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## mGluR5 positive allosteric modulators facilitate both hippocampal LTP and LTD and enhance spatial learning

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### Abstract

Highly selective positive allosteric modulators (PAMs) of metabotropic glutamate receptor subtype 5 (mGluR5) have emerged as a potential approach to treat positive symptoms associated with schizophrenia. mGluR5 plays an important role in both long term potentiation (LTP) and

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### Disclosure/Conflict of Interest

#### P. Jeffrey Conn, Ph.D.

Dr. Conn has received compensation over the past two years as a consultant from: Merck and Co., Johnson and Johnson, Hoffman La Roche, GlaxoSmithKline, Lundbeck Research USA, Epix Pharmaceuticals, Invitrogen Life Technologies, Evotec Inc., Addex Pharmaceuticals, Michael J. Fox Foundation, Seaside Therapeutics, Cephalon Inc., AstraZeneca USA, NeurOp Inc., Forest Research Institute, LEK Consulting, The Frankel Group, Prestwick Chemical Co., Millipore Corp., Genentech, IMS Health, Primary Insight, and Otsuka.

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#### Craig W. Lindsley, Ph.D.

Dr. Lindsley consults for Amgen and Eisai and receives research support that includes salary support from NIH, Michael J. Fox, and Seaside Therapeutics.

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long term depression (LTD), suggesting that mGluR5 PAMs may also have utility in improving impaired cognitive function. However, if mGluR5 PAMs shift the balance of LTP and LTD or induce a state in which afferent activity induces lasting changes in synaptic function that are not appropriate for a given pattern of activity, this could disrupt rather than enhance cognitive function. We determined the effect of selective mGluR5 PAMs on induction of LTP and LTD at the Schaffer collateral – CA1 synapse in the hippocampus. mGluR5-selective PAMs significantly enhanced threshold theta burst stimulation (TBS)-induced LTP. In addition, mGluR5 PAMs enhanced both DHPG-induced LTD and LTD induced by delivery of paired-pulse low frequency stimulation. Selective potentiation of mGluR5 had no effect on LTP induced by suprathreshold TBS or saturated LTP. The finding that potentiation of mGluR5-mediated responses to stimulation of glutamatergic afferents enhances both LTP and LTD supports the hypothesis that activation of mGluR5 by endogenous glutamate contributes to both forms of plasticity. Furthermore, two systemically active mGluR5 PAMs enhanced performance in the Morris water maze, a measure of hippocampus-dependent spatial learning. Discovery of small molecules that enhance both LTP and LTD in an activity-appropriate manner demonstrates a unique action on synaptic plasticity that may provide a novel approach for treatment of impaired cognitive function.

### Keywords

mGluR5; group I mGluR; synaptic plasticity; LTP; LTD; allosteric potentiator

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### Introduction

Schizophrenia is a complex disorder that includes positive, negative, and cognitive symptoms (Andreasen, 2000). Current medications are partially effective in treating positive symptoms but largely ineffective in treating cognitive deficits and negative symptoms. Based on this and serious adverse effects of available antipsychotic agents, there is critical need for new treatment strategies. Recently, the metabotropic glutamate receptor subtype 5 (mGluR5) has emerged as an exciting target proposed to have efficacy for treatment of schizophrenia (Marino and Conn, 2002; Conn et al., 2008). This hypothesis is based on evidence suggesting that enhanced signaling through the *N*-methyl-D-aspartate (NMDA) glutamate receptor may be antipsychotic and that mGluR5 and NMDARs are closely associated signaling partners in forebrain circuits. Activation of mGluR5 potentiates NMDAR function in multiple neuronal populations (Awad et al., 2000; Mannaioni et al., 2001; Pisani et al., 2001; Marino and Conn, 2002) and mGluR5 antagonists or genetic deletion potentiate the psychotomimetic effects of NMDAR antagonists (Henry et al., 2002; Kinney et al., 2003; Brody et al., 2004a; Brody et al., 2004b; Campbell et al., 2004; Homayoun et al., 2004; Lindsley et al., 2004; Kinney et al., 2005). Recently, we and others have discovered selective positive allosteric modulators (PAMs) for mGluR5 that have robust efficacy in animal models used to predict effectiveness in treating positive symptoms of schizophrenia (O'Brien et al., 2003; Lindsley et al., 2004; O'Brien et al., 2004; Kinney et al., 2005; Le Poul, 2005; de Paulis et al., 2006; Lindsley et al., 2006; Chen et al., 2007; Chen et al., 2008; Conn et al., 2008; Liu et al., 2008). These mGluR5 PAMs do not activate mGluR5 directly but act at an allosteric site to potentiate activation by glutamate (Chen and

Conn, 2008; Conn et al., 2009). These findings provide strong preclinical support for use of selective activators of mGluR5 as novel antipsychotic agents.

In addition to antipsychotic efficacy, mGluR5 PAMs have the potential to treat impaired cognition in schizophrenia patients. mGluR5 knockout mice have impaired NMDAR-mediated hippocampal long-term potentiation (LTP) and NMDA-dependent memory tasks (Lu et al., 1997; Jia et al., 1998). Also, the mGluR5 selective antagonist MPEP blocks theta burst stimulation (TBS)-induced LTP in area CA1 in hippocampal slices (Francesconi et al., 2004; Shalin et al., 2006) and *in vivo* (Manahan-Vaughan and Braunewell, 2005). Finally, the mGluR1/5 agonist, DHPG, primes LTP induction (Cohen et al., 1998; Raymond et al., 2000). In addition to regulation of LTP, DHPG induces an NMDAR-independent form of long-term depression (LTD) (Gasparini et al., 1999; Huber et al., 2001) and this response is absent in mGluR5 null mice and in hippocampal slices incubated with mGluR5 antagonists (Gasparini et al., 1999; Huber et al., 2001; Faas et al., 2002; Hou and Klann, 2004; Huang et al., 2004; Huang and Hsu, 2006).

In the simplest view, mGluR5 potentiation could enhance synaptic plasticity and thereby enhance some forms of cognitive function. However, previous studies of electrophysiological effects of mGluR5 PAMs relied on potentiation of exogenously applied agonists and it is not clear whether mGluR5 PAMs will enhance activation of mGluR5 by synaptically released glutamate. Thus, it is critical to determine whether mGluR5-selective PAMs enhance afferent stimulation-induced hippocampal LTP and LTD. Furthermore, if mGluR5 PAMs preferentially augment one form of synaptic plasticity and thereby disturb the LTP/LTD balance, this could disrupt rather than enhance cognitive function. Indeed, recent studies suggest that mutations associated with Fragile X Syndrome (FXS) selectively increase mGluR5-mediated hippocampal LTD (Huber et al., 2002; Bear et al., 2004; Nosyreva and Huber, 2006), while having no effect (Godfraind et al., 1996; Paradee et al., 1999; Li et al., 2002) or depressing hippocampal LTP (Lauterborn et al., 2007). This preferential enhancement of mGluR-LTD is thought to be a primary change contributing to cognitive disruption associated with FXS. Thus, we performed a series of studies to determine the effects of mGluR5 PAMs on hippocampal LTP and LTD.

We report that selective mGluR5 PAMs enhance afferent stimulation-induced LTP and LTD at the SC-CA1 synapse. Importantly, these compounds enhance both forms of synaptic plasticity while maintaining the normal patterns of presynaptic activity required to induce each, which may provide an ideal profile for agents that are used to improve some forms of cognitive function. In addition, we demonstrate that these compounds improve performance in a model of hippocampus-dependent spatial learning. This builds on studies suggesting that mGluR5 PAMs have potential utility as novel antipsychotic agents and provides direct support for the hypothesis that mGluR5 PAMs may also enhance hippocampal-dependent cognitive function.

## Methods

### Materials

4-Nitro-*N*-(1,3-diphenyl-1*H*-pyrazol-5-yl)benzamide (VU-29) was synthesized as outlined in detail by DePaulis et al. (2006). 5MPEP was synthesized as detailed by Rodriguez et al. (2005). CDPPB was synthesized as detailed by Lindsley et al. (2004). D-AP5, DHPG, glutamate, U0126, and PP 1 were purchased from Tocris (Ellisville, MO). ADX47273 was synthesized as outlined in the supplemental material.

### Animals

All animals used in these studies were cared for in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals. Experimental protocols were in accordance with all applicable guidelines regarding the care and use of animals. Animals were housed in an Association for Assessment and Accreditation of Laboratory Animal Care (AALAC) International approved facility with free access to food and water. All efforts were made to minimize animal suffering and to reduce the number of animals used.

### Measurement of Phosphoinositide Hydrolysis in Hippocampal Slices

Agonist-induced phosphoinositide (PI) hydrolysis was measured in rat hippocampal slices using a modification of the method outlined by (Berridge et al., 1982) as previously described (Berridge et al., 1982; Conn and Sanders-Bush, 1986). Briefly, cross-chopped (350  $\mu$ M  $\times$  350  $\mu$ M) slices of male (6–9 weeks) Sprague-Dawley rat hippocampus were incubated with 95% O<sub>2</sub>/5% CO<sub>2</sub> bubbled Krebs buffer (108 mM NaCl, 4.7 mM KCl, 1.2 mM MgSO<sub>4</sub>, 1.2 mM KH<sub>2</sub>PO<sub>4</sub>, 2.5 mM CaCl<sub>2</sub>, 25 mM NaHCO<sub>3</sub> and 10 mM Glucose). The tissue was allowed to recover for 30 minutes with shaking at 37°C. Following recovery, the tissue was combined, washed with warm Krebs buffer, and 25  $\mu$ L of gravity packed slices were incubated with 175  $\mu$ L Krebs containing 0.5  $\mu$ Ci [<sup>3</sup>H]myo-inositol for 45 minutes. VU-29 or vehicle controls were added and incubated for 15 minutes, followed by the addition of 10 mM LiCl and incubated for an additional 15 minutes. Finally, DHPG was added followed by an additional 45 minute incubation. The reaction was terminated by the addition of 900  $\mu$ L of chloroform:methanol (1:2). The aqueous and organic phases were separated by addition of 300  $\mu$ L chloroform and 300  $\mu$ L water, vortexing, and allowing the phases to separate by gravity. The aqueous phase was added to anion exchange columns (AG 1-X8 Resin, 100–200 mesh, formate form, BIO-RAD) and [<sup>3</sup>H]inositol phosphates were eluted and measured by liquid scintillation counting.

### Cell Culture

Secondary rat cortical astrocytes were prepared as described previously (Peavy et al., 2001; Zhang et al., 2005; Chen et al., 2007). In brief, neocortices from 2- to 4-day-old Sprague-Dawley rat pups were dissected and dissociated in DMEM by trituration with 1-ml pipette tips. The cells were then centrifuged and resuspended in DMEM [containing 1 mM sodium pyruvate, 2 mM L-glutamine, and PenStrep (100 units/ml penicillin and 0.1 mg/ml streptomycin; Invitrogen, Carlsbad, CA)] supplemented with 10% FBS in T75 tissue culture flasks; the medium was changed the next day. Cell cultures were maintained at 37°C in an

atmosphere of 95% air, 5% CO<sub>2</sub> for 6 to 8 days. Cells were shaken overnight (280–310 rpm) to remove oligodendrocytes and microgliaocytes. For ERK1/2 phosphorylation assay, the cells were then trypsinized and replated into poly-D-lysine-precoated 12-well plates at a density of about  $6 \times 10^6$  cells/well in full DMEM with 10% FBS. The second day, the medium was switched to full DMEM with G-5 supplement (Invitrogen) containing epidermal growth factor (10 ng/ml), basic fibroblast growth factor (5 ng/ml), insulin (5 µg/ml), and other factors. The cells were nearly confluent within 2 days and resembled the polygonal astrocytic appearance *in vivo*. Three days after the addition of G-5 supplement and 20 h before experiments, the medium was aspirated, and the cells were washed three times with 1x Hanks' balanced salt solution, and 1 ml of glutamine-free DMEM was added to each well.

### Cell Based Calcium Fluorescence Measurement

Rat mGluR1d was transfected into HEK293A cells using Lipofectamine as described previously (Chen et al., 2007). Rat mGluR2 and human mGluR4 were co-expressed with G<sub>qi5</sub>, which enables coupling to the calcium mobilization as previously described (Galici et al., 2006). Cells were loaded with calcium-sensitive dye according to the manufacturer's instructions (Calcium 3 kit; Molecular Devices, Sunnyvale, CA) after incubation in glutamate/glutamine-free medium (DMEM and 10% dialyzed fetal bovine serum) for 5 h. Compound A (1 ml) from Calcium 3 kit was dissolved in 20 ml of 1x Hanks' balanced salt solution (HBSS; Invitrogen) containing 2.5 mM probenecid (Sigma), adjusted to pH 7.4. Cells were loaded for 50 min at 37°C under an atmosphere of 5% carbon dioxide. Dye was then carefully removed, and cells were washed with HBSS containing probenecid. Cells were maintained in the same buffer at room temperature for the following assay. For calcium fluorescence measurement of rat cortical astrocytes, allosteric modulators were manually added 5 min before the addition of agonist. Agonist was added at a rate of 52 µl/s, and calcium flux was measured using Flexstation II (Molecular Devices) at 25°C. All of the peaks of the calcium response were normalized to the maximum response to a saturated dose of glutamate (10 µM). The submaximal concentration (EC<sub>20</sub> value) of glutamate was determined for each separate experiment, allowing for a response varying from 10 to 30% of the maximum peak.

### ERK1/2 Phosphorylation Assay

On the day of each assay, cells were first treated with CDPPB, and then stimulated with agonist. At the end of stimulation, medium containing the drug was aspirated, and 200 µl of ice-cold lysis buffer (containing 50 mM Tris-HCl, 50 mM NaCl, 5 mM EDTA, 10 mM EGTA, 1 mM Na<sub>3</sub>VO<sub>4</sub>, 2 mM Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>·10 H<sub>2</sub>O, 4 mM magnesium *para*-nitrophenyl phosphate, and 1 mM phenylmethylsulfonyl fluoride plus 10 µg/ml leupeptin and 2 µg/ml aprotinin) was added to each well. Cells were frozen at –80°C and underwent three thaw and refreeze cycles. Cells were scraped into clean tubes, the samples were centrifuged, and the supernatant was collected. Equal amounts of supernatant from each sample were mixed with 3x lithium dodecyl sulfate sample buffer, subjected to SDS-polyacrylamide gel electrophoresis, and transferred to nitrocellulose membranes. Membranes were first blocked and then stained with primary rabbit anti-p44/42 mitogen-activated protein kinase (ERK1/2) polyclonal antibody mixed with primary mouse anti-phospho-p44/42 mitogen-activated

protein kinase (phospho-ERK1/2) monoclonal antibody (Cell Signaling Technology Inc. Beverly, MA). After washing three times, membranes were subsequently stained with fluorescent dye Alexa Fluor 680-conjugated secondary goat anti-mouse IgG (H+L) (Invitrogen) mixed with fluorescent dye IRDye800-conjugated secondary goat anti-rabbit IgG (H+L) (Rockland, Gilbertsville, PA). Membranes were scanned using Odyssey Imaging System (LI-COR, Lincoln, NE). ERK1/2 phosphorylation (phosphorylated ERK1/2) is first normalized to total ERK1/2 and then expressed as percentage of maximal response or -fold above control.

### Extracellular Field Potential Recordings

Young adult (6–9 weeks) male Sprague-Dawley rats (Charles River, Wilmington, MA) were anesthetized with isoflurane, decapitated and the brains were quickly removed and submerged into ice cold cutting solution (in mM: 110 sucrose, 60 NaCl, 3 KCl, 1.25 NaH<sub>2</sub>PO<sub>4</sub>, 28 NaHCO<sub>3</sub>, 5 glucose, 0.6 (+)-sodium-L-ascorbate, 0.5 CaCl<sub>2</sub>, 7 MgCl<sub>2</sub>) continuously bubbled with 95% O<sub>2</sub>/5% CO<sub>2</sub>. The brains were then hemisected and 400 μm transverse slices were made using a vibratome (Leica VT100S). Individual hippocampi were removed from the slice and transferred to a room temperature mixture containing equal volumes of cutting solution and artificial cerebrospinal fluid (aCSF; in mM: 125 NaCl, 2.5 KCl, 1.25 NaH<sub>2</sub>PO<sub>4</sub>, 25 NaHCO<sub>3</sub>, 25 glucose, 2 CaCl<sub>2</sub>, 1 MgCl<sub>2</sub>) where they were allowed to equilibrate for 30 minutes. The hippocampi were then placed into an interface chamber perfused with oxygenated aCSF (1.5–2 mL/min) for at least 1.5 hrs at 31°C. Bipolar stimulating electrodes were placed in the stratum radiatum near the CA3-CA1 border in order to stimulate the Schaffer collaterals. Recording electrodes were pulled with a Flaming/Brown micropipette puller (Sutter Instruments, CA) to a resistance of 3–5MΩ, filled with aCSF and placed in the stratum radiatum of area CA1. Field potential recordings were acquired using either a Microelectrode AC Amplifier Model 1800 (A–M Systems) or Patch Clamp PC-505B (Warner Instruments) amplifier and pClamp 9.2 software. Input output curves were generated to determine the stimulus intensity that produced 40–50% of the maximum response prior to each experiment, which was used as the baseline stimulation. Baseline stimulation was applied at 0.05 Hz. Drugs were diluted to the appropriate concentrations in either DMSO (< 0.1%) or aCSF. Threshold LTP was induced by one train of theta burst stimulation (TBS) (nine bursts of four pulses at 100 Hz; 230 ms interburst interval). Saturated LTP was induced by 4 trains of 10 Hz TBS (nine bursts of four pulses at 100 Hz, 100 ms interburst interval). Chemically induced mGluR-LTD was induced by application of DHPG (25 – 75 μM) for 10 min. Synaptically evoked mGluR-LTD was induced by paired pulse low frequency stimulation (PP-LFS) consisting of 900 pairs of stimuli (50-ms interstimulus interval) delivered at 1Hz for 15 minutes and performed in aCSF containing in mM: 124 NaCl, 5 KCl, 1.25 NaH<sub>2</sub>PO<sub>4</sub>, 26 NaHCO<sub>3</sub>, 10 glucose, 2 CaCl<sub>2</sub>, 1 MgCl<sub>2</sub> (Kemp and Bashir, 1999; Huber et al., 2000). Synaptically evoked NMDA receptor-dependent LTD was induced by low frequency stimulation (LFS) consisting of 900 stimuli delivered at 1 Hz for 15 minutes in the same aCSF used in the PP-LFS studies. Sampled data was analyzed off-line using Clampfit 9.2. Three sequential fEPSPs were averaged and their slopes calculated. All fEPSP slopes were normalized to the average slope calculated during the predrug period (percent of baseline). Statistical significance was analyzed using GraphPad Prism and the Student's unpaired T-test unless otherwise noted.

## Whole Cell Patch-Clamp Recordings

18–24 day old Sprague Dawley rats were anesthetized with isoflurane, decapitated and the brains were quickly removed and submerged into ice cold choline chloride replacement solution (in mM: 126 Choline chloride, 2.5 KCl, 8 MgSO<sub>4</sub>, 1.3 MgCl<sub>2</sub>, 1.2 NaH<sub>2</sub>PO<sub>4</sub>, 10 glucose, 26 NaHCO<sub>3</sub>). The brains were then hemisected and 300 μm transverse slices were made using a vibratome (Vibratome 3000 Plus). Individual hippocampi were removed from the slice and transferred to a holding chamber containing artificial cerebrospinal fluid (aCSF; in mM: 130 NaCl, 3.5 KCl, 1.25 NaH<sub>2</sub>PO<sub>4</sub>, 24 NaHCO<sub>3</sub>, 10 glucose, 1.5 CaCl<sub>2</sub>, 1.5 MgCl<sub>2</sub>), incubated at 37 °C for 30 min, and then equilibrated at room temperature for at least 45 min before recording. In all experiments, 5 μM glutathione and 500 μM pyruvate were included in the choline chloride buffer and in the holding chamber ACSF. During recordings, slices were maintained fully submerged on the stage of a brain slice chamber perfused with heated (32 °C) and oxygenated ACSF at 2 ml/min. CA1 pyramidal neurons were visualized with an Olympus BX51WI upright microscope (Olympus, Lake Success, NY) coupled with a 40× water immersion objective and Hoffman optics. Borosilicate glass pipettes were pulled using a Flaming/Brown micropipette puller (Sutter Instruments, CA) to produce patch electrode resistances of 2 – 4 MΩ when filled with an intracellular solution containing (in mM): 135 K-MeSO<sub>4</sub>, 5 NaCl, 1 MgCl<sub>2</sub>, 0.025 CaCl<sub>2</sub>, 10 HEPES, 0.6 EGTA, 2 ATP, 0.2 GTP. The voltage-clamp signal was low-pass-filtered at 2 kHz, digitized at 10 kHz, and acquired using a Clampex9.2/DigiData 1332 system (Molecular Devices, Sunnyvale, CA).

## Morris Water Maze

CDPPB was suspended in a vehicle consisting of 20% w/v 2-hydroxypropyl-β-cyclodextrin. ADX47273 was dissolved in a vehicle consisting of 10% v/v Tween-80. All injections were made via the intraperitoneal (i.p.) route in a volume of 1 ml/100 g body weight. The maze apparatus consisted of a 90 cm diameter tub filled with 23 ± 1°C water that covered a 6 cm diameter submerged platform. Visual cues were placed above the rim of the tub to facilitate spatial navigation. The water was made opaque by addition of dilute nontoxic white paint. Mice were injected i.p. with the mGluR5 positive allosteric modulators CDPPB (10 mg/kg) or ADX47273 (10 mg/kg) or their corresponding vehicles 20 min prior to testing. A total of four test trials (each separated by 5 min) were conducted on each day for a total of 13 days, with 4 separate starting points utilized for each trial. The sequence of the starting point location was randomized on each day of testing. Latency to reach the platform was recorded for each trial, and a maximum swim time was set at 60 sec per trial. If the animal failed to reach the platform on any of the trials within 60 sec, it was then gently guided by the experimenter to the platform and given a score of 60 sec. Acquisition criteria were considered to be obtained when the latency to reach the platform was ≤ 15 sec on each of 4 consecutive trials. On the day following the last day of testing, a probe trial was conducted in which the platform was removed and time spent in the quadrant where the platform was previously located was recorded for each of 4 trials. Drugs were not administered on the day of the probe trial.

## Results

### Allosteric modulators of mGluR5 have predicted effects on phosphoinositide hydrolysis and ERK1/2 phosphorylation in native systems

Group I mGluRs signal through the activation of  $G_q$  proteins leading to the downstream effects of PKC activation, phosphoinositide (PI) hydrolysis and increased intracellular calcium release (Conn and Pin, 1997). Both the mGluR5 PAMs, and the neutral allosteric site ligand, 5MPEP, have been previously characterized using cell lines and recombinantly expressed mGluRs (Rodriguez et al., 2005; Chen et al., 2007). However, prior to using these compounds for studies of the role of mGluR5 in hippocampal LTP and LTD, we verified their ability to modulate mGluR5-mediated increases in PI hydrolysis in hippocampal slices. In agreement with previous studies (Sacaan et al., 1998; Gasparini et al., 1999; Johnson et al., 1999), the group I mGluR agonist, DHPG induced a concentration-dependent increase in PI hydrolysis in rat hippocampal slices (Figure 1A). Consistent with its effect in cell lines, the mGluR5 PAM, VU-29 had no effect on baseline PI hydrolysis but induced a leftward shift in the DHPG concentration response curve (CRC) and enhanced the maximum response to DHPG (Figure 1A; DHPG alone,  $EC_{50} = 8 \pm 1.6 \mu\text{M}$ ; DHPG + VU-29,  $EC_{50} = 4.5 \pm 1 \mu\text{M}$ , max response =  $138 \pm 12\%$ ,  $n = 5$ ). We then determined the effect of VU-29 on the response to a single concentration of DHPG that induced an approximate  $EC_{20}$  PI hydrolysis response. DHPG ( $3 \mu\text{M}$ ) induced a small but significant increase in PI hydrolysis compared with vehicle control (Figure 1B;  $p < 0.0001$ ,  $n = 3$ ), which was significantly potentiated by VU-29 ( $5 \mu\text{M}$ ) (Figure 1B;  $p < 0.001$ ,  $n = 3$ ). 5MPEP was previously described as a neutral allosteric site ligand at mGluR5 that is capable of selectively inhibiting effects of mGluR5 PAMs, such as VU-29 (Rodriguez et al., 2005; Chen et al., 2007; Chen et al., 2008). Consistent with this, 5MPEP ( $100 \mu\text{M}$ ) had no effect on the phosphoinositide hydrolysis response to DHPG but specifically inhibited the ability of VU-29 to potentiate the PI hydrolysis response (Figure 1B;  $p < 0.001$ ,  $n = 3$ ).

In addition, previous studies have demonstrated that multiple mGluR5 PAMs, including CPPHA and ADX47273 potentiate DHPG-induced increases in ERK1/2 phosphorylation in hippocampal slices and cortical astrocytes (Zhang et al., 2005; Liu et al., 2008). We now determined the effect of the VU-29 analog, CDPPB, on ERK1/2 phosphorylation in cortical astrocytes to confirm reports that mGluR5 PAMs belonging to this structural class also potentiate coupling of mGluR5 to this signaling pathway. As with the other mGluR5 PAMs, CDPPB induced a robust potentiation of DHPG-induced increases in ERK1/2 phosphorylation astrocytes (see supplemental material, Figure S1). Finally, we have previously shown that CPPHA potentiates mGluR5 regulation of NMDA receptor currents in hippocampal pyramidal cells (O'Brien et al., 2004) and that VU-29 and CDPPB selectively potentiate mGluR5-mediated responses in acute brain slices in the subthalamic nucleus while having no effect on mGluR1 mediated responses in the substantia nigra pars reticulata (Chen et al., 2007). Together, these data provide strong evidence that mGluR5 PAMs potentiate mGluR5 responses in multiple native systems.



### VU-29 potentiates threshold TBS-induced LTP in rat hippocampal CA1 region

To determine the effect of VU-29 on induction of LTP, extracellular field excitatory postsynaptic potentials (fEPSPs) were recorded from the dendritic layer of CA1 following stimulation of the Schaffer collaterals (SC-CA1 synapse). Theta burst stimulation (TBS) is a common stimulus protocol used to elicit robust LTP at this synapse (Larson and Lynch, 1989). Consistent with multiple previous reports, stimulation of SC afferents using a standard TBS protocol induced robust LTP at the SC-CA1 synapse (Figure 2A;  $183 \pm 10\%$  of baseline at 45 min post TBS,  $n = 6$ ). In contrast, stimulation with a modified TBS protocol, termed threshold TBS, using one train of a lower frequency of stimulus bursts (see methods) induced only a slight potentiation of fEPSPs (Figure 2B;  $115 \pm 6\%$  of baseline at 45 min post threshold TBS,  $n = 10$ ). Threshold TBS provides an ideal protocol that can be used to determine whether selective potentiation of mGluR5 with VU-29 can enhance LTP in response to a sub-maximal stimulus. Application of 500 nM VU-29 had no effect on the baseline synaptic responses as measured by fEPSP slope (Figure 2C;  $99.6 \pm 6\%$  of baseline,  $n = 8$ ). Interestingly, this same threshold TBS protocol induced robust LTP when delivered to slices preincubated with VU-29 (500 nM) for 20 min prior to delivery of the stimulus train (Figure 2D;  $152 \pm 8\%$  of baseline at 45 min post TBS;  $p < 0.05$ ,  $n = 10$ ). VU-29 did not alter paired pulse facilitation, fiber volley amplitude, or input-output curves compared with vehicle controls (see supplemental material, Figure S2). In addition, VU-29 did not alter passive membrane properties of CA1 pyramidal cells when recorded in whole cell patch-clamp mode including input resistance, action potential firing or membrane potential (see supplemental material, Figure S3).

Importantly, 5MPEP (100  $\mu$ M) completely blocked the ability of VU-29 to enhance LTP induced by threshold TBS (Figure 3A;  $92 \pm 14\%$  of baseline at 45 min post TBS;  $p > 0.05$ ,  $n = 8$ ), suggesting that the action of VU-29 is due to actions of this compound on mGluR5. In contrast, 5MPEP (100  $\mu$ M) had no effect on induction of LTP by a standard suprathreshold TBS protocol (Figure 3B;  $192 \pm 19\%$  of baseline at 45 min post TBS in the absence of 5MPEP,  $n = 8$ ;  $185 \pm 13\%$  of baseline 45 min post TBS in the presence of 5MPEP,  $n = 8$ ) or by the threshold TBS protocol (Figure 3C;  $133 \pm 14\%$  of baseline at 45 min post TBS in the absence of 5MPEP,  $n = 4$ ;  $128.3 \pm 11\%$  of baseline at 45 min post TBS in the presence of 5MPEP,  $n = 5$ ). This suggests that the effect of 5MPEP is specific to the VU-29 potentiation and excludes the possibility that 5MPEP inhibited LTP induction through blockade of signaling components other than mGluR5 that are required for LTP induction.

### Induction of LTP in the presence of VU-29 is dependent on activation of NMDA receptors and a Src-family tyrosine kinase

Previous studies suggest that induction of LTP at the SC-CA1 synapse by suprathreshold TBS is dependent on activation of NMDA receptors (Collingridge and Bliss, 1995). In addition, it has been shown that activation of mGluR5 with DHPG potentiates NMDA receptor currents (Collingridge and Bliss, 1995; Doherty et al., 1997; Jia et al., 1998; Awad et al., 2000; Doherty et al., 2000; Mannaioni et al., 2001; Gerber et al., 2007) and that this potentiation occurs in a G-protein, protein kinase C, and Src-family kinase dependent manner (Benquet et al., 2002). Furthermore, we previously reported that mGluR5 PAMs potentiate DHPG-induced enhancement of NMDA receptor currents recorded from CA1

pyramidal cells (O'Brien et al., 2004). Thus, it is possible that potentiation of synaptically activated mGluR5 by VU-29 enhances induction of normal NMDA receptor-dependent LTP. However, it is also possible that VU-29 leads to induction of a distinct form of LTP that is independent of NMDA receptor activation. To determine whether threshold TBS-LTP is dependent on NMDA receptor activation and Src-family kinases in VU-29 treated slices, we determined the effects of the NMDA receptor antagonist, D-AP5 (Figure 4A and 4B), and the Src-family kinase inhibitor, PP 1 (Figure 4B), on TBS-induced LTP in the presence of VU-29. D-AP5 (50  $\mu$ M) and PP 1 (20  $\mu$ M) completely blocked induction of LTP by the combination of VU-29 and threshold TBS (Figure 4; D-AP5:  $106.4 \pm 9\%$  of baseline,  $n = 4$ ; PP 1:  $98.5 \pm 4\%$  of baseline,  $n = 7$ ).

### VU-29 potentiates chemically induced mGluR-LTD

It is well established that activation of mGluR5 by exogenous agonist application induces a long term depression (LTD) of synaptic transmission at the SC-CA1 synapse, a phenomenon referred to as mGluR-LTD (Palmer et al., 1997; Camodeca et al., 1999; Fitzjohn et al., 1999; Huber et al., 2000; Huber et al., 2001; Kemp and Bashir, 2001; Faas et al., 2002; Tan et al., 2003; Nosyreva and Huber, 2005; Huang and Hsu, 2006; Volk et al., 2006; Kumar and Foster, 2007). Based on the clear role of mGluR5 in mGluR-LTD, we were somewhat surprised that selective potentiation of mGluR5 induced such a robust increase in threshold TBS-LTP and might have expected the opposite result. In light of this, we performed a series of studies to determine whether selective potentiation of mGluR5 would also enhance mGluR-LTD induced by application of the group I mGluR agonist DHPG. Before determining the effect of VU-29 on LTD, we established a concentration of DHPG that resulted in a significant but modest LTD response to ensure that we were working in a submaximal DHPG concentration range. At 75  $\mu$ M DHPG induced robust LTD (Figure 5A;  $49.1 \pm 7.8\%$  of baseline 80 – 90 min following washout of DHPG,  $n = 4$ ). A similar, though somewhat smaller LTD response was elicited with application of 50  $\mu$ M DHPG (Figure 5A;  $68.9 \pm 5.5\%$  of baseline,  $n = 5$ ). In contrast, 25  $\mu$ M DHPG resulted in only slight depression of fEPSP slopes ( $93 \pm 4.15\%$  of baseline,  $n = 10$ ) measured 80–90 minutes following washout of DHPG (Figure 5A). Based on these findings, we chose to use 25  $\mu$ M DHPG for all subsequent experiments. Twenty minute pretreatment of slices with VU-29 (500 nM) did not potentiate the effects of 25  $\mu$ M DHPG ( $84 \pm 6\%$  of baseline in the presence of 500 nM VU-29,  $n = 6$ ;  $89.4 \pm 2.9\%$  of baseline in the absence of 500 nM VU-29,  $n = 9$ ). We therefore increased the concentration of VU-29 to 1  $\mu$ M and found that prior application of this higher concentration resulted in a significant enhancement of the long-term depression of synaptic transmission induced by DHPG but had no effect on the initial acute depression (Figure 5B; Acute:  $64.2 \pm 3.3\%$  of baseline in the absence of VU-29;  $53.6 \pm 3.5\%$  of baseline in the presence of VU-29,  $p > 0.05$ ; LTD:  $93 \pm 4.15\%$  of baseline in the absence of VU-29,  $n = 10$ ;  $70.63 \pm 4.1\%$  of baseline in the presence of VU-29,  $n = 4$ ,  $p < 0.01$ ). As is the case for VU-29 potentiation of threshold LTP, the neutral allosteric modulator 5MPEP completely blocked the ability of VU-29 to potentiate DHPG-induced LTD (Figure 5C;  $70.63 \pm 4.1\%$  of baseline in the presence of VU-29,  $n = 4$ ;  $89.4 \pm 2.9\%$  of baseline in the absence of VU-29,  $n = 3$ ,  $p < 0.05$ ).

Previous studies indicate that the MAP kinase, ERK1/2, is phosphorylated upon induction of DHPG-LTD in the CA1 region of hippocampal slices (Berkeley and Levey, 2003; Gallagher et al., 2004; Banko et al., 2006) and that inhibitors of ERK1/2 signaling inhibit DHPG-LTD (Gallagher et al., 2004; Banko et al., 2006). These data, coupled with findings that mGluR5 PAMs potentiate mGluR5-induced ERK1/2 phosphorylation discussed above suggest that potentiation of ERK1/2 phosphorylation may be important for mGluR5 PAM enhancement of DHPG-LTD. To test the hypothesis that ERK1/2 phosphorylation is required for the enhancement of DHPG-LTD by VU-29, we performed extracellular field potential recordings in the presence of the MEK inhibitor, U0126. Prior treatment of the slice with U0126 (20  $\mu$ M) significantly inhibited the ability of VU-29 (1  $\mu$ M) to enhance LTD induced by DHPG (25  $\mu$ M) (Figure 5D; 67.2 $\pm$ 5.4% of baseline in the absence of U0126,  $n = 4$ ; 90.6 $\pm$ 5.7% of baseline in the presence of U0126,  $n = 4$ ,  $p < 0.05$ ). Taken together, these data suggest a mechanistic link between enhancement of the MEK/ERK pathway and the ability of mGluR5 PAMs to enhance mGluR-LTD.

### VU-29 potentiates stimulus-induced NMDA receptor-independent LTD

Paired-pulse low frequency stimulation (PP-LFS) induces a form of LTD that is independent of NMDA receptor activation (Kemp and Bashir, 1997, 1999; Huber et al., 2000; Kemp et al., 2000; Kumar and Foster, 2007). Initial studies using the broad spectrum mGluR antagonist LY341495 suggested that this form of synaptically-evoked LTD is dependent on mGluR activation (Bortolotto et al., 1999; Huber et al., 2000; Nosyreva and Huber, 2005). If this stimulus-induced LTD is mechanistically similar to DHPG-induced LTP and mediated by activation of mGluR5, VU-29 may potentiate the LTD response to PP-LFS stimulation. We therefore performed experiments to determine the effect of selective potentiation of mGluR5 by VU-29 on synaptically evoked LTD. Consistent with previous reports, PP-LFS induced a persistent depression of synaptic transmission at the SC-CA1 synapse that was somewhat smaller than the maximal LTD induced by DHPG (Figure 6A). Interestingly, PP-LFS-induced LTD was significantly enhanced by perfusion with VU-29 (1  $\mu$ M) (Figure 6A; 84.6  $\pm$  3.6% of baseline in the absence of VU-29,  $n = 6$ ; 70.8  $\pm$  5.3% of baseline in the presence of VU-29,  $n = 7$ ,  $p < 0.05$ ).

While mGluR-LTD is independent of NMDA receptor activation, a distinct form of LTD has also been described at the SC-CA1 synapse that is clearly dependent on NMDA receptor activation (Mulkey and Malenka, 1992). This NMDA receptor-dependent LTD is induced by a distinct stimulus protocol that includes low frequency stimulation (LFS) without the paired pulse stimulation used for induction of mGluR-LTD. However, as discussed above, mGluR5 activation potentiates NMDA receptor currents and mGluR5 PAMs enhance this effect (O'Brien et al., 2004). Thus, it is possible that VU-29 could potentiate NMDA receptor-dependent LTD in a manner similar to potentiation of NMDA receptor-dependent LTP by threshold TBS. If so, this could contribute to the enhancement seen during PP-LFS-induced LTD. However, in contrast to the effects of VU-29 on threshold TBS-LTP, enhancement of PP-LFS LTD persists in the presence of the NMDA receptor antagonist D-AP5 (Figure 6B; 90.7  $\pm$  2.2% of baseline in the absence of VU-29  $n = 5$ ; 80.5  $\pm$  1% of baseline in the presence of VU-29,  $n = 5$ ,  $p < 0.01$ ). This is consistent with previous results showing that mGluR-LTD is independent of NMDA receptor activation (Kemp and Bashir,

1997, 1999; Huber et al., 2001; Kumar and Foster, 2007). Furthermore, VU-29 does not potentiate LTD induced by the low frequency stimulation protocol used to induce NMDA receptor-dependent LTD. LFS induced a modest long lasting reduction in the slope of the fEPSP (Figure 6C). Surprisingly, we found that VU-29 was not able to potentiate this NMDA receptor-dependent form of LTD (Figure 6C;  $87.07 \pm 4\%$  of baseline in the absence of VU-29,  $n = 5$ ;  $86.88 \pm 6.7\%$  in the presence of VU-29,  $n = 4$ ,  $p > 0.05$ ).

### **VU-29 does not alter induction of suprathreshold LTP or reduce saturated LTP**

The finding that selective potentiation of mGluR5 potentiates afferent stimulation-induced LTD raises the possibility that selectively enhancing mGluR5 at glutamatergic synapses could reduce the LTP response induced by some stimulus protocols. Thus, while VU-29 potentiates threshold TBS-induced LTP, it is conceivable that this compound could dampen the maximal LTP that is induced with suprathreshold TBS stimulation or lead to depotentiation of saturated LTP in response to a stimulus protocol that normally would only lead to LTP. To address these possibilities, we determined the effect of VU-29 on LTP induced by suprathreshold stimulation. Interestingly, VU-29 had no effect on the LTP induction when a stimulus protocol (4 trains of 10 Hz TBS) that induces robust LTP in the absence of VU-29 was used (Figure 7;  $218 \pm 26\%$  of baseline at 30 min post TBS in the absence of VU-29,  $n = 6$ ;  $192 \pm 13\%$  of baseline in the presence of VU-29,  $n = 5$ ,  $p > 0.05$ ). In order to determine whether VU-29 would alter the response to TBS stimulation under conditions of saturated LTP, we applied VU-29 to slices in which a maximal LTP had been previously established. Similar to studies of the response to a single suprathreshold TBS protocol (Figure 7A), VU-29 had no effect on the response to the second suprathreshold TBS (Figure 7B:  $191 \pm 14\%$  of baseline 30 minutes post TBS,  $n = 6$ ;  $185 \pm 7\%$  of baseline 30 minutes post TBS in the presence of VU-29,  $n = 6$ ,  $p > 0.05$ ).

### **A structurally distinct mGluR5 PAM mimics the effects of VU-29 on threshold TBS-induced LTP and DHPG-LTD**

If VU-29-induced potentiation of threshold TBS-LTP and DHPG-LTD is due to selective potentiation of mGluR5 responses, it should be mimicked by a structurally distinct mGluR5 PAM. A novel mGluR5-selective PAM, termed ADX47273 that is structurally unrelated to VU-29 has recently been identified (Le Poul, 2005; de Paulis et al., 2006; Liu et al., 2008). This provides an excellent additional tool to verify the effects of mGluR5 PAMs on LTP and LTD. In addition, ADX47273 is systemically active when dosed intraperitoneally and activates mGluR5-mediated increases in ERK1/2 phosphorylation *in vivo*, making it a useful compound for behavioral studies (Liu et al., 2008). We and others (Liu et al., 2008) have established a protocol for synthesis of ADX47273 and characterized this compound to verify that it is a selective mGluR5 PAM (see supplemental material, Figure S4). In previous studies, we rigorously characterized DHPG-induced calcium mobilization in secondary cultured rat cortical astrocytes and found that this response is exclusively mediated by mGluR5 (Peavy et al., 2002) and that mGluR5 PAMs potentiate this response (Rodriguez et al., 2005; Zhang et al., 2005; Hemstapat et al., 2006; Chen et al., 2007). ADX47273 induced a robust potentiation of glutamate-induced calcium mobilization in cortical astrocytes in a manner similar to that previously reported for VU-29. Thus, ADX47273 induced a concentration-dependent potentiation of the response to an  $EC_{20} - EC_{30}$  concentration of

glutamate (300 nM) with an EC<sub>50</sub> value for ADX47273 of 108 ± 41 nM (Figure S5). Furthermore, ADX47273 (10 μM) did not affect the glutamate concentration response curves of mGluR1, mGluR2 or mGluR4 (Figure S6; p>0.05), suggesting that this compound is selective as a PAM for mGluR5 relative to these other mGluR subtypes. Data presented by Liu et al (2008) also confirms the selectivity of ADX47273 as well as demonstrating that ADX47273 competes with [<sup>3</sup>H]-MPEP binding and increases both ERK and CREB phosphorylation in the hippocampus and prefrontal cortex (Liu et al., 2008). Consistent with the effects of VU-29 (Figure 2D), ADX47273 (10 μM) induced a significant increase in threshold TBS-induced LTP of fEPSPs in the rat hippocampal CA1 region (see supplemental material, Figure S7A; 133 ± 6% of baseline at 45 min post TBS in the absence of ADX47273, n = 9; 176 ± 9% of baseline at 45 min post TBS in the presence of ADX47273, n = 12). While the control level of LTP induced by the threshold-TBS protocol is variable from day to day, the potentiation of threshold-TBS LTP induced by ADX47273 was virtually identical to that induced by threshold-TBS in the presence of VU29. Furthermore, ADX47373 (10 μM), induced a significant potentiation of DHPG-induced LTD while having no effect on acute depression (see supplemental material, Figure S7B; Acute: 64.2±3.3% of baseline in the absence of ADX47273; 63.3±4.1% of baseline in the presence of ADX47273, p>0.05; LTD: 93±4.15% of baseline in the absence of ADX47273, n = 9; 71.5±2.5% of baseline in the presence of ADX47273, n = 4, p<0.01). Taken together, these data provide strong evidence that selective potentiation of mGluR5 responses to endogenous glutamate potentiates threshold TBS-induced LTP and DHPG-induced LTD and will allow further *in vivo* characterization of mGluR5 PAMs.

### **mGluR5 PAMs enhance hippocampus-dependent learning and memory**

Collectively our data demonstrate a clear ability for mGluR5 PAMs to enhance both hippocampal LTP and LTD in a manner that maintains appropriate activity-dependence of these forms of synaptic plasticity. Theoretically, this ability to enhance both LTP and LTD in a manner that does not shift the balance of these forms of synaptic plasticity could provide an ideal profile for compounds that could be used as cognition-enhancing agents. To directly test the hypothesis that mGluR5 PAMs enhance a hippocampus-dependent form of learning, we performed studies to determine the effects of two systemically active and structurally distinct mGluR5 PAMs, CDPPB and ADX47273, on performance in the Morris water maze, a model of hippocampus-dependent spatial learning. Both CDPPB and ADX47273 (each at a dose of 10 mg/kg) enhanced performance in the Morris water maze. This augmented performance is evidenced by significant decreases in the number of days required to reach acquisition criteria (Figure 8. 10.23±0.54 in the absence of CDPPB, n = 13; 8.5±0.57 in the presence of CDPPB, n = 12, p<0.05; 8.64±0.69 in the absence of ADX47273, n = 11; 6.83±0.63 in the presence of ADX47273, n = 12, p<0.05) and increased time spent in the target quadrant during the probe trial (Figure 8. 51.5±3.5% in the absence of CDPPB, n = 13; 63.6±4.0% in the presence of CDPPB, n = 12, p<0.05; 53.4±1.9% in the absence of ADX47273, n = 11; 68.1±2.9% in the presence of ADX47273, n = 12, p<0.05). These data, coupled with the unique profile of mGluR5 PAMs on hippocampal synaptic plasticity provide strong support for the hypothesis that mGluR5 PAMs may provide efficacy as a novel approach to enhancing cognitive function *in vivo*.

## Discussion

In recent years, mGluR5 PAMs have emerged as an exciting new approach that holds promise for treatment of the positive symptoms of schizophrenia (see (Conn et al., 2008) for review). In addition, increasing evidence suggests that mGluR5 plays an important role in multiple forms of learning and memory and that selective mGluR5 PAMs may provide a novel approach to treat cognitive disturbances in patients with schizophrenia and other disorders that include impaired cognitive function (Campbell et al., 2004; Homayoun et al., 2004; O'Brien et al., 2004; Kinney et al., 2005; Balschun et al., 2006; Lecourtier et al., 2007; Bikbaev et al., 2008; Chan et al., 2008; Darrah et al., 2008).

Abundant evidence suggests that mGluR5 plays important roles in induction of both hippocampal LTP (Lu et al., 1997; Cohen et al., 1998; Raymond et al., 2000; Francesconi et al., 2004; Manahan-Vaughan and Braunewell, 2005; Shalin et al., 2006) and LTD (Gasparini et al., 1999; Huber et al., 2001; Faas et al., 2002; Hou and Klann, 2004; Huang et al., 2004; Huang and Hsu, 2006), two opposing long-lasting forms of synaptic plasticity. On the surface, the roles of mGluR5 in these forms of synaptic plasticity are consistent with potential cognition-enhancing effects of these agents. However, the potential for selective actions on these opposing forms of synaptic plasticity raises the possibility that mGluR5 PAMs could lead to inappropriate long-term changes in synaptic responses that are no longer determined by specific patterns or frequencies of synaptic activity. The most important finding of the present studies is that mGluR5 PAMs enhance both LTP and LTD but do not alter the balance or patterns of activity that induce these forms of hippocampal synaptic plasticity. This is in striking contrast to some pathological conditions that include pathophysiological changes in responses to different patterns of afferent stimulation. For instance, in animal models of Fragile X Syndrome (FXS), induction of mGluR5-dependent LTD is selectively enhanced (Huber et al., 2002; Bear et al., 2004; Nosyreva and Huber, 2006) while induction of LTP is impaired (Lauterborn et al., 2007) and this is thought to underlie the cognition-impairment seen in FXS patients. Similar findings have been reported in animal models of intense stress (Chaouloff et al., 2007, 2008) or seizure activity (Kirschstein et al., 2007). The previous findings illustrate the critical need to ensure that agents intended to enhance cognitive function do not induce pathophysiological changes in the balance of these forms of synaptic plasticity.

The unique ability of mGluR5 PAMs to enhance both LTP and LTD and maintain appropriate dependence of both forms of plasticity on specific patterns of synaptic activity, rather than inducing pathophysiological changes in the balance of LTP and LTD, suggests that these agents could have an ideal profile for use as potential cognition-enhancing agents. Consistent with this, we now report direct evidence that two structurally distinct mGluR5 PAMs enhance performance in a model of hippocampus-dependent spatial learning. These findings are consistent with multiple studies that have been reported over the past year revealing that mGluR5 PAMs enhance other aspects of cognitive function in animal models (Lecourtier et al., 2007; Chan et al., 2008; Darrah et al., 2008; Liu et al., 2008). For example, the mGluR5 PAM, DFB, induced marked improvement in spatial alternation retention (Balschun et al., 2006). In addition, CDPPB improved cognitive flexibility in a set-shifting paradigm after disruption with MK-801 (Darrah et al., 2008) and ADX47273

increased object exploration in a novel object recognition paradigm (Liu et al., 2008). Thus, our current findings are consistent with an emerging set of behavioral studies and may provide insights into the mechanisms by which mGluR5 PAMs can enhance cognitive function.

In addition to providing critical new insights into the roles of mGluR5 in both LTP and LTD, these findings illustrate a critical potential advantage of PAMs relative to traditional agonists. Unlike mGluR5 PAMs, mGluR5 agonists induce profound LTD and also lead to induction of seizure activity in hippocampal slices and in animal models (Merlin and Wong, 1997; Merlin et al., 1998; Wong et al., 1999; Kingston et al., 2002; Wong et al., 2005). Thus, traditional mGluR5 agonists have the potential to impair cognitive function by selectively inducing LTD and could induce seizure activity. This provides a potential therapeutic advantage to maintaining activity-dependence of mGluR5 signaling by using mGluR5 PAMs that selectively potentiate responses to synaptically released glutamate. Furthermore, it is intriguing that, in contrast to mGluR5 PAMs, the mGluR5-selective agonist CHPG enhances a form of LTD (induced by LFS) that is normally NMDA receptor-dependent but mGluR5-independent (Neyman and Manahan-Vaughan, 2008).

In summary, when taken together with multiple lines of evidence suggesting that these compounds have antipsychotic efficacy (O'Brien et al., 2003; Lindsley et al., 2004; O'Brien et al., 2004; Kinney et al., 2005; Le Poul, 2005; de Paulis et al., 2006; Lindsley et al., 2006; Chen et al., 2007; Chen et al., 2008; Liu et al., 2008), the present data provide strong preclinical support for use of selective mGluR5 PAMs as novel therapeutic agents capable of treating both the positive symptoms and cognitive deficits associated with schizophrenia. In addition, these data raise the possibility that mGluR5 PAMs have potential utility in treatment of other disorders that involve impairments in cognitive function.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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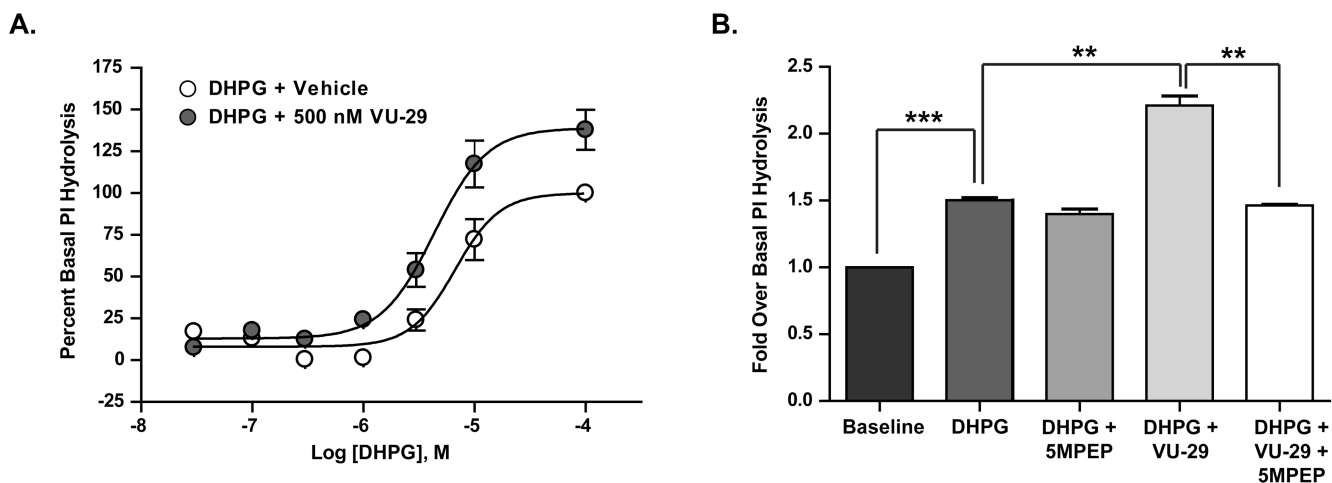


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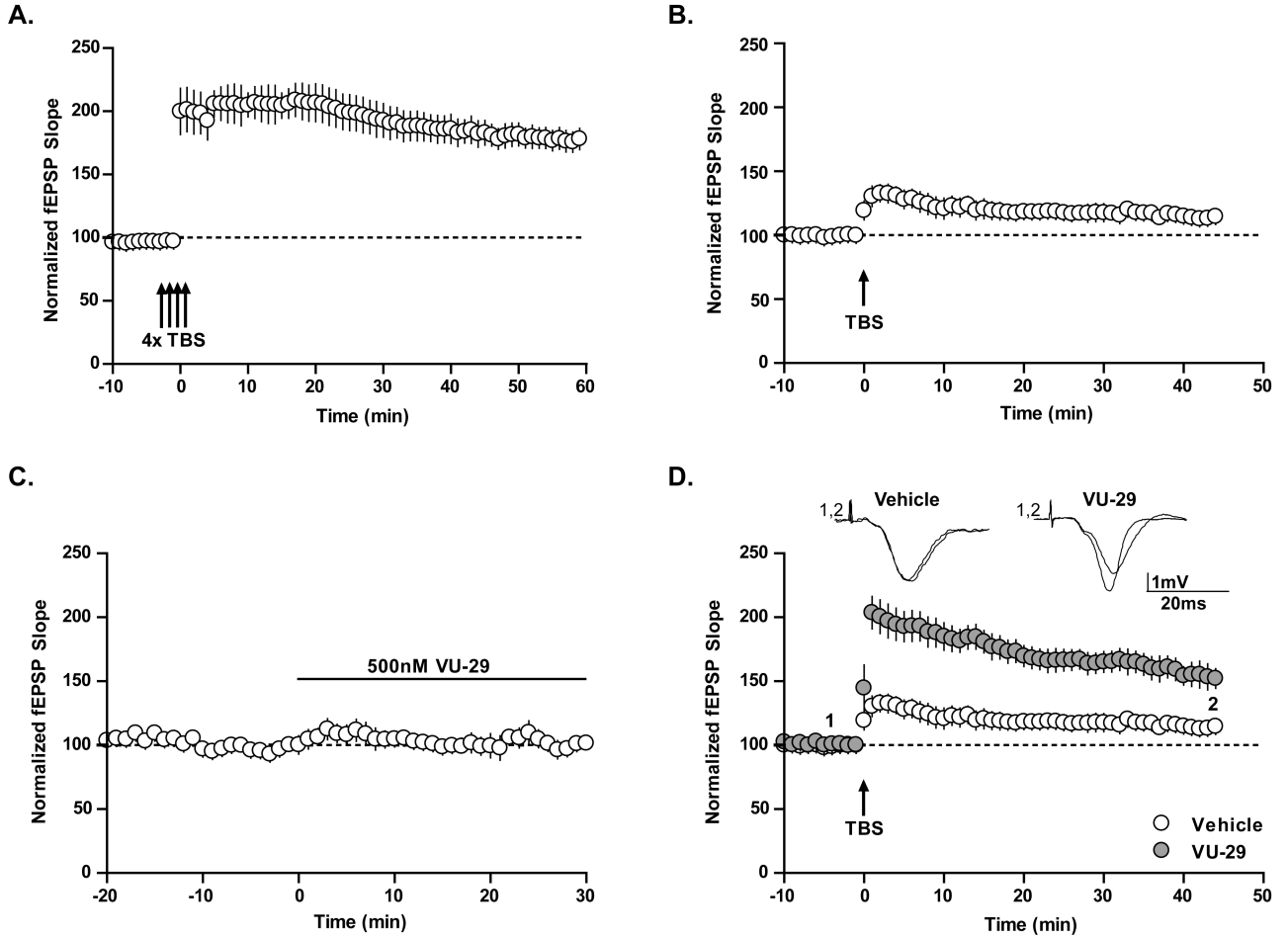
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**Figure 1. VU-29 potentiates DHPG induced increases in PI hydrolysis in rat hippocampal slices**

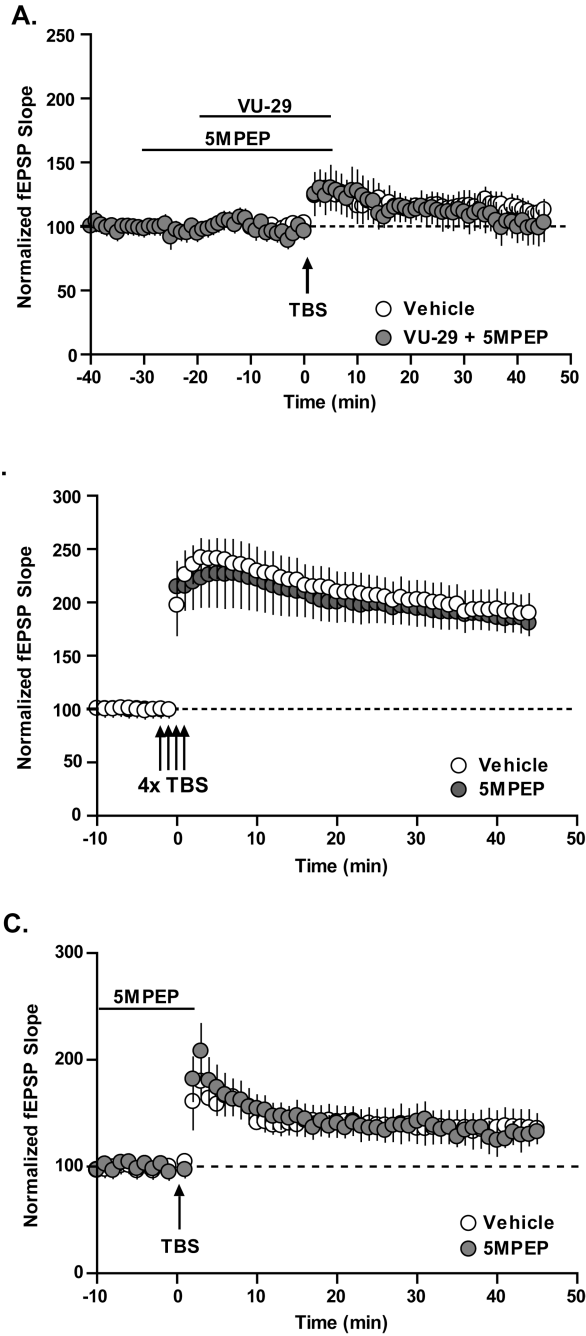
**A.** 500nM VU-29 induced a significant leftward shift in the concentration response curve of DHPG-induced PI hydrolysis in rat hippocampal slices. In the presence of VU-29 the  $EC_{50}$  of DHPG was  $4.5 \pm 1 \mu\text{M}$ , compared with  $8 \pm 1.6 \mu\text{M}$  in the absence of VU-29.

Additionally, the maximum response was enhanced to by  $38 \pm 13 \%$ . **B.** 3  $\mu\text{M}$  DHPG increased baseline PI hydrolysis, which was significantly enhanced by pre-incubation of 5  $\mu\text{M}$  VU-29 but not altered by 100  $\mu\text{M}$  5MPEP in rat hippocampal slices. 5MPEP blocked the potentiation caused by VU-29. ( $n = 5$ , experiments performed in triplicate). Error bars represent S.E.M. \*\*\*  $p < 0.0001$ , \*\*  $p < 0.001$ .



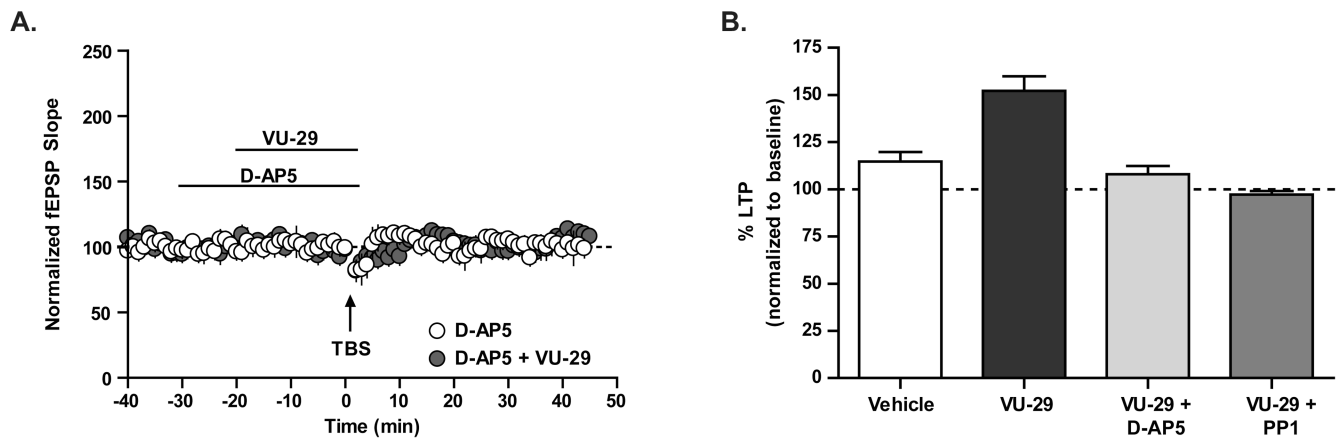
**Figure 2. The mGluR5 allosteric potentiator VU-29 facilitates the induction of LTP in area CA1 of the hippocampus**

**A.** Four trains of 10 Hz TBS induced a long lasting potentiation of the fEPSP slope at the SC-CA1 synapse. **B.** Threshold TBS induced a slight potentiation of the fEPSP slope. **C.** A 30 min incubation of 500 nM VU-29 did not alter the baseline initial fEPSP slope recorded in the rat hippocampal CA1 region ( $n = 8$ ). **D.** In control slices, threshold TBS induced a slight potentiation of fEPSP. In the presence of VU-29 (20 min pre-incubation) the same stimulation induced a significant potentiation ( $n = 12$ ;  $p < 0.05$ ). Error bars represent S.E.M.



**Figure 3. Potentiation of LTP by VU-29 is blocked by mGluR5 neutral allosteric modulator 5MPEP**

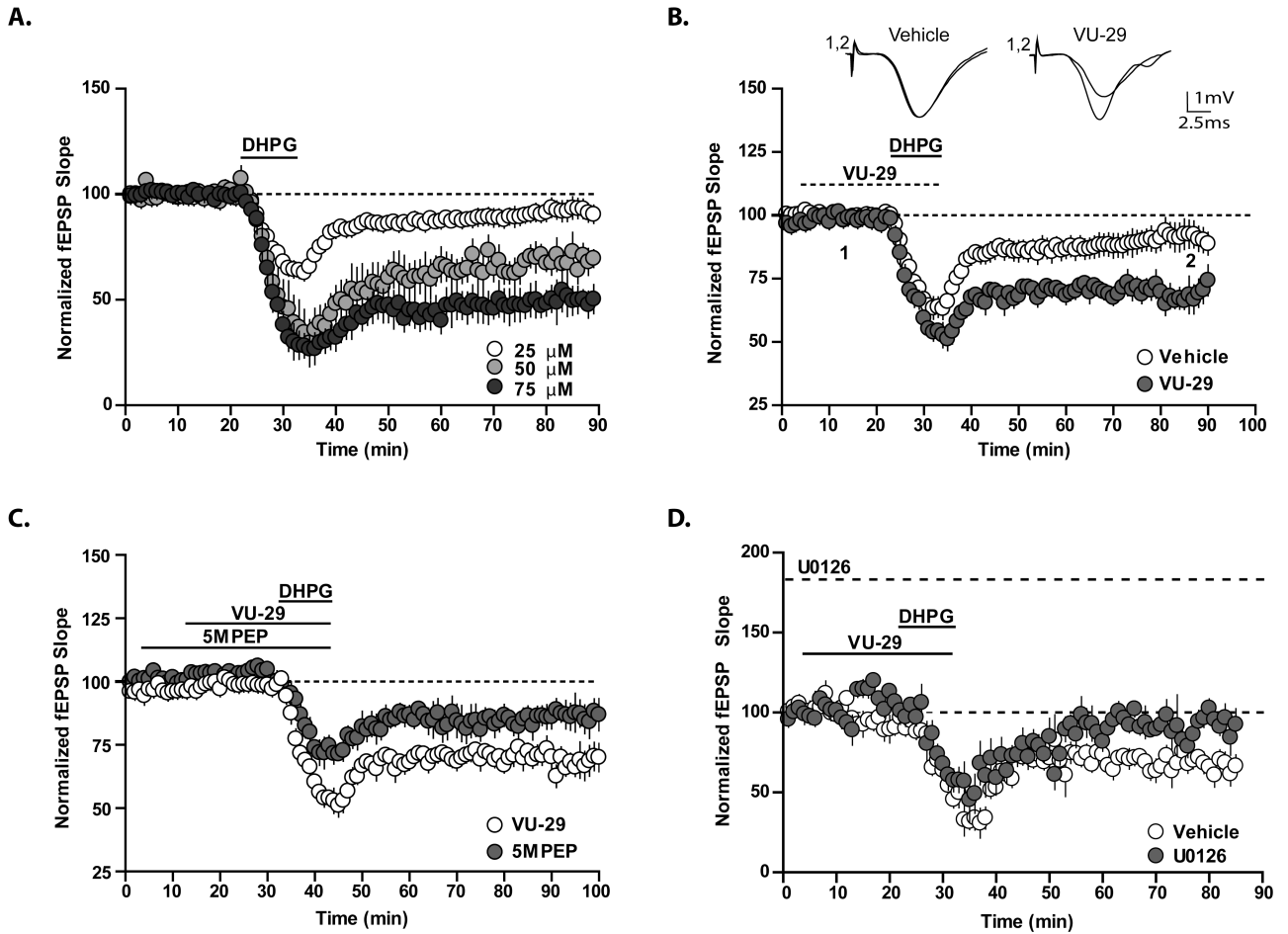
**A.** Pre-incubation of 100  $\mu$ M 5MPEP completely inhibited VU-29-facilitated TBS-induced LTP back to vehicle control ( $n = 8$ ;  $p < 0.05$ ). **B.** 10 Hz TBS induced LTP was not altered by the pre-incubation of 100  $\mu$ M 5MPEP ( $n = 8$ ;  $p > 0.05$ ). **C.** Threshold TBS induced LTP was not altered by pre-incubation of 100  $\mu$ M 5MPEP ( $n = 5$ ;  $p > 0.05$ ). Error bars represent S.E.M.



**Figure 4. The NMDA receptor antagonist, D-AP5, and the Src-family kinase inhibitor, PP 1, block VU-29-facilitated LTP of fEPSP in rat hippocampal CA1 region**

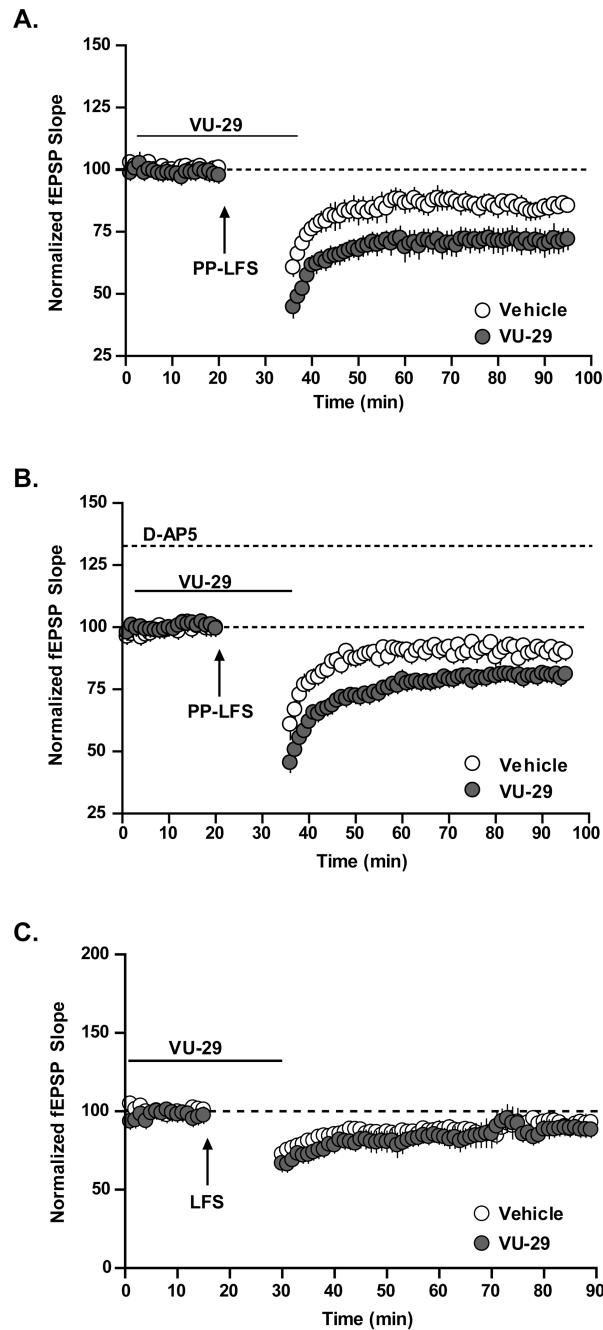
**A.** 50  $\mu$ M D-AP5 ( $n = 8$ ) completely blocked TBS-induced LTP in the presence of VU-29 ( $p > 0.05$  compared to vehicle treated slices). **B.** Bar graph depicting percent LTP induced by threshold TBS stimulation in the presence of VU-29 and 50  $\mu$ M D-AP5 ( $n = 8$ ) or 20  $\mu$ M PP 1 ( $n = 7$ ). Error bars represent S.E.M.





**Figure 5. VU-29 potentiates chemically induced mGluR-LTD in area CA1 of the rat hippocampus**

**A.** Field EPSPs are reduced upon addition of the group I mGluR agonist, DHPG, in a concentration dependent manner. **B.** In control slices, 25  $\mu$ M DHPG induced a modest level of LTD that was significantly enhanced in slices pre-incubated with 1  $\mu$ M VU-29 ( $n = 4$ ;  $p < 0.05$ ), while having no effect on acute depression ( $n = 4-6$ ;  $p > 0.05$ ). **C.** Pre-incubation of the slice with 100  $\mu$ M 5MPEP selectively inhibits the ability of VU-29 (1  $\mu$ M) to enhance DHPG-LTD ( $n = 3-4$ ;  $p < 0.05$ ) while having no effect on DHPG-LTD by itself ( $n = 3$ ;  $p > 0.05$  compared to DHPG alone). **D.** Pre-incubation of the slice with 20  $\mu$ M of the MEK inhibitor, U0126, inhibits the ability of VU-29 (1  $\mu$ M) to enhance DHPG-LTD ( $n = 3-4$ ,  $p < 0.05$ ). Error bars represent S.E.M.



**Figure 6. Selective enhancement of mGluR5 receptor signaling significantly increases stimulus induced mGluR-LTD**

**A.** Paired-pulse low frequency stimulation induces mGluR-dependent LTD that was significantly enhanced following pre-incubation of the slice with 1  $\mu$ M VU-29 ( $n = 6-7$ ;  $p < 0.05$ ). The enhancement of PP-LFS induced LTD is independent of NMDA receptor activation and selective enhancement of mGluR5 receptor signaling has no effect on NMDA receptor-dependent LTD. **B.** Incubation of the slice with 50  $\mu$ M D-AP5 did not affect the ability of 1  $\mu$ M VU-29 to potentiate PP-LFS-induced LTD ( $n=5$ ;  $p < 0.01$ ). **C.** Low frequency

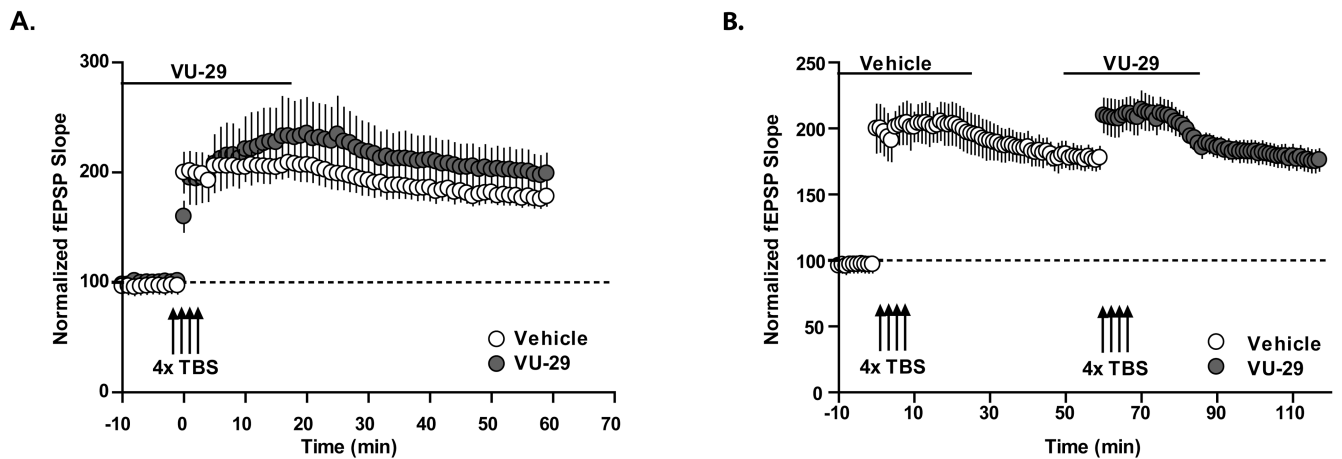
stimulation induces NMDA receptor-dependent LTD that is not enhanced in the presence of 1  $\mu$ M VU-29 ( $n = 5-6$ ;  $p > 0.05$ ). Error bars represent S.E.M.

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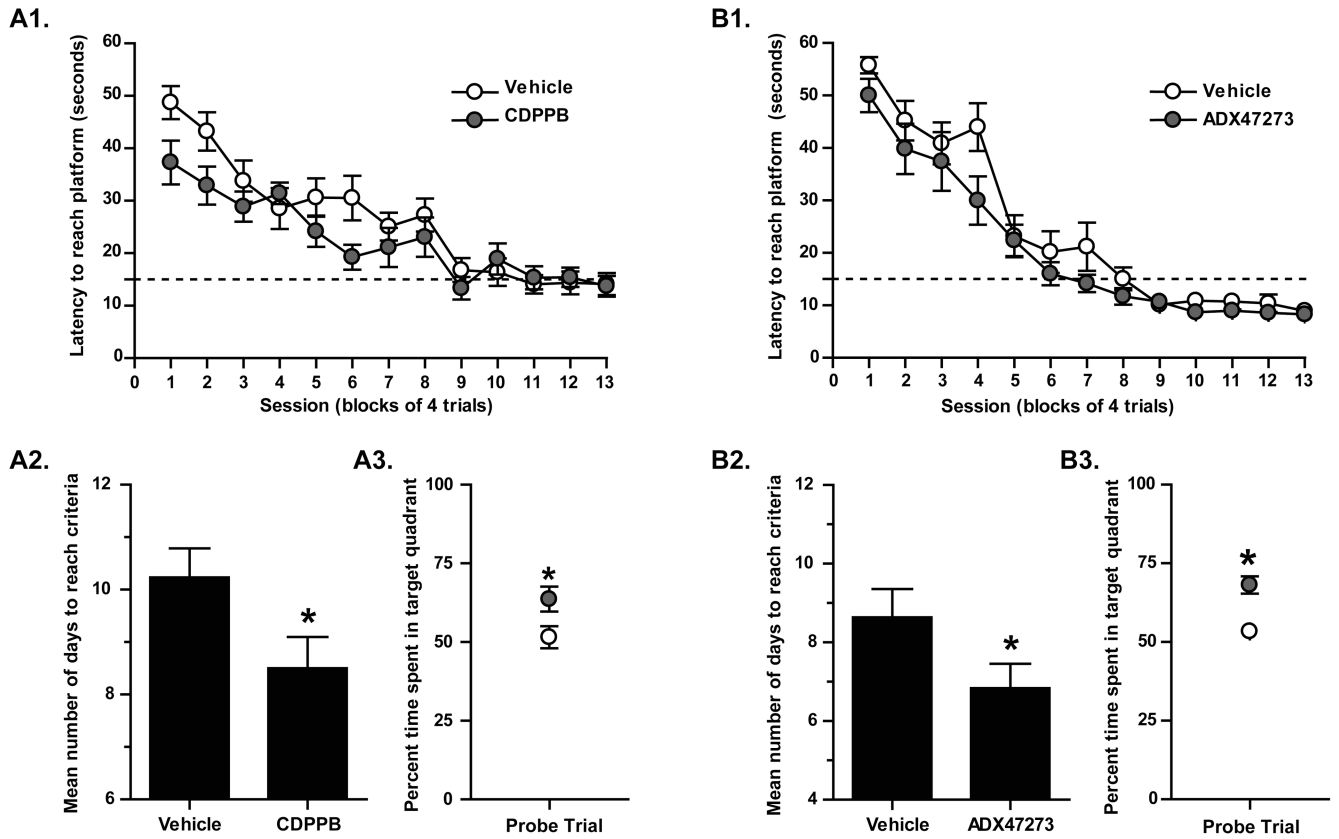
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**Figure 7. VU-29-facilitated LTP shares similar mechanisms as TBS induced LTP in area CA1 of the hippocampus**

**A.** 500 nM VU-29 did not alter LTP induced by a suprathreshold TBS protocol that induces robust LTP. In control slices, a 4X 100 Hz TBS induced robust LTP. The same stimulation in the presence of VU-29 yielded a potentiation that was not significantly different from control ( $n = 5$ ;  $p > 0.05$ ). **B.** 500nM VU-29 did not alter the induction of LTP by a suprathreshold TBS protocol in slices in which LTP was previously fully saturated. LTP was induced by 4 trains of 10 Hz TBS. After 30 minutes, the slices were incubated with 500 nM VU-29, followed by another 4 trains of 10 Hz TBS, which did not overcome the potentiation induced by the first 4 trains of TBS ( $n = 8$ ;  $p > 0.05$ ). Error bars represent S.E.M.



**Figure 8. The systemically active mGluR5 PAMs CDPBP and ADX47273 enhance performance in the Morris water maze**

**A.** CDPBP (10 mg/kg) decreases latency to reach platform (**A1**), decreases mean number of days to reach criteria (15 sec to reach platform;  $p < 0.03$ ;  $p < 0.05$ ) (**A2**), and increases time spent in target quadrant during probe trial (**A3**) ( $n = 12-13$ ;  $p < 0.05$ ). **B.** ADX47273 (10 mg/kg) also decreases latency to reach platform (**B1**), decreases mean number of days to reach criteria (15 sec to reach platform;  $p < 0.05$ ) (**B2**), and increases time spent in target quadrant during probe trial (**B3**) ( $n = 11-12$ ;  $p < 0.05$ ).