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The effects of chronic AMPK activation on hepatic triglyceride accumulation and glycerol 3-phosphate acyltransferase activity with high fat feeding

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Abstract

Background: High fat feeding increases hepatic fat accumulation and is associated with hepatic insulin resistance. AMP Activated Protein Kinase (AMPK) is thought to inhibit lipid synthesis by the acute inhibition of glycerol-3-phosphate acyltransferase (GPAT) activity and transcriptional regulation via sterol regulatory element binding protein-1c (SREBP-1c).

Methods: The purpose of this study was to determine if chronic activation of AMPK prevented an increase in GPAT1 activity in rats fed a high fat diet. Rats were fed a control (C), or a high fat (HF) diet (60% fat) for 6 weeks and injected with saline or a daily aminoimidazole carboxamide ribnucleotide (AICAR) dose of 0.5 mg/g body weight.

Results: Chronic AMPK activation by AICAR injections resulted in a significant reduction in hepatic triglyceride accumulation in both the C and HF fed animals (C, 5.5±0.7; C+AICAR, 2.7 ±0.3; HF, 21.8±3.3; and HF+AICAR, 8.0±1.8 mg/g liver). HF feeding caused an increase in total GPAT and GPAT1 activity, which was not affected by chronic AMPK activation (GPAT1 activity vs. C, C+AICAR, 92±19%; HF, 186±43%; HF+AICAR, 234±62%). Markers of oxidative capacity, including citrate synthase activity and cytochrome c abundance, were not affected by chronic AICAR treatment. Interestingly, HF feeding caused a significant increase in long chain acyl-CoA dehydrogenase or LCAD (up 66% from C), a marker of fatty acid oxidation capacity.

Conclusions: These results suggest that chronic AMPK activation limits hepatic triglyceride accumulation independent of a reduction in total GPAT1 activity.

Keywords: AMPK, GPAT1, SREBP-1c, mTOR, LCAD

Background

AMP-activated protein kinase (AMPK) is a major regulator of energy homeostasis and nutrient metabolism. AMPK is known to regulate fatty acid metabolism, protein synthesis, and glucose uptake [1,2]. Furthermore, the activation of AMPK occurs by allosteric and covalent modification of the enzyme in response to an energy deficit [3]. AMPK exerts its effects on energy metabolism by acutely

regulating enzyme activity and protein abundance as well as influencing transcription and translation of genes involved in energy metabolism [4-6]. For these reasons, AMPK is of tremendous interest in understanding the mechanisms involved in hepatic lipid accumulation.

Hepatic lipid accumulation occurs in conditions of elevated dietary fat, obesity, and decreased metabolic function associated with decreased liver function. There are a number of mechanisms that could lead to increased hepatic lipid accumulation. Simply put, hepatic lipid accumulation is the result of a greater amount of lipid uptake and/or synthesis relative to lipid oxidation and release into the circulation [7,8]. Non-alcoholic fatty liver

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disease (NAFLD) is defined as hepatic fat accumulation greater than five percent of liver weight in the absence of excessive alcoholic intake [9]. Approximately 10-30 percent of the adult population in the United States is thought to have NAFLD, making it the most common chronic liver condition among adults [10-17]. It has also extended to adolescents with one study reporting approximately 61 percent of adolescent subjects with elevated liver enzymes (a marker of NAFLD) being overweight or obese, [18]. NAFLD is strongly associated with insulin resistance and is the hepatic representation of metabolic syndrome [9,11,19,20]. If not corrected, NAFLD can lead to the development of non-alcoholic steatohepatitis (NASH), cirrhosis of the liver and hepatocellular carcinoma [21].

Consistent with AMPK's demonstrated role in energy metabolism, AMPK has been reported to increase lipid oxidation and inhibit lipid synthesis. One proposed mechanism for AMPK induced lipid regulation is in the acute inhibition of glycerol-3-phosphate acyltransferase (GPAT), an integral enzyme in triglyceride accumulation. GPAT is the rate-limiting enzyme catalyzing the first committed step in triglyceride synthesis [22,23]. Of the four predominant isoforms of GPAT, three are inhibited by *N*-Ethylmaleimide (NEM). In contrast, the isoform GPAT1 which is localized to the outer membrane of the mitochondria is resistant to NEM and accounts for 10 percent of the total GPAT activity in extra-hepatic tissues. In the liver, GPAT1 accounts for 30 to 50 percent of the total GPAT activity, making it a significant contributor to hepatic triglyceride regulation [24-26]. Chemical activation of AMPK by an AMP-analog aminoimidazole carboxamide ribonucleotide (AICAR) reduces fat accumulation in the hepatocyte by decreasing GPAT1 activity by 30 to 40 percent [5,23,26]. It is also likely that AMPK limits the fatty acid availability for triglyceride synthesis by increasing fat oxidation rates. AMPK inhibits acetyl-CoA carboxylase (ACC), an enzyme that catalyzes the formation of malonyl-CoA. Malonyl-CoA inhibits carnitine palmitoyltransferase I (CPT1) resulting in decreased beta-oxidation and increased fat synthesis. Decreasing malonyl-CoA production results in an increase in CPT1 activity [5,27-29]. Therefore, through AMPK's acute role of inhibiting GPAT1 and increasing CPT1, there is an overall increase in oxidation relative to triglyceride synthesis.

In addition to acute regulation of triglyceride synthesis enzymes, recent evidence points to a role in which AMPK influences the transcription and translation of lipid synthesis enzymes [4,30]. Sterol regulatory element binding protein-1c (SREBP-1c) increases the transcription of lipid synthesis enzymes such as ACC, fatty acid synthase (FAS), GPAT, and stearoyl-CoA desaturase (SCD1) [13,31-34]. Previous work suggests that activation of AMPK decreases promoter activity of SREBP-1c

in the liver cells, as well as decreasing the transcriptional activity of liver X receptors (LCRa), an upstream transcription factor and regulator of SREBP-1c expression, thus decreasing SREBP-1c and LXR [4]. Further, AMPK decreases SREBP-1c activity by interfering with the mammalian target of rapamycin complex (mTOR) activity. The proposed mechanism for mTOR-dependent activation of SREBP-1c is thought to be by cleavage of the SREBP-1c molecule [35]. Other studies done using cell culture models have shown that AMPK activation can inhibit mitochondrial GPAT1 abundance by decreasing SREBP-1c activity [4,36]. Thus, in addition to AMPK's role as an acute regulator, AMPK may further inhibit hepatic lipid accumulation by inhibiting SREBP-1c through transcriptional regulation reduction of mTOR activity.

While there is some evidence for AMPK dependent inhibition of lipogenic enzymes [30,37], it is not completely understood how AMPK activation mediates this effect in liver tissue. Furthermore, a greater understanding of the role of AMPK activation in the process of hepatic lipid accumulation is becoming increasingly important due to the prevalence of NAFLD and NASH as noted above. AMPK may be a valuable therapeutic target for the treatment of these conditions. Therefore, the purpose for this study was to examine the effects of chronic activation of AMPK on enzymes critical for hepatic triglyceride accumulation and lipid synthesis, specifically ACC and GPAT1. This study was designed to gain a greater understanding of the role of chronic activation of AMPK on hepatic triglyceride synthesis and accumulation.

Materials and methods

Animal care

All procedures related to animal care and use were approved by the Institutional Animal Care and Use Committee of Brigham Young University.

Diet

Male Wistar rats approximately 25 days old were divided into 4 groups of 7-10 animals each. Two groups consumed laboratory chow diet, 5053 PicoLab® Rodent Diet 20, and two groups consumed the high fat diet [38-40]. Food and water were provided *ad libitum*.

High Fat diet (g/kg of food):

116.3 g olive oil, 232.7 g flax seed oil, 87.2 g sugar, 174.6 g starch, 226.6 g casein, 4.5 g methionine, 30.7 g gelatin, 51.2 g bran, 22.5 g vitamin mix (Harlan Teklad, AIN76A), 52.2 g mineral mix (Harlan Teklad, AIN76), 1.4 g choline chloride. See Table 1 for the macronutrient composition of the two diets used in these studies.

Table 1 Macronutrient composition of diets

	Chow diet (% of calories)	High fat diet (% of calories)
Protein	23.6	20
Carbohydrate	64.5	20
Fat	12	60

Experimental design

A two-factor research design was used to examine the effects of high fat feeding as well as AMPK activation on factors important for the determination of fat accumulation in liver tissue. See Table 2 for a complete description of control and treatment groups.

AICAR injections

AICAR was dissolved in 0.9% NaCl and administered subcutaneously every morning for six weeks at a dose of 0.5 mg AICAR/g body weight. Rats were sacrificed 24 hours after the last injection was given. Throughout the six weeks, rats from Control and High Fat groups were handled daily and injected with a comparable amount of saline at the time the AICAR groups received AICAR injections.

Dissections

Rats were anesthetized with pentobarbital sodium (65 mg/kg body weight.) Liver tissue was extracted once rats were completely sedated. Liver was quickly removed and clamp frozen with liquid nitrogen chilled metal tongs then wrapped in aluminum foil and stored at -90 degrees Celsius.

Citrate synthase activity

As a measure of mitochondrial oxidative capacity, whole tissue homogenate was used to measure citrate synthase activity by the method of Srere [41]. Briefly, homogenates were prepared in 175 mM KCl, 10 mM GSH, 2 mM Ethylenediaminetetraacetic acid (EDTA), pH 7.4 (100 mg of tissue per ml.) Homogenates were further diluted in a solution of 100 mM Tris pH 8.0. Citrate synthase activity was measured and recorded for each sample.

Measurement of protein/enzyme abundance and phosphorylation

Standard bicinchoninic acid (BCA) protein assays were performed to determine protein concentration in the

Table 2 Experimental design

Chow groups	High fat groups
Chow + Daily Saline Injections	High Fat (HF) + Daily Saline Injections
Chow + Daily AICAR Injections	High Fat (HF) + Daily AICAR Injections

whole tissue or membrane fraction homogenate. The results were used to normalize protein content for Western Blots and activity assays.

Standard Western blotting procedures were performed. Briefly, aliquots of the liver homogenates were subjected to sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to a nitrocellulose membrane. The membrane was blocked with 5% milk solution followed by incubation with the primary antibody for the protein of choice overnight at 4 degrees Celsius. The next day, the membranes were rinsed with Tris-Buffered Saline Tween-20 (TBST) and incubated with the appropriate secondary antibody dissolved in 1% milk in TBST for 1 hour. This was followed again by rinses in TBST followed by one rinse in TBS. Protein content was detected by chemiluminescence. Protein band densitometry was quantified and analyzed with Image J and Alpha Ease software [4].

Primary antibodies for the following proteins were used: Total AMPK (Cell Signaling; cat. No. 2532 L), phospho-AMPK (Cell Signaling, Beverly, MA; cat. No. 2535 L), total ACC (GE Healthcare; cat. No. RPN1231V), SREBP-1c (Santa Cruz, Santa Cruz, CA; cat. No. SC13551), mTOR (Cell Signaling Technology Beverly, MA; cat. No. 2972).

In addition, we looked at 4E-binding protein (4E-BP) (Cell Signaling Technology Beverly, MA; cat no. 9452) and phosphor-raptor (Cell Signaling Technology Beverly, MA; cat no. 2083) for indications of mTOR activity and Cytochrome C (Santa Cruz, Santa Cruz, CA; cat. No. sc-13156) and LCAD (a gift from Daniel P. Kelly) for an indication of oxidative capacity.

Triglyceride assay

Liver triglycerides were measured according to the Folch method. Forty to fifty milligrams of frozen liver were homogenized with a 2:1 concentration of chloroform: methanol and agitated in a cold room overnight at 4 degrees C. One ml of 0.9% NaCl was added to each sample solution, vortexed and centrifuged for 1 hour at 1000 × g at 4 degrees C. The organic phase was removed and dried down in a lyophilizer. Samples were reconstituted with 100 µl of Tert-Butanol Triton X solution. Triglyceride content was then analyzed in each sample with the WAKO triglyceride assay kit as per manufacturer's instructions [42].

GPAT activity assay

The activity of microsomal and mitochondrial GPAT was measured using the method described previously [5,43,44]. Briefly, membrane fractions of liver tissue were formed using sample homogenates prepared with 25 mg of liver and homogenization buffer (250 mM Sucrose, 10 mM TrisHCl, 1 mM dithiothreitol (DTT), 1 mM EDTA, pH 7.4). Homogenate was centrifuged at 100,000 x g for

1 hour, after which the supernatant was removed and the pellet reconstituted with 400 μ l of the same homogenization buffer, aliquoted and frozen in -90 degree Celsius freezer. Total GPAT activity was measured using a cocktail of 800 μ M [14 C]glycerol 3-phosphate (G-3-P), 60 μ M palmitoyl Co-A, 75 mM TrisHCl, 4 mM MgCl, 2 mg/ml BSA, 8 mM NaF, 1 Mm DTT, and non-labeled G3P. Sample (40 μ g protein) was added to the solution incubated with and without 2 mM N-ethylmaleimide (NEM), an inhibitor of microsomal GPAT to isolate and measure the remaining activity, GPAT1 activity. The reaction was run for 10 min at 37°C and stopped with 0.6 ml 1% HClO₄ and chloroform-methanol (2:1). After 5 minutes on ice, another 1 ml of 1% perchloric acid and 1 ml of chloroform was added to the solution. Samples were centrifuged for 1 hour at 1000 \times g and washed 3 times with 1% perchloric acid after which 1 ml of organic phase containing the labeled G-3-P incorporated into lysophosphatidic acid was dried down using a lyophilizer. After reconstituting the samples with 2:1 tert-butanol triton \times solution, the sample with scintillation fluid was placed into scintillation tubes and counts measured with a scintillation counter. Subtracting the mitochondrial GPAT (NEM-resistant) activity from the total activity allowed for determining GPAT1's activity for each sample.

Statistical analysis

Significant differences between groups were determined using two way analysis of variance (ANOVA) and Bonferonni post hoc test for multiple comparisons. The statistical software SigmaStat (Systat Software Inc, San Jose, CA) was used. Statistical significance is defined at $p < 0.05$. Results are presented as means \pm standard error of mean (SEM).

Results

Chronic activation of AMPK limits hepatic triglyceride accumulation

We have previously reported that rats given the same treatment as in this study exhibit a significant main effect of high fat feeding on increased circulating FFA's and abdominal fat accumulation [39]. In addition, this duration of feeding as well as the dose and frequency of AICAR treatment does not result in a significant increase in body weight due to high fat feeding compared to control fed rats [39]. However, as we expected, chronic activation of AMPK using daily AICAR injections caused a reduction in hepatic triglyceride content. We verified that the subcutaneous AICAR injections at the dose we used (0.5 mg/g body weight) were sufficient to cause activation of AMPK in the liver by measuring AMPK phosphorylation one hour after acute subcutaneous injection in rats (see Figure 1). Chronic activation of

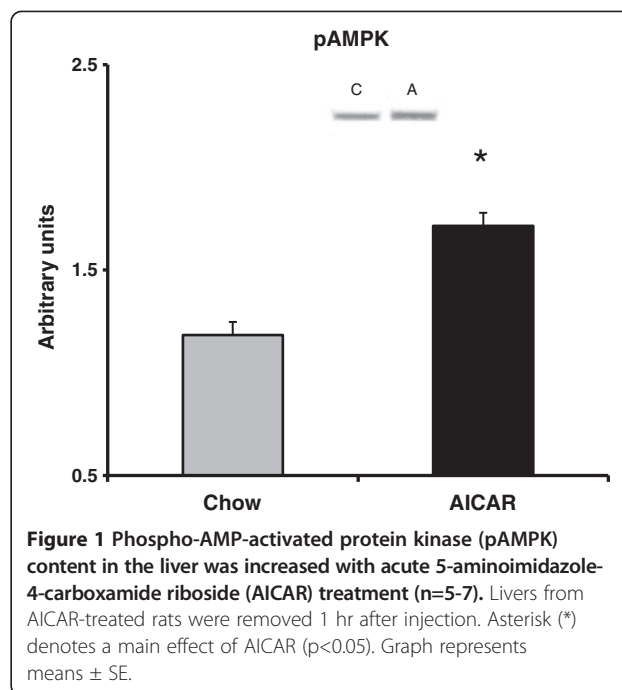


Figure 1 Phospho-AMP-activated protein kinase (pAMPK) content in the liver was increased with acute 5-aminoimidazole-4-carboxamide riboside (AICAR) treatment (n=5-7). Livers from AICAR-treated rats were removed 1 hr after injection. Asterisk (*) denotes a main effect of AICAR ($p < 0.05$). Graph represents means \pm SE.

AMPK limited the normal increase in triglyceride accumulation that occurs with high fat feeding such that it was not significantly different from the control group (See Figure 2). This finding is consistent with previous reports on the effects of AMPK activation on fat accumulation in the liver [45-47]. The mechanism by which AMPK causes this reduction in a prolonged treatment of six weeks has yet to be fully characterized.

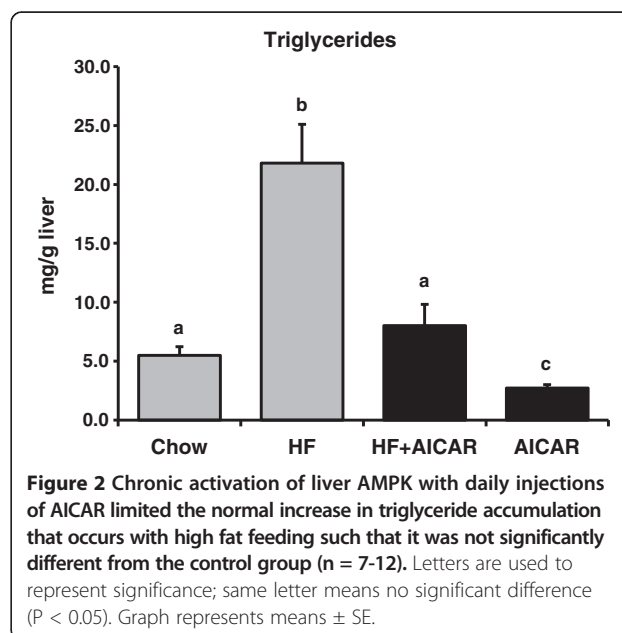
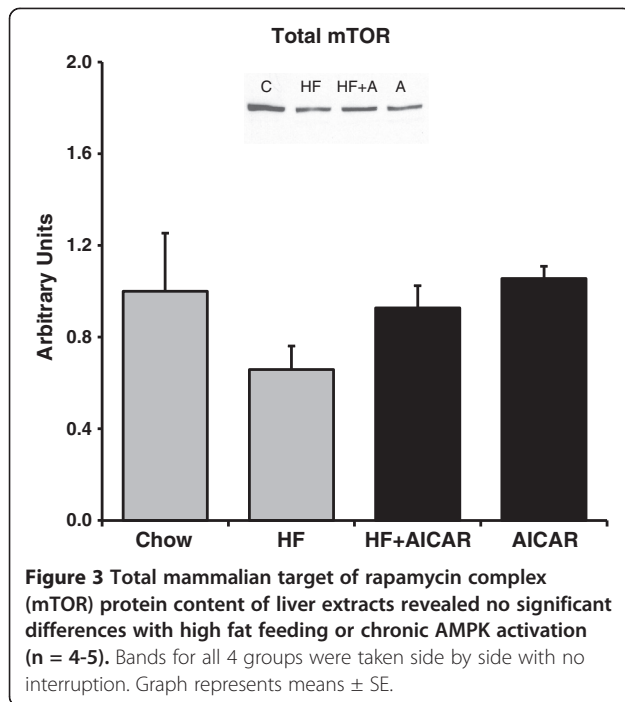


Figure 2 Chronic activation of liver AMPK with daily injections of AICAR limited the normal increase in triglyceride accumulation that occurs with high fat feeding such that it was not significantly different from the control group (n = 7-12). Letters are used to represent significance; same letter means no significant difference ($P < 0.05$). Graph represents means \pm SE.



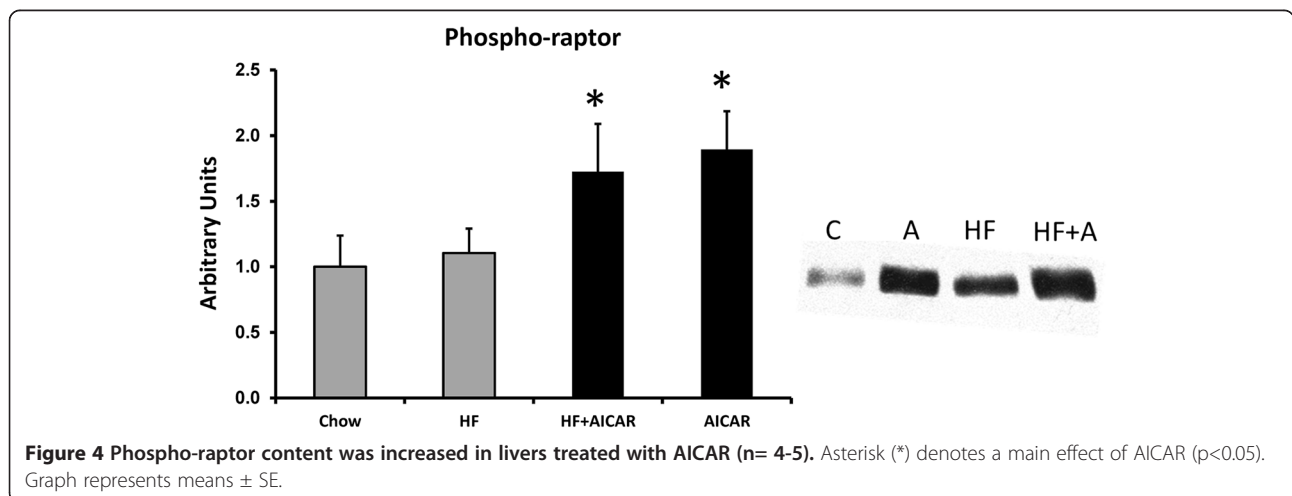
Regulation of lipid synthesis

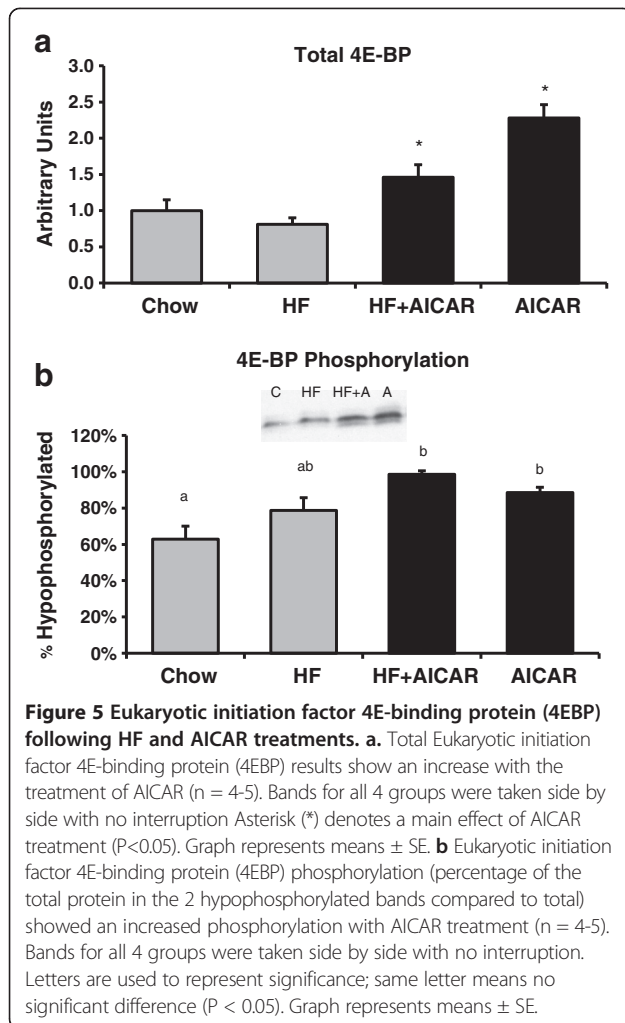
Chronic activation of AMPK decreased SREBP-1c in livers of rats fed a high fat diet

Based on previous findings, chronic AMPK activation would be expected to reduce transcription of GPAT via inhibition of mTOR and SREBP-1c [4,35]. AMPK is known to inhibit mTOR activity therefore we examined mTOR, an mTOR regulatory protein (raptor), and a downstream target of mTOR (4EBP) as an indication of mTOR activity [48]. Western blots on total mTOR complex protein in each group did not indicate a significant difference between the groups (See Figure 3). As would

be anticipated from the activation of AMPK, the total levels of phospho-raptor were significantly increased with chronic AMPK activation with both chow and high fat feeding (See Figure 4). Phosphorylation of raptor on (S792) by AMPK is important for mTOR inhibition [49-51]. The total abundance of 4E-BP was significantly increased with chronic AMPK activation with both the chow and high fat feeding (See Figure 5a.) The phosphorylation state was determined by the shift in molecular weight of total 4E-BP protein (percentage of the total protein in the two hypophosphorylated bands migrated down 1-2 kDa compared to the total at molecular weight of 20 kDa as previously documented) [48,52]. Considering the shift of 4EBP, our results indicate that there was lower amount of phosphorylated 4EBP following chronic AICAR treatment compared to the control group, which is consistent with the known inhibitory effect of AMPK on mTOR activity (See Figure 5b).

AMPK plays a major role in the activity of SREBP-1c in the liver by inhibiting mTOR complex activity [37]. SREBP-1c is positively regulated by mTOR and therefore lipogenesis is upregulated with increased mTOR activity [35]. We examined both forms of SREBP-1c in our study: the full-length (inactive) form, and cleaved (active) form [53]. High fat feeding caused a marked increase in total full-length, uncleaved SREBP-1c abundance. Consistent with the pattern observed in hepatic triglyceride accumulation, chronic activation of AMPK caused a reduction in the total full length, uncleaved SREBP-1c abundance in rats fed either chow or high fat diet (see Figure 6a). The cleaved SREBP-1c showed increases with high fat feeding and decreases with chronic AMPK activation as well but the differences were not as pronounced (See Figure 6b). Therefore, our data indicates that chronic activation of AMPK inhibits both full length and cleaved SREBP-1c protein abundance; this



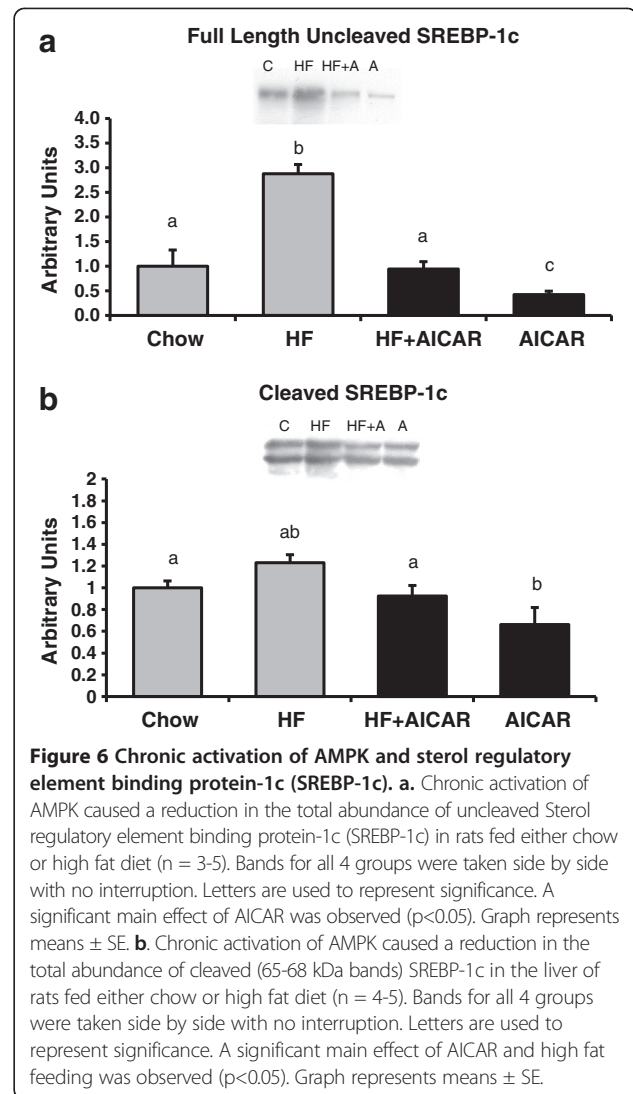


was consistent with what we observed with the mTOR dependent response as seen with 4EBP phosphorylation.

Chronic activation of AMPK had no effect on GPAT1 activity but a high fat feeding effect was present

Lipid synthesis enzymes increased by SREBP-1c include ACC and GPAT. We first examined the abundance of total ACC in response to high fat feeding and chronic AMPK activation and found that AMPK activation caused a significant reduction in total ACC protein in the chow group. Interestingly, high fat feeding did not produce a significant increase in total ACC protein (See Figure 7). These results are consistent with cleaved SREBP-1c total content.

GPAT1 activity was measured because it is another lipogenic target of SREBP-1C and is a rate-limiting enzyme for triglyceride synthesis [36]. High fat feeding caused an increase in total and NEM-sensitive GPAT activity in the liver (Figure 8a). Surprisingly, chronic activation of AMPK in either control or high fat fed animals did not cause a reduction in total or NEM-

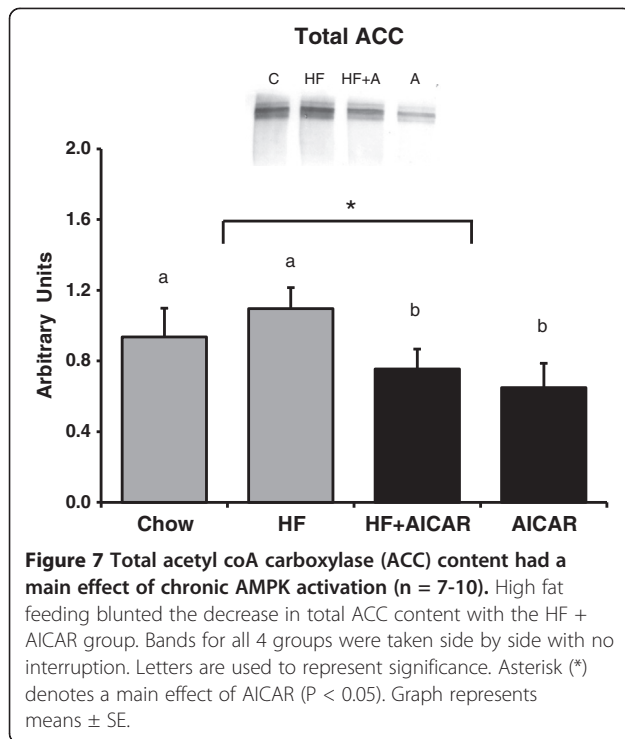


sensitive (GPAT1) activity (Figure 8b). Our results present the novel finding that there is not a direct correlation of chronic activation of AMPK with GPAT1 activity. We expected to see a reduction in GPAT1 activity based on previous results in hepatocytes regarding the acute effect of AMPK on GPAT activity [5,26]. These findings prompted further exploration of the mechanisms and regulation of fatty acid oxidation.

Lipid oxidation

Long chain acyl-CoA dehydrogenase (LCAD) was not influenced by chronic AMPK activation but was increased with high fat feeding

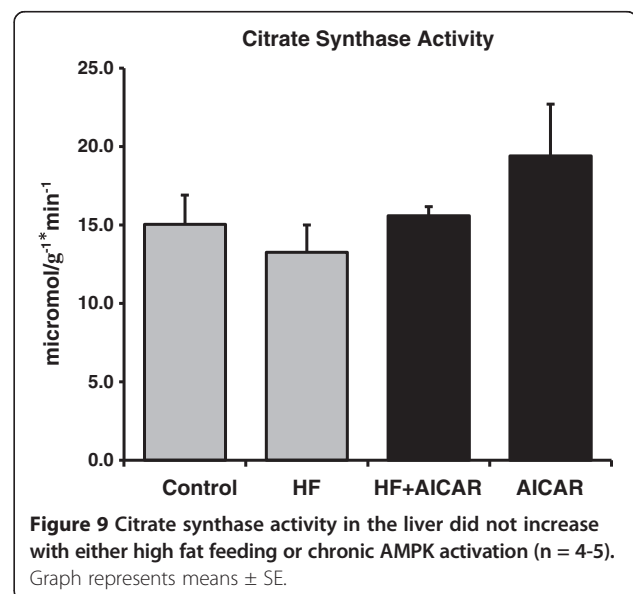
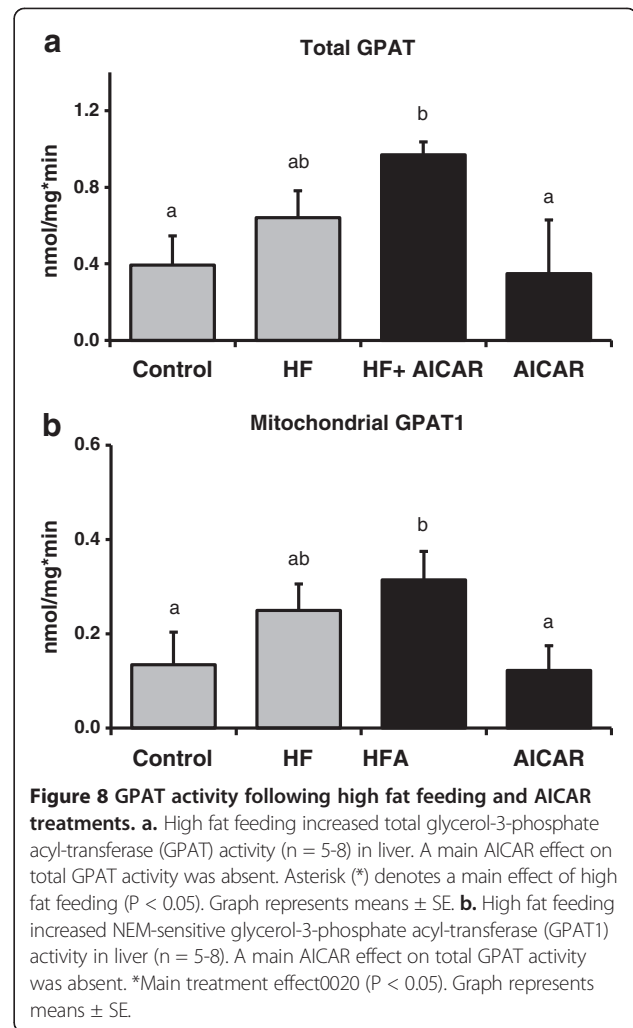
Hepatic lipid accumulation is a balance between the lipid synthesis and oxidation so two markers of mitochondrial oxidative capacity in the liver were measured. Neither high fat feeding nor chronic activation of AMPK showed statistically significant differences between groups for

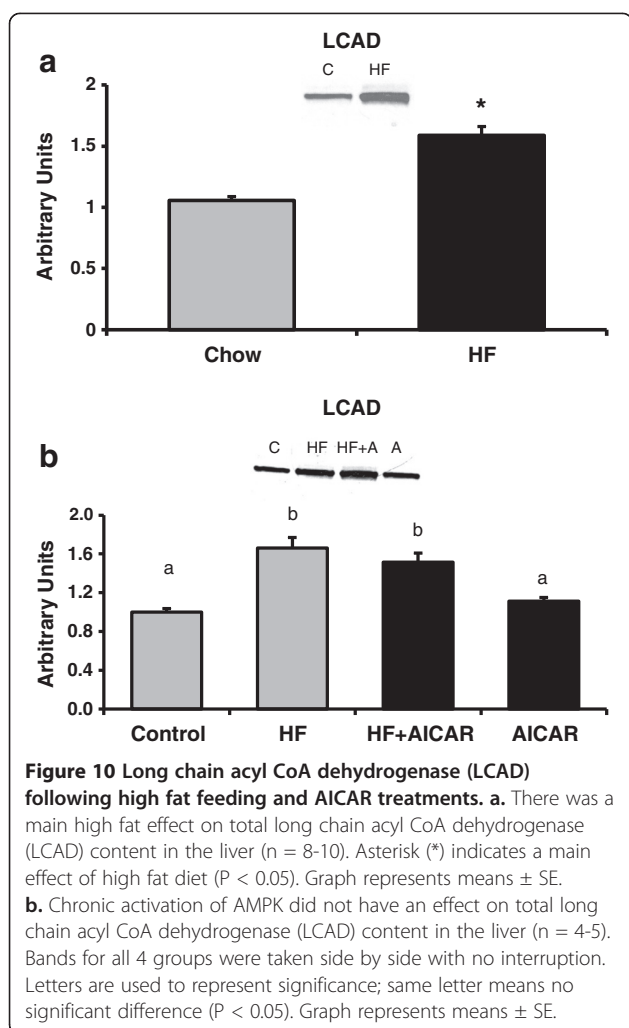


citrate synthase activity (See Figure 9) or cytochrome c content (data not shown) in the liver. Long chain acyl-CoA dehydrogenase (LCAD), a key enzyme responsible for the first step in the oxidation of long-chain fatty acyl-CoAs was measured [54]. A significant increase in LCAD was observed in response to high fat feeding, suggesting greater capacity for fat oxidation (See Figure 10a). Interestingly no effect was seen in the animals treated with AICAR (See Figure 10b). These results suggest that chronic AMPK activation does not play a significant role in changing the oxidative capacity of liver content contrary to what has been found previously in skeletal muscle [38,39,55]. Similar to the GPAT1 data, these results do not account for the observed difference in hepatic triglyceride levels in response to chronic AMPK activation.

Discussion

The purpose of this study was to examine a potential mechanism by which chronic AMPK activation limits fat accumulation in the liver. We hypothesized that AMPK would cause a reduction in GPAT1 activity, a rate limiting enzyme for triglyceride synthesis. GPAT1 has been shown to be influential in the development of NAFLD through its role in triglyceride synthesis in the liver and when overexpressed leads to excess triglyceride synthesis [22,26,56-58]. Neschen et al. and Hammond et al. demonstrated that GPAT1 knockout mice had less triglyceride accumulation when compared to wild type mice fed a high fat diet [59,60]. AMPK is thought to inhibit the transcription of GPAT by reducing the activation of





SREBP-1c. SREBP-1c is the primary transcription factor for GPAT1 and other lipogenic enzymes [31-33]. In contrast to our hypothesis, we found that chronic AMPK activation did not cause a reduction in GPAT1 activity in either the control group or the animals receiving a high fat diet. Therefore, results from this study suggest that chronic AMPK activation limits triglyceride accumulation in the liver by a mechanism other than a reduction in triglyceride synthesis capacity.

It is well documented that AMPK activation reduces hepatic triglyceride accumulation [46,47]. However the mechanisms responsible for this reduced triglyceride content in the setting of high fat feeding are not fully understood. AMPK has been best characterized as a regulator of fatty acid oxidation [61]. AMPK affects an increase in oxidation by inhibition of ACC [47]. Inhibition of ACC results in less malonyl-CoA synthesis leading to a greater activity of CPT1 due to reduced inhibition by malonyl-CoA [28,61,62]. Recently, a greater appreciation of AMPK as a regulator of triglyceride synthesis has developed. Sterol regulatory element binding

protein-1c (SREBP-1c) is a major regulator of lipogenic enzymes and AMPK reduces SREBP-1c and downstream lipogenic enzymes through an mTOR-dependent mechanism [4,47]. GPAT1 is one lipogenic enzyme that has been clearly associated with an increase in triglyceride synthesis and accumulation. The regulation of GPAT1 by SREBP-1c is evidenced by the 6.7-fold increase in GPAT1 by an overexpression of SREBP-1c in adipocytes [36]. Further, an expected increase in GPAT1 with re-feeding does not occur in liver in the absence of SREBP-1c [63]. Results from our study confirm that AMPK activation leads to a reduction in SREBP-1c abundance. Consistent with this reduction in SREBP-1c content, we observed an AMPK dependent reduction in total ACC, one of the enzymes positively regulated by SREBP-1c. GPAT1 activity assay results were unexpected and differed from the pattern observed with triglycerides, SREBP-1c and ACC. Total and NEM-sensitive (GPAT1) GPAT activity was increased with high fat feeding but chronic AMPK activation did not appear to have an inhibitory effect. Therefore our results in intact liver with chronic AMPK activation suggest an alternative regulation of total triglyceride synthesis capacity via GPAT1 than has previously been proposed in isolated hepatocytes with acute AMPK activation.

The regulation of SREBP-1c by AMPK is thought to be dependent upon inhibition of mammalian target of rapamycin (mTOR) and transcriptional activity of liver X receptor (LXR) and SREBP-1c [4,35,64]. SREBP-1c is significantly decreased by inhibitors of mTOR such as rapamycin. [65]. This indicates that through AMPK's inhibition of mTOR activity, AMPK has the effect of reducing SREBP-1c activity. Further, AMPK's role in reducing mTOR activity results in decreased protein synthesis in liver tissue [48]. The mechanism by which AMPK decreases mTOR activity was proposed by Inoki et al. to be by phosphorylation and activation of an upstream protein in the signaling cascade, tuberous sclerosis complex 1/2 (TSC1/2) [66,67]. mTOR phosphorylates downstream proteins such as eukaryotic translation initiation factor 4E-binding protein (4E-BP) and ribosomal protein p70 S6 (S6K1) thereby increasing translation of various proteins and overall protein synthesis [48,68]. Therefore, we can get an indication of the effect of chronic AMPK activation on mTOR activity by measuring the phosphorylation state (or shift) of 4E-BP [48,52]. Our study validated the effect of AMPK activation in the liver by showing a decrease in phosphorylated 4E-BP (increased hypophosphorylation) in the AICAR treated groups [52]. This suggests an inhibition of mTOR activity and explanation for the pattern seen in the SREBP-1c results.

Triglycerides accumulate in the liver particularly with chronic high fat feeding through an up-regulation of

lipogenic enzymes that enhance fatty acid and triglyceride synthesis and greater inhibition of CPT-1, a major regulator of beta-oxidation. This is evidenced by a marked decrease in beta oxidation when GPAT1 is overexpressed in hepatocytes [56,58,69] and increased beta oxidation markers when GPAT1 is knocked out in mouse myocytes [59]. There is clear evidence that a chronic high fat diet results in significantly higher hepatic weights and triglyceride levels [8,70,71]. Our study duplicated such results with an increase in triglycerides after prolonged high fat feeding. In accordance with reported results of AMPK activation in cultured hepatocyte models [45,72], the chronic AICAR treated intact liver tissue in our study had reduced levels of triglycerides in the liver to control levels. AMPK activation inhibits triglyceride accumulation by increasing beta oxidation in the cell [47,73] as well as in its proposed inhibition of mTOR and downstream targets such as SREBP-1c as noted above [67,74]. These mechanisms could explain the fat accumulation with high fat feeding and reductions with chronic AICAR treatment in the livers that was seen in our triglyceride assay results. Therefore, the reduction seen in triglyceride accumulation with chronic AMPK activation was consistent with what was expected.

Increased fat oxidation with high fat feeding could be another contributing factor to explain the conflicting findings of triglyceride content and GPAT1 data in our study. High fat states, such as ob/ob models, have shown an increase oxidative capacity with a simultaneous increase in fatty acid oxidation [75]. This high fat effect on fatty acid oxidative capacity gave reason for measuring LCAD, a marker of fatty acid oxidative capacity. Further, AMPK activation is known to influence mitochondrial biogenesis in both skeletal muscle [38,39,55] and in adipose tissue [76]. Interestingly, we did not see an increase in either citrate synthase activity or cytochrome c content with either high fat feeding or chronic activation of AMPK in the liver. However, a significant increase of LCAD with high fat feeding was observed. The increase seen in LCAD is consistent with the high fat effect expected but the chronic AMPK activation effect was not apparent from the data. Therefore, the issue of how the chronic effects of AMPK activation lead to a decrease in hepatic triglyceride accumulation remains to be resolved.

It is important to note some of the limitations in our study. First, our study did not investigate the acute regulation of GPAT by AMPK noted in other studies. AMPK has been shown to have an acute inhibitory effect on GPAT1 activity as shown in previous studies, which is likely due to phosphorylation of GPAT1 [5,24,26,77]. This acute effect was not the focus of our study, and it is not known whether this played a factor in overall trigly-

ceride accumulation. The reduction in triglycerides could be explained solely by the acute inhibition of GPAT by AMPK. Second, the fat in the high fat diet used in this study was composed of olive oil and flaxseed oil and was not a typical composition for a high diet (see Methods) due to use of tissues from animals in a companion study. This may influence fat accumulation patterns seen in our study and/or responsiveness to AMPK. Therefore more work could be done to see if our results in the chronic setting were unique to the type of fat used in our study.

Conclusions

Given the current trends in lifestyle and dietary habits, the prevalence of NAFLD and the development of NASH are likely to continue to increase. AMPK is widely recognized as a central regulator/sensor of cellular energy metabolism and when activated, is known to limit hepatic fat accumulation. Thus, AMPK is an attractive target for potential therapeutic interventions designed to treat excess fat accumulation in the liver.

In this study we examined the effect of chronic activation of AMPK via systemic administration of AICAR on hepatic GPAT1 activity. High fat feeding resulted in increased hepatic GPAT1 activity while chronic AICAR administration had no effect on GPAT1 activity over time. This was in contrast to the clear effect of AICAR treatment on triglyceride content and transcriptional regulation in the liver. Further areas of interest that may help explain our findings include research on the role of AMPK activation and other lipid regulatory mechanisms such as cellular uptake or release of fatty acids and other lipid molecules.

Abbreviations

4E-BP: 4E-binding protein; ACC: Acetyl CoA carboxylase; AICAR: Aminoimidazole carboxamide ribonucleotide; AMPK: AMP Activated Protein Kinase; BCA: Bicinchoninic acid; C: Control; CPT1: Carnitine palmitoyltransferase 1; DTT: Dithiothreitol; EDTA: Ethylenediaminetetraacetic acid; FAS: Fatty Acid Synthase; G-3-P: [¹⁴C]glycerol 3-phosphate; GPAT: Glycerol-3-phosphate acyltransferase; GPAT1: N-ethylmaleimide resistant glycerol-3-phosphate acyltransferase; HF: High Fat; LCAD: Long chain acyl-CoA dehydrogenase; LCRa: Liver x receptors; LXR: Liver x receptor; mTOR: Mammalian target of rapamycin; NAFLD: Non-alcoholic fatty liver disease; NEM: N-ethylmaleimide; S6K1: Ribosomal protein p70 S6; SCD1: Stearoyl-CoA desaturase; SDS-PAGE: Sodium dodecyl sulfate polyacrylamide gel electrophoresis; SREBP-1c: Sterol regulatory element binding protein; TBS: Tris-Buffered Saline; TBST: Tris-Buffered Saline Tween-20; TSC1/2: Tuberous sclerosis complex 1/2.

Competing interests

Bradley Henriksen, Mary Curtis, Natasha Fillmore, Brandon Cardon, David Thomson, Chad Hancock. The authors declare that they have no competing interests.

Authors' contributions

BSH participated in planning the project, conducted western blot analysis, protein and activity assays, animal care, and helped draft the manuscript. MEC also participated in planning the project, conducting protein and activity assays, and helped draft the manuscript. (Both BSH and MEC are equal contributors and are to be considered co-first authors). NF participated in the planning, animal care, and results analysis. BRC participated in the

planning, animal care, and assays. DMT participated in the planning, analysis, and interpretation of assays. CRH conceived of the study, participated in design, coordination, conduction of experiments, analysis, and helped create the manuscript.

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References

- Canto C, Auwerx J: AMP-activated protein kinase and its downstream transcriptional pathways. *Cell Mol Life Sci* 2010, **67**:3407–3423.
- Hardie DG: AMP-activated protein kinase as a drug target. *Annu Rev Pharmacol Toxicol* 2007, **47**:185–210.
- Kahn BB, Alquier T, Carling D, Hardie DG: AMP-activated protein kinase: ancient energy gauge provides clues to modern understanding of metabolism. *Cell Metab* 2005, **1**:15–25.
- Yang J, Craddock L, Hong S, Liu ZM: AMP-activated protein kinase suppresses LXR-dependent sterol regulatory element-binding protein-1c transcription in rat hepatoma McA-RH7777 cells. *J Cell Biochem* 2009, **106**:414–426.
- Muoio DM, Seefeld K, Witters LA, Coleman RA: AMP-activated kinase reciprocally regulates triacylglycerol synthesis and fatty acid oxidation in liver and muscle: evidence that sn-glycerol-3-phosphate acyltransferase is a novel target. *Biochem J* 1999, **338**(Pt 3):783–791.
- Assifi MM, Suchankova G, Constant S, Prentki M, Saha AK, Ruderman NB: AMP-activated protein kinase and coordination of hepatic fatty acid metabolism of starved/carbohydrate-refed rats. *Am J Physiol Endocrinol Metab* 2005, **289**:E794–800.
- Donnelly KL, Smith CI, Schwarzenberg SJ, Jessurun J, Boldt MD, Parks EJ: Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. *J Clin Invest* 2005, **115**:1343–1351.
- Wang Z, Yao T, Pini M, Zhou Z, Fantuzzi G, Song Z: Betaine improved adipose tissue function in mice fed a high-fat diet: a mechanism for hepatoprotective effect of betaine in nonalcoholic fatty liver disease. *Am J Physiol Gastrointest Liver Physiol* 2010, **298**:G634–642.
- Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, Ferrell LD, Liu YC, Torbenson MS, Unalp-Arida A, et al: Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 2005, **41**:1313–1321.
- Musso G, Gambino R, Cassader M: Non-alcoholic fatty liver disease from pathogenesis to management: an update. *Obes Rev* 2010, **11**:430–445.
- Rector RS, Thyfault JP, Wei Y, Ibdah JA: Non-alcoholic fatty liver disease and the metabolic syndrome: an update. *World J Gastroenterol* 2008, **14**:185–192.
- Tessari P, Coracina A, Cosma A, Tiengo A: Hepatic lipid metabolism and non-alcoholic fatty liver disease. *Nutr Metab Cardiovasc Dis* 2009, **19**:291–302.
- Nagle CA, Klett EL, Coleman RA: Hepatic triacylglycerol accumulation and insulin resistance. *J Lipid Res* 2009, **50**(Suppl):S74–79.
- Pasumarthy L, Srour J: Nonalcoholic steatohepatitis: a review of the literature and updates in management. *South Med J* 2010, **103**:547–550.
- Ruhl CE, Everhart JE: Epidemiology of nonalcoholic fatty liver. *Clin Liver Dis* 2004, **8**:501–519. vii.
- Stefanovic-Racic M, Perdomo G, Mantell BS, Sipula IJ, Brown NF, O'Doherty RM: A moderate increase in carnitine palmitoyltransferase 1a activity is sufficient to substantially reduce hepatic triglyceride levels. *Am J Physiol Endocrinol Metab* 2008, **294**:E969–977.
- Rector RS, Thyfault JP, Uptergrove GM, Morris EM, Naples SP, Borengasser SJ, Mikus CR, Laye MJ, Laughlin MH, Booth FW, Ibdah JA: Mitochondrial dysfunction precedes insulin resistance and hepatic steatosis and contributes to the natural history of non-alcoholic fatty liver disease in an obese rodent model. *J Hepatol* 2010, **52**:727–736.
- Strauss RS, Barlow SE, Dietz WH: Prevalence of abnormal serum aminotransferase values in overweight and obese adolescents. *J Pediatr* 2000, **136**:727–733.
- Marchesini G, Bugianesi E, Forlani G, Cerrelli F, Lenzi M, Manini R, Natale S, Vanni E, Villanova N, Melchionda N, Rizzetto M: Nonalcoholic fatty liver, steatohepatitis, and the metabolic syndrome. *Hepatology* 2003, **37**:917–923.
- Paschos P, Paletas K: Non alcoholic fatty liver disease and metabolic syndrome. *Hippokratia* 2009, **13**:9–19.
- Choudhury J, Sanyal AJ: Insulin resistance and the pathogenesis of nonalcoholic fatty liver disease. *Clin Liver Dis* 2004, **8**:575–594. ix.
- Cao J, Li JL, Li D, Tobin JF, Gimeno RE: Molecular identification of microsomal acyl-CoA:glycerol-3-phosphate acyltransferase, a key enzyme in de novo triacylglycerol synthesis. *Proc Natl Acad Sci USA* 2006, **103**:19695–19700.
- Coleman RA, Lee DP: Enzymes of triacylglycerol synthesis and their regulation. *Prog Lipid Res* 2004, **43**:134–176.
- Nimmo HG: Evidence for the existence of isoenzymes of glycerol phosphate acyltransferase. *Biochem J* 1979, **177**:283–288.
- Haldar D, Tso WW, Pullman ME: The acylation of sn-glycerol 3-phosphate in mammalian organs and Ehrlich ascites tumor cells. *J Biol Chem* 1979, **254**:4502–4509.
- Lewin TM, Granger DA, Kim JH, Coleman RA: Regulation of mitochondrial sn-glycerol-3-phosphate acyltransferase activity: response to feeding status is unique in various rat tissues and is discordant with protein expression. *Arch Biochem Biophys* 2001, **396**:119–127.
- Winder WW, Hardie DG: Inactivation of acetyl-CoA carboxylase and activation of AMP-activated protein kinase in muscle during exercise. *Am J Physiol* 1996, **270**:299–304.
- McGarry JD, Mannaerts GP, Foster DW: A possible role for malonyl-CoA in the regulation of hepatic fatty acid oxidation and ketogenesis. *J Clin Invest* 1977, **60**:265–270.
- Yeh YY, Zee P: Fatty acid oxidation in isolated rat liver mitochondria. Developmental changes and their relation to Hepatic levels of Carnitine and glycogen and to Carnitine acyltransferase activity. *Arch Biochem Biophys* 1979, **197**:560–569.
- Tomita K, Tamiya G, Ando S, Kitamura N, Koizumi H, Kato S, Horie Y, Kaneko T, Azuma T, Nagata H, et al: AICAR, an AMPK activator, has protective effects on alcohol-induced fatty liver in rats. *Alcohol Clin Exp Res* 2005, **29**:240S–245S.
- Shimano H, Horton JD, Shimomura I, Hammer RE, Brown MS, Goldstein JL: Isoform 1c of sterol regulatory element binding protein is less active than isoform 1a in livers of transgenic mice and in cultured cells. *J Clin Invest* 1997, **99**:846–854.
- Edwards PA, Tabor D, Kast HR, Venkateswaran A: Regulation of gene expression by SREBP and SCAP. *Biochim Biophys Acta* 2000, **1529**:103–113.
- Sekiya M, Yahagi N, Matsuzaka T, Takeuchi Y, Nakagawa Y, Takahashi H, Okazaki H, Iizuka Y, Ohashi K, Gotoda T, et al: SREBP-1-independent regulation of lipogenic gene expression in adipocytes. *J Lipid Res* 2007, **48**:1581–1591.
- Magana MM, Osborne TF: Two tandem binding sites for sterol regulatory element binding proteins are required for sterol regulation of fatty-acid synthase promoter. *J Biol Chem* 1996, **271**:32689–32694.
- Porstmann T, Santos CR, Griffiths B, Cully M, Wu M, Leevers S, Griffiths JR, Chung YL, Schulze A: SREBP activity is regulated by mTORC1 and contributes to Akt-dependent cell growth. *Cell Metab* 2008, **8**:224–236.
- Erickson J, Jackson SM, Kim JB, Spiegelman BM, Edwards PA: Identification of glycerol-3-phosphate acyltransferase as an adipocyte determination and differentiation factor 1- and sterol regulatory element-binding protein-responsive gene. *J Biol Chem* 1997, **272**:7298–7305.
- Hwang SH, Ki SH, Bae EJ, Kim HE, Kim SG: Role of adenosine monophosphate-activated protein kinase-p70 ribosomal S6 kinase-1 pathway in repression of liver X receptor-alpha-dependent lipogenic gene induction and hepatic steatosis by a novel class of dithiolethiones. *Hepatology* 2009, **49**:1913–1925.
- Hancock CR, Han DH, Chen M, Terada S, Yasuda T, Wright DC, Holloszy JO: High-fat diets cause insulin resistance despite an increase in muscle mitochondria. *Proc Natl Acad Sci USA* 2008, **105**:7815–7820.
- Fillmore N, Jacobs DL, Mills DB, Winder WW, Hancock CR: Chronic AMP-activated protein kinase activation and a high-fat diet have an

- additive effect on mitochondria in rat skeletal muscle. *J Appl Physiol* 2010, **109**:511–520.
40. Garcia-Roves P, Huss JM, Han DH, Hancock CR, Iglesias-Gutierrez E, Chen M, Holloszy JO: Raising plasma fatty acid concentration induces increased biogenesis of mitochondria in skeletal muscle. *Proc Natl Acad Sci USA* 2007, **104**:10709–10713.
41. Srere PA: Controls of citrate synthase activity. *Life Sci* 1974, **15**:1695–1710.
42. Folch J, Lees M, Sloane Stanley GH: A simple method for the isolation and purification of total lipides from animal tissues. *J Biol Chem* 1957, **226**:497–509.
43. Schlossman DM, Bell RM: Triacylglycerol synthesis in isolated fat cells. Evidence that the sn-glycerol-3-phosphate and dihydroxyacetone phosphate acyltransferase activities are dual catalytic functions of a single microsomal enzyme. *J Biol Chem* 1976, **251**:5738–5744.
44. Watt MJ, Holmes AG, Steinberg GR, Mesa JL, Kemp BE, Febbraio MA: Reduced plasma FFA availability increases net triacylglycerol degradation, but not GPAT or HSL activity, in human skeletal muscle. *Am J Physiol Endocrinol Metab* 2004, **287**:E120–127.
45. Hou X, Xu S, Maitland-Toolan KA, Sato K, Jiang B, Ido Y, Lan F, Walsh K, Wierzbicki M, Verbeuren TJ, *et al*: SIRT1 regulates hepatocyte lipid metabolism through activating AMP-activated protein kinase. *J Biol Chem* 2008, **283**:20015–20026.
46. Park KG, Min AK, Koh EH, Kim HS, Kim MO, Park HS, Kim YD, Yoon TS, Jang BK, Hwang JS, *et al*: Alpha-lipoic acid decreases hepatic lipogenesis through adenosine monophosphate-activated protein kinase (AMPK)-dependent and AMPK-independent pathways. *Hepatology* 2008, **48**:1477–1486.
47. Zhou G, Myers R, Li Y, Chen Y, Shen X, Fenyk-Melody J, Wu M, Ventre J, Doebber T, Fujii N, *et al*: Role of AMP-activated protein kinase in mechanism of metformin action. *J Clin Invest* 2001, **108**:1167–1174.
48. Reiter AK, Bolster DR, Crozier SJ, Kimball SR, Jefferson LS: Repression of protein synthesis and mTOR signaling in rat liver mediated by the AMPK activator aminoimidazole carboxamide ribonucleoside. *Am J Physiol Endocrinol Metab* 2005, **288**:E980–988.
49. Gwinn DM, Shackelford DB, Egan DF, Mihaylova MM, Mery A, Vasquez DS, Turk BE, Shaw RJ: AMPK phosphorylation of raptor mediates a metabolic checkpoint. *Mol Cell* 2008, **30**:214–226.
50. Hara K, Maruki Y, Long X, Yoshino K, Oshiro N, Hidayat S, Tokunaga C, Avruch J, Yonezawa K: Raptor, a binding partner of target of rapamycin (TOR), mediates TOR action. *Cell* 2002, **110**:177–189.
51. Kim DH, Sarbassov DD, Ali SM, King JE, Latek RR, Erdjument-Bromage H, Tempst P, Sabatini DM: mTOR interacts with raptor to form a nutrient-sensitive complex that signals to the cell growth machinery. *Cell* 2002, **110**:163–175.
52. Thomson DM, Fick CA, Gordon SE: AMPK activation attenuates S6K1, 4E-BP1, and eEF2 signaling responses to high-frequency electrically stimulated skeletal muscle contractions. *J Appl Physiol* 2008, **104**:625–632.
53. Sakai J, Nohturfft A, Goldstein JL, Brown MS: Cleavage of sterol regulatory element-binding proteins (SREBPs) at site-1 requires interaction with SREBP cleavage-activating protein. Evidence from in vivo competition studies. *J Biol Chem* 1998, **273**:5785–5793.
54. Zhang D, Liu ZX, Choi CS, Tian L, Kibbey R, Dong J, Cline GW, Wood PA, Shulman GI: Mitochondrial dysfunction due to long-chain Acyl-CoA dehydrogenase deficiency causes hepatic steatosis and hepatic insulin resistance. *Proc Natl Acad Sci USA* 2007, **104**:17075–17080.
55. Winder WW, Holmes BF, Rubink DS, Jensen EB, Chen M, Holloszy JO: Activation of AMP-activated protein kinase increases mitochondrial enzymes in skeletal muscle. *J Appl Physiol* 2000, **88**:2219–2226.
56. Linden D, William-Olsson L, Rhedin M, Asztely AK, Clapham JC, Schreyer S: Overexpression of mitochondrial GPAT in rat hepatocytes leads to decreased fatty acid oxidation and increased glycerolipid biosynthesis. *J Lipid Res* 2004, **45**:1279–1288.
57. Nagle CA, An J, Shiota M, Torres TP, Cline GW, Liu ZX, Wang S, Catlin RL, Shulman GI, Newgard CB, Coleman RA: Hepatic overexpression of glycerol-sn-3-phosphate acyltransferase 1 in rats causes insulin resistance. *J Biol Chem* 2007, **282**:14807–14815.
58. Linden D, William-Olsson L, Ahnmark A, Ekroos K, Hallberg C, Sjogren HP, Becker B, Svensson L, Clapham JC, Oscarsson J, Schreyer S: Liver-directed overexpression of mitochondrial glycerol-3-phosphate acyltransferase results in hepatic steatosis, increased triacylglycerol secretion and reduced fatty acid oxidation. *FASEB J* 2006, **20**:434–443.
59. Neschen S, Morino K, Hammond LE, Zhang D, Liu ZX, Romanelli AJ, Cline GW, Pongratz RL, Zhang XM, Choi CS, *et al*: Prevention of hepatic steatosis and hepatic insulin resistance in mitochondrial acyl-CoA:glycerol-sn-3-phosphate acyltransferase 1 knockout mice. *Cell Metab* 2005, **2**:55–65.
60. Hammond LE, Gallagher PA, Wang S, Hiller S, Kluckman KD, Posey-Marcos EL, Maeda N, Coleman RA: Mitochondrial glycerol-3-phosphate acyltransferase-deficient mice have reduced weight and liver triacylglycerol content and altered glycerolipid fatty acid composition. *Mol Cell Biol* 2002, **22**:8204–8214.
61. Merrill GF, Kurth EJ, Hardie DG, Winder WW: AICA riboside increases AMP-activated protein kinase, fatty acid oxidation, and glucose uptake in rat muscle. *Am J Physiol* 1997, **273**:1107–1112.
62. McGarry JD, Leatherman GF, Foster DW: Carnitine palmitoyltransferase I. The site of inhibition of hepatic fatty acid oxidation by malonyl-CoA. *J Biol Chem* 1978, **253**:4128–4136.
63. Shimano H, Yahagi N, Amemiya-Kudo M, Hasty AH, Osuga J, Tamura Y, Shionoiri F, Iizuka Y, Ohashi K, Harada K, *et al*: Sterol regulatory element-binding protein-1 as a key transcription factor for nutritional induction of lipogenic enzyme genes. *J Biol Chem* 1999, **274**:35832–35839.
64. Kimball SR: Interaction between the AMP-activated protein kinase and mTOR signaling pathways. *Med Sci Sports Exerc* 2006, **38**:1958–1964.
65. Li S, Brown MS, Goldstein JL: Bifurcation of insulin signaling pathway in rat liver: mTORC1 required for stimulation of lipogenesis, but not inhibition of gluconeogenesis. *Proc Natl Acad Sci USA* 2010, **107**:3441–3446.
66. Inoki K, Zhu T, Guan KL: TSC2 mediates cellular energy response to control cell growth and survival. *Cell* 2003, **115**:577–590.
67. Laplante M, Sabatini DM: An emerging role of mTOR in lipid biosynthesis. *Curr Biol* 2009, **19**:R1046–1052.
68. Kimball SR, Siegfried BA, Jefferson LS: Glucagon represses signaling through the mammalian target of rapamycin in rat liver by activating AMP-activated protein kinase. *J Biol Chem* 1999, **274**:54103–54109.
69. Lewin TM, Wang S, Nagle CA, Van Horn CG, Coleman RA: Mitochondrial glycerol-3-phosphate acyltransferase-1 directs the metabolic fate of exogenous fatty acids in hepatocytes. *Am J Physiol Endocrinol Metab* 2005, **288**:E835–844.
70. Yazdi M, Ahnmark A, William-Olsson L, Snaith M, Turner N, Osla F, Wedin M, Asztely AK, Elmgren A, Bohlooly YM, *et al*: The role of mitochondrial glycerol-3-phosphate acyltransferase-1 in regulating lipid and glucose homeostasis in high-fat diet fed mice. *Biochem Biophys Res Commun* 2008, **369**:1065–1070.
71. Kim HJ, Kim JH, Noh S, Hur HJ, Sung MJ, Hwang JT, Park JH, Yang HJ, Kim SM, Kwon DY, Yoon SH: Metabolic analysis of livers and serum from high-fat diet induced obese mice. *J Proteome Res* 2011, **10**:722–731.
72. Zang M, Zuccollo A, Hou X, Nagata D, Walsh K, Herscovitz H, Brecher P, Ruderman NB, Cohen RA: AMP-activated protein kinase is required for the lipid-lowering effect of metformin in insulin-resistant human HepG2 cells. *J Biol Chem* 2004, **279**:47898–47905.
73. Winder WW, Hardie DG: AMP-activated protein kinase, a metabolic master switch: possible roles in type 2 diabetes. *Am J Physiol* 1999, **277**:1–10.
74. Bolster DR, Crozier SJ, Kimball SR, Jefferson LS: AMP-activated protein kinase suppresses protein synthesis in rat skeletal muscle through down-regulated mammalian target of rapamycin (mTOR) signaling. *J Biol Chem* 2002, **277**:23977–23980.
75. Brady LJ, Brady PS, Romsos DR, Hoppel CL: Elevated hepatic mitochondrial and peroxisomal oxidative capacities in fed and starved adult obese (ob/ob) mice. *Biochem J* 1985, **231**:439–444.
76. Sutherland LN, Capozzi LC, Turchinsky NJ, Bell RC, Wright DC: Time course of high-fat diet-induced reductions in adipose tissue mitochondrial proteins: potential mechanisms and the relationship to glucose intolerance. *Am J Physiol Endocrinol Metab* 2008, **295**:E1076–1083.
77. Bhat BG, Wang P, Kim JH, Black TM, Lewin TM, Fiedorek FT, Coleman RA: Rat sn-glycerol-3-phosphate acyltransferase: molecular cloning and characterization of the cDNA and expressed protein. *Bba-Mol Cell Biol L* 1999, **1439**:415–423.

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