



# **Nutritional Indices for Assessing Fatty Acids: A Mini-Review**

Jiapeng Chen<sup>1</sup> and Hongbing Liu<sup>1,2,\*</sup>

- Key Laboratory of Marine Drugs, Chinese Ministry of Education, School of Medicine and Pharmacy, Ocean University of China, Qingdao 266003, China; 21180831069@stu.ouc.edu.cn
- <sup>2</sup> Laboratory for Marine Drugs and Bioproducts, Pilot National Laboratory for Marine Science and Technology (Qingdao), Qingdao 266237, China
- \* Correspondence: liuhongb@ouc.edu.cn; Tel.: +86-0532-82031823

Received: 30 June 2020; Accepted: 6 August 2020; Published: 8 August 2020



Abstract: Dietary fats are generally fatty acids that may play positive or negative roles in the prevention and treatment of diseases. In nature, fatty acids occur in the form of mixtures of saturated fatty acid (SFA), monounsaturated fatty acid (MUFA), and polyunsaturated fatty acid (PUFA), so their nutritional and/or medicinal values must be determined. Herein, we do not consider the classic indices, such as  $\sum$ SFA,  $\sum$ MUFA,  $\sum$ PUFA,  $\sum$ n-6 PUFA,  $\sum$ n-3 PUFA, and n-6 PUFA/n-3 PUFA; instead, we summarize and review the definitions, implications, and applications of indices used in recent years, including the PUFA/SFA, index of atherogenicity (IA), the index of thrombogenicity (IT), the hypocholesterolemic/hypercholesterolemic ratio (HH), the health-promoting index (HPI), the unsaturation index (UI), the sum of eicosapentaenoic acid and docosahexaenoic acid (EPA + DHA), fish lipid quality/flesh lipid quality (FLQ), the linoleic acid/ $\alpha$ -linolenic acid (LA/ALA) ratio, and *trans* fatty acid (TFA). Of these nutritional indices, IA and IT are the most commonly used to assess the composition of fatty acids as they outline significant implications and provide clear evidence. EPA + DHA is commonly used to assess the nutritional quality of marine animal products. All indices have their advantages and disadvantages; hence, a rational choice of which to use is critical.

Keywords: fatty acids; nutritional indices; human health

# 1. Introduction

Fatty acids (FAs) are organic acids with at least one carboxyl (-C(=O)OH, -COOH, or  $-CO_2H$ ) group and a long carbon chain whose links can be double bonds, as in unsaturated fatty acids, or single bonds, as in saturated fatty acids. FAs are generally derived from triglycerides and phospholipids, and are the main components of dietary fats. Most naturally occurring FAs have an unbranched chain of an even number (4–28) of carbon atoms. According to the number of double bonds, the FA catalogue includes saturated fatty acids (SFAs), monounsaturated fatty acid (MUFA), and polyunsaturated fatty acid (PUFA).

FAs are distributed to cells where they serve as fuel for muscular contraction and general metabolism. As biological compounds, FAs play critical roles in human metabolism, health, and disease. Epidemiological studies and clinical trials showed that fatty acids are associated with cardiovascular diseases [1–5], neurological diseases [6–9], non-alcoholic fatty liver disease [10–13], allergic diseases [14–16], and so on. Evidence from metabolomics experiments indicates that they participate in the metabolic pathways of related diseases [8,17–26]. For example, the free FA profile was found to be altered in both leukemia and pre-leukemic diseases, particularly C14:0, C16:0, and C18:0 [26].

FAs play positive or negative roles in the prevention and treatment of diseases. For example, SFAs may increase the risk of developing multiple sclerosis (MS) as well as disease progression,

whereas PUFAs may have beneficial effects in MS patients [7]. As another example, some essential FA metabolites may exert health effects such as anti-inflammatory and neuroprotection effects, but they can also produce negative effects such as inflammation, necrosis promoters, and atherosclerosis. In general, FAs are obtained from various dietary sources that possess characteristic FA composition and consequently influence health outcome. From this perspective, the FA composition should be assessed to determine their nutritional and/or medicinal value, especially in fatty-acid-rich foods, food supplements, and herb-based medicines.

In this mini-review, we collated the literature related to fatty acid profile analysis that was published in recent decades since 2000 to understand the implications and applications of various nutritional indices. We did not consider the classic indices such as  $\sum$ SFA,  $\sum$ MUFA,  $\sum$ PUFA,  $\sum$ n-6 PUFA,  $\sum$ n-3 PUFA, and n-6 PUFA/n-3 PUFA. The present review may help researchers to evaluate the nutritional value of fatty acids and to explore their potential usage in disease prevention and treatment. It may also help newcomers to the field of fatty acid profile analysis to quickly and accurately select appropriate indices.

### 2. Nutritional Indices

In this review, we screened articles and summarized the nutritional indices. The results are shown in Table 1.

No.	Index	Full Name	Calculation Formula	Application
1	PUFA/SFA	Polyunsaturated fatty acid/saturated fatty acid ratio	ΣPUFA/ΣSFA	Seaweeds [27–29], crops [30,31], plant oil [32,33], shellfish [34], fish [34–40], meat [41–53], and dairy products [54–57]
2	ΙΑ	Index of atherogenicity	[C12:0 + (4 × C14:0) + C16:0]/ΣUFA	Seaweeds [27–29,58,59], crops [30,31,60,61], plant oil [33,62], shellfish [63], shrimp [64], fish [36–39,65–73], meat [41–43,48–50,52,53,74–77], and dairy products [54–56,78–89]
3	IT	Index of thrombogenicity	$\begin{array}{l} (C14:0 + C16:0 + C18:0)/[(0.5 \\ \times \Sigma MUFA) + (0.5 \times \Sigma n\text{-}6 \ PUFA) + (3 \\ \times \Sigma n\text{-}3 \ PUFA) + (n\text{-}3/n\text{-}6)] \end{array}$	Seaweeds [27–29,58,59], crops [30,31,60,61], plant oil [62], shellfish [63], shrimp [64], fish [36–39,65–68,70–73], meat [43,48,50,52,53,75,77], and dairy products [54,55,78,80,86–89]
4	НН	Hypocholesterolemic /hypercholesterolemic ratio	(cis-C18:1 + ΣPUFA)/(C12:0 + C14:0 + C16:0)	Seaweeds [90], plant oil [62], shellfish [34], fish [34,36–39,72], meat [46–48,52,77,91], and dairy products [54,55,78,86,87]
5	HPI	Health-promoting index	$\Sigma UFA/[C12:0+(4 \times C14:0) + C16:0]$	Milk [92-94] and cheese [57,94,95]
6	UI	Unsaturation index	$1 \times (\% \text{ monoenoics}) + 2 \times (\% \text{ dienoics}) + 3 \times (\% \text{ trienoics}) + 4 \times (\% \text{ tetraenoics}) + 5 \times (\% \text{ pentaenoics}) + 6 \times (\% \text{ hexaenoics})$	Seaweeds [27–29,59,96–98], crops [61,99,100], meat [44,101], and milk [102]
7	EPA + DHA	Sum of eicosapentaenoic acid and docosahexaenoic acid	C22:6 n-3 + C20:5 n-3	Shellfish [34] and fish [34,36,37,40,68,103-106]
8	FLQ	Fish lipid quality/flesh Lipid quality	100 × (C22:6 n-3 + C20:5 n-3)/ΣFA	Fish [65,66,73,107,108]
9	LA/ALA	Linoleic acid /α-linolenic acid ratio	C18:2 n-6/C18:3 n-3	Lamb [43] and milk [55,109]
10	TFA	Trans fatty acid	ΣΤΓΑ	Seaweeds [90], plant oil [32,33,110], fish [35], lamb [45], and milk [78]

#### Table 1. Summary of nutritional indices.

## 2.1. Polyunsaturated Fatty Acid/Saturated Fatty Acid (PUFA/SFA)

PUFA/SFA is an index normally used to assess the impact of diet on cardiovascular health (CVH). It hypothesizes that all PUFAs in the diet can depress low-density lipoprotein cholesterol (LDL-C) and lower levels of serum cholesterol, whereas all SFAs contribute to high levels of serum cholesterol. Thus, the higher this ratio, the more positive the effect.

All that was missing was MUFA. According to Dietschy's study on dietary FAs and their regulation of plasma LDL-C concentrations in 1998, C18:1 n-9 *cis* (oleic acid), the most common MUFA in dietary food increases the activity of low-density lipoprotein receptors (LDLRs) and decreases the cholesterol concentration in serum [111]. Not all molecular species of SFAs contribute equally to serum cholesterol. C12:0, C14:0, and C16:0 can increase the cholesterol concentration in serum by inhibiting the activity of

LDLRs; C4:0, C6:0, C8:0, and C10:0 were rapidly oxidized to acetyl-CoA in the liver and could not affect the activity of LDLRs, and C18:0 appeared to be biologically neutral and have no effect on circulating LDL-C levels [111].

Notably, not all of the main classes of PUFA positively affect the prevention of cardiovascular disease (CVD). Short-term supplementation with docosahexaenoic acid (DHA)-rich fish oil may modulate the activity of peroxisome proliferators-activated receptor-gamma (PPAR- $\gamma$ ) to protect the cardiovascular system from the unhealthy effects of atherosclerotic lesions [4]. Recent clinical trials support the view that supplementation with eicosapentaenoic acid (EPA) can reduce plasma triglyceride (TG) levels and activate anti-inflammatory, anti-thrombotic, and other mechanisms to prevent atherosclerosis (AS) [2]. However, a narrative review that collated the available data showed that dietary intake of linoleic acid (LA, C18:2 n-6) is inversely correlated with CVD; however, further research is needed to clarify the underlying mechanisms [3].

PUFA/SFA is the most commonly used index for evaluating the nutritional value of dietary foods such as of seaweed (0.42–2.12, except for *Gracilaria changii*), meat (0.11–2.042), fish (0.50–1.62), shellfish (0.20–2.10), and dietary products (0.02–0.175). Chan and Matanjun determined the FA profiles of red seaweed *Gracilaria changii* a mangrove area of Malaysia, and used PUFA/SFA to assess the nutritional quality, finding a value of  $6.96 \pm 0.98$  [28]. PUFA/SFA of chicken is in the range of 0.308 to 2.042 for different dietary treatments [48]. Fernandes et al. compared the FA profile of four species of Brazilian fish, and they used the PUFA/SFA as one of the nutritional quality indices, reporting values between 1.09 to 1.47 [36]. Detailed information about the literature related to PUFA/SFA is shown in Table 2.

Materials		PUFA/SFA Value	Reference
Red seaweed	Amphiora anceps	0.42	[27]
	Kappaphycus alvarezii	0.57	[27]
	Gelidiella acerosa	0.84	[27]
	Gelidium micropterum	0.30	[27]
	Gracilaria changii	$6.96 \pm 0.98$	[28]
	Gracilaria corticata	2.12	[27]
	Gracilaria dura	1.89	[27]
	Gracilaria debilis	1.17	[27]
	Gracilaria fergusonii	0.58	[27]
	Gracilaria salicornia	0.14	[27]
	Laurencia cruciata	0.79	[27]
	Sarconema filiforme	1.71	[27]
Brown seaweed	Cystoseira indica	1.17	[27]
	Padina tetrastromatica	0.85	[27]
	Sargassum fusiforme	$0.67 \pm 0.31$	[29]
	Sargassum horneri	$0.56 \pm 0.06$	[29]
	Sargassum pallidum	$0.20 \pm 0.09$	[29]
	Sargassum swartzii	1.15	[27]
	Sargassum tenerrimum	1.18	[27]
	Sargassum thunbergii	$0.39 \pm 0.05$	[29]
	Spatoglossum asperum	1.38	[27]
Green seaweed	Caulerpa racemosa	0.44	[27]
	Caulerpa scalpeliformis	0.88	[27]
	Caulerpa veravalnensis	0.73	[27]
	Ulva fasciata	0.42	[27]
	Ulva reticulata	0.23	[27]
	Ulva rigida	0.33	[27]
Crops	Cyamopsis tetragonolobaL.	1.71	[31]
	Lupinus albus	1.53–1.97	[30]
Plant oil	Palm stearin	0.13	[32]
	Sunflower oil	4.75–4.94	[32,33]

Table 2. Application of PUFA/SFA in fatty acid evaluation \*.

Materials		PUFA/SFA Value	Reference
Shellfish	Cancer edwardsi	2.10	[34]
	Cervimunida johni	1.81	[34]
	Concholepas concholepas	1.16	[34]
	Heterocarpus reedi	1.47	[34]
	Loxechinus albus	0.20	[34]
	Mesodesma donacium	1.34	[34]
	Pleuroncodes monodon	1.68	[34]
	Pyura chilensis	1.31	[34]
	Venus antiqua	1.06	[34]
Fish	Carassius gibelio	1.62-1.70	[35]
	Cilus gilberti	1.15	[34]
	Genypterus chilensis	1.60	[34]
	Hemiramphus brasiliensis	1.09	[36]
	Hyporhamphus unifasciatus	1.11	[36]
	Kutum roach	1.02-1.79	[37]
	Lagocephalus guentheri	1.3	[38]
	Merluccius gayi	1.52	[34]
	Opisthonema oglinum	1.47	[36]
	Orechromis niloticus	0.51-0.56	[39]
	Pinguipes chilensis	0.80	[34]
	Scomber japonicus	0.92	[34]
	Scomberomorus cavalla	1.18	[36]
	Seriola lalandi	0.92	[34]
	Seriolella violacea	0.95	[34]
	Trachinotus carolinus	0.5-1.1	[40]
	Trachurus murphyi	0.95	[34]
Meat	Chicken (Caribro Vishal)	0.308-2.042	[50]
	Chicken (purchased from a hatchery and poultry farm)	0.926-0.945	[48]
	Pig (DanBred $\times$ PIC terminal line)	0.46-0.48	[49]
	Pig (Pietrain × (Duroc × Landrace))	0.85-1.29	[44]
	Lamb (Barbarine lamb)	0.13-0.37	[43,45]
	Steer (Blonded Aquitaine steer)	0.29-0.58	[42]
	Calve (75% Charolais breeds)	0.13-0.34	[51]
	Cattle (Nellore cattle)	0.11-0.20	[46]
	Yak (Phoephagus grunniens)	0.37-0.55	[41]
	Foal (Galician Mountain × Hispano-Bretón)	0.44-1.06	[52]
	Spanish dry-cured ham	0.19-0.30	[47]
	Bologna sausages	0.27-1.17	[53]
Dairy products	Cheese of Comisana ewe	0.086-0.173	[57]
	Milk of Chios sheep	0.06-0.08	[54]
	Milk of Karagouniko sheep	0.06-0.09	[54]
	Milk of Turcana dairy ewe	0.106-0.175	[55]
	Milk of Friesian × Jersey cow	0.02-0.04	[56]

Table 2. Cont.

PUFA/SFA: polyunsaturated fatty acid/saturated fatty acid ratio; \* literature from 2000 until April/2020.

## 2.2. Index of Atherogenicity (IA)

The index of atherogenicity (IA) was developed by Ulbritcht and Southgate in 1991, and characterizes the atherogenic potential of FA [112]. As the PUFA/SFA ratio is too general and unsuitable for assessing the atherogenicity of foods, Ulbritcht and Southgate proposed a new index, IA, based on PUFA/SFA considering the available evidence, and then checked whether the resulting values were in accordance. The formula for calculating IA is:

$$IA = [C12:0 + (4 \times C14:0) + C16:0] / \Sigma UFA$$
(1)

The IA indicates the relationship between the sum of SFAs and the sum of unsaturated fatty acids (UFAs). The main classes of SFAs, which include C12:0, C14:0, and C16:0, with the exception of C18:0, are considered pro-atherogenic (they favor the adhesion of lipids to cells of the circulatory and immunological systems) [67,68,113]. UFAs are considered to be anti-atherogenic as they inhibit

the accumulation of plaque and reduce the levels of phospholipids, cholesterol, and esterified fatty acids [68,113]. Therefore, the consumption of foods or products with a lower IA can reduce the levels of total cholesterol and LDL-C in human blood plasma [85].

Although the IA is more reasonable than the simple PUFA/SFA ratio for assessing the degree of atherogenicity, there are still some imperfections in the proposed IA formula, which were pointed out by Ulbritcht and Southgate. First, stearic acid (C18:0) should appear in the denominator if sufficient evidence shows that it can reduce the level of LDL-C in human blood plasma in the future. Second, not all PUFAs should be weighed equally. Third, the impact of *trans* fatty acids was not considered due to conflicting evidence [112].

The IA has been used widely for evaluating seaweeds, crops, meat, fish, dairy products, etc. Nantapo et al. analyzed the fatty acid composition of milk during different stages of lactation and found that milk with lower IA is important, and the IA ranges from 4.08 to 5.13 in different stages of lactation [56]. Akintola investigated the techniques of smoking and sun drying to understand the nutritional quality of southern pink shrimp (*Penaeus notialis*) using the IA as an index, and reported values of 0.71 to 0.82 [64].

Detailed information about the literature related to the IA is shown in Table 3. For seaweeds, the species may be the main factor influencing the IA value, which ranges from 0.03 to 3.58. The value ranges from 0.084 to 0.55 for crops, 0.21 to 1.41 for fish, and 0.165 to 1.32 for meat. For dairy products, the value ranges from 1.42 to 5.13. For ruminants, dietary treatment is the main factor influencing the IA.

Materials		IA Value	Reference
Red seaweed	Amphiora anceps	1.52	[27]
	Ceramium virgatum	$0.37 \pm 0.027$	[59]
	Corallina officinalis	$0.48 \pm 0.039$	[59]
	Gelidiella acerosa	0.80	[27]
	Gelidium micropterum	1.61	[27]
	Gracilaria changii	$0.03 \pm 0.003$	[28]
	Gracilaria corticata	0.38	[27]
	Gracilaria debilis	0.69	[27]
	Gracilaria dura	0.45	[27]
	Gracilaria fergusonii	1.34	[27]
	Gracilaria salicornia	2.87	[27]
	<i>Hymenena</i> sp.	3.58	[59]
	Kappaphycus alvarezii	0.77	[27]
	Laurencia cruciata	0.84	[27]
	Lomentaria clavellosa	$3.06 \pm 0.611$	[59]
	Polysiphonia sp.	$1.35 \pm 0.206$	[59]
	Sarconema filiforme	0.49	[27]
Brown seaweed	Cystoseira indica	0.66	[27]
	Dictyota dichotoma	$0.29 \pm 0.041$	[59]
	Laminaria ochroleuca	1.18–1.57	[58]
	Leathesia difformis	$0.48 \pm 0.021$	[59]
	Myriogloea major	$0.21 \pm 0.019$	[59]
	Padina tetrastromatica	0.81	[27]
	Sargassum fusiforme	$0.94 \pm 0.28$	[29]
	Sargassum horneri	$1.06 \pm 0.06$	[29]
	Sargassum pallidum	$1.99 \pm 0.45$	[29]
	Sargassum swartzii	0.61	[27]
	Sargassum tenerrimum	0.66	[27]
	Sargassum thunbergii	$1.16 \pm 0.10$	[29]
	Spatoglossum asperum	0.53	[27]
	Ündaria pinnatifida	0.17–0.35	[59]

Table 3. Application of IA in fatty acid evaluation \*.

Materials		IA Value	Reference
Green seaweed	Caulerpa racemosa	1.61	[27]
	Caulerpa scalpeliformis	0.86	[27]
	Caulerpa veravalnensis	1.17	[27]
	Cladophora falklandica	$0.50 \pm 0.062$	[59]
	Codium decorticatum	$0.22 \pm 0.002$	[59]
	Codium fragile	$0.29 \pm 0.020$	[59]
	Codium vermilara	$0.40 \pm 0.086$	[59]
	Ulva fasciata	1.37	[27]
	Ulva reticulata	1.54	[27]
	Ulva rigida	1.22	[27]
	Ulva sp.1	$0.20 \pm 0.055$	[59]
	Ulva sp.2	$0.08 \pm 0.004$	[59]
Crops	Cumin ( <i>Cuminum cyminum</i> )	0.46-0.53	[61] <sup>a</sup>
1	Guar seed (Cyamopsis tetragonoloba)	0.22	[31]
	White lupine (Lupinus albus)	0.084-0.107	[30]
	Scabiosa stellata	0.55	[60]
Plant oil	Camelina oil ( <i>Camelina sativa</i> )	0.05-0.07	[62]
	Sunflower oil	0.09-0.11	[33]
Shellfish	Chlamys farreri	0.31-0.37	[63]
	Patinopecten yessoensis	0.29-0.35	[63]
Shrimp	Penaeus notialis	0.71-0.82	[64]
Fish	Abramis brama	0.37-0.42	[65,66]
	Clupea harengus	$0.70 \pm 0.10$	[66]
	Cynoscion parvipinnis	1.07-1.16	[67]
	Cyprinus carpio	$0.36 \pm 0.03$	[66]
	Dicentrarchus labrax	0.40-0.42	[68]
	Esox lucius	0.43	[65]
	Hemiramphus brasiliensis	0.26	[36]
	Hyporhamphus unifasciatus	0.26	[36]
	Kutum roach	0.58-1.41	[37]
	Lagocephalus guentheri	0.43	[38]
	Leuciscus idus	$0.36 \pm 0.02$	[66]
	Limousin steers	0.70-1.14	[69]
	Micronterus salmoides	0.29-0.68	[70]
	Mugil cephalus	0.91-1.22	[70]
	Oncorhunchus mukiss	$0.33 \pm 0.01$	[66]
	Onisthonema oplinum	0.60	[36]
	Oreochromis viloticus	0.55-0.60	[39]
	Perca flumiatilis	0.37-0.44	[65 66]
	Platichthus flesus	$0.07 \ 0.11$ $0.41 \pm 0.03$	[66]
	Rutilus rutilus	0.41 ± 0.00	[65]
	Salmo trutta	0.40	[00]
	Suino truttu Scomboromorus cazalla	0.04-0.72	[72]
	Scomberomorus cuounu Sparus aurata	0.48	[36]
Maak	Chicken (Caribre Vishal)	0.165.0.624	[50]
Meat	Chicken (Caribro Visital) Chicken (nurchased from a batchery and neultry form)	0.103 - 0.034 0.272 0.200	[30]
	Respire (Currenter a lance)	0.572-0.590	[40]
	Dia (Dan Brad ) ( DIC torminal line)	0.35-0.69	[73]
	Level (Danbied × Fic terminal line)	0.27-0.31	[49]
	Lamb (Cartile di Dualia / Samana)	0.49-0.52	[43]
	Lamb (Gentile di Fuglia × Sopravissana)	0.99-1.32	[76]
	Lamb (lie de France × Pagliarola)	0.71-1.06	[76]
	Lamb (Iranian fat-tailed breed)	0.53-0.77	[74]
	Heiter (Limousin heiter)	0.50-0.57	[77]
	Steer (Blonded Aquitaine steer)	0.51-0.63	[42]
	Yak (Phoephagus grunniens)	0.37-0.43	[41]
	Foal (Galician Mountain × Hispano-Bretón)	0.59–0.62	[52]
	Bologna sausages	0.33-0.60	[53]

# Table 3. Cont.

Materials		IA Value	Reference
Dairy products	Cheese of Churra ewe	1.61-3.61	[79]
	Cheese of Holstein cow	2.38-3.72	[88]
	Cheese of Italian Friesian and Italian Red Pied cattle (Caciocavallo cheese)	2.43-2.94	[84]
	Curd of cow (Middle Rhodopes)	1.94-5.02	[78]
	Milk of Anglo-Nubian goat	1.89-2.48	[81]
	Milk of goat (market of Sardinia)	2.27-2.91	[89]
	Milk of Nubian goat	1.91-2.32	[82]
	Milk of Saanen goat	$2.77\pm0.08$	[85]
	Milk of Swedish Landrace goat	$2.47 \pm 0.07$	[85]
	Milk of Chios sheep	2.00-2.72	[54]
	Milk of Karagouniko sheep	1.76-2.57	[54]
	Milk of Churra ewe	1.71-3.39	[79]
	Milk of Lacaune ewe	1.94-2.53	[80]
	Milk of Turcana dairy ewe	1.42-1.95	[55]
	Milk of cow (Middle Rhodopes)	1.88-4.18	[78]
	Milk of Friesian × Jersey cow	4.08-5.13	[56]
	Milk of Holstein cow	1.83-2.63	[88]
	Milk of Holstein–Friesian cow	1.60-3.79	[83,109] <sup>a</sup>
	Milk of indigenous Indian cow	1.37	[109] <sup>a</sup>
	Milk of Jersey cow	2.4823-3.4360	[87]
	Milk of Sahiwal cow	2.01	[109] <sup>a</sup>
	Milk of Sahiwal × Holstein–Friesian cow	3.14	[109] <sup>a</sup>
	Milk of Italian Friesian and Italian Red Pied cattle	2.49-2.99	[84]
	Yogurt of cow milk (market of Faisalabad)	1.48-2.74	[86]
	Yogurt of sheep milk (market of Faisalabad)	1.42-2.31	[86]

Table 3. Cont.

IA: index of atherogenicity; \* literature from 2000 until April/2020; <sup>a</sup> recalculated according to the original data in the reference.

### 2.3. Index of Thrombogenicity (IT)

The index of thrombogenicity (IT) was developed by Ulbritcht and Southgate [112] together with IA in 1991. The formula is:

$$IT = (C14:0 + C16:0 + C18:0) / [(0.5 \times \Sigma MUFA) + (0.5 \times \Sigma n - 6 PUFA) + (3 \times \Sigma n - 3 PUFA) + (n - 3 / n - 6)]$$
(2)

The IT characterizes the thrombogenic potential of FAs, indicating the tendency to form clots in blood vessels and provides the contribution of different FAs, which denotes the relationship between the pro-thrombogenic FAs (C12:0, C14:0, and C16:0) and the anti-thrombogenic FAs (MUFAs and the n-3 and n-6 families) [112]. Therefore, the consumption of foods or products with a lower IT is beneficial for CVH. The IT has been used in many fatty acid composition studies to assess the degree of thrombogenicity. As with the IA formula, the proposed IT formula should be modified as our understanding of MUFA and *trans* fatty acids increases.

The IT has been used in many FA composition studies to assess the degree of thrombogenicity. Chen et al. conducted comparative studies on the fatty acid profiles of four different Chinese medicinal *Sargassum* seaweeds, where the IT was used as one of the nutritional indices to evaluate the potential effects of four *Sargassum* on CVH. The results showed that the IT was between 0.46 and 1.60 [29]. Calabrò et al. compared the fatty acid profile of three cultivars of *Lupinus albus* (Lutteur, Lublanca, and Multitalia) and the IT was used due to the correlation between fatty acids and human health [30].

Detailed information of the literature related to the IT is provided in Table 4. For seaweeds, the value ranges from 0.04 to 2.94 with the exception of *Gracilaria salicornia*, which had an IT value of 5.75 [27]. The ranges of IT values for crops, fish, meat, and dairy products are 0.139–0.56, 0.14–0.87, 0.288–1.694, and 0.39–5.04, respectively.

In brief, both the IA and the IT can be used to assess the potential effects of FA composition on CVH. A FA composition with a lower IA and IT has a better nutritional quality, and its consumption

may reduce the risk of coronary heart disease (CHD), but no organization has yet provided the recommended values for the IA and IT. As our comprehensive understanding of the function of FA molecular species deepens, the accuracies of the IA and IT formulas are expected to increase, which might be modified by taking advantage of the massive amount of available data and advanced computer technology.

Materials		IA Value	Reference
Red seaweed	Amphiora anceps	2.07	[27]
fied seaffeed	Ceramium virgatum	$0.12 \pm 0.005$	[59]
	Corallina officinalis	$0.28 \pm 0.045$	[59]
	Gelidiella acerosa	0.52	[27]
	Gelidium micronterum	1.83	[27]
	Gracilaria chanoii	$0.04 \pm 0.01$	[28]
	Gracilaria corticata	0.63	[27]
	Gracilaria dehilis	1 25	[27]
	Gracilaria dura	0.88	[27]
	Gracilaria ferousonii	2.66	[27]
	Cracilaria salicornia	5.75	[27]
	Humanana sp	2.66	[27]
	Kamanhucus alvarazii	2.00	[39]
	Lauronoia orugiata	1.17	[27]
		0.71	[27]
		$2.94 \pm 1.000$	[39]
	Polysipnoniu sp	$0.61 \pm 0.114$	[39]
	Surconemu futforme	0.55	[27]
Brown seaweed	Cystoseira indica	0.87	[27]
	Dictyota dichotoma	$0.09 \pm 0.013$	[59]
	Laminaria ochroleuca	1.06-1.89	[58]
	Leathesia difformis	$0.14 \pm 0.006$	[59]
	Myriogloea major	$0.09 \pm 0.006$	[59]
	Padina tetrastromatica	1.20	[27]
	Sargassum fusiforme	$0.46 \pm 0.21$	[29]
	Sargassum horneri	$0.65 \pm 0.07$	[29]
	Sargassum pallidum	$1.60 \pm 0.56$	[29]
	Sargassum swartzii	0.75	[27]
	Sargassum tenerrimum	0.90	[27]
	Sargassum thunbergii	$0.76 \pm 0.14$	[29]
	Spatoglossum asperum	0.50	[27]
	Undaria pinnatifida	0.08-0.26	[59]
Green seaweed	Caulerpa racemosa	1.50	[27]
	Caulerpa scalpeliformis	1.38	[27]
	Caulerpa veravalnensis	1.28	[27]
	Cladophora falklandica	$0.16 \pm 0.048$	[59]
	Codium decorticatum	$0.12 \pm 0.002$	[59]
	Codium fragile	$0.14 \pm 0.013$	[59]
	Codium vermilara	$0.30 \pm 0.080$	[59]
	Ulva fasciata	1.56	[27]
	Ulva reticulata	2.90	[27]
	Ulva rigida	1.78	[27]
	Ulva sp.1	$0.09 \pm 0.028$	[59]
	Ulva sp.2	$0.04 \pm 0.002$	[59]
Crops	Cumin (Cuminum cuminum)	0.46-0.56	[61] a
crops	Guar sood (Cuamonsis tetragonoloha)	0.40-0.50	[01]
	Scabioca stallata	0.33	[51]
	White lupine (Luninus albus)	0.139-0.180	[30]
Plant oil	Camelina oil ( <i>Camelina sativa</i> )	0.105 0.100	[62]
Challfick	Chlamus farrori	0.12 0.17	[62]
Snellfisn	Patinopecten vessoensis	0.13-0.17 0.09-0.15	[63]
Shrimp	Penaeus notialis	0.21_0.30	[64]
r	1 0.000 HOFMHO	0.21 0.00	

Table 4. Application of IT in fatty acid evaluation \*.

Materials		IA Value	Reference
Fish	Abramis brama	0.23-0.24	[65,66]
	Clupea harengus	$0.26 \pm 0.04$	[66]
	Cynoscion parvipinnis	0.18-0.29	[67]
	Cyprinus carpio	$0.31 \pm 0.03$	[66]
	Dicentrarchus labrax	0.191-0.63	[68]
	Esox lucius	0.18	[65]
	Hemiramphus brasiliensis	0.21	[36]
	Hyporhamphus unifasciatus	0.44	[36]
	Kutum roach	0.16-0.24	[37]
	Lagocephalus guentheri	0.29	[38]
	Leuciscus idus	$0.22 \pm 0.05$	[66]
	Micropterus salmoides	0.31-0.53	[70]
	Mugil cephalus	0.43-0.58	[71]
	Oncorhynchus mykiss	$0.16 \pm 0.01$	[66]
	Opisthonema oglinum	0.20	[36]
	Oreochromis niloticus	0.82-0.87	[39]
	Perca fluviatilis	0.20-0.21	[65,66]
	Platichthys flesus	$0.22 \pm 0.02$	[66]
	Rutilus rutilus	0.21	[65]
	Salmo trutta	0.21-0.30	[72]
	Scomberomorus cavalla	0.24	[36]
	Sparus aurata	0.14-0.19	[73]
Meat	Chicken (purchased from a hatchery and poultry farm)	0.755-0.784	[48]
	Chicken (Caribro Vishal)	0.288-1.694	[50]
	Rabbit (Curcuma longa)	0.83-1.12	[75]
	Lamb (Barbarine lamb)	1.1-1.15	[43]
	Heifer (Limousin heifer)	1.10-1.34	[77]
	Foal (Galician Mountain × Hispano-Bretón)	0.44 - 0.80	[52]
	Bologna sausages	0.39–1.55	[53]
Dairy products	Cheese of Holstein cow	3.22-5.04	[88]
	Curd of cow (Middle Rhodopes)	2.02-4.35	[78]
	Milk of goat (market of Sardinia)	2.70-3.20	[89]
	Milk of Chios sheep	1.24-1.46	[54]
	Milk of Karagouniko sheep	1.00 - 1.47	[54]
	Milk of Lacaune ewe	2.20-2.72	[80]
	Milk of Turcana dairy ewe	1.22-1.76	[55]
	Milk of Holstein cow	2.23-2.90	[88]
	Milk of Jersey cow	3.9813-4.6558	[87]
	Milk of cow (Middle Rhodopes)	2.05-4.03	[78]
	Yogurt of cow milk (market of Faisalabad)	0.39-1.84	[86]
	Yogurt of sheep milk (market of Faisalabad)	0.65-1.68	[86]

Table 4. Cont.

IT: index of thrombogenicity; \* literature from 2000 until April/2020; <sup>a</sup> recalculated according to the original data in the reference.

#### 2.4. Hypocholesterolemic/Hypercholesterolemic (HH) Ratio

The hypocholesterolemic/hypercholesterolemic (HH) ratio is an index used in the FA profile of lamb meat first proposed by Santos-Silva et al. in 2002 [91]. Due to the high proportion of SFA, the PUFA/SFA is normally low in lambs, so Santos-Silva et al. developed the HH as a new index to assess the effect of FA composition on cholesterol.

Basic on research about dietary FA and the regulation of plasma LDL-C [111], the HH characterizes the relationship between hypocholesterolemic fatty acid (*cis*-C18:1 and PUFA) and hypercholesterolemic FA. Because there was no C12:0 detected in the lambs, Santos-Silva et al. concluded that the formula only includes C14:0 and C16:0 in hypercholesterolemic FA. Later, Mierliță optimized the formula by adding the C12:0 in hypercholesterolemic FA during the studies of sheep milk [55]. The formula is:

$$HH = (cis - C18 : 1 + \Sigma PUFA) / (C12 : 0 + C14 : 0 + C16 : 0)$$
(3)

10 of 24

Compared with the PUFA/SFA ratio, the HH ratio may more accurately reflect the effect of the FA composition on CVD. The HH ratio has certain limitations. Similar to the IA and IT, the HH might include more kinds of fatty acids such as other molecular species of MUFA and different weights can be assigned to different molecular FA species.

The HH was first used in research on ruminants [46,77,91], which was subsequently extended to dairy products [54,55,78,86,87], marine products [34,36–39,72,90], and other fields [47,48,52,62]. Paiva et al. selected four Azorean macroalgae and used the HH as one of the indices to evaluate their nutritional and health promoting aspects, and found that the HH value ranges from 1.26 to 2.09 [90]. Ratusz et al. analyzed the FA content in 29 cold-pressed camelina (*Camelina sativa*) oils using the HH as a nutritional quality index. A relatively high HH was reported, ranging from 11.7 to 14.7, with a low IA and IT contributing to a decrease in the incidence of CHD [62].

Detailed information about the literature related to the HH is shown in Table 5. For shellfish, the HH value ranges from 1.73 to 4.75, except for *Loxechinus albus*. It is possible that the main food source of *Loxechinus albus* is algae, leading to a high proportion of SFA, so its HH is only 0.21, lower than in other species [34]. For fish, the value ranges from 1.54 to 4.83, with the exception of *Opisthonema oglinum*, which has an HH value of 0.87 [36]. For meat and dairy products, the ranges are 1.27–2.786, 0.32–1.29, respectively.

Materials		HH Value	Reference
<b>D</b> 1 1	Gelidium microdon	4.22	[90]
Red seaweed	Pterocladiella capillacea	2.09	[90]
Brown seaweed	Ulva compressa	1.90	[90]
	Ulva rigida	1.26	[90]
Plant oil	Camelina oil (Camelina sativa)	11.2–15.0	[62]
Shellfish	Cancer edwardsi	4.75	[34]
	Cervimunida johni	3.48	[34]
	Concholepas	2.52	[34]
	Heterocarpus reedi	2.91	[34]
	Loxechinus albus	0.21	[34]
	Mesodesma donacium	2.15	[34]
	Pleuroncodes monodon	3.68	[34]
	Pyura chilensis	1.73	[34]
	Venus antiqua	1.90	[34]
Fish	Cilus gilberti	1.86	[34]
	Genypterus chilensis	2.93	[34]
	Hemiramphus brasiliensis	2.46	[36]
	Hyporhamphus unifasciatus	2.43	[36]
	Kutum roach	2.04-4.83	[37]
	Lagocephalus guentheri	2.68	[38]
	Merluccius gayi	2.23	[34]
	Opisthonema oglinum	0.87	[36]
	Oreochromis niloticus	1.56-1.63	[39]
	Pinguipes chilensis	1.54	[34]
	Salmo trutta	1.88-2.16	[72]
	Scomber japonicus	2.00	[34]
	Scomberomorus cavalla	1.56	[36]
	Seriola lalandi	2.14	[34]
	Seriolella violacea	2.10	[34]
	Trachurus murphyi	1.73	[34]
Meat	Chicken (purchased from a hatchery and poultry farm)	2.658-2.786	[48]
	Lamb (Merino Branco)	1.92	[91]
	Lamb (Ile de France × Merino Branco)	2.01	[91]
	Cattle (Nellore cattle)	1.56-2.08	[46]
	Heifer (Limousin heifer)	1.27-1.87	[77]
	Foal (Galician Mountain × Hispano-Bretón)	1.76-1.98	[52]
	Spanish dry-cured ham	2.0-2.67	[47]

Table 5. Application of HH in fatty acid evaluation \*.

Materials		HH Value	Reference
Dairy products	Curd of cow (Middle Rhodopes)	0.32-0.74	[78]
	Milk of Chios sheep	0.50-0.61	[54]
	Milk of Karagouniko sheep	0.50-0.68	[54]
	Milk of Turcana dairy ewe	0.88-1.29	[55]
	Milk of cow (Middle Rhodopes)	0.34-0.75	[78]
	Milk of Jersey cow	0.4067-0.5732	[87]
	Yogurt of cow milk (market of Faisalabad)	0.54–1.12	[86]
	Yogurt of sheep milk (market of Faisalabad)	0.82-1.29	[86]

Table 5. Cont.

HH: hypocholesterolemic /hypercholesterolemic ratio; \* literature from 2000 until April/2020.

#### 2.5. Health-Promoting Index (HPI)

The health-promoting index (HPI) was proposed by Chen et al. in 2004 to assess the nutritional value of dietary fat [94], which focuses on the effect of FA composition on CVD. The formula is:

$$HPI = \Sigma UFA / [C12:0 + (4 \times C14:0) + C16:0].$$
(4)

The HPI is the inverse of the IA. It is currently mainly used in research on dairy products such as milk [92–94] and cheese [57,94,95]. Detailed information about the literature related to the HPI is provided in Table 6. Its values range from 0.16 to 0.68. Dairy products with a high HPI value are assumed to be more beneficial to human health. The HPI has the same shortcoming as the IA, and it requires reliable evidence to optimize the relevant coefficients.

Materials		HPI Value	Reference
Dairy products	Butter of Holstein cow	0.37-0.66	[93,94]
	Cheese of Red Syrian goat	0.37-0.68	[95]
	Cheese of Comisana ewe	0.42-0.50	[57]
	Cheese (Cheddar cheese) of Holstein cow	0.29-0.46	[94]
	Cheese (Provolone Cheese) of Holstein cow	0.38-0.63	[94]
	Cream of Holstein cow	0.31-0.62	[94]
	Milk of ewe (Comisana breed)	0.16-0.28	[92]
	Yogurt of Holstein cow	0.30-0.62	[94]

HPI: health-promoting index; \* literature from 2000 until April/2020.

#### 2.6. Unsaturation Index (UI)

The UI indicates the degree of unsaturation in lipids and is calculated as the sum of the percentage of each unsaturated FA multiplied by the number of double bonds within that FA [114]. The calculation formula is:

$$UI = 1 \times (\% \text{ monoenoics}) + 2 \times (\% \text{ dienoics}) + 3 \times (\% \text{ trienoics}) + 4 \times (\% \text{ tetraenoics}) + 5 \times (\% \text{ pentaenoics}) + 6 \times (\% \text{ hexaenoics})$$
(5)

Unlike  $\sum$  UFA and  $\sum$  PUFA, different unsaturated FAs have different weights in the UI. This index indicates the impact of highly unsaturated FA and does not ignore the impact of FAs that have a low degree of unsaturation. In general, the UI more comprehensively reflects the proportion of FA with different degrees of unsaturation in the total FA composition of a species.

The UI is commonly used to determine the composition of macroalgal FA. It can be used as a standard for judging the content of high-quality PUFA, in which macroalgae may be used as alternative sources of high-quality PUFA instead of fish or fish oil [98]. Colombo et al. used the UI to compare macroalgae in cold water with those in warm water, with a high UI value indicating a high degree of total unsaturation. Their results suggested that the fatty acids with a high degree of unsaturation in a membrane lipid can maintain fluidity at relatively low temperature [96].

Detailed information about the literature related to the UI is listed in Table 7. The UI value of seaweeds varies widely from 45 to 368.68, and may be closely related to their species. There is no rule at present. The disadvantage of the UI is that it only focuses on the degree of unsaturation of FAs and does not distinguish between n-6 and n-3 FA. The fatty acids in the n-6 and n-3 series have different physiological effects on the human body.

Red seaweed         Amplicia maceps         250 ± 1.01         [98]           Amplicion anceps         98.01,975         [27.98]           CallaplyIIs sp         117         [96]           Camina officinalis         202 ± 19         [97]           Caldabulit across         202 ± 19         [97]           Gelidatia across         202 ± 19         [97]           Gelidatia across         190         [27]           Gelidatia across         202 ± 10         [97]           Genolaria changii         38.68 ± 20.01         [28]           Gracharia changii         204.85, 205 ± 3.07         [27]           Gracharia changii         134.75, 135 ± 1.14         [27.98]           Gracharia dabiis         204.87, 205 ± 3.07         [27]           Gracharia dabiis         204.87, 205 ± 3.07         [27]           Gracharia dabiis         204.87, 205 ± 3.07         [27]           Gracharia dabias         204.87, 205 ± 3.07         [27]           Gracharia dabiis         204.87, 205 ± 3.07         [27]           Gracharia dabias         216 ± 1.07         [27.88]           Gracharia dabias         216 ± 1.07         [27]           Hymate masofermis         913 ± 4.11         [27.89] <t< th=""><th>Materials</th><th></th><th>UI Value</th><th>Reference</th></t<>	Materials		UI Value	Reference
Ampinon anceps         98.01,97.5         [27,98]           Callophilys p         117         [96]           Commun virgetrun         284±7         [99]           Comlina officialis         202±19         [59]           Comlina officialis         202±19         [59]           Collide acrosa         191.02         [27]           Collider incompterum         98.80         [27]           Collider incompterum         98.80         [27]           Conciloria contexta         255.07         [27]           Graciloria detalis         20.04.85,025.13.07         [27,98]           Graciloria fergusonii         134.75,135.14         [27,98]           Graciloria fergusonii         134.75,135.14         [27,98]           Graciloria fergusonii         134.75,135.14         [27,98]           Graciloria fergusonii         134.75,135.14         [27,98]           Graciloria selicorrita         22.29.8         [27,98]           Graciloria properio         9.35.44.63         [98]           Hymen experi         140.94,141.44.05         [27,98]           Hymen experi         143.85.17         [27,98]           Jacinera informe         245.54.24.61.12         [27,98]           Jacinera informe	Red seaweed	Ahnfeltia plicata	$250 \pm 1.01$	[98]
Calapitulis sp         117         [96]           Comitina officinalis         202 ± 19         [57]           Calidia acrosa         191.02         [27]           Calidia acrosa         191.02         [27]           Calidia acrosa         191.02         [27]           Calidia acrosa         191.02         [27]           Calidia acrosa         195.07         [27]           Concilaria changi         368.68 ± 20.01         [28]           Concilaria contexta         204.85, 205 ± 3.07         [27]           Gracilaria debilis         204.85, 205 ± 3.07         [27]           Gracilaria fergusonii         [134.75, 135 ± 1.14         [27,98]           Gracilaria fergusonii         [134.75, 135 ± 1.14         [27,98]           Gracilaria fergusonii         [14]         [27,98]           Gracilaria fergusonii         [10]         [21,31 ± 4.13           Hyme asperi         9.3 ± 4.11         [28]           Hyme amescfirmis         [27,98]         [24]           Laurencia curcitai         [17,29]         [27]           Laurencia curcitai         [27,98]         [27]           Softera robusia         [24,54]         [27]           Laurencia curcitai         [27]		Amphiora anceps	98.01, 97.5	[27,98]
		Callophylis sp	117	[96]
Corallina officinalis         202 ± 19         [59]           Géliditan micropterum         98,80         [27]           Géliditan micropterum         98,80         [27]           Giologiti farcala         54         [96]           Gracilaria changti         368,68 ± 20.01         [28]           Gracilaria corticata         229,10,29 ± 3.66         [27,98]           Gracilaria denia         204,01,29 ± 3.66         [27,98]           Gracilaria fergusonii         134,75, 135 ± 1.14         [27]           Gracilaria factoria         286,65 ± 20.01         [28]           Gracilaria fergusonii         134,75, 135 ± 1.14         [27,98]           Gracilaria pergisoni         93,6 ± 4.63         [98]           Hymenea speri         93,6 ± 4.63         [98]           Lawrencia cruciata         122,95,17 ± 5.64         [27,98]           Lawrencia cruciata         122,95         [73 ± 5.4]         [27,98]           Lawrencia cruciata         122,95,17 ± 5.54         [27,98]           Lawrencia cruciata         125,54,246 ± 127         [27,98]           Delegiptionia sp.         134 ± 15         [59]           Sorgassum fusiforme         125,65 ± 32.25         [29]           Delegiptionia sp.         145		Ceramium virgatum	$284 \pm 7$	[59]
Gelidialia acerssa         191.02         [27]           Gelidialia micropterum         98.80         [27]           Gloiopellis furcata         36.868         20.01         [28]           Gracilaria conticata         257.07         [27]           Gracilaria conticata         257.07         [27]           Gracilaria dultis         204.85         204.85         [27]           Gracilaria dultis         204.85         [26]         [27]           Gracilaria solicornia         294.5         [26]         [27]           Gracilaria solicornia         284.5.91         [98]         [98]           Grateloupia untiti         181.8.3.77         [98]         [98]         [14]         [14]         [27].98           Laurencia cruciata         172.95, 173.5.4.4.11         [98]         [14]         [27].98         [26]           Laurencia cruciata         172.95, 173.5.4.4.1.1         [98]         [98]         [27].98         [27].98           Laurencia cruciata         121.99         [27].98         [27].98         [27].98           Laurencia cruciata         124.91         [27].98         [27].98         [27].98           Jaurencia cruciata         199.14.44.12         [27].98         [26].99		Corallina officinalis	$202 \pm 19$	[59]
Gelidium microprenm         98.80         [27]           Gelidium microprenm         98.80         20.10           Gracellaria contexta         227.07         [27]           Gracellaria contexta         224.01.24         368.68 ± 20.01         [28]           Gracellaria contexta         224.01.24         366.61         [27.98]           Gracellaria forguessni         134.75, 135 ± 1.14         [27.98]           Gracellaria forguessni         181 ± 3.77         [98]           Gracellaria forguessni         181 ± 3.77         [98]           Gracellaria forguessni         181 ± 3.77         [98]           Gracellaria forguessni         19.34 ± 4.11         [98]           Gracellaria calcordiss         10.34 ± 4.11         [98]           Higmen seperi         9.34 ± 4.63         [98]           Laurencia papillosa         213 ± 4.89         [98]           Laurencia papillosa         712         [57]           Laurencia papillosa         712         [59]           Polysiphonia sp.         143 ± 1         [98]           Sarconenn filforme         2454,24,24 ± 1.27         [27,98]           Sarconenn filforme         2454,24 ± 1.27         [27,98]           Dictystot dichotoma         21 ± 1.0		Gelidiella acerosa	191.02	[27]
Gloipellis furcata         96, 90, 92, 93, 92, 93, 92, 93, 92, 93, 92, 93, 92, 93, 92, 93, 93, 94, 93, 94, 93, 94, 93, 94, 93, 94, 94, 93, 94, 94, 94, 94, 94, 94, 94, 94, 94, 94		Gelidium micropterum	98.80	[27]
Gracilaria changii         368.68 ± 20.01         [28]           Gracilaria corticata         257.07         [27]           Gracilaria dobilis         204.85, 205 ± 3.07         [27,98]           Gracilaria dura         249.10, 249 ± 3.66         [27,98]           Gracilaria forgusonii         134.75, 135 ± 1.14         [27,98]           Gracilaria solicorria         50.631         [27]           Grateloupia indicia         266 ± 5.91         [98]           Grateloupia capteri         93.6 ± 4.63         [98]           Hymene asperi         93.6 ± 4.63         [98]           Hymene musciformis         91.3 ± 4.11         [98]           Laurencia papilycus aturezii         140.94, 141 ± 4.05         [27,98]           Laurencia papilycus aturezii         140.94, 141 ± 4.05         [27,98]           Sarconem filiforme         24.554, 24.6 ± 1.27         [27,98]           Sarconem filiforme         24.554, 24.6 ± 1.27         [27,98]           Salera robusta         76 ± 12         [59]           Dictyota dichotoma         221 ± 1.0         [59]           Sargassum fusiformis         221 ± 1.0         [59]           Sargassum fusiformis         222.5 (2.27)         [53]           Sargassum fusiformis <t< td=""><td></td><td>Gloiopeltis furcata</td><td>54</td><td>[96]</td></t<>		Gloiopeltis furcata	54	[96]
Gracilaria corticata         227,92         227           Gracilaria duta         2494,10,249 ± 3.66         227,98           Gracilaria forgusonii         134,75,135 ± 1.14         127,98           Gracilaria silicornia         266 ± 5.91         198           Grateloupia autiti         181 ± 3.77         198           Hymenenu sep         45         199           Hymene sepri         9.3.6 ± 1.43         198           Laurencia cracitat         1122,95,173 ± 5.54         127,98           Laurencia cracitat         127,95,173 ± 5.54         127,98           Laurencia cracitat         127,95,173 ± 5.54         127,98           Laurencia cracitat         127,95,173 ± 5.54         127,98           Solicar orbusta         76 ± 1.2         159           Solicar orbusta         77         196           Brown seaweed         Cystostrin indica         195,44,195 ± 4.21         127,98           Dictyota dichotoma         321 ± 10         159           Sargassum horneri         116,16 ± 5.77         129           Sargassum horneri         161,61 ± 5.77         129           Sargassum horneri         116,16 ± 5.77         129           Sargassum horneri         116,16 ± 5.77         129		Gracilaria changii	$368.68 \pm 20.01$	[28]
Gracilaria debilis         204.85, 205 ± 3.07         22.98           Gracilaria forgusonii         249.10, 249 ± 3.66         22.98           Gracilaria forgusonii         134.75, 135 ± 1.14         12.98           Gracilaria solicornia         26.6 ± 5.91         198           Grateloupia uxutii         181 ± 3.77         198           Hymenesperi         93.6 ± 4.63         198           Hymenesperi         93.6 ± 4.63         198           Laurencia crucitat         121.5 ± 4.54         129.98           Laurencia crucitat         121.5 ± 4.54         199           Lowencia crucitat         121.5 ± 4.54         129.98           Lowencia crucitat         121.5 ± 4.54         129.98           Lowencia crucitat         121.5 ± 4.54         129.98           Lowencia crucitati         121.5 ± 4.54         121.99           Sarcorena filjorme         24.54, 24.6 ± 1.27         129.98           Sargassum fusiormis         22.1 ± 1.10         159           Lathesia difformis         22.2 ± 1.27         129.98           Sargassum fusiormis         12.6 ± 5.77         129           Sargassum fusiormis         12.6 ± 5.77         129           Sargassum fusiormis         12.02         127.98 </td <td></td> <td>Gracilaria corticata</td> <td>257.07</td> <td>[27]</td>		Gracilaria corticata	257.07	[27]
Gracilaria dura         249.10, 249 ± 3.66         [22,98]           Gracilaria salicornia         134.75, 135 ± 1.14         [22,98]           Grateloupia indica         286 ± 5.91         [98]           Grateloupia auttii         181 ± 3.77         [98]           Hymenenu sp         45         [99]           Hymene aperi         9.65 ± 4.63         [98]           Hymene aperi         9.65 ± 4.63         [98]           Hymene aperi         10.94, 141 ± 4.05         [27,98]           Laurencia crucitat         172.95, 173 ± 5.64         [27,98]           Laurencia pupilosa         213 ± 4.89         [98]           Laurencia pupilosa         76 ± 12         [59]           Soliera robusta         77         [96]           Soliera robusta         72 ± 6         [59]           Polysiphonia sp.         124, 195 ± 4.21         [27,98]           Soliera robusta         272 ± 6         [59]           Padina tetrastromatica         143 ± 15         [59]           Sargassum fusiforme         125,65 ± 32.25         [29]           Sargassum fusiformis         272 ± 6         [59]           Mariogloen major         246 ± 7         [59]           Sargassum pubildum		Gracilaria debilis	$204.85, 205 \pm 3.07$	[27,98]
Gracilaria forgusonii         134.75, 135 ± 1.14         [22,98]           Graciloupia indica         266 ± 591         [98]           Grateloupia tuditi         181 ± 3.77         [98]           Hymenena sp         93.6 ± 4.63         [98]           Hymenena spri         93.6 ± 4.63         [98]           Hymenena musciformis         91.3 ± 4.11         [98]           Kappaphycus altorrezii         140.94, 141 ± 4.05         [22,98]           Laurencia papillosa         213 ± 4.89         [98]           Lomentaria clarellosa         76 ± 12         [59]           Polysiphonia sp.         143 ± 15         [59]           Sarcorena filjormis         245.54, 246 ± 1.27         [27,98]           Sartarena filjormis         221 ± 10         [59]           Earthesia difformis         222 ± 6         [59]           Palaina tetrastromatica         154.49, 155 ± 5.50         [27,98]           Sargassum fusiformis         222 ± 6         [59]           Palaina tetrastromatica         154.49, 155 ± 5.50         [29]           Sargassum fusiformis         227 ± 6         [59]           Sargassum fusiformis         227 ± 5.57         [29]           Sargassum fusiformis         227 ± 5.50         [29]		Gracilaria dura	$249.10, 249 \pm 3.66$	[27,98]
Gracilaria silicornia         50.631         [27]           Grateloupia indica         286 ± 5.91         98           Grateloupia indica         181 ± 3.77         [98]           Hymenea sper         93 ± 4.63         [98]           Hymea esperi         93 ± 4.63         [98]           Karpaphycus alterazii         [10.94, 11 ± 4.11         [98]           Laurencia cruciata         [17.295, 173 ± 5.64         [27,98]           Laurencia papillosa         213 ± 4.89         [98]           Lonentaria clavellosa         76 ± 12         [59]           Polysiphonia sp.         143 ± 15         [59]           Sarconema filforme         245.54, 246 ± 1.27         [27,98]           Sarconema filforme         245.54, 246 ± 1.27         [27,98]           Jeathus adifformitis         227 ± 6         [59]           Myriogloca major         266 ± 7         [59]           Palinu tetrastronatica         115.49, 155 ± 5.50         [27,98]           Sargassum fuisforme         122.65 ± 32.25         [29]           Sargassum fuisforme         122.65 ± 32.25         [29]           Sargassum fuisforme         122.65 ± 32.25         [29]           Sargassum fuisforme         122.66 ± 32.25         [29] </td <td></td> <td>Gracilaria fergusonii</td> <td>134.75, 135 ± 1.14</td> <td>[27,98]</td>		Gracilaria fergusonii	134.75, 135 ± 1.14	[27,98]
Grateloupia indita         286 ± 591         [98]           Grateloupia indita         181 ± 3.7.7         [98]           Hymenena sp         45         [59]           Hymenena sp         33 ± 4.11         [98]           Hymenenia cucita         12.27,98]         [27,98]           Laurencia razinta         12.32 ± 4.09         [98]           Laurencia razinta         72.13 ± 4.04         [27,98]           Lomentria curvellosa         76 ± 1.2         [59]           Sarconema filiforme         245.54, 246 ± 1.27         [27,98]           Sarconema filiforme         195.44, 195 ± 4.21         [27,98]           Dictypta dichotana         321 ± 1.01         [27,98]           Myriogleea major         266 ± 7         [59]           Padina tetrastronatica         154.49, 155 ± 5.25         [29]           Myriogleea major         266 ± 7         [59]           Padina tetrastronatica         125.69, 225, 22         [29]           Sargassum funior         125.63, 225, 22         [29]           Sargassum funior         126.64, 25, 75         [29]           Sargassum funior         126.64, 7         [59]           Sargassum funior         127.63         [27,98]           Sargassum		Gracilaria salicornia	50.631	[27]
Grateloupia varitii         181 = 3.77         [98]           Hymena russ; offrmis         93.6 ± 4.63         [98]           Hymea musc; offrmis         91.0 ± 4.11         [98]           Kappaphycus altarezii         140.94, 141 ± 4.05         [27,98]           Laurencia cruciata         172.95, 173 ± 5.64         [27,98]           Laurencia papillosa         213 ± 4.89         [98]           Lonentaria clavellosa         76 ± 12         [59]           Polysiphonia sp.         143 ± 15         [59]           Sarcomera filiforme         245.54, 246 ± 1.27         [27,98]           Solera robusta         77         [96]           Brown seaweed         Cystoscrin india         195.44, 195 ± 4.10         [27,98]           Dictyota dichotoma         321 ± 10         [27,98]           Sargassium fusiormia         222 ± 6         [59]           Muriogloen major         266 ± 7         [59]           Palina tetrastromatica         154.49, 155 ± 5.50         [27,98]           Sargassium fusiormia         122.05         [29]           Sargassium startii         108.02         [27]           Sargassium startii         120.02         [27]           Sargassium stariti         128.02         [27]<		Grateloupia indica	$286 \pm 5.91$	[98]
Hymenena spr         45         [59]           Hypnea musciformis         91.3 ± 4.11         [98]           Kappaphyus altarezii         140.94, 141 ± 4.05         [27,98]           Laurencia cruciata         127.95, 173 ± 5.64         [27,98]           Laurencia pruillosa         213 ± 4.13         [98]           Lomentaria clavellosa         74 ± 89         [98]           Lomentaria clavellosa         74 ± 15         [59]           Sarconena filjorne         245.54, 246 ± 1.2         [27,98]           Salera robusia         77         [96]           Brown seaweed         Cystosiria inficia         195.44, 195 ± 4.21         [27,98]           Dictypta dichotoma         321 ± 10         [59]         Myriogloca major         266 ± 7         [59]           Brown seaweed         Cystosiria inficia         154.49, 155 ± 5.50         [27,98]         Sargassum horneri         116.16 ± 5.7         [29]           Sargassum horneri         116.16 ± 5.7         [29]         Sargassum horneri         182.02         [27]           Sargassum thumbergii         89.87 ± 4.47         [27,98]         Sargassum thumbergii         89.87 ± 7.44         [29]           Sargassum thumbergii         187.05, 187 ± 4.47         [27,98]         [27,98]		Grateloupia wattii	$181 \pm 3.77$	[98]
Hypne avec Hypne musciformis         913 ± 4.11         [98]           Kappaphycus alvarczii         140.94, 141 ± 4.05         [27,98]           Laurencia cruciata         172.95, 173 ± 5.64         [27,98]           Laurencia papillosa         213 ± 4.89         [98]           Lornentaria clavellosa         76 ± 12         [59]           Polysiphonia sp.         143 ± 15         [59]           Sarconema filjorme         245.54, 246 ± 1.27         [27,98]           Soliera robusta         77         [96]           Brown seaweed         Cystoceria india         195.44, 195 ± 4.21         [27,98]           Dictyota dichotoma         321 ± 10         [59]         [96]           Myriogloea major         266 ± 7         [59]         [96]           Myriogloea major         125.65 ± 32.25         [29]         [29]           Sargassum functrastromatica         154.49, 155 ± 5.50         [29]         [27]           Sargassum functrastromatica         154.49, 155 ± 5.50         [29]         [27]           Sargassum functrastromatica         187.05, 187 ± 4.47         [29]         [27]           Sargassum functrastromatica         187.05, 187 ± 4.47         [29]         [27]           Sargassum functrastromatica         1067, 1		Hymenena sp	45	[59]
Hypne nucciformis         91.3 $\pm 4.11$ [98]           Kappaphycus alvarezii         140.94, 141 $\pm 4.05$ [27,98]           Laurencia cruciata         172.95, 173 $\pm 5.64$ [27,98]           Laurencia papillosa         213 $\pm 4.89$ [98]           Lomentaria clarellosa         76 $\pm 12$ [59]           Polysiphonia sp.         143 $\pm 15$ [59]           Soliera robusta         77         [96]           Brown seaweed         Cystocim indica         195.44, 195 $\pm 4.21$ [27,98]           Dictypta dichotoma         321 $\pm 10$ [59]           Lathesia difformis         272 $\pm 6$ [59]           Myriogloa major         266 $\pm 7$ [59]           Padina tetrastromatica         154.49, 155 $\pm 5.50$ [27,98]           Sargassum fusiforme         125.05, 187 $\pm 4.47$ [27,98]           Sargassum numeri         161.6 $\pm 5.77$ [29]           Sargassum terrinum         187.05, 187 $\pm 4.47$ [27,98]		Hypnea esperi	$93.6 \pm 4.63$	[98]
Kappaphycis alcorezii         14094, 111 ± 4.05         [27,98]           Laurencia ruaciita         172,95, 173 ± 5.64         [27,98]           Laurencia papillosa         213 ± 4.89         [98]           Lomentaria clavellosa         76 ± 12         [59]           Polysiphonia sp.         143 ± 15         [59]           Soircer orbusta         77         [96]           Brown seaweed         Cystosein indica         195,44,195 ± 4.21         [27,98]           Leathesia difformis         272 ± 6         [59]           Myriogloca major         266 ± 7         [59]           Palaina tetrastromatica         154,49,155 ± 5.50         [27,98]           Sargassum fusforme         125,65 ± 32.25         [29]           Sargassum fusforme         126,65 ± 32.25         [29]           Sargassum fusforme         126,65 ± 32.25         [29]           Sargassum functria         182,02         [27]           Sargassum suratzii         182,02         [27]           Sargassum functria         182,02         [27]           Sargassum tunerzii         182,02         [27]           Sargassum tunerzii         182,02         [27]           Sargassum tunerziiitam         176,43,75         [28] </td <td></td> <td>Hypnea musciformis</td> <td><math>91.3 \pm 4.11</math></td> <td>[98]</td>		Hypnea musciformis	$91.3 \pm 4.11$	[98]
		Kappaphycus alvarezii	$140.94, 141 \pm 4.05$	[27,98]
		Laurencia cruciata	$172.95, 173 \pm 5.64$	[27,98]
		Laurencia papillosa	$213 \pm 4.89$	[98]
Polysiphonia sp.         143 ± 15         [59]           Sarconema filiforme         245.54, 246 ± 1.27         [27,98]           Soliera robusia         77         [96]           Brown seaweed         Cystoseira indica         195.44, 195 ± 4.21         [27,98]           Dictyota dichotoma         321 ± 10         [59]           Leathesia difformis         272 ± 6         [59]           Myrioglea major         266 ± 7         [59]           Padina tetrastromatica         154.49, 155 ± 5.50         [27]           Sargassum fusiforme         125.65 ± 32.25         [29]           Sargassum horneri         116.16 ± 5.77         [29]           Sargassum horneri         182.02         [27]           Sargassum horneri         182.02         [27]           Sargassum tenzrimum         187.05, 187 ± 4.47         [27,98]           Sargassum tenzrimum         187.05, 187 ± 4.47         [27,98]           Sargassum tenzrimum         187.05, 187 ± 4.47         [27,98]           Sargassum sequent         202.83, 203 ± 3.06         [27,98]           Green seaweed         Caulerpa racemosa         106.70, 107 ± 5.67         [27,98]           Caduerpa acendosa         121.67         [27]         [59]		Lomentaria clavellosa	$76 \pm 12$	[59]
Sarconem filjforme         245.54, 246 $\pm$ 1.27         [27,98]           Soliera robusta         77         [96]           Brown seaweed         Cystoscira indica         195.44, 195 $\pm$ 4.21         [27,98]           Dictyota dichotoma         321 $\pm$ 10         [59]           Lathesia diformis         272 $\pm$ 6         [59]           Mariogloa major         266 $\pm$ 7         [59]           Padina tetrastromatica         154.49, 155 $\pm$ 5.50         [27,98]           Sargassum fusiforme         125.65 $\pm$ 32.25         [29]           Sargassum plailidum         62.27 $\pm$ 15.05         [29]           Sargassum svartzii         182.02         [27]           Sargassum thurbergii         89.87 $\pm$ 7.44         [29]           Spatoglosum asperum         202.83, 203 $\pm$ 3.06         [27,98]           Stoechospernum marginatum         176 $\pm$ 3.56         [98]           Undaria pinnatifida         260-318         [59]           Green seaweed         Caulerpa carpelfornis         121.67         [27]           Caulerpa carpelfornis         121.67         [27]         [59]           Codium reminara         121.67         [27]         [59]           Codium facionitatum         219 $\pm$ 2         [59]     <		Polysiphonia sp.	$143 \pm 15$	[59]
Soliera robusia         77         [96]           Brown seaweed         Cystosein indica         195.44, 195 ± 4.21         [27,98]           Dictyota dichotoma         321 ± 10         [59]           Latthesia difformis         272 ± 6         [59]           Myriogloea major         266 ± 7         [59]           Padina tetrastromatica         154.49, 155 ± 5.50         [27,98]           Sargassum horneri         116.16 ± 5.77         [29]           Sargassum balidum         62.27 ± 15.05         [29]           Sargassum tenerrimum         182.00         [27]           Sargassum tenerrimum         182.02         [27]           Sargassum tenerrimum         182.05         [27,98]           Stoechospermum marginatum         176 ± 3.56         [98]           Undaria pinnatifida         260-318         [59]           Green seaweed         Caulerpa acamosa         106.70, 107 ± 5.67         [27,98]           Caulerpa veravalnensis         114.87, 142 ± 2.96 </td <td></td> <td>Sarconema filiforme</td> <td>245.54, 246 ± 1.27</td> <td>[27,98]</td>		Sarconema filiforme	245.54, 246 ± 1.27	[27,98]
Brown seaweed         Cystoseira indica         195.44, 195 $\pm$ 4.21         [27,98]           Dictyota dichotoma         321 $\pm$ 10         [59]           Leathesia difformis         272 $\pm$ 6         [59]           Myriogloea major         266 $\pm$ 7         [59]           Padina tetrastromatica         154.49, 155 $\pm$ 5.50         [27,98]           Sargassum fusiforme         125.65 $\pm$ 32.25         [29]           Sargassum pallidum         62.27 $\pm$ 15.05         [27]           Sargassum sourtzi         182.00         [27]           Sargassum tourtzi         182.00         [27]           Sargassum tourtzi         182.00         [27]           Sargassum tourtzi         182.00         [27]           Sargassum tourtzii         182.00         [27]           Sargassum thumbergii         89.87 $\pm$ 7.44         [29]           Spatoglossum asperum         202.83, 203 $\pm$ 3.06         [27,98]           Stoclospernum marginatum         176 $\pm$ 3.56         [98]           Undaria pinnatifida         216 7         [59]           Calderpa racemosa         106.70, 107 $\pm$ 567         [27,98]           Cladoptora falkIandica         215 $\pm$ 7         [59]           Codium decorticatum         215 $\pm$ 7 </td <td></td> <td>Soliera robusta</td> <td>77</td> <td>[96]</td>		Soliera robusta	77	[96]
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Brown seaweed	Cystoseira indica	195.44, 195 ± 4.21	[27,98]
Leathesia difformis         272 ± 6         [59]           Myriogloea major         266 ± 7         [59]           Padina tetrastromatica         154.49, 155 ± 5.50         [27,98]           Sargassum fusiforme         125.65 ± 32.25         [29]           Sargassum pallidum         62.27 ± 15.05         [29]           Sargassum pallidum         62.27 ± 15.05         [29]           Sargassum tourbergii         182.02         [27]           Sargassum thunbergii         88.87 ± 7.44         [29]           Sargassum thunbergii         88.87 ± 7.44         [29]           Sargassum thunbergii         88.87 ± 7.44         [29]           Spatoglossum asperum         202.83, 203 ± 3.06         [27,98]           Stoechospermum marginatum         176 ± 3.56         [98]           Undaria pinnatifida         260-318         [59]           Caulerpa accenosa         106.70, 107 ± 5.67         [27,98]           Caulerpa vernoalnensis         141.87, 142 ± 2.96         [27,98]           Cladophora falklandica         215 ± 7         [59]           Codium fragile         179 ± 1         [59]           Codium decorticatum         219 ± 2         [59]           Codium decortiatum         219 ± 2         [59] <td></td> <td>Dictyota dichotoma</td> <td><math>321 \pm 10</math></td> <td>[59]</td>		Dictyota dichotoma	$321 \pm 10$	[59]
Myriogloea major         266 ± 7         [59]           Padina tetrastromatica         154.49, 155 ± 5.50         [27,98]           Sargassum fusiforme         125.65 ± 32.25         [29]           Sargassum pallidum         62.27 ± 15.05         [29]           Sargassum sourtzii         116.16 ± 5.77         [29]           Sargassum sourtzii         182.02         [27]           Sargassum sourtzii         182.02         [27]           Sargassum tunubergii         89.87 ± 7.44         [29]           Spatoglossum asperum         202.83, 203 ± 3.06         [27,98]           Stoechospermum marginatum         176 ± 3.56         [98]           Undaria pinnatifida         260-318         [59]           Green seaweed         Caulerpa caemosa         106.70, 107 ± 5.67         [27,98]           Caulerpa caelpeliformis         121.67         [27]         [20]           Colium decorticatum         219 ± 2         [59]         [59]         [50]         [59]           Colium decorticatum         219 ± 2         [59]         [50]         [50]         [51]           Caulerpa calpeliformis         121.67         [27,98]         [10]         [21] ± 2         [59]           Colium decorticatum         219 ± 2		Leathesia difformis	$272 \pm 6$	[59]
Padina tetrastromatica         154.49, 155 ± 5.50         [27,98]           Sargassum fusiforme         116.16 ± 5.77         [29]           Sargassum formeri         116.16 ± 5.77         [29]           Sargassum borneri         118.102         [27]           Sargassum borneri         182.02         [27]           Sargassum tentrimum         187.05, 187 ± 4.47         [27,98]           Sargassum thunbergii         89.87 ± 7.44         [29]           Spatoglossum asperum         202.83, 203 ± 3.06         [27,98]           Stoechospernum marginatum         176 ± 3.56         [98]           Undaria pinnatifida         260-318         [59]           Green seaweed         Caulerpa racemosa         106.70, 107 ± 5.67         [27,98]           Caulerpa veravalnensis         121.67         [27]         [27]           Caulerpa veravalnensis         121.67         [27]         [29]           Codium decorticatum         219 ± 2         [59]         [59]           Codium decorticatum         219 ± 2         [59]         [59]           Codium vermilara         135 ± 16         [59]         [59]           Ultor fasciat         102.92, 103 ± 2.83         [27,98]           Ultor fasciat         102.92, 103		Myriogloea major	$266 \pm 7$	[59]
Sargassum fusiforme         125.65 ± 32.25         [29]           Sargassum pallidum         116.16 ± 5.77         [29]           Sargassum pallidum         62.27 ± 15.05         [29]           Sargassum pallidum         62.27 ± 15.05         [29]           Sargassum tenerrinum         187.05, 187 ± 4.47         [27,98]           Sargassum tenerrinum         187.05, 187 ± 4.47         [27,98]           Sargassum tenerrinum         187.05, 187 ± 4.47         [27,98]           Sargassum sourtzii         89.87 ± 7.44         [29]           Spatoglossum asperum         202.83, 203 ± 3.06         [27,98]           Stocchospermum marginatum         176 ± 3.56         [98]           Undaria primatifida         260-318         [59]           Green seaweed         Caulerpa racemosa         106.70, 107 ± 5.67         [27,98]           Caluderpa ecavalenesis         141.87, 142 ± 2.96         [27,98]           Codium decorticatum         215 ± 7         [59]           Codium decorticatum         215 ± 7         [59]           Codium decorticatum         215 ± 7         [59]           Codium decorticatum         124 ± 4.23         [98]           Ulva lactuca         87.5 ± 5.76         [98]           Ulva lactuca </td <td></td> <td>Padina tetrastromatica</td> <td><math>154.49, 155 \pm 5.50</math></td> <td>[27,98]</td>		Padina tetrastromatica	$154.49, 155 \pm 5.50$	[27,98]
Sargassum horneri         116.16 ± 5.77         [29]           Sargassum pallidum         62.27 ± 15.05         [29]           Sargassum swartzii         182.02         [27]           Sargassum swartzii         89.87 ± 7.44         [29]           Sargassum tenerrimum         187.05, 187 ± 4.47         [27,98]           Sargassum thunbergii         89.87 ± 7.44         [29]           Sargassum marginatum         106.70, 107 ± 5.67         [27,98]           Caulerpa racemosa         106.70, 107 ± 5.67         [27,98]           Caulerpa racemosa         121.67         [27]           Caulerpa racemosa         121.67         [27]           Caulerpa racemosa         121.67         [27,98]           Cladophora falklandica         215 ± 7         [59]           Codium decorticatum         219 ± 2         [59]           Codium decorticatum         135 ± 16         [59]           Ultra fasciata         102.92, 103 ± 2.83         [27,98]           Ultra intza         124 ± 4.23         [98]		Sargassum fusiforme	$125.65 \pm 32.25$	[29]
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		Sargassum horneri	$116.16 \pm 5.77$	[29]
Sargassum svartzii         182.02         [27]           Sargassum tenerrinum         187.05, 187 ± 4.47         [27,98]           Sargassum thunbergii         9.8,7 ± 7.44         [29]           Spatoglossum asperum         202.83, 203 ± 3.06         [27,98]           Stoechospermum marginatum         176 ± 3.56         [98]           Undaria pinnatifida         260-318         [59]           Green seaweed         Caulerpa acapeliformis         121.67         [27]           Caulerpa scalpeliformis         121.67         [27]           Caulerpa scalpeliformis         121.67         [27]           Caulerpa scalpeliformis         121.67         [27]           Caulerpa scalpeliformis         121.67         [27]           Codium faiklandica         215 ± 7         [59]           Codium decorticatum         219 ± 2         [59]           Codium vermilara         135 ± 16         [59]           Ulva fasciata         102.92, 103 ± 2.83         [27,98]           Ulva fasciata         102.92, 103 ± 2.83         [27,98]           Ulva rigida         93.66, 93.8 ± 5.31         [27,98]           Ulva rigida         93.66, 93.8 ± 5.31         [27,98]           Ulva rigida         93.66, 93.8 ± 5.31		Sargassum pallidum	$62.27 \pm 15.05$	[29]
Sargassum tenerrimum         187.05, 187 ± 4.47         [27,98]           Sargassum thunbergi         89.87 ± 7.44         [29]           Spatoglossum asperum         202.83, 203 ± 3.06         [27,98]           Stoechospermum marginatum         176 ± 3.56         [98]           Undaria pinnatifida         260-318         [59]           Green seaweed         Caulerpa racemosa         106.70, 107 ± 5.67         [27,98]           Caulerpa scalpeliformis         121.67         [27]           Caulerpa scalpeliformis         121.67         [27]           Caulerpa veravalnensis         141.87, 142 ± 2.96         [27,98]           Cladophora falklandica         215 ± 7         [59]           Codium decorticatum         219 ± 2         [59]           Codium vermilara         135 ± 16         [59]           Ulva fasciata         102.92, 103 ± 2.83         [27,98]           Ulva lactuca         87.5 ± 5.76         [98]           Ulva reticulata         70.87, 5.9 ± 5.33         [27,98]           Ulva reticulata         70.87, 5.9 ± 5.33         [27,98]           Ulva tubulosa         99.6 ± 3.23         [98]           Ulva sp.1         209 ± 2.0         [59]           Ulva sp.2         288 ± 10		Sargassum swartzii	182.02	[27]
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		Sargassum tenerrimum	$187.05, 187 \pm 4.47$	[27,98]
$ \begin{array}{c cccc} Spatoglossum asperum & 202.83, 203 \pm 3.06 & [27,98] \\ Stoechospernum marginatum & 176 \pm 3.56 & [98] \\ Undaria pinnatifida & 260-318 & [59] \\ \hline \\ Green seaweed & Caulerpa racemosa & 106.70, 107 \pm 5.67 & [27,98] \\ Caulerpa veravalnensis & 121.67 & [27] \\ Caulerpa veravalnensis & 141.87, 142 \pm 2.96 & [27,98] \\ Cladophora falklandica & 215 \pm 7 & [59] \\ Codium decorticatum & 219 \pm 2 & [59] \\ Codium vermilara & 135 \pm 16 & [59] \\ Codium vermilara & 135 \pm 16 & [59] \\ Ulva fasciata & 102.92, 103 \pm 2.83 & [27,98] \\ Ulva fasciata & 102.92, 103 \pm 2.83 & [27,98] \\ Ulva linza & 124 \pm 4.23 & [98] \\ Ulva rigida & 93.96, 93.8 \pm 5.31 & [27,98] \\ Ulva rigida & 93.96, 93.8 \pm 5.31 & [27,98] \\ Ulva rigida & 99.96 \pm 3.23 & [98] \\ Ulva sp. & 76.3 \pm 5.40 & [98] \\ Ulva sp. & 76.3 \pm 5.40 & [98] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva regida & 99.6 \pm 3.23 & [98] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva regida & 99.6 \pm 3.23 & [99] \\ Meat & Pig (Pietrain \times (Duroc \times Landrace)) & 111-124 & [44] \\ Dry-cured ham (Landrace \times Large White (25\% Pietrain) pig) & 73 \pm 6 & [101] \\ Dairy products & Milk of (New Zealand × California) white rabbit & 86-120 & [102] \\ \end{array}$		Sargassum thunbergii	$89.87 \pm 7.44$	[29]
Stoechospermum marginatum Undaria pinnatifida         176 ± 3.56 260-318         [98] 260-318           Green seaweed         Caulerpa racemosa Caulerpa scalpeliformis         106.70, 107 ± 5.67         [27,98] Caulerpa scalpeliformis           Caulerpa ceravalnensis         141.87, 142 ± 2.96         [27,98] Caulerpa veravalnensis         141.87, 142 ± 2.96         [27,98] Caulerpa veravalnensis           Caulerpa veravalnensis         141.87, 142 ± 2.96         [27,98] Codium decorticatum         219 ± 2         [59] Codium fragile         [59]           Codium fragile         179 ± 11         [59]         [59]         [59]         [59]           Codium cermilara         135 ± 16         [59]         [59]         [59]         [59]           Ulva fasciata         102.92, 103 ± 2.83         [27,98]         [10a lactuca         [8]         [178]           Ulva linza         124 ± 4.23         [98]         [10a lactuca         [93.96, 93.8 ± 5.31         [27,98]           Ulva reticulata         70.87, 70.9 ± 5.33         [27,98]         [10a regida         [93.96, 93.8 ± 5.31         [27,98]           Ulva regida         93.96, 93.8 ± 5.31         [27,98]         [10a regida         [93.96, 93.8 ± 5.31         [27,98]           Ulva sp.1         102 vertubuosa         99.6 ± 3.23         [98]         [10a sp.1]		Spatoglossum asperum	$202.83, 203 \pm 3.06$	[27,98]
Undaria pinnatifida         260-318         [59]           Green seaweed         Caulerpa racemosa Caulerpa scalpeliformis         106.70, 107 ± 5.67         [27,98]           Caulerpa veravalnensis         121.67         [27]           Caulerpa veravalnensis         141.87, 142 ± 2.96         [27,98]           Cladophora falklandica         215 ± 7         [59]           Codium decorticatum         219 ± 2         [59]           Codium decorticatum         135 ± 16         [59]           Codium vermilara         135 ± 16         [59]           Ulva fasciata         102.92, 103 ± 2.83         [27,98]           Ulva triculata         70.87, 70.9 ± 5.33         [27,98]           Ulva reticulata         70.87, 70.9 ± 5.33         [27,98]           Ulva reticulata         70.87, 70.9 ± 5.33         [27,98]           Ulva sp.         76.3 ± 5.40         [98]           Ulva sp.         76.3 ± 5.40         [98]           Ulva sp.1         209 ± 20         [59]           Ulva sp.2         288 ± 10		Stoechospermum marginatum	$176 \pm 3.56$	[98]
Green seaweedCaulerpa racemosa106.70, 107 $\pm$ 5.67[27,98] Caulerpa scalpeliformisCaulerpa scalpeliformis121.67[27] Caulerpa veravalnensis141.87, 142 $\pm$ 2.96[27,98] Caulerpa veravalnensisCladophora falklandica215 $\pm$ 7[59] Codium decorticatum219 $\pm$ 2[59] Codium decorticatum[59] Codium vernilara135 $\pm$ 16[59] Ulva fasciata[59] Ulva fasciata[27,98] (59] Ulva fasciata[27,98] (59] Codium vernilara[219 $\pm$ 2[59] (59] Ulva fasciata[27,98] (59] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] (20] (20][27,98] 		Undaria pinnatifida	260-318	[59]
Caulerpa scalpeliformis121.67[27]Caulerpa veravalnensis141.87, 142 $\pm$ 2.96[27,98]Cladophora falklandica215 $\pm$ 7[59]Codium decorticatum219 $\pm$ 2[59]Codium regile179 $\pm$ 11[59]Codium vermilara135 $\pm$ 16[59]Ulva fasciata102.92, 103 $\pm$ 2.83[27,98]Ulva fasciata102.92, 103 $\pm$ 2.83[27,98]Ulva lactuca $87.5 \pm 5.76$ [98]Ulva reiculata70.87, 70.9 $\pm$ 5.33[27,98]Ulva rigida93.69, 93.8 $\pm$ 5.31[27,98]Ulva rigida99.6 $\pm$ 3.23[98]Ulva sp.76.3 $\pm$ 5.40[98]Ulva sp.1209 $\pm$ 20[59]Ulva sp.2288 $\pm$ 10[59]CropsCumin (Cuminum cyminum)125.21-133.10[61]Soybean (Glycine max)148–155[99]MeatPig (Pietrain × (Duroc × Landrace))111–124[44]Dry-cured ham (Landrace × Large White (25% Pietrain) pig)73 $\pm$ 6[101]Dairy productsMilk of (New Zealand × California) white rabbit86–120[102]	Green seaweed	Caulerpa racemosa	106.70, 107 $\pm 5.67$	[27,98]
Caulerpa veravalnensis141.87, 142 $\pm$ 2.96[27,98]Cladophora falklandica215 $\pm$ 7[59]Codium decorticatum219 $\pm$ 2[59]Codium decorticatum119 $\pm$ 1[59]Codium vermilara135 $\pm$ 16[59]Ulva fasciata102.92, 103 $\pm$ 2.83[27,98]Ulva lactuca87.5 $\pm$ 5.76[98]Ulva reticulata70.87, 70.9 $\pm$ 5.33[27,98]Ulva reticulata70.87, 70.9 $\pm$ 5.33[27,98]Ulva reticulata93.96, 93.8 $\pm$ 5.31[27,98]Ulva reticulata99.6 $\pm$ 3.23[98]Ulva sp.76.3 $\pm$ 5.40[98]Ulva sp.1209 $\pm$ 20[59]Ulva sp.2288 $\pm$ 10[59]CropsCumin (Cuminum cyminum)125.21-133.10[61]Soybean (Glycine max)148-155[99]MeatPig (Pietrain × (Durce × Landrace))111-124[44]Dry-cured ham (Landrace × Large White (25% Pietrain) pig)73 $\pm$ 6[101]Dairy productsMilk of (New Zealand × California) white rabbit86-120[102]		Caulerpa scalpeliformis	121.67	[27]
Cladophora falklandica $215 \pm 7$ [59]Codium decorticatum $219 \pm 2$ [59]Codium fragile $179 \pm 11$ [59]Codium vermilara $135 \pm 16$ [59]Ulva fasciata $102.92, 103 \pm 2.83$ [27,98]Ulva lactuca $87.5 \pm 5.76$ [98]Ulva lactuca $87.5 \pm 5.33$ [27,98]Ulva reticulata $70.87, 70.9 \pm 5.33$ [27,98]Ulva reticulata $70.87, 70.9 \pm 5.33$ [27,98]Ulva rigida $93.96, 93.8 \pm 5.31$ [27,98]Ulva sp. $76.3 \pm 5.40$ [98]Ulva sp. $209 \pm 20$ [59]Ulva sp.1 $209 \pm 20$ [59]Ulva sp.2 $288 \pm 10$ [59]CropsCumin (Cuminum cyminum) $125.21-133.10$ [61]Soybean (Glycine max) $148-155$ [99]MeatPig (Pietrain × (Duroc × Landrace)) $111-124$ [44]Dry-cured ham (Landrace × Large White (25% Pietrain) pig) $73 \pm 6$ [101]Dairy productsMilk of (New Zealand × California) white rabbit $86-120$ [102]		Caulerpa veravalnensis	$141.87, 142 \pm 2.96$	[27,98]
Codium decorticatum $219 \pm 2$ [59]Codium fragile $179 \pm 11$ [59]Codium vermilara $102.92, 103 \pm 2.83$ [27,98]Ulva fasciata $102.92, 103 \pm 2.83$ [27,98]Ulva lactuca $87.5 \pm 5.76$ [98]Ulva iniza $124 \pm 4.23$ [98]Ulva reticulata $70.87, 70.9 \pm 5.33$ [27,98]Ulva rigida $93.96, 93.8 \pm 5.31$ [27,98]Ulva rigida $93.96, 93.8 \pm 5.31$ [27,98]Ulva sp. $76.3 \pm 5.40$ [98]Ulva sp.1 $209 \pm 20$ [59]Ulva sp.2 $288 \pm 10$ [59]CropsCumin (Cuminum cyminum) $125.21-133.10$ [61]Soybean (Glycine max) $148-155$ [99]MeatPig (Pietrain × (Durce × Landrace)) $111-124$ [44]Dry-cured ham (Landrace × Large White (25% Pietrain) pig) $73 \pm 6$ [101]Dairy productsMilk of (New Zealand × California) white rabbit $86-120$ [102]		Cladophora falklandica	$215 \pm 7$	[59]
$ \begin{array}{c cccc} Codium fragile & 179 \pm 11 & [59] \\ Codium vermilara & 135 \pm 16 & [59] \\ Ulva fasciata & 102.92, 103 \pm 2.83 & [27,98] \\ Ulva lactuca & 87.5 \pm 5.76 & [98] \\ Ulva linza & 124 \pm 4.23 & [98] \\ Ulva reticulata & 70.87, 70.9 \pm 5.33 & [27,98] \\ Ulva reticulata & 70.87, 70.9 \pm 5.33 & [27,98] \\ Ulva rigida & 93.96, 93.8 \pm 5.31 & [27,98] \\ Ulva sp. & 76.3 \pm 5.40 & [98] \\ Ulva sp. & 76.3 \pm 5.40 & [98] \\ Ulva sp. & 76.3 \pm 5.40 & [98] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 2152.1-133.10 & [61] \\ Soybean (Glycine max) & 148-155 & [99] \\ \end{array} $		Codium decorticatum	$219 \pm 2$	[59]
$ \begin{array}{c cccc} Codium vermilara & 135 \pm 16 & [59] \\ Ulva fasciata & 102.92, 103 \pm 2.83 & [27,98] \\ Ulva lactuca & 87.5 \pm 5.76 & [98] \\ Ulva linza & 124 \pm 4.23 & [98] \\ Ulva reticulata & 70.87, 70.9 \pm 5.33 & [27,98] \\ Ulva regida & 93.96, 93.8 \pm 5.31 & [27,98] \\ Ulva tubulosa & 99.6 \pm 3.23 & [98] \\ Ulva sp. & 76.3 \pm 5.40 & [98] \\ Ulva sp. & 76.3 \pm 5.40 & [98] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 209 \pm 20 & [59] \\ Ulva sp. & 288 \pm 10 & [59] \\ \end{array} $		Codium fragile	$179 \pm 11$	[59]
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Codium vermilara	$135 \pm 16$	[59]
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Ulva fasciata	$102.92, 103 \pm 2.83$	[27,98]
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Ulva lactuca	$87.5 \pm 5.76$	[98]
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Ulva linza	$124 \pm 4.23$	[98]
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Ulva reticulata	$70.87, 70.9 \pm 5.33$	[27,98]
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Ulva rigida	93.96, 93.8 ± 5.31	[27,98]
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Ulva tubulosa	99.6 ± 3.23	[98]
$ \begin{array}{c c} Ulva {\rm sp.1} & 209 \pm 20 & [59] \\ Ulva {\rm sp.2} & 288 \pm 10 & [59] \\ \hline {\rm Crops} & {\rm Cumin} ({\rm Cuminum} {\rm cyminum}) & 125.21-133.10 & [61] \\ {\rm Soybean} ({\rm Glycine} {\rm max}) & 148-155 & [99] \\ \hline {\rm Meat} & {\rm Pig} ({\rm Pietrain} \times ({\rm Duroc} \times {\rm Landrace})) & 111-124 & [44] \\ {\rm Dry-cured} {\rm ham} ({\rm Landrace} \times {\rm Large} {\rm White} (25\% {\rm Pietrain}) {\rm pig}) & 73 \pm 6 & [101] \\ \hline {\rm Dairy} {\rm products} & {\rm Milk of} ({\rm New Zealand} \times {\rm California}) {\rm white rabbit} & 86-120 & [102] \\ \hline \end{array} $		Ulva sp.	$76.3 \pm 5.40$	[98]
Ulva sp.2288 $\pm$ 10[59]CropsCumin (Cuminum cyminum) Soybean (Glycine max)125.21-133.10 148-155[61] [99]MeatPig (Pietrain × (Duroc × Landrace)) Dry-cured ham (Landrace × Large White (25% Pietrain) pig)111-124 73 $\pm$ 6[101] [101]Dairy productsMilk of (New Zealand × California) white rabbit86-120[102]		Ulva sp.1	$209 \pm 20$	[59]
		Ulva sp.2	$288 \pm 10$	[59]
Soybean (Glycine max)148–155[99]MeatPig (Pietrain × (Duroc × Landrace))111–124[44]Dry-cured ham (Landrace × Large White (25% Pietrain) pig)73 ± 6[101]Dairy productsMilk of (New Zealand × California) white rabbit86–120[102]	Crops	Cumin (Cuminum cyminum)	125.21-133.10	[61]
MeatPig (Pietrain × (Duroc × Landrace))111–124[44]Dry-cured ham (Landrace × Large White (25% Pietrain) pig) $73 \pm 6$ [101]Dairy productsMilk of (New Zealand × California) white rabbit $86-120$ [102]		Soybean (Glycine max)	148-155	[99]
Dry-cured ham (Landrace × Large White (25% Pietrain) pig) $73 \pm 6$ [101]Dairy productsMilk of (New Zealand × California) white rabbit $86-120$ [102]	Meat	Pig (Pietrain × (Duroc × Landrace))	111-124	[44]
Dairy products         Milk of (New Zealand × California) white rabbit         86–120         [102]		Dry-cured ham (Landrace × Large White (25% Pietrain) pig)	73 ± 6	[101]
	Dairy products	Milk of (New Zealand × California) white rabbit	86-120	[102]

**Table 7.** Application of UI in fatty acid evaluation \*.

UI: unsaturation index; \* literature from 2000 until April/2020.

#### 2.7. Sum of Eicosapentaenoic Acid and Docosahexaenoic Acid (EPA + DHA)

EPA and DHA are n-3 long-chain PUFAs that play essential roles in biological processes in the human body. They can reduce the risk of CVD, hypertension, and inflammation. DHA is a critical component of the retina and the neuronal system and is involved in visual functioning and cognitive functioning in humans [115,116]. The American Heart Association summarized the preventive effect of n-3 PUFA from seafood on CVD in the 2015–2020 Dietary Guidelines for Americans [1].

EPA and DHA can be synthesized from  $\alpha$ -linolenic acid in the human body, but exogenous supplementation is still needed when insufficient.  $\alpha$ -linolenic acid (ALA; C18:3 n-3) can be converted to EPA and DHA by desaturase and elongase, respectively. EPA and DHA can be supplemented by ingesting ALA. Burdge et al. [115] studied the capacity of humans to convert ALA to EPA and DHA. In a carbon isotope labeling experiment, six young male subjects orally received <sup>13</sup>C-ALA as a part of their habitual diet. The results indicated that the subjects had a limited capacity to convert ALA to EPA, and <sup>13</sup>C-labeling of DHA was not detected [115]. Brenna et al. summarized related studies and reached a similar conclusion [117]. Although the conversion of ALA to EPA and DHA was observed in tracer studies in all age groups, regardless of whether the study participant was male or female, the efficiency of directly supplementing with EPA to increase the level of EPA was found to be 15-fold that of supplementing with high levels of ALA. The conversion of ALA to EPA and DHA in infants is only 1%, and is even lower in adults [117]. Therefore, the rate of conversion of ALA to EPA and DHA that is required for health is far from sufficient; direct intake of EPA and DHA is more effective.

EPA + DHA is an index that is recognized worldwide. Recommendations for EPA + DHA intake can be found in various dietary guidelines. According to the Food and Agriculture Organization of the United Nations (UN FAO), the recommended amount is 0.250–2 g/day. Due to the low EPA and DHA contents in terrestrial plants and animals, this index is mostly used to evaluate the nutritional value of seafood and seafood products, particularly fish, which makes it an important nutritional index for seafood. Rincón-Cervera et al. studied the fatty acid composition of fish and shellfish captured in the South Pacific, and the results showed that EPA + DHA ranged between 115.15 and 1370.67 mg/100 g in all studied fish species and between 63.61 and 522.68 mg/100 g in all studied shellfish species [34]. Detailed information about the literature related to EPA + DHA is shown in Table 8. The species of fish and shellfish as well as their nutrition intake are key factors influencing the EPA + DHA value.

### 2.8. Fish Lipid Quality/Flesh Lipid Quality (FLQ)

FLQ was originally use for fish lipid quality [107,108] or flesh lipid quality [65,66,73]. The purpose of FLQ is similar to that of the EPA + DHA index, but it calculates the sum of EPA and DHA as a percentage of total fatty acids. The formula is:

$$FLQ = 100 \times (C22:6 n - 3 + C20:5 n - 3) / \Sigma FA$$
(6)

FLQ is more suitable for marine products given their higher proportions of EPA and DHA. This index may be considered a supplement to EPA + DHA since the absolute quantity for EPA and DHA is more important. Until now, FLQ has only been used to assess the quality of lipids in fish. Senso et al. examined the fatty acid profile of the fillet of farmed sea bream (*Sparus aurata*) harvested in different seasons using FLQ as the lipid quality index. FLQ was lowest in April [73]. Detailed information about the literature related to FLQ is provided in Table 9. The value ranges from 13.01 to 36.37 for closely related species.

Materials		EPA + DHA Value	Reference
Shellfish	Cancer edwardsi	$205.62 \pm 6.19 \text{ mg}/100 \text{ g}$	[34]
	Cervimunida johni	$162.90 \pm 2.83 \text{ mg}/100 \text{ g}$	[34]
	Concholepas concholepas	$63.61 \pm 0.42 \text{ mg}/100 \text{ g}$	[34]
	Heterocarpus reedi	$186.98 \pm 3.88 \text{ mg}/100 \text{ g}$	[34]
	Loxechinus albus	$208.55 \pm 10.28 \text{ mg}/100 \text{ g}$	[34]
	Mesodesma donacium	$216.96 \pm 9.76$ mg/100 g	[34]
	Pleuroncodes monodon	$189.83 \pm 3.74 \text{ mg}/100 \text{ g}$	[34]
	Pyura chilensis	$522.68 \pm 28.02 \text{ mg}/100 \text{ g}$	[34]
	Venus antiqua	$214.34 \pm 7.52 \text{ mg}/100 \text{ g}$	[34]
Fish	Cilus gilberti	294.57 ± 8.76 mg/100 g	[34]
	Dicentrarchus labrax	270–480 mg/100 g	[68]
	Genypterus chilensis	$115.15 \pm 6.16$ mg/100 g	[34]
	Kutum roach	96–250 mg/100 g	[61] <sup>a</sup>
	Merluccius gayi gayi	$309.38 \pm 6.81 \text{ mg}/100 \text{ g}$	[34]
	Pinguipes chilensis	$507.60 \pm 25.32 \text{ mg}/100 \text{ g}$	[34]
	Scomber japonicus	1370.67 ± 55.79 mg/100 g	[34]
	Seriola lalandi	915.76 ± 19.68 mg/100 g	[34]
	Seriolella violacea	$304.04 \pm 14.15 \text{ mg}/100 \text{ g}$	[34]
	Trachinotus carolinus	621–941 mg/100 g	[40]
	Trachurus murphyi	786.90 ± 11.44 mg/100 g	[34]
	Epinephelus coioides	19.9–25.4%	[103]
	Hemiramphus brasiliensis	$16.71\% \pm 0.07\%$	[36]
	Hyporhamphus unifasciatus	$15.53\% \pm 0.07\%$	[36]
	Megalobrama amblycephala	5.52-7.36%	[106]
	Opisthonema oglinum	$40.86\% \pm 0.07\%$	[36]
	Salmo salar	11.80-11.81%	[104]
	Scomberomorus cavalla	$35.06\% \pm 0.07\%$	[36]
	Sparidentex hasta	45.8–230.4 mg/g lipid	[105]

Table 8. Application of EPA + DHA in fatty acid evaluation \*.

EPA + DHA: sum of eicosapentaenoic acid and docosahexaenoic acid; \* literature from 2000 until April/2020; <sup>a</sup> recalculated according to the original data in the reference.

*.

Materials		FLQ Value	Reference
Fish	Abramis brama	24.46-30.14	[65,66]
	Clupea harengus	$13.01 \pm 0.77$	[66]
	Cyprinus carpio	$13.99 \pm 2.15$	[66]
	Esox Lucius	36.37	[65]
	Leuciscus idus	$24.32 \pm 2.47$	[66]
	Oncorhynchus mykiss	$17.97 \pm 2.46$	[66]
	Perca fluviatilis	30.14-33.22	[65,66]
	Platichthys flesus	$20.25 \pm 2.30$	[66]
	Rutilus	28.41	[65]
	Sparus aurata	19.35–31.27	[73]

FLQ: fish lipid quality/flesh Lipid quality; \* literature from 2000 until April/2020.

## 2.9. The Linoleic Acid/α-Linolenic Acid (LA/ALA) Ratio

The linoleic acid (LA, C18:2 n-6)/ $\alpha$ -linolenic acid (ALA, C18:3 n-3) ratio was developed for guiding infant formula. LA and ALA compete for the same desaturase and elongase enzymes, which they use to synthesize long-chain unsaturated fatty acids. Due to the low conversion rate of ALA, reducing the LA/ALA ratio only provides a modest improvement in the levels of some n-3 long-chain PUFAs; however, the balance may be the most important factor when long-chain PUFAs are not present in infant formulas.

The Definitions & Nutrient Composition section of the Guidelines for Infant Formula published by Food Standards Australia New Zealand (FSANZ) sets the minimum and maximum proportions of LA and ALA, and specifies an LA/ALA ratio within 5:1–15:1.

The LA/ALA ratio has a higher reference value when judging the nutritional value of baby food and infant formula. Tissues of adults have a lower rate of synthesis of n-3 long-chain PUFAs than those of infants, so the LA/ALA ratio in the diet does not have too much of an impact on adults. In the literature we reviewed, the LA/ALA ratio was used in research on ruminants and dairy products as well [43,55,109]. Majdoub-Mathlouthi et al. compared the meat fatty acid composition of Barbarine lambs raised on rangelands and those reared indoors. The results showed that the grazing lambs had lower LA/ALA [43]. Sharma et al. compared the fatty acid profile of indigenous Indian cow milk with exotic and crossbred counterparts. LA/ALA was used to reflect the quality of milk [109]. LA/ALA in indigenous cattle was found to be lower than others, providing scientific data for the superiority of indigenous cow milk [109]. Detailed information of the literature related to LA/ALA is listed in Table 10. Turcana dairy ewe milk has a low LA/ALA value due to the high content of ALA given the inclusion of hemp seed in the diet [55].

Materials		LA/ALA Value	Reference
Meat	Lamb (Barbarine lamb)	6.78–10.05	[43]
Dairy products	Milk of Turcana dairy ewe	0.98-1.36	[55]
	Milk of Sahiwal cow	$3.313 \pm 0.262$	[109]
	Milk of Holstein–Friesian cow	$3.446 \pm 0.196$	[109]
	Milk of Sahiwal $ imes$ Holstein–Friesian cow	$3.065 \pm 0.093$	[109]
	Milk of indigenous Indian cow	$2.464\pm0.147$	[109]

Fable 10.	Application	of LA/ALA	in fatty acid	evaluation *
	11	'	2	

#### LA/ALA: linoleic acid/α-linolenic acid ratio; \* literature from 2000 until April/2020.

# 2.10. Trans Fatty Acid (TFA)

Most unsaturated FA in the human diet have a *cis* configuration. However, *trans* fatty acid (TFA) is present in the human diet as well. According to the Food and Drug Administration (FDA), TFA is defined as the sum of all unsaturated fatty acids that contain one or more isolated (i.e., non-conjugated) double bond(s) in a *trans* configuration [118,119]. The European Food Safety Authority (EFSA) gives a different definition of TFA, which are also present as either *trans*-MUFA or *trans*-PUFA. *Trans*-PUFAs have at least one *trans* double bond and may therefore also have double bonds in the *cis* configuration. Conjugated fatty acid (CLA) is separated from TFA as an independent section by the EFSA. CLAs may have health benefits that are different from those of TFAs, such as anti-cancer [120,121] and anti-atherosclerosis [122] activities, so it is appropriate to exclude CLA from the definition of TFA.

According to the EFSA, TFAs may originate from various sources, including of bacterial conversion of unsaturated fatty acids in the rumen of ruminants, industrial hydrogenation (used to produce semi-liquid and solid fats; can be used to produce margarine, shortening, biscuits, etc.), deodorization of unsaturated vegetable oils (or occasionally fish oils) with a high content of polyunsaturated fatty acids (a necessary step of refining), and heating and frying oil at excessively high temperatures (>220 °C) [123].

TFA does not play a positive role in any vital functions. On the contrary, the intake of TFA may harm human health. Evidence suggests that ruminant-derived TFA has similar adverse effects on blood lipids and lipoproteins as TFA from industrial sources. Sufficient evidence is still needed to reveal whether a difference exists between equivalent amounts of ruminant and industrially produced TFA in terms of risk of CHD [123]. *Trans*-MUFA is the most common TFA in the human diet. A few clinical trials with normotensive subjects proved that *trans*-MUFA from hydrogenated oil has no effect on systolic or diastolic blood pressure [124]. Prospective cohort studies showed that a consistent relationship exists between higher TFA intake and increased risk of CHD. Conversely, a daily intake of 3.6 g of TFA from milk fat for five weeks did not affect blood pressure or isobaric arterial elasticity [125].

According to population nutrient intake goals from the World Health Organization (WHO)/FAO, the intake of TFA should constitute less than 1% of total energy. For pregnancy and lactation, the lowest possible intake of industrially-produced TFAs is required. According to the EFSA, TFA in the diet is provided by several sources that contain essential FAs and other nutrients [124], Therefore, the EFSA panel concluded that the intake of TFA should be sufficiently reduced within a nutritionally adequate diet to lower the intake of TFA while ensuring the nutrient intake [124]. The 2015–2020 Dietary Guidelines for Americans emphasize that individuals should reduce their intake of *trans* fatty acid to as low as possible by limiting their consumption of foods that contain small quantities of natural TFA from the diet. In the United Kingdom, the recommended intake of TFA is less than 2% of total daily energy or 5 g/day.

The TFA index is currently used in seaweed [90], lamb [45], milk [78], fish [35], and plant oil [32,33,110]. Skałecki et al. compared the fatty acid profiles of Prussian Carp fish (*Carassius gibelio*) fillets with and without skin; the share of TFA was the same in both types [35]. Mishra and Sharma monitored the changes occurring in rice bran oil and its blend with sunflower oil during repeated frying cycles of potato chips with different moisture contents (0.5% and 64.77%) [110]. The results showed that blended oil was better when used to fry dried potato chips, as TFA was the lowest after deep fat frying (increased from 1.15% to 1.80%) [110]. Detailed information of the literature related to TFA is listed in Table 11.

Materials		TFA Value	Reference
Red seaweed	Gelidium microdon Pterocladiella capillacea	$\begin{array}{c} 1.34\% \pm 0.20\% \\ 1.47\% \pm 0.09\% \end{array}$	[90] [90]
Brown seaweed	Ulva compressa Ulva rigida	$\begin{array}{l} 7.35\% \pm 0.63\% \\ 4.89\% \pm 0.26\% \end{array}$	[90] [90]
Plant oil	Palm stearin Rice bran oil Sunflower oil	0.6% 1.27–2.91% 0.2%, 0.84–1.71%	[32] [110] [32,33]
Fish	Carassius gibelio	1.06% ± 0.06%, 10.58–37.15 mg/100 g	[35]
Meat	Lamb (Barbarine lamb)	2.23-2.83%	[45]
Dairy products	Curd of cow (Middle Rhodopes) Milk of cow (Middle Rhodopes)	340–1090 mg/100 g 110–210 mg/100 g	[78] [78]

Table 11. Application of TFA in fatty acid evaluation \*.

TFA: Trans fatty acid; \* literature from 2000 until April/2020.

# 3. Conclusions

In this review, we summarized 10 FA indices that have been commonly used in the literature to characterized FA composition. Among them, PUFA/SFA, IA, IT, HH, HPI, and UI are the most frequently used indices and are widely used to evaluate a variety of research materials, mainly related to CVH. PUFA/SFA is a basic index that simply considers  $\Sigma$ PUFA and  $\Sigma$ SFA. IA, IT, HH, HPI, and UI were derived based on revising PUFA/SFA, which consider the contribution of different molecular species of SFA, as well as MUFA. However, all of these six indices do not reflect the influence of different molecular species of PUFA. For instance, n-3 PUFA and n-6 PUFA exhibit different effects on CVH. EPA + DHA and FLQ are used in the analysis of fish or shellfish, which are rich in n-3 PUFA. The LA/ALA ratio is an important index for baby food and infant formula. TFA is an indicator of food safety because it has a negative effect on many vital functions. Due to the lack of systematic integration of clinical evidence and literature data related to FA, suggesting ideas and proposals for the update of indices is difficult. Besides, CVH is the main assessment of FA indices used at present. As FA functions continue to be revealed, more indices that can be used for other diseases are expected.

With the present review, we aimed to help researchers evaluate the nutritional value of FAs and to explore their potential usage in disease prevention and treatment, and to help newcomers to the field

of FA analysis to quickly and accurately select appropriate indices. The human body is complex, so a reasonable selection of indices can help researchers to more comprehensively evaluate the research materials. The purpose of using an index is only to assess the potential nutritional and/or medicinal value of the research materials; they should not be considered gold standards. The indices should not be used indiscriminately, and the results obtained with the indices should be interpreted with caution. After a reasonable assessment using the indices, a more systematic and complex research process should be used to reach a conclusion about the nutritional effect of the research object on the human body. We recommend that researchers apply these indices to help compare several research objects to select one or more objects of interest for further in-depth research.

Author Contributions: Conceptualization, H.L. and J.C.; writing—original draft preparation, J.C.; writing—review and editing, H.L.; funding acquisition, H.L. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the National Natural Science Foundation of China, grant number 81973433, and the Marine S&T Fund of Shandong Province for Pilot National Laboratory for Marine Science and Technology (Qingdao), grant number 2018SDKJ0405.

Conflicts of Interest: The authors declare no conflicts of interest.

#### Abbreviations

ALA	α-linolenic acid
AS	Atherosclerosis
CHD	Coronary heart disease
CLA	Conjugated fatty acids
CVD	Cardiovascular disease
CVH	Cardiovascular health
DHA	Docosahexaenoic acid
EFSA	European Food Safety Authority
EPA	Eicosapentaenoic acid
FA	Fatty acid
FAO	Food and Agriculture Organization
FDA	Food and Drug Administration
FLQ	Fish lipid quality/flesh lipid quality
FSANZ	Food Standards Australia New Zealand
HH	Hypocholesterolemic/hypercholesterolemic ratio
HPI	Health-promoting index
IA	Index of atherogenicity
IT	Index of thrombogenicity
LA	Linoleic acid
LDL-C	Low-density lipoprotein cholesterol
LDLR	Low-density lipoprotein receptors
MS	Multiple sclerosis
MUFA	Monounsaturated fatty acid
PPAR-γ	Peroxisome proliferators-activated receptor-gamma
PUFA	Polyunsaturated fatty acid
SFA	Saturated fatty acid
TFA	Trans fatty acid
TG	Triglycerides
UFA	Unsaturated fatty acid
UI	Unsaturation index
WHO	World Health Organization

## References

- Rimm, E.B.; Appel, L.J.; Chiuve, S.E.; Djoussé, L.; Engler, M.B.; Kris-Etherton, P.M.; Mozaffarian, D.; Siscovick, D.S.; Lichtenstein, A.H. Seafood long-chain n-3 polyunsaturated fatty acids and cardiovascular disease: A science advisory from the American Heart Association. *Circulation* 2018, 138, 35–47. [CrossRef] [PubMed]
- 2. Wu, H.; Xu, L.; Ballantyne, C.M. Dietary and pharmacological fatty acids and cardiovascular health. *J. Clin. Endocrinol. Metab.* **2020**, *105*, 1030–1045. [CrossRef] [PubMed]
- 3. Marangoni, F.; Agostoni, C.; Borghi, C.; Catapano, A.L.; Cena, H.; Ghiselli, A.; La Vecchia, C.; Lercker, G.; Manzato, E.; Pirillo, A. Dietary linoleic acid and human health: Focus on cardiovascular and cardiometabolic effects. *Atherosclerosis* **2020**, *292*, 90–98. [CrossRef] [PubMed]
- Naeini, Z.; Toupchian, O.; Vatannejad, A.; Sotoudeh, G.; Teimouri, M.; Ghorbani, M.; Nasli-Esfahani, E.; Koohdani, F. Effects of DHA-enriched fish oil on gene expression levels of p53 and NF-κB and PPAR-γ activity in PBMCs of patients with T2DM: A randomized, double-blind, clinical trial. *Nutr. Metab. Cardiovasc.* 2020, 30, 441–447. [CrossRef]
- 5. Bird, J.K.; Calder, P.C.; Eggersdorfer, M. The role of n-3 long chain polyunsaturated fatty acids in cardiovascular disease prevention, and interactions with statins. *Nutrients* **2018**, *10*, 775. [CrossRef]
- 6. Tomata, Y.; Larsson, S.C.; Hägg, S. Polyunsaturated fatty acids and risk of Alzheimer's disease: A Mendelian randomization study. *Eur. J. Nutr.* **2020**, *59*, 1763–1766. [CrossRef]
- 7. Langley, M.R.; Triplet, E.M.; Scarisbrick, I.A. Dietary influence on central nervous system myelin production, injury, and regeneration. *BBA-Mol. Basis Dis.* **2020**, *1866*, 165779. [CrossRef]
- 8. Zhou, Y.; Tao, X.; Wang, Z.; Feng, L.; Wang, L.; Liu, X.; Pan, R.; Liao, Y.; Chang, Q. Hippocampus metabolic disturbance and autophagy deficiency in olfactory bulbectomized rats and the modulatory effect of fluoxetine. *Int. J. Mol. Sci.* **2019**, *20*, 4282. [CrossRef]
- 9. Chang, J.P.; Chang, S.; Yang, H.; Chen, H.; Chien, Y.; Yang, B.; Su, H.; Su, K. Omega-3 polyunsaturated fatty acids in cardiovascular diseases comorbid major depressive disorder-results from a randomized controlled trial. *Brain Behav. Immun.* **2020**, *85*, 14–20. [CrossRef]
- 10. Konstantynowicz-Nowicka, K.; Berk, K.; Chabowski, A.; Kasacka, I.; Bielawiec, P.; Łukaszuk, B.; Harasim-Symbor, E. High-fat feeding in time-dependent manner affects metabolic routes leading to nervonic acid synthesis in NAFLD. *Int. J. Mol. Sci.* **2019**, *20*, 3829. [CrossRef]
- Chen, L.; Wang, Y.; Xu, Q.; Chen, S. Omega-3 fatty acids as a treatment for non-alcoholic fatty liver disease in children: A systematic review and meta-analysis of randomized controlled trials. *Clin. Nutr.* 2018, 37, 516–521. [CrossRef] [PubMed]
- 12. Chen, Y.; Chen, H.; Huang, B.; Chen, Y.; Chang, C. Polyphenol rich extracts from *Toona sinensis* bark and fruit ameliorate free fatty acid-induced lipogenesis through AMPK and LC3 pathways. *J. Clin. Med.* **2019**, *8*, 1664. [CrossRef] [PubMed]
- Wu, K.; Zhao, T.; Hogstrand, C.; Xu, Y.; Ling, S.; Chen, G.; Luo, Z. FXR-mediated inhibition of autophagy contributes to FA-induced TG accumulation and accordingly reduces FA-induced lipotoxicity. *Cell Commun. Signal.* 2020, 18, 1–16. [CrossRef] [PubMed]
- 14. Tobias, T.A.; Wood, L.G.; Rastogi, D. Carotenoids, fatty acids and disease burden in obese minority adolescents with asthma. *Clin. Exp. Allergy* **2019**, *49*, 838–846. [CrossRef]
- 15. Monga, N.; Sethi, G.S.; Kondepudi, K.K.; Naura, A.S. Lipid mediators and asthma: Scope of therapeutics. *Biochem. Pharmacol.* **2020**, in press. [CrossRef]
- Magnusson, J.; Ekström, S.; Kull, I.; Håkansson, N.; Nilsson, S.; Wickman, M.; Melén, E.; Risérus, U.; Bergström, A. Polyunsaturated fatty acids in plasma at 8 years and subsequent allergic disease. *J. Allergy Clin. Immunol.* 2018, 142, 510–516. [CrossRef]
- 17. Miranda-Gonçalves, V.; Lameirinhas, A.; Henrique, R.; Baltazar, F.; Jerónimo, C. The metabolic landscape of urological cancers: New therapeutic perspectives. *Cancer Lett.* **2020**, 477, 76–87. [CrossRef]
- 18. Tripathi, R.K.P. A perspective review on fatty acid amide hydrolase (FAAH) inhibitors as potential therapeutic agents. *Eur. J. Med. Chem.* **2020**, *188*, 111953. [CrossRef]
- 19. Vadell, A.K.; Bärebring, L.; Hulander, E.; Gjertsson, I.; Lindqvist, H.M.; Winkvist, A. Anti-inflammatory diet in rheumatoid arthritis (ADIRA)-a randomized, controlled crossover trial indicating effects on disease activity. *Am. J. Clin. Nutr.* **2020**, *111*, 1203–1213. [CrossRef]

- Dos Santos Simon, M.I.S.; Dalle Molle, R.; Silva, F.M.; Rodrigues, T.W.; Feldmann, M.; Forte, G.C.; Marostica, P.J.C. Antioxidant micronutrients and essential fatty acids supplementation on cystic fibrosis outcomes: A systematic review. J. Acad. Nutr. Diet. 2020, 120, 1016–1033. [CrossRef]
- 21. Kjølbæk, L.; Benítez-Páez, A.; Del Pulgar, E.M.G.; Brahe, L.K.; Liebisch, G.; Matysik, S.; Rampelli, S.; Vermeiren, J.; Brigidi, P.; Larsen, L.H. Arabinoxylan oligosaccharides and polyunsaturated fatty acid effects on gut microbiota and metabolic markers in overweight individuals with signs of metabolic syndrome: A randomized cross-over trial. *Clin. Nutr.* **2020**, *39*, 67–79. [CrossRef] [PubMed]
- 22. Song, Y.; Hogstrand, C.; Ling, S.; Chen, G.; Luo, Z. Creb-Pgc1α pathway modulates the interaction between lipid droplets and mitochondria and influences high fat diet-induced changes of lipid metabolism in the liver and isolated hepatocytes of yellow catfish. *J. Nutr. Biochem.* **2020**, *80*, 108364. [CrossRef] [PubMed]
- 23. Mozaffari, H.; Daneshzad, E.; Larijani, B.; Bellissimo, N.; Azadbakht, L. Dietary intake of fish, n-3 polyunsaturated fatty acids, and risk of inflammatory bowel disease: A systematic review and meta-analysis of observational studies. *Eur. J. Nutr.* **2020**, *59*, 1–17. [CrossRef] [PubMed]
- 24. Min, S.Y.; Learnard, H.; Kant, S.; Gealikman, O.; Rojas-Rodriguez, R.; DeSouza, T.; Desai, A.; Keaney, J.F.; Corvera, S.; Craige, S.M. Exercise rescues gene pathways involved in vascular expansion and promotes functional angiogenesis in subcutaneous white adipose tissue. *Int. J. Mol. Sci.* **2019**, *20*, 2046. [CrossRef]
- 25. Syren, M.; Turolo, S.; Marangoni, F.; Milani, G.P.; Edefonti, A.; Montini, G.; Agostoni, C. The polyunsaturated fatty acid balance in kidney health and disease: A review. *Clin. Nutr.* **2018**, *37*, 1829–1839. [CrossRef]
- 26. Khalid, A.; Siddiqui, A.J.; Huang, J.; Shamsi, T.; Musharraf, S.G. Alteration of serum free fatty acids are indicators for progression of pre-leukaemia diseases to leukaemia. *Sci. Rep.* **2018**, *8*, 1–10. [CrossRef]
- 27. Kumar, M.; Kumari, P.; Trivedi, N.; Shukla, M.K.; Gupta, V.; Reddy, C.; Jha, B. Minerals, PUFAs and antioxidant properties of some tropical seaweeds from Saurashtra coast of India. *J. Appl. Phycol.* **2011**, *23*, 797–810. [CrossRef]
- 28. Chan, P.T.; Matanjun, P. Chemical composition and physicochemical properties of tropical red seaweed, *Gracilaria changii*. *Food Chem.* **2017**, *221*, 302–310. [CrossRef]
- 29. Chen, Z.; Xu, Y.; Liu, T.; Zhang, L.; Liu, H.; Guan, H. Comparative studies on the characteristic fatty acid profiles of four different Chinese medicinal *Sargassum* seaweeds by GC-MS and chemometrics. *Mar. Drugs* **2016**, *14*, 68. [CrossRef]
- 30. Calabrò, S.; Cutrignelli, M.I.; Lo Presti, V.; Tudisco, R.; Chiofalo, V.; Grossi, M.; Infascelli, F.; Chiofalo, B. Characterization and effect of year of harvest on the nutritional properties of three varieties of white lupine (*Lupinus albus L.*). *J. Sci. Food Agric.* **2015**, *95*, 3127–3136. [CrossRef]
- 31. Chiofalo, B.; Lo Presti, V.; D'Agata, A.; Rao, R.; Ceravolo, G.; Gresta, F. Qualitative profile of degummed guar (*Cyamopsis tetragonoloba L.*) seeds grown in a Mediterranean area for use as animal feed. *J. Anim. Physiol. Anim. Nutr.* **2018**, *102*, 260–267. [CrossRef] [PubMed]
- 32. Farajzadeh Alan, D.; Naeli, M.H.; Naderi, M.; Jafari, S.M.; Tavakoli, H.R. Production of *trans*-free fats by chemical interesterified blends of palm stearin and sunflower oil. *Food Sci. Nutr.* **2019**, *7*, 3722–3730. [CrossRef] [PubMed]
- 33. Filip, S.; Hribar, J.; Vidrih, R. Influence of natural antioxidants on the formation of *trans* fatty acid isomers during heat treatment of sunflower oil. *Eur. J. Lipid Sci. Technol.* **2011**, *113*, 224–230. [CrossRef]
- Rincón-Cervera, M.Á.; González-Barriga, V.; Romero, J.; Rojas, R.; López-Arana, S. Quantification and distribution of omega-3 fatty acids in south pacific fish and shellfish species. *Foods* 2020, *9*, 233. [CrossRef] [PubMed]
- 35. Skałecki, P.; Kaliniak-Dziura, A.; Domaradzki, P.; Florek, M.; Kępka, M. Fatty acid composition and oxidative stability of the lipid fraction of skin-on and skinless fillets of prussian carp (*Carassius gibelio*). *Animals* **2020**, 10, 778. [CrossRef]
- Fernandes, C.E.; Da Silva Vasconcelos, M.A.; de Almeida Ribeiro, M.; Sarubbo, L.A.; Andrade, S.A.C.; de Melo Filho, A.B. Nutritional and lipid profiles in marine fish species from Brazil. *Food Chem.* 2014, 160, 67–71. [CrossRef]
- 37. Hosseini, H.; Mahmoudzadeh, M.; Rezaei, M.; Mahmoudzadeh, L.; Khaksar, R.; Khosroshahi, N.K.; Babakhani, A. Effect of different cooking methods on minerals, vitamins and nutritional quality indices of *kutum roach (Rutilus frisii kutum)*. *Food Chem.* **2014**, *148*, 86–91. [CrossRef]

- Sreelakshmi, K.R.; Rehana, R.; Renjith, R.K.; Sarika, K.; Greeshma, S.S.; Minimol, V.A.; Ashokkumar, K.; Ninan, G. Quality and shelf life assessment of puffer fish (*Lagocephalus guentheri*) fillets during chilled storage. *J. Aquat. Food Prod. Technol.* 2019, 28, 25–37. [CrossRef]
- 39. Tonial, I.B.; Oliveira, D.F.; Coelho, A.R.; Matsushita, M.; Coró, F.A.G.; De Souza, N.E.; Visentainer, J.V. Quantification of essential fatty acids and assessment of the nutritional quality indexes of lipids in tilapia alevins and juvenile tilapia fish (*Oreochromis niloticus*). *J. Food Res.* **2014**, *3*, 105–114. [CrossRef]
- 40. Rombenso, A.N.; Trushenski, J.T.; Schwarz, M.H. Fish oil replacement in feeds for juvenile Florida Pompano: Composition of alternative lipid influences degree of tissue fatty acid profile distortion. *Aquaculture* **2016**, 458, 177–186. [CrossRef]
- 41. Hao, L.; Xiang, Y.; Degen, A.; Huang, Y.; Niu, J.; Sun, L.; Chai, S.; Zhou, J.; Ding, L.; Long, R. Adding heat-treated rapeseed to the diet of yak improves growth performance and tenderness and nutritional quality of the meat. *Anim. Sci. J.* **2019**, *90*, 1177–1184. [CrossRef] [PubMed]
- 42. Castro, T.; Cabezas, A.; De la Fuente, J.; Isabel, B.; Manso, T.; Jimeno, V. Animal performance and meat characteristics in steers reared in intensive conditions fed with different vegetable oils. *Animal* **2016**, *10*, 520–530. [CrossRef]
- 43. Majdoub-Mathlouthi, L.; Saïd, B.; Kraiem, K. Carcass traits and meat fatty acid composition of Barbarine lambs reared on rangelands or indoors on hay and concentrate. *Animal* **2015**, *9*, 2065–2071. [CrossRef]
- 44. Realini, C.E.; Pérez-Juan, M.; Gou, P.; Díaz, I.; Sárraga, C.; Gatellier, P.; García-Regueiro, J.A. Characterization of *Longissimus thoracis, Semitendinosus* and *Masseter* muscles and relationships with technological quality in pigs. 2. Composition of muscles. *Meat Sci.* **2013**, *94*, 417–423. [CrossRef] [PubMed]
- 45. Brogna, D.M.; Nasri, S.; Salem, H.B.; Mele, M.; Serra, A.; Bella, M.; Priolo, A.; Makkar, H.; Vasta, V. Effect of dietary saponins from *Quillaja saponaria L.* on fatty acid composition and cholesterol content in muscle *Longissimus dorsi* of lambs. *Animal* **2011**, *5*, 1124–1130. [CrossRef] [PubMed]
- Correa, L.B.; Zanetti, M.A.; Del Claro, G.R.; de Melo, M.P.; Rosa, A.F.; Netto, A.S. Effect of supplementation of two sources and two levels of copper on lipid metabolism in Nellore beef cattle. *Meat Sci.* 2012, *91*, 466–471. [CrossRef] [PubMed]
- Fernández, M.; Ordóñez, J.A.; Cambero, I.; Santos, C.; Pin, C.; de la Hoz, L. Fatty acid compositions of selected varieties of Spanish dry ham related to their nutritional implications. *Food Chem.* 2007, 101, 107–112. [CrossRef]
- 48. Winiarska-Mieczan, A.; Kwiecień, M.; Kwiatkowska, K.; Baranowska-Wójcik, E.; Szwajgier, D.; Zaricka, E. Fatty acid profile, antioxidative status and dietary value of the breast muscle of broiler chickens receiving glycine-Zn chelates. *Anim. Prod. Sci.* **2020**, *60*, 1095–1102. [CrossRef]
- 49. Alvarenga, A.; Sousa, R.V.; Parreira, G.G.; Chiarini-Garcia, H.; Almeida, F. Fatty acid profile, oxidative stability of pork lipids and meat quality indicators are not affected by birth weight. *Animal* **2014**, *8*, 660–666. [CrossRef]
- Mir, N.A.; Tyagi, P.K.; Biswas, A.K.; Tyagi, P.K.; Mandal, A.B.; Kumar, F.; Sharma, D.; Biswas, A.; Verma, A.K. Inclusion of flaxseed, broken rice, and distillers dried grains with solubles (DDGS) in broiler chicken ration alters the fatty acid profile, oxidative stability, and other functional properties of meat. *Eur. J. Lipid Sci. Technol.* 2018, 120, 1700470. [CrossRef]
- 51. Turner, T.; Hessle, A.; Lundström, K.; Pickova, J. Silage-concentrate finishing of bulls versus silage or fresh forage finishing of steers: Effects on fatty acids and meat tenderness. *Acta Agric. Scand.* **2011**, *61*, 103–113. [CrossRef]
- 52. Lorenzo, J.M.; Crecente, S.; Franco, D.; Sarriés, M.V.; Gómez, M. The effect of livestock production system and concentrate level on carcass traits and meat quality of foals slaughtered at 18 months of age. *Animal* **2014**, *8*, 494–503. [CrossRef] [PubMed]
- 53. Pires, M.A.; Rodrigues, I.; Barros, J.C.; Carnauba, G.; de Carvalho, F.A.; Trindade, M.A. Partial replacement of pork fat by *Echium* oil in reduced sodium bologna sausages: Technological, nutritional and stability implications. *J. Sci. Food Agric.* **2020**, *100*, 410–420. [CrossRef] [PubMed]
- 54. Sinanoglou, V.J.; Koutsouli, P.; Fotakis, C.; Sotiropoulou, G.; Cavouras, D.; Bizelis, I. Assessment of lactation stage and breed effect on sheep milk fatty acid profile and lipid quality indices. *Dairy Sci. Technol.* **2015**, *95*, 509–531. [CrossRef]
- 55. Mierliță, D. Effects of diets containing hemp seeds or hemp cake on fatty acid composition and oxidative stability of sheep milk. *S. Afr. J. Anim. Sci.* **2018**, *48*, 504–515. [CrossRef]

- 56. Nantapo, C.; Muchenje, V.; Hugo, A. Atherogenicity index and health-related fatty acids in different stages of lactation from Friesian, Jersey and Friesian × Jersey cross cow milk under a pasture-based dairy system. *Food Chem.* **2014**, *146*, 127–133. [CrossRef]
- 57. Bonanno, A.; Di Grigoli, A.; Mazza, F.; De Pasquale, C.; Giosuè, C.; Vitale, F.; Alabiso, M. Effects of ewes grazing sulla or ryegrass pasture for different daily durations on forage intake, milk production and fatty acid composition of cheese. *Animal* **2016**, *10*, 2074–2082. [CrossRef]
- Otero, P.; López-Martínez, M.I.; García-Risco, M.R. Application of pressurized liquid extraction (PLE) to obtain bioactive fatty acids and phenols from *Laminaria ochroleuca* collected in Galicia (NW Spain). *J. Pharm. Biomed.* 2019, 164, 86–92. [CrossRef]
- 59. Dellatorre, F.G.; Avaro, M.G.; Commendatore, M.G.; Arce, L.; de Vivar, M.E.D. The macroalgal ensemble of Golfo Nuevo (Patagonia, Argentina) as a potential source of valuable fatty acids for nutritional and nutraceutical purposes. *Algal Res.* **2020**, *45*, 101726. [CrossRef]
- 60. Rahmouni, N.; Pinto, D.C.; Santos, S.A.; Beghidja, N.; Silva, A.M. Lipophilic composition of *Scabiosa stellata L*.: An underexploited plant from Batna (Algeria). *Chem. Pap.* **2018**, *72*, 753–762. [CrossRef]
- 61. Pandey, S.; Patel, M.K.; Mishra, A.; Jha, B. Physio-biochemical composition and untargeted metabolomics of cumin (*Cuminum cyminum L.*) make it promising functional food and help in mitigating salinity stress. *PLoS ONE* **2015**, *10*, e0144469. [CrossRef]
- 62. Ratusz, K.; Symoniuk, E.; Wroniak, M.; Rudzińska, M. Bioactive Compounds, nutritional quality and oxidative stability of cold-pressed Camelina (*Camelina sativa L.*) oils. *Appl. Sci.* **2018**, *8*, 2606. [CrossRef]
- 63. Wu, Z.X.; Hu, X.P.; Zhou, D.Y.; Tan, Z.F.; Liu, Y.X.; Xie, H.K.; Rakariyatham, K.; Shahidi, F. Seasonal variation of proximate composition and lipid nutritional value of two species of scallops (*Chlamys farreri* and *Patinopecten yessoensis*). *Eur. J. Lipid Sci. Technol.* **2019**, *121*, 1800493. [CrossRef]
- 64. Akintola, S.L. Effects of smoking and sun-drying on proximate, fatty and amino acids compositions of southern pink shrimp (*Penaeus notialis*). *J. Food Sci. Technol.* **2015**, *52*, 2646–2656. [CrossRef] [PubMed]
- Łuczyńska, J.; Paszczyk, B. Health risk assessment of heavy metals and lipid quality indexes in freshwater fish from lakes of Warmia and Mazury region, Poland. *Int. J. Environ. Res. Public Health* 2019, *16*, 3780. [CrossRef]
- 66. Łuczyńska, J.; Paszczyk, B.; Nowosad, J.; Łuczyński, M.J. Mercury, fatty acids content and lipid quality indexes in muscles of freshwater and marine fish on the polish market. Risk assessment of fish consumption. *Int. J. Environ. Res. Public Health* **2017**, *14*, 1120. [CrossRef]
- 67. González-Félix, M.L.; Maldonado-Othón, C.A.; Perez-Velazquez, M. Effect of dietary lipid level and replacement of fish oil by soybean oil in compound feeds for the shortfin corvina (*Cynoscion parvipinnis*). *Aquaculture* **2016**, 454, 217–228. [CrossRef]
- Monteiro, M.; Matos, E.; Ramos, R.; Campos, I.; Valente, L.M. A blend of land animal fats can replace up to 75% fish oil without affecting growth and nutrient utilization of European seabass. *Aquaculture* 2018, 487, 22–31. [CrossRef]
- 69. Benhissi, H.; García-Rodríguez, A.; de Heredia, I.B. The effects of rapeseed cake intake during the finishing period on the fatty-acid composition of the *longissimus* muscle of Limousin steers and changes in meat colour and lipid oxidation during storage. *Anim. Prod. Sci.* **2020**, *60*, 1103–1110. [CrossRef]
- Subhadra, B.; Lochmann, R.; Rawles, S.; Chen, R. Effect of dietary lipid source on the growth, tissue composition and hematological parameters of largemouth bass (*Micropterus salmoides*). *Aquaculture* 2006, 255, 210–222. [CrossRef]
- Bouzgarrou, O.; El Mzougui, N.; Sadok, S. Smoking and polyphenols' addition to improve freshwater mullet (*Mugil cephalus*) fillets' quality attributes during refrigerated storage. *Int. J. Food Sci. Technol.* 2016, 51, 268–277. [CrossRef]
- 72. Dal Bosco, A.; Mugnai, C.; Roscini, V.; Castellini, C. Fillet fatty acid composition, estimated indexes of lipid metabolism and oxidative status of wild and farmed brown trout (*Salmo trutta L.*). *Ital. J. Food Sci.* **2013**, *25*, 83–89.
- Senso, L.; Suárez, M.D.; Ruiz-Cara, T.; García-Gallego, M. On the possible effects of harvesting season and chilled storage on the fatty acid profile of the fillet of farmed gilthead sea bream (*Sparus aurata*). *Food Chem.* 2007, 101, 298–307. [CrossRef]

- Ghafari, H.; Rezaeian, M.; Sharifi, S.D.; Khadem, A.A.; Afzalzadeh, A. Effects of dietary sesame oil on growth performance and fatty acid composition of muscle and tail fat in fattening Chaal lambs. *Anim. Feed. Sci. Technol.* 2016, 220, 216–225. [CrossRef]
- 75. Peiretti, P.G.; Masoero, G.; Meineri, G. Effects of replacing palm oil with maize oil and *Curcuma longa* supplementation on the performance, carcass characteristics, meat quality and fatty acid profile of the perirenal fat and muscle of growing rabbits. *Animal* **2011**, *5*, 795–801. [CrossRef]
- Salvatori, G.; Pantaleo, L.; Di Cesare, C.; Maiorano, G.; Filetti, F.; Oriani, G. Fatty acid composition and cholesterol content of muscles as related to genotype and vitamin E treatment in crossbred lambs. *Meat Sci.* 2004, 67, 45–55. [CrossRef]
- 77. Wójciak, K.M.; Stasiak, D.M.; Ferysiuk, K.; Solska, E. The influence of sonication on the oxidative stability and nutritional value of organic dry-fermented beef. *Meat Sci.* **2019**, *148*, 113–119. [CrossRef]
- 78. Ivanova, S.; Angelov, L. Assessment of the content of dietary *trans* fatty acids and biologically active substances in cow's milk and curd. *Generations* **2017**, *4*, 5.
- Bodas, R.; Manso, T.; Mantecon, A.R.; Juarez, M.; De la Fuente, M.A.; Gómez-Cortés, P. Comparison of the fatty acid profiles in cheeses from ewes fed diets supplemented with different plant oils. *J. Agric. Food Chem.* 2010, *58*, 10493–10502. [CrossRef]
- Casamassima, D.; Nardoia, M.; Palazzo, M.; Vizzarri, F.; D Alessandro, A.G.; Corino, C. Effect of dietary extruded linseed, verbascoside and vitamin E supplements on yield and quality of milk in Lacaune ewes. *J. Dairy Res.* 2014, *81*, 485–493. [CrossRef]
- Kholif, A.E.; Morsy, T.A.; Abd El Tawab, A.M.; Anele, U.Y.; Galyean, M.L. Effect of supplementing diets of Anglo-Nubian goats with soybean and flaxseed oils on lactational performance. *J. Agric. Food Chem.* 2016, 64, 6163–6170. [CrossRef] [PubMed]
- Kholif, A.E.; Gouda, G.A.; Olafadehan, O.A.; Abdo, M.M. Effects of replacement of Moringa oleifera for berseem clover in the diets of Nubian goats on feed utilisation, and milk yield, composition and fatty acid profile. *Animal* 2018, 12, 964–972. [CrossRef] [PubMed]
- Rutkowska, J.; Białek, M.; Bagnicka, E.; Jarczak, J.; Tambor, K.; Strzałkowska, N.; Jóźwik, A.; Krzyżewski, J.; Adamska, A.; Rutkowska, E. Effects of replacing extracted soybean meal with rapeseed cake in corn grass silage-based diet for dairy cows. J. Dairy Res. 2015, 82, 161–168. [CrossRef] [PubMed]
- Esposito, G.; Masucci, F.; Napolitano, F.; Braghieri, A.; Romano, R.; Manzo, N.; Di Francia, A. Fatty acid and sensory profiles of Caciocavallo cheese as affected by management system. *J. Dairy Sci.* 2014, 97, 1918–1928. [CrossRef] [PubMed]
- 85. Yurchenko, S.; Sats, A.; Tatar, V.; Kaart, T.; Mootse, H.; Jõudu, I. Fatty acid profile of milk from Saanen and Swedish Landrace goats. *Food Chem.* **2018**, 254, 326–332. [CrossRef] [PubMed]
- Ahmad, N.; Manzoor, M.F.; Shabbir, U.; Ahmed, S.; Ismail, T.; Saeed, F.; Nisa, M.; Anjum, F.M.; Hussain, S. Health lipid indices and physicochemical properties of dual fortified yogurt with extruded flaxseed omega fatty acids and fibers for hypercholesterolemic subjects. *Food Sci. Nutr.* 2019, *8*, 273–280. [CrossRef]
- Salles, M.S.; D Abreu, L.F.; Júnior, L.C.R.; César, M.C.; Guimarães, J.G.; Segura, J.G.; Rodrigues, C.; Zanetti, M.A.; Pfrimer, K.; Netto, A.S. Inclusion of sunflower oil in the bovine diet improves milk nutritional profile. *Nutrients* 2019, *11*, 481. [CrossRef]
- Vargas-Bello-Pérez, E.; Íñiguez-González, G.; Fehrmann-Cartes, K.; Toro-Mujica, P.; Garnsworthy, P.C. Influence of fish oil alone or in combination with hydrogenated palm oil on sensory characteristics and fatty acid composition of bovine cheese. *Anim. Feed Sci. Technol.* 2015, 205, 60–68. [CrossRef]
- 89. Pittau, D.; Panzalis, R.; Spanu, C.; Scarano, C.; De Santis, E.P. Survey on the fatty acids profile of fluid goat milk. *Ital. J. Food Saf.* **2013**, *2*, 33. [CrossRef]
- 90. Paiva, L.; Lima, E.; Neto, A.I.; Marcone, M.; Baptista, J. Health-promoting ingredients from four selected Azorean macroalgae. *Food Res. Int.* **2016**, *89*, 432–438. [CrossRef]
- 91. Santos-Silva, J.; Bessa, R.; Santos-Silva, F. Effect of genotype, feeding system and slaughter weight on the quality of light lambs: II. Fatty acid composition of meat. *Livest. Prod. Sci.* **2002**, *77*, 187–194. [CrossRef]
- Bonanno, A.; Di Grigoli, A.; Vitale, F.; Alabiso, M.; Giosuè, C.; Mazza, F.; Todaro, M. Legume grain-based supplements in dairy sheep diet: Effects on milk yield, composition and fatty acid profile. *Anim. Prod. Sci.* 2016, 56, 130–140. [CrossRef]

- Bobe, G.; Zimmerman, S.; Hammond, E.G.; Freeman, A.E.; Porter, P.A.; Luhman, C.M.; Beitz, D.C. Butter composition and texture from cows with different milk fatty acid compositions fed fish oil or roasted soybeans. *J. Dairy Sci.* 2007, *90*, 2596–2603. [CrossRef] [PubMed]
- 94. Chen, S.; Bobe, G.; Zimmerman, S.; Hammond, E.G.; Luhman, C.M.; Boylston, T.D.; Freeman, A.E.; Beitz, D.C. Physical and sensory properties of dairy products from cows with various milk fatty acid compositions. *J. Agric. Food Chem.* 2004, *52*, 3422–3428. [CrossRef] [PubMed]
- 95. Giorgio, D.; Di Trana, A.; Di Napoli, M.A.; Sepe, L.; Cecchini, S.; Rossi, R.; Claps, S. Comparison of cheeses from goats fed 7 forages based on a new health index. *J. Dairy Sci.* **2019**, *102*, 6790–6801. [CrossRef]
- 96. Colombo, M.L.; Rise, P.; Giavarini, F.; De Angelis, L.; Galli, C.; Bolis, C.L. Marine macroalgae as sources of polyunsaturated fatty acids. *Plant Foods Hum. Nutr.* **2006**, *61*, 64–69. [CrossRef]
- Poerschmann, J.; Spijkerman, E.; Langer, U. Fatty acid patterns in *Chlamydomonas* sp. as a marker for nutritional regimes and temperature under extremely acidic conditions. *Microb. Ecol.* 2004, 48, 78–89. [CrossRef]
- 98. Kumari, P.; Kumar, M.; Gupta, V.; Reddy, C.; Jha, B. Tropical marine macroalgae as potential sources of nutritionally important PUFAs. *Food Chem.* **2010**, *120*, 749–757. [CrossRef]
- Ghassemi-Golezani, K.; Farhangi-Abriz, S. Changes in oil accumulation and fatty acid composition of soybean seeds under salt stress in response to salicylic acid and jasmonic acid. *Russ. J. Plant Physiol.* 2018, 65, 229–236. [CrossRef]
- 100. Gomes-Laranjo, J.; Peixoto, F.; Sang, H.W.W.F.; Torres-Pereira, J. Study of the temperature effect in three chestnut (*Castanea sativa* Mill.) cultivars' behaviour. *J. Plant Physiol.* **2006**, *163*, 945–955. [CrossRef]
- Segura, J.; Escudero, R.; de Ávila, M.R.; Cambero, M.I.; López-Bote, C.J. Effect of fatty acid composition and positional distribution within the triglyceride on selected physical properties of dry-cured ham subcutaneous fat. *Meat Sci.* 2015, 103, 90–95. [CrossRef] [PubMed]
- 102. Rodríguez, M.; García-García, R.M.; Arias-Álvarez, M.; Millán, P.; Febrel, N.; Formoso-Rafferty, N.; López-Tello, J.; Lorenzo, P.L.; Rebollar, P.G. Improvements in the conception rate, milk composition and embryo quality of rabbit does after dietary enrichment with n-3 polyunsaturated fatty acids. *Animal* 2018, 12, 2080–2088. [CrossRef] [PubMed]
- 103. Lin, H.Z.; Liu, Y.J.; He, J.G.; Zheng, W.H.; Tian, L.X. Alternative vegetable lipid sources in diets for grouper, *Epinephelus coioides* (Hamilton): Effects on growth, and muscle and liver fatty acid composition. *Aquac. Res.* 2007, 38, 1605–1611. [CrossRef]
- 104. Larsson, T.; Koppang, E.O.; Espe, M.; Terjesen, B.F.; Krasnov, A.; Moreno, H.M.; Rørvik, K.; Thomassen, M.; Mørkøre, T. Fillet quality and health of Atlantic salmon (*Salmo salar L.*) fed a diet supplemented with glutamate. *Aquaculture* 2014, 426, 288–295. [CrossRef]
- 105. Mozanzadeh, M.T.; Marammazi, J.G.; Yavari, V.; Agh, N.; Mohammadian, T.; Gisbert, E. Dietary n-3 LC-PUFA requirements in silvery-black porgy juveniles (*Sparidentex hasta*). *Aquaculture* **2015**, *448*, 151–161. [CrossRef]
- 106. Xu, W.; Qian, Y.; Li, X.; Li, J.; Li, P.; Cai, D.; Liu, W. Effects of dietary biotin on growth performance and fatty acids metabolism in blunt snout bream, *Megalobrama amblycephala* fed with different lipid levels diets. *Aquaculture* 2017, 479, 790–797. [CrossRef]
- 107. Abrami, G.; Natiello, F.; Bronzi, P.; McKenzie, D.; Bolis, L.; Agradi, E. A comparison of highly unsaturated fatty acid levels in wild and farmed eels (*Anguilla anguilla*). *Comp. Biochem. Physiol.* **1992**, *101*, 79–81. [CrossRef]
- 108. Krajnović-Ozretic, M.; Najdek, M.; Ozretić, B. Fatty acids in liver and muscle of farmed and wild sea bass (*Dicentrarchus labrax L.*). *Comp. Biochem. Physiol.* **1994**, *109*, 611–617. [CrossRef]
- 109. Sharma, R.; Ahlawat, S.; Aggarwal, R.; Dua, A.; Sharma, V.; Tantia, M.S. Comparative milk metabolite profiling for exploring superiority of indigenous Indian cow milk over exotic and crossbred counterparts. *J. Food Sci. Technol.* 2018, 55, 4232–4243. [CrossRef]
- 110. Mishra, R.; Sharma, H.K. Effect of frying conditions on the physico-chemical properties of rice bran oil and its blended oil. *J. Food Sci. Technol.* **2014**, *51*, 1076–1084. [CrossRef]
- 111. Dietschy, J.M. Dietary fatty acids and the regulation of plasma low density lipoprotein cholesterol concentrations. *J. Nutr.* **1998**, *128*, 444–448. [CrossRef] [PubMed]
- 112. Ulbricht, T.; Southgate, D. Coronary heart disease: Seven dietary factors. Lancet 1991, 338, 985–992. [CrossRef]

- 113. Omri, B.; Chalghoumi, R.; Izzo, L.; Ritieni, A.; Lucarini, M.; Durazzo, A.; Abdouli, H.; Santini, A. Effect of dietary incorporation of linseed alone or together with tomato-red pepper mix on laying hens' egg yolk fatty acids profile and health lipid indexes. *Nutrients* **2019**, *11*, 813. [CrossRef] [PubMed]
- 114. Logue, J.A.; De Vries, A.L.; Fodor, E.; Cossins, A.R. Lipid compositional correlates of temperature-adaptive interspecific differences in membrane physical structure. *J. Exp. Biol.* **2000**, *203*, 2105–2115. [PubMed]
- 115. Burdge, G.C.; Jones, A.E.; Wootton, S.A. Eicosapentaenoic and docosapentaenoic acids are the principal products of α-linolenic acid metabolism in young men. *Br. J. Nutr.* **2002**, *88*, 355–363. [CrossRef] [PubMed]
- 116. Johnson, E.J.; Schaefer, E.J. Potential role of dietary n-3 fatty acids in the prevention of dementia and macular degeneration. *Am. J. Clin. Nutr.* **2006**, *83*, 1494–1498. [CrossRef]
- 117. Brenna, J.T.; Salem, N., Jr.; Sinclair, A.J.; Cunnane, S.C. α-Linolenic acid supplementation and conversion to n-3 long-chain polyunsaturated fatty acids in humans. *Prostaglandins Leukot. Essent. Fat. Acids* 2009, 80, 85–91. [CrossRef]
- 118. Wayland, M.M. Final determination regarding partially hydrogenated oils. Fed. Regist. 2015, 80, 116.
- Food and Drug Administration, H. Final Determination Regarding Partially Hydrogenated Oils. Notification; declaratory order; extension of compliance date. *Fed. Regist.* 2018, 83, 23358–233589.
- 120. Lock, A.L.; Corl, B.A.; Barbano, D.M.; Bauman, D.E.; Ip, C. The anticarcinogenic effect of *trans*-11 18: 1 is dependent on its conversion to *cis*-9, *trans*-11 CLA by Δ9-desaturase in rats. *J. Nutr.* 2004, 134, 2698–2704. [CrossRef]
- 121. Corl, B.A.; Barbano, D.M.; Bauman, D.E.; Ip, C. *cis-9, trans-*11 CLA derived endogenously from *trans-*11 18: 1 reduces cancer risk in rats. *J. Nutr.* **2003**, *133*, 2893–2900. [CrossRef] [PubMed]
- 122. Kritchevsky, D.; Tepper, S.A.; Wright, S.; Czarnecki, S.K.; Wilson, T.A.; Nicolosi, R.J. Conjugated linoleic acid isomer effects in atherosclerosis: Growth and regression of lesions. *Lipids* **2004**, *39*, 611. [CrossRef] [PubMed]
- 123. European Food Safety Authority (EFSA). Scientific opinion on dietary reference values for fats, including saturated fatty acids, polyunsaturated fatty acids, monounsaturated fatty acids, *trans* fatty acids, and cholesterol. *EFSA J.* **2010**, *8*, 1461.
- 124. European Food Safety Authority (EFSA). Opinion of the scientific panel on dietetic products, nutrition and allergies on a request from the commission related to the presence of *trans* fatty acids in foods and the effect on human health of the consumption of *trans* fatty acids. *EFSA J.* **2004**, *81*, 1–49.
- Raff, M.; Tholstrup, T.; Sejrsen, K.; Straarup, E.M.; Wiinberg, N. Diets rich in conjugated linoleic acid and vaccenic acid have no effect on blood pressure and isobaric arterial elasticity in healthy young men. *J. Nutr.* 2006, 136, 992–997. [CrossRef] [PubMed]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).