


Article

# Chloro-Furanocembranolides from *Leptogorgia* sp. Improve Pancreatic Beta-Cell Proliferation

Amalia B. Gallardo <sup>1,2</sup>, Ana R. Díaz-Marrero <sup>1,†</sup> , José M. de la Rosa <sup>1</sup>, Luis D’Croz <sup>3,4</sup>, Germán Perdomo <sup>5</sup>, Irene Cózar-Castellano <sup>6</sup>, José Darías <sup>1</sup> and Mercedes Cueto <sup>1,\*</sup>

<sup>1</sup> Instituto de Productos Naturales y Agrobiología (IPNA-CSIC), Avenida Astrofísico F. Sánchez, 3, 38206 La Laguna, Tenerife, Spain; amalia.gallardo@umag.cl (A.B.G.); adiazmar@ull.edu.es (A.R.D.-M.); jmrosa@ull.es (J.M.d.l.R.); dariasjose4@gmail.com (J.D.)

<sup>2</sup> Departamento de Química, Facultad de Ciencias, Universidad de Magallanes, Avenida Bulnes 01855, Punta Arenas 6200000, Chile

<sup>3</sup> Departamento de Biología Marina y Limnología, Universidad de Panamá, Panama City P.O. Box 3366, Panama; dcrozlc@gmail.com

<sup>4</sup> Smithsonian Tropical Research Institute, STRI, Balboa P.O. Box 0843-03092, Panama

<sup>5</sup> Facultad de Ciencias de la Salud, Universidad de Burgos, 09001 Burgos, Spain; gmperdomo@ubu.es

<sup>6</sup> Instituto de Biología y Genética Molecular, University of Valladolid-CSIC, 47005 Valladolid, Spain; cozarirene@gmail.com

\* Correspondence: mcueto@ipna.csic.es; Tel.: +34-922-250-144

† Present address: University Institute of Bio-Organic Chemistry “Antonio González” (CIBICAN), University of La Laguna, 38206 Tenerife, Spain.

Received: 20 December 2017; Accepted: 29 January 2018; Published: 2 February 2018

**Abstract:** Two new chloro-furanocembranolides (**1**, **2**) and two new 1,4-diketo cembranolides (**3**, **4**) were isolated from the crude extract of *Leptogorgia* sp. together with a new *seco*-furanocembranolide (**5**) and the known *Z*-deoxypukalide (**6**), rubifolide (**7**), scabrolide D (**8**) and epoxylophodione (**9**). Their structures were determined based on spectroscopic evidence. Four compounds: **1**, **2**, **7** and **8** were found to activate the proliferation of pancreatic insulin-producing (beta) cells.

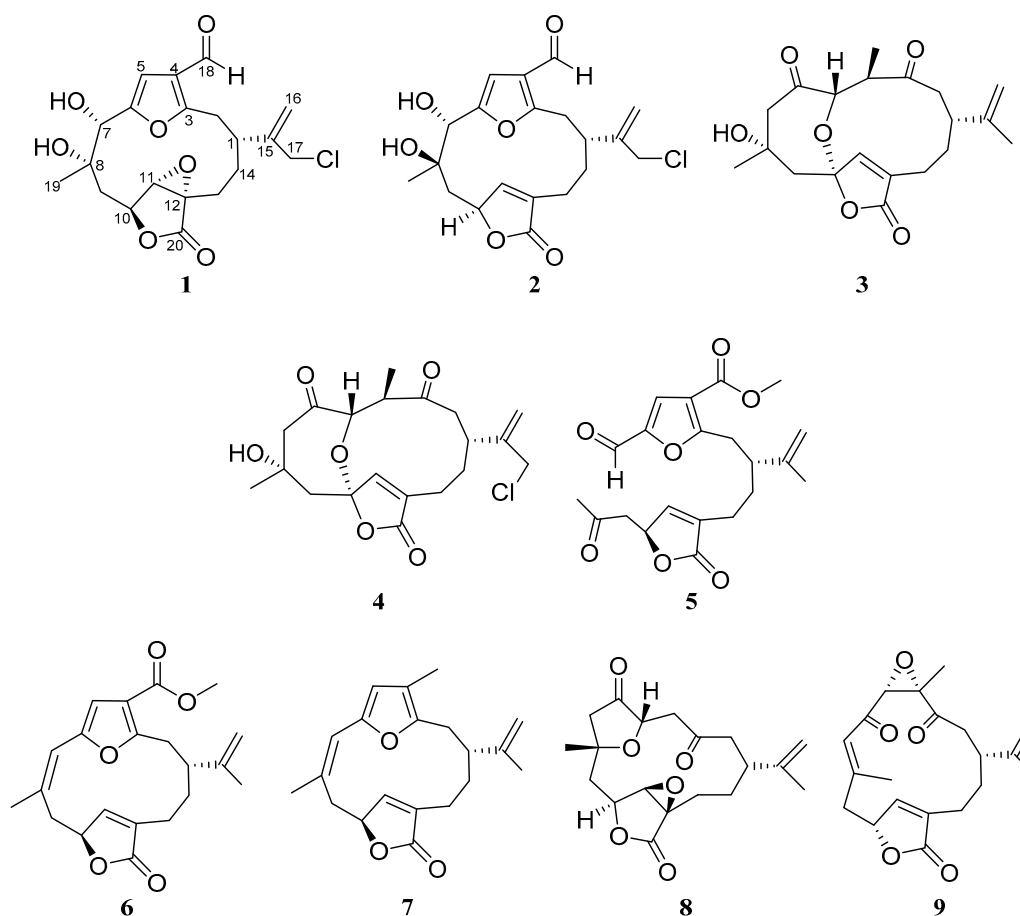
**Keywords:** *Leptogorgia*; cembranolides; furanocembranolides; diketocembranolides; *seco*-furanocembranolides; chloro-furanocembranolides; pancreatic beta-cells

## 1. Introduction

Octocorals of the genus *Leptogorgia* biosynthesize highly oxygenated diterpenoids based on the cembrane skeleton: (a) furanocembranolides into which a  $\gamma$ -lactone subunit is embedded; (b) 1,4-diketocembranoids produced by oxidative cleavage of the furan ring; and (c) *nor*-1,4-diketocembranolides that lack the C-18 isoprenic methyl group [1,2].

It has been suggested that activation of pancreatic beta-cell proliferation is a strategy to maintain functional beta-cell mass in diabetes mellitus [3]. Regarding this, we have shown that furanocembranoids such as epoxyypukalide, pukalide, *Z*-deoxypukalide and leptolide improve beta-cell proliferation [4–6]. This prompted us to study a new extract of *Leptogorgia* sp. collected by SCUBA diving off Aleta (Panama), from which compounds 1–9 were obtained after flash chromatography followed by HPLC (Figure 1). The unusual halogenated compounds **1** and **2**, together with the known *Z*-desoxyypukalide (**6**) [1] and rubifolide (**7**) [7] belong to the furanocembranolide structural class. The remaining six compounds typify three different subclasses that involve several types of rearrangements from a furanocembranolide precursor: (a) **3**, **4** and the previously isolated epoxylophodione (**9**) [8] are 1,4-diketocembranolides, **4** being the only haloderivative of the genus *Leptogorgia* of this subclass; (b) compound **5** is the first *seco*-furanocembranolide isolated from genus

*Leptogorgia*, which showed a similar ring scission pattern to *seco*-bipinnatin J, the only *seco*-derivative [9] isolated from *Pseudopterogorgia*; and (c) scabrolide D (8) [10] is a *nor*-1,4-diketocembranolide which lacks the C-18 isoprenic methyl group.



**Figure 1.** Novel chloro-furanocembranolides (1, 2), 1,4-diketocembranolides (3, 4), *seco*-furanocembranolide (5) and the known *Z*-deoxypukalide (6), rubifolide (7), scabrolide D (8) and epoxylophodione (9) from *Leptogorgia* sp.

## 2. Results

Compound 1 was obtained as an oil whose EIMS spectrum showed peaks at  $m/z$   $[M - 1]^+$  409/411, with relative intensities suggesting one chlorine atom. These peaks correspond to the molecular formula  $C_{20}H_{23}ClO_7$  (HREIMS) ( $m/z$  409.1041  $[M - 1]^+$ , calcd. for  $C_{20}H_{22}^{35}ClO_7$  409.1054). The diterpenic nature of 1 is corroborated by the  $^{13}C$  NMR spectrum (in Supplementary Materials), which displayed correlations in the HSQC spectrum indicative of seven quaternary carbons, six methines, six methylenes and one methyl (Table 1). Absorptions for a hydroxyl group at  $3476\text{ cm}^{-1}$  and carbonyl groups at  $1776$  and  $1676\text{ cm}^{-1}$  were observed in the IR spectrum.

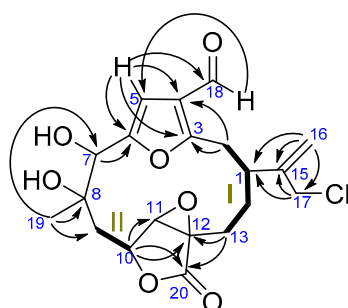
Notable  $^1H$  and  $^{13}C$  NMR signals are: an aldehyde group [ $\delta_{H-18}$  9.89 (1H, s),  $\delta_{C-18}$  184.7], a trisubstituted furane ring [ $\delta_{H-5}$  6.70 (1H, s),  $\delta_{C-5}$  106.5;  $\delta_{C-3}$  161.3;  $\delta_{C-6}$  156.0;  $\delta_{C-4}$  123.4], a  $\alpha,\beta$ -epoxy- $\gamma$ -lactone ring [ $\delta_{H-11}$  3.73 (1H, s),  $\delta_{C-11}$  62.9;  $\delta_{C-20}$  172.1;  $\delta_{H-10}$  4.85 (1H, dd,  $J = 5.2, 11.3$  Hz),  $\delta_{C-10}$  74.3 and  $\delta_{C-12}$  60.6] and a chloromethylene [ $\delta_{H-17}$  4.18 (2H, br s) y  $\delta_{C-17}$  46.6].

Connectivity information obtained from COSY, HSQC and HMBC experiments unambiguously determined the planar structure of compound 1 as a furanocembranolide, containing a C-18 oxidized to aldehyde, a vicinal diol at C-7–C-8, a C-10–C-20  $\alpha,\beta$ -epoxy- $\gamma$ -lactone moiety and a chloroisoprenyl group at C-1.

$^1\text{H}$ - $^1\text{H}$ -COSY experiments established two spin systems:  $\text{H}_2$ -2- $\text{H}_2$ -13 (fragment I) and  $\text{H}_2$ -9- $\text{H}$ -10 (fragment II) (Figure 2). The HMBC correlations  $\text{H}_2$ -16/ $\text{C}$ -17,  $\text{C}$ -15,  $\text{C}$ -1 and  $\text{H}_2$ -17/ $\text{C}$ -16,  $\text{C}$ -15,  $\text{C}$ -1 locate a chloroisopropenyl group at  $\text{C}$ -1 of fragment I, whereas the correlations  $\text{H}_3$ -19/ $\text{C}$ -7,  $\text{C}$ -8 and  $\text{C}$ -9 allowed us to lengthen fragment II by adding a vicinal dihydroxyl moiety bonded to  $\text{C}$ -9. The north end of both fragments II and I are connected together by insertion of a furane ring, in agreement with the HMBC correlations  $\text{H}$ -5/ $\text{C}$ -3,  $\text{C}$ -4,  $\text{C}$ -6 and  $\text{H}_2$ -2 with  $\text{C}$ -4. An epoxy lactone binds the southern ends of fragments I and II between  $\text{C}$ -13 and  $\text{C}$ -10, as deduced from the HMBC correlations  $\text{H}$ -10/ $\text{C}$ -11,  $\text{C}$ -12,  $\text{C}$ -20;  $\text{H}_2$ -13/ $\text{C}$ -20,  $\text{C}$ -11,  $\text{C}$ -12 and  $\text{H}_2$ -14 with  $\text{C}$ -12. Thus, the structure of **1**, with nine degrees of unsaturation, has been established.

**Table 1.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopic data [500 and 125 MHz,  $\text{CDCl}_3$ ] of compounds **1** and **2**.

No.	1		2	
	$\delta_{\text{C}}$ , Type	$\delta_{\text{H}}$ (J in Hz)	$\delta_{\text{C}}$ , Type	$\delta_{\text{H}}$ (J in Hz)
1	37.6, CH	3.39, m	39.7, CH	2.46, dddd (2.2, 2.5, 9.5, 11.7)
2	32.9, $\text{CH}_2$	3.15, m	32.7, $\text{CH}_2$	a: 2.97, dd (2.5, 14.8) b: 3.19, dd (11.9, 15.1)
3	161.3, C	-	162.1, C	-
4	123.4, C	-	122.7, C	-
5	106.5, CH	6.70, s	106.5, CH	6.80, s
6	156.0, C	-	154.3, C	-
7	74.0, CH	5.29, s	75.6, CH	4.59, s
8	74.3, C	-	73.6, C	-
9	40.5, $\text{CH}_2$	1.51, m 1.87, dd (5.2, 15.0)	43.0, CH	b: 1.89, dd (11.7, 14.8) a: 2.59, dd (4.2, 14.8)
10	74.3, CH	4.85, dd (5.2, 11.3)	78.4, CH	4.97, m
11	62.9, CH	3.73, s	148.6, C	5.86, s
12	60.6, C	-	136.2, C	-
13	22.5, $\text{CH}_2$	a: 1.45, m b: 2.44, dd (11.5, 14.7)	21.7, $\text{CH}_2$	a: 2.14, m b: 2.35, ddd (2.8, 11.9, 15.1)
14	30.4, $\text{CH}_2$	1.42, m 2.04, m	30.2, $\text{CH}_2$	1.60, ddd (2.8, 2.8, 15.1) 2.02, m
15	144.2, C	-	146.9, C	-
16	118.0, $\text{CH}_2$	5.37, s; 5.47, s	117.0, $\text{CH}_2$	5.19, s; 5.36, s
17	46.6, $\text{CH}_2$	4.18, br s	47.2, $\text{CH}_2$	4.12, br s
18	184.7, CH	9.89, s	184.3, CH	9.97, s
19	22.4, $\text{CH}_3$	1.35, s	19.6, $\text{CH}_3$	1.41, s
20	172.1, C	-	173.5, C	-



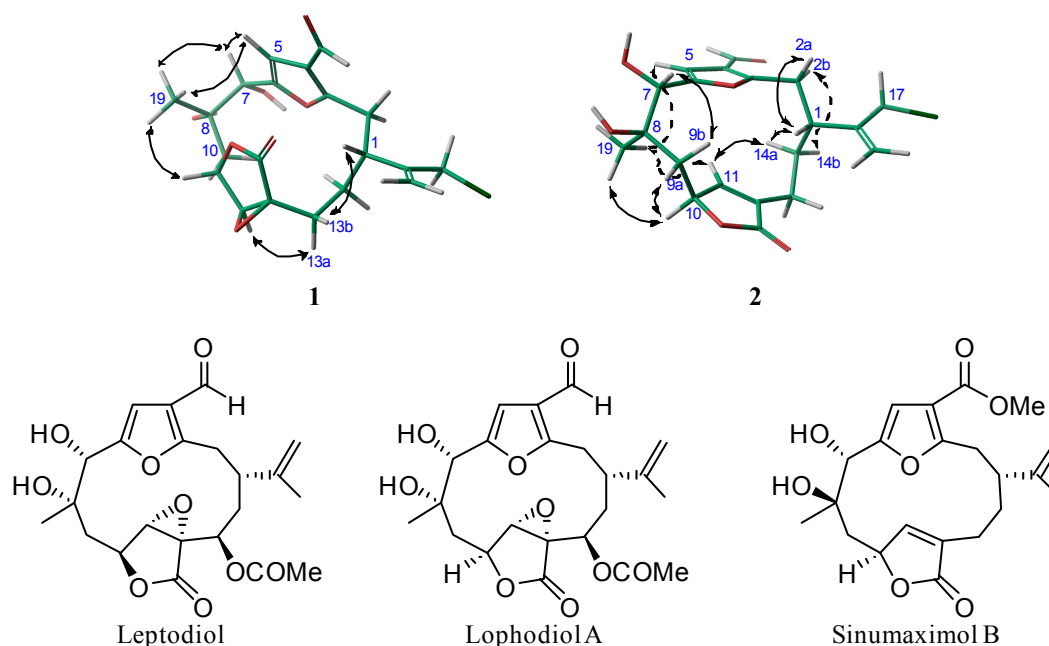
**1**

**Figure 2.**  $^1\text{H}$ - $^1\text{H}$ -COSY (—), HMBC (→) correlations of **1**.

Compound **2** was obtained as an oil whose EIMS spectrum showed a molecular ion at  $m/z$   $[M]^+$  394/396, with an isotopic pattern for a chlorine atom in the molecular formula  $C_{20}H_{23}ClO_6$  (HREIMS) ( $m/z$  394.1181  $[M]^+$ , calcd. for  $C_{20}H_{23}^{35}ClO_6$  394.1183). The  $^{13}C$  NMR spectrum and correlations in the HSQC spectrum indicated seven quaternary carbons, six methines, six methylenes and one methyl (Table 1). Absorptions for a hydroxyl group at  $3558\text{ cm}^{-1}$  and carbonyl group at  $1747\text{ cm}^{-1}$  were observed in its IR spectrum.

$^1H$  and  $^{13}C$  NMR data resemble those of **1**. The principal differences lie in the chemical shifts of H-7, H<sub>2</sub>-9, C-9 and C-19, which were:  $\delta_{H-7}$  4.59 (1H, s),  $\delta_{H-9}$  1.89 (1H, m); 2.59 (1H, dd);  $\delta_{C-9}$  43.0 and  $\delta_{C-19}$  19.6, compared with those of **1** (Table 1). These shift values suggested that compounds **1** and **2** differ in the configuration of C-7 and C-8. Also, their molecular formulas diverge by 16 amu of oxygen. This suggests that the  $\gamma$ -lactone ring system is devoid of the epoxide ring on **2**. This absence is confirmed by the new signals observed in the  $^1H$  and  $^{13}C$  NMR spectra for a proton ( $\delta_{H-11}$  5.86 (1H, s)) of a disubstituted olefin ( $\delta_{C-11}$  148.6;  $\delta_{C-12}$  136.2). Thus, the structure of **2** with nine degrees of unsaturation was established as shown in Figure 1 by COSY, HSQC and HMBC experiments.

The relative configurations of compounds **1** and **2** were ascertained by NOESY experiments, molecular mechanics [11], chemical shift studies and comparison of their spectroscopic data with those of previously described cembranoids, leptodiol [1], lophodiol A [12] and sinumaximol B [13] (Figure 3).



**Figure 3.** Selected NOE effects ( $\leftrightarrow$ ) of **1** and **2** and leptodiol, lophodiol A and sinumaximol B.

In compound **1**, the observed NOEs of H-5 with H-7 and H<sub>3</sub>-19, together with those of H-7 with H<sub>3</sub>-19 suggested that the adjacent hydroxyl groups at C-7–C-8 should be in a *cis* relationship. In compound **2**, the observed NOEs of H<sub>3</sub>-19 with H-5 and H-10 indicate that these protons and Me-19 must be on same side of the molecule, whereas the NOEs of H<sub>3</sub>-19 with H-9a and of H-7 with H-9b indicates that H-7 and Me-19, so the vicinal diols on C-7–C-8, have a *trans*-relationship. Therefore, the relative configuration of C-8 is opposite to that on compound **1**.

The configurations of C-7–C-8 vicinal diols were corroborated by comparison of the  $^1H$  and  $^{13}C$  NMR chemical shifts around the diol moiety C-7–C-8 of compounds **1** and **2** with those of the related diols leptodiol, lophodiol A and sinumaximol B, shown in Table 2. The chemical shifts of C-19 and H-7 of compounds **1** and **2** present strong differences ( $\Delta\delta_{C-19} = 2.8$  ppm and  $\Delta\delta_{H-7} = 0.7$ ). The chemical shifts of C-19 and H-7 of leptodiol ( $\delta_{C-19}$  22.7;  $\delta_{H-7}$  5.12) and lophodiol A ( $\delta_{C-19}$  22.7;  $\delta_{H-7}$  5.24), both with the C-7–C-8 diols in an  $\alpha$ -*cis*-relationship, are very similar to those of compound **1**. Whereas the

chemical shifts of C-19 and H-7 of sinumaximol B ( $\delta_{C-19}$  19.8;  $\delta_{H-7}$  4.52), whose C-7–C-8 diols show a *trans*-relationship, present strong differences (upfield  $\Delta\delta_{C-19} \approx 2.6$  ppm and downfield  $\Delta\delta_{H-7} \approx 0.8$ ) which are very similar to those presented by compound **2** in comparison to compound **1**.

In compound **1**, a striking  $^1\text{H}$  NMR signal is the singlet observed for H-11 ( $\delta$  3.73, s) due to the roughly  $90^\circ$  dihedral angle formed between H-10 and H-11. This results in a small  $J_{H-10, H-11}$  that confirms the relative configuration of C-10 and C-11 as represented in the energetically favourable conformation shown in Figure 3.

Finally, in compound **1**, the relative configuration of C-1 was secured by the NOESY correlation of H-13b with H-1 and H-13a with H-11, as shown in the 3D model in Figure 3. In compound **2**, the observed NOEs of H-1 with H-2a and H-14a, as well as of H-14a with H-11 and of H-2b with H-14b, indicate that the isopropenyl group of **2** is situated on the alpha side of the molecule. Therefore, both compounds belong to the furanocembranolide of the  $\alpha$ -series and their relative configurations are:  $1R^*$ ,  $7S^*$ ,  $8S^*$ ,  $10S^*$ ,  $11S^*$  and  $12S^*$  for compound **1** and  $1R^*$ ,  $7S^*$ ,  $8R^*$  and  $10S^*$  for compound **2**.

**Table 2.** Selected  $^1\text{H}$  and  $^{13}\text{C}$  NMR data [ $\text{CDCl}_3$ ] of leptodiol, lophodiol A, **1**, sinumaximol B and **2**.

No.	Leptodiol	Lophodiol A	<b>1</b>	Sinumaximol B	<b>2</b>
$\delta_{H-7}$	5.12, br s	5.24, s	5.29, s	4.52, s	4.59, s
$\delta_{H-9}$	1.61, dd (8.8, 14.5)	1.55, m	1.55, m	1.85, dd (11.5, 14.5)	1.89, dd (11.7, 14.8)
$\delta_{H-19}$	1.38, s	1.40 s	1.40, s	1.38, s	1.41, s
$\delta_{C-19}$	22.7	22.7	22.4	19.8	19.6
$\delta_{C-7}$	73.5	73.4	74.0	76.1	75.6
$\delta_{C-9}$	41.1	40.9	40.5	43.2	43.0

Compound **3** was obtained as an oil whose EIMS spectrum showed a peak at  $m/z$  362  $[\text{M}]^+$ , which corresponds to the molecular formula  $\text{C}_{20}\text{H}_{26}\text{O}_6$  (HREIMS) ( $m/z$  362.1737  $[\text{M}]^+$ , calcd. for  $\text{C}_{20}\text{H}_{26}\text{O}_6$  362.1729). These data are in agreement with the  $^{13}\text{C}$  NMR spectrum, which displayed correlations in the HSQC spectrum indicative of seven quaternary carbons, four methines, six methylenes and three methyls. Absorptions for a hydroxyl group at  $3475\text{ cm}^{-1}$  and carbonyl groups at  $1751$ ,  $1721$  and  $1701\text{ cm}^{-1}$  were observed in their IR spectrum. According to the degree of unsaturation given by the  $^{13}\text{C}$  NMR data, **3** must be a tricyclic compound.

In the  $^1\text{H}$  and  $^{13}\text{C}$  NMR experiments (Table 3), signals were found for:  $\alpha,\beta$ -unsaturated- $\gamma$ -lactone ring [ $\delta_{H-11}$  6.76 (1H, s),  $\delta_{C-11}$  155.9;  $\delta_{C-20}$  170.6;  $\delta_{C-10}$  90.5]; isopropenyl group [ $\delta_{H-16}$  4.68 (1H, br s),  $\delta_{H-16}$  4.70 (1H, br s);  $\delta_{C-16}$  109.7] and [ $\delta_{H-17}$  1.69 (3H, s);  $\delta_{C-17}$  20.9]; methyl group on a quaternary carbon bonded to oxygen [ $\delta_{H-19}$  1.37 ( $\text{CH}_3$ , s) and  $\delta_{C-8}$  74.3]; oximethine proton bonded to a secondary carbon [ $\delta_{H-5}$  3.31 (1H, d, 10.4 Hz) and  $\delta_{C-5}$  60.8] and two carbonyls [ $\delta_{C-3}$  215.4 and  $\delta_{C-6}$  203.5].

$^1\text{H}$ - $^1\text{H}$ -COSY experiments established two spin systems:  $\text{H}_2$ -2– $\text{H}_2$ -13 (fragment **I**) and  $\text{H}_3$ -18– $\text{H}$ -5 (fragment **II**) (Figure 4). The HMBC correlations of  $\text{H}_2$ -16/C-1, C-17 and  $\text{H}_3$ -17/C-16, C-15, C-1 locates the isopropenyl group at C-1 in fragment **I**, whereas the HMBC correlations of  $\text{H}_2$ -2 and  $\text{H}_3$ -18 with C-3 connects fragments **I** and **II** through C-3. The HMBC correlations  $\text{H}_3$ -19/ C-7, C-8, C-9, along with those of H-5 and  $\text{H}_2$ -7 with C-6, allowed us to extend fragment **II** by connecting it with substructure **III** through C-6. Both ends of the fragment **I** and substructure **III** (C-6–C-9) are connected by inserting an  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone bonded to C-13 and C-9, respectively, due to the HMBC correlations  $\text{H}_2$ -13/C-20, C-11, C-12 and those of H-11/C-10 and  $\text{H}_2$ -9/C-10, C-11. The quaternary feature of C-10 comes from the oxygen linkage between C-5 and C-10 supported by an HMBC correlation of H-5 with C-10. Therefore, the tricyclic structure of **3** has been established as depicted in Figure 4.

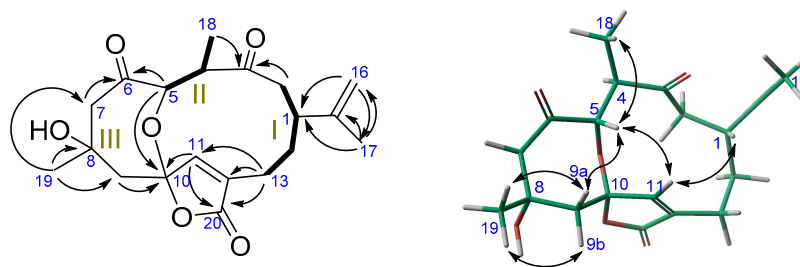


Figure 4.  $^1\text{H}$ - $^1\text{H}$ -COSY (—), HMBC (---) and selected NOE effects ( $\leftrightarrow$ ) of **3**.

Table 3.  $^1\text{H}$  and  $^{13}\text{C}$  NMR Spectroscopic data [500 and 125 MHz,  $\text{CDCl}_3$ ] of compounds **3**, **4** and **5**.

No.	3		4		5	
	$\delta_{\text{C}}$ , Mult.	$\delta_{\text{H}}$ (J in Hz)	$\delta_{\text{C}}$ , Mult.	$\delta_{\text{H}}$ (J in Hz)	$\delta_{\text{C}}$ , Mult.	$\delta_{\text{H}}$ (J in Hz)
1	39.7 CH	2.64, ddd (4.2, 7.1, 7.1)	35.4 CH	2.90, m	45.7, CH	2.70, m
2	49.7, $\text{CH}_2$	2.44, m 2.46, m	49.9, $\text{CH}_2$	2.53 m 2.53, m	32.3, $\text{CH}_2$	3.12, dd (6.6, 14.2) 3.22, dd (8.5, 14.2)
3	215.4, C	-	215.3, C	-	166.6, C	-
4	41.9, CH	2.89, ddd (6.6, 6.6, 10.6)	41.7, CH	2.90, m	116.7, C	-
5	60.8, CH	3.31, d (10.4)	60.9, CH	3.31, d (10.3)	122.0, CH	7.44, s
6	203.5, C	-	205.8, C	-	150.6, C	-
7	55.9, $\text{CH}_2$	2.68, m	55.9, $\text{CH}_2$	2.69, m	177.1, CH	9.54, s
8	74.3, C	-	74.3, C	-	204.3, C	-
9	45.6, $\text{CH}_2$	b: 1.92, m a: 2.34, d (15.1)	45.6, $\text{CH}_2$	b: 1.90, dd (2.6, 15.4) a: 2.32, d (15.1)	46.5, $\text{CH}_2$	2.63, dd (7.3, 17.7) 2.99, dd (6.7, 17.7)
10	90.5, C	-	90.3, C	-	76.6, CH	5.27, m
11	155.9, CH	6.76, s	156.0, CH	6.79, s	147.7, CH	7.11, m
12	131.4, C	-	131.1, C	-	134.3, C	-
13	24.0, $\text{CH}_2$	a: 2.27, ddd (5.4, 9.3, 14.3) b: 2.50, m	22.8, $\text{CH}_2$	a: 2.28, m b: 2.49, m	23.1, $\text{CH}_2$	2.16, m 2.25, m
14	31.7, $\text{CH}_2$	1.89, m	31.5, $\text{CH}_2$	2.00, m	30.1, $\text{CH}_2$	1.64, m 1.72, m
15	149.4, CH	-	149.3, C	-	144.8, C	-
16	109.7, $\text{CH}_2$	4.68, br s; 4.70, br s	114.7, $\text{CH}_2$	5.00, s; 5.20, dd (0.9, 0.9)	113.6, $\text{CH}_2$	4.64, br s 4.73, dd (1.6, 1.6)
17	20.9, $\text{CH}_3$	1.69, s	47.6, $\text{CH}_2$	4.03, m	18.0, $\text{CH}_3$	1.68, s
18	17.2, $\text{CH}_3$	1.04, d (6.6)	17.3, $\text{CH}_3$	1.04, d (6.6)	162.9, C	-
19	30.1, $\text{CH}_3$	1.37, s	30.1, $\text{CH}_3$	1.37, s	30.5, $\text{CH}_3$	2.21, s
20	170.6, C	-	171.6, C	-	172.9, C	-
OMe	-	-	-	-	51.8, $\text{CH}_3$	3.86, s

Compound **4** was obtained as an oil whose EIMS spectrum showed peaks at  $m/z$  396/398  $[\text{M}]^+$ , which correspond to the formula  $\text{C}_{20}\text{H}_{25}\text{ClO}_6$  (HREIMS) ( $m/z$  396.1324  $[\text{M}]^+$ , calcd. for  $\text{C}_{20}\text{H}_{25}\text{O}_6^{35}\text{Cl}$ , 396.1340). Considering the HSQC correlations, signals observed in the  $^{13}\text{C}$  NMR spectrum indicate seven quaternary carbons, four methines, seven methylenes and two methyls. Absorptions for a hydroxyl group at  $3420\text{ cm}^{-1}$  and carbonyl groups at  $1727$ ,  $1690$  and  $1647\text{ cm}^{-1}$  were observed in their IR spectrum.

$^1\text{H}$  and  $^{13}\text{C}$  NMR data (Table 3) resemble those of **3**. The molecular formula of **4** showed that one proton of **3** is substituted by a chlorine atom in **4**. Its corresponding data from both  $^1\text{H}$  and  $^{13}\text{C}$  NMR reveal that the substitution is situated on the isopropenyl appendage, where the methyl group of **3** changed to chloromethylene in **4**. This substitution was confirmed by the fragment ( $m/z$  321.1335

$[M - C_3H_4Cl]^+$ , calcd. for  $C_{17}H_{21}O_6$ , 321.1338) observed in HREIMS. The planar structure of **4** was confirmed as the 17-chloro derivative of compound **3** by COSY, HSQC and HMBC experiments.

NOESY experiments, studies of coupling constants and molecular mechanics calculations suggest that **3** and **4** have the same relative stereochemistry (Figure 1). In both compounds, NOEs were observed for H-5 with H-9a and H-11 and for H<sub>3</sub>-19 with H-9a and H-9b, defining a relative configuration for C-5 and C-10 and establishing H<sub>3</sub>-19 on C-8 in a *pseudo*-equatorial disposition and therefore coplanar to H-5. Also, the large coupling constants of H-5 ( $J = 10.4$  Hz) in **3** and H-5 ( $J = 10.3$  Hz) in **4** indicate that H-5 and H-4 are *trans*-diaxial, as the observed NOE between H-5 and H<sub>3</sub>-18 corroborates. Finally, the NOE observed between H-11 and H-1 places the isopropenyl group on the opposite side to the Me-18. Therefore, the overall relative configuration for **3** and **4** should be  $1R^*$ ,  $4R^*$ ,  $5S^*$ ,  $8R^*$  and  $10S^*$ .

Compound **5** was obtained as an oil with an EIMS spectrum peak at  $m/z$  388  $[M]^+$ , which corresponds to the molecular formula  $C_{21}H_{24}O_7$  (HREIMS) ( $m/z$  388.1524  $[M]^+$ , calcd. for  $C_{21}H_{24}O_7$ , 388.1522). These data are in agreement with the  $^{13}C$  NMR spectrum, which displayed correlations in the HSQC spectrum indicating eight quaternary carbons, five methines, five methylenes and three methyls (Table 3). Absorptions for carbonyl groups at 1655, 1650 and 1638  $cm^{-1}$  were noted in the IR spectrum.

In addition to the  $^1H$  and  $^{13}C$  NMR data registered for an isopropenyl group, a furan ring and a  $\alpha,\beta$ -unsaturated- $\gamma$ -lactone, other notable key signals were detected for the following functional groups: aldehyde [ $\delta_{H-7}$  9.54 (1H, s),  $\delta_{C-7}$  177.1]; methyl ketone [ $\delta_{H-19}$  2.21 (3H, s),  $\delta_{C-8}$  204.3] and methyl ester [ $\delta_{H-21}$  3.86 (3H, s),  $\delta_{C-18}$  162.9]. The 21 carbon atoms given by the molecular formula suggested that the isoprenic methyl group C-18 of a regular furanocembranolide is oxidized to a methyl ester and that the methyl-ketone and the aldehyde might come from the oxidative cleavage of the C-7–C-8 bond. According to the connectivity information from experiments COSY, HSQC and HMBC, compound **5** should be a *seco*-furanocembranolide.

$^1H$ - $^1H$ -COSY experiments established two spin systems: H<sub>2</sub>-2–H<sub>2</sub>-13 (fragment I) and H<sub>2</sub>-9–H-10 (fragment II) (Figure 5). The HMBC correlations of H<sub>2</sub>-16/C-1, C-17 and H<sub>3</sub>-17/C-16, C-15, C-1 locate the isopropenyl group at C-1 of fragment I and also those of H-5/C-3, C-4, C-6 and H<sub>2</sub>-2 with C-3 connected fragment I at the furan ring. The HMBC correlations of H<sub>2</sub>-13/C-11, C-12, C-20 and H-11/C-10 and H<sub>2</sub>-9/C-11 showed that fragments I and II are linked through an  $\alpha,\beta$ -unsaturated- $\gamma$ -lactone. HMBC correlations of H<sub>3</sub>-19/C-8, C-9 allowed us to place the methyl-ketone. Finally, the aldehyde must be located at C-6, in good agreement with the chemical shift observed for the aldehyde group of *seco*-bipinnatin J ( $\delta_{H-7}$  9.52 (1H, s),  $\delta_{C-7}$  177.5), the only *seco*-derivative [9] isolated from *Pseudopterogorgia*, which showed the same scission pattern.

Compound **5** is the first *seco*-furanocembranolide isolated from genus *Leptogorgia*. *Z*-deoxypukalide [1], also isolated in this work, can be considered a biogenetic precursor of **5** by oxidative cleavage of the corresponding  $\Delta^{7,8}$ . Since *Z*-deoxypukalide belongs to the  $\alpha$ -cembranolide series, we assign the same relative configurations  $1R^*$ ,  $10S^*$  to **5**.

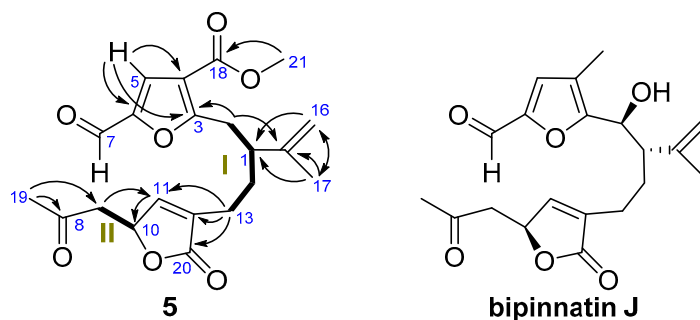


Figure 5.  $^1H$ - $^1H$ -COSY (—) and HMBC (---) correlations of **5** and bipinnatin J.

It should be expected that compounds **1–5** belong to the same enantiomeric series as *Z*-deoxypukalide, (**6**) whose absolute configuration we have previously determined using an NMR-based method using Pirkle's reagent [1].

#### Activation of Pancreatic Beta-Cell Proliferation

Several strategies have been proposed to recover functional beta-cell mass loss in diabetes mellitus onset; one of them is to activate beta-cell proliferation [3]. In previous work, we showed that furanocembranolides such as epoxy-pukalide, pukalide, *Z*-deoxypukalide and leptolide augment beta-cell proliferation [4–6]. In order to acquire detailed knowledge of the proliferation effect induced by furanocembranolides, compounds **1**, **2** and rubifolide (**7**) were used to treat synchronized INS-1 cells and proliferation was then measured. INS-1 cells were preincubated with 0.1  $\mu$ M of each product and proliferation was measured by BrdU incorporation (Table 4), showing a 2–3-fold increase in proliferation. Although it is difficult to reach a conclusion regarding the functional groups that could modulate this proliferation activity, these results also support chloro-furanocembranolides being potential activators of pancreatic beta-cell proliferation.

Furthermore, synchronized INS-1 cells were treated with the *nor*-cembranolide, scabrolide D (**8**), (Table 4), showing a  $2.8 \pm 0.69$ -fold change above untreated cells (1.0). We therefore consider it of interest in searching for compounds of the furanocembranolide and *nor*-1,4-diketocembranolide families, in order to develop a new class of antidiabetic agents.

**Table 4.** Beta-cell proliferation measurement after treating INS-1 beta-cells with each compound.

Compound (0.1 $\mu$ M)	Proliferation Ratio <sup>a</sup>
<b>1</b>	$2.5 \pm 0.65$
<b>2</b>	$2.0 \pm 0.61$
Rubifolide ( <b>7</b> )	$3.3 \pm 0.80$ <sup>b</sup>
Scabrolide D ( <b>8</b> )	$2.8 \pm 0.69$ <sup>b</sup>

<sup>a</sup> Fold change above untreated cells (1.0). A threshold over 1.5-fold was considered to increase proliferation.

<sup>b</sup>  $p < 0.05$  versus control.

### 3. Experimental Section

#### 3.1. General Experimental Procedures

Optical rotations were measured on a Perkin-Elmer model 343 Plus polarimeter (Perkin-Elmer, Rodgau, Germany) using a Na lamp at 20 °C. IR spectra were recorded on a Perkin-Elmer 1650/FTIR spectrometer (Perkin-Elmer, Rodgau, Germany). <sup>1</sup>H NMR and <sup>13</sup>C NMR, HSQC, HMBC and COSY spectra were measured employing a Bruker AMX 500 instrument (Bruker, Karlsruhe, Germany) operating at 500 MHz for <sup>1</sup>H NMR and at 125 MHz for <sup>13</sup>C NMR. All <sup>13</sup>C and <sup>1</sup>H NMR spectra were internally referenced to the residual solvent signal (CDCl<sub>3</sub>:  $\delta_C$  77.0 ppm,  $\delta_H$  7.25 ppm). Two-dimensional NMR spectra were obtained using the standard Bruker software (TOpSpin 2.1, Bruker, Karlsruhe, Germany). The EIMS data were obtained on a Waters Vg-Micromass spectrometer (Waters, Manchester, UK) model Zab 2F. HPLC separations were performed on an Agilent 1200 Series Quaternary LC system (Agilent Technologies, Waldbronn, Germany) apparatus equipped with a UV detector (DAD G1315D, Agilent Technologies, Waldbronn, Germany) and an Ascentis<sup>®</sup> C18 semi-preparative column (5  $\mu$ m, 25 cm  $\times$  21.2 mm, Supelco, Bellefonte, PA, USA) eluted with CH<sub>3</sub>CN-H<sub>2</sub>O mixtures. Size-exclusion chromatography used Sephadex LH-20 as stationary phase and hexane-MeOH-CH<sub>2</sub>Cl<sub>2</sub> (3:1:1) as solvent system. The spray reagent used to develop TLC plates was H<sub>2</sub>SO<sub>4</sub>-H<sub>2</sub>O-AcOH (1:4:20).



### 3.2. Collection, Extraction and Isolation

*Leptogorgia* sp. was collected by SCUBA diving off Aleta (Panama) at –10 m. A voucher specimen has been deposited at the Smithsonian Tropical Research Institute (Panama City, Panama) with code 200708. Specimens of *Leptogorgia* sp. (458.3 g) were extracted with acetone at room temperature and were concentrated to give a dark gum (10.1 g). C-18 reversed-phase flash chromatography of the crude extract gave fractions 4 (290.4 mg; 2:3 H<sub>2</sub>O/MeOH) and 5 (1119.1 mg; 1:4 H<sub>2</sub>O/MeOH) containing cembranolides, as indicated by their <sup>1</sup>H NMR spectra. Fraction 4 was further chromatographed by molecular exclusion LH-20 to give two sub-fractions of interest, 4<sub>1</sub> (39.4 mg) and 4<sub>5</sub> (18.6 mg). C-18 reversed-phase HPLC of 4<sub>1</sub> using a gradient from H<sub>2</sub>O-CH<sub>3</sub>CN (7:3) to CH<sub>3</sub>CN (100%) afforded compound 1 (1.3 mg, t<sub>R</sub> 61 min) and scabrolide D (8) (6.5 mg, t<sub>R</sub> 64 min). From sub-fraction 4<sub>5</sub>, compounds 2 (5.4 mg; t<sub>R</sub> 101.5 min) and 4 (0.5 mg; t<sub>R</sub> 83 min) were separated after C-18 reversed-phase HPLC using a gradient from H<sub>2</sub>O-CH<sub>3</sub>CN (7:3) to CH<sub>3</sub>CN (100%). Fraction 5 was chromatographed by molecular exclusion LH-20 to give five sub-fractions of interest: 5<sub>1</sub> (87.6 mg), 5<sub>2</sub> (89.5 mg), 5<sub>3</sub> (82.9 mg), 5<sub>4</sub> (59.1 mg) and 5<sub>5</sub> (50.3 mg). All these fractions were chromatographed separately by C-18 reversed-phase HPLC using a gradient from H<sub>2</sub>O-CH<sub>3</sub>CN (7:3) to CH<sub>3</sub>CN (100%), to afford compounds 3 (1.6 mg; t<sub>R</sub> 83.1 min), 5 (2.1 mg; t<sub>R</sub> 52.3 min), *Z*-deoxyypukalide (6) (5.7 mg; t<sub>R</sub> 111.2 min), rubifolide (7) (6.0 mg; t<sub>R</sub> 74 min) and epoxylophodione (9) (0.4 mg; t<sub>R</sub> 82.3 min).

**Compound 1:** Colourless oil;  $[\alpha]_D^{20}$  –20.7 (c 0.03, CH<sub>2</sub>Cl<sub>2</sub>); IR (film)  $\nu_{\max}$  3476, 2932, 1776, 1676 cm<sup>–1</sup>; <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.35 (3H, s, H-19), 1.42 (1H, m, H-14a), 1.45 (1H, m, H-13a), 1.51 (1H, m, H-9), 1.87 (1H, dd, *J* = 5.2, 15.0 Hz, H-9), 2.04 (1H, m, H-14b), 2.44 (1H, dd, *J* = 11.5, 14.7 Hz, H-13b), 3.15 (2H, m, H-2), 3.39 (1H, m, H-1), 3.73 (1H, s, H-11), 4.18 (2H, br s, H-17), 4.85 (1H, dd, *J* = 5.2, 11.3 Hz, H-10), 5.29 (1H, s, H-7), 5.37 (1H, s, H-16a), 5.47 (1H, s, H-16b), 6.70 (1H, s, H-5), 9.89 (1H, s, H-18); <sup>13</sup>C NMR (125 MHz CDCl<sub>3</sub>)  $\delta$  22.4 (CH<sub>3</sub>, C-19), 22.5 (CH<sub>2</sub>, C-13), 30.4 (CH<sub>2</sub>, C-14), 32.9 (CH<sub>2</sub>, C-2), 37.6 (CH, C-1), 40.5 (CH<sub>2</sub>, C-9), 46.6 (CH<sub>2</sub>, C-17), 60.6 (C, C-12), 62.9 (CH, C-11), 74.0 (CH, C-7), 74.3 (C, C-8), 74.3 (CH, C-10), 106.5 (CH, C-5), 118.0 (CH<sub>2</sub>, C-16), 123.4 (C, C-4), 144.2 (C, C-15), 156.0 (C, C-6), 161.3 (C, C-3), 172.0 (C, C-20), 184.7 (CH, C-18); EIMS *m/z* 409/411 [M – 1]<sup>+</sup>, 395/397 [M – CH<sub>3</sub>]<sup>+</sup>, 393/395 [M – OH]<sup>+</sup>, 355 [M – C<sub>3</sub>H<sub>4</sub>Cl]<sup>+</sup>; HREIMS *m/z* [M – 1]<sup>+</sup> 409.1041 (calcd. for C<sub>20</sub>H<sub>22</sub><sup>35</sup>ClO<sub>7</sub> 409.1054), 411.1027 (calcd. for C<sub>20</sub>H<sub>22</sub><sup>37</sup>ClO<sub>7</sub>, 411.1025), 395.0887 (calcd. for C<sub>19</sub>H<sub>20</sub><sup>35</sup>ClO<sub>7</sub>, 395.0898), 395.1069 (calcd. for C<sub>20</sub>H<sub>22</sub><sup>37</sup>ClO<sub>6</sub>, 395.1075).

**Compound 2:** Colourless oil;  $[\alpha]_D^{20}$  –11.0 (c 0.10, CH<sub>2</sub>Cl<sub>2</sub>); IR (film)  $\nu_{\max}$  3558, 2935, 1747, 1674 cm<sup>–1</sup>; <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.41 (3H, s, H-19), 1.60 (1H, ddd, *J* = 2.8, 2.8, 15.1 Hz, H-14a), 1.89 (1H, dd, *J* = 11.7, 14.8 Hz, H-9b), 2.02 (1H, m, H-14b), 2.14 (1H, m, H-13a), 2.35 (1H, ddd, *J* = 2.8, 11.9, 15.1 Hz, H-13b), 2.46 (1H, dddd, *J* = 2.2, 2.5, 9.5, 11.7 Hz, H-1), 2.59 (1H, dd, *J* = 4.1, 14.8 Hz, H-9a), 2.97 (1H, dd, *J* = 2.5, 14.8 Hz, H-2a), 3.19 (1H, dd, *J* = 11.9, 15.1 Hz, H-2b), 4.12 (2H, br s, H-17), 4.59 (1H, s, H-7), 4.97 (1H, m, H-10), 5.19 (1H, s, H-16), 5.36 (1H, s, H-16), 5.86 (1H, s, H-11), 6.80 (1H, s, H-5), 9.97 (1H, s, H-18); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  19.6 (CH<sub>3</sub>, C-19), 21.7 (CH<sub>2</sub>, C-13), 30.2 (CH<sub>2</sub>, C-14), 32.7 (CH<sub>2</sub>, C-2), 39.7 (CH, C-1), 43.0 (CH<sub>2</sub>, C-9), 47.2 (CH<sub>2</sub>, C-17), 73.6 (C, C-8), 75.6 (CH, C-7), 78.4 (CH, C-10), 106.5 (CH, C-5), 117.0 (CH<sub>2</sub>, C-16), 122.7 (C, C-4), 136.2 (C, C-12), 146.9 (C, C-15), 148.6 (CH, C-11), 154.3 (C, C-6), 162.1 (C, C-3), 173.5 (C, C-20), 184.3 (CH, C-18); EIMS *m/z* 394/396 [M]<sup>+</sup>, 377/379 [M – OH]<sup>+</sup>; HREIMS *m/z* [M]<sup>+</sup> 394.1181 (calcd. for C<sub>20</sub>H<sub>23</sub>O<sub>6</sub><sup>35</sup>Cl, 394.1183), 396.1162 (calcd. for C<sub>20</sub>H<sub>23</sub>O<sub>6</sub><sup>37</sup>Cl, 396.1154), 377.1146 (calcd. for C<sub>20</sub>H<sub>22</sub>O<sub>5</sub><sup>35</sup>Cl, 377.1156), 379.1133 (calcd. for C<sub>20</sub>H<sub>22</sub>O<sub>5</sub><sup>37</sup>Cl, 379.1126).

**Compound 3:** Colourless oil;  $[\alpha]_D^{20}$  +10.0 (c 0.16, CH<sub>2</sub>Cl<sub>2</sub>); IR (film)  $\nu_{\max}$  3475, 2933, 1751, 1721, 1701 cm<sup>–1</sup>; <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.04 (3H, d, *J* = 6.6 Hz, H-18), 1.37 (3H, s, H-19), 1.69 (3H, s, C-17), 1.89 (2H, m, H-14), 1.92 (1H, m, H-9b), 2.27 (2H, ddd, *J* = 5.4, 9.3, 14.3 Hz, H-13a), 2.34 (1H, d, *J* = 15.1 Hz, H-9a), 2.44 (1H, m, H-2a), 2.46 (1H, m, H-2b), 2.50 (1H, m, H-13b), 2.64 (1H, ddd, *J* = 4.2, 7.1, 7.1 Hz, H-1), 2.68 (2H, m, H-7), 2.89 (1H, ddd, *J* = 6.6, 6.6, 10.6 Hz, H-4), 3.31 (1H, d, *J* = 10.4 Hz, H-5), 4.68 (1H, br s, H-16a), 4.70 (1H, br s, H-16b), 6.76 (1H, s, H-11); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  17.2 (CH<sub>3</sub>, C-18), 20.9 (CH<sub>3</sub>, C-17), 24.0 (CH<sub>2</sub>, C-13), 30.1 (CH<sub>3</sub>, C-19), 31.7 (CH<sub>2</sub>, C-14), 39.7 (CH, C-1), 41.9 (CH, C-4),

45.6 (CH<sub>2</sub>, C-9), 49.7 (CH<sub>2</sub>, C-2), 55.9 (CH<sub>2</sub>, C-7), 60.8 (CH, C-5), 74.3 (C, C-8), 90.5 (C, C-10), 109.7 (CH<sub>2</sub>, C-16), 131.4 (C, C-12), 149.4 (C, C-15), 155.9 (CH, C-11), 170.5 (C, C-20), 203.5 (C, C-6), 215.4 (C, C-3); EIMS *m/z* 362 [M]<sup>+</sup>, 347 [M – CH<sub>3</sub>]<sup>+</sup>, 345 [M – OH]<sup>+</sup>; HREIMS *m/z* [M]<sup>+</sup> 362.1737 (calcd. for C<sub>20</sub>H<sub>26</sub>O<sub>6</sub>, 362.1729), [M – CH<sub>3</sub>]<sup>+</sup> 347.1509 (calcd. for C<sub>19</sub>H<sub>23</sub>O<sub>6</sub>, 347.1495), 345.1708 (calcd. for C<sub>20</sub>H<sub>25</sub>O<sub>5</sub>, 345.1702).

Compound 4: Colourless oil; [ $\alpha$ ]<sub>D</sub><sup>20</sup> +66.0 (c 0.05, CH<sub>2</sub>Cl<sub>2</sub>); IR (film)  $\nu_{\max}$  3420, 2956, 1727, 1690, 1647 cm<sup>-1</sup>; <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.04 (3H, d, *J* = 6.6 Hz, H-18), 1.37 (3H, s, H-19), 1.90 (1H, dd, *J* = 2.6, 15.4 Hz H-9b), 2.00 (2H, m, H-14), 2.28 (1H, m, H-13a), 2.32 (1H, d, *J* = 15.1 Hz, H-9b), 2.49 (1H, m, H-13b), 2.53 (2H, m, H-2), 2.69 (2H, m, H-7), 2.90 (1H, m, H-1), 2.90 (1H, m, H-4), 3.31 (1H, d, *J* = 10.3 Hz, H-5), 4.03 (2H, m, H-17), 5.00 (1H, s, H-16b), 5.20 (1H, dd, *J* = 0.9, 0.9 Hz, H-16a), 6.79 (1H, s, H-11); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  17.3 (CH<sub>3</sub>, C-18), 22.8 (CH<sub>2</sub>, C-13), 30.1 (CH<sub>3</sub>, C-19), 31.5 (CH<sub>2</sub>, C-14), 35.4 (CH, C-1), 41.7 (CH, C-4), 45.6 (CH<sub>2</sub>, C-9), 47.6 (CH<sub>2</sub>, C-17), 49.9 (CH<sub>2</sub>, C-2), 55.9 (CH<sub>2</sub>, C-7), 60.9 (CH, C-5), 74.3 (C, C-8), 90.3 (C, C-10), 114.7 (CH<sub>2</sub>, C-16), 131.1 (C, C-12), 149.3 (C, C-15), 156.0 (CH, C-11), 171.6 (C, C-20), 205.8 (C, C-6), 215.3 (C, C-3); EIMS *m/z* 396/398 [M]<sup>+</sup>, 378/380 [M – H<sub>2</sub>O]<sup>+</sup>, 361 [M – Cl]<sup>+</sup>, 321 [M – C<sub>3</sub>H<sub>4</sub>Cl]<sup>+</sup>; HREIMS *m/z* [M]<sup>+</sup> 396.1324 (calcd. for C<sub>20</sub>H<sub>25</sub>O<sub>6</sub><sup>35</sup>Cl, 396.1340), 398.1315 (calcd. for C<sub>20</sub>H<sub>25</sub>O<sub>6</sub><sup>37</sup>Cl, 398.1310), 361.1642 (calcd. for C<sub>20</sub>H<sub>25</sub>O<sub>6</sub>, 361.1651), 321.1335 (calcd. for C<sub>17</sub>H<sub>21</sub>O<sub>6</sub>, 321.1338).

Compound 5: Colourless oil; [ $\alpha$ ]<sub>D</sub><sup>20</sup> –2.11 (c 0.21, CH<sub>2</sub>Cl<sub>2</sub>); IR (film)  $\nu_{\max}$  2100, 1655, 1650, 1638 cm<sup>-1</sup>; <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.64 (1H, m, H-14), 1.68 (3H, s, H-17), 1.72 (1H, m, H-14), 2.16 (1H, m, H-13a), 2.21 (3H, s, H-19), 2.25 (1H, m, H-13b), 2.63 (1H, dd, *J* = 7.3, 17.7 Hz, H-9a), 2.70 (1H, m, H-1), 2.99 (1H, dd, *J* = 6.7, 17.7 Hz, H-9b), 3.12 (1H, dd, *J* = 6.6, 14.2 Hz, H-2), 3.22 (1H, dd, *J* = 8.5, 14.2 Hz, H-2), 3.86 (3H, s, H-21), 4.64 (1H, br s, H-16a), 4.73 (1H, dd, *J* = 1.6, 1.6 Hz, H-16b), 5.27 (1H, m, H-10), 7.11 (1H, m, H-11), 7.44 (1H, s, H-5), 9.54 (1H, s, H-7); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  18.0 (CH<sub>3</sub>, C-17), 23.1 (CH<sub>2</sub>, C-13), 30.1 (CH<sub>2</sub>, C-14), 30.5 (CH<sub>3</sub>, C-19), 32.3 (CH<sub>2</sub>, C-2), 45.7 (CH, C-1), 46.5 (CH<sub>2</sub>, C-9), 51.8 (CH<sub>3</sub>, C-21), 76.6 (CH, C-10), 113.6 (CH<sub>2</sub>, C-16), 116.7 (C, C-4), 122.0 (CH, C-5), 134.3 (C, C-12), 144.8 (C, C-15), 147.7 (CH, C-11), 150.6 (C, C-6), 162.9 (C, C-18), 166.6 (C, C-3), 172.9 (C, C-20), 177.1 (CH, C-7), 204.3 (C, C-8); EIMS *m/z* 388 [M]<sup>+</sup>, 357 [M – OCH<sub>3</sub>]<sup>+</sup>; ESMS *m/z* [M]<sup>+</sup> 388.1524 (calcd. for C<sub>21</sub>H<sub>24</sub>O<sub>7</sub>, 388.1522), 357.1341 (calcd. for C<sub>20</sub>H<sub>21</sub>O<sub>6</sub>, 357.1338).

### 3.3. Activation of Pancreatic Beta-Cell Proliferation

INS-1 cells (insulinoma rat cells) were seeded at a density of 20,000 cells per well in 96-well plates. Compounds **1**, **2**, rubifolide (**7**) and scabrolide (**8**) were assayed at a final concentration of 0.1  $\mu$ M in culture medium supplemented with 5.5 mM glucose. Proliferation was measured after 24 h using the BrdU kit (Roche, Mannheim, Germany), following manufacturer's instructions. At least three independent experiments in triplicate were performed per compound.

**Supplementary Materials:** The following are available online at [www.mdpi.com/1660-3397/16/2/49/s1](http://www.mdpi.com/1660-3397/16/2/49/s1), Figure S1: <sup>1</sup>H NMR of **1** in CDCl<sub>3</sub>; Figure S2: <sup>13</sup>C NMR spectrum of **1** in CDCl<sub>3</sub>; Figure S3: <sup>1</sup>H NMR of **2** in CDCl<sub>3</sub>; Figure S4: <sup>13</sup>C NMR spectrum of **2** in CDCl<sub>3</sub>; Figure S5: <sup>1</sup>H NMR of **3** in CDCl<sub>3</sub>; Figure S6: <sup>13</sup>C NMR spectrum of **3** in CDCl<sub>3</sub>; Figure S7: <sup>1</sup>H NMR of **4** in CDCl<sub>3</sub>; Figure S8: <sup>13</sup>C NMR spectrum of **4** in CDCl<sub>3</sub>; Figure S9: <sup>1</sup>H NMR of **5** in CDCl<sub>3</sub>; Figure S10: <sup>13</sup>C NMR spectrum of **5** in CDCl<sub>3</sub>.

**Acknowledgments:** This work was supported by the Ministerio de Ciencia e Innovación (SAF2009-0839 and RTA 2015-00010-C03-02). ARDM acknowledges funding from IMBRAIN project (FP7-REGPOT-2012-CT2012-31637-IMBRAIN) and from Cabildo de Tenerife (Agustín de Betancourt Programme). A.B.G. would like to thank Convenio Universidad de Magallanes (Chile) and CSIC, project 2009CL0031, for financial support.

**Author Contributions:** A.B.G., J.M.d.I.R., G.P. and I.C.-C. performed the experiments, analysed the data and contributed to discussion; L.D. contributed biological material; A.R.D.-M., J.D. and M.C. supervised experiments, analysed the data and wrote the manuscript. All authors approved the final version of this manuscript.

**Conflicts of Interest:** The authors declare no conflict of interest.

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