



OPEN

Combined effects of nucleotide-binding domain-like receptor protein 3 polymorphisms and environmental metals exposure on chronic kidney disease

Yu-Mei Hsueh^{1,2}, Wei-Jen Chen³, Ying-Chin Lin^{1,4,5}, Ya-Li Huang², Horng-Sheng Shiu⁶, Yuh-Feng Lin^{7,8}, Ru-Lan Hsieh^{9,10} & Hsi-Hsien Chen^{11,12}✉

Chronic inflammation is the cause of chronic kidney disease (CKD). The nucleotide-binding domain-like receptor protein 3 (NLRP3) inflammasome plays a vital role in the inflammation process and is associated with the regulatory effects of *NLRP3* gene polymorphisms. This study evaluated the association between *NLRP3* gene polymorphisms and CKD, and further explored whether the association of environmental metals with CKD varied by the *NLRP3* genotypes. A total of 218 CKD patients and 427 age- and sex-matched healthy controls were recruited in this clinic-based case-control study. Patients were identified as having CKD if their estimated glomerular filtration rate (eGFR) < 60 mL/min/1.73 m² and stage 3–5 for at least 3 months. We examined the genotypes of fifteen common single-nucleotide polymorphisms in *NLRP3* genes. Concentrations of total urinary arsenic were examined by summing of urinary inorganic arsenic species. Concentrations of selenium, cadmium, and lead were measured from blood samples. Associations between *NLRP3* polymorphisms, environmental metals exposure, and CKD were evaluated using multivariable logistic regression while controlling for confounders. We observed that the odds of carrying *NLRP3* rs4925650 GA/AA genotypes, *NLRP3* rs1539019 CA/AA genotypes, and *NLRP3* rs10157379 CT/TT genotypes were significantly higher among CKD cases compared to controls, with the adjusted odds ratio (95% confidence interval) were 1.54 (1.01–2.36), 1.56 (1.04–2.33), and 1.59 (1.05–2.38), respectively. The significant multiplicative interactions were identified between high levels of blood lead and *NLRP3* rs4925650 GA/AA genotypes; high levels of blood cadmium or low levels of plasma selenium and the *NLRP3* haplotype (rs4925648, rs4925650, rs12048215, and rs10754555) C-A-A-C multiplicatively interacted to increase the risk of CKD. Our results imply that *NLRP3* polymorphisms may play an important role in the development of environmental metals exposure related CKD.

Chronic kidney disease (CKD) affects 8–16% of the world's population¹, resulting in CKD being a common public health problem worldwide². Using estimated glomerular filtration rate (eGFR) < 60 mL/min/1.73 m² to

¹Department of Family Medicine, Wan Fang Hospital, Taipei Medical University, Taipei, Taiwan. ²Department of Public Health, School of Medicine, College of Medicine, Taipei Medical University, Taipei, Taiwan. ³Department of Medicine, Section of Epidemiology and Population Sciences, Baylor College of Medicine, Houston, TX, USA. ⁴Department of Family Medicine, School of Medicine, College of Medicine, Taipei Medical University, Taipei, Taiwan. ⁵Department of Geriatric Medicine, School of Medicine, College of Medicine, Taipei Medical University, Taipei, Taiwan. ⁶Department of Chinese Medicine, College of Medicine, Chang Gung University, Taoyuan, Taiwan. ⁷Graduate Institute of Clinical Medicine, School of Medicine, College of Medicine, Taipei Medical University, Taipei, Taiwan. ⁸Division of Nephrology, Department of Internal Medicine, Shuang Ho Hospital, New Taipei, Taiwan. ⁹Department of Physical Medicine and Rehabilitation, Shin Kong Wu Ho-Su Memorial Hospital, Taipei, Taiwan. ¹⁰Department of Physical Medicine and Rehabilitation, School of Medicine, College of Medicine, Taipei Medical University, Taipei, Taiwan. ¹¹Division of Nephrology, Department of Internal Medicine, School of Medicine, College of Medicine, Taipei Medical University, Taipei, Taiwan. ¹²Division of Nephrology, Department of Internal Medicine, Taipei Medical University Hospital, Taipei, Taiwan. ✉email: 570713@yahoo.com.tw

define CKD, the prevalence of CKD in Taiwan was 11.9%, and only 3.5% of patients aware their disease³. As the incidence of end-stage renal disease in Taiwan (426/10⁶ in 2012) was the highest globally⁴, CKD is an extremely important health issue in Taiwan.

Our previous study has reported a significantly increased odds of high total urinary arsenic levels and low plasma lycopene levels among the CKD cases compared to the health controls⁵. We also found an increased odds of high blood cadmium and lead levels among CKD cases compared to the controls, whereas a decreased odds of high plasma selenium levels were observed among CKD cases compared to the controls⁶. Recently, a study has shown that plasma concentrations of arsenic and lead were significantly related to the decline of renal function⁷. In addition, a study reported that increasing concentrations of blood cadmium and lead were associated with an increased risk of proteinuria and related to decreased eGFR⁸. These studies suggested that low levels of selenium and high levels of arsenic, cadmium, and lead may increase the risk of CKD. However, the mechanism underlying the effect of these metals on CKD remains unclear.

Inflammation is the cause of several kinds of kidney diseases, such as acute kidney injury and CKD⁹. The nucleotide-binding domain-like receptors 3 (NLRP3) inflammasome is a multi-protein complex that plays an important role in the inflammation process¹⁰. A previous study has indicated that NLRP3-induced inflammation may promote kidney inflammation and causes CKD¹¹. Also, a study has reported that the NLRP3 inflammasome may be involved in the pathogenesis of acute kidney injury, CKD, diabetic nephropathy, and crystal-related nephropathy¹². Recent evidence has been built to implicate arsenic, cadmium, lead, and selenium on NLRP3 related inflammatory. A study has reported that arsenic activates the NLRP3 inflammasome and induces inflammatory cell death¹³. However, another study showed that arsenic inhibits the secretion of interleukin (IL)-1 β and IL-18, which was caused by activating the NLRP3 inflammasomes in macrophages; thus, indicating that exposure to arsenic may affect inflammasome-mediated inflammation¹⁴. These inconsistent findings reveal an unclear relationship between arsenic exposure and NLRP3 inflammasomes. In addition to arsenic exposure, a carp experiment showed that cadmium exposure induced apoptosis of the anterior spleen and splenic lymphocytes by activating NLRP3¹⁵. Moreover, lead may activate the NLRP3 signaling pathway to cause oxidative stress and inflammation in chicken testicles, thereby reducing testicular function¹⁶. In contrast, selenium has a mitigating effect by modifying activation of the NLRP3 signaling pathway in chicken testes caused by lead¹⁶.

Variations in the *NLRP3* gene may affect mRNA stability and NLRP3 performance¹⁷. The *NLRP3* gene has nine exons within its 32.9 kb sequence¹⁷ and is located on chromosome 1q44¹⁸. There are 60 common single nucleotide polymorphisms (SNPs) that have been identified in the *NLRP3* gene¹⁹. Several studies have suggested that *NLRP3* polymorphisms were associated with the risk of cardiovascular diseases. A study has found that the 50-year-old subjects with *NLRP3* rs7512998 CC or CT genotypes had a higher blood pressure levels compared to those with TT genotype²⁰. In Chinese Han population, *NLRP3* rs10754556 CC genotype was associated with the occurrence of coronary artery disease compared to those with CG or GG genotypes²¹. The *NLRP3* rs4612666 T allele is associated with an increased risk of aorta sclerosis-like ischemic stroke²². To date, few studies have explored the association between *NLRP3* gene polymorphisms and CKD. The present study aimed to examine the association between *NLRP3* genotypes and CKD, and to further explore whether *NLRP3* gene polymorphisms may modify the associations of total urinary arsenic, blood cadmium and lead, and plasma selenium with CKD.

Materials and methods

Study subjects. This study was a clinic-based case-control study. In total, 218 clinically confirmed CKD patients and 427 age- and sex-matched controls who volunteered to participate in this study were recruited previously²³. All participants measured serum creatinine concentrations by isotope dilution mass spectrometry (IDMS). The eGFR (mL/min/1.73 m²) was calculated using the equation: $186.3 \times (\text{serum creatinine})^{-1.154} \times (\text{age})^{-0.203} \times (0.742 \text{ for females})$ ²⁴. CKD patients were clinically diagnosed at the Department of Internal Medicine/Nephrology, Taipei Medical University Hospital and Taipei Municipal Wan Fang Hospital, with an eGFR < 60 mL/min/1.73 m² for at least 3 months (stages 3–5) and without hemodialysis according to KDIGO 2012 Clinical Practice Guideline for the Evaluation and Management of Chronic Kidney Disease²⁵. Matched controls, with no CKD diagnosis, were recruited from receiving adult and senior citizen health examinations in the Department of Family Medicine. The Research Ethics Committee of Taipei Medical University approved this study (TMU-Joint Institutional Review Board, N201912058) which was conducted in accordance with the Declaration of Helsinki. All participants provided informed consent before the questionnaire interview and specimens collection.

Interview and specimen collection. The questionnaire interview and the urine and blood samples collection were described previously^{5,23}. We measured the arsenic species concentrations in single spot urine samples. EDTA-vacuum syringes were used to collect 5–8 mL peripheral blood samples, and the plasma, red blood cell, and buffy coat were separated. Buffy coat was separated for DNA extraction and to determine *NLRP3* polymorphisms. We analyzed the cadmium and lead concentrations from red blood cells, and measured selenium concentrations from plasma.

Environmental metals exposure measurement. Urinary concentrations of inorganic-related arsenic species, including trivalent arsenite (As^{III}), pentavalent arsenate (As^V), monomethylarsonic acid (MMA^V), and dimethylarsinic acid (DMA^V) were measured as described previously²⁶. We assessed total urinary arsenic concentrations by summing the urinary concentrations of As^{III}, As^V, MMA^V, and DMA^V. The measure of arsenic in urine specimens was a direct method of excluding nontoxic organic arsenic that contributed to total arsenic exposure²⁷. Urinary concentrations were adjusted for urinary creatinine concentrations due to variations in the hydration state²⁸. In addition, concentrations of blood cadmium and lead and plasma selenium were determined

as described previously⁶. The validity and reliability of the environmental metals exposure measurement are shown in Supplementary Table S1.

Determination of the genetic polymorphisms. DNA was extracted by digestion with proteinase K followed by phenol and chloroform. We selected 15 common *NLRP3* SNPs based on their minor-allele frequencies (≥ 0.2) in the Han Chinese in the Beijing HapMap database. The Agena Bioscience MassARRAY iPLEX system was used according to the manufacturer's instructions (National Genome Medicine Center, Taipei, Taiwan) to determine the genotypes of 15 SNPs, including rs4925654, rs4925650, rs12239046, rs4925648, rs10925025, rs10925019, rs1539019, rs3806265, rs10925026, rs10157379, rs12143966, rs10754555, rs3806268, rs12048215, and rs12137901. Among them, rs12137901 did not meet the Hardy–Weinberg equilibrium and was excluded from the statistical analysis. Linkage disequilibrium (LD) strength was determined by calculating D' and r^2 of the Lewontin using Haploview 4.1 software²⁹.

Statistical analysis. Differences in the continuous variables were compared between the two groups using the Wilcoxon rank-sum test. The Kruskal–Wallis test was used to compare continuous variables of more than two groups. Multiple logistic regression models were performed estimating odds ratios (ORs) and 95% confidence intervals (CIs) to evaluate the associations between *NLRP3* polymorphism, environmental metal exposure, and CKD while adjusting for confounders. The measures of metal concentrations were categorized into three groups based on the tertile distribution of concentrations among controls. We further treated each tertile group as an ordinal variable in the models to conduct the trend test. Additionally, multiple linear regression models were used to assess associations between *NLRP3* polymorphisms and eGFR while adjusting for confounders. Confounding was informed by prior knowledge and met the criterion that changed the ORs of exposure variables at least by 10% when adding to models assessing between environmental metal exposure and CKD⁵. We explored the interaction by estimating the combined effects of environmental metals exposure (median of concentrations among controls as a cutoff point) and *NLRP3* genotypes on CKD. The product term was added to the logistic regression models to conduct for the multiplicative interaction between metal and *NLRP3* genotypes. The SAS 9.4 (SAS Institute, Cary, NC, USA) was used for statistical analyses. A two-sided p value < 0.05 was considered statistically significant.

Results

The sociodemographic characteristics, eGFR, lifestyle, and disease histories of diabetes and hypertension between CKD cases and controls are shown in Table 1. The mean and standard deviation of age and eGFR in the 218 CKD patients and 427 controls were 65.11 ± 13.52 and 64.21 ± 12.49 years, and 31.54 ± 14.57 and 84.21 ± 15.62 mL/min/1.73 m² respectively. The CKD cases had a lower level of education. Most cases were less likely to have habits of drinking coffee, tea, and alcohol than controls. The CKD patients had significantly increased odds of regularly used analgesics compared to controls, with OR (95% CI) = 2.94 (1.58–5.44). The odds of having disease histories of diabetes and hypertension were 3–5-fold increase among cases compared to controls.

Table 2 presents the association between 14 *NLRP3* gene polymorphisms and CKD. We observed that the odds of carrying *NLRP3* rs4925650 GA/AA genotypes were 1.54-fold (95% CI 1.15–3.34) increased among CKD cases compared to controls after adjusting for covariates. In addition, participants with CKD had a 1.56–1.79 fold increased odds of carrying *NLRP3* rs12239046 CT/TT, *NLRP3* rs10925025 AG/GG, *NLRP3* rs1539019 CA/AA, *NLRP3* rs10925026 CA/AA, and *NLRP3* rs10157379 CT/TT genotypes compared to controls. After additionally adjusting environmental metals exposure, we observed that *NLRP3* rs4925650 [GA/AA vs. GG, OR (95% CI) = 1.89 (0.98–3.35)], *NLRP3* rs1539019 [CA/AA vs. CC, OR (95% CI) = 1.52 (0.95–2.44)], and *NLRP3* rs10157379 [CT/TT vs. CC, OR (95% CI) = 1.50 (0.94–2.41)] were associated with CKD though the confidence interval marginally included the null value. No association was found between other *NLRP3* genotypes and CKD after adjusting for covariates and metals concentrations. *NLRP3* rs4925650, *NLRP3* rs1539019, and *NLRP3* rs10157379 genotypes were selected for further analyzed the effect modification of gene and metal on CKD.

LD and haplotype analyses revealed that the *NLRP3* genes exhibited the four haplotype blocks shown in Supplementary Figure S1. D' of Lewontin of the haplotype *NLRP3* block 1 (*NLRP3* rs4925648, *NLRP3* rs4925650, *NLRP3* rs12048215, and *NLRP3* rs10754555), *NLRP3* block 2 (*NLRP3* rs3806265 and *NLRP3* rs3806268), *NLRP3* block 3 (*NLRP3* rs10925019 and *NLRP3* rs4925654), and *NLRP3* block 4 (*NLRP3* rs1539019, *NLRP3* rs10925025, *NLRP3* rs12143966, *NLRP3* rs12239046, *NLRP3* rs10925026, and *NLRP3* rs10157379) ranged from 0.99 to 1.00. The association between the *NLRP3* gene haplotypes and CKD is shown in Table 3. We observed that the odds of C–G–A–C and C–G–A–G haplotypes in the *NLRP3* block 1 respectively decreased by 47% and 52% among the CKD cases compared to the controls. In addition, the combined T–G–G–G, C–G–A–C, C–G–A–G, and C–G–G–G haplotypes in the *NLRP3* block 1 were significantly inversely associated with CKD compared to the C–A–A–C haplotype. These associations remained statistically significant after further adjusting for metals concentrations in the models. The odds of C–A haplotype in *NLRP3* block 3 were decreased by 35% among the CKD cases compared to the controls. No association was observed between *NLRP3* blocks 2 and 4 and CKD.

Associations between *NLRP3* rs4925650, *NLRP3* rs1539019, and *NLRP3* rs10157379 genotypes and eGFR are shown in Table 4. Participants who carried the *NLRP3* rs4925650 AA genotype decreased 7.62 mL/min/1.73 m² of eGFR when compared to those carrying the GG genotype. Participants who carried the *NLRP3* rs1539019 A allele and the *NLRP3* rs10157379 T allele decreased eGFR by 6 mL/min/1.73 m² when compared with those carrying the *NLRP3* rs1539019 C allele and the *NLRP3* rs10157379 C allele. These results showed no changes after adjusting for blood cadmium and lead or plasma selenium levels in models.

Environmental metals exposure was found to be associated with CKD in our study (Supplementary Table S2). As the levels of total urinary arsenic and blood cadmium and lead increased, the OR of CKD increased

Variables	CKD cases (N = 218)	Controls (N = 427)	Age-sex adjusted OR (95% CI)
Sex			
Male	133 (61.01)	263 (61.59)	1.00
Female	85 (38.99)	164 (38.41)	1.04 (0.74–1.46) ^a
Age	65.11 ± 13.52	64.22 ± 12.49	1.01 (0.99–1.02) ^b
eGFR (mL/min/1.73m ²)	31.54 ± 14.57	84.21 ± 15.62	0.35 (0.20–0.60) ^{***}
Educational level			
Illiterate/elementary school	90 (41.28)	97 (22.83)	1.00 [§]
Junior/senior high school	72 (32.03)	150 (35.13)	0.49 (0.33–0.74) ^{***}
College and above	56 (25.69)	180 (42.15)	0.31 (0.20–0.48) ^{***}
Cigarette smoking			
Non-smoker	160 (73.39)	311 (72.83)	1.00
Former smoker	33 (15.14)	74 (17.33)	0.85 (0.53–1.39)
Current smoker	25 (11.47)	42 (9.84)	1.21 (0.69–2.12)
Alcohol consumption			
Never	179 (82.11)	274 (64.17)	1.00
Occasional or frequently	39 (17.89)	153 (35.83)	0.36 (0.24–0.55) ^{***}
Coffee consumption			
Never	170 (77.98)	218 (51.05)	1.00
Occasional or frequently	48 (22.02)	209 (48.95)	0.29 (0.20–0.43) ^{***}
Tea consumption			
Never	123 (56.42)	149 (34.89)	1.00
Occasional or frequently	95 (43.58)	278 (65.11)	0.41 (0.29–0.58) ^{***}
Analgesic usage			
No/yes as-needed basis	192 (88.07)	408 (95.55)	1.00
Yes, routinely	26 (11.93)	19 (4.45)	2.94 (1.58–5.44) ^{***}
Diabetes			
No	132 (60.55)	383 (89.70)	1.00
Yes	86 (39.45)	44 (10.30)	5.71 (3.77–8.66) ^{***}
Hypertension			
No	94 (43.12)	298 (69.79)	1.00
Yes	124 (56.88)	129 (30.21)	3.14 (2.22–4.44) ^{***}

Table 1. Sociodemographic characteristics, lifestyle and disease histories, and eGFR between the CKD cases and controls, and the ORs of these variables for CKD. Values are expressed as mean ± standard deviation or number (%) of cases and controls. CKD chronic kidney disease, eGFR estimated glomerular filtration rate, OR odds ratio, CI confidence interval. ^{***} $p < 0.001$. [§] $p < 0.05$ for the trend test. ^aAge adjusted OR and 95% CI. ^bSex adjusted OR and 95% CI.

significantly in a dose–response manner. In contrast, as the levels of plasma selenium increased, the OR of CKD decreased significantly in a dose–response relationship. No difference was observed in comparing concentrations of environmental metals exposure by different genotypes of *NLRP3* rs4925650, *NLRP3* rs1539019, and *NLRP3* rs10157379.

Figure 1 shows the combined effect of *NLRP3* rs4925650, *NLRP3* rs1539019, *NLRP3* rs10157379, and levels of environmental metals exposure on the CKD. The trend analysis showed that the OR of CKD increased gradually with exposure to no risk factors, one risk factor, or two risk factors (risk genotypes, high levels of arsenic, cadmium, and lead, or low levels of selenium). We observed that the odds of carrying *NLRP3* rs4925650 GA/AA genotypes and high levels of blood lead (> 37.40 µg/L) were 5.03-fold increased (95% CI 2.46–10.27) among CKD cases compared to controls (Fig. 1C). The p-value of the interaction term of *NLRP3* rs4925650 and levels of blood lead was 0.0229, which indicated a multiplicative interaction between *NLRP3* rs4925650 and blood lead on CKD. In addition, we observed that *NLRP3* block 1 (risk haplotype: C-A-A-C) interacted with total urinary arsenic, blood lead and cadmium, and plasma selenium to significantly enhance the OR of CKD, respectively (Fig. 2). High levels of blood cadmium and the *NLRP3* block 1 C-A-A-C haplotype, and low levels of plasma selenium and the *NLRP3* block 1 C-A-A-C haplotype significantly and multiplicatively interacted to increase the OR of CKD, respectively.

Discussion

To the best of our knowledge, this study is the first to evaluate the associations between *NLRP3* polymorphisms, environmental metals exposure, and CKD. We found that the odds of carrying *NLRP3* rs4925650 GA/AA genotypes, *NLRP3* rs1539019 CA/AA genotypes, and *NLRP3* rs10157379 CT/TT genotypes were significantly higher among CKD cases compared to controls. In addition, certain *NLRP3* genotypes were interacting

<i>NLRP3</i> genotypes	CKD ases	Controls	Age-sex adjusted ORs (95% CI)	Multivariate adjusted ORs (95% CI) ^a
rs4925654 G>A				
GG	159 (73.27)	303 (71.13)	1.00	1.00 [§]
GA	54 (24.88)	113 (26.53)	0.90 (0.62–1.32)	0.72 (0.46–1.14)
AA	4 (1.84)	10 (2.35)	0.75 (0.23–2.43)	0.36 (0.08–1.51)
GA/AA versus GG	58 (26.73)	123 (28.87)	0.88 (0.62–1.25)	0.70 (0.45–1.08)
rs4925650 G>A				
GG	59 (27.06)	133 (31.44)	1.00 [§]	1.00 [§]
GA	101 (48.52)	210 (49.65)	1.08 (0.73–1.59)	1.37 (0.87–2.16)
AA	58 (21.53)	80 (18.91)	1.62 (1.02–2.56)*	1.96 (1.15–3.34)*
GA/AA versus GG	159 (72.94)	290 (68.56)	1.23 (0.85–1.77)	1.54 (1.01–2.36)*
rs12239046 C>T				
CC	70 (32.11)	166 (38.88)	1.00	1.00
CT	114 (52.29)	189 (44.26)	1.45 (1.01–2.09)*	1.77 (1.16–2.73)**
TT	34 (15.60)	72 (18.86)	1.12 (0.68–1.84)	1.10 (0.62–1.96)
CT/TT versus CC	148 (67.89)	261 (61.12)	1.36 (0.96–1.92)*	1.56 (1.04–2.34)*
rs4925648 C>T				
CC	124 (56.88)	234 (54.80)	1.00	1.00
CT	78 (35.78)	162 (37.94)	0.92 (0.65–1.30)	0.94 (0.62–1.41)
TT	16 (7.34)	31 (7.26)	0.98 (0.52–1.87)	0.98 (0.47–2.04)
CT/TT versus CC	94 (43.12)	193 (45.20)	0.93 (0.67–1.29)	0.95 (0.64–1.39)
rs10925025 G>A				
AA	70 (31.94)	165 (38.73)	1.00	1.00
AG	113 (52.07)	189 (44.37)	1.43 (0.99–2.06)*	1.77 (1.15–2.72)**
GG	34 (15.67)	72 (16.90)	1.12 (0.68–1.83)	1.10 (0.62–1.95)
AG/GG versus AA	147 (67.74)	261 (61.27)	1.34 (0.95–1.90)*	1.56 (1.04–2.33)*
rs10925019 C>T				
CC	102 (46.79)	199 (46.60)	1.00	1.00
CT	101 (46.33)	182 (42.62)	1.09 (0.77–1.53)	0.98 (0.65–1.46)
TT	15 (6.89)	46 (10.77)	0.64 (0.34–1.20)	0.78 (0.38–1.61)
CC/CT versus TT	203 (93.12)	381 (89.23)	0.62 (0.34–1.13)	0.79 (0.40–1.59)
rs1539019 C>A				
CC	72 (33.03)	166 (39.06)	1.00	1.00
CA	113 (51.83)	189 (44.47)	1.40 (0.97–2.01)*	1.76 (1.15–2.71)**
AA	33 (15.14)	70 (16.47)	1.09 (0.66–1.79)	1.09 (0.61–1.96)
CA/AA versus CC	146 (66.97)	259 (60.94)	1.31 (0.93–1.85)	1.56 (1.04–2.33)*
rs3806265 T>C				
TT	68 (31.19)	111 (26.06)	1.00	1.00
TC	108 (49.54)	215 (50.47)	0.79 (0.51–1.24)	0.79 (0.51–1.24)
CC	42 (19.27)	100 (23.47)	0.70 (0.41–1.19)	0.70 (0.41–1.19)
TC/CC versus TT	150 (68.81)	315 (73.94)	0.76 (0.50–1.16)	0.76 (0.50–1.16)
rs10925026 A>C				
CC	70 (32.11)	166 (39.06)	1.00	1.00
CA	114 (52.29)	187 (44.00)	0.83 (0.56–1.21)	1.79 (1.16–2.74)**
AA	34 (15.60)	72 (16.94)	0.69 (0.43–1.11)	1.10 (0.62–1.95)
CA/AA versus CC	148 (68.35)	259 (60.94)	0.79 (0.55–1.13)	1.57 (1.05–2.35)*
rs10157379 T>C				
CC	69 (31.65)	164 (38.68)	1.00	1.00
CT	114 (52.29)	189 (44.58)	1.45 (1.01–2.10)*	1.79 (1.16–2.75)**
TT	35 (16.06)	71 (16.75)	1.18 (0.72–1.93)	1.15 (0.65–2.04)
CT/TT versus CC	149 (68.35)	259 (60.94)	1.38 (0.97–1.95)*	1.59 (1.05–2.38)*
rs12143966 A>G				
GG	57 (26.15)	125 (29.83)	1.00	1.00
GA	112 (51.38)	191 (45.58)	1.29 (0.87–1.95)	1.41 (0.89–2.21)
AA	49 (22.48)	103 (24.58)	1.04 (0.66–1.66)	1.14 (0.67–1.96)
GA/AA versus GG	161 (73.85)	294 (70.17)	1.20 (0.83–1.74)	1.31 (0.86–2.01)
rs10754555 C>G				
Continued				

<i>NLRP3</i> genotypes	CKD ases	Controls	Age-sex adjusted ORs (95% CI)	Multivariate adjusted ORs (95% CI) ^a
CC	85 (38.99)	154 (36.15)	1.00	1.00
CG	102 (46.79)	199 (46.71)	0.93 (0.65–1.35)	0.87 (0.57–1.32)
GG	31 (14.22)	73 (17.14)	0.78 (0.47–1.28)	0.69 (0.39–1.23)
CG/GG versus CC	133 (61.01)	271 (63.85)	0.89 (0.64–1.15)	0.82 (0.55–1.21)
rs3806268 A>G				
AA	68 (31.19)	113 (26.59)	1.00	1.00
AG	108 (49.54)	212 (49.88)	0.85 (0.58–1.25)	0.84 (0.54–1.31)
GG	42 (19.27)	100 (23.53)	0.71 (0.44–1.13)	0.73 (0.43–1.26)
AG/GG versus AA	150 (68.81)	312 (73.41)	0.81 (0.56–1.15)	0.80 (0.53–1.22)
rs12048215 A>G				
AA	103 (47.25)	192 (44.96)	1.00	1.00
AG	93 (42.66)	185 (43.33)	0.94 (0.67–1.33)	0.93 (0.62–1.41)
GG	22 (10.09)	50 (11.71)	0.83 (0.47–1.44)	0.95 (0.50–1.77)
AG/GG versus AA	115 (52.75)	235 (55.02)	0.92 (0.66–1.28)	0.94 (0.64–1.38)

Table 2. The association between *NLRP3* gene polymorphisms and CKD. *NLRP3* rs3806265 and rs10754555 were missing for one participant. *NLRP3* rs4925654, rs10925025, rs1539019, rs10925026, and rs3806268 were missing for two participants. *NLRP3* rs10157379 was missing for three participants. *NLRP3* rs4925650 was missing for four participants. *NLRP3* rs12143966 was missing for nine participants. CKD chronic kidney disease, *NLRP3* nucleotide-binding domain-like receptors 3, OR odds ratio, CI confidence interval. ⁺0.05 ≤ *p* < 0.1, ^{*}*p* < 0.05, ^{**}*p* < 0.01. [§]*p* < 0.05 for the trend test. ^aAdjusted for age, sex, educational level, alcohol, coffee and tea consumption, analgesic usage, and disease histories of diabetes and hypertension.

with environmental metals exposure on the risk of CKD. Specifically, high levels of blood lead and *NLRP3* rs4925650 GA/AA genotypes; high levels of blood cadmium and *NLRP3* block 1 C-A-A-C haplotype, and low levels of plasma selenium and *NLRP3* block 1 C-A-A-C haplotype significantly and multiplicatively interacted to increase the risk of CKD.

Persistent inflammation and activation of the innate immune system is a chronic kidney damage phenomenon, which is important for the development of CKD³⁰. The mechanism of inflammation-induced CKD is still unknown. The pattern recognition receptors, such as nucleotide oligomerization domain-like receptors (NLRs), C-type lectin-like receptors, toll-like receptors, and retinoic acid-inducible gene I-like receptors act as sensors of the innate immune system³¹. The most characteristic inflammasome is NLRP3, which responds to endogenous damage signals caused by entry of pathogen or tissue damage³². The NLRP3 inflammasome is believed to play a key role in the underlying inflammatory response in many chronic diseases, including CKD³³.

Genetic variation in the *NLRP3* gene may be an important determinant of the degree of the immune inflammatory response, which affects susceptibility to inflammatory diseases^{34,35}. In the present study, *NLRP3* rs1539019 CA/AA genotypes significantly increased with the risk of CKD after adjusting for confounders. Various studies have evaluated the association of *NLRP3* rs1539019 polymorphism with several health-related outcomes. Patients with chronic hepatitis C virus and the *NLRP3* rs1539019 AA genotype do not respond to interferon therapy³⁶, and the *NLRP3* rs1539019 TT genotype is related to pneumoconiosis in Chinese coal workers³⁷. In addition, it has been reported that the *NLRP3* rs1539019 A allele is related to circulating fibrinogen concentration and therefore to the risk of cardiovascular disease³⁸. The rs1539019 G>T locus is close to a 12-nucleotide sequence identified as a consensus binding site for epidermal growth factor 1 that may influence the vertebrate blood coagulation network³⁹. *NLRP3* rs1539019 A allele has been indicated to increase circulating fibrinogen levels³⁸, an indicator of inflammation, which may be associated with blood coagulation, followed by leading to CKD. However, another study found no significant association between the *NLRP3*rs1539019 polymorphism and the risk of essential hypertension in a Japanese population⁴⁰. The *NLRP3* rs1539019 polymorphism was not associated with primary gouty arthritis in the Chinese Han population⁴¹. *NLRP3* rs1539019 is an intronic polymorphism and it is unclear how genetic variations in introns affect gene function. However, one study reported that many transcription factors bind to intron sites, and these intron sites may play a role regulating gene expression⁴².

In addition to *NLRP3* rs1539019 polymorphism, we also found that *NLRP3* rs4925650 GA/AA genotypes and *NLRP3* rs10157379 CT/TT genotypes significantly increased the risk of CKD after adjusting for confounders. Hence, the effect of *NLRP3* rs4925650 GA/AA genotypes and *NLRP3* rs10157379 CT/TT genotypes on susceptibility to CKD appears to be independent of age, sex, educational level, consumption of tea, alcohol, and coffee, analgesic usage, and disease histories of diabetes and hypertension. The present study found an association between the *NLRP3* rs10157379 CT/TT genotypes and CKD in the Taiwanese population, which is in accordance with our previous research showed that the *NLRP3* rs10157379 T allele has a borderline association with renal cell carcinoma⁴³. These results imply that subjects with *NLRP3* rs10157379 T allele may be associated with renal damage than those with C allele. In addition, a recent study found that the *NLRP3* rs10157379 CT genotype was associated with the severity of severe acute respiratory syndrome (SARS)⁴⁴. *NLRP3* may also be related to host immunity and susceptibility to inflammation disorders⁴⁵. Further, NLRP3 can interact with thioredoxin-interacting protein (TXNIP), a protein involved in insulin resistance. TXNIP deficiency may impair

<i>NLRP3</i> haplotypes	CKD Cases	Controls	Age-sex adjusted ORs (95% CI)	Multivariate adjusted ORs (95% CI) ^a
<i>NLRP3</i> block 1: rs4925648, rs4925650, rs12048215, and rs10754555				
C-A-A-C	215 (49.65)	370 (43.94)	1.00	1.00
T-G-G-G	108 (24.94)	224 (26.60)	0.84 (0.63–1.11)	0.79 (0.57–1.10)
C-G-A-C	56 (12.93)	133 (15.80)	0.73 (0.51–1.04) ⁺	0.63 (0.42–0.95) [*]
C-G-A-G	27 (6.24)	62 (7.36)	0.75 (0.46–1.22)	0.48 (0.27–0.85) [*]
C-G-G-G	27 (6.24)	53 (6.29)	0.79 (0.49–1.28)	0.82 (0.47–1.43)
T-G-G-G/C-G-A-C/C-G-A-G/C-G-G-G versus C-A-A-C	218 (50.35)	472 (56.06)	0.79 (0.63–0.99) [*]	0.70 (0.54–0.92) [*]
<i>NLRP3</i> block 2: rs3806265 and rs3806268				
T-A	242 (55.50)	437 (51.41)	1.00	1.00
C-G	190 (43.58)	411 (48.35)	0.84 (0.67–1.06)	0.85 (0.64–1.11)
C-A	3 (0.69)	1 (0.12)	0.61 (0.06–5.93)	0.35 (0.03–4.70)
T-G	1 (0.23)	1 (0.12)	1.67 (0.10–27.00)	3.72 (0.23–61.18)
C-G/C-A/T-G versus T-A	192 (44.04)	413 (48.59)	0.84 (0.67–1.06)	0.85 (0.65–1.11)
<i>NLRP3</i> block 3: rs10925019 and rs4925654				
C-G	242 (55.76)	445 (52.11)	1.00	1.00
T-G	130 (29.95)	275 (32.20)	0.88 (0.67–1.14)	0.85 (0.62–1.14)
C-A	62 (14.29)	134 (15.69)	0.85 (0.60–1.19)	0.65 (0.43–0.97) [*]
T-G/C-A versus C-G	192 (44.24)	409 (47.89)	0.87 (0.69–1.09)	0.78 (0.59–1.02) [*]
<i>NLRP3</i> block 4: rs1539019, rs10925025, rs12143966, rs12239046, rs10925026, and rs10157379				
C-G-A-C-A-T	222 (51.39)	440 (52.38)	1.00	1.00
A-A-G-T-C-C	177 (40.97)	322 (38.33)	1.09 (0.85–1.39)	1.15 (0.87–1.53)
C-G-G-C-A-T	27 (6.25)	72 (8.57)	0.74 (0.46–1.18)	0.85 (0.43–1.49)
C-A-G-T-C-C	4 (0.93)	3 (0.36)	2.71 (0.60–12.24)	1.47 (0.26–8.34)
C-G-A-C-A-C	1 (0.23)	2 (0.24)	0.97 (0.09–10.79)	1.25 (0.11–14.52)
A-G-A-C-A-T	0	1 (0.12)	–	–
A-G-G-C-A-T	1 (0.23)	0	–	–
A-A-G-T-C-C/C-G-G-C-A-T/C-A-G-T-C-C/C-G-A-C-A-C/A-G-A-C-A-T/A-G-G-C-A-T versus C-G-A-C-A-T	210 (48.61)	400 (47.62)	1.04 (0.82–1.31)	1.11 (0.85–1.45)

Table 3. The association between *NLRP3* gene haplotypes and CKD. CKD chronic kidney disease, *NLRP3* nucleotide-binding domain-like receptors 3, OR odds ratio, CI confidence interval. ⁺0.05 ≤ *p* < 0.1, ^{*}*p* < 0.05. ^aAdjusted for age, sex, educational level, alcohol, coffee and tea consumption, analgesic usage, and disease histories of diabetes and hypertension.

<i>NLRP3</i> genotypes	Genotypes/Alleles	β (SE) ^a	<i>p</i> values
rs4925650 G>A	GA versus GG	−1.32 (2.40)	0.582
	AA versus GG	−7.62 (2.90)	0.009
	GA/AA versus GG	−3.30 (2.26)	0.145
	A versus G	−3.06 (2.26)	0.175
rs1539019 C>A	CA versus CC	−7.47 (2.26)	0.001
	AA versus CC	−0.54 (3.05)	0.859
	CA/AA versus CC	−5.68 (2.14)	0.008
	A versus C	−5.93 (2.14)	0.006
rs10157379 T>C	CT versus CC	−7.09 (2.27)	0.002
	TT versus CC	−1.32 (3.03)	0.664
	CT/TT versus CC	−5.58 (2.14)	0.009
	T versus C	−5.84 (2.14)	0.007

Table 4. The association between *NLRP3* gene polymorphisms and eGFR. eGFR, estimated glomerular filtration rate (mL/min/1.73 m²); β, Regression coefficient; SE, Standard error of regression coefficient. ^aAdjusted for age, sex, educational level, alcohol, coffee and tea consumption, analgesic usage, disease histories of diabetes and hypertension, and total urinary arsenic.

the activation of the *NLRP3* inflammasome and subsequent secretion of interleukin 1β, which was involved in the pathogenesis of diabetes⁴⁶. As diabetes is a recognized risk factor for CKD, *NLRP3* genes may be involved in diabetes-related CKD. Additionally, the haplotype analyses were performed showing that the *NLRP3* block

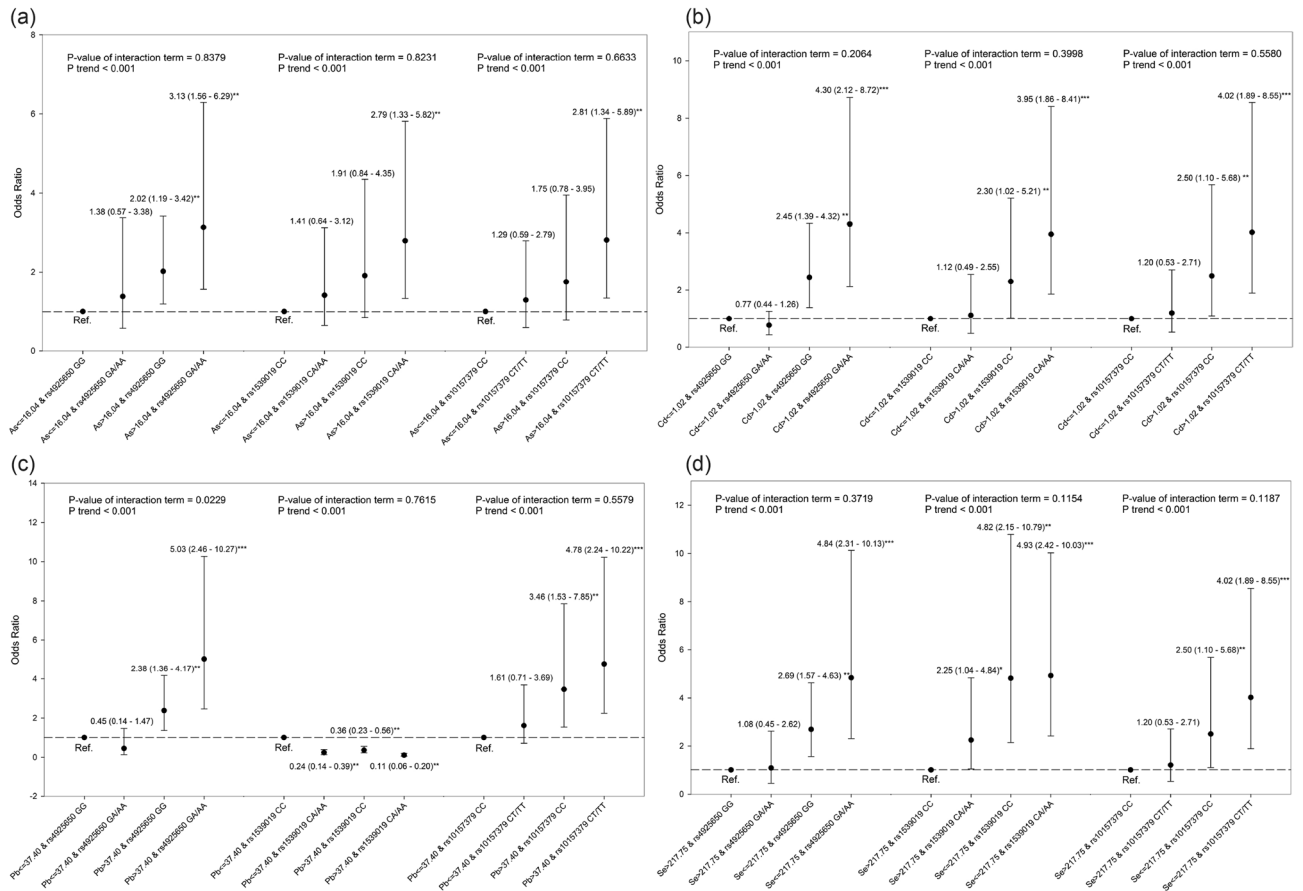


Figure 1. The combined effect of *NLRP3* rs4925650, *NLRP3*rs 1,539,019, *NLRP3* rs10157379, and levels of environmental metals exposure on the CKD. (a) Total urinary arsenic; (b) Blood cadmium; (c) Blood lead; (d) Plasma selenium. The estimates of OR were adjusted for age, sex, educational level, alcohol, coffee and tea consumption, analgesic usage, disease histories of diabetes and hypertension, and levels of other metals.

1 C-G-A-C, C-G-A-G, C-G-G-G, or T-G-G-G haplotypes significantly decreased the OR of CKD compared to that of the C-A-A-C haplotype (which includes the *NLRP3* rs4925650 A allele). To date, epidemiological studies evaluating the effect of *NLRP3* polymorphisms on CKD are limited, further studies are needed to explore the possible mechanism of the associations between these genotypes, the haplotype, and CKD.

Several in vitro and in vivo studies have indicated that metals exposure may affect *NLRP3* functional changes. An in vitro study has reported that the insulin resistance induced by NaAsO_2 is due to activation of the *NLRP3* inflammasome⁴⁷. Chicken experiments have shown that the *NLRP3* signaling pathway is activated by lead-induced oxidative stress after lead administration, which causes testicular damage¹⁶. Other animal studies have shown that cadmium chloride induces testicular injury⁴⁸ or liver injury⁴⁹ in mice by activating the *NLRP3* signaling pathway. A selenium-rich basal diet may inhibit lipopolysaccharide-induced inflammation in chicken liver by suppressing the toll like receptor 4-nuclear factor κB -*NLRP3* signaling pathway⁵⁰. Also, a study has reported that dietary selenium attenuates *Staphylococcus aureus* mastitis in mice by inhibiting the *NLRP3* inflammasome⁵¹. Our study found no difference when comparing concentrations of environmental metals exposure by different genotypes of *NLRP3* rs4925650, *NLRP3* rs1539019, and *NLRP3* rs10157379, which suggests that metals exposure and the *NLRP3* genes have independent effects on CKD. Additional studies are needed to better understand the effect of environmental metals exposure in *NLRP3* function and its mechanism.

In the present study, we observed that high levels of blood lead and *NLRP3* rs4925650 GA/AA genotypes significantly interacted to increase the risk of CKD after multivariate adjustment. This may be because lead in the blood can induce alterations of inflammatory marker *NLRP3* inflammasome activation¹⁶, and reduced eGFR⁶, leading to an increase in the risk of CKD. We also found that high levels of blood cadmium and the *NLRP3* block 1 haplotype C-A-A-C multiplicatively interacted to increase the risk of CKD after adjusting for multiple risk factors. Evidence has shown that cadmium in the blood may inhibit heme oxygenase 1 and nuclear factor erythroid 2-related factor 2, and activate the *NLRP3* inflammasome⁴⁹ or increase reactive oxygen species to activate the *NLRP3* inflammasome⁴⁸, and decreased eGFR⁶, which may jointly cause CKD pathogenesis⁵². In addition, we observed that low plasma selenium level and the *NLRP3* block 1 haplotype C-A-A-C multiplicatively interacted to increase the risk of CKD after adjusting for multiple risk factors. Studies have found that a low level of plasma selenium may not inhibit the expression of *NLRP3*⁵¹, or downregulate the toll-like receptor 4-nuclear factor- κB -*NLRP3* signaling pathway⁵⁰, which may increase kidney inflammation to increase the risk of CKD⁵³. Our study did not precisely measure the levels of serum *NLRP3* inflammasome. Therefore, whether the *NLRP3*

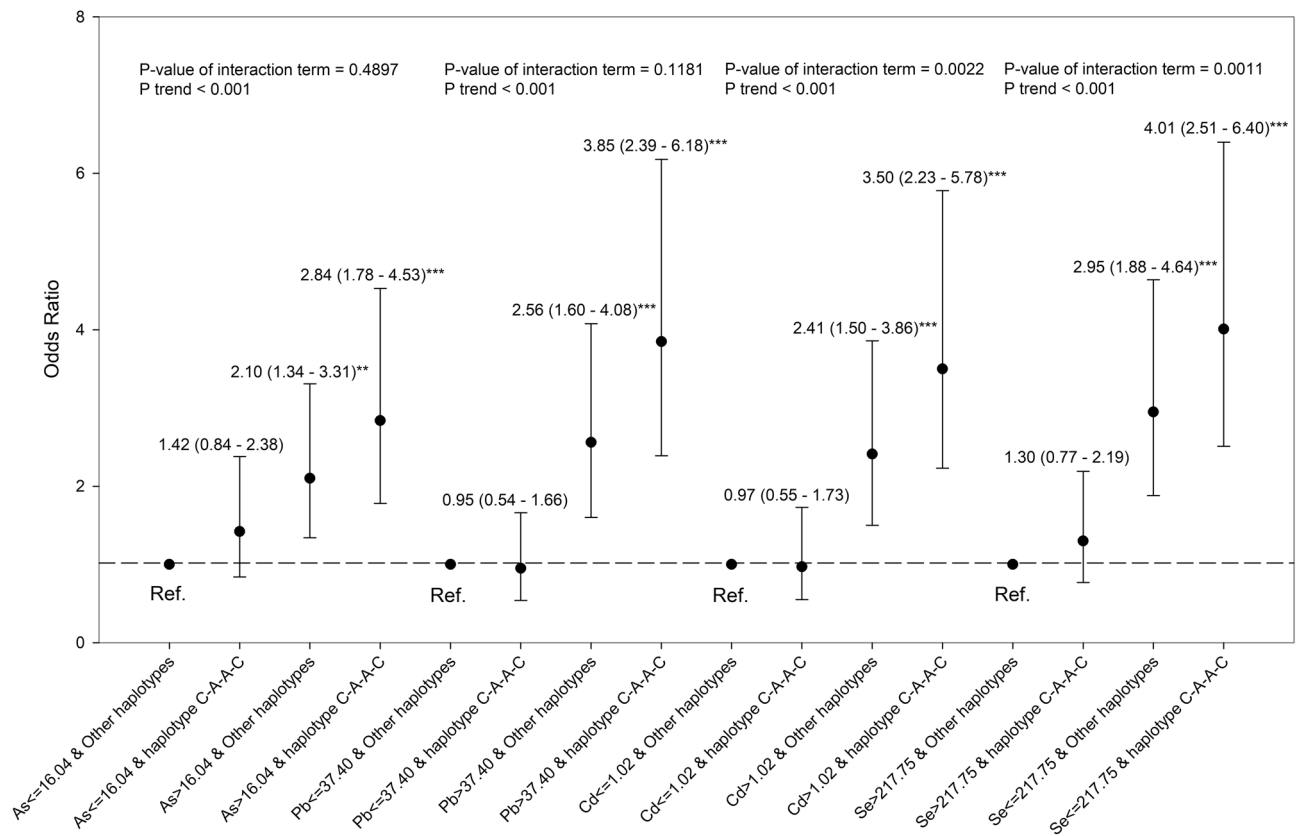


Figure 2. The combined effect of the *NLRP3* block 1 haplotypes (rs4925648, rs4925650, rs12048215, and rs10754555) and levels of total urinary arsenic, blood lead and cadmium, and plasma selenium on CKD. The estimates of OR were adjusted for age, sex, educational level, alcohol, coffee and tea consumption, analgesic usage, disease histories of diabetes and hypertension, and levels of other metals.

inflammasome was affected by levels of blood cadmium and lead, low levels of plasma selenium or the *NLRP3* polymorphisms remain unknown. The underlying mechanism of joint effect of environmental metals exposure and *NLRP3* polymorphisms affecting CKD needs further exploration.

The sample size of this study was small with limited statistical power. Further studies with larger sample size are needed to improve the precision of point estimates when assessing the effect modification of *NLRP3* gene polymorphisms and environmental metals exposure on CKD. Additionally, unmeasured factors such as blood pressure medication and treatment might have potentially influenced our observed results. Future studies should consider these factors in multiple regression models. The analysis of 15 *NLRP3* gene polymorphisms may not represent those of the entire gene functions. Our study did not analyze gene polymorphisms that regulate *NLRP3* expression differently according to the disease conditions. Further studies should be carried out to evaluate the function of *NLRP3* and its related gene polymorphisms in order to determine their role in CKD development.

Conclusions

In conclusion, the present study found evidence that the *NLRP3* rs4925650 GA/AA genotypes or *NLRP3* block 1 haplotype C-A-A-C altered the risk of CKD related to high levels of blood lead and cadmium, or low levels of plasma selenium. Future studies are warranted to measure the levels of serum *NLRP3* inflammasome, to elucidate the biological mechanism underlying the associations between *NLRP3* polymorphisms, environmental metals exposure, and CKD.

Received: 20 September 2021; Accepted: 1 April 2022

Published online: 15 April 2022

References

- Jha, V. *et al.* Chronic kidney disease: Global dimension and perspectives. *Lancet* **382**, 260–272 (2013).
- Couser, W. G., Remuzzi, G., Mendis, S. & Tonelli, M. The contribution of chronic kidney disease to the global burden of major noncommunicable diseases. *Kidney Int.* **80**, 1258–1270 (2011).
- Wen, C. P. *et al.* All-cause mortality attributable to chronic kidney disease: A prospective cohort study based on 462 293 adults in Taiwan. *Lancet* **371**, 2173–2182 (2008).
- Lin, Y. C. *et al.* Incidence and prevalence of ESRD in Taiwan Renal Registry Data System (TWRDS): 2005–2012. *Acta Nephrol.* **28**, 65–66 (2014).

5. Hsueh, Y. M. *et al.* Urinary arsenic species and CKD in a Taiwanese population: A case–control study. *Am J Kidney Dis* **54**, 859–870 (2009).
6. Wu, C. Y. *et al.* The association between plasma selenium and chronic kidney disease related to lead, cadmium and arsenic exposure in a Taiwanese population. *J. Hazard. Mater.* **375**, 224–232 (2019).
7. Liu, Y. *et al.* Associations of plasma metal concentrations with the decline in kidney function: A longitudinal study of Chinese adults. *Ecotoxicol. Environ. Saf.* **189**, 110006 (2020).
8. Lee, J. *et al.* Environment-wide association study of CKD. *Clin. J. Am. Soc. Nephrol.* **15**, 766–775 (2020).
9. Anders, H. J. & Muvuru, D. A. The inflammasomes in kidney disease. *J. Am. Soc. Nephrol.* **22**, 1007–1018 (2011).
10. Zhang, C. *et al.* Activation of Nod-like receptor protein 3 inflammasomes turns on podocyte injury and glomerular sclerosis in hyperhomocysteinemia. *Hypertension* **60**, 154–162 (2012).
11. Vilaysane, A. *et al.* The NLRP3 inflammasome promotes renal inflammation and contributes to CKD. *J. Am. Soc. Nephrol.* **21**, 1732–1744 (2010).
12. Hutton, H. L., Ooi, J. D., Holdsworth, S. R. & Kitching, A. R. The NLRP3 inflammasome in kidney disease and autoimmunity. *Nephrology* **21**, 736–744 (2016).
13. Qiu, T. *et al.* Taurine attenuates arsenic-induced pyroptosis and nonalcoholic steatohepatitis by inhibiting the autophagic-inflammasomal pathway. *Cell Death. Dis.* **9**, 946 (2018).
14. Ahn, H. *et al.* Mercury and arsenic attenuate canonical and non-canonical NLRP3 inflammasome activation. *Sci. Rep.* **8**, 13659 (2018).
15. Zhang, Y., Liu, Q., Yin, H. & Li, S. Cadmium exposure induces pyroptosis of lymphocytes in carp pronephros and spleens by activating NLRP3. *Ecotoxicol. Environ. Saf.* **202**, 110903 (2020).
16. Huang, H. *et al.* Anti-inflammatory effect of selenium on lead-induced testicular inflammation by inhibiting NLRP3 inflammasome activation in chickens. *Theriogenology* **155**, 139–149 (2020).
17. Zhang, A. Q. *et al.* Clinical relevance of single nucleotide polymorphisms within the entire NLRP3 gene in patients with major blunt trauma. *Crit. Care* **15**, R280 (2011).
18. Zaki, M. H., Lamkanfi, M. & Kanneganti, T. D. The Nlrp3 inflammasome: Contributions to intestinal homeostasis. *Trends Immunol.* **32**, 171–179 (2011).
19. The International HapMap Project. The international HapMap project. *Nature* **426**, 789–796 (2003).
20. Kunnas, T., Maatta, K. & Nikkari, S. T. NLR family pyrin domain containing 3 (NLRP3) inflammasome gene polymorphism rs7512998 (C>T) predicts aging-related increase of blood pressure, the TAMRISK study. *Immun. Ageing* **12**, 19 (2015).
21. Zhou, D. *et al.* The NLRP3 rs10754558 polymorphism is associated with the occurrence and prognosis of coronary artery disease in the Chinese Han population. *Biomed. Res. Int.* **2016**, 3185397 (2016).
22. Cheng, L., Yin, R., Yang, S., Pan, X. & Ma, A. Rs4612666 polymorphism of the NLRP3 gene is associated with the occurrence of large artery atherosclerotic ischemic strokes and microembolic signals. *Biomed. Res. Int.* **2018**, 6345805 (2018).
23. Chen, W. J. *et al.* Renin–angiotensin–aldosterone system related gene polymorphisms and urinary total arsenic is related to chronic kidney disease. *Toxicol. Appl. Pharmacol.* **279**, 95–102 (2014).
24. National Kidney Foundation. K/DOQI clinical practice guidelines for chronic kidney disease: Evaluation, classification, and stratification. *Am. J. Kidney Dis.* **39**, S1–266 (2002).
25. KDIGO. Clinical practice guideline for the evaluation and management of chronic kidney disease. *Kidney Int. Suppl.* **2013**(3), 1–150 (2012).
26. Hsueh, Y. M. *et al.* Serum beta-carotene level, arsenic methylation capability, and incidence of skin cancer. *Cancer Epidemiol. Biomark. Prev.* **6**, 589–596 (1997).
27. National Research Council Arsenic in drinking water (1999).
28. Barr, D. B. *et al.* Urinary creatinine concentrations in the U.S. population: Implications for urinary biologic monitoring measurements. *Environ. Health Perspect.* **113**, 192–200 (2005).
29. Barrett, J. C., Fry, B., Maller, J. & Daly, M. J. Haploview: Analysis and visualization of LD and haplotype maps. *Bioinformatics* **21**, 263–265 (2005).
30. Granata, S., Dalla, G. A., Bellin, G., Lupo, A. & Zaza, G. Transcriptomics: A step behind the comprehension of the polygenic influence on oxidative stress, immune deregulation, and mitochondrial dysfunction in chronic kidney disease. *Biomed. Res. Int.* **2016**, 9290857 (2016).
31. Wang, X. & Yi, F. Implication of pattern-recognition receptors in cardiovascular diseases. *Antioxid. Redox. Signal.* **22**, 1130–1145 (2015).
32. Guo, H., Callaway, J. B. & Ting, J. P. Inflammasomes: Mechanism of action, role in disease, and therapeutics. *Nat. Med.* **21**, 677–687 (2015).
33. Masood, H., Che, R. & Zhang, A. Inflammasomes in the pathophysiology of kidney diseases. *Kidney Dis.* **1**, 187–193 (2015).
34. Paramel, G. V., Sirsjo, A. & Fransen, K. Role of genetic alterations in the NLRP3 and CARD8 genes in health and disease. *Mediat. Inflamm.* **2015**, 846782 (2015).
35. Verma, D. *et al.* Gene polymorphisms in the NALP3 inflammasome are associated with interleukin-1 production and severe inflammation: Relation to common inflammatory diseases?. *Arthritis Rheum.* **58**, 888–894 (2008).
36. Estfanous, S. Z. K., Ali, S. A., Seif, S. M., Soror, S. H. A. & Abdelaziz, D. H. A. Inflammasome genes’ polymorphisms in Egyptian chronic hepatitis C patients: Influence on vulnerability to infection and response to treatment. *Mediat. Inflamm.* **2019**, 3273645 (2019).
37. Ji, X. *et al.* Polymorphisms in inflammasome genes and risk of coal workers’ pneumoconiosis in a Chinese population. *PLoS ONE* **7**, e47949 (2012).
38. Dehghan, A. *et al.* Association of novel genetic Loci with circulating fibrinogen levels: A genome-wide association study in 6 population-based cohorts. *Circ. Cardiovasc. Genet.* **2**, 125–133 (2009).
39. Davidson, C. J., Tuddenham, E. G. & McVey, J. H. 450 million years of hemostasis. *J. Thromb. Haemost.* **1**, 1487–1494 (2003).
40. Omi, T. *et al.* An intronic variable number of tandem repeat polymorphisms of the cold-induced autoinflammatory syndrome 1 (CIAS1) gene modifies gene expression and is associated with essential hypertension. *Eur. J. Hum. Genet.* **14**, 1295–1305 (2006).
41. Zhang, Q. B., Qing, Y. F., He, Y. L., Xie, W. G. & Zhou, J. G. Association of NLRP3 polymorphisms with susceptibility to primary gouty arthritis in a Chinese Han population. *Clin. Rheumatol.* **37**, 235–244 (2018).
42. Rose, A. B. Introns as gene regulators: A brick on the accelerator. *Front. Genet.* **9**, 672 (2018).
43. Chung, C. J. *et al.* Polymorphism of nucleotide binding domain-like receptor protein 3 (NLRP3) increases susceptibility of total urinary arsenic to renal cell carcinoma. *Sci. Rep.* **10**, 6640 (2020).
44. Maes, M. *et al.* In COVID-19, NLRP3 inflammasome genetic variants are associated with critical disease and these effects are partly mediated by the sickness symptom complex: A nomothetic network approach. *Mol. Psychiatry* <https://doi.org/10.1038/s41380-021-01431-4> (2022).
45. Fritz, J. H., Ferrero, R. L., Philpott, D. J. & Girardin, S. E. Nod-like proteins in immunity, inflammation and disease. *Nat. Immunol.* **7**, 1250–1257 (2006).
46. Zhou, R., Tardivel, A., Thorens, B., Choi, I. & Tschopp, J. Thioredoxin-interacting protein links oxidative stress to inflammasome activation. *Nat. Immunol.* **11**, 136–140 (2010).

47. Jia, X. *et al.* Arsenic induces hepatic insulin resistance via mtROS-NLRP3 inflammasome pathway. *J. Hazard. Mater.* **399**, 123034 (2020).
48. Han, C. *et al.* Protective effect of *Polygonatum sibiricum* against cadmium-induced testicular injury in mice through inhibiting oxidative stress and mitochondria-mediated apoptosis. *J. Ethnopharmacol.* **261**, 113060 (2020).
49. Liu, C. *et al.* Cadmium induces acute liver injury by inhibiting Nrf2 and the role of NF-kappaB, NLRP3, and MAPKs signaling pathway. *Int. J. Environ. Res. Public Health* **17**, 138 (2019).
50. Qu, J., Wang, W., Zhang, Q. & Li, S. Inhibition of lipopolysaccharide-induced inflammation of chicken liver tissue by selenomethionine via TLR4-NF-kappaB-NLRP3 signaling pathway. *Biol. Trace Elem. Res.* **195**, 205–214 (2020).
51. Bi, C. L. *et al.* Selenium plays an anti-inflammatory role by regulation NLRP3 inflammasome in *Staphylococcus aureus*-infected mouse mammary gland. *Biol. Trace Elem. Res.* **199**, 604–610 (2020).
52. Mulay, S. R. Multifactorial functions of the inflammasome component NLRP3 in pathogenesis of chronic kidney diseases. *Kidney Int.* **96**, 58–66 (2019).
53. Li, L., Tang, W. & Yi, F. Role of inflammasome in chronic kidney disease. *Adv. Exp. Med. Biol.* **1165**, 407–421 (2019).

Acknowledgements

This study was supported by grants from the Taipei Medical University-Wanfang Hospital Research Project (110TMU-WFH-01) and the Ministry of Science and Technology of Taiwan (MOST 103-2314-B-038-021-MY2 (1-2), MOST 103-2314-B-038-021-MY2 (2-2), MOST 105-2314-B-038-082, MOST 106-2314-B-038-066, MOST 107-2320-B-039-010, MOST 106-2314-B-002-235-MY3, MOST 107-2314-B-038-073, MOST 108-2314-B-038-089, MOST 109-2314-B-038-081- and MOST 109-2314-B-038-067-).

Author contributions

Y.L.H., H.S.S., and R.L.H. partly contributed to the conception and design of the work, and H.H.C., Y.F.L., and Y.C.L. recruited the study subjects; W.J.C. has done the experiment; Y.M.H. contributed to the statistical analysis and analyzed the data. Y.M.H. wrote the manuscript; W.J.C. reviewed and editing the manuscript; H.H.C. performed the study design and executed the whole research plan.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1038/s41598-022-10098-y>.

Correspondence and requests for materials should be addressed to H.-H.C.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022