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SPECIALTY SECTION This article was submitted to Medicinal and Pharmaceutical Chemistry, a section of the journal Frontiers in Chemistry

RECEIVED 23 February 2022 ACCEPTED 05 July 2022 PUBLISHED 09 August 2022

#### CITATION

Hasanvand Z, Motahari R, Nadri H, Moghimi S, Foroumadi R, Ayati A, Akbarzadeh T, Bukhari SNA and Foroumadi A (2022), Novel 3aminobenzofuran derivatives as multifunctional agents for the treatment of Alzheimer's disease. *Front. Chem.* 10:882191. doi: 10.3389/fchem.2022.882191

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# Novel 3-aminobenzofuran derivatives as multifunctional agents for the treatment of Alzheimer's disease

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A novel multifunctional series of 3-aminobenzofuran derivatives **5a-p** were designed and synthesized as potent inhibitors of acetylcholinesterase (AChE) and butyrylcholinesterase (BuChE). The target compounds **5a-p** were prepared *via* a three-step reaction, starting from 2-hydroxy benzonitrile. *In vitro* anticholinesterase activity exhibited that most of the compounds had potent acetyl- and butyrylcholinesterase inhibitory activity. In particular, compound **5f** containing 2-fluorobenzyl moiety showed the best inhibitory activity. Furthermore, this compound showed activity on self- and AChE-induced Aβ-aggregation and MTT assay against PC12 cells. The kinetic study revealed that compound **5f** can be considered as a novel multifunctional structural unit against Alzheimer's disease.

#### KEYWORDS

Alzheheimer's disease, acetylcholinesterase, neuroprotection, docking studies, abaggregation, 3-amino benzofuran

### 1 Introduction

Alzheimer's disease (AD) is the most prominent neurodegenerative illness of the Central Nervous System (CNS) progressed by abnormal cholinergic neurons loss, and deposition of extracellular amyloid- $\beta$  and tau-proteins into plaques and neurofibrillary tangles (Scarpini et al., 2003; McKhann et al., 2011; Li et al., 2017). This debilitating disease is mainly characterized by a deterioration of memory, and changes in learning and personality (Lai et al., 2012). Despite the implications of many factors, still there are unclear points in the pathogenesis and etiology of AD (Gauthier et al., 2006; Poprac et al., 2017; Talesa, 2021).

Acetylcholine (ACh), is a cholinergic neurotransmitter and its deficiency is responsible for the cognitive decline in AD patients which has been considered a target to designing novel agent with an AChE-inhibition profile to increase the cerebral acetylcholine level (Lane et al., 2004; Marco-Contelles et al., 2006). Cholinesterase has two types involving acetylcholinesterase (AChE, EC 3.1.1.7) and butyrylcholinesterase (BuChE, EC 3.1.1.8) catalyzing the hydrolysis of ACh to choline and acetate (Mesulam and Asunction, 1987; Weinstock, 1999). Due to the evidence about the activity of BuChE in the pathogenesis of AD, the use of nonselective inhibitors targeting both BuChE and AChE would be more beneficial in AD patients compared to selective ones. Quite a few selective and reversible anti-AChE drugs (donepezil, rivastigmine, and galantamine) have been approved for the treatment of AD. However, these drugs suffer from severe side effects including vomiting, nausea, diarrhea, bradycardia, abnormal dreams, and fatigue (Smith et al., 1997; Forette et al., 1999; Schneider 2000; Cummings et al., 2014). Considering the complex pathophysiology of this disease, there is a high demand for the discovery and development of efficient new therapeutic agents with minimal side effects to effectively combat AD.

The development of multifunctional molecules which are capable of influencing a number of targets has represented considerable attention for the treatment of AD (Cavalli et al., 2008; Camps et al., 2008; León et al., 2013; Rosini et al., 2014). Previous studies revealed that the crystal structure of AChE consists of two binding sites, the peripheral anionic site (PAS) at the entrance and the catalytic active site (CAS) at the bottom of the gorge (Muñoz-Ruiz et al., 2005). It is proposed that the peripheral anionic site of AChE could play an important role in the aggregation of insoluble forms of  $A\beta$  plaques in the brain. So, inhibition of the self-assembled  $A\beta$  peptide formation would be a promising therapeutic strategy for AD treatment (Inestrosa et al., 1996; Castro and Martinez 2001). In addition, considerable evidence implicated the important role of oxidative stress in the development and progression of AD pathogenesis. Therefore, the neuroprotection approach against oxidative stress has beneficial effects on AD (Longo and Massa 2004; Greenough et al., 2013). In order to explain the pathogenesis of AD, the cholinergic hypothesis was first proposed and is relatively accepted, indicating that the decreased levels of ACh are related to the pathogenesis of AD. As the main cholinesterase (ChE), 80% of hydrolytic activity in the brain is performed by acetylcholinesterase (AChE), so inhibiting AChE which will result in increased levels of ACh in the brain has been considered as a good strategy to treat AD. While inhibition of BuChE might be considered a promising strategy for the treatment of AD, the development of AChE inhibitors is still the most important strategy to treat AD.

Benzofuran derivatives have attracted considerable attention in numerous natural products and therapeutic

agents due to their interesting biological and pharmacological properties, including anti-Alzheimer's, analgesic, anti-inflammatory, anti-hyperlipidemic, antiviral, antimicrobial, anti-inflammatory, and antitumor (Alper-Hayta et al., 2008; Choi et al., 2011; Al-Qirim et al., 2012; Rizzo et al., 2013; Nevagi et al., 2015). Synthesis and evaluation of different benzofuran derivatives have been considered by our research group for many years (Pouramiri et al., 2016; Mehrabi et al., 2017). In terms of the AD research, we intend to design and develop multifunctional small molecules, based on this valuable pharmacophore which can be regarded as a mimic of the indanone part of donepezil (Choi et al., 2004). The Aß antiaggregants and site interactions with AB were recognized in the literature. Substitution of the C-2 position of benzofuran was done to increase the inhibitory activity towards AChE (Goyal et al., 2018). Moreover, many reports in the literature with substitution at the C-3 position of benzofuran were done to evaluate their activity as anti-Alzheimer's agents exhibiting increased inhibitory activity towards AB (Howlett et al., 1999; Byun et al., 2008). Donepezil is an FDA-approved drug showing dualbinding site interactions. Docking studies indicated the interactions of N-benzyl piperidine residue with the catalytic active site (CAS) and interactions of hydrophobic aromatic part of the molecule with the peripheral anionic site (PAS) of AChE (Koellner et al., 2000; Saxena et al., 2003; Kapková et al., 2006). Up to now, quite a few numbers of aromatic rings have been reported as PAS binding scaffolds. In the light of the aforementioned and following our previous attempts (Nadri et al., 2010; Baharloo et al., 2015), in this study, we describe the design, synthesis, and biological evaluation of a novel series of 3aminobenzofuran-based derivatives as multifunctional ligands targeting acetylcholinesterase, butyrycholinesterase, and  $\beta$ -Amyloid aggregation (Figure 1).

# 2 Results and discussion

### 2.1 Chemistry

The 3-aminobenzofuran derivatives **5a-p** were prepared according to Scheme 1. The pathway was started from the reaction of commercially available 2-hydroxybenzonitrile **1** with 4-(bromomethyl) pyridine **2** in the presence of  $K_2CO_3$ , afforded 2-(pyridin-4-ylmethoxy)benzonitrile **3**. The cyclization reaction of compound **3** in the presence of *t*-BuOK in DMF at 80°C afforded 4-(3-aminobenzofuran-2-yl) pyridine **4** (Chen et al., 2013; Loidreau et al., 2013). Accordingly, the final compounds **5a-p** were prepared by the reaction of compound **4** with different benzyl chloride derivatives in dry acetonitrile under reflux conditions (1–6 h). After the completion of the

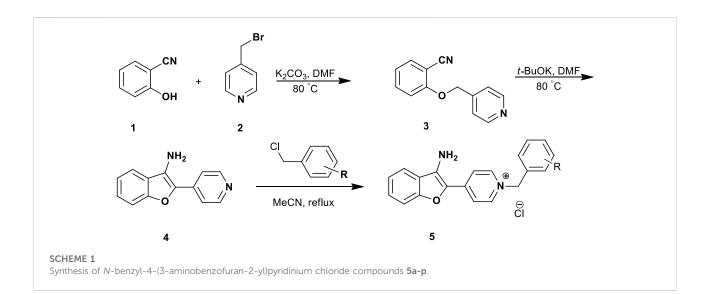
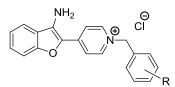


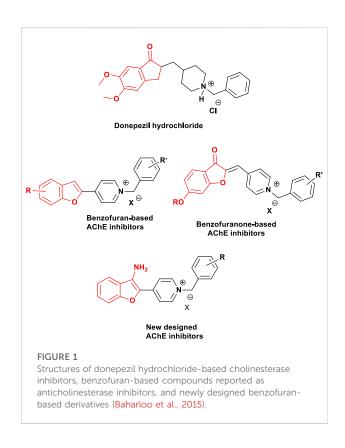
TABLE 1 Inhibitory activity of the target compounds 5a-p against AChE and BuChE.



| Compounds | R                | $IC_{50} \ (\mu M)^a \ AChE$ | IC <sub>50</sub> (µM) BuChE |
|-----------|------------------|------------------------------|-----------------------------|
| 5a        | Н                | 0.81 ± 0.02                  | 0.82 ± 0.07                 |
| 5b        | 2-Me             | $3.29 \pm 0.01$              | 17.35 ± 2.45                |
| 5c        | 3-Me             | $3.32 \pm 0.04$              | 12.89 ± 3.56                |
| 5d        | 4-Me             | 29.31 ± 0.82                 | 25.33 ± 1.91                |
| 5e        | $3\text{-OCH}_3$ | $81.06 \pm 1.61$             | >100                        |
| 5f        | 2-F              | $0.64 \pm 0.04$              | $0.55 \pm 0.07$             |
| 5g        | 3-F              | $1.68 \pm 0.02$              | $2.89 \pm 0.42$             |
| 5h        | 4-F              | $0.88 \pm 0.04$              | $5.05 \pm 0.73$             |
| 5i        | 2-Cl             | $0.70 \pm 0.01$              | $0.99 \pm 0.05$             |
| 5j        | 3-Cl             | $2.99 \pm 0.03$              | 18.77 ± 1.11                |
| 5k        | 4-Cl             | $16.60 \pm 0.52$             | 31.18 ± 2.56                |
| 51        | 2-Br             | $1.34 \pm 0.01$              | $10.54 \pm 0.19$            |
| 5m        | 3-Br             | $4.47 \pm 0.02$              | 13.95 ± 1.20                |
| 5n        | 4-Br             | $15.84 \pm 0.91$             | 22.55 ± 3.33                |
| 50        | $2-NO_2$         | $7.84 \pm 0.05$              | 28.63 ± 0.13                |
| 5p        | $4-NO_2$         | $20.55 \pm 0.8$              | $28.26 \pm 0.48$            |
| Donepezil | _                | $0.016 \pm 0.001$            | 3.99 ± 0.27                 |

 $^{\rm a}{\rm The}$  concentration (mean  $\pm$  SEM of three experiments) required for 50% inhibition. The bold value was the most active compound.

reaction, the final products were collected by filtration and washed with *n*-hexane to afford target compounds in good yields ranging from 56% to 74%.



### 2.2 Biological screenings

#### 2.2.1 Cholinesterase inhibition

The in-vitro cholinesterase inhibitory activity of synthesized 3aminobenzofuran-based compounds **5a-p** was evaluated by using the Ellman's method (Ellman et al., 1961). This method is based on the reaction of acetylthiocholine with 5,5'-dithio-bis-(2-

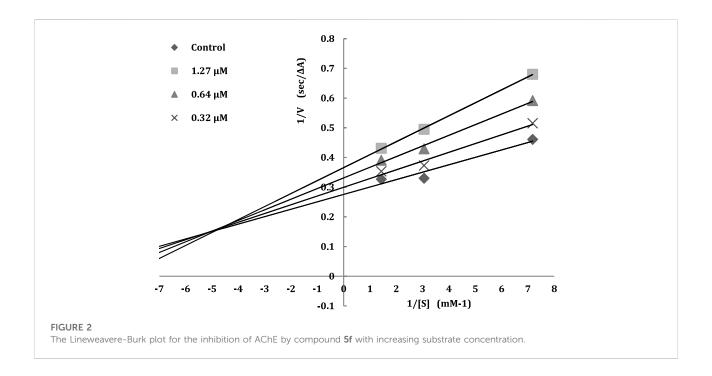


TABLE 2 Inhibition of self-induced and AChE-induced  $A\beta_{1-42}$  aggregation by selected compounds.

| Compounds | Inhibition of A $\beta$ aggregation (%) |                           |  |
|-----------|---|---------------------------|--|
|           | Self-induced <sup>a</sup>               | AChE-induced <sup>b</sup> |  |
| 5a        | 17.6 ± 3.8                              | 22.4 ± 1.6                |  |
| 5f        | 29.8 ± 1.8                              | $30.1 \pm 1.9$            |  |
| 5h        | 38.8 ± 2.8                              | 35.6 ± 1.5                |  |
| 5i        | 24.8 ± 1.5                              | $26.5 \pm 3.8$            |  |
| 51        | 25.7 ± 2.7                              | $22.4 \pm 2.3$            |  |
| Donepezil | 14.9 ± 2.5                              | 25.7 ± 1.9                |  |

<sup>a</sup>Inhibition of A $\beta_{1-42}$  aggregation was produced by the tested compound at 10  $\mu$ M concentration. Values are expressed as means  $\pm$  SEM of three experiments. <sup>b</sup>Co-aggregation inhibition of A $\beta_{1-42}$  and AChE (0.01 u/ml) by the tested compounds at 100  $\mu$ M concentration was detected by the ThT assay. Values are expressed as means  $\pm$  SEM of three experiments.

nitrobenzoic) acid (DTNB) to yield the colored product. The obtained IC<sub>50</sub> values are summarized in Table 1 and compared with donepezil as the standard drug. Accordingly, almost all 3-aminobenzofuran derivatives exhibited moderate to good inhibitory activity with IC<sub>50</sub>s ranging from 0.64 to 81.06  $\mu$ M. Among the tested compounds, analog **5f** was the most effective inhibitor against AChE and BuChE, while compound **5e** containing 3-methoxybenzyl moiety showed the least inhibitory activity on both. The nonpolar group (OCH<sub>3</sub>) has decreased the AChE inhibitory potency due to increased lipophilic characteristics.

Based on the structure-activity relationship (SAR) study, the potency of compounds was influenced by the type and the

TABLE 3 Effects of compounds 5a, 5f, 5h, 5i, and 5l on cell viability in PC12 cells.

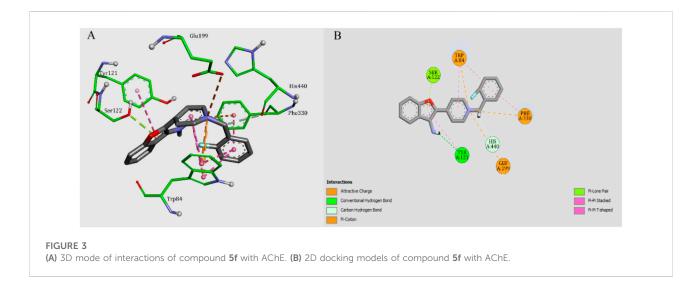
| Compounds | Viability<br>(%) of PC12 cells <sup>a,b</sup> |
|-----------|---|
| 5a        | 85.9 ± 1.5                                    |
| 5f        | 86.4 ± 2.6                                    |
| 5h        | $88.2 \pm 2.4$                                |
| 5i        | $90.2 \pm 4.1$                                |
| 51        | 87.1 ± 2.9                                    |
|           |   |

<sup>a</sup>Cell viability is expressed as the mean percentage of viable cells.

 $^{\rm b}$  Mean percentage of viable cells compared with the untreated cells was determined to be 56.2  $\pm$  1.96%.

position of the substitutions. The unsubstituted derivative **5a** showed potent activity against the AChE enzyme with  $IC_{50} = 0.81 \,\mu$ M, however, the attachment of electron-donating groups such as methyl and methoxy significantly reduced the inhibitory activities of compounds. Those compounds containing electron-withdrawing groups (fluoro, chloro, and bromo) showed better inhibitory potencies than the compounds containing electron-donating groups **5b-5e**.

The SAR analysis indicated that fluorobenzyl-containing compounds **5f-5h** had promising enzymatic inhibitory activity (IC<sub>50</sub>s ranging from 0.64 to 1.68  $\mu$ M) and the presence of fluoro substituent at *ortho* and *para* positions of the benzyl moiety led to better anti-AChE activity than *meta* substituted one **5g**. In the chloro and bromo substituted derivatives, compounds containing *ortho* and *metha* substitutions (**5i, 5j, 5l,** and **5m**) showed moderate inhibitory



activities, however, the attachment of these substitutions to *para* position dramatically decreased inhibitory activity of compounds (**5k** and **5n**). Among the electron-withdrawing-containing compounds, nitro-containing derivatives showed weak inhibitory potencies, and the compound with 2-nitrobenzyl group **50** showed better activity than that of 4-nitrobenzyl derivative **5p**.

The obtained results showed that compounds bearing fluorobenzyl moiety were more effective than other benzyl derivatives. In addition, *ortho* position was more favorable than *meta-* or *para-* substituted analogs. All compounds except **5e** showed BuChE inhibition. By comparing the IC<sub>50</sub> values against AChE and BuChE, it could be concluded that nearly the same trend was observed. The potent inhibitory activity toward BuChE was observed in **5f**. This compound showed good inhibition with an IC<sub>50</sub> value of 0.55  $\mu$ M. The movement of substituents involving fluorine, chlorine, bromine, and methyl from *ortho* to *para* resulted in decreased inhibitory activity.

#### 2.2.2 Kinetics studies

The inhibition model of acetylcholinesterase can be categorized into irreversible-type inhibition and reversible-type inhibition. Therefore, to achieve further insight into the mode of interaction of designed compounds on AChE, the rate of enzyme activity was measured at different concentrations of the most potent compound **5f** 0.32, 0.64, 1.27  $\mu$ M and in the presence of substrate (ATCh). The type of inhibition can be concluded with a graphical analysis of the Lineweaver-Burk reciprocal plots (Baharloo et al., 2015). The higher concentration of inhibitor showed increasing slopes and intercepts. This proves a mixed-type inhibitory pattern on AChE (Figure 2).

# 2.2.3 Inhibition of $\beta\text{-amyloid}$ aggregation of selected compounds

The aggregation of the  $\beta$ -amyloid peptide is a negative event in the pathogenesis of Alzheimer's disease, so, anti-amyloid therapy is

an important factor in the treatment of Alzheimer's. To evaluate the anti-aggregating activity of designed compounds, the five most potent derivatives, namely 5a, 5f, 5h, 5i, and 5l were selected and evaluated against self-induced and AChE-induced AB aggregation. For this purpose, we used the thioflavin T (ThT) assay. Donepezil was used as a reference compound. The obtained results showed that the tested 3-aminobenzofuran compounds (5a, 5f, 5h, 5i, and 5l) at 10  $\mu M$  concentration exhibited good inhibitory activity of self-induced  $A\beta_{1-42}$ aggregation (17.6%, 29.8%, 38.8%, 24.8%, and 25.7% inhibition, respectively) compared to donepezil as the reference drug (14.9%). Compounds 5f and 5h were about 2-folds more effective than donepezil in the inhibition of A $\beta$  aggregation (Table 2). At 100  $\mu$ M concentration, compounds 5a, 5f, 5h, 5i, and 5l showed moderate to good activity to inhibit AChE-induced AB aggregation. Notably, compounds 5f and 5h were more potent (30.1% and 35.6, respectively) than the reference drug (25.7%). The ThT assay indicated that target compounds could inhibit the selfinduced  $A\beta_{1-42}$  aggregation and AChE-induced A $\beta$  aggregation.

# 2.2.4 Neuronal cell treatment and viability measurements

Safety is an important factor for CNS agents. In order to evaluate the cytotoxic effect of selected compounds **5a**, **5f**, **5h**, **5i**, and **5l**, the in-vitro cell viability assay was investigated on the PC12 cell line using the MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay. The PC12 cells were incubated with varying concentrations (0.01–100  $\mu$ M) of the test compounds for 24 h. The obtained results revealed that all tested compounds were nontoxic to PC12 cells (Table 3). The results are expressed as the percentage of viable cells.

### 2.2.5 Molecular docking studies

To get a better insight into the binding patterns between AChE and our new 3-aminobenzofuran derivatives, docking simulation

were performed for the most active compound **5f** in the active site of AChE (PDB code: 1EVE) by using AutoDock software. The two- and three-dimensional modes of interactions were prepared and illustrated in Figure 3. Compound **5f** was anchored in the mid-gorge by some attractive charge among the Trp84, Phe330, and Glu199. Moreover, the pyridine ring was stacked against Trp84. The pendant 2-fluouropheny ring was sandwiched in between Trp84 and Phe330. The amino benzofuran ring made a T-shape stacking with Tyr121. A hydrogen bond between amino function and Tyr121 could stabilize such interaction.

### **3** Experimental

### 3.1 Chemistry

All reagents and solvents were obtained from commercial suppliers and used without purification. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were acquired on a Bruker 500, 300 MHz NMR spectrometer and referenced to TMS. IR (KBr-disc) spectra were obtained by using a Bruker spectrometer. Melting points were determined by using a Kofler hot stage apparatus. All Reactions were monitored with thin layer TLC, using silica gel (60 Å) purchased from Merck. Mass analysis was carried out using an Agilent mass spectrometer. The elemental analysis for C, H, and N was carried out with an ElementarAnalysen system GmbH VarioEL.

### 3.1.1 General procedure for the preparation of 2-((pyridine-4-yl) methoxy) benzonitrile 3

4-(Bromomethyl) pyridine 2 (10 mmol) was added to a mixture of 2-hydroxybenzonitrile 1 (10 mmol) and  $K_2CO_3$  (20 mmol) in DMF (10 ml) and stirred at 80°C for 8 h. After completion of the reaction, the reaction mixture was slowly cooled down to room temperature and poured into ice and water. The precipitated solid was filtered, washed, and recrystallized from EtOH to afford 2-((pyridine-4-yl) methoxy) benzonitrile intermediate 3.

White solid, yield: (83%). IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3388, 2220, 1591, 1484, 1444, 1384, 1285, 1156, 876, 754. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 5.22 (s, 2H, CH<sub>2</sub>), 6.95 (d, J = 8.5 Hz, 2H, Ar-H), 7.06 (t, J = 7.5 Hz, 2H, Ar-H), 7.40 (d, J = 6.0 Hz, 2H, Ar-H), 7.50–7.54 (m, 1H, Ar-H), 7.62 (dd, J = 7.5, 1.5 Hz, 1H, Ar-H).

### 3.1.2 General procedure for the preparation of 2-(pyridine-4-yl) 3-aminobenzofuran derivative 4

A mixture of intermediate **3** (3 mmol) and potassium *tert*butoxide (5 mmol) in DMF (5 ml) was stirred at 80°C for 5 h. After the completion of the reaction, the reaction mixture was cooled down to room temperature and was poured into ice and water. The precipitated solid was filtered, washed, and recrystallized from EtOH to afford 2-(pyridine-4-yl) 3-aminobenzofuran **4**.

Yellow solid, yield: (73%). IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3360, 3311, 3191, 1596, 1409, 1355, 1274, 1212, 1166, 1143, 993, 822, 734,

682, 600, 527. <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 5.83 (s, 2H, CH<sub>2</sub>), 7.24 (t, *J* = 10.0 Hz, 1H, Ar-H), 7.38 (t, *J* = 10.0 Hz, 1H, Ar-H), 7.48 (d, *J* = 10.0 Hz, 1H, Ar-H), 7.65 (d, *J* = 5.0 Hz, 2H, Ar-H), 7.91 (d, *J* = 10.0 Hz, 1H, Ar-H), 8.53 (d, *J* = 5.0 Hz, 2H, Ar-H).

# 3.1.3 General procedure for the preparation of *N*-benzyl-4-(3-aminobenzofuran-2-yl) pyridinium bromides (**5a-p**)

A mixture of 2-(pyridine-4-yl)-3-aminobenzofuran **4** (1 mmol) and the appropriate benzyl chloride derivatives (1.2 mmol) in dry acetonitrile (7 ml) was heated under reflux for 1–6 h. The precipitated solid was collected by filtration and washed with *n*-hexane (5 ml) to give **5a-p** in 56%–74% yields.

#### 3.1.3.1 4-(3-Aminobenzofuran-2-yl)-1-benzylpyridin-1ium chloride (**5a**)

White solid, yield: (72%); mp 160–162°C. IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3293, 3158, 3033, 1631, 1592, 1542, 1312, 1215, 1160, 1100, 1030, 875, 834, 753, 690. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 5.66 (s, 2H, CH<sub>2</sub>), 7.29 (t, *J* = 6.5 Hz, 1H, Ar-H), 7.43–7.46 (m, 5H, Ar-H), 7.51 (s, 4H, Ar-H), 7.97 (s, 2H, NH<sub>2</sub>), 8.10 (d, *J* = 7.5 Hz, 1H, Ar-H), 8.79 (d, *J* = 6.0 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 60.7, 111.5, 116.7, 122.9, 128.2, 128.3, 128.6, 129.0, 130.4, 135.1, 140.1, 140.3, 142.1, 142.6, 142.7, 154.6. Anal. Calcd. for C<sub>20</sub>H<sub>17</sub>ClN<sub>2</sub>O, C, 71.32; H, 5.09; N, 8.32; Found C, 71.60; H, 4.79; N, 8.10.

# 3.1.3.2 4-(3-Aminobenzofuran-2-yl)-1-(2-methylbenzyl) pyridin-1-ium chloride (**5b**)

White solid, yield: (67%); mp 182–184°C. IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3326, 3145, 1722, 1633, 1592, 1541, 1452, 1315, 1197, 1149, 1102, 1031, 968, 837, 746, 655. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 2.31 (s, 3H, CH<sub>3</sub>), 5.70 (s, 2H, CH<sub>2</sub>), 7.11 (d, *J* = 6.5 Hz, 1H, Ar-H), 7.25–7.30 (m, 4H, Ar-H), 7.43 (s, 2H, Ar-H), 7.54 (d, *J* = 12.0 Hz, 2H, Ar-H), 8.00 (s, 2H, NH<sub>2</sub>), 8.11 (d, *J* = 7.0 Hz, 1H, Ar-H), 8.62 (d, *J* = 4.0 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 18.7, 61.3, 111.7, 116.6, 116.8, 121.9, 122.1, 122.7, 126.5, 128.2, 128.8, 130.7, 133.2, 136.5, 140.2, 140.4, 142.2, 142.9, 154.6. Anal. Calcd. for C<sub>21</sub>H<sub>19</sub>ClN<sub>2</sub>O, C, 71.89; H, 5.46; N, 7.98; Found C, 72.06; H, 5.22; N, 8.15.

# 3.1.3.3 4-(3-Aminobenzofuran-2-yl)-1-(3-methylbenzyl) pyridin-1-ium chloride (**5c**)

White solid, yield: (56%); mp 190–192°C. IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3122, 1974, 1639, 1542, 1226, 1162, 1048, 844, 762, 653. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 2.41 (s, 3H, CH<sub>3</sub>), 5.59 (s, 2H, CH<sub>2</sub>), 7.40 (d, *J* = 7.0 Hz, 2H, Ar-H), 7.47 (d, *J* = 7.0 Hz, 2H, Ar-H), 7.50 (d, *J* = 10.5 Hz, 2H, Ar-H), 7.70 (t, *J* = 8.0 Hz, 1H, Ar-H), 7.74 (d, *J* = 8.0 Hz, 1H, Ar-H), 7.80 (d, *J* = 4.5 Hz, 1H, Ar-H), 8.22 (s, 2H, NH<sub>2</sub>), 8.35 (d, *J* = 8.0 Hz, 1H, Ar-H), 8.96 (d, *J* = 7.0 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 20.9, 60.7, 116.6, 116.8, 122.1, 122.7, 125.4, 128.1, 128.7, 128.8, 129.5, 130.4, 135.1, 138.4, 140.3, 140.4, 141.7, 142.1, 154.6. Anal. Calcd. for C<sub>21</sub>H<sub>19</sub>ClN<sub>2</sub>O, C, 71.89; H, 5.46; N, 7.98; Found C, 72.02; H, 5.83; N, 7.77.

# 3.1.3.4 4-(3-Aminobenzofuran-2-yl)-1-(4-methylbenzyl) pyridin-1-ium chloride (**5d**)

White solid, yield: (57%); mp 220–222°C. IR (KBr) ( $\nu_{max}/$  cm<sup>-1</sup>): 3313, 3136, 3038, 1643, 1537, 1350, 1154, 1028, 829, 752, 666. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta = 2.29$  (s, 3H, CH<sub>3</sub>), 5.58 (s, 2H, CH<sub>2</sub>), 7.23 (d, J = 7.5 Hz, 2H, Ar-H), 7.28 (t, J = 7.0 Hz, 2H, Ar-H), 7.41 (d, J = 7.5 Hz, 2H, Ar-H), 7.50 (t, J = 8.0 Hz, 1H, Ar-H), 7.54 (d, J = 8.0 Hz, 1H, Ar-H), 7.59 (s, 1H, Ar-H), 8.01 (s, 2H, NH<sub>2</sub>), 8.15 (d, J = 8.0 Hz, 1H, Ar-H), 8.76 (d, J = 6.5 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta = 20.6$ , 60.6, 116.6, 122.3, 122.4, 122.7, 128.1, 128.3, 129.5, 130.4, 132.2, 138.4, 140.2, 142.0, 142.4, 142.5, 154.5. Anal. Calcd. for C<sub>21</sub>H<sub>19</sub>ClN<sub>2</sub>O, C, 71.89; H, 5.46; N, 7.98; Found C, 71.57; H, 5.80; N, 7.61.

### 3.1.3.5 4-(3-Aminobenzofuran-2-yl)-1-(3methoxybenzyl) pyridin-1-ium chloride (**5e**)

White solid, yield: (61%); mp 210–212°C. IR (KBr) ( $\nu_{max}$ / cm<sup>-1</sup>): 3134, 1638, 1594, 1541, 1310, 1265, 1223, 1160, 1103, 1042, 840, 754, 694. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 3.74 (s, 3H, OCH<sub>3</sub>), 5.61 (s, 2H, CH<sub>2</sub>), 6.93 (d, J = 8.5 Hz, 1H, Ar-H), 7.08 (d, J = 7.0 Hz, 1H, Ar-H), 7.18 (s, 1H, Ar-H), 7.24 (t, J = 7.5 Hz, 1H, Ar-H), 7.32 (t, J = 7.5 Hz, 1H, Ar-H), 7.45 (d, J = 8.0 Hz, 1H, Ar-H), 7.49 (d, J = 8.0 Hz, 1H, Ar-H), 7.69 (s, 2H, Ar-H), 8.00 (s, 2H, NH<sub>2</sub>), 8.19 (d, J = 7.5 Hz, 1H, Ar-H), 8.82 (d, J = 6.5 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 55.2, 60.5, 111.5, 114.2, 116.6, 120.4, 122.3, 122.4, 122.7, 128.1, 130.3, 136.6, 140.3, 140.5, 141.9, 142.5, 142.6, 154.6, 159.5. Anal. Calcd. for C<sub>21</sub>H<sub>19</sub>ClN<sub>2</sub>O<sub>2</sub>, C, 68.76; H, 5.22; N, 7.64; Found C, 69.05; H, 5.51; N, 7.30.

# 3.1.3.6 4-(3-Aminobenzofuran-2-yl)-1-(2-fluorobenzyl) pyridin-1-ium chloride (**5f**)

White solid, yield: (73%); mp 253–255°C. IR (KBr) ( $\nu_{max}$ / cm<sup>-1</sup>): 3175, 2982, 1635, 1541, 1314, 1225, 1159, 1107, 1040, 833, 757, 648. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 5.74 (s, 2H, CH<sub>2</sub>), 7.26–7.32 (m, 3H, Ar-H), 7.46–7.55 (m, 3H, Ar-H), 7.58 (d, J = 7.5 Hz, 1H, Ar-H), 7.69 (s, 2H, Ar-H), 8.02 (s, 2H, NH<sub>2</sub>), 8.17 (d, J = 8.0 Hz, 1H, Ar-H), 8.68 (d, J = 6.5 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 55.3, 115.8, 116.6, 122.1, 122.5, 125.1, 128.1, 130.5, 130.9, 131.5, 140.7, 141.8, 142.1, 142.7, 143.6, 154.7, 159.3, 161.3 (d, J = 240 Hz). Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>ClFN<sub>2</sub>O, C, 67.70; H, 4.55; N, 7.90; Found C, 67.44; H, 4.24; N, 7.55.

# 3.1.3.7 4-(3-Aminobenzofuran-2-yl)-1-(3-fluorobenzyl) pyridin-1-ium chloride (**5g**)

White solid, yield: (65%); mp 267–269°C. IR (KBr) ( $\nu_{max}/$  cm<sup>-1</sup>): 3298, 3167, 2981, 1630, 1543, 1455, 1313, 1253, 1158, 1098, 1036, 879, 835, 750, 692. <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 5.66 (s, 2H, CH<sub>2</sub>), 7.24 (t, *J* = 8.0 Hz, 1H, Ar-H), 7.29 (t, *J* = 7.0 Hz, 1H, Ar-H), 7.36 (d, *J* = 7.0 Hz, 1H, Ar-H), 7.43–7.47 (m, 3H, Ar-H), 7.48–7.54 (m, 3H, Ar-H), 7.97 (s, 2H, NH<sub>2</sub>), 8.10 (d, *J* = 8.0 Hz, 1H, Ar-H), 8.80 (d, *J* = 6.5 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 60.0, 115.3, 115.5, 115.7, 116.7, 122.0, 122.6, 124.4, 128.2, 130.5, 131.2, 137.3, 140.3, 140.4, 142.2, 142.6,

142.7, 159.6 (d, *J* = 237 Hz). Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>ClFN<sub>2</sub>O, C, 67.70; H, 4.55; N, 7.90; Found C, 67.48; H, 4.24; N, 7.66.

# 3.1.3.8 4-(3-Aminobenzofuran-2-yl)-1-(4-fluorobenzyl) pyridin-1-ium chloride (**5h**)

White solid, yield: (71%); mp 258–260°C. IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3296, 3159, 2977, 1631, 1543, 1313, 1226, 1099, 1032, 876, 826, 753, 658. <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 5.63 (s, 2H, CH<sub>2</sub>), 7.29 (t, *J* = 9.0 Hz, 3H, Ar-H), 7.40 (d, *J* = 6.0 Hz, 2H, Ar-H), 7.50 (d, *J* = 8.0 Hz, 1H, Ar-H), 7.54 (d, *J* = 7.5 Hz, 1H, Ar-H), 7.62 (t, *J* = 6.5 Hz, 2H, Ar-H), 7.96 (s, 2H, NH<sub>2</sub>), 8.09 (d, *J* = 7.5 Hz, 1H, Ar-H), 8.79 (d, *J* = 7.0 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 59.9, 115.8, 116.0, 116.8, 122.6, 128.1, 130.4, 130.8, 131.3, 140.2, 140.3, 142.1, 142.5, 142.6, 154.6, 161.3 (d, *J* = 237 Hz). Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>ClFN<sub>2</sub>O, C, 67.70; H, 4.55; N, 7.90; Found C, 67.45; H, 4.89; N, 8.12.

# 3.1.3.9 4-(3-Aminobenzofuran-2-yl)-1-(2-chlorobenzyl) pyridin-1-ium chloride (**5i**)

White solid, yield: (64%); mp 135–137°C. IR (KBr) ( $\nu_{max}/$  cm<sup>-1</sup>): 3347, 3290, 3171, 3081, 1637, 1592, 1545, 1467, 1314, 1216, 1158, 1101, 1042, 871, 827, 751, 671. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 5.76 (s, 2H, CH<sub>2</sub>), 7.30 (t, J = 7.0 Hz, 1H, Ar-H), 7.41 (d, J = 7.0 Hz, 2H, Ar-H), 7.45 (d, J = 8.0 Hz, 2H, Ar-H), 7.52 (t, J = 8.0 Hz, 2H, Ar-H), 7.57 (t, J = 8.0 Hz, 2H, Ar-H), 7.99 (s, 2H, NH<sub>2</sub>), 8.11 (d, J = 7.5 Hz, 1H, Ar-H), 8.65 (d, J = 6.5 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 58.7, 115.7, 116.4, 122.2, 122.6, 128.0, 128.2, 130.0, 130.6, 130.7, 132.4, 132.8, 140.6, 140.7, 142.2, 142.9, 143.0, 154.7. Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>Cl<sub>2</sub>N<sub>2</sub>O, C, 64.70; H, 4.34; N, 7.55; Found C, 64.56; H, 4.02; N, 7.34.

### 3.1.3.10 4-(3-Aminobenzofuran-2-yl)-1-(3chlorobenzyl) pyridin-1-ium chloride (**5**j)

White solid, yield: (59%); mp 180–182°C. IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3291, 3139, 1635, 1540, 1313, 1208, 1160, 1097, 1032, 875, 833, 759, 688. <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 5.64 (s, 2H, CH<sub>2</sub>), 7.29 (t, *J* = 7.5 Hz, 1H, Ar-H), 7.43 (d, *J* = 5.5 Hz, 2H, Ar-H), 7.45–7.54 (m, 5H, Ar-H), 7.68 (s, 1H, Ar-H), 7.97 (d, *J* = 5.5 Hz, 2H, Ar-H), 8.09 (d, *J* = 7.5 Hz, 1H, Ar-H), 8.80 (d, *J* = 6.5 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 59.9, 116.8, 121.9, 122.1, 122.6, 127.1, 128.2, 128.8, 130.5, 130.9, 133.5, 137.4, 140.3, 140.4, 142.2, 142.6, 142.7, 154.6. Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>Cl<sub>2</sub>N<sub>2</sub>O, C, 64.70; H, 4.34; N, 7.55; Found C, 65.00; H, 4.63; N, 7.24.

#### 3.1.3.11 4-(3-Aminobenzofuran-2-yl)-1-(4chlorobenzyl) pyridin-1-ium chloride (**5k**)

Yellow solid, yield: (69%); mp 192–194°C. IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3145, 1640, 1593, 1541, 1317, 1217, 1164, 1098, 1027, 825, 761. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta = 5.64$  (s, 2H, CH<sub>2</sub>), 7.28 (t, J = 7.5 Hz, 1H, Ar-H), 7.50 (d, J = 7.6 Hz, 2H, Ar-H), 7.53 (d, J = 7.6 Hz, 2H, Ar-H), 7.56–7.60 (m, 3H, Ar-H), 7.62 (d, J = 4.0 Hz, 1H, Ar-H), 8.02 (s, 2H, NH<sub>2</sub>), 8.15 (d, J = 8.0 Hz, 1H, Ar-H), 8.79 (d, J = 6.5 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta = 59.8$ , 116.8, 122.1,

122.3, 122.5, 128.1, 129.0, 130.3, 133.7, 134.1, 140.5, 140.6, 142.1, 142.6, 142.7, 154.7. Anal. Calcd. for  $C_{20}H_{16}Cl_2N_2O$ , C, 64.70; H, 4.34; N, 7.55; Found C, 64.45; H, 4.09; N, 7.78.

### 3.1.3.12 4-(3-Aminobenzofuran-2-yl)-1-(2bromobenzyl) pyridin-1-ium chloride (**5**l)

White solid, yield: (61%); mp 191–193°C. IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3293, 3171, 3076, 1636, 1544, 1463, 1314, 1214, 1155, 1101, 1032, 870, 826, 750, 652. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 5.73 (s, 2H, CH<sub>2</sub>), 7.31 (t, *J* = 8.0 Hz, 2H, Ar-H), 7.39 (t, *J* = 7.5 Hz, 2H, Ar-H), 7.47 (d, *J* = 6.5 Hz, 2H, Ar-H), 7.51 (d, *J* = 10.0 Hz, 1H, Ar-H), 7.54 (d, *J* = 8.5 Hz, 1H, Ar-H), 7.57 (d, *J* = 8.0 Hz, 1H, Ar-H), 7.99 (s, 2H, NH<sub>2</sub>), 8.11 (d, *J* = 8.0 Hz, 1H, Ar-H), 8.64 (d, *J* = 7.0 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 60.9, 116.6, 122.0, 122.1, 122.6, 122.9, 128.2, 128.6, 130.3, 130.9, 133.2, 134.0, 140.6, 140.7, 142.3, 143.1, 143.9, 154.7. Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>BrClN<sub>2</sub>O, C, 57.78; H, 3.88; N, 6.74; Found C, 57.45; H, 3.57; N, 6.47.

### 3.1.3.13 4-(3-Aminobenzofuran-2-yl)-1-(3bromobenzyl) pyridin-1-ium chloride (**5m**)

White solid, yield: (74%); mp 270–272°C. IR (KBr) ( $\nu_{max}/$  cm<sup>-1</sup>): 3292, 3159, 1634, 1539, 1313, 1160, 1098, 1032, 834, 757, 671. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta = 5.63$  (s, 2H, CH<sub>2</sub>), 7.30 (t, J = 7.0 Hz, 1H, Ar-H), 7.41 (d, J = 6.5 Hz, 2H, Ar-H), 7.51–7.57 (m, 3H, Ar-H), 7.61 (d, J = 7.5 Hz, 1H, Ar-H), 7.81 (s, 1H, Ar-H), 7.97 (s, 2H, NH<sub>2</sub>), 8.09 (d, J = 7.5 Hz, 2H, Ar-H), 8.79 (d, J = 6.0 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta = 59.9$ , 115.6, 116.8, 121.9, 122.1, 122.6, 127.3, 128.2, 130.6, 131.2, 131.8, 137.6, 138.1, 140.3, 142.2, 142.6, 142.7, 154.6. Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>BrClN<sub>2</sub>O, C, 57.78; H, 3.88; N, 6.74; Found C, 58.03; H, 3.61; N, 6.38.

### 3.1.3.14 4-(3-Aminobenzofuran-2-yl)-1-(4bromobenzyl) pyridin-1-ium chloride (**5n**)

White solid, yield: (68%); mp 254–257°C. IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3171, 1636, 1592, 1540, 1314, 1214, 1162, 1100, 1015, 823, 759. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 5.63 (s, 2H, CH<sub>2</sub>), 7.29 (d, J = 7.5 Hz, 1H, Ar-H), 7.43 (d, J = 5.5 Hz, 2H, Ar-H), 7.49 (t, J = 8.0 Hz, 2H, Ar-H), 7.53 (d, J = 7.5 Hz, 2H, Ar-H), 7.64 (d, J = 7.5 Hz, 2H, Ar-H), 7.96 (s, 2H, NH<sub>2</sub>), 8.10 (d, J = 8.0 Hz, 1H, Ar-H), 8.77 (d, J = 6.5 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 59.9, 116.6, 116.8, 121.9, 122.1, 122.3, 122.6, 128.1, 130.5, 132.0, 134.5, 140.3, 140.4, 142.1, 142.7, 154.6. Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>BrClN<sub>2</sub>O, C, 57.78; H, 3.88; N, 6.74; Found C, 57.59; H, 4.02; N, 6.52.

# 3.1.3.15 4-(3-Aminobenzofuran-2-yl)-1-(2-nitrobenzyl) pyridin-1-ium chloride (**50**)

Yellow solid, yield: (67%); mp 148–150°C. IR (KBr) ( $\nu_{max}$ /cm<sup>-1</sup>): 3167, 2986, 1707, 1639, 1525, 1337, 1170, 1107, 1045, 854, 789, 736. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta = 6.02$  (s, 2H, CH<sub>2</sub>), 7.13 (d, J = 7.5 Hz, 1H, Ar-H), 7.32 (t, J = 8.0 Hz, 2H, Ar-H), 7.53 (d, J = 8.0 Hz, 2H, Ar-H), 7.57 (t, J = 7.0 Hz, 1H, Ar-H), 7.71 (d, J = 7.5 Hz, 1H, Ar-H), 7.80 (d, J = 7.5 Hz, 1H, Ar-H), 8.04 (s, 2H, NH<sub>2</sub>), 8.12 (d, J = 8.0 Hz, 1H, Ar-H), 8.66 (d, J = 6.5 Hz,

2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 58.2, 116.7, 122.0, 122.2, 122.6, 125.5, 128.2, 129.1, 129.9, 130.3, 130.6, 134.8, 140.7, 142.4, 143.3, 144.0, 147.3, 154.7. Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>ClN<sub>3</sub>O<sub>3</sub>, C, 62.91; H, 4.22; N, 11.01; Found C, 63.11; H, 3.95; N, 11.35.

# 3.1.3.16 4-(3-Aminobenzofuran-2-yl)-1-(4-nitrobenzyl) pyridin-1-ium chloride (**5p**)

Yellow solid, yield: (73%); mp 173–175°C. IR (KBr) ( $\nu_{max}$ / cm<sup>-1</sup>): 3302, 3178, 3104, 1637, 1533, 1340, 1213, 1165, 1103, 1031, 827, 753, 689. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  = 5.81 (s, 2H, CH<sub>2</sub>), 7.31 (t, J = 7.0 Hz, 1H, Ar-H), 7.49–7.57 (m, 3H, Ar-H), 7.74 (d, J = 8.0 Hz, 2H, Ar-H), 7.99 (s, 2H, NH<sub>2</sub>), 8.10 (d, J = 7.5 Hz, 2H, Ar-H), 8.28 (d, J = 7.5 Hz, 2H, Ar-H), 8.79 (d, J = 6.0 Hz, 2H, Ar-H). <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ ):  $\delta$  = 59.7, 116.8, 122.0, 122.2, 122.5, 123.9, 124.1, 128.2, 129.4, 130.6, 140.7, 142.2, 142.3, 142.8, 147.6, 154.7. Anal. Calcd. for C<sub>20</sub>H<sub>16</sub>ClN<sub>3</sub>O<sub>3</sub>, C, 62.91; H, 4.22; N, 11.01; Found C, 62.68; H, 4.51; N, 10.83.

### 3.2 In vitro biological evaluations

#### 3.2.1 Cholinesterase inhibition assay

The method described by Ellman was used to evaluate the inhibitory activity of compounds **5a-p** against AChE, according to procedures previously reported. AChE from E. electricus, and horse serum BuChE (eqBuChE, Sigma-Aldrich) 5,5-Dithiobis-(2-nitrobenzoic acid) (DTNB), and acetylthiocholine iodide (ATC) were purchased from Sigma Aldrich. Donepezil was used as a standard drug and all experiments were assayed in triplicate. In this procedure, five different concentrations of each compound were used at 25°C. To determine the IC<sub>50</sub>, 100  $\mu$ l of DTNB, 3 ml phosphate buffer (0.1 M, pH = 8.0), 50  $\mu$ l of the enzyme, and 50  $\mu$ l of each compound were incubated in 24 well plates for 5 min followed by the addition of 20  $\mu$ l of the substrate (acetylthiocholine iodide) and absorbance was detected at 415 nm for 6 min, and then the percent inhibition was plotted from inhibition curves. For the invitro BuChE assay similar method has been used.

#### 3.2.2 Kinetics study

To understand further insight into the inhibition model of compounds, the kinetic study was performed for the most active compound **5f** according to procedures previously reported (Baharloo et al., 2015). The experiment was assayed in triplicate.

# 3.2.3 *In vitro* inhibition of self-induced and AChE-induced $A\beta_{1-42}$ aggregation

To assess the inhibitory effect of compounds **5a**, **5f**, **5h**, **5i**, and **5l** on the  $\beta$ -Amyloid aggregation, we used a thioflavin T (ThT)based fluorescence assay to determine the self-induced and AChEinduced A $\beta_{1-42}$  aggregation (Levine, 1993). A $\beta_{1-42}$  (Sigma A9810) 50  $\mu$ M was dissolved in ammonium hydroxide (1%  $\nu/\nu$ ) for prefibrillation and incubated for 72 h at 37°C. A $\beta_{1-42}$  (10  $\mu$ l) in the absence and presence of human recombinant AChE (0.01 u/mL, Sigma C1682) were added to phosphate buffer (0.05 M, pH = 7.4) and incubated at 37°C for 48 h with or without target compounds **5a, 5f, 5h, 5i,** and **5l** (100  $\mu$ M). Next, 50  $\mu$ l of thioflavin T (ThT, 200  $\mu$ M) containing 50 mM glycine-NaOH buffer (pH 8.5) was added. Fluorescences were recorded ( $\lambda_{exc}$  = 446 nm;  $\lambda_{em}$  = 490 nm) by Microplate Reader (Spectra Max). Donepezil (100  $\mu$ M, Sigma D-6821) was used as a reference drug. The aggregation inhibition of the tested compounds was calculated on AChE or self-induced Aβ-aggregation according to the equation (1–IF<sub>i</sub>/IF<sub>c</sub>) × 100, where IF<sub>i</sub> and IFc are the fluorescence intensities in the presence of inhibitors and in the absence of inhibitors, respectively.

#### 3.2.4 Cell viability assay

In order to evaluate cell viability of **5a**, **5f**, **5h**, **5i**, and **5l**, the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay was performed using PC12 cell line. Cells were seeded in 96-well plates at a density of 5,000 cells/well for 24 h, and then cells were treated with tested compounds and glutamate (5 mM) and incubated at 37°C for 24 h. After that, the medium was removed and 20  $\mu$ l of the MTT reagent (5 mg/ml) was added and incubated at 37 °C for 4 h. Then, the medium was replaced with 150  $\mu$ l DMSO to solve formazan crystals. The absorbance was recorded by microplate reader apparatus (Biotek, Winooski, VT, United States) at 570 nm. The assay was performed in three independent experiments and triplicate.

#### 3.2.5 Docking simulations

The molecular docking studies were performed using the program Autodock Vina (Trott and Olson 2010). The crystal structure of acetyl cholinesterase (1EVE) was derived from the Protein Data Bank (http://www.rcsb.org/). The co-crystallized ligand and water molecules were deleted from the protein structure. The top 3-aminocoumarin derivative **5f** was selected for the docking study. The selected ligand was constructed using Marvin Sketch, 2012, Chem Axon (http://www.chemaxon.com). The center of the grid box were set as follow; x = 2.071, y = 63.616, z = 67.737, and the active site box were set at 15 Å × 15 Å × 15 Å. Molecular visualizations were carried out by Discovery Studio 4. 5 client software (Accelrys, Inc., San Diego, CA, United States).

### 4 Conclusion

In the present study, a novel series of 3-aminobenzofuran derivatives with *N*-benzylpyridinium moiety were designed, synthesized, and evaluated as novel agents for the treatment of AD. The in-vitro assays revealed that most target compounds had moderate to good anti-AChE and -BuChE activity with no toxicity against PC12 cells. The best result was obtained from 2-fluorobenzyl derivative **5f** which exhibited potent AChE and BuChE inhibitory activity with IC<sub>50</sub> = 0.64 and 0.55  $\mu$ M, respectively. Moreover, compounds **5f** and **5h** have remarkable anti-aggregation activity compared to donepezil as the reference drug. Docking's study

demonstrated that 3-aminobenzofuran (hydrophobic aromatic fragment) can be positioned in the peripheral anionic site and the *N*-benzylpyridinium fragment bound to the catalytic anionic site of AChE. These results showed that compound **5f** could be considered a promising multifunctional derivative for further studies in the field of new anti-Alzheimer agents.

### Data availability statement

The original contributions presented in the study are included in the article/Supplementary Materials; further inquiries can be directed to the corresponding author.

### Author contributions

ZH and RM performed synthesis and characterization of the target compounds. SNAB evaluated anti-A $\beta$  aggregation. HN and TA performed AChE, BuChE, and kinetics studies. RF participated in the ChEs inhibition assay. SM and AA wrote the first draft of the manuscript and performed docking studies. AF controlled the synthetic results and reviewed the manuscript for proofreading. All authors reviewed and agreed to the content of the manuscript.

# Funding

This work was supported and funded by Tehran University of Medical Sciences, Grant No. 99-3-104-51406.

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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### Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fchem. 2022.882191/full#supplementary-material Al-Qirim, T., Shattat, G., Sweidan, K., El-Huneidi, W., Sheikha, G. A., Khalaf, R. A., et al. (2012). *In vivo* antihyperlipidemic activity of a new series of N-(Benzoylphenyl) and N-(Acetylphenyl)-1-benzofuran-2-carboxamides in rats. *Arch. Pharm. Weinh.* 345, 401–406. doi:10.1002/ardp. 201100225

Alper-Hayta, S., Arisoy, M., Temiz-Arpaci, Ö., Yildiz, I., Aki, E., Özkan, S., et al. (2008). Synthesis, antimicrobial activity, pharmacophore analysis of some new 2-(substitutedphenyl/benzyl)-5-[(2-benzofuryl)carboxamido]benzoxazoles. *Eur. J. Med. Chem.* 43, 2568–2578. doi:10.1016/j.ejmech.2007.12.019

Baharloo, F., Moslemin, M. H., Nadri, H., Asadipour, A., Mahdavi, M., Emami, S., et al. (2015). Benzofuran-derived benzylpyridinium bromides as potent acetylcholinesterase inhibitors. *Eur. J. Med. Chem.* 93, 196–201. doi:10.1016/j. ejmech.2015.02.009

Byun, J. H., Kim, H., Kim, Y., Mook-Jung, I., Kim, D. J., Lee, W. K., et al. (2008). Aminostyrylbenzofuran derivatives as potent inhibitors for Aβ fibril formation. *Bioorg. Med. Chem. Lett.* 18, 5591–5593. doi:10.1016/j.bmcl.2008. 08.111

Camps, P., Formosa, X., Galdeano, C., Gómez, T., Muñoz-Torrero, D., Scarpellini, M., et al. (2008). Novel donepezil-based inhibitors of acetyl- and butyrylcholinesterase and acetylcholinesterase-induced  $\beta$ -amyloid aggregation. J. Med. Chem. 51, 3588–3598. doi:10.1021/jm8001313

Castro, A., and Martinez, A. (2001). Peripheral and dual binding site acetylcholinesterase inhibitors: Implications in treatment of alzheimer's disease. *Mini Rev. Med. Chem.* 1, 267–272. doi:10.2174/1389557013406864

Cavalli, A., Bolognesi, M. L., Minarini, A., Rosini, M., Tumiatti, V., Recanatini, M., et al. (2008). Multi-target-Directed ligands to combat neurodegenerative diseases. J. Med. Chem. 51, 347–372. doi:10.1021/jm7009364

Chen, W., Deng, X.-Y., Li, Y., Yang, L.-J., Wan, W.-C., Wang, X.-Q., et al. (2013). Synthesis and cytotoxic activities of novel hybrid 2-phenyl-3-alkylbenzofuran and imidazole/triazole compounds. *Bioorg. Med. Chem. Lett.* 23, 4297–4302. doi:10.1016/j. bmcl.2013.06.001

Choi, H. D., Seo, P. J., Son, B. W., and Kang, B. W. (2004). Synthesis of 2-(4hydroxyphenyl)benzofurans and their application to -amyloid aggregation inhibitor. *Arch. Pharm. Res.* 27, 19. doi:10.1007/BF02980039

Choi, M.-J., Jung, K. H., Kim, D., Lee, H., Zheng, H.-M., Park, B. H., et al. (2011). Anti-cancer effects of a novel compound HS-113 on cell growth, apoptosis, and angiogenesis in human hepatocellular carcinoma cells. *Cancer Lett.* 306, 190–196. doi:10.1016/j.canlet.2011.03.005

Cummings, J. L., Morstorf, T., and Zhong, K. (2014). Alzheimer's disease drugdevelopment pipeline: Few candidates, frequent failures. *Alzheimer's Res. Ther.* 6, 37–43. doi:10.1186/alzrt269

Ellman, G. L., Courtney, K. D., Andres, V., and Featherstone, R. M. (1961). A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol.* 7, 88–95. doi:10.1016/0006-2952(61)90145-9

Forette, F. R., Anand, G., and Gharabawi, A. (1999). A phase II study in patients with Alzheimer's disease to assess the preliminary efficacy and maximum tolerated dose of rivastigmine (ExelonR). *Eur. J. Neurol.* 6, 423–429. doi:10.1046/j.1468-1331. 1999.640423.x

Gauthier, S., Reisberg, B., Zaudig, M., Petersen, R. C., Ritchie, K., Broich, K., et al. (2006). Mild cognitive impairment. *Lancet* 367, 1262–1270. doi:10.1016/S0140-6736(06)68542-5

Goyal, D., Kaur, A., and Goyal, B. (2018). Benzofuran and indole: Promising scaffolds for drug development in Alzheimer's disease. *ChemMedChem* 13, 1275–1299. doi:10.1002/cmdc.201800156

Greenough, M. A., Camakaris, J., and Bush, A. I. (2013). Metal dyshomeostasis and oxidative stress in Alzheimer's disease. *Neurochem. Int.* 62, 540–555. doi:10. 1016/j.neuint.2012.08.014

Howlett, D. R., Perry, A. E., Godfrey, F., Swatton, J. E., Jennings, K. H. C., Wood, S. J., et al. (1999). Inhibition of fibril formation in  $\beta$ -amyloid peptide by a novel series of benzofurans. *Biochem. J.* 340, 283–289. doi:10.1042/bj3400283

Inestrosa, N. C., Alvarez, A., Pérez, C. A., Moreno, R. D., Vicente, M., Linker, C., et al. (1996). Acetylcholinesterase accelerates assembly of amyloid-beta-peptides into Alzheimer's fibrils: Possible role of the peripheral site of the enzyme. *Neuron* 16, 881–891. doi:10.1016/s0896-6273(00)80108-7

Kapková, P., Alptüzün, V., Frey, P., Erciyas, E., and Holzgrabe, U. (2006). Search for dual function inhibitors for Alzheimer's disease: Synthesis and biological activity of acetylcholinesterase inhibitors of pyridinium-type and their A $\beta$  fibril formation inhibition capacity. *Bioorg. Med. Chem.* 14, 472–478. doi:10.1016/j.bmc.2005.08.034

Koellner, G., Kryger, G., Millard, C. B., Silman, I., Sussman, J. L., Steiner, T., et al. (2000). Active-site gorge and buried water molecules in crystal structures of acetylcholinesterase from *Torpedo californica*. J. Mol. Biol. 296, 713–735. doi:10. 1006/jmbi.1999.3468

Lai, R., Albala, B., Kaplow, J. M., Aluri, J., YenSatlin, M. A., and Satlin, A. (2012). First-in-human study of E2609, a novel BACE1 inhibitor, demonstrates prolonged reductions in plasma beta-amyloid levels after single dosing. *Alzheimers Dement.* 8, P96. doi:10.1016/j.jalz.2012.05.237

Lane, R. M., Kivipelto, M., and Greig, N. H. (2004). Acetylcholinesterase and its inhibition in Alzheimer disease. *Clin. Neuropharmacol.* 27, 141–149. doi:10.1097/00002826-200405000-00011

León, R., Garcia, A. G., and Marco-Contelles, J. (2013). Recent advances in the multitarget-directed ligands approach for the treatment of Alzheimer's disease. *Med. Res. Rev.* 33, 139–189. doi:10.1002/med.20248

Levine, H. (1993). Thioflavine T interaction with synthetic Alzheimer's disease  $\beta$ -amyloid peptides: Detection of amyloid aggregation in solution. *Protein Sci.* 2, 404–410. doi:10.1002/pro.5560020312

Li, F., Wu, J.-J., Wang, J., Yang, X.-L., Cai, P., Liu, Q.-H., et al. (2017). Synthesis and pharmacological evaluation of novel chromone derivatives as balanced multifunctional agents against Alzheimer's disease. *Bioorg. Med. Chem.* 25, 3815–3826. doi:10.1016/j.bmc.2017.05.027

Loidreau, Y., Dubouilh-Benard, C., Marchand, P., Nourrisson, M.-R., Duflos, M., Buquet, C., et al. (2013). Efficient new synthesis of N-Arylbenzo[b]furo[3, 2-d] pyrimidin-4-amines and their benzo[b]thieno[3, 2-d]pyrimidin-4-amine analogues via a microwave-assisted dimroth rearrangement. *J. Heterocycl. Chem.* 50, 1187–1197. doi:10.1002/jhet.1716

Longo, F. M., and Massa, S. M. (2004). Neuroprotective strategies in Alzheimer's disease. *NeuroRX* 1, 117–127. doi:10.1602/neurorx.1.1.117

Marco-Contelles, J., León, R., de los Ríos, C., Guglietta, A., Terencio, J., López, M. G., et al. (2006). Novel multipotent Tacrine–Dihydropyridine hybrids with improved acetylcholinesterase inhibitory and neuroprotective activities as potential drugs for the treatment of alzheimer's disease. *J. Med. Chem.* 49, 7607–7610. doi:10.1021/jm061047j

McKhann, G. M., Knopman, D. S., Chertkow, H., Hyman, B. T., Jack, C. R., Kawas, C. H., et al. (2011). The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's*. & amp. Dement. 7, 263–269. doi:10.1016/j.jalz.2011.03.005

Mehrabi, M., Pourshojaei, Y., Moradi, A., Sharifzadeh, M., Khosravani, L., Sabourian, R., et al. (2017). Design, synthesis, molecular modeling and anticholinesterase activity of benzylidene-benzofuran-3-ones containing cyclic amine side chain. *Future Med. Chem.* 9, 659–671. doi:10.4155/fmc-2016-0237

Mesulam, M. M., and Asunction, M. M. (1987). Cholinesterases within neurofibrillary tangles related to age and Alzheimer's disease. *Ann. Neurol.* 22, 223–228. doi:10.1002/ana.410220206

Muñoz-Ruiz, P., Rubio, L., García-Palomero, E., Dorronsoro, I., del Monte-Millán, M., Valenzuela, R., et al. (2005). Design, synthesis, and biological evaluation of dual binding site acetylcholinesterase inhibitors: New diseasemodifying agents for alzheimer's disease. *J. Med. Chem.* 48, 7223–7233. doi:10.1021/jm0503289

Nadri, H., Pirali-Hamedani, M., Shekarchi, M., Abdollahi, M., Sheibani, V., Amanlou, M., et al. (2010). Design, synthesis and anticholinesterase activity of a novel series of 1benzyl-4-((6-alkoxy-3-oxobenzofuran-2(3H)-ylidene) methyl) pyridinium derivatives. *Bioorg. Med. Chem.* 18, 6360–6366. doi:10.1016/j.bmc.2010.07.012

Nevagi, R. J., Dighe, S. N., and Dighe, S. N. (2015). Biological and medicinal significance of benzofuran. *Eur. J. Med. Chem.* 97, 561–581. doi:10.1016/j.ejmech. 2014.10.085

Poprac, P., Jomova, K., Simunkova, M., Kollar, V., Rhodes, C. J., Valko, M., et al. (2017). Targeting free radicals in oxidative stress-related human diseases. *Trends Pharmacol. Sci.* 38, 592–607. doi:10.1016/j.tips.2017.04.005

Pouramiri, B., Mahdavi, M., Moghimi, S., Firoozpour, L., Nadri, H., Moradi, A., et al. (2016). Synthesis and antiacetylcholinesterase activity evaluation of new 2-aryl benzofuran derivatives. *Lett. Drug Des. Discov.* 13, 897–902. doi:10.2174/1570180813666160610124637

Rizzo, S., Tarozzi, A., Bartolini, M., Da Costa, G., Bisi, A., Gobbi, S., et al. (2012). 2-Arylbenzofuran-based molecules as multipotent Alzheimer's disease modifying agents. *Eur. J. Med. Chem.* 58, 519–532. doi:10.1016/j.ejmech.2012. 10.045

Rosini, M., Simoni, E., Minarini, A., and Melchiorre, C. (2014). Multi-target design strategies in the context of alzheimer's disease: Acetylcholinesterase inhibition and NMDA receptor antagonism as the driving forces. *Neurochem. Res.* 39, 1914–1923. doi:10.1007/s11064-014-1250-1

Saxena, A., Fedorko, J. M., Vinayaka, C. R., Medhekar, R., Radic, Z., Taylor, P., et al. (2003). Aromatic amino-acid residues at the active and peripheral anionic sites control the binding of E2020 (AriceptR) to cholinesterases. *Eur. J. Biochem.* 270, 4447–4458. doi:10.1046/j.1432-1033.2003.03837.x

Scarpini, E., Scheltens, P., and Feldman, H. (2003). Treatment of alzheimer's disease: Current status and new perspectives. *Lancet Neurol.* 2, 539–547. doi:10. 1016/s1474-4422(03)00502-7

Schneider, L. S. (2000). A critical review of cholinesterase inhibitors as a treatment modality in Alzheimer's disease. *Dialogues Clin. Neurosci.* 2, 111–128. doi:10.31887/DCNS.2000.2.2/lschneider

Smith, C. P., Bores, G. M., Petko, W., Li, M., Selk, D. E., Rush, D. K., et al. (1997). Pharmacological activity and safety profile of P10358, a novel, orally active acetylcholinesterase inhibitor for Alzheimer's disease. *J. Pharmacol. Exp. Ther.* 280, 710–720.

Talesa, V. N. (2021). Acetylcholinesterase in Alzheimer's disease. Mech. Ageing Dev. 122, 1961–1969. doi:10.1016/s0047-6374(01)00309-8

Trott, O., and Olson, A. J. (2010). AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. J. Comput. Chem. 31, 455–461. doi:10.1002/jcc.21334

Weinstock, M. (1999). Selectivity of cholinesterase inhibition: Clinical implications for the treatment of Alzheimer's disease. *CNS Drugs* 12, 307–323. doi:10.2165/00023210-199912040-00005