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Identification of SARS-CoV-2 in post-mortem nasopharyngeal swabs



Dear Editor, we would like to share the ideas on “Forensic evaluation of two nucleic acid extraction systems and validation of a RT-qPCR protocol for identification of SARS-CoV-2 in post-mortem nasopharyngeal swabs [1].” Barrio et al. concluded that *“Both the automated and the manual RNA extraction procedures showed good efficiency, but the automated virus extraction by bio-robot produced more reproducible results than the manual extraction [1].”* Generally, a robotic RNA purification protocol usually has a better threshold for detection of RNA of virus [2]. However, there are still many considerations on automated extraction. In general, the automatic extraction has to be based on commercial kit. The commercial RNA extraction kits may be insufficient when there is a large demand during outbreak [3]. Additionally, comparing to manual approach, there are problems of in-valid rate and problem of negative agreement of automatic approach [4]. Barrio et al. mentioned for cross reactivity study but there is no specific study on cross reactivity to other viruses [1]. In a complete assessment of an automated extraction approach, it is necessary to perform a specific assessment on many respiratory viruses. In a recent report, Li et al. reported a similar evaluation with a complete cross reactivity analysis was performed in 20 other respiratory viruses [5]. Additionally, the reproducibility is also associated with stability. The effect of transport media on stability of virus should be mentioned [6]. Using of a lysis buffer supplemented with nucleic acid stabilization mix can also help extend stability [7]. To conclude on the stability property of new automated extraction should recognize the effect of transportation and specimen preparation.

Author statement

Both authors have equal contribution is idea giving, drafting, writing, revising and approving for submission.

Conflict of interest

None.

References

- [1] P.A. Barrio, A. Fernández-Rodríguez, P. Martín, C. Fernández, L. Fernández, A. Alonso, Forensic evaluation of two nucleic acid extraction systems and validation of a RT-qPCR protocol for identification of SARS-CoV-2 in post-mortem nasopharyngeal swabs, *Forensic Sci. Int.* 323 (2021) 110775, <https://doi.org/10.1016/j.forsciint.2021.110775> (Online ahead of print).
- [2] S. Perelle, L. Cavellini, C. Burger, S.B. Boisseau, C.H. Collette, H. Merle, P. Fach, Use of a robotic RNA purification protocol based on the NucliSens easyMAG for real-time RT-PCR detection of hepatitis A virus in bottled water, *J. Virol. Methods* 157 (1) (2009) 80–83.
- [3] S. Klein, T.G. Müller, D. Khalid, V. Sonntag-Buck, A.M. Heuser, B. Glass, M. Meurer, I. Morales, A. Schillak, A. Freistaedter, I. Ambiel, S.L. Winter, L. Zimmermann, T. Naumoska, F. Bubeck, D. Kirrmair, S. Ullrich, I. Barreto Miranda, S. Anders, D. Grimm, P. Schnitzler, M. Knap, H.G. Kräusslich, V.L. Dao Thi, K. Börner, P. Chlanda, SARS-CoV-2 RNA extraction using magnetic beads for rapid large-scale testing by RT-qPCR and RT-LAMP, *Viruses* 12 (8) (2020) 863.
- [4] D. Nörz, N. Fischer, A. Schultz, S. Kluge, U.M. Runge, M. Aepfelbacher, S. Pfefferle, M. Lütgehetmann, Clinical evaluation of a SARS-CoV-2 RT-PCR assay on a fully automated system for rapid on-demand testing in the hospital setting, *J. Clin. Virol.* 128 (2020) 104390.
- [5] Y. Li, J. Li, Y. Zhang, L. Dai, L. Li, J. Liu, S. Zhang, X. Wu, Y. Hu, C. Qin, T. Jiang, X. Kang, Development of an automatic integrated gene detection system for novel severe acute respiratory syndrome-related coronavirus (SARS-CoV2), *Emerg. Microbes Infect.* 9 (1) (2020) 1489–1496.
- [6] Y. Penrod, D. e Garcia, S.T. Dunn, Evaluation of transport media for laboratory detection of SARS-CoV-2 in upper respiratory tract swab specimens, *J. Med Virol.* 93 (5) (2021) 2774–2781.
- [7] O. Erster, O. Shkedi, G. Benedek, E. Zilber, I. Varkovitzky, R. Shirazi, D.O. Shorka, Y. Cohen, T. Bar, R. Yechiel, M.T. Oikawa, D. Venkert, M. Linial, E.O. Djian, M. Mandelboim, Z. Livneh, G.S. Saltzman, E. Mendelson, D. Wolf, M.S. Cohen, O. Mor, Y. Lewis, D. Zeevi, Improved sensitivity, safety, and rapidity of COVID-19 tests by replacing viral storage solution with lysis buffer, *PLoS One* 16 (3) (2021) e0249149.

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