

Original Article



OPEN ACCESS

Received: Apr 26, 2018

Revised: Aug 8, 2018

Accepted: Sep 1, 2018

Correspondence to

Murat Cakir

Department of Pediatric Gastroenterology
Hepatology and Nutrition, Faculty of Medicine,
Karadeniz Technical University, Trabzon 61080,
Turkey.

E-mail: muratcak@hotmail.com

Copyright © 2019 by The Korean Society of
Pediatric Gastroenterology, Hepatology and
Nutrition

This is an open-access article distributed
under the terms of the Creative Commons
Attribution Non-Commercial License ([https://
creativecommons.org/licenses/by-nc/4.0/](https://creativecommons.org/licenses/by-nc/4.0/))
which permits unrestricted non-commercial
use, distribution, and reproduction in any
medium, provided the original work is properly
cited.

Funding

The study was funded by the Karadeniz
Technical University Scientific Research
Projects Unit (study ID: 6704 and code
number: 11560).

Conflict of Interest

The authors have no financial conflicts of
interest.

Neurocognitive Functions in Infants with Malnutrition; Relation with Long-chain Polyunsaturated Fatty Acids, Micronutrients Levels and Magnetic Resonance Spectroscopy

**Murat Cakir,¹ Sukran Senyuva,² Sibel Kul,³ Elif Sag,¹ Ali Cansu,⁴
Fulya Balaban Yucesan,⁵ Serap Ozer Yaman,⁵ and Asim Orem⁵**

¹Department of Pediatric Gastroenterology Hepatology and Nutrition, Faculty of Medicine, Karadeniz Technical University, Trabzon, Turkey

²Department of Pediatrics, Faculty of Medicine, Karadeniz Technical University, Trabzon, Turkey

³Department of Radiology, Faculty of Medicine, Karadeniz Technical University, Trabzon, Turkey

⁴Department of Pediatric Neurology, Faculty of Medicine, Karadeniz Technical University, Trabzon, Turkey

⁵Department of Biochemistry, Faculty of Medicine, Karadeniz Technical University, Trabzon, Turkey

ABSTRACT

Purpose: Malnutrition may influence neurocognitive development in children by directly affecting the brain structural development, or indirectly by affecting the children's cognition experience. Malnutrition alters the cell numbers, cell migration, synaptogenesis, and neurotransmission due to inadequate availability of necessary micronutrients to support cell growth. We aimed to analyze neurocognitive development in infants with malnutrition and its association with long chain polyunsaturated fatty acids (LC-PUFA), micronutrients levels and magnetic resonance spectroscopy (MRS) findings.

Methods: The study included two groups; group 1, infants with malnutrition (n=24), group 2; healthy infants (n=21). Peripheral blood was obtained from the participants for studying micronutrients and LC-PUFA levels. The neurocognitive development was analyzed by the use of an Ankara Developmental Screening Inventory test. MRS were performed on all infants.

Results: All parameters of neurocognitive development and serum calcium (9.6±0.9 mg/dL vs. 10.4±0.3 mg/dL, $p<0.05$) and magnesium (2.02±0.27 mg/dL vs. 2.2±0.14 mg/dL, $p<0.05$) levels were noted as being low in infants with marked malnutrition. No difference was found in LC-PUFA levels between healthy and malnourished infants. Thalamic choline/creatine levels were significantly high in infants with malnutrition (1.33±0.22 vs. 1.18±0.22, $p<0.05$). Total neurocognitive development in infants was positively correlated with serum calcium levels ($p<0.05$, $r=0.381$).

Conclusion: Calcium supplementation may improve neurocognitive development in malnourished infants.

Keywords: Malnutrition; Neurocognitive; Calcium; Fatty acids, unsaturated; Spectroscopy

INTRODUCTION

Infancy is a vulnerable period for brain development, and nutritional inadequacy, in another words malnutrition, in this period may negatively influence the neurocognitive development in the later life of a developing child. Malnutrition may influence neurocognitive development by directly affecting the brain structural development, or indirectly by affecting children's experience and motor behavior. The incidence of malnutrition also alters the cell numbers, cell migration, myelination, synaptogenesis, hippocampal formation and neurotransmission of a child's developing brain, due to inadequate availability of energy, protein and micronutrients lacking during this critical brain development process in children. Additionally, it may affect the overall health and well-being of a child, by reducing immune competence and resistance to infections, and affect the prolonged duration of recovering from the infections. Chronic and frequent infections may affect the child's physical activity and interactions with the environment [1-3].

Micronutrients deficiencies are a widespread problem in children throughout the globe, especially iron deficiency anemia (25% under the age of 3 years), inadequate zinc, iodine intake and vitamin B12 deficiency. Micronutrients are involved in the brain development, and deficiencies are traditionally associated with cognitive and motor impairment [4]. There has been an increasing interest regarding the role of long chain polyunsaturated fatty acids (LC-PUFA) on the neurocognitive development in children in recent years. These fatty acids are critical for the brain development and function in children, and play a central functional role in brain tissue development, which are precursors for the mediators that play a role in the immune reactions, inflammation and the influence of neuronal signal processing and transmission in children [5].

Magnetic resonance spectroscopy (MRS) is a novel advanced imaging technique used as a tool to identify and categorize non-invasive information about the biochemical composition of imaged tissue. The clinical uses in pediatrics of this test include the diagnosis of brain tumors and inherited metabolic diseases and prognosis of hypoxic-ischemic encephalopathy, traumatic brain injury, demyelinating conditions and infectious brain lesions [6]. Neurocognitive impairment is expected in infants with malnutrition. But its association with micronutrients, LC-PUFA levels and MRS findings was not widely evaluated in the past. Therefore, we aimed to analyze the neurocognitive functions in infants with malnutrition and its association with LC-PUFA, micronutrients levels and MRS findings.

MATERIALS AND METHODS

The study included two groups of infants aged between 2–36 months; group 1, infants with malnutrition (n=24) and group 2; age and sex matched healthy infants (n=21). The diagnosis of malnutrition was made according to the standard World Health Organization (WHO) guidelines. Severe malnutrition is defined as the presence of weight for height Z score <3 standard deviation or presence of bilateral nutritional edema or visible severe wasting [7]. Infants with prematurity, intrauterine growth retardation, neurometabolic diseases, and structural central nervous system anomalies or diseases associated with neurocognitive impairment (such as congenital hypothyroidism) were not included into the group 1. None of the infants in group 2 had known chronic diseases, prematurity or intrauterine growth retardation.

After informant and written consent from the parents of the infants, anthropometric measurement was performed. Then, the sample of peripheral blood was obtained for studying the complete blood count, albumin, lipid and micronutrients level including calcium, magnesium, iron, folic acid, zinc, vitamin A and B12 levels and LC-PUFA levels (arachidonic acid [AA] and docosahexaenoic acid [DHA]). Then, a neurocognitive development assessment and MRS were performed for all infants.

Assessment of neurocognitive development

The neurocognitive development of the infants was analyzed with the use of an Ankara Developmental Screening Inventory (ADSI) test. This tool was developed by Erol et al. [8] as a culturally relevant device to yield an index of the developmental status of children. The ADSI is a measurement tool that evaluates five component scales (general development, social and self-care skills, fine motor and gross motor developments, and language-cognitive language), and total points (T points) of children between the age of 1 month and 6 years. The inventory consisted of 154 items answered by the mothers of the children as either response of a “yes” or “no.” If a child received a score of 20%–30% lower than the average score for his or her age in at least two subtests, he or she was classified as having poor neurocognitive development [9].

MRS analysis

Multivoxel MRS (TR/TE, 1.500/135) was performed on all infants with a standard head coil and 1.5-T MRI system (Magnetom Symphony; Siemens Healthcare, Erlangen, Germany). Three orthogonal baseline images were obtained with automated magnetic field shimming. A volume of interest with a thickness of 15 mm was placed on the centrum semiovale directly over the corpus callosum, and parallel to the sphenoidal plane. After optimal water signal suppression with chemical shift-selective techniques, the MRS data were collected. The entire evaluation was completed within 7 minutes. Three specific voxels were selected from the bilateral thalamus, basal ganglia and from the white matter within the volume of interest. Spectra were generated by the use of a Fourier transformation from the data collected, from these eight specific voxels within the volume of interest. After manual baseline correction (near zero) procedures, the metabolite peaks were determined. Three dominant spectra (choline [Cho] at 3.21 parts per million, creatine [Cr] at 3.04 parts per million, and N-acetylaspartate [NAA] at 2.02 parts per million) were analyzed.

Determination of LC-PUFA in erythrocyte membranes

Erythrocyte membranes were isolated from red blood cells according to the method of Hamaguchi et al. [10]. Preparation of fatty acid methyl esters in erythrocyte membranes was performed by using the method of Sattler et al. [11], with some modifications. Fatty acid composition of the erythrocyte membrane was determined by a gas chromatographic method (Agilent Technologies 6890N; Agilent Technologies, Waldbronn, Germany) using a flame ionization detector. Experimental conditions for gas chromatographic flame ionization detector were as follows; column: HP-88 capillary column 100 m×0.250 mm×0.20 µm nominal, inlet temperature: 250°C, injection volume: 1 µL, carrier gas: helium, head pressure: 230 kPa constant pressure (33 cm/s at 50°C), oven temperature: 120°C for 1 minute, 10°C/min to 175°C, hold at 175°C for 10 minutes, 5°C/min to 210°C, hold at 210°C for 5 minutes, 5°C/min to 230°C, hold at 230°C for 10 minutes. Analytical standard (Supelco 37-component FAME mixture, 47885-U; Sigma-Aldrich Co., Dorset, UK) was used for the verification of retention times. DHA and AA in erythrocyte membranes were reported as weight percentages of the total fatty acids analyzed.

Statistical analysis

Statistical analysis was performed using the Statistical Package for Social Sciences version 23 (IBM Corp., Armonk, NY, USA). The noted differences between the groups were calculated using an independent samples *t*-test (Student's *t*-test) for the normally distributed data and the Mann-Whitney *U* test for data that was not normally distributed. A χ^2 test was used for the comparisons of qualitative data. Correlations between the variables were calculated using a linear regression. The values of $p < 0.05$ were considered significant.

Informed consent for participating in the study was obtained from parents of all cases, and the study was approved by the ethics committee (2013/129). The study was funded by the Karadeniz Technical University Scientific Research Projects Unit (study ID: 6704 and code number: 11560).

RESULTS

The demographic and clinic findings of the patients and control group are shown in **Table 1**. No difference was found between the groups in terms of age, gender and duration of breastfeeding ($p > 0.05$ for all). As expected, anthropometric parameters including weight, weight Z score, height and height Z score were lower in patients with malnutrition ($p < 0.05$ for all). According to the WHO guidelines, 10 patients (41.7%) had severe malnutrition.

Neurocognitive functions of the infants with malnutrition and who were in the control group are shown in **Table 2**. All neurocognitive functions were found to be lower in infants with malnutrition ($p < 0.05$ for all).

Laboratory parameters of the patients and control group are shown in **Table 3**. Total protein, albumin, total cholesterol, Ca and Mg levels were significantly low in patients with malnutrition ($p < 0.05$ for all). No significant difference was found in LC-PUFA levels within the groups.

Table 1. Demographic and clinic findings of the patients and control group

Parameters	Group 1 (n=24)	Group 2 (n=21)
Age (mo)	10.8±7.6	11.1±9.6
Gender (female)	15 (62.5)	14 (66.7)
Duration of breastfeeding (mo)	7.1±5.7	8.1±3.1
Etiology of malnutrition		
Primary	12 (50)	
Secondary	12 (50)	
Congenital heart disease	3	
Celiac disease	2	
Short bowel syndrome	2	
Cystic fibrosis	2	
Multiple food allergy	2	
Congenital cleft plate and lip	1	
Severe malnutrition	10 (41.7)	
Edema	3 (12.5)	
Hepatomegaly	2 (8.3)	
Weight (g)	5,889±2,209 ^a	9,395±2,684 ^b
Weight Z score	-3.66±1.32 ^c	0.08±0.68 ^d
Height (cm)	64.9±9.9 ^e	74.7±11.5 ^f
Height Z score	-2.68±1.41 ^g	0.17±0.79 ^h

Values are presented as mean±standard deviation or number (%).

^{a-h} $p < 0.05$.

Table 2. Neurocognitive functions of the patients with malnutrition and healthy controls

Parameters	Group 1 (n=24)	Group 2 (n=21)
Language cognitive	-0.77±2.08 ^a	0.81±1.15 ^b
Fine motor	-0.77±1.87 ^c	0.71±0.81 ^d
Gross motor	-0.79±1.94 ^e	0.55±0.85 ^f
Social and self-care skills	-0.88±2.07 ^g	0.90±1.09 ^h
General development	-0.87±1.94 ⁱ	0.88±0.98 ^j
T points	44.17±8.39 ^k	54.24±4.76 ^l
“Poor” neurocognitive development	6 (25)	0

Values are presented as mean±standard deviation or number (%).

T points: total points.

a-b, c-d, e-f, g-h, i-j, k-l $p < 0.05$.

Table 3. Laboratory parameters of the patients and control group

Parameters	Group 1 (n=24)	Group 2 (n=21)
Hemoglobin (g/dL)	10.9±1.3	11.3±1.1
Anemia	12 (50)	8 (38.1)
Total protein (g/dL)	5.8±0.9 ^a	6.5±0.5 ^b
Albumin (g/dL)	3.7±0.7 ^c	4.2±0.3 ^d
Hypoalbuminemia	5 (20.8)	0
Total cholesterol (mg/dL)	124.5±37 ^e	153.9±30.9 ^f
Triglyceride (mg/dL)	150.1±53.2	174.1±100.6
HDL (mg/dL)	33.5±12.0	40.4±12.9
LDL (mg/dL)	80.8±35.6	97.9±34.4
Ferritin (ng/mL)	102.2±150.8	48.1±64.8
Micronutrients		
Ca (mg/dL)	9.6±0.9 ^g	10.4±0.3 ^h
Mg (mg/dL)	2.02±0.27 ⁱ	2.2±0.14 ^j
Fe (µg/dL)	50.3±42.1	48.8±32.9
Vitamin B12 (pg/mL)	455.1±295.7	389.5±177.6
Folic acid (ng/mL)	19.6±7.6	21.9±5.2
Zn (µg/dL)	133.3±80.3	115.5±20
Vitamin A (µg/L)	249.2±114.2	278.4±160.4
LC-PUFA (%)		
AA	15.0±1.8	15.3±2.0
DHA	5.3±1.4	5.1±1.4
AA/DHA	3.1±1.8	3.2±1.1

Values are presented as mean±standard deviation or number (%).

HDL: high-density lipoprotein, LDL: low-density lipoprotein, Ca: calcium, Mg: magnesium, Fe: iron, Zn: zinc, LC-PUFA: long-chain polyunsaturated fatty acids, AA: arachidonic acid, DHA: docosahexaenoic acid.

a-b, c-d, e-f, g-h, i-j $p < 0.05$.

The MRS findings of the patients and control group is shown in **Fig. 1**. It is noted that the Cho/Cr ratio was higher in all regions in infants with malnutrition, but only thalamic Cho/Cr ratio showed a significant difference (1.33±0.22 vs. 1.18±0.22, $p < 0.05$) (**Fig. 2**).

Correlation analysis of parameters such as micronutrients and LC-PUFA levels with total T points was revealed that the T points was only correlated with serum Ca levels ($p < 0.05$, $r = 0.381$) (**Fig. 3**). No correlation was found between the LC-PUFA and total T points. Additionally, no correlation was found between the MRS findings and total T points.

DISCUSSION

The association between malnutrition and neurocognitive development in children have been extensively researched in animal models and human studies in previous studies [12-17]. In this study, we analyzed neurocognitive functions and its associations with micronutrients,

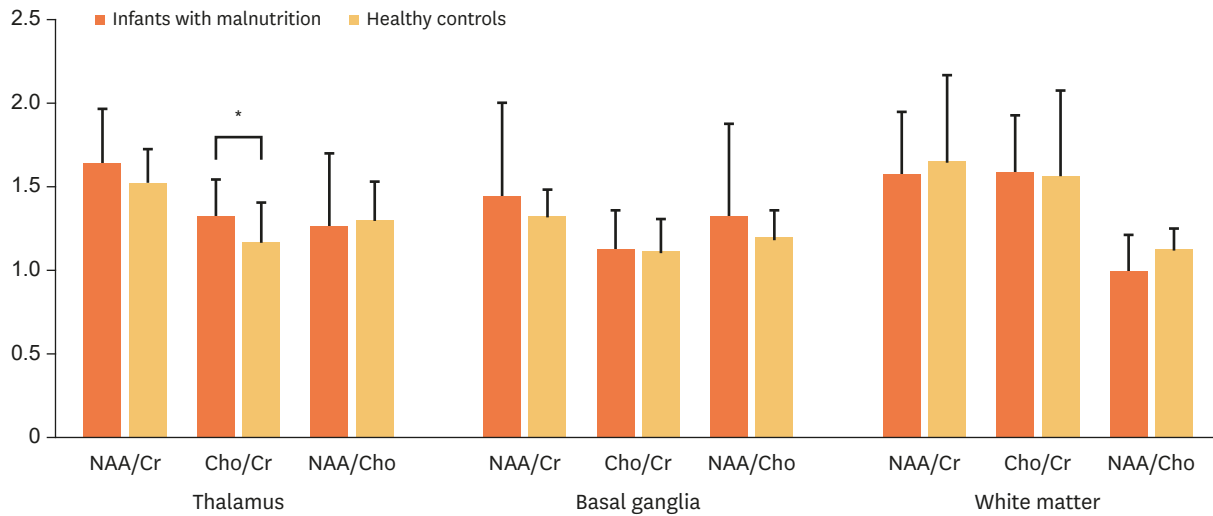


Fig. 1. MRS findings of malnourished and healthy infants. Note thalamic Cho/Cr ratio was higher in infants with malnutrition (1.33±0.22 vs. 1.18±0.22). MRS: magnetic resonance spectroscopy, Cho: choline, Cr: creatine, NAA: N-acetylaspartate. *p<0.05.

LC-PUFA levels and MRS findings in infants with malnutrition. We found that 1) all parameters of neurocognitive functions and serum total cholesterol, Ca and Mg levels were low in infants with malnutrition, 2) no difference was found in LC-PUFA levels between healthy and malnourished infants, 3) thalamic Cho/Cr levels were significantly high in infants with malnutrition, and 4) total neurocognitive development (T points) in infants was positively correlated with serum Ca levels.

Serum lipid profiles were studied in malnourished children previously, and similar to our study total cholesterol levels were found to be low in malnourished children compared to healthy controls in all studies [18-21]. Additionally, they also found that low-density lipoprotein and high-density lipoprotein levels were low and triglyceride levels were high in malnourished children. Disturbance of the lipid metabolism in malnourished children were attributed to low lipoprotein lipase activity in malnourished infants. Hypocalcemia was found

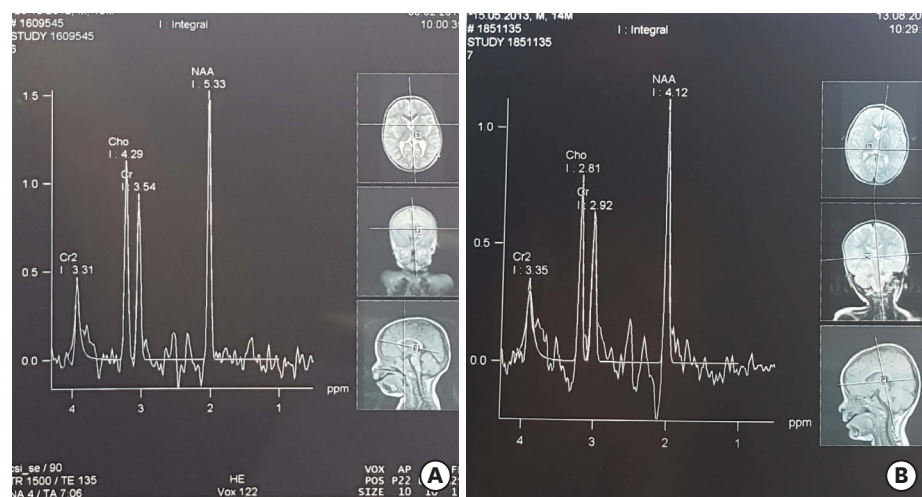


Fig. 2. MRS image of thalamic Cho/Cr ratio of a patient (A) and an infant in the control group (B). MRS: magnetic resonance spectroscopy, Cho: choline, Cr: creatine.

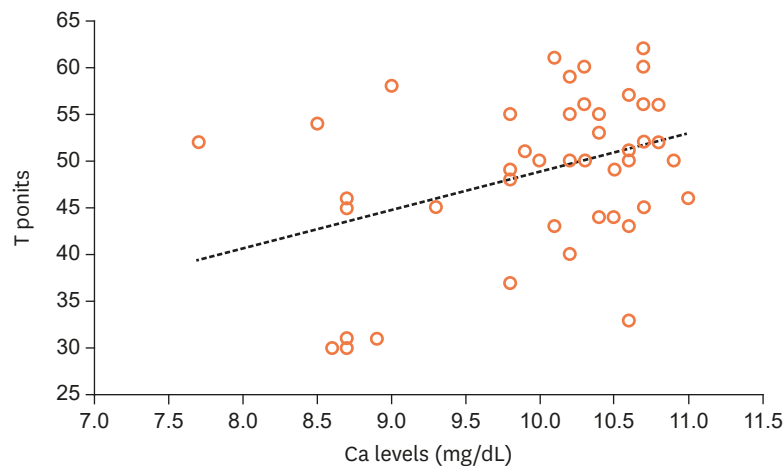


Fig. 3. Correlation of serum Ca levels with T points ($p < 0.05$, $r = 0.381$).
Ca: calcium, T points: total points.

in 26% of the infants with malnutrition at initial admission and found to be associated with an increased risk of mortality on the follow-up [22]. Hypocalcemia was mainly related in this case, with dietary deficiencies or increased metabolic losses. Similar to Ca, Mg levels were also found low in children with malnutrition, and this result was associated with an incidence of lethargy and refractory convulsions in the children [23].

We did not find any difference in the serum LC-PUFA levels between malnourished and healthy infants. Additionally, no association was found between LC-PUFA levels and neurocognitive functions. Contrary to our study, El-khayat et al. [24], showed that LC-PUFA levels were low in children with protein energy malnutrition, and neurocognitive functions were positively correlated with LC-PUFA levels. Neurocognitive functions were improved after nutritional rehabilitation with LC-PUFA enriched enteral nutrition. Additionally, it was shown that LC-PUFA levels have an important function for the intestinal repair process after malnutrition in the early infancy [25]. LC-PUFAs are critical for the brain development and function, but efficiency of LC-PUFA supplementation for the improvement or positive effects on the long-term neurocognitive development of premature or mature infants are still inconsistent [16,26-28]. Some studies showed beneficial effects of LC-PUFA supplementation, whereas some studies or meta-analysis showed no effect [26-29]. Inconsistent results may be associated with differences of duration and doses of LC-PUFA used in the studies. No significant difference was found in the duration of breastfeeding among the groups in our study, but we did not analyze the levels of LC-PUFA in mothers of the infants, therefore LC-PUFA levels may be similar in 2 groups.

MRS analysis especially assessing for motor and neurocognitive outcomes in premature infants with hypoxic-ischemic encephalopathy are widely used in recent years. The combination of Cho/Cr and NAA/Cho ratios in the white matter at term-equivalent age were found to be predictive of motor outcome at 1 year in premature infants [30]. Increased levels of thalamic Cho/Cr ratio were also shown to be useful for the early identification of hypoxic-ischemic encephalopathy in full term neonates [31]. MRS studies in infants with malnutrition are lacking at this time. In our study, we found that Cho/Cr ratio was higher in all regions in infants with malnutrition, but that only thalamic Cho/Cr ratio showed a significant difference. Contrary to our study, Nassar et al. [32], studied the brain metabolites in infants

with protein energy malnutrition (n=16) and healthy controls (n=9), and they found that all metabolites were decreased in infants with protein energy malnutrition and an improvement were obtained after nutritional rehabilitation. Similar to our study, it was shown that Cho/Cr ratio especially in the white matter of the brain, was increased in children older than 2 years with developmental delay, as compared to healthy controls [33].

Finally, we evaluated the parameters associated with neurocognitive function in infants and we found that only serum Ca levels were positively correlated with neurocognitive functions (T points). No correlation was found with the other parameters. The association between maternal vitamin D deficiency and fetal development including bone, lung and brain development have been well defined in previous studies [34]. Maternal vitamin D deficiency were found to be associated with neurocognitive difficulties in 10-year-old offspring. It was shown that the presence of vitamin D has benefits when found in brain receptors in widespread regions of the brain including the hippocampus, prefrontal cortex, cingulate gyrus, basal forebrain, caudate/putamen, thalamus, hypothalamus and cerebellum, and it contributes to the neuroprotection by modulating the production of nerve growth and neuroprotective factors. Decreased levels of vitamin D receptors were shown in the elderly patients with dementia and Alzheimer's disease. There has been no study about the association of serum Ca levels with the neurocognitive development, but it was shown that Ca is essential for synaptic plasticity, a neuronal process that entails gene expression, and which is presumably the cellular counterpart of cognitive brain functions such as learning and memory [34,35].

The limitations of our study were as follows: 1) we were not able to study the vitamin D levels, and 2) neurocognitive development in a child, which is also associated with environmental factors such as cultural, genetic and socio-economic factors that we were not able to analyze these parameters in our study.

In conclusion, our study revealed important clues for the pathogenesis of neurocognitive impairment in infants with malnutrition. Changes in MRS after nutritional rehabilitation and long-term consequences of high thalamic Cho/Cr ratio must be prospectively evaluated. Our findings about the LC-PUFA levels need to be supported with large-number of patients.

REFERENCES

1. Agostoni C, Manzoni P. Nutrition and neurocognitive development. *Early Hum Dev* 2013;89 Suppl 1:S1-3.
[PUBMED](#) | [CROSSREF](#)
2. Prado EL, Dewey KG. Nutrition and brain development in early life. *Nutr Rev* 2014;72:267-84.
[PUBMED](#) | [CROSSREF](#)
3. Grantham-McGregor S, Cheung YB, Cueto S, Glewwe P, Richter L, Strupp B; International Child Development Steering Group. Developmental potential in the first 5 years for children in developing countries. *Lancet* 2007;369:60-70.
[PUBMED](#) | [CROSSREF](#)
4. Black MM. Micronutrient deficiencies and cognitive functioning. *J Nutr* 2003;133:3927S-31S.
[PUBMED](#) | [CROSSREF](#)
5. Uauy R, Dangour AD. Nutrition in brain development and aging: role of essential fatty acids. *Nutr Rev* 2006;64:S24-33.
[PUBMED](#) | [CROSSREF](#)
6. Ulmer S, Backens M, Ahlhelm FJ. Basic principles and clinical applications of magnetic resonance spectroscopy in neuroradiology. *J Comput Assist Tomogr* 2016;40:1-13.
[PUBMED](#) | [CROSSREF](#)

7. World Health Organization. Severe acute malnutrition [Internet]. Geneva: World Health Organization; 2009 [cited 2017 Dec 10]. Available from: <http://www.who.int/nutrition/topics/malnutrition/en/>.
8. Erol N, Sezgin N, Savasir I. Validity studies for development-screening inventory. *Turk J Psychol* 1993;8:16-22.
9. Savasir I, Sezgin N, Erol N. Handbook of Ankara development-screening inventory. 2nd ed. Ankara: Rekmay; 1998.
10. Hamaguchi H, Cleve H. Solubilization of human erythrocyte membrane glycoproteins and separation of the MN glycoprotein from a glycoprotein with I, S, and A activity. *Biochim Biophys Acta* 1972;278:271-80.
[PUBMED](#) | [CROSSREF](#)
11. Sattler W, Puhl H, Hayn M, Kostner GM, Esterbauer H. Determination of fatty acids in the main lipoprotein classes by capillary gas chromatography: BF₃/methanol transesterification of lyophilized samples instead of Folch extraction gives higher yields. *Anal Biochem* 1991;198:184-90.
[PUBMED](#) | [CROSSREF](#)
12. Granados-Rojas L, Serrano N, Gutiérrez-Ospina G, Díaz-Cintra S. Prenatal protein malnutrition differentially affects the volume of the granule layer and mossy fibers in young male and female rats. *Proc West Pharmacol Soc* 2002;45:53-4.
[PUBMED](#)
13. Mathangi DC, Namasivayam A. Effect of chronic protein restriction on motor co-ordination and brain neurotransmitters in albino rats. *Food Chem Toxicol* 2001;39:1039-43.
[PUBMED](#) | [CROSSREF](#)
14. Alamy M, Bengelloun WA. Malnutrition and brain development: an analysis of the effects of inadequate diet during different stages of life in rat. *Neurosci Biobehav Rev* 2012;36:1463-80.
[PUBMED](#) | [CROSSREF](#)
15. Benítez-Bribiesca L, De la Rosa-Alvarez I, Mansilla-Olivares A. Dendritic spine pathology in infants with severe protein-calorie malnutrition. *Pediatrics* 1999;104:e21.
[PUBMED](#) | [CROSSREF](#)
16. Nyaradi A, Li J, Hickling S, Foster J, Oddy WH. The role of nutrition in children's neurocognitive development, from pregnancy through childhood. *Front Hum Neurosci* 2013;7:97.
[PUBMED](#) | [CROSSREF](#)
17. Engle PL, Fernández PD. INCAP studies of malnutrition and cognitive behavior. *Food Nutr Bull* 2010;31:83-94.
[PUBMED](#) | [CROSSREF](#)
18. Isik Y, Kalyoncu M, Okten A. Serum leptin levels in marasmic children and the relationship between leptin and lipid profile. *Ann Nutr Metab* 2004;48:259-62.
[PUBMED](#) | [CROSSREF](#)
19. Akuyam A, Isah HS, Ogala WN. Serum lipid profile in malnourished Nigerian children in Zaria. *Niger Postgrad Med J* 2008;15:192-6.
[PUBMED](#)
20. Fageer SA, Karar TA, Hag SM. Assessment of plasma levels of proteins, total cholesterol and high density. *J Sci Technol* 2013;14:35-43.
21. Veiga GR, Ferreira HS, Sawaya AL, Calado J, Florêncio TM. Dyslipidaemia and undernutrition in children from impoverished areas of Maceió, state of Alagoas, Brazil. *Int J Environ Res Public Health* 2010;7:4139-51.
[PUBMED](#) | [CROSSREF](#)
22. Chisti MJ, Salam MA, Ashraf H, Faruque AS, Bardhan PK, Shahid AS, et al. Prevalence, clinical predictors, and outcome of hypocalcaemia in severely-malnourished under-five children admitted to an urban hospital in Bangladesh: a case-control study. *J Health Popul Nutr* 2014;32:270-5.
[PUBMED](#)
23. Singla PN, Chand P, Kumar A, Kachhawaha JS. Serum magnesium levels in protein-energy malnutrition. *J Trop Pediatr* 1998;44:117-9.
[PUBMED](#) | [CROSSREF](#)
24. El-khayat H, Shaaban S, Emam EK, Elwakkad A. Cognitive functions in protein-energy malnutrition: in relation to long chain-polyunsaturated fatty acids. *Pak J Biol Sci* 2007;10:1773-81.
[PUBMED](#) | [CROSSREF](#)
25. Gil A, Ramirez M, Gil M. Role of long-chain polyunsaturated fatty acids in infant nutrition. *Eur J Clin Nutr* 2003;57 Suppl 1:S31-4.
[PUBMED](#) | [CROSSREF](#)
26. Schulzke SM, Patole SK, Simmer K. Long-chain polyunsaturated fatty acid supplementation in preterm infants. *Cochrane Database Syst Rev* 2011:CD000375.
[PUBMED](#)

27. Smithers LG, Gibson RA, Makrides M. Maternal supplementation with docosahexaenoic acid during pregnancy does not affect early visual development in the infant: a randomized controlled trial. *Am J Clin Nutr* 2011;93:1293-9.
[PUBMED](#) | [CROSSREF](#)
28. McCann JC, Ames BN. Is docosahexaenoic acid, an n-3 long-chain polyunsaturated fatty acid, required for development of normal brain function? An overview of evidence from cognitive and behavioral tests in humans and animals. *Am J Clin Nutr* 2005;82:281-95.
[PUBMED](#) | [CROSSREF](#)
29. Simmer K, Patole SK, Rao SC. Long-chain polyunsaturated fatty acid supplementation in infants born at term. *Cochrane Database Syst Rev* 2008:CD000376.
[PUBMED](#)
30. Kendall GS, Melbourne A, Johnson S, Price D, Bainbridge A, Gunny R, et al. White matter NAA/Cho and Cho/Cr ratios at MR spectroscopy are predictive of motor outcome in preterm infants. *Radiology* 2014;271:230-8.
[PUBMED](#) | [CROSSREF](#)
31. Guo L, Wang D, Bo G, Zhang H, Tao W, Shi Y. Early identification of hypoxic-ischemic encephalopathy by combination of magnetic resonance (MR) imaging and proton MR spectroscopy. *Exp Ther Med* 2016;12:2835-42.
[PUBMED](#) | [CROSSREF](#)
32. Nassar M, Shaaban S, Abbas Y, Galal A, Naguib A, Kamal El-Deen A. Proton Magnetic Resonance Spectroscopy in protein energy malnutrition patients. *J Pediatr Gastroenterol Nutr* 2011;52 Suppl 1:e63.
33. Filippi CG, Uluğ AM, Deck MD, Zimmerman RD, Heier LA. Developmental delay in children: assessment with proton MR spectroscopy. *AJNR Am J Neuroradiol* 2002;23:882-8.
[PUBMED](#)
34. Hart PH, Lucas RM, Walsh JP, Zosky GR, Whitehouse AJ, Zhu K, et al. Vitamin D in fetal development: findings from a birth cohort study. *Pediatrics* 2015;135:e167-73.
[PUBMED](#) | [CROSSREF](#)
35. Schlögl M, Holick MF. Vitamin D and neurocognitive function. *Clin Interv Aging* 2014;9:559-68.
[PUBMED](#)