Saudi Journal of Biological Sciences 30 (2023) 103718

Contents lists available at ScienceDirect

Saudi Journal of Biological Sciences

journal homepage: www.sciencedirect.com

Original article

Are iron ore microparticles toxic for the European clam Ruditapes decussatus? Response elements from biomarker activities and in silico modeling



^a University of Nouakchott, Faculty of Sciences and Technology, New University Campus, BP 5026, Nouakchott, Mauritania

^b Laboratory of Environment Biomonitoring, Coastal Ecology Unit, Faculty of Sciences of Bizerte, University of Carthage, Zarzouna, Tunisia

^c Department of Biology, College of Science, Princess Nourah bint Abdulrahman University, P.O. Box 84428, Riyadh 11671, Saudi Arabia

^d Biology Department, College of Science, Imam Mohammad Ibn Saud Islamic University (IMSIU), Riyadh 11623, Saudi Arabia

ARTICLE INFO

Article history: Received 12 May 2023 Revised 14 June 2023 Accepted 23 June 2023 Available online 28 June 2023

Keywords: Ruditapes decussatus Bioindicators Biomarkers Bioavailability Microparticles Iron ore

ABSTRACT

Inevitably, high concentrations of iron, the most widely produced ore globally, can be found in aquatic environments. To assess the toxicity of iron on aquatic organisms, Ruditapes decussatus specimens were subjected to microparticles derived from two types of iron ore (hematite and magnetite) at four different concentrations (0.5, 1, 1.5, and 5 g/L). The findings revealed that both types of iron ore were absorbed by clams in a concentration-dependent manner. Biomarkers analysis demonstrated significant and organspecific impacts on the health of the clams caused by these microparticles, which was further supported by computational analyses on bioavailability. Within seven days of exposure, changes were observed in the activities of several enzymes, including catalase, acetylcholinesterase, and glutathione S-transferases, as well as in the rate of lipid peroxidation in both the digestive gland and gills. This study provides an environmental perspective on the toxicological effects of iron ore microparticles.

© 2023 The Author(s). Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

Global reports have extensively documented humans' uptake of heavy metals through food chains (Mulvad et al., 1996; Kumar and Achyuthan, 2007: Achary et al., 2017). These heavy metals, characterized by their persistent nature and inability to degrade, tend to accumulate in vital organs like the liver, stomach, and kidneys. Consequently, their entry into the body poses significant health risks (Duruibe et al., 2007). Each heavy metal exhibits specific toxicity: Cd primarily affects the kidneys and bones, Hg is particularly detrimental to the human nervous system, Pb has neurotoxic

Peer review under responsibility of King Saud University.



effects on children and pregnant women, and long-term exposure to As increases the risk of skin cancer, dermatomycosis, and hyperpigmentation (Khanam, 2014). The mining of metals contributes to environmental pollution, particularly the contamination of water caused by mining processes and waste discharge. Several studies. such as those conducted by Carlsson et al. (2002), Alvarez et al. (2008), and Mataís (2008), have highlighted the significance of mining waste deposits in close proximity to mining areas as a major and easily transportable source of contamination. The movement of these waste deposits depends on critical factors such as wind speed, direction, waste volume, waste type, sulfide concentration, porosity, degree of compaction, and more. The residues generated from metal mining have a substantial negative impact on water and sediment quality, as indicated by research conducted by Lottermoser (2010) and Younger et al. (2002). In addition to the physical disturbances caused by the high discharge and excessive deposition of mining waste on the seafloor, the presence of metals and metalloids can result in toxic effects, as evidenced by studies conducted by Ramirez-Llodra et al. (2015), Morello et al. (2016), and Jordi et al. (2021). Most studies on mining activities focus on sites affected by sulfide ore residues, as demonstrated by the works of Sternal et al. (2017) and Pedersen et al. (2018).

https://doi.org/10.1016/j.sjbs.2023.103718

1319-562X/© 2023 The Author(s). Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).





^{*} Corresponding author.

E-mail addresses: melainineaillal@una.mr (M. Aillal), khazri.abdzlhafidh@fsb. u-carthage.tn (A. Khazri), nialhoshani@pnu.edu.sa (N. Al-Hoshani), faboufahja@ imamu.edu.sa (F. Boufahja), hamouda.beyrem@fsb.ucar.tn (H. Beyrem), lafdal@ environnement.gov.mr (M. Yahya Lafdal).

The extraction of iron ore is crucial for the modern economy as it is an essential resource for many metal mines. Iron ore is extracted in various countries, including Australia, Brazil, Mauritania, and China (WA, 2019), is the latter being the world's leading producer. However, the iron mining industry is also characterized by significant waste production. In the Quadrilátero Ferrífero region of Brazil, approximately 40% of the extracted mass of iron ore is rejected for every metric ton extracted (Mendes et al., 2019). The National Mining Industries Company (SNIM), which operates iron ore mining, is mostly positioned in the wilayas of Tiris Zemour (Zouerate) and Dakhlet Nouadhibou in the southern part of the Cape Blanc in Mauritania. It has a mineral port where iron ore is loaded onto large ships for transportation to international markets. During this operation, significant amounts of mine dust invade the coast. The Cap Blanc site has the highest metal content, as it receives most of the discharge from the country's largest iron ore processing industry (SNIM). Several studies indicate that spatial variations of trace metal elements analyzed inside mussels' tissues (Perna perna) along the Mauritanian coasts show heterogeneous and relatively low trends in the concentration of different metals analyzed in this species (Bilal, 2017; IMROP, 2019; Sidoumou et al., 1999; Legraa, 2019).

Bivalves are commonly used as biological indicators in environmental programs and policies due to their bioaccumulation of various pollutants, particularly trace metals (Sellami et al., 2017; Oliveira et al., 2018). Ecotoxicological studies using molluscs often rely on monitoring biomarker responses (Volland et al., 2017). The biological material used in the current study was the benthic filterfeeding clam species *Ruditapes decussatus*, which is largely present in the Mediterranean Sea and Northeast Atlantic Ocean, and its usefulness in the quality assessment of water and sediments in coastal areas (Sobral and Widdows, 1997). In silico bioavailability and toxicokinetics, based on the physicochemical properties, were assessed to support the experimental findings.

2. Material and methods

2.1. Chemistry of iron ore

X-ray fluorescence analysis has been adopted by the SNIM company to determine the elements present (Iron, SiO_2 , AI_2O_3 , P, H_2O , and other elements that have not been disclosed) in the two types of iron ore, XFA (Hematite ore), and GMAB (Magnetite ore) (Table 1).

The exploitation of hematite (Fe_2O_3) and magnetite (Fe_3O_4) iron ore represents a significant source of heavy metal emissions such as Ni, Pb, Cu, Co, Cd, and Zn. These heavy metals, found in iron ore dust, are then deposited on surfaces near the mines at different distances depending on wind speed and particle size (Simpson and Spadaro, 2016; Embile et al., 2018; Mendes et al., 2019; WA, 2019). In particular, the high occurrence of iron is common in polluted sites (Berrow and Mitchell, 1991; Colbeck, 1995; Adamson, 2000; Manceau et al., 2002), and can cause significant interferences, such as:

- Fe/Cu: the iron line tends to absorb that of copper,

Table 1Composition of iron ore.

	Iron ore fine XFA (%)	Iron ore fine GMAB (%)
Iron	58	66
SiO ₂	14	7
Al_2O_3	1	0.35
Р	0.075	0.015
H ₂ O	1.5	1.5
Other elements	25.43	23.135

- Co/Fe: overestimation of the iron values when measuring those of cobalt and *vice versa* (Sammut, 2007).

In light of these interferences, it has been decided to perform heavy metal analysis on the fine fraction of the iron ore using Flame Atomic Absorption Spectroscopy (attacked by HCl + HF followed by HNO_3 + HCl and recovery by HCl, then analyzed by SAA-Flame) for Co, Cu, Ni, and Pb (Fig. 1).

2.2. Sampling of animals and implementation of contamination experiments

European clams *R. decussatus* of approximately identical sizes (3.25-4.1 cm) were gathered next to Menzel Jemil City (GPS coordinates: $37^{\circ}13'31.6''$ N $9^{\circ}55'40.9''$ E) at Bizerte lagoon (Tunisia) because of its pristine aspect with no close iron discharges. In the field, bivalves were collected arbitrarily, nonetheless of sex and size, and placed in 2 L glass aquariums full of seawater collected previously at the collection biotope. The selected clams are transferred to coolers filled with seawater until arriving at the laboratory. Bivalves were first acclimatized for three days before exposure to nine different experimental conditions, with ten individuals per aquarium. The experimental conditions included a control group (20 clams not exposed to iron ores) and eight groups exposed to varying concentrations of hematite and magnetite ores.

The experimental conditions correspond to the following treatments for both hematite ore and magnetite ore:

- An untreated control (20 clams).
- A concentration C1 (10 clams exposed to 0.5 g/L of ore).
- A concentration of C2 (10 clams exposed to 1 g/L of ore).
- A concentration C3 (10 clams exposed to 1.5 g/L of ore).
- A concentration of C4 (10 clams exposed to 5 g/L of ore).

Clams exposed to iron ore were exposed for 7 days. The selected time intervals were determined to evaluate the impact of salinity over time. Following a 7-day exposure period, the water in each glass beaker was removed and replaced every 48 h with natural seawater containing equivalent salinity levels. This process aimed to refresh the water and maintain a stable salinity environment. During that period, the aquaria were full of natural seawater and is renewed every couple of days. A constant temperature was adopted during the experiment (18 ± 1 °C); the dissolved oxygen and the salinity were kept at 6.25 mg/L, and 32 PSU, respectively. No loss of life was noticed at the time of the experiment and bivalves were fed ordinarily.

2.3. Evaluation of biochemical markers' activities

Following the dissection of each group of five bivalves, their organs (gills and digestive glands) were carefully collected and placed in a stake box filled with ice to maintain their freshness. The total protein extraction process was then conducted using a buffer solution consisting of distilled water (500 mL), sucrose (500 mM), EDTA (1 mM), Tris/HCl (10 mM), and PMSF (1 mM), with a constant pH value of 7.4. The tissues were crushed with the assistance of an Ultra-Turrax homogenizer (IKA T18 Basic) in the prepared buffer, and subsequently, they were centrifuged for 30 min at 9000 rpm (4 °C) to separate the proteins, which were quantified in the resulting supernatant according to the method described by Bradford (1976).

Catalase is implicated in cell protection in opposition to Reactive Oxygen Species. Herein, the evaluation of catalase activities (CAT) was based on the method reported by Aebi (1974).

Glutathione S-transferase (GST) is an antioxidant biomarker that is a binding enzyme used in xenobiotics' detoxification. Its activity was evaluated using the protocol mentioned in Habig



Fig. 1. Heavy metals' (Co, Cu, Ni, and Pb) contents measured in the two types of iron ore, namely Hematite ore and Magnetite ore.

et al. (1974). According to these authors, the substrates used are 1-chloro-2,4-dinitrobenzene and glutathione.

Malondialdehyde (MDA) rate is a lipid peroxidation marker of atherogenic lipoproteins. The measurement of MDA content was conducted as described by Haque et al. (2019).

Neurotoxicity was assessed by variations in acetylcholinesterase (AChE) activity, an enzymatic biomarker in charge of acetylcholine hydrolysis. AChE activity was evaluated by spectrophotometry after mixing 5,5'-dithiobis-2-nitrobenzoic acid and thiocholine to produce 5-thio-2-nitrobenzoic acid (Ellman et al. (1961).

2.4. Bioavailability and toxicokinetic properties

Bioavailability and toxicokinetic properties were investigated computationally, based on the physicochemical factors of hematite- and magnetite ore as reported earlier (Badraoui et al., 2023; Jedli et al., 2022; Zammel et al., 2022).

2.5. Data processing

The outcomes of the bioassay conducted were given as average \pm SD. Significant differences between the controls and treatments were tested by one-way ANOVA. Whenever ANOVA outcomes are significant (p less than 0.05), multiple comparisons were made with the aid of Tukey's HSD test. All the data processing was accomplished using the program STATISTICA v.8.0.

3. Results

3.1. Chemistry of iron ore

The presence of high charges of iron, as shown in Table 1, was common in contaminated sites and can cause significant interference with heavy metals. Iron ores, such as hematite and magnetite, were analyzed using the SAA-Flame technique (Fig. 1). The Hematite (H) iron ore contains approximately 68.52 mg/kg of Co, 17.42 mg/kg of Cu, 28.28 mg/kg of Ni, and 26.75 mg/kg of Pb. On the other hand, the Magnetite (M) iron ore contains about 73.73 mg/kg of Co, 21.29 mg/kg of Cu, 43.51 mg/kg of Ni, and 44.59 mg/kg of Pb. The metal concentrations in Hematite-type iron ore microparticles follow thus the decreasing order of Co > Ni > Pb > Cu, with respective concentrations of 68.52 mg/kg, 28.28 mg/kg, 26.75 mg/kg, and 17.42 mg/kg. For Magnetite-type iron ore microparticles, the order is Co > Pb > Ni > Cu, with respective concentrations of 73.73 mg/kg, 44.59 mg/kg, 43.51 mg/kg, and 21.29 mg/kg.

3.2. Biomarker changes in R. decussatus

The activity variations of CAT, GST, and AChE, and levels of MDA, in both gills and digestive glands of *R. decussatus* contaminated by the two types of iron minerals (Hematite and Magnetite), are shown in Figs. 2, 3, 4, and 5, respectively.

3.2.1. Catalase (CAT) activity

Following exposure to two types of iron minerals (Hematite and Magnetite), the effect on CAT activity in clams is presented in Fig. 2. Compared to controls, CAT activity enhanced significantly in the gills and also the digestive glands of *R. decussatus* following the exposure to the two types of ore from concentration C3 (1.5 g/L). *R. decussatus* clams treated with 0.5 and 1 g/L (C1 and C2) revealed that microparticles of both types of iron ore had no significant impact on CAT activity in the two organs considered. However, the presence of in the experimental units 1.5 and 3 g/L (C3 and C4) was followed by a discernible augmentation in CAT activity in both organs for both types of iron ore microparticles.

The effect of magnetite microparticles at concentration C3, showing significant changes, was considerably superior in gills compared to digestive glands, while the effect of hematite microparticles at the same concentration C3 on both organs was similar.

3.2.2. Glutathione S-transferase (GST) activity

At the end of the bioassay, a significant increase was noticed in the GST activities (Fig. 3) in both organs considered for *R. decussatus* contaminated with hematite and magnetite iron ore microparticles compared to controls. For experimental units enriched with hematite microparticles, GST activity increased in comparison to controls in the gills and digestive glands of the European clams starting from C3 (1.5 g/L). For magnetite microparticles, a discernible increase in GST activity at gills was reported starting from C2 (1 g/L), while in the digestive glands, it was revealed starting from concentration C3 (1.5 g/L). The effect of both types of ore microparticles (hematite and magnetite) on GST activity was greater at gills compared to digestive glands.

3.2.3. Malondialdehyde (MDA) levels

For control specimens, the MDA levels showed no noticeable difference in the gills in comparison to the digestive glands (Fig. 4). These low MDA values in the controls confirm the stability of the experimental conditions during the experimental period. No significant difference was recorded in the clams contaminated with low doses (C1 and C2) compared to the controls. This result shows the absence of cellular damage in individuals contaminated with both types of microparticles of magnetite and hematite iron ore during the exposure period. Except for the gills of individuals



Fig. 2. Catalase (CAT) activities in gills (A and B) and digestive glands (A' and B') of *Ruditapes decussatus* contaminated with hematite and magnetite iron ore microparticles at four concentrations (C1-C4). Results are given as average values (n = 5) ± Standard Error (SE); Statistical significance is indicated by different letters (a, b, c).



Fig. 3. Glutathione S-transferase (GST) activities in gills (A and B) and digestive glands (A' and B') of *Ruditapes decussatus* contaminated with hematite and magnetite iron ore microparticles at four concentrations (C1-C4). Results are given as average values (n = 5) ± Standard Error (SE); Statistical significance is indicated by different letters (a, b, c).

contaminated with magnetite-type iron ore where the MDA level began to increase from concentration C2 (1 g/L). Concentration C3 significantly modulated the MDA level at both gills and digestive glands after the two types of micro-particles of magnetite and hematite iron ore were introduced.

3.2.4. Acetylcholinesterase (AChE) activity

The exposure to hematite and magnetite microparticles caused a discernible inhibition in AChE activity (Fig. 5). The AChE activity measured after seven days from the start showed a significant inhibition for concentrations of iron ore microparticles equal to 0.5 g/L (C1), 1 g/L (C2), and 1.5 g/L (C3). Particularly, no significant modification was detected in comparison to the control under the treatment C4 (5 g/L) after a week of exposure.

3.3. In silico findings

The bioavailability and toxicokinetic features of iron ore are given in Table 2. The bioavailability polygons prove that hematite and magnetite ore possessed acceptable oral bioavailability and supported their potential toxicological outcomes. Fig. 6 exhibits that most of the physicochemical properties (flexibility, lipophilicity, etc.) allow iron ore to stand in the most suitable oral bioavailability, except unsaturation. While both pan assay interference structures and Brenk structural alert showed 0 alerts for hematite and magnetite ore, TPSA values were 43.37 and 36.92 respectively.



Fig. 4. Malondialdehyde (MDA) content in gills (A and B) and digestive glands (A' and B') of *Ruditapes decussatus* contaminated with hematite and magnetite iron ore microparticles at four concentrations (C1-C4). Results are given as average values (*n* = 5) ± Standard Error (SE); Statistical significance is indicated by different letters (a, b, c).



Fig. 5. Acetylcholinesterase (AChE) activities in gills (A and B) and digestive glands (A' and B') of *Ruditapes decussatus* contaminated with hematite and magnetite iron ore microparticles at four concentrations (C1-C4). Results are given as average values (*n* = 5) ± Standard Error (SE); Statistical significance is indicated by different letters (a, b, c).

Table 2	
Bioavailability and toxicokinetic properties of hematite ore and magnetite ore.	

Entry / Parameter Hematite ore Magnetite ore Molar Refractivity 2.46 4.34 No. heavy atoms 5 7 0 No. rotatable bonds 2 36.92 TPSA (Å²) 43.37 Pan Assay Interference Structures 0 alert 0 alert Brenk Structural Alert 0 alert 0 alert

4. Discussion

The company SNIM exploits iron ore microparticles that have high metal contents for all measured metals. These results suggest that Magnetite-type iron ore microparticles contain higher metal concentrations than Hematite-type. Several research works confirmed already the detrimental impact of heavy metals on bivalves living in aquatic habitats. These filter-feeding organisms are known to absorb heavy metals from water, as evidenced by Chandurvelan et al. (2015) and Khazri et al. (2019). Once a high Reactive Oxygen Species (ROS) is observed, that is thought to be the cause of the harmful impact of heavy metals (Trevisan et al., 2014; Freitas et al., 2018). To mitigate the damage due to ROS,



Fig. 6. Bioavailability hexagons of hematite ore and magnetite ore. LIPO: Lipophilicity, SIZE: Molecular size, POLAR: Polarity, INSOLU: Insolubility, INSATU: Insaturation, FLEX: Flexibility, The pink area represents the most suitable area for oral bioavailability. Note that regardless insaturation, both hematite ore and magnetite ore stand in the pink area, which is the most suitable for oral bioavailability.

organisms have developed defence tools such as antioxidant enzymes and non-enzymatic antioxidants. However, when ROS levels become too high, an oxidative stress status start, and biomolecule harm to proteins and lipids and have negative impacts on cells. According to the study of Shanker and Aschner (2003), the existence of metals in aquatic habitats may cause a significant modification of ROS and lead to oxidative stress. The current experiment aimed to examine the effects of exposure to hematite and magnetite iron ore microparticles on diverse biochemical markers linked to oxidative stress. For this purpose, the R. decussatus clams were considered as a biological model to evaluate the harmful impacts of iron ore microparticles on specific biomarkers. The detection of changes in markers of oxidative stress (enzymatic or not) within this bivalve indicates its exposure to this kind of pollutant, implying a potential hazard arising from its toxic properties. CAT, an enzyme, aids in the elimination of hydrogen peroxide (H₂O₂) that arises during oxidative stress in cells, according to multiple studies (Modesto and Martinez, 2010, and references herein). Its function is to accelerate the translation of H₂O₂ to H₂O and O₂ (Faria et al., 2010). Heavy metals can stimulate CAT and, thus, may be used as an oxidative stress biomarker (Ozkan et al., 2017). The results presented here indicate a discernible augmentation in the CAT activity in both organs tested (gills and digestive glands) of R. decussatus after the enrichment of its close environment with ore microparticles compared to controls (see Fig. 2). The noticeable susceptibility of mussels' gills, as opposed to their digestive gland, to stress caused by heavy metals, can be clarified by the fact that gills serve as the primary organs through which these invertebrates interact with water. They perform essential functions such as filtration, respiration, and ion exchange, and act as the primary interface for contaminants. The distinct reactions observed in this study among mussels exposed to varying concentrations provide a novel understanding regarding the species' environmental adaptability mechanisms, particularly concerning heavy metals, as well as the interplay between these changes and the levels of exposure. The outcomes of the bioassay conducted are in accordance with the conclusions of Valko et al. (2005) that the presence of metals can cause oxidative stress. Proportionality between the levels of copper present in the iron ore microparticles and catalase activity was also observed, suggesting that the augmentation in this activity followed exposure to copper. Copper has been previously identified as a toxic metal by Geracitano and Monserrat (2002) and Ferreira-Cravo et al. (2009). The discernible augmentation in CAT noticed at a C3 concentration resulted from the production of H₂O₂ as part of the antioxidant protection mechanisms that aim to defend cells against oxidative damage. Metal particles can cause oxidative stress by crossing the cell membrane barrier (Vale et al., 2016), leading to an inequality in pathways of oxidants and antioxidants (Falfushynska et al., 2016). The antioxidant phase II defence mechanism is characterized by the presence of GST enzymes, which are known for their substrates. According to the reports by Damiens et al. (2007) and Durou et al. (2007), GSTs are efficient to detoxify aquatic organisms with various xenobiotics (metals PAHs, PCBs, etc.). In D. trunculus living on the coast of Taghazout, a clear increase in GST activity was noticed, which is probably due to the presence of metals that stimulate the antioxidant protection processes (Radwan et al., 2010). This suggests that the detoxification pathway was induced by oxidative forces controlled by this enzyme, as reported by Elia et al. (2007). The presence of toxic metals and persistent organic xenobiotics can trigger the activation of GST, whether in the field- (Durou et al., 2007) or in laboratory conditions (Livingstone et al., 1990). Nevertheless, in the case of exposure to iron ore microparticles, the noticeable increase in GST activity compared to controls (Fig. 4) is considered a protective response against oxidative stress. According to Valavanidis et al. (2006), when there is too much ROS, it can overpower antioxidant systems and cause oxidative stress. This status can lead to an increase in lipid peroxidation at cell membranes, as noted in their study. The considerable elevation of MDA levels in the gills indicates that heavy metals induce cellular damage via a mechanism involving free radicals. The oxidative harm in the gills could additionally hinder the respiratory mechanisms by disrupting the normal functioning of these organs. Consequently, it implies that more energy is required to maintain homeostasis, particularly concerning osmotic balance, and there are alterations in the metabolic rate associated with the production of pro-oxidants. In relation to R. decussatus clams that were studied here within, it was found that exposure to both types of ore microparticles (hematite and magnetite) was followed by a discernible augmentation in lipid peroxidation at both organs considered after seven days (Fig. 5). The concentration of microparticles also correlated with an intensification in lipid peroxidation in these tissues. Couillard et al. (1995) found similar results when studying transplanted P. grandis in the field after exposure to heavy metals. Siwela et al. (2010) demonstrated that exposure to contaminated sediments and water with (Pb, Cu, Zn, Cd, Fe, and Ni) was followed by a significant augmentation in lipid peroxidation in the common pond snail Lymnaea

natalensis. Acetylcholinesterase plays a crucial role in nerve signal transmission in living organisms by breaking down acetylcholine in the synaptic gap and ending its excitatory effect on the postsynaptic membrane. This helps to ensure that nerve signals are transmitted normally. According to Soreq and Seidman (2001), AChE is an enzyme with a central character in the neurotransmission process. Inhibition of AChE, as stated by Kuhr and Dorough (1976), may result in a variety of behavioral changes, such as hyperactivity, loss of coordination, convulsions, and paralysis. The study assessed how the AChE activity in *Mytella charruana* was impacted by various ionic forms of heavy metals, including As³⁺, Hg²⁺, Cd²⁺, Cu²⁺, Pb²⁺, and Zn²⁺. The results indicated that the inhibitory power of the metals on AChE in *M. charruana* followed a decreasing ranking: $Hg^{2+} > Pb^{2+} > Cd^{2+} > As^{3+} > Cu^{2+} > Zn^{2+}$, as evidenced by their respective IC₅₀ values (Dos Santos et al., 2022). As shown in Fig. 3, exposure to hematite and magnetite microparticles caused a change in the AChE activity of the European clam. AChE activity measured a week from the start of the current experiment showed significant inhibition for concentrations less than 1.5 g/L (C1, C2, and C3). The reduction of AChE activity is triggered by the presence of heavy metals, which have been found to have an inhibitory effect on this enzyme in various species, including common carpCyprinus carpio (Suresh et al., 1992) and the green crab Carcinus maenas (Elumalai et al., 2007), as consistent with the outcomes of the current bioassay. The negative impact on the health of clams, as reported by disruption in the assessed biomarkers, MDA levels, and decreased AChE activity might be the consequence of the bioavailability and toxicokinetic properties of iron ore. Such results might include interactions with the cytochrome P450 (Cyp) isoforms such as Cyp1A2, Cyp3A4, and CypC19... (Ishak et al., 2022, Badraoui et al., 2023). Interactions with Cyp are commonly assessed in toxicological approaches (Badraoui et al., 2023; Hedfi et al., 2022) as they are usually associated with mild to severe toxicological outcomes (Allouche et al., 2022; Hedfi et al., 2022).

5. Conclusions

This study provides a comprehensive summary of the initial findings regarding biomarkers observed in the gills and digestive glands of *R. decussatus* when the habitat was enriched with microparticles of iron ore, including hematite and magnetite. The results clearly demonstrate that exposure to these microparticles had detrimental effects on the health of the clams, as indicated by significant alterations in biomarker activities and the MDA rate. Specifically, concentrations C3 and C4 prominently stimulated GST, CAT, and MDA levels in both organs examined. However, a decrease in AChE activity was noticeable at the lowest tested concentration, C1, for both types of ore. The biomarker responses observed in clams exposed to both types of ore differed from those of the control group, and the impact of the ore was more pronounced on the gills compared to the digestive glands under both conditions.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgments

The authors extend their appreciation to the Princess Nourah bint Abdulrahman University Researchers Supporting Project number (PNURSP2023R437), Princess Nourah bint Abdulrahman University, Riyadh, Saudi Arabia.

References

- Achary, M.S., Satpathy, K., Panigrahi, S., Mohanty, A., Padhi, R., Biswas, S., Prabhu, R., Vijayalakshmi, S., Panigrahy, R.J.F.C., 2017. Concentration of heavy metals in the food chain components of the nearshore coastal waters of Kalpakkam, southeast coast of India. Food Control 72, 232–243.
- Adamson, I., Prieditis, H., Hedgecock, C., Vincent, R., 2000. Zinc is the toxic factor in the lung response to an atmospheric particulate sample. Toxicol. Appl. Pharmacol. 166, 111–119.
- Aebi, H., 1974. Catalase. In: Bergmayer, H.U. (Ed.), Methods of Enzymatic Analysis. Academic Press, London, pp. 671–684.
- Allouche, M., Ishak, S., Ben Ali, M., Hedfi, A., Almalki, M., Karachle, P.K., et al., 2022. Molecular interactions of polyvinyl chloride microplastics and beta-blockers (diltiazem and bisoprolol) and their effects on marine meiofauna: combined *in vivo* and modeling study. J. Hazard. Mat. 431, 128609.
- Badraoui, R., Allouche, M., El Ouaer, D., Siddiqui, A.J., Ishak, S., Hedfi, A., Beyrem, H., Pacioglu, O., Rudayni, H.A., Boufahja, F., 2023. Ecotoxicity of chrysene and phenanthrene on meiobenthic nematodes with a case study of *Terschellingia longicaudata*: Taxonomics, toxicokinetics, and molecular interactions modelling, Environ. Pollut. 316, 120459.
- Berrow, M.L., Mitchell, R.L., 1991. Location of trace elements in soil profiles: total contents of particles-size separates. Earth Sci. Trans. Roy. Soc. Edin. Earth Sci. 82, 195–209.
- Bilal, S., 2017. Contribution à l'étude de la qualité des eaux du Banc d'Arguin (Mauritanie) : Dosage des métaux lourds chez les Mollusques (Anadara senilis), les Poissons (Tilapia : Sarotherodon melanotheron) et dans le Sédiment. Ibn Tofail, Maroc, Faculty of Sciences of Kénitra.
- Bradford, M.M., 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal. Biochem. 72, 248–254.
- Carlsson, E., Thunberg, J., Ohlander, B., Holmstrom, H., 2002. Sequential extraction of sulfide-rich tailings remediated by the application of till cover, Kristineberg mine, northern Sweden. Sci. Total Environ. 299 (1–3), 207–226.
- Chandurvelan, R., Marsden, I.D., Glover, C.N., Gaw, S., 2015. Assessment of a mussel as a metal bioindicator of coastal contamination: relationships between metal bioaccumulation and multiple biomarker responses. Sci. Total Environ. 511, 663–675.
- Colbeck, I. 1995. Particle emission from outdoor and indoor sources. In Airbone particulate matter; Samara, K., Ed. P 1-34.
- Couillard, Y., Campbell, P., Auclair, J., Pellerin-Massicotte, J., 1995. Field transplantation of a freshwater bivalve Pyganodon grandis, across a metal contamination gradient. II. Metallothionein response to Cd and Zn exposure, evidence for cytotoxicity, and links to effects at higher levels of biological organization. Can. J. Fish. Aquat. Sci. 52 (4), 703–715.
- Damiens, G., Gnassia-Barelli, M., Loquès, F., Roméo, M., Salbert, V., 2007. Integrated biomarker response index as a useful tool for environmental assessment evaluated using transplanted mussels. Chemosphere 66 (3), 574–583.
- Dos Santos, G.P.C., de Assis, C.R.D., Oliveira, V.M., Cahu, T.B., Silva, V.L., Santos, J.F., Yogui, G.T., Bezerra, R.S., 2022. Acetylcholinesterase from the charru mussel *Mytella charruana*: Kinetic characterization, physicochemical properties and potential as in vitro biomarker in environmental monitoring of mollusk extraction areas. Comp. Biochem. Physiol. C: Toxicol. Pharmacol. 252, 109225.
- Durou, C., Poirier, L., Amiard, J.C., Budzinski, H., Gnassia-Barelli, M., Lemenach, K., et al., 2007. Biomonitoring in a clean and a multi-contaminated estuary based on biomarkers and chemical analyses in the endobenthic worm *Nereis diversicolor*. Environ. Pollut. 148 (2), 445–458.
- Duruibe, J.O., Ogwuegbu, M., Egwurugwu, J.J.I.J., 2007. Heavy metal pollution and human biotoxic effects. Int. J. Phys. Sci. 2 (5), 112–118.
- Elia, A.C., Galarini, R., Dörr, A.J.M., Taticchi, M.I., 2007. Heavy metal contamination and antioxidant response of a freshwater bryozoan (*Lophopus crystallinus* Pall., Phylactolaemata). Ecotoxicol. Environ. Saf. 66 (2), 188–194.
- Ellman, G.L., Courtney, K.D., Andres, V., Featherstone, R.M., 1961. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem. Pharmacol. 7 (2), 88–95.
- Elumalai, M., Antunes, C., Guilhermino, L., 2007. Enzymatic biomarkers in the crab Carcinus maenas from the Minho River estuary (NW Portugal) exposed to zinc and mercury. Chemosphere 66, 1249–1255.
- Embile Jr., R.F., Walder, I.F., Schuh, C., Donatelli, J.L., 2018. Cu, Pb and Fe release from sulfide-containing tailings in seawater: results from laboratory simulation of submarine tailings disposal. Mar. Pollut. Bull. 137, 582–592.
- Falfushynska, H., Gnatyshyna, L., Fedoruk, O., Sokolova, I.M., Stoliar, O., 2016. Endocrine activities and cellular stress responses in the marsh frog *Pelophylax ridibundus* exposed to cobalt, zinc and their organic nanocomplexes. Aquat. Toxicol. 170, 62–71.
- Faria, M., Huertas, D., Soto, D.X., Grimalt, J.O., Catalan, J., Riva, M.C., Barata, C., 2010. Contaminant accumulation and multi-biomarker responses in feld collected zebra mussels (*Dreissena polymorpha*) and crayfsh (*Procambarus clarkii*), to evaluate toxicological efects of industrial hazardous dumps in the Ebro River (NE Spain). Chemosphere 78 (3), 232–240.
- Ferreira-Cravo, M., Ventura-Lima, J., Sandrini, J.Z., Amado, L.L., Geracitano, L.A., Rebelo, M., Monserrat, J.M., et al., 2009. Antioxidant responses in different body

M. Aillal, A. Khazri, N. Al-Hoshani et al.

regions of the polychaeta *Laeonereis acuta* (Nereididae) exposed to copper. Ecotoxicol. Environ. Saf. 72 (2), 388–393.

- Freitas, R., Coppola, F., Marchi, L.D., Codela, V., Pretti, C., Chiellini, F., Morelli, A., Polese, G., Soares, A.M.V.M., Figueira, E., 2018. The influence of Arsenic on the toxicity of carbon nanoparticles in bivalves. J. Hazard. Mat. 358, 484–493.
- Geracitano, L., Monserrat, J.M., Bianchini, A., 2002. Physiological and antioxidant enzyme responses to acute and chronic exposure of *Laeonereis acuta* (Polychaeta, Nereididae) to copper. J. Exp. Mar. Biol. Ecol. 277 (2), 145–156.
- Habig, W.H., Pabst, M.J., Jakoby, W.B., 1974. Glutathione S-transferases. The first enzymatic step in mercapturic acid formation. J. Biol. Chem. 25 (249), 7130– 7139.
- Haque, M.N., Eom, H.J., Nam, S.E., Shin, Y., Rhee, J.S., 2019. Chlorothalonil induces oxidative stress and reduces enzymatic activities of Na+ /K+ - ATPase and acetylcholinesterase in gill tissues of marine bivalves. PLoS ONE 14 (4), e0214236.
- Hedfi, A., Ali, M.B., Korkobi, M., Allouche, M., Harrath, A.H., Beyrem, H., et al., 2022. The exposure to polyvinyl chloride microplastics and chrysene induces multiple changes in the structure and functionality of marine meiobenthic communities. J. Hazard. Mat. 436, 129161.
- IMROP, 2019. 9ème édition du groupe de travail de l'Institut Mauritanien sur l'Aménagement des ressources halieutiques et gestion de la biodiversité au service du développement durable. 32 p.
- Ishak, S., Allouche, M., Hedfi, A., Ben Ali, M., Albogami, B., Almalki, M., Plavan, G., et al., 2022. Effect of the antidepressant paroxetine on migratory behaviour of meiobenthic nematodes: computational and open experimental microcosm approach. Mar. Pollut. Bull. 177, 113558.
- Jedli, O., Ben-Nasr, H., Zammel, N., Rebai, T., Saoudi, M., Elkahoui, S., et al. 2022. Attenation of ovalbumin-induced inflammation and lung oxidative injury in asthamtic rats by Zingiber officinale extract : combined in silico and in vivo study on antioxidant potential, STAT6 and TNF- pathways. 3 Biotech. 12(9), 191.
- Jordi, P., Robert, B. F., Francesco, O., Jordi, U., Josep, M.S., Jordi, C., Bernhard, D., 2021. Release of trace elements during bioreductive dissolution of magnetite from metal mine tailings: Potential impact on marine environments. pp. 9.
- Khanam, S.J.T.B.B.S., 2014. Heavy metal contamination and health hazards: a review. Trends Biotechnol. Biol. Sci. 1, 5–8.
- Khazri, A., Mezni, A., Sellami, B., Touaylia, S., Noiyri, I., Beyrem, H., Mahmoudi, E., 2019. Protective properties of filamentous blue–green alga Spirulina against the oxidative stress induced by cadmium in freshwater mussel *Unio ravoisieri*. Chem. Ecol. 35 (9), 825–834.
- Kuhr, R.J., Dorough, H.W., 1976. Carbamate insecticides: chemistry, biochemistry, and toxicology. CRC Press, Inc., p. 301.Kumar, K.A., Achyuthan, H.J.J., 2007. Heavy metal accumulation in certain marine
- Kumar, K.A., Achyuthan, H.J.J., 2007. Heavy metal accumulation in certain marine animals along the East Coast of Chennai, Tamil Nadu, India. J. Environ. Biol. 28, 637–643.
- Legraa, M.E.H.M., 2019. Evaluation de la vulnérabilité des côtes de la Mauritanie face aux pollutions. Thèse de doctorat, 255.
- Livingstone, D.R., Martinez, P.G., Michel, X., Narbonne, J.F., O'hara, S., Ribera, D., Winston, G.W., 1990. Oxyradical production as a pollution mediated mechanism of toxicity in the common mussel, *Mytilus edulis* L, and other molluscs. Funct. Ecol. 4(3),415–424.
- Lottermoser, B.G., 2010. Mine Wastes: Characterization, Treatment and Environmental Impacts, Mine Wastes: Characterization, Treatment and Environmental Impacts. Springer-Verlag, Berlin, Berlin, p. 400.
- Manceau, A., Lanson, B., Drits, V.A., 2002. Structure of heavy metal sorbed birnessite. Part III: Results from powder and polarized extended X-ray absorption fine structure spectroscopy. Geochim. Cosmochim. Acta 66, 2639–2663.
- Matías, M.S.A., 2008. Contamination en métaux lourds des eaux de surface et des sédiments du Val de Milluni (Andes Boliviennes) par des déchets miniers. Approches géochimique, minéralogique et hydrochimique. PhD thesis. pp. 378.
- Mendes, B.C., Pedroti, L.G., Fontes, M.P.F., Ribeiro, J.C.L., Vieira, C.M.F., Pacheco, A.A., Azevedo, A.R.G., 2019. Technical and environmental assessment of the incorporation of iron ore tailings in construction clay bricks. Constr. Build. Mater. 227, 116669.
- Modesto, K.A., Martinez, C.B.R., 2010. Roundup causes oxidative stress in liver and inhibits acetylcholinesterase in muscle and brain of the fish Prochilodus lineatus. Chemosphere 78, 294–299.
- Morello, E.B., Haywood, M.D.E., Brewer, D.T., Apte, S.C., Asmund, G., Kwong, Y.T.J., Dennis, D., 2016. The ecological impacts of submarine tailings placement. In: Hughes, R.N., Hughes, D.J., Smith, I.P., Dale, A.C. (Eds.), Oceanography and Marine Biology: An Annual Review. Oceanogr. Mar. Biol. 54, 315-366.
- Mulvad, G., Pedersen, H., Hansen, J., Dewailly, E., Jul, E., Pedersen, M., Bjerregaard, P., Malcom, G., Deguchi, Y., Middaugh, J.J.S., 1996. Exposure of Greenlandic Inuit to organochlorines and heavy metals through the marine food-chain: an international study. Sci. Total Environ. 186 (1–2), 137–139.

- Oliveira, P., Lírio, A.V., Canhoto, C., Guilhermino, L., 2018. Toxicity of mercury and post-exposure recovery in *Corbicula fluminea*: neurotoxicity, oxidative stress and oxygen consumption. Ecol. Indicat. 91, 503–510.
- Ozkan, D., Dagdeviren, M., Katalay, S., Guner, A., Yavasoglu, N.U.K., 2017. Multibiomarker responses after exposure to pollution in the Mediterranean mussels (*Mytilus galloprovincialis* L.) in the Aegean Coast of Turkey. Bull. Environ. Contam. Toxicol. 98, 46–52.
- Pedersen, K.B., Jensen, P.E., Sternal, B., Ottosen, L.M., Henning, M.V., Kudahl, M.M., Junttila, J., Skirbekk, K., Frantzen, M., 2018. Long-term dispersion and availability of metals from submarine mine tailing disposal in a fjord in Arctic Norway. Environ. Sci. Pollut. Res. Int. 25, 32901–32912.
- Radwan, M.A., El-Gendy, K.S., Gad, A.F., 2010. Biomarkers of oxidative stress in the land snail, Theba pisana for assessing ecotoxicological efects of urban metal pollution. Chemosphere 79 (1), 40–46.
- Ramirez-Llodra, E., Trannum, H.C., Evenset, A., Levin, L.A., Andersson, M., Finne, T.E., Hilario, A., Flem, B., Christensen, G., Schaanning, M., Vanreusel, A., 2015. Submarine and deep-sea mine tailing placements: a review of current practices, environmental issues, natural analogs and knowledge gaps in Norway and internationally. Mar. Pollut. Bull 97, 13–35.
- Sammut, M., 2007. Spéciation du cadmium, du plomb et du zinc dans les poussières d'émissions atmosphériques d'origine sidérurgique Approche de l'impact toxicologique des poussières. Paul Cezanne university, p. 254. PhD thesis.
- Sellami, B., Mezni, A., Khazri, A., Bouzidi, I., Saidani, W., Sheehan, D., Beyrem, H., 2017. Toxicity assessment of ZnO-decorated Au nanoparticles in the Mediterranean clam Ruditapes decussatus. Aquat. Toxicol. 188, 10–19.
- Shanker, G., Syversen, T., Aschner, M., 2003. Astrocyte-mediated methylmercury neurotoxicity. Biol. Trace Elem. Res. 95, 1–10.
- Sidoumou, Z., Gnassia-Barelli, M., Roméo, M., 1999. Étude des métaux lourds chez deux mollusques prélevés sur la cote mauritanienne, *Crassostrea gigas* et *Perna perna*. J. Rech. Océan. 24, 13–18.
- Simpson, S.L., Spadaro, D.A., 2016. Bioavailability and chronic toxicity of metal sulfide minerals to benthic marine invertebrates: implications for Deep Sea exploration, mining and tailings disposal. Environ. Sci. Technol. 50, 4061–4070.
- Siwela, A.H., Nyathi, C.B., Naik, Y.S., 2010. A comparison of metal levels and antioxidant enzymes in freshwater snails, Lymnaea natalensis, exposed to sediment and water collected fromWright Dam and Lower Mguza Dam, Bulawayo. Zimbabwe. Ecotoxicol. Environ. Saf. 73 (7), 1728–1732.
- Sobral, P., Widdows, J., 1997. Effects of elevated temperatures on the scope for growth and resistance to air exposure of the clam *Ruditapes decussatus* (L.) from southern Portugal. Sci. Mar. 61, 163–171.
- Soreq, H., Seidman, S., 2001. Acetylcholinesterase-new roles for an old actor. Nat. Rev. Neurosci. 2, 294–302.
- Sternal, B., Junttila, J., Skirbekk, K., Forwick, M., Carroll, J., Pedersen, K.B., 2017. The impact of submarine copper mine tailing disposal from the 1970s on Repparfjorden, northern Norway. Mar. Pollut. Bull. 120, 136–153.
- Suresh, A., Sivaramakrishna, B., Victoriamma, P.C., Radhakrishnaiah, K., 1992. Comparative study on the inhibition of acetylcholinesterase activity in the freshwater fish *Cyprinus carpio* by mercury and zinc. Biochem. Int. 26 (2), 367– 375.
- Trevisan, R., Delapedra, G., Mello, D.F., Arl, M., Schmidt, E.C., Meder, F., Monopoli, M., Cargnin-Ferreira, E., Bouzon, Z.L., Fisher, A.S., Sheehan, D., Dafre, A.L., 2014. Gills are an initial target of zinc oxide nanoparticles in oysters Crassostrea gigas, leading to mitochondrial disruption and oxidative stress. Aqua. Toxicol. 153, 27–38.
- Valavanidis, A., Vlahogianni, T., Dassenakis, M., Scoullos, M., 2006. Molecular biomarkers of oxidative stress in aquatic organisms in relation to toxic environmental pollutants. Ecotoxicol. Environ. Saf. 64 (2), 178–189.
- Vale, G., Mehennaoui, K., Cambier, S., Libralato, G., Jomini, S., Domingos, R.F., 2016. Manufactured nanoparticles in the aquatic environment-biochemical responses on freshwater organisms: a critical overview. Aquat. Toxicol. 170, 162–174.
- Valko, M.M.H.C.M., Morris, H., Cronin, M.T.D., 2005. Metals, toxicity and oxidative stress. Curr. Med. Chem. 12 (10), 1161–1208.
 Volland, M., Blasco, J., Hampel, M., 2017. Validation of reference genes for RT-qPCR
- Volland, M., Blasco, J., Hampel, M., 2017. Validation of reference genes for RT-qPCR in marine bivalve ecotoxicology: systematic review and case study using copper treated primary Ruditapes philippinarum hemocytes. Aquat. Toxicol. 185, 86– 94.
- WA Iron Ore profile, 2019. Western Australia Iron Ore Profile. Western Australian Department of State Development, Australia Available in. https://www.jtsi. wa.gov.au/docs/ default-source/default-document-library/wa-iron-ore-profile-0519.pdf?sfvrsn= db8701c_4.
- Younger, P.L., Banwart, S.A., Hedin, R.S., 2002. Mine water hydrology, pollution, remediation. Springer, Netherlands, p. 63.
- Zammel, N., Jedli, O., Rebai, T., Hamadou, W.S., Elkahoui, S., Jamal, A., Alam, J.M., et al. 2022. Kidney injury and oxidative damage alleviation by Zingiber officinale: Phramacokinetics and protective approach in a combined murine model of osteoporosis. 3 Biotech. 12(5), 112.