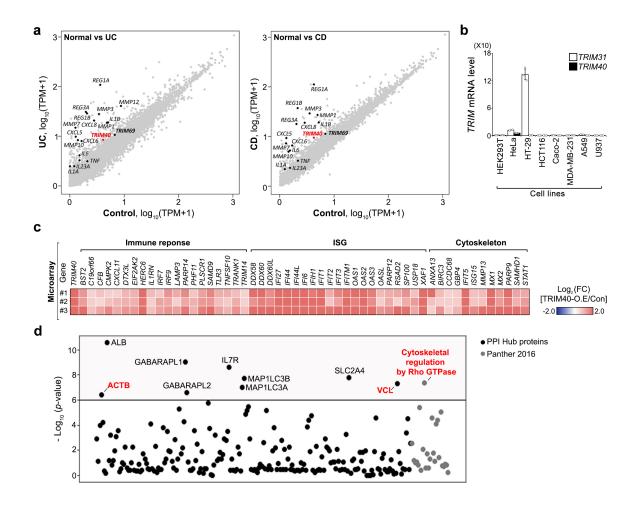
TRIM40 is a pathogenic driver of inflammatory bowel disease subverting intestinal barrier integrity Sujin Kang¹, Jaekyung Kim¹, Areum Park¹, Minsoo Koh¹, Wonji Shin¹, Gayoung Park¹, Taeyun A. Lee¹, Hyung Jin Kim¹, Heonjong Han^{1,2}, Yongbo Kim², Myung Kyung Choi¹, Jae Hyung Park³, Eunhye Lee¹, Hyun-Soo Cho¹, Hyun Woo Park³, Jae Hee Cheon^{4,5}, Sungwook Lee², Boyoun Park¹ **Supplementary Figures Supplementary Figures Supplementary Figure legends**



Supplementary Fig. 1. Aberrant TRIM40 upregulation is linked to IBD. a, Scatter plots representing the highly inducible signature genes associated with UC or CD (fold-change [FC] > 2) compared to healthy individuals (GSE117993). b, qPCR showing relative mRNA expression of TRIM40 and TRIM31 in various human cell lines (HEK293T [embryonic kidney], HeLa [cervix], HT-29 [colon], HCT116 [colon], Caco-2 [colon], MDA-MB-231 [breast], A549 [lung], and U937 [lymphocyte]). Data are representative of three independent experiments. c, Heat map representing relative log₂ values for fold-change of differentially up-regulated genes measured by microarray in HT-29 cells stably expressing Myc-TRIM40 compared with control. Red or blue colors represent high or low fold-change, respectively. Microarray results were obtained in three independent experiments (#1~#3). d, Manhattan plots showing analysis and classification of TRIM40-interacting proteins. Co-immunoprecipitated proteins with anti-Myc antibody in HT-29 cells expressing control vector or Myc-TRIM40 were separated and visualized by SDS-PAGE and silver staining, and subsequently analyzed by liquid chromatography with tandem mass spectrometry (LC-MS/MS). Data were analyzed by using protein-protein interaction (PPI) Hub proteins and Panther 2016 database. Black or gray circles represent PPI Hub proteins or Panther 2016 results, respectively. Red-letters indicate the cytoskeleton regulation-related proteins. P values are determined by fisher's exact test.

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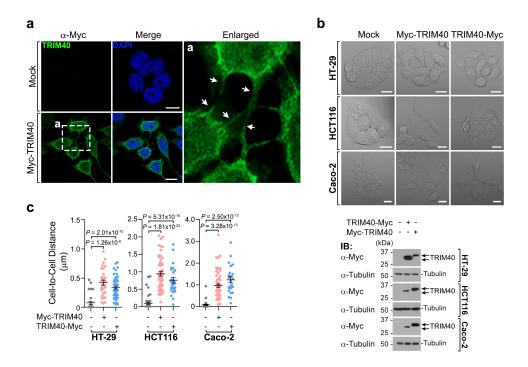
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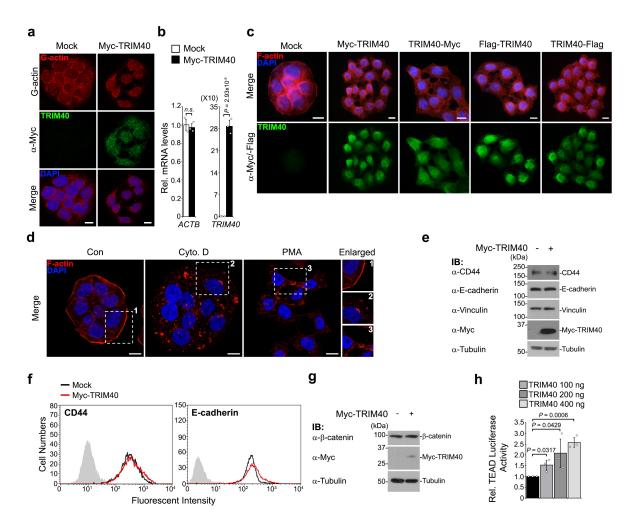
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Supplementary Fig. 2. TRIM40 overexpression disrupts cell-to-cell contacts. a, Confocal microscopic analysis of TRIM40 (green) in HT-29 cells expressing control vector and Myc-TRIM40. Nuclei were stained with DAPI (blue). Enlarged views of the regions denoted by the white dashed squares. White arrows represent filamentous stretches of TRIM40. Scale bars, 10 µm. b, Differential interference contrast images showing morphological deformation in HT-29, HCT116, and Caco-2 cells expressing Myc-TRIM40 or TRIM40-Myc compared with control. Scale bars, 10 µm. The degree of TRIM40 overexpression was analyzed by immunoblot (bottom). Tubulin was used as a loading control. c, Graphs showing comparison of cell-to-cell distance in different epithelial cell lines, HT-29, HCT116, or Caco-2 cells, stably expressing Myc-TRIM40 or control vector. HT-29, control (n = 33), Myc-TRIM40 (n = 29), and TRIM40-Myc (n = 55); HCT116, control (n = 58), Myc-TRIM40 (n = 56), and TRIM40-Myc (n = 58) 25); Caco-2, control (n = 34), Myc-TRIM40 (n = 65), and TRIM40-Myc (n = 23). Distances were analyzed by ImageJ software (V1.8.0). P values are determined by unpaired two-tailed t test. (n, biologically independent measurements of the distance between cells; mean \pm SD). All Data are representative of three independent experiments and source data are provided as a Source Data file.



Supplementary Fig. 3. TRIM40 overexpression affects cortical actin stability. a, Confocal microscopic analysis of G-actin distribution in HT-29 cells expressing control vector and Myc-TRIM40. Cells were stained with DNase I (G-actin, red) or anti-Myc antibodies (TRIM40, green), and nuclei were stained with DAPI (blue). Scale bars, 10 μm. b, qPCR showing relative mRNA levels of ACTB (β-actin) or TRIM40 in HT-29 cells expressing control vector and Myc-TRIM40. n.s.; not significant. c, Immunofluorescence assay of F-actin in HT-29 cells expressing Myc- or Flag-tagged at either N- or C-terminus of TRIM40. Nuclei were stained with DAPI (blue). Scale bars, 10 μm. d, Confocal microscopic analysis of F-actin in HT-29 cells after treatment with 0.25 µM cytochalasin D (Cyto. D) or 100 ng/ml phorbol 12-myristate 13-acetate (PMA) for 2h or 4 h, respectively. Cells were stained with phalloidin (F-actin, red) and nuclei were stained with DAPI (blue). Scale bars, 10 μm. e, Immunoblot showing protein expression levels of CD44, E-cadherin, or vinculin in HT-29 cells expressing control vector and Myc-TRIM40. Cell lysates were immunoblotted with indicated antibodies. Tubulin was used as a loading control. f, The cell surface expression level of junctional proteins, CD44 and Ecadherin, in HT-29 cells expressing control vector and Myc-TRIM40. Cell surface expression levels of CD44 and E-cadherin were assessed by flow cytometry analysis. Forward (FSC) and side scatter (SSC) were used for gating of mononuclear cells to exclude non-viable cells. g, β-

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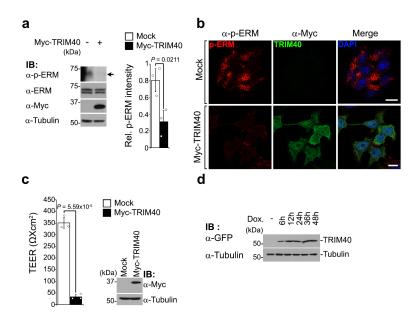
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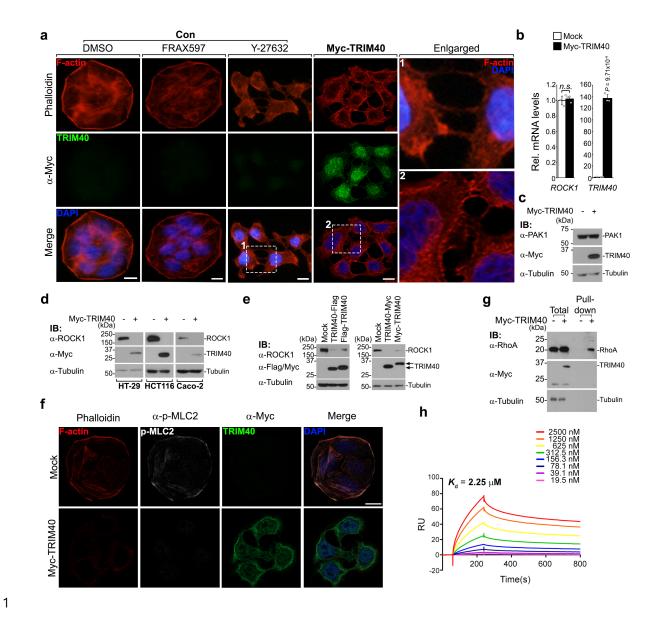
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catenin protein levels in HT-29 cells expressing control vector and Myc-TRIM40. Cell lysates were immunoblotted with the indicated antibodies. Tubulin was used as a loading control. h, Graph representing an increase in TEAD activity by TRIM40 overexpression. Transcriptional activity of TEAD measured by luciferase assay with 8 × GTIIC-reporter plasmid containing tandem TEAD-binding sites in 293A cells transiently expressing different concentrations of TRIM40 plasmid. P values are determined by unpaired two-tailed t test in $\bf b$ and $\bf h$. (n=3biological independent experiments, mean ± SD) All Data are representative of three independent experiments and source data are provided as a Source Data file.

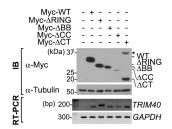


Supplementary Fig. 4. TRIM40 inhibits ERM phosphorylation. **a**, Immunoblots showing decreased phosphorylated ERM (p-ERM) levels in HT-29 cells expressing Myc-TRIM40 compared with the control vector. Quantification of the band intensity of p-ERM (right graph). Tubulin was used as a loading control. **b**, confocal fluorescent images of Myc-TRIM40 (green) and p-ERM (red) in HT-29 cells expressing control vector or Myc-TRIM40. Nuclei were stained with DAPI (blue). Scale bars, 10 μ m. **c**, Transepithelial electrical resistance (TEER) values of Caco-2 cells expressing control vector or Myc-TRIM40 grown on transwell inserts. Myc-TRIM40 overexpression in Caco-2 cells was confirmed by immunoblot analysis (right blots). Tubulin was used as a loading control. **d**, Dox-inducible GFP-TRIM40 overexpression in HT-29 cells was confirmed by immunoblot analysis. Tubulin was used as a loading control. *P* values are determined by unpaired two-tailed *t* test in **a** and **c**. (*n* = 3 biologically independent experiments, mean \pm SD) Data are representative of three independent experiments and source data are provided as a Source Data file.

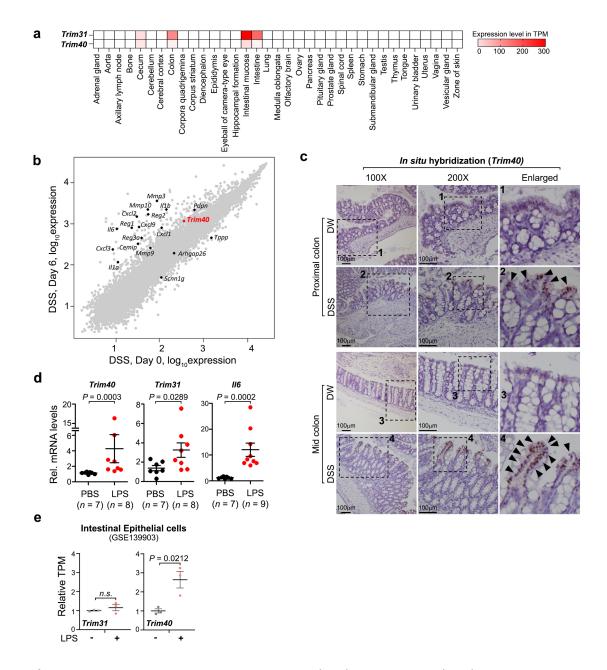


Supplementary Fig. 5. TRIM40 regulates RhoA-ROCK1 signaling by reducing ROCK1 protein levels. **a**, Immunofluorescence assay of F-actin distribution after inhibiting PAK1 or ROCK1. HT-29 cells were treated with PAK inhibitor (FRAX597, 5 μM) or ROCK1 inhibitor (Y-27632, 10 μM) for 4 h or 6 h, respectively. F-actin (red) and TRIM40 (green) were stained with phalloidin and anti-Myc antibodies, respectively. Nuclei were stained with DAPI (blue). Scale bars, 10 μm. Enlarged views of the regions denoted by the white dashed squares. **b**, qPCR showing relative mRNA levels of *ROCK1* and *TRIM40* in HT-29 cells stably expressing control vector or Myc-TRIM40. *P* values are determined by unpaired two-paired *t* test. (n = 3 biological independent experiments, mean ± SD) n.s.; not significant. **c**, PAK1 proteins levels in HT-29 cells stably expressing control vector or Myc-TRIM40. Cells were lysed and immunoblotted with the indicated antibodies. **d**, Evaluating ROCK1 protein levels in HT-29, HCT116, and Caco-2 cells expressing control vector or Myc-TRIM40. Cells were lysed and

then immunoblotted with the indicated antibodies. e, ROCK1 proteins levels in HT-29 cells expressing control vector, Myc- or Flag-tagged at either N- or C-terminus of TRIM40. Cells were lysed and then immunoblotted with the indicated antibodies. Tubulin was used as a loading control in c, d, e. f, Confocal microscopic analysis of F-actin and phosphorylated MLC2 (p-MLC2) in HT-29 cells expressing control vector or Myc-TRIM40. F-actin was stained with phalloidin (F-actin, red). p-MLC2 (white) and TRIM40 (green) were stained with anti-p-MLC2 and anti-Myc antibodies, respectively. Nuclei were stained with DAPI (blue). Scale bars, 10 µm. g, Rhotekin RBD (Rho-binding domain) pull down assay from HT-29 cells stably expressing control vector or Myc-TRIM40. The cell lysates were immunoprecipitated with glutathione resin, and the purified proteins were immunoblotted with the indicated antibodies. Tubulin was used as a loading control. h, Surface plasmon resonance (SPR) assay of the ROCK1 kinase domain (1~415 amino acids) concentration dependent binding to the TRIM40, representing a direct interaction of TRIM40 with ROCK1 ($K_d = 2.25 \mu M$). Data are representative of three independent experiments and source data are provided as a Source Data file.



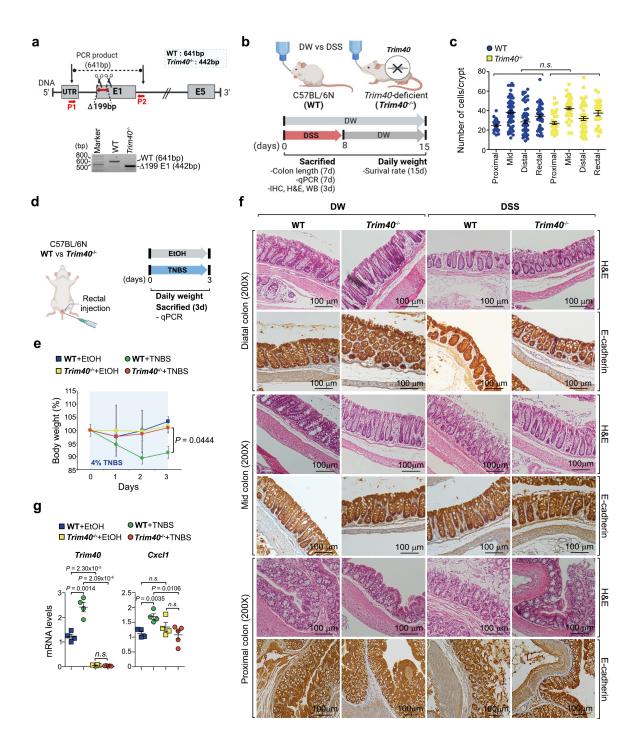
Supplementary Fig. 6. Protein and mRNA expression levels of wild-type TRIM40 and its deletion mutants. RT-PCR and immunoblot showing mRNA or protein expression levels of WT-TRIM40 or TRIM40 deletion mutants in HT-29 cells. HT-29 cells expressing control vector, WT-TRIM40, or TRIM40 deletion mutants were lysed and then immunoblotted with the indicated antibodies. PCR was performed with cDNA and the indicated primers. *; dimeric forms of TRIM40 Δ CT. Tubulin or *GAPDH* was used as a loading control. Data are representative of three independent experiments and source data are provided as a Source Data file.



Supplementary Fig. 7. *Trim40* mRNA expression levels are increased in the DSS- or LPS-induced inflammation. **a**, The gene expression level of *Trim40* and *Trim31* in mouse tissues from RNA-seq datasets of FANTOM5 project. **b**, Microarray analysis showing increased expression of *Trim40* in colon tissues from DSS-treated mice compared with control (GSE22307). **c**, RNAscope *in situ* hybridization (ISH) showing *Trim40* mRNA expression in the proximal and mid colon tissues from WT male mice after 2% DSS or DW administration for 7 days. Brown staining (black arrow heads) indicates positive *Trim40* mRNA. Black dashed squares are shown the enlarged region (#1-4). Scale bars, 100 μ m. **d**, qPCR showing relative mRNA levels of *Trim40*, *Trim31*, and *Il6* in distal colon from male mice intraperitoneally challenged with LPS (10 mg/kg body weight) (PBS group, n = 7; LPS group, n = 8 to 9). *P* values are determined by unpaired two-tailed t test (n, numbers of mice; mean t SD). **e**, The

cells of mice treated with PBS or LPS (GSE139903). P values are determined by unpaired two-tailed t test (n, numbers of samples; mean ± SEM) Data are representative of three independent experiments and source data are provided as a Source Data file.

gene expression level of Trim40 and Trim31 in RNA-seq dataset from intestinal epithelial



Supplementary Fig. 8. *Trim40*-dificient mice are protective to chemically induced colitis. **a**, A schematic representation showing CRISPR/Cas9-mediated gene-targeting strategy to generate $Trim40^{-/-}$ male mice. E; exon, P1/P2 and red arrows; PCR primers recognizing 5'-UTR or Intron 1 positioned in Trim40. Bottom gel, PCR products amplified from the targeted region of genomic DNA revealed genotypes of mice. **b**, Experimental design of DSS-induced colitis in WT and $Trim40^{-/-}$ male mice (image created with BioRender.com). **c**, Graph showing comparison of cell numbers in crypt from distal, mid, proximal, and rectal colon between WT (Proximal, n = 28; Mid, n = 71; Distal, n = 61; Rectal, n = 42) and $Trim40^{-/-}$ (Proximal, n = 40;

Mid, n = 46; Distal, n = 52; Rectal, n = 26) mice. P values are determined by unpaired two-1 2 tailed t test. (n, numbers of cells, mean ± SD) n.s.; not significant. d, Experimental design of TNBS-induced colitis in WT and Trim40^{-/-} male mice (image created with BioRender.com). e, 3 4 Relative loss of body weight in male mice for 3 days after 4% TNBS rectal injection. WT mice (EtOH group, n = 4; TNBS group, n = 4), $Trim40^{-/-}$ mice (EtOH group, n = 4; TNBS group, n = 5). 5 6 P values are determined by unpaired two-tailed t test. (n, numbers of mice, mean \pm SEM) f, Representative H&E staining and IHC staining of distal, mid, and proximal colon at day 3 after 7 DSS exposure. WT and *Trim40*^{-/-} male mice were administrated to 1% DSS in drinking water 8 for 3 days and euthanized on day 3. Histological changes and difference between WT and 9 *Trim40*^{-/-} mice were examined by H&E staining and IHC for E-cadherin. ($n \ge 3$ biological 10 independent experiments) g, qPCR showing relative mRNA levels of Trim40 or Cxcl1 in 4% 11 12 TNBS-induced colitis for 3 days. The expressions of *Trim40* or *Cxcl1* were analyzed in the distal colon tissue from WT male mice (EtOH group, n = 4; TNBS group, n = 4) and $Trim40^{-/-}$ 13 14 male mice (EtOH group, n = 4; TNBS group, n = 5). P values are determined by unpaired twotailed t test. (n, numbers of mice, mean \pm SD) n.s.; not significant. Source data are provided 15 16 as a Source Data file.

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