

The Love-Hate Relationship Between TGF-β Signaling and the Immune System During Development and Tumorigenesis

Baode Chen^{1†}, Chenglin Mu^{2†}, Zhiwei Zhang², Xuelin He^{3*} and Xia Liu^{2*}

¹ Department of Laboratory Medicine, Key Laboratory of Clinical In Vitro Diagnostic Techniques of Zhejiang Province, The First Affiliated Hospital, College of Medicine, Zhejiang University, Hangzhou, China, ² Institute for Intelligent Bio/Chem Manufacturing (iBCM), Zhejiang University (ZJU)-Hangzhou Global Scientific and Technological Innovation Center, Hangzhou, China, ³ Kidney Disease Center, The First Affiliated Hospital, College of Medicine, Zhejiang University, Hangzhou, China

OPEN ACCESS

Edited by:

Juan Francisco Santibanez, University of Belgrade, Serbia

Reviewed by:

Theresa L. Whiteside, University of Pittsburgh, United States David A. Horwitz, University of Southern California, United States

*Correspondence:

Xuelin He hexuelin2018@zju.edu.cn Xia Liu I210136@zju.edu.cn [†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Immunology

Received: 07 March 2022 Accepted: 25 April 2022 Published: 26 May 2022

Citation:

Chen B, Mu C, Zhang Z, He X and Liu X (2022) The Love-Hate Relationship Between TGF-β Signaling and the Immune System During Development and Tumorigenesis. Front. Immunol. 13:891268. doi: 10.3389/fimmu.2022.891268 Since TGF- β was recognized as an essential secreted cytokine in embryogenesis and adult tissue homeostasis a decade ago, our knowledge of the role of TGF- β in mammalian development and disease, particularly cancer, has constantly been updated. Mounting evidence has confirmed that TGF- β is the principal regulator of the immune system, as deprivation of TGF- β signaling completely abrogates adaptive immunity. However, enhancing TGF- β signaling constrains the immune response through multiple mechanisms, including boosting Treg cell differentiation and inducing CD8⁺ T-cell apoptosis in the disease context. The love-hate relationship between TGF- β signaling TGF- β , especially for cancer treatment. Nonetheless, recent work on combination therapies of TGF- β inhibition and immunotherapy have provide insights into the development of TGF- β -targeted therapies, with favorable outcomes in patients with advanced cancer. Hence, we summarize the entanglement between TGF- β and the immune system in the developmental and tumor contexts and recent progress on hijacking crucial TGF- β signaling pathways as an emerging area of cancer therapy.

Keywords: TGF- β , immune system, tumor progression, tumor microenvironment, cancer therapy

INTRODUCTION

The immune system's integrity and function are critical determinants of normal development and disease outcomes, particularly cancer. All immune system cells, including lymphocytic T, B, NK, and myeloid cells, originate from the hematopoietic stem cells and undergo multiple developmental stages, and TGF- β is imperative in all developmental stages. In the cancer context, the immune system and tumors evolve with unique features, making it more complicated to elucidate the multifunctional features of TGF- β during tumorigenesis. The undesirable clinical results of inhibiting the TGF- β pathway pose significant challenges for targeted therapy in cancer therapeutics. Therefore, we mainly summarize the crosstalk between TGF- β and the immune system in the context of homeostasis and disease, particularly cancer.

In both the embryonic and postnatal development stages, nearly all cells respond to TGF- β family signals and make fate decisions under the influence of TGF- β signals (1, 2). As the main components of adaptive immunity, T cells and B cells respond to TGF- β signals in different developmental stages, thus resulting in the progression or blockage of development. When mature T and B cells migrate into the periphery, they maintain peripheral immune homeostasis until they are activated by foreign antigens and differentiate into cell subsets, mostly with TGF- β involvement.

The tumor and tumor microenvironment (TME) evolve coordinately with reciprocal signaling from the tumor bulk, which consists of tumor cells, tumor epithelial cells (TECs), carcinomaassociated fibroblasts (CAFs), and immune cells. Different cells within the tumor context respond to TGF-B stimulation in a context-dependent manner and have considerable signaling heterogeneity, which is an impediment to therapeutic approaches for patients with cancer. A comprehensive understanding of the function of TGF- β in oncogenesis requires sufficient knowledge of the complicated responses of different cell types in the tumor to TGF- β . Here, we summarize current knowledge on TGF- β signaling functions in individual cellular components enclosed by the TME. Clarifying the immunosuppressive role of the TGF-B signaling pathway within tumors (3-5) and converting the tumorsuppressive microenvironment by remodeling TGF-B-initiated transmembrane signaling have spurred therapeutic progress in TGF-β-related drugs, including molecular blockers, CAR-T cells, and bispecific antibodies. We also highlight future challenges and directions in combining established regimens with anti-TGF-B to further enhance therapeutic efficacy.

BIOLOGICAL ACTIVITY AND SIGNALING MODELS OF THE TGF-B-RELATED PATHWAY

The biology of TGF- β signaling has been extensively investigated in several invaluable reviews (3, 6). Figure 1 presents a flow diagram of TGF- β secretion and the downstream signal pathway activation process. Overall, the mammalian genome encodes three functionally overlapping TGF- β isoforms, and each isoform binds noncovalently to the latency-associated peptide (LAP) at the N-terminal portion (7, 8). LAPs associate with the large ECM protein LTBP (latent TGF-B1 binding protein) by disulfide bonds to compose a tripartite complex termed the large latent complex (LLC). The primary biological function of LAP in LLC is to confer latency to TGF- β by preventing the binding of TGF- β to TGFR1 and TGFR2, whereas LTBP mainly functions to tether bona fide latent ligand to the ECM and assist with the proper folding and secretion of TGF- β (9). The release of TGF- β from the ECM mainly occurs through extracellular proteolytic cleavage from the LAP domain dependent on proteases (7), as well as matrix metalloproteinases in the TME (10).

Released TGF- β triggers cellular signaling by engaging with the tetrameric receptor complex, which is composed of TGF- β RI and TGF- β RII. Then, TGF- β RII binds to TGF- β and recruits and

phosphorylates TGF- β RI in the form of a heterotetrameric complex. Activation of the TGF- β receptor complex triggers Smad-dependent or non-Smad-mediated cascade events. In the canonical Smad-mediated model, TGF- β binds to transmembrane receptors and then recruits and phosphorylates intracellular Smad2 and Smad3 proteins, forming heterotrimeric complexes with SMAD4, which then translocates into the nucleus to activate or repress target gene transcription (11).

The SMAD-independent noncanonical TGF- β signaling pathway includes several context-dependent downstream pathways, including the ERK/MAPK, PI3K/AKT, and MKK/ p38 pathways (12, 13). For example, during the epithelialmesenchymal transition (EMT), TGF- β -modulated fibroblastic lineage reprogramming and cell emigration are dependent on p38 mitogen-activated protein kinase (14). The kinetics and functions of the ERK/MAPK pathway when encountering TGF- β stimulation in epithelial cells, fibroblasts, and cancer cells are tissue specific (15).

THE TGF-B SIGNALING PATHWAY IS INVOLVED IN THE DEVELOPMENT OF THE HEMATOPOIETIC AND IMMUNE SYSTEMS

Hematopoietic stem cells (HSCs) sustain the lifelong provision of immune cells. The immune system is a complex network of biological processes that protect individuals from infection and disease. TGF- β plays essential roles in different stages of immune system development and maintains immune tolerance and cellular homeostasis by exerting specific functions on various immune cell components, as summarized in **Figure 2**.

Hematopoietic Stem Cells

The earliest evidence supporting the role of TGF- β in mouse development was obtained by mutation of the TGF-B1 gene in embryonic stem cells, which led to mice dying by 3-4 weeks of age (16, 17). HSCs in the bone marrow were shown to preserve quiescence and infrequent division to maintain the continuous replenishment of the mature peripheral immune cell pool. Numerous subsequent studies have confirmed the role of TGF- β in the hematopoietic system in both humans and mice (18–20). In bone marrow, the ablation of megakaryocytes (MKs) significantly increases HSC proliferation through TGF-B-SMAD signaling (21). Furthermore, several groups provided evidence for the physiological influence of TGF- β in maintaining HSC quiescence, including increased HSC cycling and reduced regenerative capacity upon transplantation in TGF TGF-B RII-deficient HSCs (22), impaired HSC homing upon transplantation in TGF-B1-deficient neonates (23), and improved hematopoietic regeneration after TGF-B blockade with a neutralized TGF- β antibody (1D11) accompanied by chemotherapy (24). Interestingly, the self-renewal and differentiation capacity of HSCs do not differ between steadystate and stress conditions in mice deficient in TGF-BRI (25, 26). The higher expression level of TGF-BRII within HSCs and other

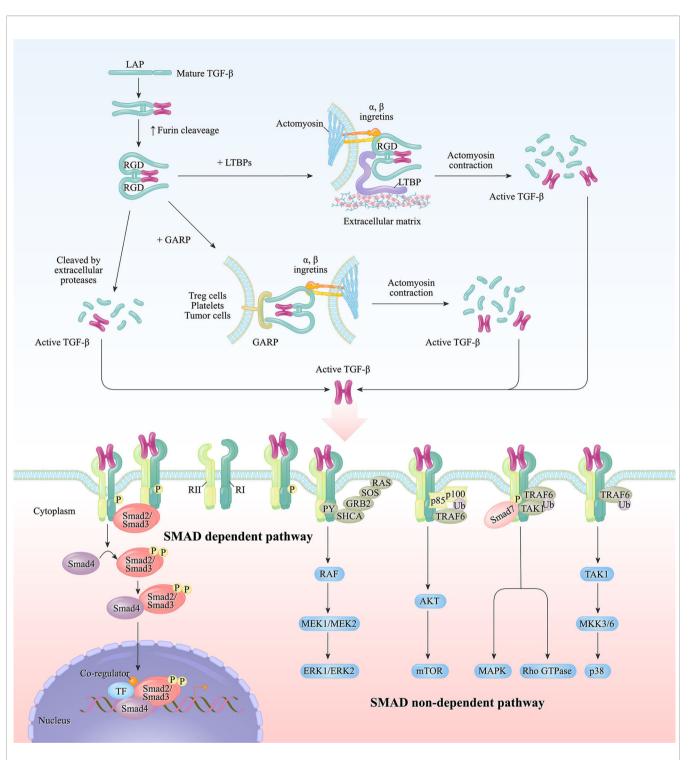
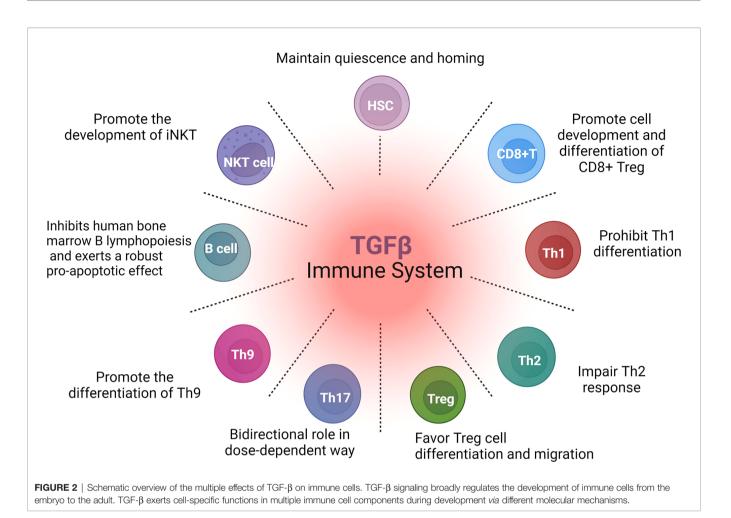


FIGURE 1 | Canonical and non-canonical TGF- β signaling. Engagement between TGF- β and its receptor initiates canonical and non-canonical signaling pathways. The mature fragment of TGF- β 1 remains associated with latency-associated peptide (LAP) at N-terminal propeptide. LTBPs (latent transforming growth factor β binding proteins) from the extracellular matrix (ECM) form the large latent complex (LLC) with TGF- β in the endoplasmic reticulum. When TGF- β was released and recognized by receptors, it subsequently stimulates the canonical and non-canonical pathways in cells through separate mechanisms. In canonical signaling, which is also called the SMAD-dependent pathway, the receptor activation triggers a cascade of SMAD proteins phosphorylation and translocation into the nucleus, thus promoting the downstream gene expression. While in non-canonical signaling, the receptor ligation leads to SMAD non-dependent pathway activation, including MEK/ERK pathway, AKT pathway, MAPK, and p38 pathways.



TGF- β RI-independent pathways may explain the differences detected between the two mouse models. Overall, TGF- β interferes with growth mainly through the upregulation of p57^{Kip2} in the most primitive HSC compartment (27–29). In addition, TGF- β 1-deficient mice display enhanced myelopoiesis, suggesting a negative regulatory role of TGF- β in myelopoiesis (30). The above observations indicate that TGF- β is a pivotal regulator of the host hematopoietic system.

CD8⁺ T Cells

T and B cells derived from HSCs are essential components of the adaptive immune system. As the thymus is the primary site for T-cell development, early studies primarily focused on TGF- β regulation of intrathymic T-cell development. Notably, most intrathymic $\alpha\beta$ T lymphocytes undergo a negative selection process, during which a majority of $\alpha\beta$ T lymphocytes with high affinity for autonomous antigens are eradiated to avoid self-attack. Strikingly, TGF- β signaling-deficient DP thymocytes bypass negative selection, leading to substantial accumulation in the periphery and induction of autoimmune lesions in several organs (31). In addition, when intrathymic CD8⁺ T cells, TGF- β regulates the cell surface level of IL-7R α on CD8⁺ thymocytes

and promotes CD8⁺ T cell lineage commitment by suppressing Gfi-1, a known IL-7Rα transcriptional repressor (32). A subset of CD8⁺Foxp3⁺ Treg cells that possess repressive function in the thymus are induced by TGF- β stimulation from CD8⁺Foxp3⁻ T cells (33, 34). Likewise, sustained commitment of thymusderived CD4⁺FoxP3⁺ regulatory T cells (Treg cells) requires TGF- β signaling (35, 36). In summary, TGF- β instructs the intrathymic development of conventional CD8⁺ T cells and intrathymic differentiation of CD8⁺ Treg and CD4⁺ Treg populations. To interpret the specific role of TGF- β in peripheral mature T cells, Richard A Flavell's laboratory generated a transgenic mouse model in which TGF-B signaling was exclusively abolished in T cells. The mice showed high infiltration of inflammatory cells in diverse organs, automatic activation of T cells, and autoimmune antibody secretion, confirming the role of TGF- β in modulating immunological balance (37). Subsequently, other laboratories constructed mice with TGF- βRII or TGF- βRI deficiency governed by the CD4 promoter, which showed severe developmental defects in T-cell lineages and autoimmune-associated lesions (35, 38). In addition, when major histocompatibility complex (MHC) class II molecules were mutated in TGF- β 1-null mice, the mutation rescued the inflammatory phenotype (30), suggesting that TGF-

 β 1 deficiency results in an uncontrolled T-cell response. Collectively, the above studies convincingly identify a dominant role for TGF- β acting directly during the T-cell development process by maintaining immunological homeostasis, affirming the significance of TGF- β in regulating T-cell activity.

CD4⁺ T Cells

TGF- β signaling maintains cellular homeostasis in a similar way in CD4⁺ T cells by promoting the expression of IL-7R α . However, due to the multifaceted nature of differentiation activity, TGF- β also exerts different functions on individual CD4⁺ T subsets, including Th1, Treg, Th17, and Th9 cells.

T Helper 1 (Th1) Cells

Th1-cell differentiation is driven by the master transcription factor T-bet, which is potently inhibited by TGF- β (39). Ectopic expression of TGF- β in developing Th1 cells abolish the inhibitory effect of TGF- β (40), confirming the imperative role of TGF- β in limiting Th1-cell differentiation. Low Th1-cell activity and reduced immune cytotoxicity can forecast negative outcomes in colorectal cancer patients (41). In particular, microsatellite-instable (MSI) colon cancer patients exhibit an elevated ratio of Th1-to-naive T cells, which is inversely correlated with *tgfb* gene expression (42).

T Helper 2 (Th2) Cells

The role of TGF- β in Th2 cells is controversial. In an early study, adding TGF- β to naive T cells led to the inhibition of Th2 cell differentiation by inducing Sox4, a transcription factor that negatively regulated the Th2 master regulator GATA-3. In addition, TGF- β was able to directly prevent Th2-associated cytokine secretion, including IL-5 secretion (43, 44). Intriguingly, when murine T cells lack both T-bet and TGF- β RII expression, they also spontaneously exhibit inflammation, which is associated with reinforced Th2 cell differentiation (45). In addition, in a TGF- β -rich context, such as the mucosa of T. muris-infected mice, the protein mina may act as an important counterbalance to the induction of Relm β by the Th2 cytokines IL-4 and IL-13 (46). In cancer, a study that TGF- β signaling hinders Th2 cell responses that reconstruct the tumor vasculature and restrain tumor advancement (47).

Treg Cells

Several studies have elaborated the major role of TGF- β signaling in Treg cells (36, 48). In essence, TGF- β signaling supports thymus-derived Treg cell survival by suppressing T-cell clonal deletion and promoting induced Treg (iTreg) cell differentiation in the periphery by inducing Foxp3 expression. Additionally, TGF- β directs the movement or retention of Treg cells in inflammatory tissue through several different molecular mechanisms, such as GPR15-mediated homing into the large intestine mucosa (49). Mechanistically, the engagement of TGF- β signaling promotes the binding of Smad3 to the enhancer region of Foxp3, which is called CNS1 (50, 51), to modulate Foxp3 expression. In addition, Smad2 cooperatively interacts with Smad3 in iTreg cell production (52).

T Helper 17 (Th17) Cells

The signaling mechanisms underlying the role of TGF- β in the differentiation of the Th17-cell subset are controversial. Dan R Littman's group proposed a dose-dependent theory in which a low concentration of TGF- β promotes Foxp3 expression, while a high concentration of TGF- β upregulates IL-23 receptor levels on CD4⁺ T cells and promotes RORyt+ Th17-cell commitment (53). As a TGF- β family member, BMP receptor 1 α was demonstrated to suppress Th17-cell differentiation from CD4⁺ T cells because the loss of this receptor promotes the differentiation of Th17 cells and exacerbates colitis in a mouse model. By contrast, another study found that the ALK5 (TGF-B type I receptor kinase) inhibitor SB-505124 potently inhibited human Th17 differentiation in vitro by decreasing the gene expression of *il-17* and *roryt* genes, along with the protein level of IL-17 (54). These pieces of evidence suggest that TGF-β family members may perform individual tasks during Th17cell differentiation.

T Helper 9 (Th9) Cells

T helper 9 (Th9) cells are CD4⁺ effector T cells that exert robust antitumor activities that are as strong as those of Th1 cells (55). Several studies have confirmed that TGF- β , in conjunction with IL-4, controls the differentiation of Th9 cells (56, 57). The underlying molecular mechanism includes downregulation of the expression of the DNA-binding inhibitor Id3 by TGF-β1 in conjunction with IL-4, which accelerates the elevated binding of E2A and GATA-3 to the promoter region of IL9, thus resulting in increased Il9 gene transcription and Th9 cell differentiation (58). Th9 cell differentiation depends upon TGF- β along with IL-4, which is also explained by the molecular induction of the transcription factor PU.1 (59, 60). In addition, a recent study by Yichuan Xiao's group reported that TGF-B collaborates with BFAR (a bifunctional apoptosis regulator) to regulate the antitumor function of Th9 cells, as BFAR-overexpressing Th9 cells display favorable antitumor efficacy. By contrast, BFAR KO significantly inhibits TGF-BR1 ubiquitination and Th9 differentiation, hence inhibiting the antitumor function of Th9 cells (61). Taken together, the above studies emphasize the importance of TGF- β in Th9 cell differentiation and function.

B Cells

The involvement of TGF- β in B-cell development has been demonstrated extensively. In the early developmental stage, BMP-6 participates in fine-tuning of human bone marrow B lymphopoiesis by upregulating two important Smad targets, Id1 and Id3 (62). Moreover, TGF-B1 inhibits kappa acquisition in murine pre-B-cell clones (63), thereby regulating the transition between the pre-B-cell stage and the mature plasma cell stage with immunoglobulin-secreting ability. Antiproliferative effects of TGF- β in both murine and human mature B cells treated with exogenous TGF- β in vitro were subsequently reported (64, 65). Simultaneously, TGF- β also exerts a robust pro-apoptotic effect in human and murine B cells through Smad3-mediated Bim expression (pro-apoptotic member) (66, 67). B-cell-specific knockout mice with TGF-BRII deficiency show dramatically increased proliferation of peripheral B cells in response to a usually fragile immunogen, resulting in disrupted lifespan and

conventional B cell function (68). Interestingly, mice with B-cellspecific Smad2 deficiency exhibit normal B-cell development but an impaired switch to IgA and expanded peritoneal B1a cells in Peyer's patches (48), indicating a distinct context-dependent role of TGF- β in B-cell equilibrium. In summary, TGF- β impacts the establishment of the B-cell-based immune system from the early developmental stage to the mature functional stage.

iNKT Cells

iNKT cells and conventional T cells arise from the same doublepositive progenitor thymocytes, but unlike conventional T cells, iNKT cells mainly recognize endogenous and exogenous lipid antigens, which are presented by atypical MHC class I-like CD1d molecules. TGF- β signaling fine-tunes iNKT cell survival and development. A study in conditional knockout mice showed that TGF- β signaling regulates the precursor cell differentiation of iNKT cells, therefore impairing the maturation of iNKT cells in the thymus and in the periphery (69). A subsequent study identified the role of the TGF-B/SMAD4 axis in controlling RORyt⁺ iNKT subset development and operation during infection conditions (70). The role of the miR-17~92 family in regulating iNKT cell development and maturation by regulating TGF- β signaling was revealed by another group (71). Overall, TGF- β drives the intrathymic growth of iNKT cells and plays an essential role in maintaining the function of mature iNKT cells in the periphery.

JANUS-FACED REGULATION OF TGF-B SIGNALING IN TUMOR PROGRESSION

The dual role of TGF- β in tumor progression is highly context dependent, with a tumor-suppressing role in the beginning stages of carcinogenesis and a tumor-promoting role during subsequent tumor progression. In the early stage, TGF- β signals predominantly inhibit cell proliferation, promote apoptosis (72), and maintain genome stability as a tumor suppressor (73). TGF- β expression is low in epithelial cells but increases in hyperplastic and neoplastic tissues. In noncancerous and premalignant cells, TGF- β promotes robust retardation of cell cycle progression by inhibiting late G1 phase activation by increasing the expression of CDK inhibitors, including p15^{INK4} and p21^{\tilde{C} IP1} (12, 27, 74, 75). Additionally, TGF- β has been postulated to stimulate apoptosis through various mechanisms; for example, in HCC tumor models, the TGF-β/SMAD axis prompts c-Myc-induced apoptosis, resulting in the abolishment of tumor initiation (76). Moreover, TGF- β triggers cell apoptosis by SMAD-modulated production of death-associated protein kinase (40) and repression of the ID family members ID1, ID2, and ID3, which determine the efficiency of cell amplification and differentiation (77, 78). In addition, oncogenic stress from RAS is involved in TGF-β triggering cell death in premalignant cells. In a murine model of aging, TBRII deficiency leads to enhanced keratinocyte motility with a decline in apoptosis (79).

Paradoxically, the TGF- β signaling pathway is highly expressed in many advanced cancers and is correlated with

poor prognosis (80). TGF- β stimulates tumorigenesis via several mechanisms, including EMT, cell invasion, tumor metastasis, and immune suppression. EMT is a biological process in which epithelial progenitor cells undergo biochemical alteration and gradually lose polarity, resulting in enhanced migratory capacity, downregulation of cell-cell adhesion, and increased stem cell-like features (81). To induce EMT, TGF- β signaling activates the p38 and JNK pathways (5, 14) or the ERK1 and ERK2 pathways (82). Simultaneously, TGF- β signals foster cancer evolution and metastasis by stimulating tumor angiogenesis and the antitumor function of cancerassociated fibroblasts, allowing the tumor to evade antitumor immune responses in the TME (73). In bone metastases of breast cancer and prostate cancer, TGF- β elevates the expression of metastasis-related genes, including cxcr4, mmp1, and jag1 (83, 84). In mouse models of pancreatic and colon cancer, TGF- β exerts its antitumor functions primarily by delaying the transition of premalignant cells to malignant cells (12). Additionally, in a KrasG12D-mutant mouse model, the loss of SMAD4 accelerates progression to PDA by causing apoptosis in pancreatic progenitors (85). The overexpression of SMAD4 in SMAD4-deficient tumor cells inhibits tumorigenesis (13). Additionally, T-cell SMAD4 deficiency induces the automatic development of epithelial cancers in the gastrointestinal tract due to abnormally high levels of proinflammatory cytokines (86).

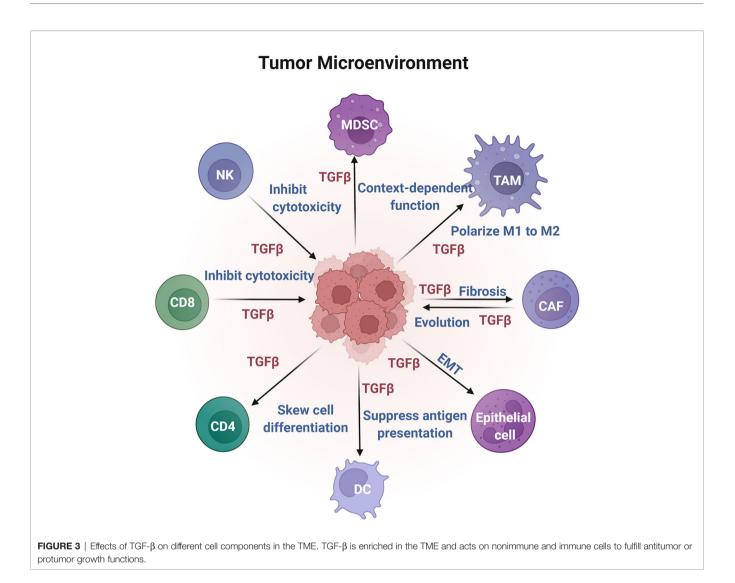
In summary, TGF- β functions as a tumor suppressor during the initial stage of tumor formation but gradually shows its evil side as a tumor promoter with tumor progression and metastasis. In recent years, growing evidence has reinforced the immunesuppressive role of TGF- β in the TME *via* multiple mechanisms, breaking the deadlock of utilizing TGF- β inhibitors for therapeutic purposes and providing new insights and strategies for targeting TGF- β as an immunotherapy. Considering the complexity of cell components in the TME, we have summarized the function of TGF- β in individual cell subsets in the TME (shown in **Figure 3**).

TGF-B EFFECTS ON NONIMMUNE CELLS IN THE TME

In addition to cancer cells, TGF- β is strongly associated with the regulation of CAF cells, epithelial cells, and immune cells in the TME (87). The regulation of nonimmune cells is discussed in the following sections.

CAFs

The TME is often characterized by an abundance of fibroblasts, termed CAFs, which are also the primary producers of TGF- β in various tumor types. Researchers have focused on TGF- β signaling as one of the underlying mechanisms of tissue fibrosis and tumorigenesis (88). CAFs originate from fibroblasts, which are responsive to inflammatory and tumor-derived signals (13, 89, 90). CAFs contribute to the synthesis of collagen along with the secretion of an array of soluble factors that promote tumor formation, invasiveness, metastasis (91–93),



and even chemoresistance (94, 95). Elevated numbers of CAFs are regularly found in tumor patients and are negatively correlated with disease prognosis (96, 97) and immunotherapy efficacy (98, 99). In colorectal cancer patients, all colorectal cancer subtypes with poor prognosis coexpress similar gene patterns induced by TGF- β in CAFs (100, 101). A colorectal cancer tumor-bearing mouse model confirmed that increased TGF- β levels produced by CAFs in the TME represent a fundamental mechanism of immune evasion that blocks the aggregation of CD4⁺ T helper and cytotoxic CD8⁺ T cells (42). Furthermore, TGF- β blockade perturbs fibroblast activity in the TME and augments T-cell penetration and activation (102).

Mechanistically, TGF- β signaling promotes fibroblastmyofibroblast transdifferentiation through either SMAD or non-SMAD signaling pathways (103, 104). On the one hand, p53 acts as a coactivator with p-SMAD3 to induce myofibroblast production and fibrosis under TGF- β 1 stimulation (105). On the other hand, the noncanonical Hippo signaling effectors YAP/TAZ cooperate with p-SMAD2/3 to drive renal fibrosis (106). In pancreatic cancers, TGF- β signaling antagonizes IL1-induced JAK/STAT activation by downregulating IL-1 receptor 1 (IL-1R1) expression and drives the commitment of iCAFs (inflammatory CAFs) to myoCAFs (myofibroblastic CAFs) (107). The crosstalk between CAFs and tumor cells through TGF- β may favor tumor progression. Although the function of TGF- β signaling in affecting CAFs has been clearly annotated, understanding the heterogeneity of the response of CAF cells to TGF- β or how to convert myCAFs into iCAFs may provide more valuable insights for unleashing the full potential of immunotherapies.

Epithelial Cells

Resting epithelial cells rarely show TGF- β expression; however, both TGF- β 1 and TGF- β receptor levels tend to be enhanced in hyperplasia and neoplasia (108, 109), indicating a tumor-promoting role of TGF- β in cancer advancement. Within tumors, elevated glucose uptake and hyperglycemia induce AKT activation and promote the externalization of TGF- β receptors, thus enhancing the cell surface presence of TGF- β receptors and TGF- β responsiveness (6). Autocrine and paracrine TGF- β represses epithelial characteristics, instructs the transdifferentiation of

TGF- β and Immune Cell Development

nontransformed epithelial cells toward a mesenchymal phenotype, and inhibits normal epithelial cell multiplication by blocking the cell division checkpoint at G1 phase (110). In response to TGF-β, epithelial cells acquire CSC-like phenotypes by concurrent hypomethylation and hypermethylation of EMT-regulating genes (8). Simultaneously, epithelial cells promote tumor progression by mitigating their epithelial characteristics and enhancing their migration and invasiveness. Moreover, TGF-B promotes epithelial plasticity by reprogramming gene expression patterns, especially enhancing EMT signature transcription factors such as Snail1, Snail2 ZEB1, and ZEB2, which cooperatively work with the SMAD3/4 complex to induce mesenchymal genes and restrain epithelial genes (111). Hence, most carcinoma cells gain proliferative properties by inactivating their epithelial antiproliferative properties and taking advantage of enhanced TGF- β signaling through effects on gene expression in epithelial plasticity.

TGF-B REGULATES THE IMMUNE CELL RESPONSE IN THE TME

The cellular sources of TGF- β in tumors are mainly carcinoma cells, stromal cells, and immune cells. Although most cells located in the tumor context respond to TGF- β in a context-dependent way, extensive evidence suggests that enriched TGF- β expression in the TME may compromise antitumor immunity and limit the efficacy of immunotherapy (80, 112). In the TME, TGF- β critically employs immunosuppressive functions by regulating immune populations, including adaptive B and T cells, innate natural killer cells, dendritic cells, myeloid tumor-associated macrophages, and myeloid-derived suppressor cells, as shown in **Figure 3**.

NK Cells

TGF- β signaling within the complex tumor environment restrains the antitumor activities of assorted cell subsets, including innate immune cells and adaptive immune cells. As innate immune cells, NK cells exhibit tumor-suppressing activity by activating receptors such as NKG2D and NKp30, which are silenced by TGF- β 1 directly and indirectly (113). Additionally, TGF- β silences IFN- γ and T-bet expression in NK cells (114). In TGF-β-rich TME models, TGF- β promotes the transition of NK cells into ILC1s, which are devoid of cytotoxic function (115). Moreover, SMAD4 blocks the transition of NK cells into ILC1s through noncanonical TGF-β signaling, as NK-cell-specific knockout of SMAD4 results in damaged effector NK cells and loss of metastasis control (116). Correspondingly, when tumor cells are inoculated into mice with TGF-B receptor 2 depletion in NK cells, knockout mice display significantly fewer metastases than control mice (117). In human metastatic breast cancer, TGF- β drives the metabolic malfunction of circulatory NK cells in patients (118). Blocking TGF- β and/or GARP can recover the metabolic condition and activity of NK cells, suggesting a promising strategy for enhancing NK-cell-based immunotherapies by targeting the GARP-TGF- β axis. Notably, TGF- β intensively disturbs the function of NK cells (119).

The critical metabolic checkpoint kinase mTOR is one of the targets underlying TGF- β signaling in NK cells (117). In addition, pharmaceutically targeting the αv integrin/TGF- β axis in combination with allogeneic NK cells in a glioblastoma stem cell (GSC)-engrafted mouse model promotes the antitumor function of NK cells and tumor growth (120). Further studies exploring the molecular mechanisms underlying the effects of TGF- β on NK cells will provide new ideas for improving NK-cell-based therapeutics (121)

DCs

DCs are the most powerful antigen-presenting cells and bridge the innate and adaptive immune responses by processing tumor antigens and presenting peptides to either CD4⁺ or CD8⁺ T cells. In vitro and in vivo studies have shown that TGF- β and IL-10 together suppress the maturation and activation of DCs (121). Adding TGF-B1 during the differentiation of DCs significantly amplifies the expression of DC-associated genes (122, 123) and promotes DC differentiation from progenitor CDP cells toward an mDC subset (124). In Smad7-deficient murine DC cells, the development of splenic CD8⁺CD103⁺ DCs is impaired, leading to an increased Treg cell number and resistance to autoimmune disease development (125). In mouse models of breast cancer and melanoma, increased TGF- β signaling suppresses tumor progression by increasing indoleamine 2,3-dioxygenase (IDO) in pDCs and increasing secretion of the myeloid cell attractor CCL22 (126). Moreover, blockade of both TGF- β receptor and IL-10 expression by DCs markedly enhances T-cell cytolytic activity toward cancer cells (127).

T Cells

TGF- β impacts multiple phases of the T cell response, including activation, migration, differentiation and proliferation in both the TME and tumor-draining lymph nodes. TGF-β directly reduces CXCR3 expression on CTLs by increasing the binding of SMAD2 to the CXCR3 promoter; hence, deletion of TGF- β receptor I in CD8⁺ T cells upregulates CXCR3 expression and improves CD8⁺ T-cell trafficking into tumors (128). In addition, TGF- β reduces the cytotoxicity of CD8⁺ T cells by impairing their secretion of perforin, granzyme, and IFN γ (129), which are imperative for CTL-mediated tumor killing (130). One model proposed that TGF- β induces the phosphorylation of ITIMs, which aid the recruitment of the inhibitory protein phosphatases SHP-1 and/or SHP-2 to attenuate TCR signaling (131). This model is further supported by the multifocal lymphoproliferative inflammation phenotype exhibited by mice with TGF- β depletion in T cells (132). In addition, TGF- β impedes the secretion of IL-2, which is necessary for the proliferation and response of CD8⁺ T cells. Additionally, TGF-B1 upregulates PD-1 levels on tumor antigen-specific TILs, further providing conclusive evidence supporting the tumor-promoting role of TGF- β through the regulation of CD8⁺ T cells (133).

The role of CD4⁺ T cells in tumor eradication has drawn increasing attention in recent years. Depleting TGF- β R2 in CD4⁺ T cells halts cancer development due to tissue repair and rebuilding of the tumor vasculature (47). In a recent study, Ming O Li's group formulated a bispecific receptor decoy

named the CD4 TGF- β Trap that was able to specifically function on CD4⁺ T cells instead of CD8⁺ T cells and exert a pronounced antitumor effect (134). Among all CD4⁺ Th subsets, Th1 cells are responsible for tumor killing by cytotoxic activity. TGF- β signaling suppresses the Th1 effector regulators T-BET and STAT4 (135, 136). Furthermore, TGF- β significantly promotes the expression of the transcription factor FoxP3 and drives regulatory T (Treg) cell development (38, 45), which suppresses CTL function in the antitumor response. Moreover, TGF- β from tumor effusions mediates the inhibition of mitochondrial respiration and the generation of IFN- γ in human CD4⁺ T subsets (137). In addition, ablating TGF- β RII in CD4⁺ T cells dramatically halts cancer progression (47). Most importantly, TGF- β skews the differentiation direction of TH1 cells into TH2 and TH17 cells (138).

TAMs

Myeloid cells are the main component of tumor-infiltrating leukocytes involved in tumorigenesis and are termed tumorassociated macrophages (TAMs). Importantly, TGF-B also affects the myeloid cell repertoire within tumors, including macrophages, MDSCs, and neutrophils. Accumulating evidence strongly indicates that the TME polarizes macrophages from the M1 to the M2 immunosuppressive protumoral subset, leading to immunosuppression and tumorigenesis. Additionally, TAMs can promote tumor cell migration and metastasis via the TGF-β2/NF-κB/Kindlin-2 pathway (139, 140). Therefore, TAMs are potential targets for tumor therapy (141). Malignant cells in the tumors generate TGF- β , thus upregulating CXCR4 on the monocyte cell surface, whereas perivascular fibroblasts produce the CXCR4 ligand CXCL12 to target these monocytes to the tumor, followed by monocytes differentiating into perivascular macrophages (142). Additionally, TGF- β signaling represses the anti-inflammatory role of macrophages through degradation of MYD88 to inactivate NF- κ B signaling (143). Moreover, genetic mice with myeloid cell-specific depletion of Tgfbr2 display less tumor metastasis (144).

MDSCs

Patients or mice bearing tumors always exhibit an increased number of circulating neutrophils, which are associated with a poor prognosis (145). The phenotypes of MDSCs (myeloidderived suppressive cells) are similar to those of monocytes and immature neutrophils in chronic infection, inflammation, or cancer. The functional difference between MDSCs and neutrophils is that T-cell proliferation suppression is mediated by MDSCs and not neutrophils (146). Therefore, it is broadly accepted that MDSCs significantly repress the cytotoxic function of effector cells and promote tumor angiogenesis and metastasis (147). The function of TGF- β on MDSCs in tumor evolution has been reported by several groups. In one study, TGF- β fine-tuned MDSC accumulation and activation in tumors by inducing microRNA-494 expression (147). Additionally, TGF- β directly affects monocytic-MDSC (Mo-MDSC) expansion and MDSC functions (148). This latter finding was further supported by results in melanoma and breast cancer mouse models, which showed that TGF- β affects the generation of the CXCR3 ligand CCL9 in MDSCs, leading to less immune cell accumulation and cancer cell survival (149). In addition, conditional knockout mice with Tgfbr2 deletion in myeloid cells are resistant to tumor metastasis as a result of MDSC dysfunction, implying a critical role of TGF- β in MDSCs during tumor metastasis (144). However, restricting TGF- β signaling in the tumor epithelium induces prominent accumulation of CCR1⁺ immature myeloid cells, which increase extension and penetration by cancer cells (150). Similarly, abrogating Tgfbr2 expression in carcinoma cells results in direct recruitment of migrating MDSCs into tumors *via* the upregulation of the SDF-1/CXCR4 and CXCL5/CXCR2 axes (151). The context-dependent function of TGF- β signaling in tumor-infiltrating MDSCs remains controversial, and the molecular mechanism needs to be explored.

Preclinical Studies Targeting TGF- β as a Cancer Therapy

The TGF-B/SMAD signaling pathway has been demonstrated to be an essential immune envision mechanism in both hematopoietic and solid tumors (152). In childhood B-ALL patients, TGF-B1 induced NK cell dysfunction to mediate escape from immune surveillance (153). In addition, TGF- β signaling blockade inhibited the proliferation of leukemia stem cells (154). In a mouse model with cancer-related anemia, pharmaceutically blockade of TGF- β signaling mitigated disease progression, suggesting a likely therapeutic target for alleviating hematopoiesis disease. Superfluous TGF- β in the bone marrow microenvironment impaired the bone marrow niches, which maintain the stemness and function of hematopoietic stem cells (155). CAR-T cell therapy has essentially reformed the therapeutic regimen of hematological malignancies (156). By constructing novel CD19 CAR-tTRII-I7R-T cells, which convert the TGF-β signaling into immuneactivating IL-7 signaling, the tumor-killing efficacy of modified CAR-T cells was significantly better than in the control group (157). The above studies proposed that TGF- β signaling prohibition could be a potential therapeutic approach for relieving defects in hematopoiesis.

Solid tumors show substantial antigen heterogeneity (156). Moreover, solid tumor cells are organized into multiple compartments and are often surrounded by other tissues, where they are less accessible to T cells. Tumor-bearing mouse models show that combined treatment with TGF- β inhibition and immune checkpoint antibodies such as anti-PD-L1 were able to induce better tumor regression and more prolonged survival (42, 99), leading the way to combining TGF- β inhibitors with ICB drugs clinically for advanced cancer patients. Besides, as a newly developed fusion protein targeting both TGF-B and PD-L1, Bintrafusp alfa (M7824) was demonstrated to effectively reduce the breast tumor and colon tumor growth in mice model (158). Hence, the clinical efficacy of M7824 in different advanced malignancies was evaluated in several ongoing clinical trials. Moreover, bifunctional antibody-ligand traps (Y-traps), which target both CTLA-4 and, resulted in significantly superior antitumor efficacy compared to CTLA-4 antibody monotherapy (159). When combining specific TGF- β 1 inhibitor SRK181mIgG1 with anti–PD-1 antibody in multiple mouse models of cancer, the combo therapy significantly induced the intratumoral infiltration of CD8⁺ T cells with less dose-limiting toxicology (160). These preclinical results provide a principle for exploring TGF β inhibition to work synergistically with ICB in cancer patients.

Current Clinical Trials Targeting TGF-Beta in the Tumor Microenvironment

With a growing number of preclinical studies demonstrating the role of TGF- β in suppressing immune response in different tumor types, more clinical trials have been conducted recently to evaluate the therapeutic safety and efficacy of targeting TGF- β in advanced cancer patients (73, 161). Up to now, however, both promising and undesirable outcomes have been revealed. We have summarized the finished clinical trials and ongoing trials that target TGF-B signals in advanced tumor patients in recent ten years (as shown in Tables 1, 2). Among them, Galunisertib (LY2157299) is a selective molecule inhibitor of RI kinase, which reduces signaling in the -rich immunosuppressive tumor microenvironment (162). A Phase I Study of Galunisertib in advanced tumor patients evaluated the safety and recommended the dose for the phase II study (163). Nevertheless, the co-administration of Galunisertib and anti-PD-L1 antibody in recurrent/refractory metastatic pancreatic cancer showed no apparent clinical activity (164). Another selective smallmolecule inhibitor Vactosertib (TEW-7197), which serves as a TGF-B R1 inhibitor, was well-assessed in its pharmacokinetics (165). In a Phase Ib trial for relapsed multiple myeloma, the combo therapy with vactosertib and pomalidomide showed better efficacy assessment than historical control, indicating the further application of Vactosertib in clinical trial multiple myeloma (166). In addition, several multi-center phase 2 studies for subjects with advanced or metastatic tumors are ongoing to estimate the safety and efficacy of vactosertib in association with pembrolizumab (NCT04515979) in lung cancer patients or vactosertib plus imatinib (NCT03802084) in desmoid tumor. Moreover, a pan-anti- neutralizing antibody, NIS793, has overcome the resistance of checkpoint blockade immunotherapy in the treatment of squamous cell carcinomas (167), which paves the way for the clinical trial of NIS793/ α -PD-1 combination therapy (NCT02947165). Meanwhile, this antibody is presently being evaluated together with gemcitabine/nab-paclitaxel chemotherapy and anti-PD-1 antibody for patients with metastatic pancreatic ductal adenocarcinoma in a phase II clinical trial (NCT04390763). Another pan- antibody SAR439459 is being estimated for its safety and antitumor activity either as monotherapy or together with the anti-PD-1antibody in patients with advanced solid tumors (NCT03192345).

Although significant progress was achieved, suspension or failure of anti- related clinical studies occurs in certain conditions. LY3022859 is a human anti-RII IgG1 monoclonal antibody, which significantly inhibited cancer cell growth and metastasis in preclinical models (168). However, in a phase I study of advanced tumor patients, owing to the burst of cytokine release syndrome, the study was stopped without determining the maximum tolerated dose (169). Moreover, a first-in-class bifunctional fusion protein Bintrafusp alfa (M7824), which targets both TGF- β and PD-L1, showed disease control regardless of PD-L1 in recurrent glioblastoma patients (170). Also, in biliary tract cancer patients, the M7824 monotherapy showed encouraging efficacy with durable responses (171). However, in the following phase III lung cancer clinical trials, treatment with Bintrafusp alfa was unable to show compelling efficacy, accompanied by the termination of three related trials till now. Even so, more than ten clinical trials related to Bintrafusp alfa combination therapy are ongoing in different cancer patients, indicating cancer context-dependent efficacy may be observed.

Immune checkpoint blockade therapies have achieved massive success in treating a variety of cancers. However, checkpoint inhibitors work to rejuvenate the body's immune activity instead of removing the immunosuppressive barriers in the tumor microenvironment (159). Subsequently, the efficacy of ICB therapies is limited in a minority of patients. Hence, the TGF- β pathway inactivation has emerged as a working partner for cancer patients with ICB resistance (160).

DISCUSSION

The determining function of TGF- β in maintaining immune system integrity is to preserve immune homeostasis and tolerance by regulating immune cell development, proliferation, differentiation, and survival. A competent immune system must maintain an assorted pool of naïve immune cells with the companion of various cytokines, including TGF- β . Malfunctions of TGF- β block immune cell development and function, resulting in cancer progression.

Because of the pleiotropic effects of TGF- β on both normal physiological function and tumorigenesis, long-term blockade of TGF- β and the related signaling pathways may have adverse effects. Furthermore, the biology of solid tumors is complex. Therefore, the current significant challenge in translating anti-TGF- β inhibition into clinical treatment is to explore the various function of the TGF- β signal pathway acting on different cell components in the tumors, thus searching for a precise targeting approach with less toxicity and other side effects.

Importantly, TGF- β serves as a primary immune evasion mechanism in various malignancies by building a tolerogenic immune environment. One hallmark of TGF- β 's evasion mechanism is promoting the amplification and aggregation of Treg cells in the tumors, which inhibit the cytotoxicity of CD8⁺T and Th1 cells. Besides, the TGF- β can be secreted and functions on stromal cells by an autocrine pathway and promote myelofibrosis and angiogenesis. Lately, the metabolic regulating role of TGF- β in inducing CAF and endothelial cell generation in cancer was revealed, indicating a novel role of TGF- β signaling in reprogramming the metabolic landscape of the tumor environment.

Cancer immunotherapy has indeed benefited patients who cannot receive surgical therapy or are resistant to chemotherapy. Repressing TGF- β signaling has shown a synergistic effect with immune checkpoint inhibitors in preclinical models and certain patients, providing a new solution for patients who are insensitive to

TABLE 1 Completed clinical trials to evaluate TGF- β pathway antagonists.

Target	Agent	Tumor type	Clinical efficacy	Starting date	Identifier
TGF-β2	AP12009 Temozolomide PCV	Glioblastoma Anaplastic Astrocytoma	Not reported	2007-02-06	NCT00431561
TGF-β2	Lucanix	Non-small cell lung cancer	Mos:20 versus 17 m	2008-05-13	NCT00676507
TGF-β2	AP 12009	Pancreatic Neoplasms Melanoma Colorectal Neoplasms	Not reported	2009-02-13	NCT00844064
СТ	LY2157299	Malignant Glioma	mOS:18.2 versus 17.9 m	2010-10-13	NCT01220271
RT	Radiation				
TβRI	Temozolomide				
TβRI	LY2157299 Sorafenib Ramucirumab	Hepatocellular Carcinoma	mPFS 2.7 m part A and 4.2 m part B	2010-11-24	NCT01246986
ΤβRΙ	Galunisertib Gemcitabine	Advanced or Metastatic Unresectable Pancreatic Cancer	mOS 8.9 versus 7.1 m	2011-06-14	NCT01373164
TGF-β1 TGF-β2	Placebo Fresolimumab RT	Refractory breast cancer	ORR 0%	2011-07-25	NCT01401062
TGF-β3 TβRI	LY2157299	Glioblastoma		2012-04-20	NCT01582269
	monohydrate Lomustine Placebo				
TβRII	LY3022859	Advance solid tumors	Not reported	2012-07-20	NCT01646203
ΤβRΙ	LY2157299 Gemcitabine	Inoperable or metastatic pancreatic cancer	ORR 0%		NCT02154646
TβRI	TEW-7197	Advanced Stage Solid Tumors		2014-06-10	NCT02160106
TβRI	LY2157299 Sorafenib	Metastatic hepatocellular carcinoma	ORR 9%	2014-09-15	NCT02240433
TGF-β RI	LY2157299 Lomustine	Glioma	ORR 14%		NCT01682187
TGF-β RI	Galunisertib Durvalumab	Metastatic Pancreatic Cancer	ORR 3%		NCT02734160
TGF-β RII PD-L1	M7824	Human Papilloma VirusCervical Cancer Oropharyngeal CancerAnal Cancer Vaginal or Penile Cancer	ORR 39%	2018-02-09	NCT03427411
TGF-β RII PD-L1	M7824	Advanced Adenocarcinoma of the Pancreas	Study was closed after one treatment related death.	2018-03-02	NCT03451773
TGF-β RII PD-L1	MSB0011359C	Metastatic or Locally Advanced Solid Tumors		2015-08-07	NCT02517398
TGF-β RII/PD- L1		Pre-treated cervical tumors	ORR 28%		NCT02517398
TGF-β RII/PD- L1		Refractory head and neck cancer	ORR 22%		NCT02517398
TGF-β RII/PD- L1		Pre-treated NSCLC	PD-L1 > 1%, ORR 40%, PD-L1 > 80%, ORR 71%		NCT02517398
TGF-β RII PD-L1	M7824	Pre-treated esophageal adenocarcinoma	ORR 20%		NCT02517398
TGF-β RII PD-L1	M7824	Pre-treated gastric cancer	ORR 22%		NCT02517398
TGF-β RII PD-L1	M7824	Pre-treated biliary tract cancer	ORR 23%		NCT02517398
TGF-β RII PD-L1	M7824	Refractory colorectal cancer	ORR 3.4%		NCT02517398
Vaccine Anti-PD-1	Vigil Pembrolizumab	Melanoma		2015-10-14	NCT02574533

The underlined NCT Number represents the National Clinical Trial number, which is an identification that ClinicalTrials.gov assigns a study when it is registered. The NCT number is assigned when the study is registered.

ICB treatment. Moreover, modified CAR-T cell therapy by depleting the TGF- β signal in CAR-T cells also demonstrates potent efficacy in treating cancers, indicating a promising field by generating more robust and less toxic CAR-T cells by modifying the TGF- β signal pathway.

Given the essential role of the TGF- β signal pathway in physiological and pathological conditions, it's expected that the integral blockade of the TGF- β signal pathway resulted in adverse effects that restrict the therapeutic progression. Hence, new therapeutic modalities with more precise

TABLE 2 | Ongoing clinical trials to evaluate TGF- β pathway antagonists.

Target	Agent	Tumor type	Starting date	identifier
TβRI	Galunisertib	Metastatic Castration-resistant Prostate Cancer	2015-05-22	NCT02452008
AR	Enzalutamide			
α-TGF-β	NIS793	Breast Cancer	2016-10-27	NCT02947165
α- PD - 1	PDR001	Lung Cancer		
		Hepatocellular Cancer		
		Colorectal Cancer		
		Pancreatic Cancer		
		Renal Cancer		
TGF-βR1	Vactosertib	Multiple Myeloma	2017-05-08	NCT03143985
ALK5	Pomalidomide			
TGF-β	M7824	Pretreated MSI-H mCRC	2018-02-19	NCT03436563
PD-L1			2010 02 10	110100100000
PD-L1	M7825	Breast Cancer	2018-05-14	NCT03524170
TGF-β	1017 823	Dieast Galicei	2010-00-14	<u>NO103324170</u>
•	M7004	TNPO	0010 07 06	
PDL1	M7824	TNBC	2018-07-06	NCT03579472
TGF-β	Eribulin Mesylate			10700554470
PD-L1	M7824	SCLC	2018-06-13	NCT03554473
TGF-β	Topotecan			
	Temozolomide			
TGF-β	Vactosertib	Desmoid Tumor	2019-01-14	NCT03802084
	imatinib			
TGF-β	Platinum-based regimen +	Metastatic NSCLC	2019-02-15	NCT03840915
PD-L1	M7824			
TGF-β	M7824 With cCRT	NSCLC	2019-02-15	NCT03840902
PD-L1				
TGF-βR1	Pembrolizumab	Colorectal Cancer	2019-02-18	NCT03844750
ion pitt	vactosertib	Resectable Hepatic Metastases	2010 02 10	10100011100
TGF-βR1	Vactosertib	Myeloproliferative Neoplasm	2019-09-25	NCT04103645
PD-L1/TGF-β	M7824	Local-Regionally Recurrent Head and Neck Squamous Cell Carcinoma	2020-01-07	NCT04220775
PD-L1		Cervical Cancer		
	M7824	Cervical Caricer	2020-01-29	NCT04246489
TGF-β		First line Materiatic Descuration Dustel Adams and since	0000 05 10	NOTO 4000700
TGF-β	Gemcitabine	First-line Metastatic Pancreatic Ductal Adenocarcinoma	2020-05-18	NCT04390763
PD-1	nab-paclitaxel			
	NIS793			
	Spartalizumab			
PDL1	M7824	Thymoma	2020-06-05	NCT04417660
TGF-β		Thymic Carcinoma		
PD-L1	M7824	Thymic Epithelial Tumor	2020-06-05	NCT04417660
TGF-β				
PD-L1	M7824	Recurrent Thymoma	2020-06-05	NCT04417660
TGF-β				
PD-L1	M7824	Thymic Cancer	2020-06-05	NCT04417660
TGF-β			2020 00 00	110101111000
PD-L1	PRGN-2009	Cervical cancers	2020-06-16	NCT04432597
	M7824	p16+ Oropharyngeal cancers	2020-00-10	10104402001
TGF-β	11/1 024	Anal cancers		
		Other locally advanced or metastatic solid tumors (e.g. lung, esophagus) that are		
		known HPV+.	0000	
TGF-βR1	Vactosertib 300 mg BID	Non-Small-Cell Lung	2020-08-17	NCT04515979
ALK5	pembrolizumab 200 mg IV			
PD-L1	M7824	Cervical	2021-01-14	NCT04708470
TGF-β	NHS-IL12	Oropharyngeal		
	Entinostat	Anal		
		Vulvar		
		Vaginal		
		<u> </u>		

The underlined NCT Number represents the National Clinical Trial number, which is an identification that ClinicalTrials.gov assigns a study when it is registered. The NCT number is assigned when the study is registered.

targeting or more assorted regimen design still need further exploration. In the future, specific targeting TGF- β in certain types of immunosuppressive cell components may reduce the incidence and hardness of adverse effects and increase beneficial efficacy.

TGF- β signal pathway was hyperactivated in colon cancer and pancreatic cancer. However, different tumors harbor diverse tumor microenvironments, either with low levels or high levels of TGF- β . Accordingly, more studies are needed to explore whether the tumor architecture or landscape influences the efficacy of TGF- β inhibition. Besides, whether genetic, epigenetic, and microbiota differences in different cancer types define the efficacy of TGF- β signal pathway blockade also needs to be studied. All things considered, we will be able to reach a more precise and personalized modality for treating cancers by anti-TGF- β pathway.

REFERENCES

- Jurberg AD, Vasconcelos-Fontes L, Cotta-de-Almeida V. Tale From TGF-Beta Superfamily for Thymus Ontogeny and Function. *Front Immunol* (2015) 6:442. doi: 10.3389/fimmu.2015.00442
- Letterio JJ, Roberts AB. Regulation of Immune Responses by TGF-Beta. Annu Rev Immunol (1998) 16:137–61. doi: 10.1146/annurev.immunol.16.1.137
- David CJ, Massague J. Contextual Determinants of TGFbeta Action in Development, Immunity and Cancer. Nat Rev Mol Cell Biol (2018) 19:419– 35. doi: 10.1038/s41580-018-0007-0
- Chung JY, Chan MK, Li JS, Chan AS, Tang PC, Leung KT, et al. TGF-Beta Signaling: From Tissue Fibrosis to Tumor Microenvironment. *Int J Mol Sci* (2021) 22(14):7575. doi: 10.3390/ijms22147575
- Kim BN, Ahn DH, Kang N, Yeo CD, Kim YK, Lee KY, et al. TGF-Beta Induced EMT and Stemness Characteristics Are Associated With Epigenetic Regulation in Lung Cancer. *Sci Rep* (2020) 10:10597(570):eaav5183. doi: 10.1038/s41598-020-67325-7
- Derynck R, Budi EH. Specificity, Versatility, and Control of TGF-Beta Family Signaling. *Sci Signal* (2019) 12(570):eaav5183. doi: 10.1126/ scisignal.aav5183
- Travis MA, Sheppard D. TGF-Beta Activation and Function in Immunity. *Annu Rev Immunol* (2014) 32:51–82. doi: 10.1146/annurev-immunol-032713-120257
- Dongre A, Weinberg RA. New Insights Into the Mechanisms of Epithelial-Mesenchymal Transition and Implications for Cancer. *Nat Rev Mol Cell Biol* (2019) 20:69–84. doi: 10.1038/s41580-018-0080-4
- Tauriello DVF, Sancho E, Batlle E. Overcoming TGFbeta-Mediated Immune Evasion in Cancer. *Nat Rev Cancer* (2021) 22(1):25–44. doi: 10.1038/s41568-021-00413-6
- Yu Q, Stamenkovic I. Cell Surface-Localized Matrix Metalloproteinase-9 Proteolytically Activates TGF-Beta and Promotes Tumor Invasion and Angiogenesis. *Genes Dev* (2000) 14:163–76. doi: 10.1101/gad.14.2.163
- Wang Q, Zou Y, Nowotschin S, Kim SY, Li QV, Soh CL, et al. The P53 Family Coordinates Wnt and Nodal Inputs in Mesendodermal Differentiation of Embryonic Stem Cells. *Cell Stem Cell* (2017) 20:70–86. doi: 10.1016/j.stem.2016.10.002
- Batlle E, Massague J. Transforming Growth Factor-Beta Signaling in Immunity and Cancer. *Immunity* (2019) 50:924–40. doi: 10.1016/ j.immuni.2019.03.024
- Duda DG, Sunamura M, Lefter LP, Furukawa T, Yokoyama T, Yatsuoka T, et al. Restoration of SMAD4 by Gene Therapy Reverses the Invasive Phenotype in Pancreatic Adenocarcinoma Cells. Oncogene (2003) 22:6857–64. doi: 10.1038/sj.onc.1206751
- van der Heide LP, van Dinther M, Moustakas A, ten Dijke P. TGF Beta Activates Mitogen- and Stress-Activated Protein Kinase-1 (MSK1) to Attenuate Cell Death. J Biol Chem (2011) 286:5003–11. doi: 10.1074/ jbc.M110.167379
- Valcourt U, Kowanetz M, Niimi H, Heldin CH, Moustakas A. TGF-Beta and the Smad Signaling Pathway Support Transcriptomic Reprogramming During Epithelial-Mesenchymal Cell Transition. *Mol Biol Cell* (2005) 16:1987–2002. doi: 10.1091/mbc.e04-08-0658
- Kulkarni AB, Huh CG, Becker D, Geiser A, Lyght M, Flanders KC, et al. Transforming Growth Factor Beta 1 Null Mutation in Mice Causes Excessive Inflammatory Response and Early Death. *Proc Natl Acad Sci USA* (1993) 90:770–4. doi: 10.1073/pnas.90.2.770
- Shull MM, Ormsby I, Kier AB, Pawlowski S, Diebold RJ, Yin M, et al. Targeted Disruption of the Mouse Transforming Growth Factor-Beta 1 Gene Results in Multifocal Inflammatory Disease. *Nature* (1992) 359:693–9. doi: 10.1038/359693a0

AUTHOR CONTRIBUTIONS

BC, CM, and ZZ wrote the manuscript. XH and XL wrote and critically revised the manuscript. All authors contributed to the article and approved the submitted version.

- Junker U, Knoefel B, Nuske K, Rebstock K, Steiner T, Wunderlich H, et al. Transforming Growth Factor Beta 1 Is Significantly Elevated in Plasma of Patients Suffering From Renal Cell Carcinoma. *Cytokine* (1996) 8:794–8. doi: 10.1006/cyto.1996.0105
- Batard P, Monier MN, Fortunel N, Ducos K, Sansilvestri-Morel P, Phan T, et al. TGF-(Beta)1 Maintains Hematopoietic Immaturity by a Reversible Negative Control of Cell Cycle and Induces CD34 Antigen Up-Modulation. *J Cell Sci* (2000) 113:383–90:(Pt 3). doi: 10.1242/jcs.113.3.383
- 20. Sitnicka E, Ruscetti FW, Priestley GV, Wolf NS, Bartelmez SH. Transforming Growth Factor Beta 1 Directly and Reversibly Inhibits the Initial Cell Divisions of Long-Term Repopulating Hematopoietic Stem Cells. *Blood* (1996) 88:82-8. doi: 10.1182/blood.V88.1.82. bloodjournal88182
- Zhao M, Perry JM, Marshall H, Venkatraman A, Qian P, He XC, et al. Megakaryocytes Maintain Homeostatic Quiescence and Promote Post-Injury Regeneration of Hematopoietic Stem Cells. *Nat Med* (2014) 20:1321–6. doi: 10.1038/nm.3706
- 22. Yamazaki S, Ema H, Karlsson G, Yamaguchi T, Miyoshi H, Shioda S, et al. Nonmyelinating Schwann Cells Maintain Hematopoietic Stem Cell Hibernation in the Bone Marrow Niche. *Cell* (2011) 147:1146–58. doi: 10.1016/j.cell.2011.09.053
- Capron C, Lacout C, Lecluse Y, Jalbert V, Chagraoui H, Charrier S, et al. A Major Role of TGF-Beta1 in the Homing Capacities of Murine Hematopoietic Stem Cell/Progenitors. *Blood* (2010) 116:1244–53. doi: 10.1182/blood-2009-05-221093
- Brenet F, Kermani P, Spektor R, Rafii S, Scandura JM. TGFbeta Restores Hematopoietic Homeostasis After Myelosuppressive Chemotherapy. J Exp Med (2013) 210:623–39. doi: 10.1084/jem.20121610
- Larsson J, Blank U, Helgadottir H, Bjornsson JM, Ehinger M, Goumans MJ, et al. TGF-Beta Signaling-Deficient Hematopoietic Stem Cells Have Normal Self-Renewal and Regenerative Ability In Vivo Despite Increased Proliferative Capacity In Vitro. Blood (2003) 102:3129–35. doi: 10.1182/ blood-2003-04-1300
- Larsson J, Blank U, Klintman J, Magnusson M, Karlsson S. Quiescence of Hematopoietic Stem Cells and Maintenance of the Stem Cell Pool Is Not Dependent on TGF-Beta Signaling *In Vivo. Exp Hematol* (2005) 33:592–6. doi: 10.1016/j.exphem.2005.02.003
- Scandura JM, Boccuni P, Massague J, Nimer SD. Transforming Growth Factor Beta-Induced Cell Cycle Arrest of Human Hematopoietic Cells Requires P57kip2 Up-Regulation. *Proc Natl Acad Sci USA* (2004) 101:15231–6. doi: 10.1073/pnas.0406771101
- Yamazaki S, Iwama A, Takayanagi S, Morita Y, Eto K, Ema H, et al. Cytokine Signals Modulated *via* Lipid Rafts Mimic Niche Signals and Induce Hibernation in Hematopoietic Stem Cells. *EMBO J* (2006) 25:3515–23. doi: 10.1038/sj.emboj.7601236
- Yamazaki S, Iwama A, Takayanagi S, Eto K, Ema H, Nakauchi H. TGF-Beta as a Candidate Bone Marrow Niche Signal to Induce Hematopoietic Stem Cell Hibernation. *Blood* (2009) 113:1250–6. doi: 10.1182/blood-2008-04-146480
- Letterio JJ, Geiser AG, Kulkarni AB, Dang H, Kong L, Nakabayashi T, et al. Autoimmunity Associated With TGF-Beta1-Deficiency in Mice Is Dependent on MHC Class II Antigen Expression. J Clin Invest (1996) 98:2109–19. doi: 10.1172/JCI119017
- McCarron MJ, Irla M, Serge A, Soudja SM, Marie JC. Transforming Growth Factor-Beta Signaling in Alphabeta Thymocytes Promotes Negative Selection. Nat Commun (2019) 10:5690. doi: 10.1038/s41467-019-13456-z
- Park JH, Yu Q, Erman B, Appelbaum JS, Montoya-Durango D, Grimes HL, et al. Suppression of IL7Ralpha Transcription by IL-7 and Other Prosurvival Cytokines: A Novel Mechanism for Maximizing IL-7-Dependent T Cell Survival. *Immunity* (2004) 21:289–302. doi: 10.1016/j.immuni.2004.07.016

- 33. Mayer CT, Floess S, Baru AM, Lahl K, Huehn J, Sparwasser T. CD8+ Foxp3+ T Cells Share Developmental and Phenotypic Features With Classical CD4+ Foxp3+ Regulatory T Cells But Lack Potent Suppressive Activity. *Eur J Immunol* (2011) 41:716–25. doi: 10.1002/eji.201040913
- 34. Robb RJ, Lineburg KE, Kuns RD, Wilson YA, Raffelt NC, Olver SD, et al. Identification and Expansion of Highly Suppressive CD8(+)FoxP3(+) Regulatory T Cells After Experimental Allogeneic Bone Marrow Transplantation. *Blood* (2012) 119:5898–908. doi: 10.1182/blood-2011-12-396119
- Liu Y, Zhang P, Li J, Kulkarni AB, Perruche S, Chen W. A Critical Function for TGF-Beta Signaling in the Development of Natural CD4+CD25+Foxp3+ Regulatory T Cells. *Nat Immunol* (2008) 9:632–40. doi: 10.1038/ni.1607
- Ouyang W, Beckett O, Ma Q, Li MO. Transforming Growth Factor-Beta Signaling Curbs Thymic Negative Selection Promoting Regulatory T Cell Development. *Immunity* (2010) 32:642–53. doi: 10.1016/j.immuni.2010.04.012
- Gorelik L, Flavell RA. Abrogation of TGFbeta Signaling in T Cells Leads to Spontaneous T Cell Differentiation and Autoimmune Disease. *Immunity* (2000) 12:171–81. doi: 10.1016/s1074-7613(00)80170-3
- Marie JC, Liggitt D, Rudensky AY. Cellular Mechanisms of Fatal Early-Onset Autoimmunity in Mice With the T Cell-Specific Targeting of Transforming Growth Factor-Beta Receptor. *Immunity* (2006) 25:441–54. doi: 10.1016/j.immuni.2006.07.012
- O'Garra A, Gabrysova L, Spits H. Quantitative Events Determine the Differentiation and Function of Helper T Cells. *Nat Immunol* (2011) 12:288–94. doi: 10.1038/ni.2003
- Jang CW, Chen CH, Chen CC, Chen JY, Su YH, Chen RH. TGF-Beta Induces Apoptosis Through Smad-Mediated Expression of DAP-Kinase. *Nat Cell Biol* (2002) 4:51–8. doi: 10.1038/ncb731
- 41. Mlecnik B, Bindea G, Kirilovsky A, Angell HK, Obenauf AC, Tosolini M, et al. The Tumor Microenvironment and Immunoscore Are Critical Determinants of Dissemination to Distant Metastasis. *Sci Transl Med* (2016) 8:327ra326. doi: 10.1126/scitranslmed.aad6352
- Tauriello DVF, Palomo-Ponce S, Stork D, Berenguer-Llergo A, Badia-Ramentol J, Iglesias M, et al. TGFbeta Drives Immune Evasion in Genetically Reconstituted Colon Cancer Metastasis. *Nature* (2018) 554:538–43. doi: 10.1038/nature25492
- Sad S, Mosmann TR. Single IL-2-Secreting Precursor CD4 T Cell Can Develop Into Either Th1 or Th2 Cytokine Secretion Phenotype. J Immunol (1994) 153:3514–22. doi: 10.1016/1074-7613(95)90051-9
- 44. Kuwahara M, Yamashita M, Shinoda K, Tofukuji S, Onodera A, Shinnakasu R, et al. The Transcription Factor Sox4 Is a Downstream Target of Signaling by the Cytokine TGF-Beta and Suppresses T(H)2 Differentiation. *Nat Immunol* (2012) 13:778–86. doi: 10.1038/ni.2362
- Li MO, Sanjabi S, Flavell RA. Transforming Growth Factor-Beta Controls Development, Homeostasis, and Tolerance of T Cells by Regulatory T Cell-Dependent and -Independent Mechanisms. *Immunity* (2006) 25:455–71. doi: 10.1016/j.immuni.2006.07.011
- Pillai MR, Lian S, Bix M. Mina: A Th2 Response Regulator Meets TGFbeta. Curr Opin Immunol (2014) 31:38–43. doi: 10.1016/j.coi.2014.09.005
- Liu M, Kuo F, Capistrano KJ, Kang D, Nixon BG, Shi W, et al. TGF-Beta Suppresses Type 2 Immunity to Cancer. *Nature* (2020) 587:115–20. doi: 10.1038/s41586-020-2836-1
- Liu M, Li S, Li MO. TGF-Beta Control of Adaptive Immune Tolerance: A Break From Treg Cells. *Bioessays* (2018) 40:e1800063. doi: 10.1002/ bies.201800063
- Kim SV, Xiang WV, Kwak C, Yang Y, Lin XW, Ota M, et al. GPR15-Mediated Homing Controls Immune Homeostasis in the Large Intestine Mucosa. *Science* (2013) 340:1456–9. doi: 10.1126/science.1237013
- Zheng Y, Josefowicz S, Chaudhry A, Peng XP, Forbush K, Rudensky AY. Role of Conserved Non-Coding DNA Elements in the Foxp3 Gene in Regulatory T-Cell Fate. *Nature* (2010) 463:808–12. doi: 10.1038/nature08750
- Tone Y, Furuuchi K, Kojima Y, Tykocinski ML, Greene MI, Tone M. Smad3 and NFAT Cooperate to Induce Foxp3 Expression Through Its Enhancer. *Nat Immunol* (2008) 9:194–202. doi: 10.1038/ni1549
- Takimoto T, Wakabayashi Y, Sekiya T, Inoue N, Morita R, Ichiyama K, et al. Smad2 and Smad3 Are Redundantly Essential for the TGF-Beta-Mediated Regulation of Regulatory T Plasticity and Th1 Development. *J Immunol* (2010) 185:842–55. doi: 10.4049/jimmunol.0904100

- Zhou L, Lopes JE, Chong MM, Ivanov II, Min R, Victora GD, et al. TGF-Beta-Induced Foxp3 Inhibits T(H)17 Cell Differentiation by Antagonizing RORgammat Function. *Nature* (2008) 453:236–40. doi: 10.1038/ nature06878
- 54. Aarts J, Van Caam A, Helsen M, Walgreen B, Vitters E, Van de Loo F, et al. Ab0082 Inhibition Of Tgfβ Signaling Using Sb-505124 Blocks Th17 Differentiation And Restores The Th17/Treg Balance *In Vivo*, But Does Not Suppress Experimental Arthritis. *Ann Rheumatic Dis* (2020) 79:1341–1. doi: 10.1136/annrheumdis-2020-eular.1301
- 55. Xue G, Jin G, Fang J, Lu Y. IL-4 Together With IL-1beta Induces Antitumor Th9 Cell Differentiation in the Absence of TGF-Beta Signaling. *Nat Commun* (2019) 10:1376. doi: 10.1038/s41467-019-09401-9
- Putheti P. Polarizing Cytokines for Human Th9 Cell Differentiation. Methods Mol Biol (2017) 1585:73–82. doi: 10.1007/978-1-4939-6877-0_6
- Tsuda M, Hamade H, Thomas LS, Salumbides BC, Potdar AA, Wong MH, et al. A Role for BATF3 in TH9 Differentiation and T-Cell-Driven Mucosal Pathologies. *Mucosal Immunol* (2019) 12:644–55. doi: 10.1038/s41385-018-0122-4
- Nakatsukasa H, Zhang D, Maruyama T, Chen H, Cui K, Ishikawa M, et al. The DNA-Binding Inhibitor Id3 Regulates IL-9 Production in CD4(+) T Cells. Nat Immunol (2015) 16:1077–84. doi: 10.1038/ni.3252
- Wilhelm C, Hirota K, Stieglitz B, Van Snick J, Tolaini M, Lahl K, et al. An IL-9 Fate Reporter Demonstrates the Induction of an Innate IL-9 Response in Lung Inflammation. *Nat Immunol* (2011) 12:1071–7. doi: 10.1038/ni.2133
- Chang HC, Sehra S, Goswami R, Yao W, Yu Q, Stritesky GL, et al. The Transcription Factor PU.1 Is Required for the Development of IL-9-Producing T Cells and Allergic Inflammation. *Nat Immunol* (2010) 11:527–34. doi: 10.1038/ni.1867
- Pei S, Huang M, Huang J, Zhu X, Wang H, Romano S, et al. BFAR Coordinates TGFbeta Signaling to Modulate Th9-Mediated Cancer Immunotherapy. J Exp Med (2021) 218(7):e20202144. doi: 10.1084/jem.20202144
- Kersten C, Dosen G, Myklebust JH, Sivertsen EA, Hystad ME, Smeland EB, et al. BMP-6 Inhibits Human Bone Marrow B Lymphopoiesis–Upregulation of Id1 and Id3. *Exp Hematol* (2006) 34:72–81. doi: 10.1016/ j.exphem.2005.09.010
- Lee G, Ellingsworth LR, Gillis S, Wall R, Kincade PW. Beta Transforming Growth Factors Are Potential Regulators of B Lymphopoiesis. J Exp Med (1987) 166:1290–9. doi: 10.1084/jem.166.5.1290
- Petit-Koskas E, Genot E, Lawrence D, Kolb JP. Inhibition of the Proliferative Response of Human B Lymphocytes to B Cell Growth Factor by Transforming Growth Factor-Beta. *Eur J Immunol* (1988) 18:111–6. doi: 10.1002/eji.1830180117
- Kehrl JH, Taylor A, Kim SJ, Fauci AS. Transforming Growth Factor-Beta is a Potent Negative Regulator of Human Lymphocytes. Ann N Y Acad Sci (1991) 628:345–53. doi: 10.1111/j.1749-6632.1991.tb17267.x
- Lomo J, Blomhoff HK, Beiske K, Stokke T, Smeland EB. TGF-Beta 1 and Cyclic AMP Promote Apoptosis in Resting Human B Lymphocytes. *J Immunol* (1995) 154:1634–43.
- Wildey GM, Patil S, Howe PH. Smad3 Potentiates Transforming Growth Factor Beta (TGFbeta)-Induced Apoptosis and Expression of the BH3-Only Protein Bim in WEHI 231 B Lymphocytes. J Biol Chem (2003) 278:18069– 77. doi: 10.1074/jbc.M211958200
- Cazac BB, Roes J. TGF-Beta Receptor Controls B Cell Responsiveness and Induction of IgA In Vivo. Immunity (2000) 13::443–51. doi: 10.1016/s1074-7613(00)00044-3
- Doisne JM, Bartholin L, Yan KP, Garcia CN, Duarte N, Le Luduec JB, et al. iNKT Cell Development is Orchestrated by Different Branches of TGF-Beta Signaling. J Exp Med (2009) 206:1365–78. doi: 10.1084/jem.20090127
- Havenar-Daughton C, Li S, Benlagha K, Marie JC. Development and Function of Murine RORgammat+ iNKT Cells Are Under TGF-Beta Signaling Control. *Blood* (2012) 119:3486–94. doi: 10.1182/blood-2012-01-401604
- Fedeli M, Riba M, Garcia Manteiga JM, Tian L, Vigano V, Rossetti G, et al. miR-17 Approximately 92 Family Clusters Control iNKT Cell Ontogenesis via Modulation of TGF-Beta Signaling. Proc Natl Acad Sci U S A (2016) 113: E8286–95. doi: 10.1073/pnas.1612024114
- Drabsch Y, ten Dijke P. TGF-Beta Signalling and Its Role in Cancer Progression and Metastasis. *Cancer Metastasis Rev* (2012) 31:553-68. doi: 10.1007/s10555-012-9375-7

- Colak S, Ten Dijke P. Targeting TGF-Beta Signaling in Cancer. Trends Cancer (2017) 3:56-71. doi: 10.1016/j.trecan.2016.11.008
- Seoane J, Le HV, Shen L, Anderson SA, Massague J. Integration of Smad and Forkhead Pathways in the Control of Neuroepithelial and Glioblastoma Cell Proliferation. *Cell* (2004) 117:211–23. doi: 10.1016/s0092-8674(04)00298-3
- Sandhu C, Garbe J, Bhattacharya N, Daksis J, Pan CH, Yaswen P, et al. Transforming Growth Factor Beta Stabilizes P15ink4b Protein, Increases P15ink4b-Cdk4 Complexes, and Inhibits Cyclin D1-Cdk4 Association in Human Mammary Epithelial Cells. *Mol Cell Biol* (1997) 17:2458–67. doi: 10.1128/MCB.17.5.2458
- 76. Wang H, Wang P, Xu M, Song X, Wu H, Evert M, et al. Distinct Functions of Transforming Growth Factor-Beta Signaling in C-MYC Driven Hepatocellular Carcinoma Initiation and Progression. *Cell Death Dis* (2021) 12:200. doi: 10.1038/s41419-021-03488-z
- 77. Kowanetz M, Valcourt U, Bergstrom R, Heldin CH, Moustakas A. Id2 and Id3 Define the Potency of Cell Proliferation and Differentiation Responses to Transforming Growth Factor Beta and Bone Morphogenetic Protein. *Mol Cell Biol* (2004) 24:4241–54. doi: 10.1128/MCB.24.10.4241-4254.2004
- Papaspyridonos M, Matei I, Huang Y, do Rosario Andre M, Brazier-Mitouart H, Waite JC, et al. Id1 Suppresses Anti-Tumour Immune Responses and Promotes Tumour Progression by Impairing Myeloid Cell Maturation. *Nat Commun* (2015) 6:6840. doi: 10.1038/ncomms7840
- Guasch G, Schober M, Pasolli HA, Conn EB, Polak L, Fuchs E. Loss of TGFbeta Signaling Destabilizes Homeostasis and Promotes Squamous Cell Carcinomas in Stratified Epithelia. *Cancer Cell* (2007) 12:313–27. doi: 10.1016/j.ccr.2007.08.020
- Akhurst RJ, Hata A. Targeting the TGFbeta Signalling Pathway in Disease. Nat Rev Drug Discov (2012) 11:790–811. doi: 10.1038/nrd3810
- 81. Valastyan S, Weinberg RA. Tumor Metastasis: Molecular Insights and Evolving Paradigms. *Cell* (2011) 147:275–92. doi: 10.1016/j.cell.2011.09.024
- Lee MK, Pardoux C, Hall MC, Lee PS, Warburton D, Qing J, et al. TGF-Beta Activates Erk MAP Kinase Signalling Through Direct Phosphorylation of ShcA. *EMBO J* (2007) 26:3957–67. doi: 10.1038/sj.emboj.7601818
- Kang Y, He W, Tulley S, Gupta GP, Serganova I, Chen CR, et al. Breast Cancer Bone Metastasis Mediated by the Smad Tumor Suppressor Pathway. *Proc Natl Acad Sci USA* (2005) 102:13909–14. doi: 10.1073/pnas.0506517102
- Yin JJ, Selander K, Chirgwin JM, Dallas M, Grubbs BG, Wieser R, et al. TGF-Beta Signaling Blockade Inhibits PTHrP Secretion by Breast Cancer Cells and Bone Metastases Development. J Clin Invest (1999) 103:197–206. doi: 10.1172/JCI3523
- Bardeesy N, Cheng KH, Berger JH, Chu GC, Pahler J, Olson P, et al. Smad4 is Dispensable for Normal Pancreas Development Yet Critical in Progression and Tumor Biology of Pancreas Cancer. *Genes Dev* (2006) 20:3130–46. doi: 10.1101/gad.1478706
- Hahn JN, Falck VG, Jirik FR. Smad4 Deficiency in T Cells Leads to the Th17-Associated Development of Premalignant Gastroduodenal Lesions in Mice. *J Clin Invest* (2011) 121:4030–42. doi: 10.1172/JCI45114
- Kim BG, Malek E, Choi SH, Ignatz-Hoover JJ, Driscoll JJ. Novel Therapies Emerging in Oncology to Target the TGF-Beta Pathway. J Hematol Oncol (2021) 14:55. doi: 10.1186/s13045-021-01053-x
- Huang CY, Chung CL, Hu TH, Chen JJ, Liu PF, Chen CL. Recent Progress in TGF-Beta Inhibitors for Cancer Therapy. *BioMed Pharmacother* (2021) 134:111046. doi: 10.1016/j.biopha.2020.111046
- Kalluri R. The Biology and Function of Fibroblasts in Cancer. Nat Rev Cancer (2016) 16:582–98. doi: 10.1038/nrc.2016.73
- Yoshida GJ, Azuma A, Miura Y, Orimo A. Activated Fibroblast Program Orchestrates Tumor Initiation and Progression; Molecular Mechanisms and the Associated Therapeutic Strategies. *Int J Mol Sci* (2019) 20(9):2256. doi: 10.3390/ijms20092256
- Akhurst RJ, Derynck R. TGF-Beta Signaling in Cancer–a Double-Edged Sword. *Trends Cell Biol* (2001) 11:S44–51. doi: 10.1016/s0962-8924(01)02130-4
- 92. Boire A, Covic L, Agarwal A, Jacques S, Sherifi S, Kuliopulos A. PAR1 Is a Matrix Metalloprotease-1 Receptor That Promotes Invasion and Tumorigenesis of Breast Cancer Cells. *Cell* (2005) 120:303–13. doi: 10.1016/j.cell.2004.12.018
- 93. Orimo A, Gupta PB, Sgroi DC, Arenzana-Seisdedos F, Delaunay T, Naeem R, et al. Stromal Fibroblasts Present in Invasive Human Breast Carcinomas

Promote Tumor Growth and Angiogenesis Through Elevated SDF-1/ CXCL12 Secretion. Cell (2005) 121:335-48. doi: 10.1016/j.cell.2005.02.034

- Hessmann E, Patzak MS, Klein L, Chen N, Kari V, Ramu I, et al. Fibroblast Drug Scavenging Increases Intratumoural Gemcitabine Accumulation in Murine Pancreas Cancer. *Gut* (2018) 67:497–507. doi: 10.1136/gutjnl-2016-311954
- Su S, Chen J, Yao H, Liu J, Yu S, Lao L, et al. CD10(+)GPR77(+) Cancer-Associated Fibroblasts Promote Cancer Formation and Chemoresistance by Sustaining Cancer Stemness. *Cell* (2018) 172:841–56.e816. doi: 10.1016/ j.cell.2018.01.009
- Henry LR, Lee HO, Lee JS, Klein-Szanto A, Watts P, Ross EA, et al. Clinical Implications of Fibroblast Activation Protein in Patients With Colon Cancer. *Clin Cancer Res* (2007) 13:1736–41. doi: 10.1158/1078-0432.CCR-06-1746
- Tsujino T, Seshimo I, Yamamoto H, Ngan CY, Ezumi K, Takemasa I, et al. Stromal Myofibroblasts Predict Disease Recurrence for Colorectal Cancer. *Clin Cancer Res* (2007) 13:2082–90. doi: 10.1158/1078-0432.CCR-06-2191
- Dominguez CX, Muller S, Keerthivasan S, Koeppen H, Hung J, Gierke S, et al. Single-Cell RNA Sequencing Reveals Stromal Evolution Into LRRC15 (+) Myofibroblasts as a Determinant of Patient Response to Cancer Immunotherapy. *Cancer Discov* (2020) 10:232–53. doi: 10.1158/2159-8290.CD-19-0644
- 99. Mariathasan S, Turley SJ, Nickles D, Castiglioni A, Yuen K, Wang Y, et al. TGFbeta Attenuates Tumour Response to PD-L1 Blockade by Contributing to Exclusion of T Cells. *Nature* (2018) 554:544–8. doi: 10.1038/nature25501
- 100. Calon A, Lonardo E, Berenguer-Llergo A, Espinet E, Hernando-Momblona X, Iglesias M, et al. Stromal Gene Expression Defines Poor-Prognosis Subtypes in Colorectal Cancer. *Nat Genet* (2015) 47:320–9. doi: 10.1038/ng.3225
- 101. Isella C, Terrasi A, Bellomo SE, Petti C, Galatola G, Muratore A, et al. Stromal Contribution to the Colorectal Cancer Transcriptome. *Nat Genet* (2015) 47:312–9. doi: 10.1038/ng.3224
- 102. Grauel AL, Nguyen B, Ruddy D, Laszewski T, Schwartz S, Chang J, et al. TGF β -Blockade Uncovers Stromal Plasticity in Tumors by Revealing the Existence of a Subset of Interferon-Licensed Fibroblasts. *Nat Commun* (2020) 11:6315. doi: 10.1038/s41467-020-19920-5
- Penke LR, Peters-Golden M. Molecular Determinants of Mesenchymal Cell Activation in Fibroproliferative Diseases. *Cell Mol Life Sci* (2019) 76:4179– 201. doi: 10.1007/s00018-019-03212-3
- 104. Carthy JM, Sundqvist A, Heldin A, van Dam H, Kletsas D, Heldin CH, et al. Tamoxifen Inhibits TGF-Beta-Mediated Activation of Myofibroblasts by Blocking Non-Smad Signaling Through ERK1/2. J Cell Physiol (2015) 230:3084–92. doi: 10.1002/jcp.25049
- Higgins SP, Tang Y, Higgins CE, Mian B, Zhang W, Czekay RP, et al. TGF-Beta1/P53 Signaling in Renal Fibrogenesis. *Cell Signal* (2018) 43:1–10. doi: 10.1016/j.cellsig.2017.11.005
- 106. Szeto SG, Narimatsu M, Lu M, He X, Sidiqi AM, Tolosa MF, et al. YAP/TAZ Are Mechanoregulators of TGF-Beta-Smad Signaling and Renal Fibrogenesis. J Am Soc Nephrol (2016) 27:3117–28. doi: 10.1681/ASN.2015050499
- 107. Biffi G, Oni TE, Spielman B, Hao Y, Elyada E, Park Y, et al. IL1-Induced JAK/ STAT Signaling Is Antagonized by TGFbeta to Shape CAF Heterogeneity in Pancreatic Ductal Adenocarcinoma. *Cancer Discov* (2019) 9:282–301. doi: 10.1158/2159-8290.CD-18-0710
- Oft M, Akhurst RJ, Balmain A. Metastasis is Driven by Sequential Elevation of H-Ras and Smad2 Levels. *Nat Cell Biol* (2002) 4:487–94. doi: 10.1038/ ncb807
- 109. Ito Y, Sarkar P, Mi Q, Wu N, Bringas PJr., Liu Y, et al. Overexpression of Smad2 Reveals Its Concerted Action With Smad4 in Regulating TGF-Beta-Mediated Epidermal Homeostasis. *Dev Biol* (2001) 236:181–94. doi: 10.1006/ dbio.2001.0332
- 110. Zhang Y, Alexander PB, Wang XF. TGF-Beta Family Signaling in the Control of Cell Proliferation and Survival. *Cold Spring Harb Perspect Biol* (2017) 9(4):a022145. doi: 10.1101/cshperspect.a022145
- 111. Lamouille S, Xu J, Derynck R. Molecular Mechanisms of Epithelial-Mesenchymal Transition. Nat Rev Mol Cell Biol (2014) 15:178–96. doi: 10.1038/nrm3758
- 112. Miao Y, Yang H, Levorse J, Yuan S, Polak L, Sribour M, et al. Adaptive Immune Resistance Emerges From Tumor-Initiating Stem Cells. *Cell* (2019) 177:1172–86.e1114. doi: 10.1016/j.cell.2019.03.025

- 113. Castriconi R, Cantoni C, Della Chiesa M, Vitale M, Marcenaro E, Conte R, et al. Transforming Growth Factor Beta 1 Inhibits Expression of NKp30 and NKG2D Receptors: Consequences for the NK-Mediated Killing of Dendritic Cells. *Proc Natl Acad Sci USA* (2003) 100:4120–5. doi: 10.1073/ pnas.0730640100
- 114. Laouar Y, Sutterwala FS, Gorelik L, Flavell RA. Transforming Growth Factor-Beta Controls T Helper Type 1 Cell Development Through Regulation of Natural Killer Cell Interferon-Gamma. *Nat Immunol* (2005) 6:600–7. doi: 10.1038/ni1197
- 115. Gao Y, Souza-Fonseca-Guimaraes F, Bald T, Ng SS, Young A, Ngiow SF, et al. Tumor Immunoevasion by the Conversion of Effector NK Cells Into Type 1 Innate Lymphoid Cells. *Nat Immunol* (2017) 18:1004–15. doi: 10.1038/ni.3800
- 116. Cortez VS, Ulland TK, Cervantes-Barragan L, Bando JK, Robinette ML, Wang Q, et al. SMAD4 Impedes the Conversion of NK Cells Into ILC1-Like Cells by Curtailing non-Canonical TGF-Beta Signaling. *Nat Immunol* (2017) 18:995–1003. doi: 10.1038/ni.3809
- 117. Viel S, Marcais A, Guimaraes FS, Loftus R, Rabilloud J, Grau M, et al. TGF-Beta Inhibits the Activation and Functions of NK Cells by Repressing the mTOR Pathway. *Sci Signal* (2016) 9:ra19. doi: 10.1126/scisignal.aad1884
- Slattery K, Woods E, Zaiatz-Bittencourt V, Marks S, Chew S, Conroy M, et al. TGFbeta Drives NK Cell Metabolic Dysfunction in Human Metastatic Breast Cancer. J Immunother Cancer (2021) 9(2):e002044. doi: 10.1136/jitc-2020-002044
- Regis S, Dondero A, Caliendo F, Bottino C, Castriconi R. NK Cell Function Regulation by TGF-Beta-Induced Epigenetic Mechanisms. *Front Immunol* (2020) 11:311. doi: 10.3389/fimmu.2020.00311
- 120. Shaim H, Shanley M, Basar R, Daher M, Gumin J, Zamler DB, et al. Targeting the Alpha V Integrin/TGF-Beta Axis Improves Natural Killer Cell Function Against Glioblastoma Stem Cells. J Clin Invest (2021) 131(14): e142116. doi: 10.1172/jci142116
- 121. Strobl H, Knapp W. TGF-Beta1 Regulation of Dendritic Cells. Microbes Infect (1999) 1:1283–129. doi: 10.1016/s1286-4579(99)00256-7
- 122. Sere KM, Lin Q, Felker P, Rehage N, Klisch T, Ortseifer I, et al. Dendritic Cell Lineage Commitment is Instructed by Distinct Cytokine Signals. *Eur J Cell Biol* (2012) 91:515–23. doi: 10.1016/j.ejcb.2011.09.007
- 123. Felker P, Sere K, Lin Q, Becker C, Hristov M, Hieronymus T, et al. TGF-Beta1 Accelerates Dendritic Cell Differentiation From Common Dendritic Cell Progenitors and Directs Subset Specification Toward Conventional Dendritic Cells. J Immunol (2010) 185:5326–35. doi: 10.4049/ jimmunol.0903950
- 124. Sere K, Felker P, Hieronymus T, Zenke M. TGFbeta1 Microenvironment Determines Dendritic Cell Development. Oncoimmunology (2013) 2:e23083. doi: 10.4161/onci.23083
- 125. Lukas D, Yogev N, Kel JM, Regen T, Mufazalov IA, Tang Y, et al. TGF-Beta Inhibitor Smad7 Regulates Dendritic Cell-Induced Autoimmunity. *Proc Natl* Acad Sci USA (2017) 114:E1480–9. doi: 10.1073/pnas.1615065114
- 126. Hanks BA, Holtzhausen A, Evans KS, Jamieson R, Gimpel P, Campbell OM, et al. Type III TGF-Beta Receptor Downregulation Generates an Immunotolerant Tumor Microenvironment. J Clin Invest (2013) 123:3925–40. doi: 10.1172/JCI65745
- 127. Thepmalee C, Panya A, Junking M, Chieochansin T, Yenchitsomanus PT. Inhibition of IL-10 and TGF-Beta Receptors on Dendritic Cells Enhances Activation of Effector T-Cells to Kill Cholangiocarcinoma Cells. *Hum Vaccin Immunother* (2018) 14:1423–31. doi: 10.1080/21645515.2018.1431598
- Gunderson AJ, Yamazaki T, McCarty K, Fox N, Phillips M, Alice A, et al. TGFbeta Suppresses CD8(+) T Cell Expression of CXCR3 and Tumor Trafficking. *Nat Commun* (2020) 11:1749. doi: 10.1038/s41467-020-15404-8
- Flavell RA, Sanjabi S, Wrzesinski SH, Licona-Limon P. The Polarization of Immune Cells in the Tumour Environment by TGFbeta. *Nat Rev Immunol* (2010) 10:554–67. doi: 10.1038/nri2808
- 130. Thomas DA, Massague J. TGF-Beta Directly Targets Cytotoxic T Cell Functions During Tumor Evasion of Immune Surveillance. *Cancer Cell* (2005) 8:369–80. doi: 10.1016/j.ccr.2005.10.012
- 131. Arumugam V, Bluemn T, Wesley E, Schmidt AM, Kambayashi T, Malarkannan S, et al. TCR Signaling Intensity Controls CD8+ T Cell Responsiveness to TGF-Beta. J Leukoc Biol (2015) 98:703-12. doi: 10.1189/jlb.2HIMA1214-578R

- 132. Gorelik L, Flavell RA. Immune-Mediated Eradication of Tumors Through the Blockade of Transforming Growth Factor-Beta Signaling in T Cells. *Nat Med* (2001) 7:1118–22. doi: 10.1038/nm1001-1118
- 133. Park BV, Freeman ZT, Ghasemzadeh A, Chattergoon MA, Rutebemberwa A, Steigner J, et al. TGFbeta1-Mediated SMAD3 Enhances PD-1 Expression on Antigen-Specific T Cells in Cancer. *Cancer Discov* (2016) 6:1366–81. doi: 10.1158/2159-8290.CD-15-1347
- 134. Li S, Liu M, Do MH, Chou C, Stamatiades EG, Nixon BG, et al. Cancer Immunotherapy via Targeted TGF-Beta Signalling Blockade in TH Cells. Nature (2020) 587:121–5. doi: 10.1038/s41586-020-2850-3
- 135. Gorelik L, Constant S, Flavell RA. Mechanism of Transforming Growth Factor Beta-Induced Inhibition of T Helper Type 1 Differentiation. J Exp Med (2002) 195:1499–505. doi: 10.1084/jem.20012076
- 136. Lin JT, Martin SL, Xia L, Gorham JD. TGF-Beta 1 Uses Distinct Mechanisms to Inhibit IFN-Gamma Expression in CD4+ T Cells at Priming and at Recall: Differential Involvement of Stat4 and T-Bet. *J Immunol* (2005) 174:5950–8. doi: 10.4049/jimmunol.174.10.5950
- 137. Dimeloe S, Gubser P, Loeliger J, Frick C, Develioglu L, Fischer M, et al. Tumor-Derived TGF-Beta Inhibits Mitochondrial Respiration to Suppress IFN-Gamma Production by Human CD4(+) T Cells. *Sci Signal* (2019) 12 (599):eaav3334. doi: 10.1126/scisignal.aav3334
- 138. Marshall HD, Ray JP, Laidlaw BJ, Zhang N, Gawande D, Staron MM, et al. The Transforming Growth Factor Beta Signaling Pathway Is Critical for the Formation of CD4 T Follicular Helper Cells and Isotype-Switched Antibody Responses in the Lung Mucosa. *Elife* (2015) 4:e04851. doi: 10.7554/ eLife.04851
- 139. Wang Z, Yang Y, Cui Y, Wang C, Lai Z, Li Y, et al. Tumor-Associated Macrophages Regulate Gastric Cancer Cell Invasion and Metastasis Through TGFbeta2/NF-Kappab/Kindlin-2 Axis. *Chin J Cancer Res* (2020) 32:72–88. doi: 10.21147/j.issn.1000-9604.2020.01.09
- 140. Shen J, Li J, Wang B, Jin H, Wang M, Zhang Y, et al. Deletion of the Transforming Growth Factor Beta Receptor Type II Gene in Articular Chondrocytes Leads to a Progressive Osteoarthritis-Like Phenotype in Mice. Arthritis Rheum (2013) 65:3107–19. doi: 10.1002/art.38122
- 141. Choi J, Gyamfi J, Jang H, Koo JS. The Role of Tumor-Associated Macrophage in Breast Cancer Biology. *Histol Histopathol* (2018) 33:133–45. doi: 10.14670/HH-11-916
- 142. Arwert EN, Harney AS, Entenberg D, Wang Y, Sahai E, Pollard JW, et al. A Unidirectional Transition From Migratory to Perivascular Macrophage Is Required for Tumor Cell Intravasation. *Cell Rep* (2018) 23:1239–48. doi: 10.1016/j.celrep.2018.04.007
- 143. Lee YS, Park JS, Kim JH, Jung SM, Lee JY, Kim SJ, et al. Smad6-Specific Recruitment of Smurf E3 Ligases Mediates TGF-Beta1-Induced Degradation of MyD88 in TLR4 Signalling. *Nat Commun* (2011) 2:460. doi: 10.1038/ ncomms1469
- 144. Pang Y, Gara SK, Achyut BR, Li Z, Yan HH, Day CP, et al. TGF-Beta Signaling in Myeloid Cells Is Required for Tumor Metastasis. *Cancer Discov* (2013) 3:936–51. doi: 10.1158/2159-8290.CD-12-0527
- 145. Gentles AJ, Newman AM, Liu CL, Bratman SV, Feng W, Kim D, et al. The Prognostic Landscape of Genes and Infiltrating Immune Cells Across Human Cancers. *Nat Med* (2015) 21:938–45. doi: 10.1038/nm.3909
- Gabrilovich DI. Myeloid-Derived Suppressor Cells. Cancer Immunol Res (2017) 5:3–8. doi: 10.1158/2326-6066.CIR-16-0297
- 147. Liu Y, Lai L, Chen Q, Song Y, Xu S, Ma F, et al. MicroRNA-494 Is Required for the Accumulation and Functions of Tumor-Expanded Myeloid-Derived Suppressor Cells via Targeting of PTEN. J Immunol (2012) 188:5500–10. doi: 10.4049/jimmunol.1103505
- 148. Lee CR, Lee W, Cho SK, Park SG. Characterization of Multiple Cytokine Combinations and TGF-Beta on Differentiation and Functions of Myeloid-Derived Suppressor Cells. Int J Mol Sci (2018) 19(3):869. doi: 10.3390/ ijms19030869
- 149. Yan HH, Jiang J, Pang Y, Achyut BR, Lizardo M, Liang X, et al. CCL9 Induced by TGFbeta Signaling in Myeloid Cells Enhances Tumor Cell Survival in the Premetastatic Organ. *Cancer Res* (2015) 75:5283–98. doi: 10.1158/0008-5472.CAN-15-2282-T
- 150. Kitamura T, Kometani K, Hashida H, Matsunaga A, Miyoshi H, Hosogi H, et al. SMAD4-Deficient Intestinal Tumors Recruit CCR1+ Myeloid Cells That Promote Invasion. *Nat Genet* (2007) 39:467–75. doi: 10.1038/ng1997

- 151. Yang L, Huang J, Ren X, Gorska AE, Chytil A, Aakre M, et al. Abrogation of TGF Beta Signaling in Mammary Carcinomas Recruits Gr-1+CD11b+ Myeloid Cells That Promote Metastasis. *Cancer Cell* (2008) 13:23–35. doi: 10.1016/j.ccr.2007.12.004
- Derynck R, Turley SJ, Akhurst RJ. TGFbeta Biology in Cancer Progression and Immunotherapy. Nat Rev Clin Oncol (2021) 18:9–34. doi: 10.1038/ s41571-020-0403-1
- 153. Rouce RH, Shaim H, Sekine T, Weber G, Ballard B, Ku S, et al. The TGF-Beta/SMAD Pathway Is an Important Mechanism for NK Cell Immune Evasion in Childhood B-Acute Lymphoblastic Leukemia. *Leukemia* (2016) 30:800–11. doi: 10.1038/leu.2015.327
- 154. Shingai Y, Yokota T, Okuzaki D, Sudo T, Ishibashi T, Doi Y, et al. Autonomous TGF Beta Signaling Induces Phenotypic Variation in Human Acute Myeloid Leukemia. Stem Cells (2021) 39:723–36. doi: 10.1002/stem.3348
- 155. Wang B, Wang Y, Chen H, Yao S, Lai X, Qiu Y, et al. Inhibition of TGFbeta Improves Hematopoietic Stem Cell Niche and Ameliorates Cancer-Related Anemia. Stem Cell Res Ther (2021) 12(1):65. doi: 10.1186/s13287-020-02120-9
- 156. The Lancet O: CAR T-Cell Therapy for Solid Tumours. Lancet Oncol (2021) 22:893. doi: 10.1016/s1470-2045(21)00353-3
- 157. Noh KE, Lee JH, Choi SY, Jung NC, Nam JH, Oh JS, et al. TGF-Beta/IL-7 Chimeric Switch Receptor-Expressing CAR-T Cells Inhibit Recurrence of CD19-Positive B Cell Lymphoma. *Int J Mol Sci* (2021) 22(16):8706. doi: 10.3390/ijms22168706
- 158. Knudson KM, Hicks KC, Luo X, Chen JQ, Schlom J, Gameiro SR. M7824, a Novel Bifunctional Anti-PD-L1/TGFbeta Trap Fusion Protein, Promotes Anti-Tumor Efficacy as Monotherapy and in Combination With Vaccine. Oncoimmunology (2018) 7:e1426519. doi: 10.1080/2162402X.2018.1426519
- 159. Ravi R, Noonan KA, Pham V, Bedi R, Zhavoronkov A, Ozerov IV, et al. Bifunctional Immune Checkpoint-Targeted Antibody-Ligand Traps That Simultaneously Disable TGFbeta Enhance the Efficacy of Cancer Immunotherapy. Nat Commun (2018) 9:741. doi: 10.1038/s41467-017-02696-6
- 160. Martin CJ, Datta A, Littlefield C, Kalra A, Chapron C, Wawersik S, et al. Selective Inhibition of TGF Beta 1 Activation Overcomes Primary Resistance to Checkpoint Blockade Therapy by Altering Tumor Immune Landscape. Sci Trans Med (2020) 12(536):eaay8456. doi: 10.1126/scitranslmed.aay8456
- 161. Ciardiello D, Elez E, Tabernero J, Seoane J. Clinical Development of Therapies Targeting TGFbeta: Current Knowledge and Future Perspectives. Ann Oncol (2020) 31:1336–49. doi: 10.1016/j.annonc.2020.07.009
- 162. Holmgaard RB, Schaer DA, Li Y, Castaneda SP, Murphy MY, Xu X, et al. Targeting the TGFbeta Pathway With Galunisertib, a TGFbetaRI Small Molecule Inhibitor, Promotes Anti-Tumor Immunity Leading to Durable, Complete Responses, as Monotherapy and in Combination With Checkpoint Blockade. J Immunother Cancer (2018) 6(1):47. doi: 10.1186/ s40425-018-0356-4
- 163. Yap TA, Vieito M, Baldini C, Sepulveda-Sanchez JM, Kondo S, Simonelli M, et al. First-In-Human Phase I Study of a Next-Generation, Oral, TGFbeta Receptor 1 Inhibitor, LY3200882, in Patients With Advanced Cancer. *Clin Cancer Res* (2021) 27:6666–76. doi: 10.1158/1078-0432.CCR-21-1504
- 164. Melisi D, Oh DY, Hollebecque A, Calvo E, Varghese A, Borazanci E, et al. Safety and Activity of the TGFbeta Receptor I Kinase Inhibitor Galunisertib

Plus the Anti-PD-L1 Antibody Durvalumab in Metastatic Pancreatic Cancer. *J Immunother Cancer* (2021) 9(3):e002068. doi: 10.1136/jitc-2020-002068

- 165. Jung SY, Yug JS, Clarke JM, Bauer TM, Keedy VL, Hwang S, et al. Population Pharmacokinetics of Vactosertib, a New TGF-Beta Receptor Type Iota Inhibitor, in Patients With Advanced Solid Tumors. *Cancer Chemother Pharmacol* (2020) 85:173–83. doi: 10.1007/s00280-019-03979-z
- 166. Malek E, Hwang S, Caimi PF, Metheny LL, Tomlinson BK, Cooper BW, et al. Phase Ib Trial of Vactosertib in Combination With Pomalidomide in Relapsed Multiple Myeloma: A Corticosteroid-Free Approach by Targeting TGF-β Signaling Pathway. J Clin Oncol (2021) 39:8039–9. doi: 10.1200/ JCO.2021.39.15_suppl.8039
- 167. Dodagatta-Marri E, Meyer DS, Reeves MQ, Paniagua R, To MD, Binnewies M, et al. Alpha-PD-1 Therapy Elevates Treg/Th Balance and Increases Tumor Cell Psmad3 That Are Both Targeted by Alpha-TGFbeta Antibody to Promote Durable Rejection and Immunity in Squamous Cell Carcinomas. *J Immunother Cancer* (2019) 7:62. doi: 10.1186/s40425-018-0493-9
- 168. Zhong Z, Carroll KD, Policarpio D, Osborn C, Gregory M, Bassi R, et al. Anti-Transforming Growth Factor Beta Receptor II Antibody Has Therapeutic Efficacy Against Primary Tumor Growth and Metastasis Through Multieffects on Cancer, Stroma, and Immune Cells. *Clin Cancer Res* (2010) 16:1191–205. doi: 10.1158/1078-0432.CCR-09-1634
- 169. Tolcher AW, Berlin JD, Cosaert J, Kauh J, Chan E, Piha-Paul SA. Et Al: A Phase 1 Study of Anti-TGFbeta Receptor Type-II Monoclonal Antibody LY3022859 in Patients With Advanced Solid Tumors. *Cancer Chemother Pharmacol* (2017) 79:673–80. doi: 10.1007/s00280-017-3245-5
- 170. Khasraw M, Weller M, Lorente D, Kolibaba K, Lee CK, Gedye C, et al. Bintrafusp Alfa (M7824), a Bifunctional Fusion Protein Targeting TGF-Beta and PD-L1: Results From a Phase I Expansion Cohort in Patients With Recurrent Glioblastoma. *Neurooncol Adv* (2021) 3:vdab058. doi: 10.1093/ noajnl/vdab058
- 171. Yoo C, Oh DY, Choi HJ, Kudo M, Ueno M, Kondo S, et al. Phase I Study of Bintrafusp Alfa, a Bifunctional Fusion Protein Targeting TGF-Beta and PD-L1, in Patients With Pretreated Biliary Tract Cancer. J Immunother Cancer (2020) 8(1):e000564. doi: 10.1136/jitc-2020-000564

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Chen, Mu, Zhang, He and Liu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.