criteria donors after brain death. Transpl Int 2011;24:548-54.
- 27. Smith TB, Nicholson ML, Hosgood SA. Advances in hypothermic and normothermic perfusion in kidney transplantation. Transplantology 2021;2:460-77.
- 28. Jochmans I, O'Callaghan JM, Pirenne J, Ploeg RJ. Hypothermic machine perfusion of kidneys retrieved

from standard and high-risk donors. Transpl Int 2015;28:665-76.

- 29. Brat A, de Vries KM, van Heurn EW, Huurman VA, de Jongh W, Leuvenink HG, et al. Hypothermic machine perfusion as a national standard preservation method for deceased donor kidneys. Transplantation 2021 Jun 23 [Epub]. https://doi.org/10.1097/ TP.000000000003845.
- Maathuis MH, Manekeller S, van der Plaats A, Leuvenink HG, Hart NA, Lier AB, et al. Improved kidney graft function after preservation using a novel hypothermic machine perfusion device. Ann Surg 2007;246:982-8.
- Watson CJ, Wells AC, Roberts RJ, Akoh JA, Friend PJ, Akyol M, et al. Cold machine perfusion versus static cold storage of kidneys donated after cardiac death: a UK multicenter randomized controlled trial. Am J Transplant 2010;10:1991-9.
- 32. Ciancio G, Gaynor JJ, Sageshima J, Roth D, Kupin W, Guerra G, et al. Machine perfusion following static cold storage preservation in kidney transplantation: donor-matched pair analysis of the prognostic impact of longer pump time. Transpl Int 2012;25:34-40.
- Gallinat A, Efferz P, Paul A, Minor T. One or 4 h of "inhouse" reconditioning by machine perfusion after cold storage improve reperfusion parameters in porcine kidneys. Transpl Int 2014;27:1214-9.
- O'Neill S, Srinivasa S, Callaghan CJ, Watson CJ, Dark JH, Fisher AJ, et al. Novel organ perfusion and preservation strategies in transplantation: where are we going in the United Kingdom? Transplantation 2020;104:1813-24.
- Weissenbacher A, Vrakas G, Nasralla D, Ceresa CD. The future of organ perfusion and re-conditioning. Transpl Int 2019;32:586-97.
- Carrel A, Lindbergh CA. The culture of whole organs. Science 1935;81:621-3.
- Jochmans I, Moers C, Smits JM, Leuvenink HG, Treckmann J, Paul A, et al. Machine perfusion versus cold storage for the preservation of kidneys donated after cardiac death: a multicenter, randomized, controlled trial. Ann Surg 2010;252:756-64.
- Hosgood SA, Brown RJ, Nicholson ML. Advances in kidney preservation techniques and their application in clinical practice. Transplantation 2021;105:e202-14.
- DiRito JR, Hosgood SA, Tietjen GT, Nicholson ML. The future of marginal kidney repair in the context of normothermic machine perfusion. Am J Transplant

2018;18:2400-8.

- 40. Zulpaite R, Miknevicius P, Leber B, Strupas K, Stiegler P, Schemmer P. Ex-vivo kidney machine perfusion: therapeutic potential. Front Med (Lausanne) 2021;8:808719.
- 41. Baan CC. Repairing and regenerating organs for transplantation has become a reality. Transplantation 2019;103:224-6.
- 42. Wilms H. Renocorticale Sauerstoffversorgung und Nierendurchblutung nach autologer Nierentransplantation beim Hund (thisis). Freiburg: Albert Ludwigs University of Freiburg; 1978.
- 43. Minor T, Paul A, Efferz P, Wohlschlaeger J, Rauen U, Gallinat A. Kidney transplantation after oxygenated machine perfusion preservation with Custodiol-N solution. Transpl Int 2015;28:1102-8.
- 44. Le Meur Y, Delpy E, Renard F, Hauet T, Badet L, Rerolle JP, et al. HEMO2life® improves renal function independent of cold ischemia time in kidney recipients: a comparison with a large multicenter prospective cohort study. Artif Organs 2021 Dec 24 [Epub]. http:// doi.org/10.1111/aor.14141.
- Le Meur Y, Badet L, Essig M, Thierry A, Büchler M, Drouin S, et al. First-in-human use of a marine oxygen carrier (M101) for organ preservation: a safety and proof-of-principle study. Am J Transplant 2020;20:1729-38.
- 46. Fontes P, Lopez R, van der Plaats A, Vodovotz Y, Minervini M, Scott V, et al. Liver preservation with machine perfusion and a newly developed cell-free oxygen carrier solution under subnormothermic conditions. Am J Transplant 2015;15:381-94.
- 47. Opelz G. Collaborative Transplant Study [Internet]. Heidelberg: Collaborative Transplant Study; 2022 [cited 2022 Mar 8]. Available from: https://www.ctstransplant.org/.
- 48. Feng S. Donor intervention and organ preservation: where is the science and what are the obstacles? Am J Transplant 2010;10:1155-62.
- 49. de Vries EE, Hoogland ER, Winkens B, Snoeijs MG, van Heurn LW. Renovascular resistance of machine-perfused DCD kidneys is associated with primary nonfunction. Am J Transplant 2011;11:2685-91.
- 50. Guy AJ, Nath J, Cobbold M, Ludwig C, Tennant DA, Inston NG, et al. Metabolomic analysis of perfusate during hypothermic machine perfusion of human cadaveric kidneys. Transplantation 2015;99:754-9.
- 51. Moers C, Varnav OC, van Heurn E, Jochmans I, Kirste GR, Rahmel A, et al. The value of machine perfusion

perfusate biomarkers for predicting kidney transplant outcome. Transplantation 2010;90:966-73.

- Parikh CR, Hall IE, Bhangoo RS, Ficek J, Abt PL, Thiessen-Philbrook H, et al. Associations of perfusate biomarkers and pump parameters with delayed graft function and deceased donor kidney allograft function. Am J Transplant 2016;16:1526-39.
- Niekisch MB, Von Elverfeldt D, El Saman A, Hennig J, Kirste G. Improved pretransplant assessment of renal quality by means of phosphorus-31 magnetic resonance spectroscopy using chemical shift imaging. Transplantation 2004;77:1041-5.
- 54. Bon D, Billault C, Thuillier R, Hebrard W, Boildieu N, Celhay O, et al. Analysis of perfusates during hypothermic machine perfusion by NMR spectroscopy: a potential tool for predicting kidney graft outcome. Transplantation 2014;97:810-6.
- 55. Thuillier R, Allain G, Celhay O, Hebrard W, Barrou B, Badet L, et al. Benefits of active oxygenation during hypothermic machine perfusion of kidneys in a preclinical model of deceased after cardiac death donors. J Surg Res 2013;184:1174-81.
- 56. Weissenbacher A, Lo Faro L, Boubriak O, Soares MF, Roberts IS, Hunter JP, et al. Twenty-four-hour normothermic perfusion of discarded human kidneys with urine recirculation. Am J Transplant 2019;19:178-92.
- 57. Kaths JM, Hamar M, Echeverri J, Linares I, Urbanellis P, Cen JY, et al. Normothermic ex vivo kidney perfusion for graft quality assessment prior to transplantation. Am J Transplant 2018;18:580-9.
- 58. Kaths JM, Cen JY, Chun YM, Echeverri J, Linares I, Ganesh S, et al. Continuous normothermic ex vivo kidney perfusion is superior to brief normothermic perfusion following static cold storage in donation after circulatory death pig kidney transplantation. Am J Transplant 2017;17:957-69.
- Kron P, Schlegel A, Muller X, Gaspert A, Clavien PA, Dutkowski P. Hypothermic oxygenated perfusion: a simple and effective method to modulate the immune response in kidney transplantation. Transplantation 2019;103:e128-36.
- 60. Consortium for Organ Preservation in Europe (COPE). COPE trials [Internet]. Oxford: COPE; 2019 [cited 2022 Mar 7]. Available from: https://cope-eu.com/work%20 programme/trials.
- 61. van Leeuwen LL, Leuvenink HG, Olinga P, Ruigrok MJ. Shifting paradigms for suppressing fibrosis in kidney transplants: supplementing perfusion solu-

tions with anti-fibrotic drugs. Front Med (Lausanne) 2022;8:806774.

- 62. Hamelink TL, Ogurlu B, De Beule J, Lantinga VA, Pool MB, Venema LH, et al. Renal normothermic machine perfusion: the road toward clinical implementation of a promising pretransplant organ assessment tool. Transplantation 2022;106:268-79.
- 63. Manara A, Shemie SD, Large S, Healey A, Baker A, Badiwala M, et al. Maintaining the permanence principle for death during in situ normothermic regional perfusion for donation after circulatory death organ recovery: a United Kingdom and Canadian proposal. Am J Transplant 2020;20:2017-25.
- 64. Domínguez-Gil B, Ascher N, Capron AM, Gardiner D, Manara AR, Bernat JL, et al. Expanding controlled donation after the circulatory determination of death: statement from an international collaborative. Intensive Care Med 2021;47:265-81.
- 65. Dalsgaard FF, Moeslund N, Zhang ZL, Pedersen M, Qerama E, Beniczky S, et al. Clamping of the aortic arch vessels during normothermic regional perfusion after circulatory death prevents the return of brain activity in a porcine model. Transplantation 2022 Jan 18 [Epub]. https://doi.org/10.1097/ TP.0000000000004047.
- 66. Shemie SD, Gardiner D. Circulatory arrest, brain arrest and death determination. Front Cardiovasc Med 2018;5:15.
- 67. Oniscu GC, Randle LV, Muiesan P, Butler AJ, Currie IS, Perera MT, et al. In situ normothermic regional perfusion for controlled donation after circulatory death: the United Kingdom experience. Am J Transplant 2014;14:2846-54.
- Watson CJ, Hunt F, Messer S, Currie I, Large S, Sutherland A, et al. In situ normothermic perfusion of livers in controlled circulatory death donation may prevent ischemic cholangiopathy and improve graft survival. Am J Transplant 2019;19:1745-58.
- 69. Nasralla D, Coussios CC, Mergental H, Akhtar MZ, Butler AJ, Ceresa CD, et al. A randomized trial of normothermic preservation in liver transplantation. Nature 2018;557:50-6.
- 70. Markmann JF, Abouljoud MS, Ghobrial RM, Bhati CS, Pelletier SJ, Lu AD, et al. Impact of portable normothermic blood-based machine perfusion on outcomes of liver transplant: the OCS Liver PROTECT randomized clinical trial. JAMA Surg 2022;157:189-98.
- 71. op den Dries S, Karimian N, Sutton ME, Westerkamp

AC, Nijsten MW, Gouw AS, et al. Ex vivo normothermic machine perfusion and viability testing of discarded human donor livers. Am J Transplant 2013;13:1327-35.

- 72. Mergental H, Laing RW, Kirkham AJ, Perera MT, Boteon YL, Attard J, et al. Transplantation of discarded livers following viability testing with normothermic machine perfusion. Nat Commun 2020;11:2939.
- 73. Minor T, Efferz P, Fox M, Wohlschlaeger J, Lüer B. Controlled oxygenated rewarming of cold stored liver grafts by thermally graduated machine perfusion prior to reperfusion. Am J Transplant 2013;13:1450-60.
- 74. Watson CJ, Kosmoliaptsis V, Randle LV, Russell NK, Griffiths WJ, Davies S, et al. Preimplant normothermic liver perfusion of a suboptimal liver donated after circulatory death. Am J Transplant 2016;16:353-7.
- 75. Laing RW, Mergental H, Yap C, Kirkham A, Whilku M, Barton D, et al. Viability testing and transplantation of marginal livers (VITTAL) using normothermic machine perfusion: study protocol for an open-label, non-randomised, prospective, single-arm trial. BMJ Open 2017;7:e017733.
- Jamieson RW, Zilvetti M, Roy D, Hughes D, Morovat A, Coussios CC, et al. Hepatic steatosis and normothermic perfusion-preliminary experiments in a porcine model. Transplantation 2011;92:289-95.
- 77. Fondevila C, Hessheimer AJ, Maathuis MH, Muñoz J, Taurá P, Calatayud D, et al. Hypothermic oxygenated machine perfusion in porcine donation after circulatory determination of death liver transplant. Transplantation 2012;94:22-9.
- Op den Dries S, Sutton ME, Karimian N, de Boer MT, Wiersema-Buist J, Gouw AS, et al. Hypothermic oxygenated machine perfusion prevents arteriolonecrosis of the peribiliary plexus in pig livers donated after circulatory death. PLoS One 2014;9:e88521.
- Liu Q, Nassar A, Buccini L, Iuppa G, Soliman B, Pezzati D, et al. Lipid metabolism and functional assessment of discarded human livers with steatosis undergoing 24 hours of normothermic machine perfusion. Liver Transpl 2018;24:233-45.
- Ben Mosbah I, Roselló-Catafau J, Franco-Gou R, Abdennebi HB, Saidane D, Ramella-Virieux S, et al. Preservation of steatotic livers in IGL-1 solution. Liver Transpl 2006;12:1215-23.
- 81. van Rijn R, Schurink IJ, de Vries Y, van den Berg AP, Cortes Cerisuelo M, Darwish Murad S, et al. Hypothermic machine perfusion in liver transplantation: a randomized trial. N Engl J Med 2021;384:1391-401.

- 82. Dutkowski P, Schlegel A, de Oliveira M, Müllhaupt B, Neff F, Clavien PA. HOPE for human liver grafts obtained from donors after cardiac death. J Hepatol 2014;60:765-72.
- Schlegel A, Dutkowski P. Role of hypothermic machine perfusion in liver transplantation. Transpl Int 2015;28:677-89.
- Fondevila C, Hessheimer AJ, Maathuis MH, Muñoz J, Taurá P, Calatayud D, et al. Superior preservation of DCD livers with continuous normothermic perfusion. Ann Surg 2011;254:1000-7.
- 85. Weeder PD, van Rijn R, Porte RJ. Machine perfusion in liver transplantation as a tool to prevent non-anastomotic biliary strictures: rationale, current evidence and future directions. J Hepatol 2015;63:265-75.
- Bruinsma BG, Yeh H, Ozer S, Martins PN, Farmer A, Wu W, et al. Subnormothermic machine perfusion for ex vivo preservation and recovery of the human liver for transplantation. Am J Transplant 2014;14:1400-9.
- Kollmann D, Linares-Cervantes I, Ganesh S, Rosales R, Hamar M, Goto T, et al. Normothermic ex vivo liver perfusion prevents intrahepatic platelet sequestration after liver transplantation. Transplantation 2020;104:1177-86.
- Mabrut JY, Lesurtel M, Muller X, Dubois R, Ducerf C, Rossignol G, et al. Ex vivo liver splitting and hypothermic oxygenated machine perfusion: technical refinements of a promising preservation strategy in split liver transplantation. Transplantation 2021;105:e89-90.
- Gilbo N, Fieuws S, Meurisse N, Nevens F, van der Merwe S, Laleman W, et al. Donor hepatectomy and implantation time are associated with early complications after liver transplantation: a single-center retrospective study. Transplantation 2021;105:1030-8.
- 90. Gilbo N, Fieuws S, Jochmans I, Monbaliu D. Reply to: "The Impact of donor hepatectomy time on liver transplantation outcomes". Transplantation 2022;106:e175-6.
- 91. Hamaoui K, Gowers S, Sandhu B, Vallant N, Cook T, Boutelle M, et al. Development of pancreatic machine perfusion: translational steps from porcine to human models. J Surg Res 2018;223:263-74.
- 92. Branchereau J, Renaudin K, Kervella D, Bernadet S, Karam G, Blancho G, et al. Hypothermic pulsatile perfusion of human pancreas: preliminary technical feasibility study based on histology. Cryobiology 2018;85:56-62.
- 93. Barlow AD, Hamed MO, Mallon DH, Brais RJ, Gribble

KJT[<]

FM, Scott MA, et al. Use of ex vivo normothermic perfusion for quality assessment of discarded human donor pancreases. Am J Transplant 2015;15:2475-82.

- 94. Fridell JA, Mangus RS, Powelson JA. Histidine-tryptophan-ketoglutarate for pancreas allograft preservation: the Indiana University experience. Am J Transplant 2010;10:1284-9.
- Hamed A, Tsui S, Huber J, Lin R, Poggio EC, Ardehali A. Serum lactate is a highly sensitive and specific predictor of post cardiac transplant outcomes using the organ care system. J Heart Lung Transplant 2009;28:S71.
- 96. Yeter R, Hübler M, Pasic M, Hetzer R, Knosalla C. Organ preservation with the organ care system. Appl Cardiopulm Pathophysiol 2011;15:202-12.
- 97. Tenderich G, Tsui S, El-Banayosy A, Dhital K, Schulte-Eistrup S, Schulz U, et al. The 1-year follow-up results of the PROTECT patient population using the organ care system. J Heart Lung Transplant 2008;27:S166.
- García Sáez D, Zych B, Sabashnikov A, Bowles CT, De Robertis F, Mohite PN, et al. Evaluation of the organ care system in heart transplantation with an adverse donor/recipient profile. Ann Thorac Surg 2014;98:2099-105.
- 99. Schroder JN, D'Alessandro D, Esmailian F, Boeve T, Tang P, Liao K, et al. Successful utilization of extended criteria donor (ECD) hearts for transplantation - results of the OCS[™] heart EXPAND trial to evaluate the effectiveness and safety of the OCS heart system to preserve and assess ECD hearts for transplantation. J Heart Lung Transplant 2019;38:S42.
- 100. Rojas SV, lus F, Schibilsky D, Kaufeld T, Sommer W, Benk C, et al. Cardiac transplantation in higher risk patients: is ex vivo heart perfusion a safe preservation technique? A two center experience. J Heart Lung Transplant 2019;38:S43.
- 101. Messer S, Page A, Berman M, Colah S, Dunning J, Pavlushkov E, et al. First to 50: early outcomes following heart transplantation at Royal Papworth Hospital from donation after circulatory determined death (DCD) donors. J Heart Lung Transplant 2019;38:S43.
- 102. Iyer A, Gao L, Doyle A, Rao P, Cropper JR, Soto C, et al. Normothermic ex vivo perfusion provides superior organ preservation and enables viability assessment of hearts from DCD donors. Am J Transplant 2015;15:371-80.
- 103. Iyer A, Gao L, Doyle A, Rao P, Jayewardene D, Wan B,

et al. Increasing the tolerance of DCD hearts to warm ischemia by pharmacological postconditioning. Am J Transplant 2014;14:1744-52.

- 104. Feizpour CA, Gauntt K, Patel MS, Carrico B, Vagefi PA, Klassen D, et al. The impact of machine perfusion of the heart on warm ischemia time and organ yield in donation after circulatory death. Am J Transplant 2022 Jan 10 [Epub]. https://doi.org/10.1111/ ajt.16952.
- 105. Tenderich G, El-Banayosy A, Rosengard B, Tsui S, Wallwork J, Hetzer R, et al. Prospective multi-center European trial to evaluate the safety and performance of the Organ Care System for heart transplants (PRO-TECT). J Heart Lung Transplant 2007;26:S64.
- 106. McCurry K, Jeevanandam V, Mihaljevic T, Couper G, Elanwar M, Saleh H, et al. Prospective Multi-Center Safety and Effectiveness Evaluation of the Organ Care System Device for Cardiac Use (PROCEED). J Heart Lung Transplant 2008;27:S166.
- 107. Stamp NL, Shah A, Vincent V, Wright B, Wood C, Pavey W, et al. Successful heart transplant after ten hours out-of-body time using the TransMedics Organ Care System. Heart Lung Circ 2015;24:611-3.
- 108. de Perrot M, Liu M, Waddell TK, Keshavjee S. Ischemia-reperfusion-induced lung injury. Am J Respir Crit Care Med 2003;167:490-511.
- 109. Loor G, Warnecke G, Villavicencio MA, Smith MA, Kukreja J, Ardehali A, et al. Portable normothermic ex-vivo lung perfusion, ventilation, and functional assessment with the Organ Care System on donor lung use for transplantation from extended-criteria donors (EXPAND): a single-arm, pivotal trial. Lancet Respir Med 2019;7:975-84.
- 110. Mohite PN, Sabashnikov A, García Sáez D, Pates B, Zeriouh M, De Robertis F, et al. Utilization of the Organ Care System Lung for the assessment of lungs from a donor after cardiac death (DCD) before bilateral transplantation. Perfusion 2015;30:427-30.
- 111. Bozso S, Vasanthan V, Luc JG, Kinaschuk K, Freed D, Nagendran J. Lung transplantation from donors after circulatory death using portable ex vivo lung perfusion. Can Respir J 2015;22:47-51.
- 112. de Vries RJ, Tessier SN, Banik PD, Nagpal S, Cronin SEJ, Ozer S, et al. Supercooling extends preservation time of human livers. Nat Biotechnol 2019;37:1131-6.