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Article The Recovery Benefit on Skin Blood Flow Using Vibrating Foam Rollers for Postexercise Muscle Fatigue in Runners

Yi-Horng Lai ¹, Ai-Yi Wang ², Chia-Chi Yang ³ and Lan-Yuen Guo ^{2,3,4,5,*}

- ¹ School of Mechanical and Electrical Engineering, Xiamen University Tan Kah Kee College, Zhangzhou 363105, China; lai81@xujc.com
- ² Department of Sports Medicine, College of Medicine, Kaohsiung Medical University, Kaohsiung 807, Taiwan; aiyi0703@gmail.com
- ³ The Master Program of Long-Term Care in Aging, College of Nursing, Kaohsiung Medical University, Kaohsiung 807, Taiwan; chiachiyang@kmu.edu.tw
- ⁴ Program in Biomedical Engineering, College of Medicine, Kaohsiung Medical University, Kaohsiung 807, Taiwan
- ⁵ Department of Medical Research, Kaohsiung Medical University Hospital, Kaohsiung 807, Taiwan
- * Correspondence: yuen@kmu.edu.tw; Tel.: +886-7-3121101 (ext. 2737/614)

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Abstract: Purpose: To determine the effect of vibrating rollers on skin blood flow after running for recovery from muscle fatigue. Method: 23 healthy runners, aged between 20 to 45 years, participated in a crossover trial. Muscle fatigue was induced by running, and recovery using a vibrating roller was determined before and after the intervention. Each subject was measured at three time points (prerun, postrun, and postroller) to compare skin blood flow perfusion and blood flow oscillation at the midpoint of the dominant gastrocnemius muscle. The results show that blood perfusion is greater when a vibrating roller is used than a foam roller, but there is no statistical difference. The analysis of blood flow oscillation shows that vibrating rollers induce 30% greater endothelial activation than a foam roller. Vibrating rollers significantly stimulate the characteristic frequency for myogenic activation (p < 0.05); however, the effect size is conservative.

Keywords: vibrating roller; recovery of muscle fatigue; skin blood flow; blood flow oscillation

1. Introduction

For prolonged exercise, leg muscles undergo repetitive contraction patterns over a long time. During long-term exercise, the main mechanism for muscle damage is the constant contraction and relaxation of muscle fibers, which damages muscle filaments, destroys the sarcoplasmic reticulum, increases lactic acid accumulation, and decreases ATP energy. This type of exercise pattern often leads to physical and mental fatigue [1,2].

Nervous system fatigue is defined in terms of central nervous system fatigue or peripheral nerve fatigue. The main mechanism involves interaction between the nervous system and muscle cells. Psychological stress and endocrine factors affect motor nerve fibers, muscle fibers, and muscle sensory receptors. The reaction in the motor cortex decreases the excitement of nerve impulses that are transmitted to the spine, and there are fewer potassium ions in nerve cells [3].

Manual massage after exercise can reduce muscle fatigue [4–6]. Physical stimulation of muscles by massage increases local blood circulation and removes metabolic waste. As manual massage is limited in terms of the execution angle for massage and efficiency, massage tools (e.g., massage balls, yoga columns, and foam rollers) have been developed. A foam roller (FR) can be used for self-myofascial

release, which is relatively convenient, effective, and affordable [7–11]. On the other hand, in [12], the effects of massage were rather small and only relevant in the short term. The performance recovery of massage remains partly unclear.

Self myofascial release uses "autogenic inhibition" [13]. During the rolling massage period, the pressure that is applied to the muscles stimulates the sensory receptors (i.e., Golgi's body) at the junction of the muscles and the muscle bonds. Golgi's body detects the variation in muscle tension, and the muscle spindle regulates the length of muscle fibers to relax the muscles. The protective mechanism of autogenic inhibition prevents injuries to the muscles. The experiment results for self myofascial release show that the VAS pain scale scores, muscle tension, and stiffness are reduced, and the range of joint motion, muscle elasticity, and softness of fascia is increased [14].

Vibration therapy stimulates the H reflex of muscles, recruits more motor units, and activates nerve receptors. Studies have confirmed that vibration stimulation reduces postexercise fatigue and muscle soreness [15,16]. In addition, the acute physiological reaction mechanism for vibration increases blood circulation in the local muscles [17]. Vibration stimulation may also improve jumping performance and agility [18].

Combining the benefits of foam rollers and vibration stimulation, vibrating rollers (VRs) are used for self-myofascia relaxation. Vibration stimulation stimulates Golgi's keys, inhibits muscle contraction, and promotes muscle relaxation. Studies involving VRs show that VRs also increase the flexibility of muscles and the angle of joint movement and reduce pain [19–27].

For microcirculatory, skin blood flow (SBF) provides tissues with vital oxygen and nutrients while removing waste products and distributing signaling molecules around the organism. Previous studies have shown that vibration stimulation increases muscle temperature and skin blood. In addition, SBF reduces delayed muscle soreness after exercise. This acute physiological reaction mechanism is involved in muscle recovery [28–33]. Furthermore, blood flow oscillation (BFO) reflects the current functional state of blood flow regulation systems. BFO analysis shows that characteristic frequencies reflect endothelial activity (0.08–0.02 Hz), sympathetic neurogenic activity (0.02–0.06 Hz), myogenic activity (0.6–2.3 Hz) [34–39].

At present, most studies involving FRs focus on deep fascia massage. Whether the use of VRs to stimulate local soft tissue can promote micro blood flow and accelerate muscle fatigue recovery is still unknown. In this study, we hypothesize that vibration stimulation can stimulate Golgi's bond, inhibit muscle contraction, and promote muscle relaxation. Therefore, VRs may be more able to relax the muscle fascia than FRs. In addition, VRs may have lower pain intensity than FRs when performing self fascia relaxation. This study uses laser Doppler flowmetry measurements to determine the effects of VRs on local SBF and compares the effects of FRs and VRs on BFO. The effectiveness of muscle recovery after fatigue due to exercise is also determined.

2. Materials and Methods

All experimental procedures were approved by the Institutional Review Board of Kaohsiung Medical University Chung-Ho Memorial Hospital (No. KMUHIRB-E(I)-20190022). The protocol for experiments complies with the relevant guidelines of the Declaration of Helsinki. Written informed consent was obtained from each voluntary participant.

2.1. Participants

A total of 23 healthy novice runners (11 women and 12 men) were recruited for this study. The criteria for subjects included being aged between 20 and 45 years old and running regularly from 1 to 3 times a week at a speed of at least 6 km/h for 5–10 km and/or 30–50 min each time. Exclusion criteria included pulmonary disease, acute infection, cardiovascular disease, musculoskeletal injuries within 6 months, leg fracture surgery, diseases with neurological symptoms, varicose veins, and lower limb pain. Table 1 summarizes the demographic data for participants.

Title 1	Male	Female
Age (years)	26.2 ± 5.2	26.6 ± 7.8
Height (m)	169.5 ± 3.9	156.8 ± 5.1

Table 1. Characteristics of the demographic data.

2.2. Examination Protocol

This study uses a crossover experiment. The flow diagram of the protocol is shown in Figure 1. The experimental group and the control group are the same group of subjects. The order of experimental intervention was decided randomly, so there are no individual differences between the two groups. The protocol includes a baseline, fatiguing exercise and recovery with intervention. Skin blood perfusion was measured before (prerun) and after (postrun) a 50-min fatigue-inducing run [2], whereby the participant begans to walk briskly on a treadmill (without a gradient) at a speed of 6 km/h. After warming up for 5 min, the subject accelerated at a rate of 1 km/h every 2 min until Borg's score reached 13 (somewhat hard). Then, the subject ran at a steady speed for 50 min until Borg's score reached 17 (very hard) or 90% of the maximum heart rate (i.e., HRmax = 200 - age). Lastly, the subject continued to run for another 2 min, before slowing and finishing the run.



Figure 1. The flow diagram of the protocol.

After completing postrun measurements, participants performed 6 min of VR or FR, which was randomized for recovery intervention and SBF measurement (postroller). The participants had a one-week interval to recover from muscle fatigue between crossover experiments.

For roller recovery treatment, participants were supported on the ground with their arms and pushed their bodies forward and backward from the knee fossa to the Achilles tendon for 3 s and then reversed from the Achilles tendon to the knee fossa. The entire calf gastrocnemius muscle and the medial and lateral muscles of the gastrocnemius muscles of both legs were massaged. Each leg was

massaged for 3 min (Figure 2). The visual analog scale (VAS) was used to evaluate the pain intensity of subjects during the intervention.



Figure 2. The procedure of roller treatment.

2.3. Examination Instrument and Outcome

2.3.1. Vibrating Roller Technique

The VR (FE07-VIBR50), designed by LAIN HONG SHING YEH CO, Taiwan, has a 5-stage preset vibration frequency (20, 25, 32, 40, and mixed frequency). The vibration frequency selected in this study is the mixed frequency mode (i.e., $20 \sim 40$ Hz: 20 Hz $\rightarrow 25$ Hz $\rightarrow 32$ Hz $\rightarrow 40$ Hz $\rightarrow 32$ Hz $\rightarrow 25$ Hz $\rightarrow 20$ Hz $\rightarrow 25$ Hz; automatic adjustment every 10 s) (Figure 3).



Figure 3. The vibrating roller.

2.3.2. Laser Doppler Flowmetry

SBF perfusion was measured using a laser Doppler flowmeter (Oxford Optronix Ltd., Abingdon, UK). The sampling frequency is 256 Hz. The measured signals are in relative units (blood perfusion units (BPU)). Each subject was subject to a 2-min SBF measurement before running, after running, and after using a VR or FR. The subjects were supine and kept a fixed posture during the measurements to reduce artificial interference. The measurement site was estimated visually at the midpoint of the calf gastrocnemius muscle of the dominant leg (Figure 4).

2.3.3. Blood Flow Perfusion and Oscillation Analysis

Using Matlab 2016 (MathWorks Inc., Natick, MA, USA), the signal noise was filtered and the SBF perfusion was calculated. Similar to previous studies [34–38], a Morlet wavelet transform was used to determine BFO. Five characteristic frequencies in the frequency interval between 0.0095 and 2 Hz were identified. The outcomes of BFO analysis reflect the activity of metabolic endothelial-related controls (0.008–0.02 Hz), sympathetic neurogenic systems (0.02–0.06 Hz), myogenic activity (0.06–0.2 Hz), respiratory movements (0.2–0.6 Hz), and cardiac movements (0.6–2.3 Hz).

Examples of SBF perfusion and BFO are shown in Figure 5. The frequency corresponding to the scale of the Morlet wavelet parameter ranged from 0.007 to 2.62 Hz over 50 intervals. The respective scale boundaries for the characteristic frequency were at 306, 1033, 3506, and 10530. Vibrating intervention affects only a local area of skin and causes no local heating. We anticipate that the interventions are local mechanisms of reactive hyperemia. Only three characteristic frequencies (endothelial, neurogenic, and myogenic) were analyzed.



Figure 4. The skin blood flow measurement.



Figure 5. An example of typical records of skin blood flow perfusion (**a**) and blood flow oscillation spectrum (**b**).

2.4. Statistical Analysis

Data are expressed as mean and standard deviation. All results were analyzed using SPSS 25 (SPSS Inc., Chicago, IL, USA) statistical software. A two-way repeated-measures ANOVA and a posthoc analysis were conducted to determine the effects of different conditions on dependent outcomes. Any significant interaction × time was identified and a posthoc analysis was used to determine the effect.

Furthermore, paired *t*-tests within each group were conducted to determine the significant effects of the intervention. In addition, the relative changes in normalized measures from prerun to postrun and from postrun to postroller were analyzed using a paired *t*-test. The significance difference (α) was set at 0.05.

3. Results

Using MATLAB and data analysis, the data for SBF perfusion and BFO at three time points (i.e., prerun, postrun, and postroller) are calculated.

3.1. Skin Blood Flow Perfusion

3.1.1. Two-Way Repeated-Measures ANOVA and Posthoc Analysis in Skin Blood Flow Perfusion

The results for SBF perfusion prerun, postrun, and postroller are shown in Table 2 and Figure 6. The ANOVA result is shown in Table 3. The posthoc analysis results are shown in Table 4.

Table 2. The experimental result of skin blood flow (SBF; A.U.) and normalized energy of blood flow oscillation (BFO; %).

		Prerun	Postrun	Postroller
SBF	Foam roller	35.1 (17.4)	55.6 (37.8)	40.7 (20.8)
	Vibrating roller	33.3 (16.8)	46.2 (23.0)	50.1 (41.6)
BFO	Foam roller	11.1 (6.0)	11.0 (5.3)	11.4 (5.4)
endothelial	Vibrating roller	10.5 (6.1)	10.6 (4.2)	12.5 (7.9)
BFO	Foam roller	7.6 (3.0)	6.5 (4.1)	7.0 (4.0)
neurogenic	Vibrating roller	7.0 (4.3)	5.6 (2.4)	6.3 (2.9)
BFO	Foam roller	15.9 (5.0)	16.0 (6.0)	18.0 (6.8)
myogenic	Vibrating roller	13.6 (5.8)	16.4 (5.1)	17.8 (6.9)

Values are mean(standard deviation).

Table 3.	Two-way	ANOVA	analysis of	f SBF and	normalized	energy c	of BFO.
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		F Value	df	р	η^2
	Roller	0.018	1	0.894	0.0001
SBF	Time	9.810	2	0.001 *	0.1294
	Roller \times Time	1.128	2	0.343	0.0168
PEO	Roller	0.163	1	0.690	0.0012
DFU an dath alial	Time	0.891	2	0.425	0.0133
endothelial	Roller \times Time	0.283	2	0.756	0.0043
BFO neurogenic	Roller	2.931	1	0.101	0.0217
	Time	1.338	2	0.284	0.0199
	Roller \times Time	0.031	2	0.970	0.0005
BFO myogenic	Roller	0.733	1	0.401	0.0055
	Time	4.439	2	0.025 *	0.0630
	Roller \times Time	1.298	2	0.294	0.0193

^{*} Significant difference (p < 0.05).

Table 4. Two-way ANOVA posthoc analysis of SBF (A.U.).

Roller	Section (I)	Section (J)	Difference (J-I)	р
Foam roller	Prerun	Postrun	20.536 (7.130)	0.026 *
	Prerun	Postroller	5.579 (3.762)	0.457
	Postrun	Postroller	-14.957 (7.981)	0.223
Vibrating roller	Prerun	Postrun	12.988 (5.306)	0.068
	Prerun	Postroller	16.903 (7.890)	0.130
	Postrun	Postroller	3.915 (8.680)	1

* Significant difference (p < 0.05).

After using VR, SBF perfusion was higher for the VR group than the FR group. There were larger individual differences in the SBF perfusion, so the difference between the two types of rollers was not significant. The ANOVA analysis shows a significant time effect (p < 0.001). The posthoc test results show a significant increase between prerun and postrun measurements for the FR group (Table 4).



Figure 6. The SBF trend between prerun, postrun, and post-roller.

3.1.2. Analysis of Relative Changes in Normalized Skin Blood Flow Perfusion

The paired *t*-tests for relative changes in normalized SBF perfusion for the two types of rollers are shown in Table 5. For the VR group, there is a significant effect of relative change in normalized perfusion between prerun and postroller measurements. However, the relative change in normalized perfusion between postrun and postroller measurements is not significant. For the FR group, the results in Tables 4 and 5 show a significant improvement between prerun and postrun measurements. As the intervention time increases, the relative change in normalized perfusion gradually decreases.

		Intervention	Pair Difference	df	p
SBF _	Prerun vs. Postroller	Foam roller Vibrating roller	27.90% (78.44) 79.23%(137.33)	22	0.04 *
	Postrun vs. Postroller	Foam roller Vibrating roller	-2.21% (97.52) 19.00%(91.59)	22	0.24
BFO endothelial [—]	Prerun vs. Postroller	Foam roller Vibrating roller	33.10% (79.17) 46.47% (102.10)	22	0.32
	Postrun vs. Postroller	Foam roller Vibrating roller	19.38% (60.35) 53.08% (138.63)	22	0.12
BFO neurogenic —	Prerun vs. Postroller	Foam roller Vibrating roller	-4.11% (58.59) -7.16% (50.73)	22	0.42
	Postrun vs. Postroller	Foam roller Vibrating roller	12.77% (55.25) 23.61% (67.43)	22	0.27
BFO myogenic —	Prerun vs. Postroller	Foam roller Vibrating roller	13.52% (34.34) 55.19% (99.48)	22	0.03 *
	Postrun vs. Postroller	Foam roller Vibrating roller	21.16% (72.93) 17.65% (45.58)	22	0.43

Table 5. The analysis of relative changes in normalized SBF and BFO (%).

* Significant difference (p < 0.05).

3.2. Blood Flow Oscillation

3.2.1. Two-Way Repeated-Measures ANOVA and Posthoc Analysis in Blood Flow Oscillation

The spectrum energy of BFO for prerun, postrun, and postroller is shown in Table 2. The trend plot of normalized energy in myogenic frequency band is shown in Figure 7. The ANOVA analysis

is shown in Table 3. In terms of endothelial and neurogenic energy, the ANOVA results are not significant for either the FR group or the VR group. However, for myogenic energy, the time effect has a significant effect.



Figure 7. The trend plot of normalized energy in myogenic frequency band.

3.2.2. The Analysis of Relative Changes in Normalized Blood Flow Oscillation

The results for a paired *t*-test for the relative change in normalized BFO for the two types of rollers are shown in Table 5.

For normalized endothelial and neurogenic energy, the paired *t*-test results are not significant for either the FR group or the VR group. However, between postrun and postroller sections, the VR group shows a greater relative change (33%) in normalized energy than the FR group.

In terms of normalized myogenic energy, there is a significant effect of relative change between prerun and postroller measurements for the VR group.

3.3. Visual Analog Scale (VAS)

The result of VAS for FRs was 6.2 (2.11); for VRs, it was 5.72 (2.49). The pair *t*-test result was not significant. In addition, taking the number of people as the unit, during the intervention, 11 (48%) subjects felt less pain when using VRs, 7 (30%) subjects felt less pain when using FRs, and 5 (22%) felt the same pain intensity.

4. Discussion

The results for skin blood flow perfusion (Table 2) show that SBF perfusion for the FR group decreases for the postroller measurement. There is decreased perfusion for the FR group because when FR involves insufficient compression, the effect of self-myofascia relaxation is not consistent. Excessive muscle tension inhibits blood supply and slows muscle recovery [7]. For this study, a larger number of subjects reported that when using VRs for self-myofascia relaxation massage, the pain intensity during muscle pressing was less than that of FRs. This result is different from that of a previous study, in which FR and VR groups exhibited a higher pain threshold than the control group [12,20]. Some studies have noted that VRs or FRs can increase the muscle pain threshold, but only VRs achieve statistically significant improvement benefits [26], which is also demonstrated by this study.

For the VR group, blood flow perfusion increases, but not significantly. SBF that is induced by vibration has been reported by many previous studies [17,28,29,31,32,40], but the parameters for vibration (frequency, device mode, intervention time) are not constant. One previous study showed that after 5 min intervention with 30 Hz vibration, SBF gradually decreased. Different frequencies of

vibration have a specific effect on SBF during the recovery period. A frequency of 50 Hz increases SBF, and a frequency of 30 Hz decreases SBF [17]. This study uses a 6-min protocol of VR intervention to determine the adaptability of participants, whereby the frequency of VR automatically changes every ten seconds (20 Hz \rightarrow 25 Hz \rightarrow 32 Hz \rightarrow 40 Hz \rightarrow 32 Hz \rightarrow 25 Hz \rightarrow 20 Hz \rightarrow 25 Hz). This may explain why a local vibration device has no significant effect on SBF.

In one study [40], passive intervention using vibration was shown to increase SBF, but there was no increase in blood flow in the femoral artery or in skeletal muscle. The previous findings cannot be applied to explain the hemodynamics of muscle fatigue recovery. In this condition, blood flows away from the integumentary system and is redirected to the musculature. Another vibration study concluded that nitric oxide (NO) synthase in endothelial cells increases SBF. However, NO is only one of the endogenous factors that induce skin vasodilation. Epinephrine, norepinephrine, and histamine are also activated by vibration to promote a vasodilatory response [41]. For muscle fatigue exercise, the dominant regulation mechanism for SBF changes from sympathetic activation to hypothalamus regulation. Skin vasodilation opens all capillaries in skeletal muscles, and skeletal muscle blood flow increases to 80% of cardiac output [30]. The complicated relationship between endothelial, neurogenic, and myogenic activity on skin vasodilation can be determined using BFO analysis.

The wavelet analysis results for BFO show no significant difference between postrun and postroller results. However, the relative energy for endothelial activation increases by nearly 33% for the VR group. During muscle exercise, the balance between oxygen supply, oxygen demand, and the control of blood flow is determined by metabolic factors [33]. The results of this study are in agreement with those of past studies [34,41] in that the characteristic frequency of endothelial activation is 0.008 to 0.02 Hz and promotes SBF perfusion.

Myogenic activity is associated with the rhythmic oscillations of capillaries, so the decrease in myogenic activity in wavelet power reflects a decrease in SBF [35]. This study determines that VRs significantly stimulate the characteristic frequency for myogenic activation. The myogenic response, which is most common in smaller resistance arteries, refers to the contraction that is caused by the muscle cell itself. It is used to regulate organ blood flow and peripheral resistance. When the blood vessels are in the precontraction state, myogenic activation can cause additional contraction or expansion, which increases or decreases blood flow [42].

All participants for this study were healthy novice runners, and the procedure of fatigue-inducing running is exhausting, so there were few participants. There were no comparisons with a control group (a group without roller intervention) for this study. Second, pulse rate was measured during the measurement of SBF, but the pulse rate and respiration of subjects during the 50-min fatigue running were not, so the characteristic frequencies for respiratory movements (0.2–0.6 Hz) and cardiac movements (0.6–2.3 Hz) in BFO cannot be verified. Moreover, because of the conservative effect size, the result of the current work is exploratory and not confirmative. For these reasons, if the reader wants to translate the result into practice, the present findings should be interpreted with caution.

5. Conclusions

In this study, there was no significant difference in the SBF perfusion trend between FR and VR intervention for muscle fatigue recovery. Using a variable frequency VR, the relative change in normalized perfusion was not significant. There was greater myogenic activation in BFO after VR intervention. However, due to the conservative effect size, further research is needed to elucidate whether the variable frequency VR for self-fascia relaxation massage can accelerate recovery from fatigue.

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References

- 1. Allen, D.G.; Lamb, G.D.; Westerblad, H. Skeletal muscle fatigue: Cellular mechanisms. *Physiol. Rev.* 2008, *88*, 287–332. [CrossRef]
- 2. Koblbauer, I.F.; Schooten, K.S.V.; Verhagen, E.A.; Dien, J.H.V. Kinematic changes during running-induced fatigue and relations with core endurance in novice runners. *J. Sci. Med. Sport* **2014**, *17*, 419–424. [CrossRef]
- 3. Gandevia, S.C. Spinal and supraspinal factors in human muscle fatigue. *Physiol. Rev.* **2001**, *81*, 1725–1789. [CrossRef]
- 4. Portillo-Soto, A.; Eberman, L.E.; Demchak, T.J.; Peebles, C. Comparison of blood flow changes with soft tissue mobilization and massage therapy. *J. Altern. Complement. Med.* **2014**, *20*, 932–936. [CrossRef] [PubMed]
- Albin, S.R.; Koppenhaver, S.; Bailey, B.; Blommel, H.; Fenter, B.; Lowrimore, C.; Smith, A.C.; McPoil, T.G. The effect of manual therapy on gastrocnemius muscle stiffness in healthy individuals. *Foot* 2019, *38*, 70–75. [CrossRef] [PubMed]
- Nunes, G.S.; Bender, P.U.; De Menezes, F.S.; Yamashitafuji, I.; Vargas, V.Z.; Wageck, B.B. Massage therapy decreases pain and perceived fatigue after long-distance Ironman triathlon: A randomised trial. *J. Physiother.* 2016, 62, 83–87. [CrossRef]
- Baumgart, C.; Freiwald, J.; Kühnemann, M.; Hotfiel, T.; Hüttel, M.; Hoppe, M.W. Foam rolling of the calf and anterior thigh: Biomechanical loads and acute effects on vertical jump height and muscle stiffness. *Sports* 2019, 7, 27. [CrossRef] [PubMed]
- 8. Behm, D.G.; Wilke, J. Do self-myofascial release devices release myofascia? Rolling mechanisms: A narrative review. *Sports Med.* **2019**, *49*, 1173–1181. [CrossRef]
- Freiwald, J.; Baumgart, C.; Kühnemann, M.; Hoppe, M.W. Foam-rolling in sport and therapy—Potential benefits and risks: Part 2—Positive and adverse effects on athletic performance. *Sports Orthop. Traumatol.* 2016, 32, 267–275. [CrossRef]
- Hendricks, S.; Hill, H.; Hollander, S.D.; Lombard, W.; Parker, R. Effects of foam rolling on performance and recovery: A systematic review of the literature to guide practitioners on the use of foam rolling. *J. Bodyw. Mov. Ther.* 2020, 24, 151–174. [CrossRef]
- Schroder, J.F.; Lueders, L.; Schmidt, M.; Braumann, K.-M.; Hollander, K. Foam rolling effects on soft tissue tone, elasticity and stiffness in the time course of recovery after weight training. *Sports Orthop. Traumatol.* 2018, 35, 171–177. [CrossRef]
- 12. Poppendieck, W.; Wegmann, M.; Ferrauti, A.; Kellmann, M.; Pfeiffer, M.; Meyer, T. Massage and performance recovery: A meta-analytical review. *Sports Med.* **2016**, *46*, 183–204. [CrossRef] [PubMed]
- 13. Beardsley, C.; Škarabot, J. Effects of self-myofascial release: A systematic review. *J. Bodyw. Mov. Ther.* **2015**, 19, 747–758. [CrossRef] [PubMed]
- 14. Torres, R.; Ribeiro, F.; Duarte, J.A.; Cabri, J.M. Evidence of the physiotherapeutic interventions used currently after exercise-induced muscle damage: Systematic review and meta-analysis. *Phys. Ther. Sport* **2012**, *13*, 101–114. [CrossRef] [PubMed]
- 15. Fagnani, F.; Giombini, A.; Di Cesare, A.; Pigozzi, F.; Di Salvo, V. The effects of a whole-body vibration program on muscle performance and flexibility in female athletes. *Am. J. Phys. Med. Rehabil.* **2006**, *85*, 956–962. [CrossRef]
- 16. Imtiyaz, S.; Veqar, Z.; Shareef, M.Y. To compare the effect of vibration therapy and massage in prevention of Delayed Onset Muscle Soreness (DOMS). *J. Clin. Diagn. Res.* **2014**, *8*, 133–136. [CrossRef]
- 17. Maloney-Hinds, C.; Petrofsky, J.S.; Zimmerman, G. The effect of 30 Hz vs. 50 Hz passive vibration and duration of vibration on skin blood flow in the arm. *Med. Sci. Monit.* **2008**, *14*, 112–116.
- 18. Kurt, C. Alternative to traditional stretching methods for flexibility enhancement in well-trained combat athletes: Local vibration versus whole-body vibration. *Biol. Sport* **2015**, *32*, 225–233. [CrossRef]
- García-Gutiérrez, M.T.; Guillén-Rogel, P.; Cochrane, D.J.; Marín, P.J. Cross transfer acute effects of foam rolling with vibration on ankle dorsiflexion range of motion. *J. Musculoskelet. Neuronal Interact.* 2018, 18, 262–267.

- 20. Cheatham, S.W.; Stull, K.R.; Kolber, M.J. Comparison of a vibration roller and a nonvibration roller intervention on knee range of motion and pressure pain threshold: A randomized controlled trial. *J. Sport Rehabil.* **2019**, *28*, 39–45. [CrossRef]
- 21. Han, S.-W.; Lee, Y.-S.; Lee, N.-J. The influence of the vibration form roller exercise on the pains in the muscles around the hip joint and the joint performance. *J. Phys. Ther. Sci.* **2017**, *29*, 1844–1847. [CrossRef] [PubMed]
- 22. Lee, C.-J.; Chu, I.-H.; Lyu, B.-J.; Chang, W.-D.; Chang, N.-J. Comparison of vibration rolling, nonvibration rolling, and static stretching as a warm-up exercise on flexibility, joint proprioception, muscle strength, and balance in young adults. *J. Sports Sci.* **2018**, *36*, 2575–2582. [CrossRef] [PubMed]
- 23. Lim, J.-H.; Park, C.-B. The immediate effects of foam roller with vibration on hamstring flexibility and jump performance in healthy adults. *J. Exerc. Rehabil.* **2019**, *15*, 50–54. [CrossRef]
- 24. Lim, J.-H.; Park, C.-B.; Kim, B.-G. The effects of vibration foam roller applied to hamstring on the quadriceps electromyography activity and hamstring flexibility. *J. Exerc. Rehabil.* **2019**, *15*, 560–565. [CrossRef]
- 25. Lin, W.-C.; Lee, C.-L.; Chang, N.-J. Acute effects of dynamic stretching followed by vibration foam rolling on sports performance of badminton athletes. *J. Sports Sci. Med.* **2020**, *19*, 420–428. [PubMed]
- Romero-Moraleda, B.; González-García, J.; Cuéllar-Rayo, Á.; Balsalobre-Fernández, C.; Muñoz-García, D.; Morencos, E. Effects of vibration and non-vibration foam rolling on recovery after exercise with induced muscle damage. *J. Sports Sci. Med.* 2019, *18*, 172–180. [PubMed]
- 27. De Benito, A.M.; Valldecabres, R.; Ceca, D.; Richards, J.D.; Barrachina-Igual, J.; Pablos, A. Effect of vibration vs non-vibration foam rolling techniques on flexibility, dynamic balance and perceived joint stability after fatigue. *PeerJ* **2019**, *7*, e8000. [CrossRef] [PubMed]
- Fuller, J.T.; Thomson, R.L.; Howe, P.R.C.; Buckley, J.D. Effect of vibration on muscle perfusion: A systematic review. *Clin. Physiol. Funct. Imaging* 2012, 33, 1–10. [CrossRef]
- Menéndez, H.; Martín-Hernández, J.; Ferrero, C.; Figueroa, A.; Herrero, A.J.; Marín, P.J. Influence of isolated or simultaneous application of electromyostimulation and vibration on leg blood flow. *Eur. J. Appl. Physiol.* 2015, *115*, 1747–1755. [CrossRef]
- Lohman, E.B.; Bains, G.S.; Lohman, T.; DeLeon, M.; Petrofsky, J.S. A comparison of the effect of a variety of thermal and vibratory modalities on skin temperature and blood flow in healthy volunteers. *Med. Sci. Monit.* 2011, 17, 72–81. [CrossRef]
- 31. Lohman, E.; Sackiriyas, K.S.B.; Bains, G.S.; Calandra, G.; Lobo, C.; Nakhro, D.; Malthankar, G.; Paul, S. A comparison of whole body vibration and moist heat on lower extremity skin temperature and skin blood flow in healthy older individuals. *Med. Sci. Monit.* **2012**, *18*, 415–424. [CrossRef] [PubMed]
- 32. Tzen, Y.-T.; Weinheimer-Haus, E.M.; Corbiere, T.F.; Koh, T.J. Increased skin blood flow during low intensity vibration in human participants: Analysis of control mechanisms using short-time Fourier transform. *PLoS ONE* **2018**, *13*, e0200247. [CrossRef] [PubMed]
- 33. Zange, J.; Molitor, S.; Illbruck, A.; Müller, K.; Schönau, E.; Kohl-Bareis, M.; Rittweger, J. In the unloaded lower leg, vibration extrudes venous blood out of the calf muscles probably by direct acceleration and without arterial vasodilation. *Graefe's Arch. Clin. Exp. Ophthalmol.* **2014**, *114*, 1005–1012. [CrossRef] [PubMed]
- 34. Cui, R.F.; Luo, S.T.; Lu, L.Q.; Zhou, W.W.; Li, Z.Y. A method for extracting characteristic frequency components of blood flow signals based on wavelet transform. *Appl. Mech. Mater.* **2013**, *313*, 1221–1224. [CrossRef]
- 35. Hodges, G.J.; Mallette, M.M.; Martin, Z.T.; Del Pozzi, A.T. Effect of sympathetic nerve blockade on low-frequency oscillations of forearm and leg skin blood flow in healthy humans. *Microcirculation* **2017**. [CrossRef]
- 36. Grinevich, A.A.; Tankanag, A.; Tikhonova, I.; Chemeris, N. A new approach to the analysis of skin blood flow oscillations in human. *Microvasc. Res.* **2019**, *126*, 103889–103896. [CrossRef]
- 37. Mizeva, I.A.; Vetrova, D.V. Pulsations of cutaneous blood flow during local heating. *Russ. J. Biomech.* **2014**, *18*, 447–454.
- Jan, Y.-K.; Liao, F.; Rice, L.A.; Woods, J.A. Using reactive hyperemia to assess the efficacy of local cooling on reducing sacral skin ischemia under surface pressure in people with spinal cord injury: A preliminary report. *Arch. Phys. Med. Rehabil.* 2013, 94, 1982–1989. [CrossRef]
- 39. Harkikh, E.; Mizeva, I.; Makovik, I.; Dremin, V.; Zherebtsov, E.; Potapova, E.; Dunaev, A. Blood flow oscillations as a signature of microvascular abnormalities. *Proc. SPIE* **2018**, *10685*, 1–7.
- 40. Lohman, E.; Petrofsky, J.S.; Maloney-Hinds, C.; Betts-Schwab, H.; Thorpe, D. The effect of whole body vibration on lower extremity skin blood flow in normal subjects. *Med. Sci. Monit.* **2007**, *13*, 71–76.

- 41. Mitchell, U.H.; Johnson, P.K. Vibration and skin blood flow changes in subjects with restless legs syndrome. *J. Parkinsonism Restless Legs Syndr.* **2014**, *4*, 9–16. [CrossRef]
- 42. Jan, Y.-K.; Struck, B.D.; Foreman, R.D.; Robinson, C. Wavelet analysis of sacral skin blood flow oscillations to assess soft tissue viability in older adults. *Microvasc. Res.* **2009**, *78*, 162–168. [CrossRef] [PubMed]

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