

RESEARCH ARTICLE

Increased growth rates of stream salamanders following forest harvesting

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Funding information

Arkansas Science and Technology Authority; Weyerhaeuser Company; University of Arkansas; National Council for Air and Stream Improvement

Abstract

Timber harvesting can influence headwater streams by altering stream productivity, with cascading effects on the food web and predators within, including stream salamanders. Although studies have examined shifts in salamander occupancy or abundance following timber harvest, few examine sublethal effects such as changes in growth and demography. To examine the effect of upland harvesting on growth of the stream-associated Ouachita dusky salamander (*Desmognathus brimleyorum*), we used capture–mark–recapture over three years at three headwater streams embedded in intensely managed pine forests in west-central Arkansas. The pine stands surrounding two of the streams were harvested, with retention of a 14- and 21-m-wide forested stream buffer on each side of the stream, whereas the third stream was an unharvested control. At the two treatment sites, measurements of newly metamorphosed salamanders were on average 4.0 and 5.7 mm larger post-harvest compared with pre-harvest. We next assessed the influence of timber harvest on growth of post-metamorphic salamanders with a hierarchical von Bertalanffy growth model that included an effect of harvest on growth rate. Using measurements from 839 individual *D. brimleyorum* recaptured between 1 and 6 times (total captures, $n = 1229$), we found growth rates to be 40% higher post-harvest. Our study is among the first to examine responses of individual stream salamanders to timber harvesting, and we discuss mechanisms that may be responsible for observed shifts in growth. Our results suggest timber harvest that includes retention of a riparian buffer (i.e., stream-side management zone) may have short-term positive effects on juvenile stream salamander growth, potentially offsetting negative sublethal effects associated with harvest.

KEYWORDS

Arkansas, before–after control–impact, capture–mark–recapture, *Desmognathus brimleyorum*, interior highlands, Ouachita dusky salamander, Ouachita Mountains, streamside management zone

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1 | INTRODUCTION

Headwater streams are strongly influenced by harvesting of timber within the surrounding watershed (Webster et al., 1992). The most evident direct effect of harvesting on forest streams is the removal of shading vegetation, which alters stream microclimates (Olson et al., 2007) and results in both increased average stream temperatures (Reiter et al., 2015) and reduction in allochthonous inputs (i.e., leaf litter; Webster & Waide, 1982). Consequently, following harvest, streams are less light-limited and filamentous green algae often increase in abundance (Lowe et al., 1986), increasing total primary production (Webster et al., 1983). Accompanying this shift in the stream energy base is often a switch in dominant benthic invertebrates (Gurtz & Wallace, 1984; Wallace, 1988; Wallace & Gurtz, 1986) from shredders to scrapers and collectors that feed on algae (Wallace et al., 1997; Webster et al., 1992). Other potential effects of forest harvesting include short-term increases in stream flow with less evapotranspiration and potential alterations to nutrient processing (i.e., nitrogen and phosphorus loss; Webster et al., 1992).

Stream salamanders are a primary vertebrate predator within headwater stream systems where they consume both aquatic and terrestrial invertebrate detritivores (Johnson & Wallace, 2005; Southerland et al., 2004) and can attain extremely high densities and biomass (e.g., 11,294 salamanders/ha; Peterman et al., 2008). As such, they can exert direct and indirect biotic control of prey species and influence ecosystem processes along grazer and detrital pathways (reviewed in Davic & Welsh, 2004). These trophic associations may ultimately influence the breakdown of leaf litter and transfer of nutrients (Davic & Welsh, 2004; Walker et al., 2018; Wyman, 1998) and, importantly, can be influenced by forest harvesting, particularly when allochthonous inputs of leaf litter are reduced (Johnson & Wallace, 2005; Wallace et al., 1997).

Numerous studies across North America report that salamander populations decline for a period of time after timber harvesting (e.g., Ash, 1997; Connette & Semlitsch, 2015; Herbeck & Larsen, 1999; Petranka et al., 1994; Reichenbach & Sattler, 2007). However, recent research has suggested that forestry best management practices (BMPs), specifically implementation of riparian buffers (referred to as streamside management zones/SMZs in some U.S. regions; Lee et al., 2004), may ameliorate negative effects on salamander movement (Johnston & Frid, 2002), abundance (Halloran et al., 2021; Maigret et al., 2014; Perkins & Hunter, 2006; Peterman & Semlitsch, 2009), and species richness and occupancy (Guzy et al., 2019; Kroll et al., 2008). Yet, it is unclear whether harvesting can affect fitness surrogates such as stream salamander growth and reproduction. Little research has examined salamander response to forestry activities at the individual level (i.e., mark-recapture approaches; but see Cecala et al., 2014; Chazal & Niewiarowski, 1998; Connette & Semlitsch, 2015), and to our knowledge, none have examined changes in individual salamander growth rates in response to timber harvesting, either for woodland or for stream-associated salamanders. Although a few studies have explored the influence of forest management on endpoints such as body condition (Hocking et al., 2013; Homyack

et al., 2011; Karraker & Welsh, 2006), examining growth at the individual level is necessary to address alternative explanations for changes in population demography such as size-biased mortality.

Measuring growth as a potential response to harvest is particularly important because body size influences survival and fecundity of salamanders and thus contributes to individual fitness and population growth (Hernández-Pacheco et al., 2021; Tilley, 1968). Energetic requirements of salamanders may vary with differences in the thermal environment of harvested areas (Homyack et al., 2011), influencing metabolic rates, growth, and ultimately body size. Similarly, because stream salamander growth has been correlated with prey biomass (Huntsman et al., 2011; Johnson & Wallace, 2005), changes in the stream invertebrate community caused by harvesting within the watershed may influence salamander growth and body size (e.g., Bumpers et al., 2017).

In this study, we used intensive capture-mark-recapture at three streams to examine the effect of upland forest harvesting on growth of a stream salamander species, the Ouachita dusky salamander (*Desmognathus brimleyorum*). To reduce the influence of stochastic differences among sites and years, we used a before-after control-impact (BACI) design, which allowed for comparisons within the same sites before and after harvest, and comparisons with a designated control site through time. Although our streams contained riparian buffers (SMZs), we predicted that stream salamander growth rate would be faster and body condition would be higher immediately post-harvest, due to short-term increased productivity resulting from canopy reduction or potential nutrient increases.

2 | METHODS

2.1 | Study species

Desmognathus brimleyorum occurs in west-central Arkansas and southwest Oklahoma (Means, 1999) and is one of the least studied *Desmognathus* species (Petranka, 1998). Oviposition typically occurs during summer months (Trauth et al., 1988), and has been reported to peak in late June and early July in the Cossatot Mountains (Means, 1975), but may occur anytime from March to September (Trauth et al., 1990). The larval period has not been well established but is thought to last approximately 10 months to a year (Means, 1974; Trauth et al., 1990). Our observations of 148 individuals indicate that metamorphs range from ~20–30 mm SVL. Within west-central Arkansas, average length for adults has been reported from the Cossatot Mountains as 64 mm SVL (females) and 71 mm SVL (males) (Means, 1999). Here, we considered individuals >45 mm SVL at time of capture as adults, and based on 349 individuals, average adult length for our study area is 56 mm (SD 7.6 mm); we observed a maximum of 80 and 79 mm SVL for females and males, respectively. Based on our mark-recapture dataset, individuals may live to at least seven years. Females have been reported to reproduce at >63 mm SVL (Means, 1974; Trauth et al., 1990), and in our study, gravid females were on average 61 mm ($n = 144$), although we observed

gravid females as small as 49–55 mm SVL ($n = 10$). In a concurrent study, Halloran et al. (2021) reported average net movement (i.e., distance between furthest upstream and furthest downstream capture locations) to be less than 20 m with a slight upstream movement bias, although a few individuals moved up to 164 m within the stream. However, movement was increased following upland clear-cut timber harvest; 35% of 1030 *D. brimleyorum* at the control site had a net movement of 4 m or less, but at treatment sites 19% of the 1423 individuals had net in-stream movements less than 4 m between pre-harvest and post-harvest surveys (Halloran et al., 2021).

2.2 | Study sites

This study was conducted in northeast Howard County, in west-central Arkansas, USA, within 7–23 km of the Cossatot Mountains, in the southernmost subdivision of the Ouachita Mountains (Figure 1), and occurred within even-aged loblolly pine (*Pinus taeda*) forest managed by Weyerhaeuser Company. To assess the influence of timber harvest on the stream-dwelling Ouachita dusky salamander (*Desmognathus brimleyorum*), we selected three 1st-order, intermittent

headwater streams based on similar size, morphology, and silvicultural history. Sites were located within 16 km of each other in the Little Missouri River Watershed, with elevations ranging from 190 to 300 m above sea level. Each stream drained a small watershed (0.41–1.15 km²) within a mature (29–35 years old) loblolly pine stand.

2.3 | Study design

We used a BACI study design to examine the effects of timber harvest on salamander growth. Specifically, we conducted intensive capture–mark–recapture of salamanders at one “control” (unharvested) site and two before–after sites (hereafter “BA1” and “BA2”) that were clear-cut-harvested during the study, with an SMZ retained along each stream. We conducted salamander surveys at the control and BA1 sites from 2014 to 2016, during March, June, and October of each year; during each month, each site was sampled three times (approximately one week apart; Figure 1). The same survey schedule was implemented for BA2; however, surveys began one year later in March 2015. At each site, we established a 200 m stream transect at the most downstream section of each stream. The BA1 and

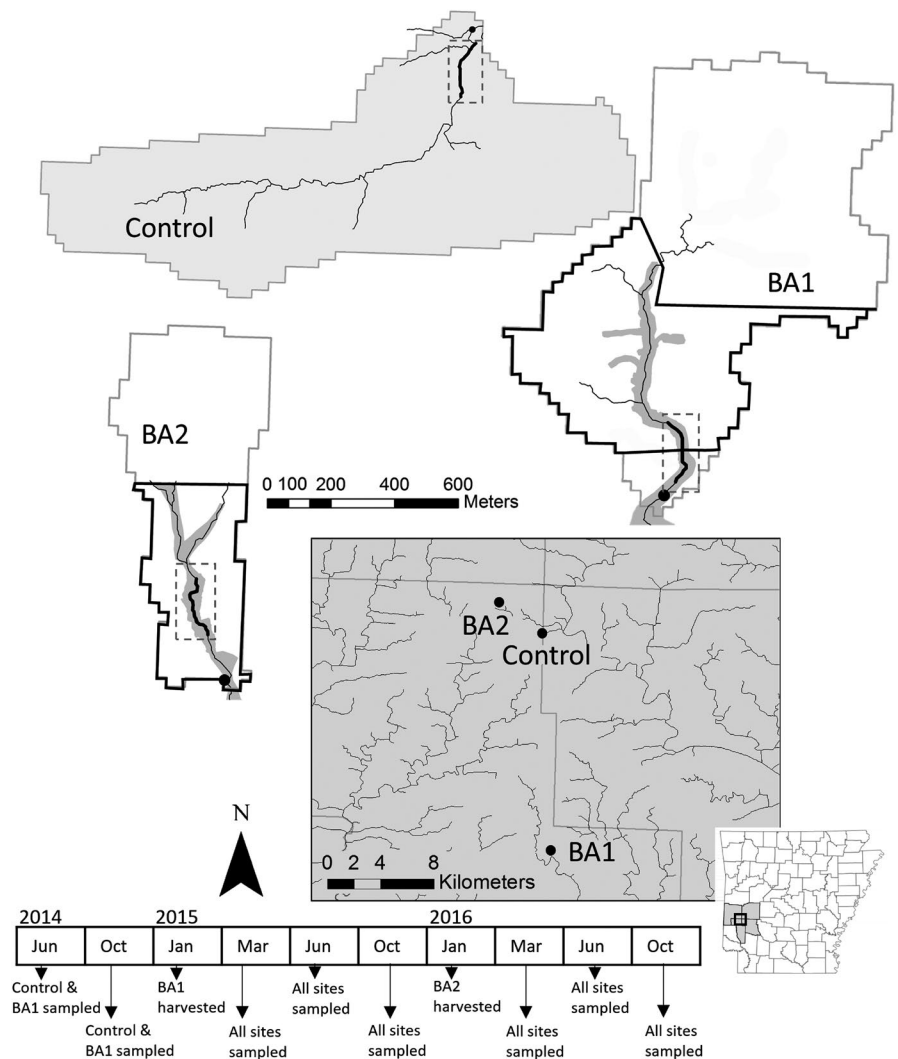


FIGURE 1 Location of study sites in northeast Howard County, Arkansas, USA, and timeline of timber harvest and sampling schedule. Stream watersheds are outlined in gray. Approximate streamside management zone (SMZ) boundary for treatment sites (BA1, BA2) is shaded in gray. Bold black lines enclosed in a dashed box indicate the 200-m sampling transect where salamanders were sampled. At before–after sites, the harvested section of the watershed is outlined in black

BA2 sites were clear-cut-harvested in January 2015 and January 2016, respectively, with a 28-m and a 42-m SMZ retained along the length of the stream (Figure 1). In concordance with state BMPs, some overstory pine trees were harvested from the SMZ of BA1 to promote hardwood regeneration. The SMZ at BA2 along with the riparian forest surrounding the control site was comprised of an oak-hickory (*Quercus* and *Carya* spp.) overstory with a cedar (*Juniperus virginiana*) and holly (*Ilex opaca*) understory. The SMZ of the BA1 site was dominated by loblolly pine in the overstory and holly, muscledwood (*Carpinus caroliniana*), and hophornbeam (*Ostrya virginiana*) in the understory. Based on 16–28 measures of canopy cover taken with a concave spherical densiometer (Lemmon, 1956) in the center of each stream transect before and after harvest after leaf-out (i.e., May), canopy cover decreased by 18% (100 vs. 82%) at BA1 and by 24% (99% vs. 75%) at BA2 following harvesting, but remained constant at 98.9%–100% at the control site.

2.4 | Field methods

Each salamander survey began approximately one hour after sunset and consisted of a thorough visual search (i.e., turning over rocks and debris) of the streambed for the length of each 200-m transect. Post-metamorphic *Desmognathus brimleyorum* were captured using dip-nets and placed in separate containers, and each individual's location was marked with a flag. The following day, we processed captured salamanders in the laboratory by anesthetizing each individual with a solution of 1 g Orajel™-20% benzocaine/1 L of de-chlorinated tap water (Cecala et al., 2007) and recording body metrics using a digital scale and calipers [i.e., mass (g), total length (mm), snout-vent length (SVL; mm)]. All salamanders were anesthetized regardless of recapture status to allow for precise measurements of length and mass. Following measurement, each newly captured individual was given a unique identification mark using a subcutaneous injection of visible implant elastomer (VIE; Northwest Marine Technologies; Grant, 2008). We ventrally marked individuals using a combination of four colors (pink, orange, blue, and yellow) and 6 marking locations (posterior to each limb and anterior to each hind limb) with a 0.5-ml Micro-Fine™ insulin syringe (28-gauge/0.35 mm). To ensure reliable identification, each salamander was marked at a minimum of two positions using at least two colors. Any recently metamorphosed individuals (less than a year since metamorphosis; <45 mm SVL) were labeled as juveniles and were not given marks anterior to each hind limb, as we have observed the cutaneous layers in this region are too thin to reliably hold marks in place. *Desmognathus* salamanders are not consistently sexually dimorphic, and determining sex of most individuals requires sacrificing the animal for internal inspection of the gonads. However, when possible during the breeding season we recorded the sex of adult salamanders based on secondary sexual characters including mental gland and papillose vent in males and plicate vent in females (Noble, 1931); limited sample size precluded incorporation of sex into growth models. Salamanders were returned to their exact capture location within ~2 days after capture,

but occasionally, salamanders were held up to 5 days to avoid releasing during unusually high flow events. For more details on sampling, see Halloran et al. (2021). All research was conducted with approval by the University of Arkansas Institutional Animal Care and Use Committee (AUP 14032).

2.5 | Data analysis

2.5.1 | Body size

We examined variation in body size of recently metamorphosed salamanders (i.e., less than a year since metamorphosis, <45 mm SVL) from June of each year at control and treatment sites with box plots. We then compared body condition of salamanders at control and treatment sites with a one-way ANOVA on ranked residuals (Welsh et al., 2008) where the log of mass (g) depended on the following predictors: log of snout-vent length (mm) and treatment (i.e., log (Mass) ~ log (SVL) + treatment). Salamanders included in the body condition analysis are approximately the same subset of those included in the growth analysis (below) but excluded gravid females. All analyses and figures were constructed in RStudio using R version 3.6.0 (R Core Team, 2021). Model assumption was verified by plotting residuals versus fitted values.

2.5.2 | Growth

We assessed growth of post-metamorphic *D. brimleyorum* with Wang's (1998) parameterization of the Fabens method for estimating von Bertalanffy growth model parameters. The Wang (1998) model is parameterized in terms of the growth increment, Z , as:

$$Z = [l_{\infty} + \beta \{X - E(X)\} - X] (1 - e^{-kT}) + \varepsilon$$

where Z is the change in salamander SVL; $l_{\infty} + \beta\{X - E(X)\}$ is a first-order approximation to the asymptotic SVL (Wang, 1998), where l_{∞} (the population mean asymptotic length) and β are parameters to be estimated, X is the salamander SVL at the beginning of the interval, and $E(X)$ is the sample mean SVL; k is the growth coefficient; T is the interval between recaptures; and ε is a term for model and measurement error. The final dataset excluded captures <21 days apart (within primary sampling intervals), as we assumed growth to be negligible relative to the resolution of our measurements within this time interval. We expanded the Wang (1998) model by including log-normal random intercepts for site and year to account for spatial and temporal differences in k . Because of sparse data in some site-by-year combinations, we truncated the random site and year effects to be ± 2 , corresponding to a sevenfold increase or decrease relative to the mean growth rate for a given site or year. We further expanded the model to estimate the effect of harvest on $\log(k)$ using a binary indicator for harvest (0 = pre-harvest or control, 1 = post-harvest). The submodel for k was therefore:

$$\log(k_i) = \alpha_0 + \alpha \times \text{treatment}_i + \eta_{\text{site } i} + \eta_{\text{year } i}, \text{ where}$$

$$\eta_{\text{site } i} \sim \text{Gaussian}(0, \sigma_{\text{site}}), \text{ and}$$

$$\eta_{\text{year } i} \sim \text{Gaussian}(0, \sigma_{\text{year}})$$

We used vague priors for all model parameters: uniform(min. = 0, max = 0.1) for k , Gaussian(mean = 0, SD = 10) for the log-scale effect of treatment on k , Gaussian(0, 1) for β , uniform(0, 100) for l_{∞} , and exponential($\lambda = 1$) for all standard deviations. For growth intervals that spanned both pre- and post-harvest conditions, we specified the harvest covariate as missing and gave the missing data a Bernoulli(probability = 0.5) prior. Similarly, for intervals that spanned multiple years, we integrated model results over the interval by drawing the year effect from a categorical distribution with equal probability given to each year spanned by the interval between captures. To assess model fit, we used a posterior predictive check by simulating data under the model and calculating a Bayesian p -value using sum of squares for the observed and simulated data (Kéry, 2010).

We implemented the model in a Bayesian framework using the software Just Another Gibbs Sampler version 4.3.0 (JAGS; Plummer, 2015) as called from R version 3.5.1 (R Core Team, 2021) using the package “jagsUI” (Kellner, 2016). We sampled from the posterior distribution using five independent chains of 1,000,000 iterations each after a burn-in period of 200,000 iterations, and thinned chains by a factor of 50 to base inference on 100,000 samples from the posterior distribution. We used the Gelman and Rubin statistic (Gelman & Rubin, 1992) and examination of history plots to assess convergence; we observed no evidence for lack of convergence (all $\hat{R} < 1.03$ and history plots appeared well-mixed with no trends). Unless indicated otherwise, posterior distributions are summarized as median (0.025 quantile – 0.975 quantile).

3 | RESULTS

We measured and marked 1,509 individual *D. brimleyorum* during this study. A subset of 839 individuals (control, $n = 342$; BA1, $n =$

135; BA2, $n = 362$) were recaptured between 1 and 6 times for a total of 1,229 captures (by site: control, $n = 571$; BA1, $n = 178$; BA2, $n = 480$; by year: 2014, $n = 86$; 2015, $n = 533$; 2016, $n = 610$), and these data were used to estimate growth before and after clear-cut timber harvest. Based on raw data for June of each year, shortly after metamorphosis, mean body sizes of recently metamorphosed *D. brimleyorum* were greater in all three post-harvest site-years than in the five pre-harvest/control site-years (Figure 2). On average, salamanders were 5.7 and 4.0 mm larger post-harvest at BA1 and BA2, respectively (Figure 2). Our body condition analysis included 1,103 captures (control = 738, treatment = 368), and body condition was not influenced by treatment (Figure 3; $R^2 = 0.20$, $F_{1,1103} = 2.99$, $p = .084$).

The growth model fit our data well (Bayesian p -value = .49). We found a positive effect of treatment (mean alpha = 0.29, 95% CRI -0.03 to 0.48) on salamander growth rate, k , with k 1.4 (95% credible interval 0.98–1.6) times greater following harvest in treated sites (posterior probability of a positive effect = 0.97; Figure 4). Variation in k was similar among sites ($\sigma_{\text{site}} = 0.35$ [0.12 to 2.58]) and years ($\sigma_{\text{year}} = 0.46$ [0.16 to 3.05]). Model-estimated mean asymptotic length among salamanders in this study was 60.3 (58.8–62.2)-mm SVL (Figure 5). Model-based expected individual asymptotic lengths ranged from 52 to 75 mm, with positive growth increments estimated for salamanders up to 73 mm SVL (x-intercept of Figure 5).

4 | DISCUSSION

We conducted a three-year capture-mark-recapture study examining growth of 839 *D. brimleyorum*, before and after clear-cut timber harvesting. Across both treatment sites and years, growth rates were 40% higher after harvest. Our study is among the first to examine stream salamander responses to timber harvesting at the individual level, an

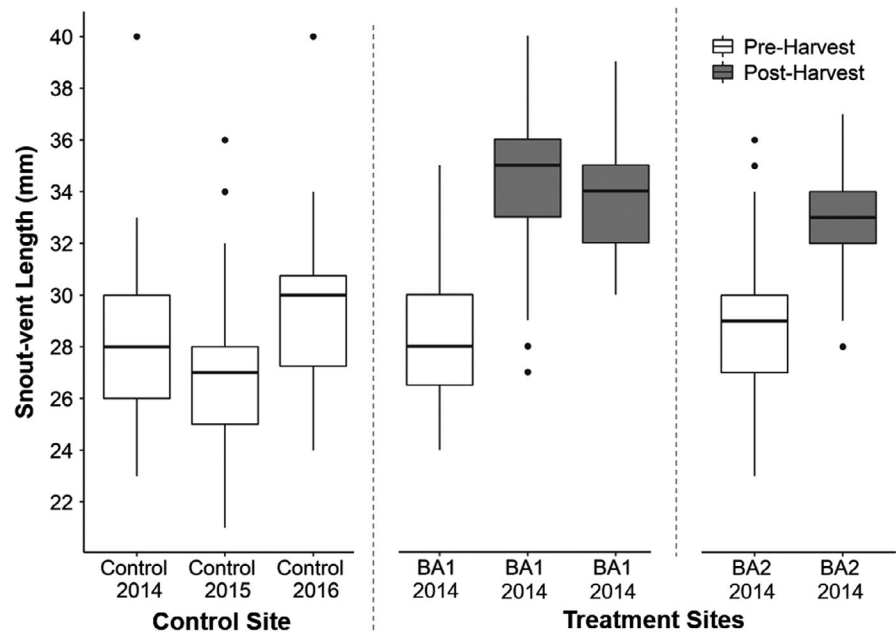


FIGURE 2 Pre- and post-harvest mean body size (snout-vent length) for recently metamorphosed (i.e., less than a year since metamorphosis, ≤ 45 mm SVL) *Desmognathus brimleyorum* captured in June each year at control and treatment sites. In each box plot, the horizontal bar is the median, boxes correspond to the first and third quartiles, and whiskers extend to the highest value within 1.5*interquartile range; data beyond the whiskers are plotted as points

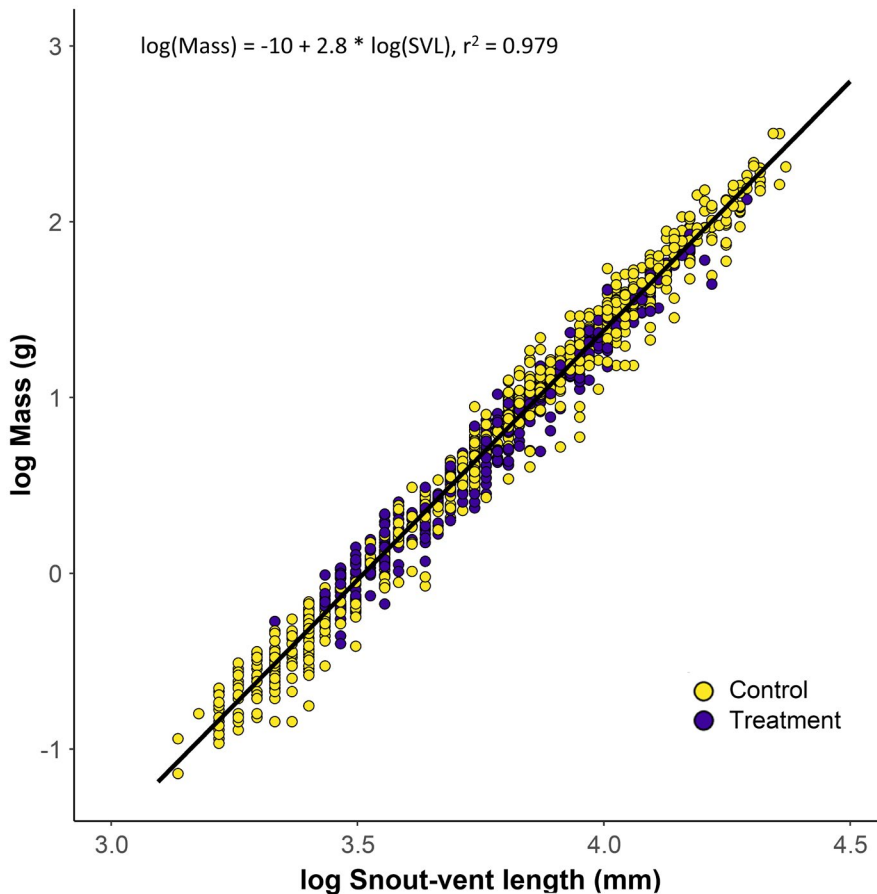


FIGURE 3 Relationship between the log of snout-vent length and log of mass for *Desmognathus brimleyorum* for control (unharvested and pre-harvest; yellow circles) and treatment (post-harvest; indigo circles) sites

approach that is particularly notable as sublethal effects caused by harvesting may influence salamander population dynamics.

Increased growth rates of salamanders post-harvest may influence *D. brimleyorum* populations in several ways. For *Desmognathus* species, body size and egg production are positively correlated, such that larger individuals are more fecund (Tilley, 1968), a common relationship in salamanders (Kaplan & Salthe, 1979; Salthe, 1969). Salamanders with faster growth rates could reach sexual maturity earlier or have higher fecundity at first reproduction, which may have significant effects on individual lifetime fitness and emergent population dynamics (Bernardo, 1993; Homyack & Haas, 2009; Tilley, 1977, 1980). Additionally, *Desmognathus* salamanders are territorial and prone to cannibalism, and salamander assemblages are often structured by competition and intraguild predation (Camp & Lee, 1996; Hairston, 1986). Thus, larger size may result in competitive and survival advantages over conspecifics. Finally, increased growth rates may reduce predation risk, as many important predators of salamanders in headwater streams (e.g., fish, frogs, snakes, other salamanders) are gape-limited.

Several non-mutually exclusive factors may explain increased growth of *D. brimleyorum* following harvest of the surrounding stand. In conjunction with faster growth rates post-harvest (Figure 4), average body size (SVL) of newly metamorphosed salamanders in June was ~4–5 mm longer post-harvest compared with pre-harvest (Figure 2), suggesting variation in growth is initiated during the larval stage. Larval salamander growth rates can be affected by density of conspecifics

and competition for resources (e.g., Morin et al., 1983; Petranka & Sih, 1986; Semlitsch, 1987). However, timber harvest did not have a negative effect on abundance or apparent survival of *D. brimleyorum* at either treatment site during our study (Halloran et al., 2021). Thus, release from intraspecific competition is not likely to be the primary mechanism driving the differences in growth rate we observed.

Alternatively, a likely mechanism driving increased growth rates may be a shift in the quality, quantity, or composition of invertebrate prey available. At our treatment sites, canopy cover decreased ~20% post-harvest. Following overstory harvest, there is typically a temporary increase in light, stimulating primary production in streams (e.g., Webster et al., 1983) even among streams that retain riparian buffers (Kiffney et al., 2004). As a consequence, there is often an increase in productivity of grazer macroinvertebrate assemblages that feed on algal growth (Duncan & Brusven, 1985; Murphy, 1998; Murphy et al., 1981; Price et al., 2003; Silsbee & Larson, 1983; Wallace & Gurtz, 1986). In headwater streams, scraper and collector–gatherer functional feeding groups (i.e., biofilm consumers) are typically higher in nutrient content than are shredder taxa, although this pattern is highly variable (Cross et al., 2003; Frost et al., 2006). Importantly, algae are the primary source of fatty acids in aquatic food webs; thus, increases in biofilm consumers could lead to increased intake of essential fatty acids (Ballantyne et al., 2003; Brett & Muller-Navarra, 1997), which are important for salamander growth (Fitzpatrick, 1976). Additionally, chitin content of macroinvertebrates varies (Cauchie, 2002), and prey may be assimilated

FIGURE 4 Relationship between growth increment and snout-vent length of *Desmognathus brimleyorum* under control (unharvested and pre-harvest; yellow dashed lines and yellow circles) and treatment (post-harvest; solid indigo lines and indigo circles) conditions. Bold lines represent posterior modes, narrow lines represent 95% highest posterior density intervals, the intensity of shading represents the posterior probability density, and points represent observed values. The horizontal gray dotted line along the x-axis at zero represents a daily growth increment of zero

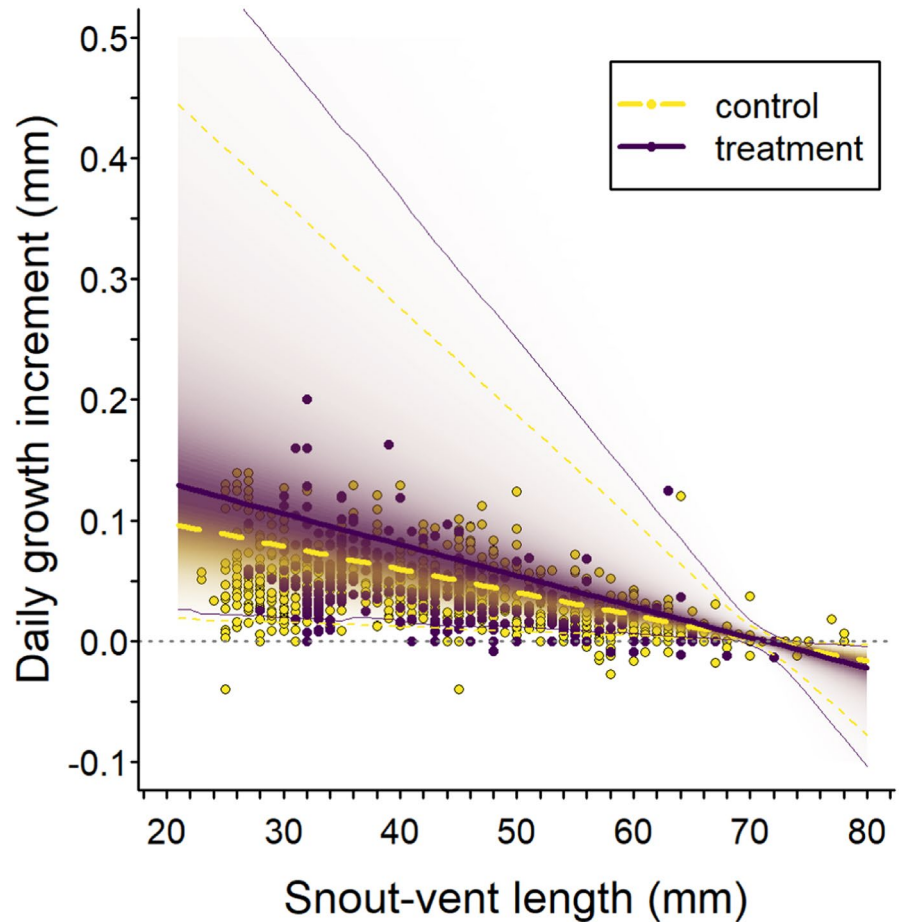
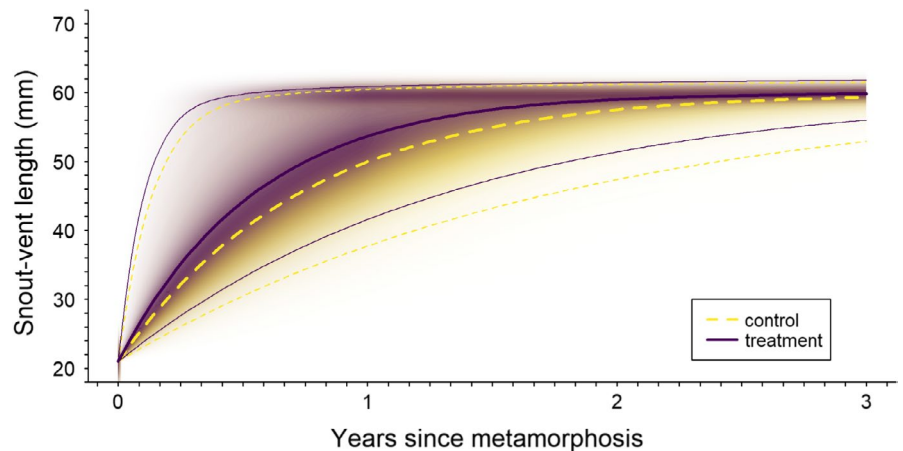


FIGURE 5 Population mean growth curves for *Desmognathus brimleyorum* under control (unharvested and pre-harvest; dashed yellow lines) and treatment (post-harvest; solid indigo lines) conditions. Bold lines represent posterior modes, light lines represent 95% highest posterior density intervals, and the intensity of shading represents the posterior probability density



differently depending on digestibility. Thus, it is possible that post-harvest conditions favor production of invertebrate prey with increased assimilation (i.e., more energy dense and/or easier to digest), contributing to increased salamander growth rates.

Increased invertebrate prey quantity may also influence increased salamander growth rates post-harvest. Studies have shown that macroinvertebrate abundance and density increase immediately post-harvest (Haggerty et al., 2004; Kiffney et al., 2003; Wallace & Ely, 2014), even at sites with riparian buffers (Kiffney et al., 2003). Local resources influence growth in many populations, and greater prey availability increased salamander growth in several studies (Bernardo, 1994; Bernardo & Agosta, 2003; Tilley, 1974). For

example, Bumpers et al. (2017) documented increased *Desmognathus quadramaculatus* growth due to increased abundance of invertebrate prey, because of experimental phosphorus enrichment of headwater streams. Following timber harvesting, the amount of dissolved nutrients leached from soil to the stream may increase until vegetation becomes reestablished, though riparian buffers minimize overland flow of water into streams (Swank, 1988; Swank et al., 2001; Webster et al., 1992). Fertilizer application to newly planted stands may further increase nutrient inputs to streams (Binkley et al., 1999), although riparian buffers can minimize inputs (Kastendick et al., 2012; Secoges et al., 2013). Similarly, disturbance to the upland overstory may add a brief pulse of nutrients in the form of organic matter to

streams after harvest. Thus, it is likely that nutrient enrichment was at least partially responsible for the effects we observed and there is potential for this effect to be magnified under management regimes that include fertilization of newly planted timber stands.

Importantly, although we observed faster growth rates of salamanders post-harvest, possibly attributed to shifts in availability of invertebrate prey, we did not find evidence of a larger body condition (i.e., mass relative to body length) after harvest, and thus, energy stores were similar within two years post-harvest. Conversely, studies of terrestrial salamanders in managed forests of the Pacific Northwest and Eastern United States have found reduced body condition in recently harvested forests (Homyack et al., 2011; Welsh et al., 2008). Small sample sizes of previous studies in conjunction with a focus on terrestrial species (Homyack et al., 2011; Welsh et al., 2008) complicate comparisons across forest management regimes, particularly because harvested streams in our study had riparian buffers.

Because salamanders are ectothermic, a possible mechanism driving increased growth rates post-harvest may be changes to metabolism because of increased air and water temperatures. In laboratory experiments, salamanders grow faster under warmer conditions (Beachy, 1995), and recent work has suggested a link between warming climate and increased salamander body sizes (McCarthy et al., 2017). Numerous studies have established that harvesting of riparian vegetation increases stream temperature (Herunter et al., 2004; Johnson & Jones, 2000; Wilkerson et al., 2006). However, the magnitude of stream temperature response to harvest may vary with inclusion of riparian buffers. Riparian buffers at our treatment sites were 14 and 21 m wide, and studies of streams in British Columbia with similar buffer widths documented 1–4°C increases in stream temperatures following harvest of the surrounding stand (Herunter et al., 2004; Kiffney et al., 2003; Macdonald et al., 2003). At streams with riparian buffers, increased stream temperatures following harvest have been observed to persist for 5 years (Macdonald et al., 2003). It is important to consider the potential biological consequences of even small changes in thermal regime, as temperature influences nearly every aspect of the physiology of ectotherms, including salamanders (Rome et al., 1992). Additionally, warmer temperatures can influence seasonal activity of *Desmognathus* salamanders (Shealy, 1975), resulting in a slightly extended activity season during cooler months, which may increase juvenile salamander growth rates following harvest.

5 | CONCLUSIONS

Our intensive capture–recapture study of >800 individuals contributes to improved understanding of effects of timber harvesting on stream salamanders. Compared with control/pre-harvest sites, we documented higher post-harvest growth rates of *D. brimleyorum* at two treatment sites up to two years post-harvest. Our study did not measure stream and air temperatures or prey availability and consumption before and after harvest; therefore, we are limited in our ability to identify mechanisms responsible for these patterns. However, given that timber harvesting did not affect salamander abundance or apparent

survival (Halloran et al., 2021), a release from intraspecific competition is not a likely mechanism. Alternatively, juvenile salamanders may have different behavioral responses to harvesting or may capitalize on post-harvest resource pulses resulting from a shift in the quality, quantity, or composition of invertebrate prey available. Additionally, changes to metabolism from potentially increased air and water temperatures post-harvest may favor increased growth or result in a slightly extended activity season during cooler months. Addressing these potential mechanisms was beyond the scope of this study, and low replication (i.e., two treatment sites) reduces our ability to determine how robust our findings are across managed forests. However, our results suggest that harvesting may have short-term positive effects on growth as has been seen with fish (e.g., Wilzbach et al., 2005), and thus may reduce predation risk or offset potential negative sublethal effects associated with harvest. However, this conclusion is predicated on the fact that ~20 m riparian buffers (i.e., SMZs) were retained along each side of our streams during harvest. Forestry BMPs for our study region recommend minimum buffers of 11–24 m (Arkansas Forestry Commission, 2002) to protect water quality (Cristan et al., 2016). To develop more focused and efficient management approaches, future studies may seek to determine the mechanistic relationships driving changes in growth rates post-harvest, which could include monitoring shifts in the abiotic environment, invertebrate community, and salamander bioenergetics before and after harvest.

ACKNOWLEDGMENTS

This study was funded by Weyerhaeuser Company®, the National Council for Air and Stream Improvement, the Arkansas Science and Technology Authority, and the University of Arkansas, with additional support from the U.S. Geological Survey Ecosystem Mission Area. We thank Paula Szczerba and Shane Szczerba for logistical support during data collection, and Meredith Swartwout, Chelsea Kross, Jennifer Mortensen, Logan Estes, Mitchell Pruitt, Brittany Furtado, and Philip Vogrinc for assistance with data collection. We thank the Ouachita Mountains Biological Station (OMBS) and Kay Johnson for providing housing and workspace during field seasons. We are grateful to Jennifer Mortensen and Teofil Nakov for assistance with data manipulation, and to Doug Leasure for assistance with geospatial analyses informing site selection. For comments that improved this manuscript, we thank Mark Henderson and two anonymous reviewers. For input of project design and management, we thank Michelle Evans-White, Dan Magoulick, Darren Miller, and T. Bently Wigley. Any use of trade, firm, or names is for descriptive purposes only and does not imply endorsement by the U.S. Government. This is contribution 810 of the U.S. Geological Survey Amphibian Research and Monitoring Initiative.

CONFLICT OF INTEREST

None declared.

AUTHOR CONTRIBUTIONS

Jacquelyn C. Guzy: Conceptualization (equal); data curation (lead); formal analysis (supporting); investigation (lead); methodology (lead); project administration (lead); visualization (equal); writing—original draft (lead); writing—review and editing (lead). **Brian J.**

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DATA AVAILABILITY STATEMENT

Salamander growth data and model code from this study can be accessed from the Dryad Digital Repository at <https://doi.org/10.5061/dryad.2fqz612q3>.

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How to cite this article: Guzy, J. C., Halstead, B. J., Halloran, K. M., Homyack, J. A., & Willson, J. D. (2021). Increased growth rates of stream salamanders following forest harvesting. *Ecology and Evolution*, *11*, 17723–17733. <https://doi.org/10.1002/ece3.8238>