

Downregulated lncRNA SNHG18 Suppresses the Progression of Hepatitis B Virus-Associated Hepatocellular Carcinoma and Mediates the Antitumor Effect of Oleanolic Acid

Wenqi Song^{1,*}, Xuemei Zhang^{2,*}, Lin Feng¹, Yongwei Lai¹, Tongtong Li¹, Pengxia Zhang¹

¹ Key Laboratory of Microecology-immune Regulatory Network and Related Diseases, School of Basic Medicine, Jiamusi University, Jiamusi, Heilongjiang, People's Republic of China; ²Department of Gastroenterology, The First Affiliated Hospital of Jiamusi University, Jiamusi, Heilongjiang, People's Republic of China

*These authors contributed equally to this work

Correspondence: Pengxia Zhang, Key Laboratory of Microecology-immune Regulatory Network and Related Diseases, School of Basic Medicine, Jiamusi University, 148 Xuefu Street, Jiamusi, Heilongjiang, 154000, People's Republic of China, Email zhangpengxiajms@163.com

Purpose: Oleanolic acid (OA) has been widely reported to possess antitumor effects, but the specific molecular mechanism underlying its inhibition of hepatocellular carcinoma (HCC) progression remains unclear. This study aims to uncover the mechanism of OA antitumor effect on HBV-associated HCC and identify a potential biomarker for tumor progression.

Patients and Methods: The effect of OA on major cellular processes of HBV-associated HCC cells was evaluated by CCK8 and Transwell assay. The potential molecular mechanism was assessed by cell transfection. This study also enrolled 111 HCC patients infected with HBV to evaluate the prognostic potential of lncRNA SNHG18 (SNHG18) in HBV-associated HCC.

Results: The inhibitory effect of OA was observed in the critical cellular processes of HBV-associated HCC cells, which depend on OA concentration. Downregulated SNHG18 in HBV-associated HCC was demonstrated to be involved in disease development and predict patients' prognosis. The downregulation of SNHG18 dramatically promoted cellular processes of HBV-associated HCC could reverse the inhibitory effect of OA.

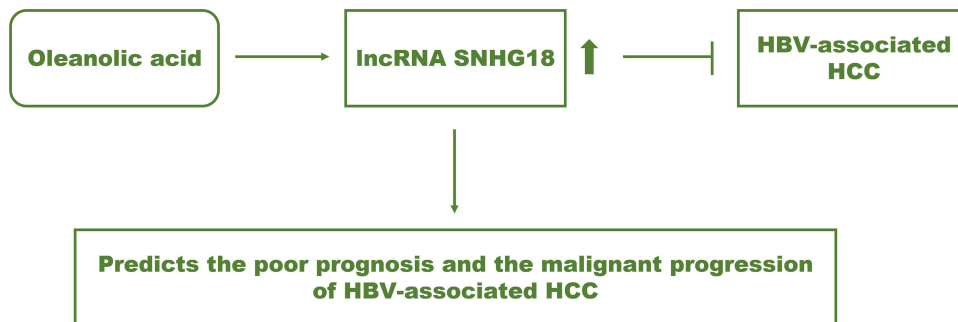
Conclusion: SNHG18 served as a tumor suppressor and prognostic biomarker of HBV-associated HCC. Enhancing SNHG18 might be the mechanism underlying the antitumor effect of OA in HBV-associated HCC.

Keywords: HBV, HCC, oleanolic acid, lncRNA SNHG18, prognosis, progression

Introduction

Hepatocellular carcinoma (HCC) is a frequent type of liver cancer and is also one of the most common causes of cancer-related mortality. Chronic hepatitis B virus (HBV) infection is closely correlated with HCC onset and progression.¹ Over 80% of HCC patients are HBV carriers in China.² The lack of specific symptoms results in the hysteresis of HBV-associated HCC diagnosis and the unsatisfying 5-year overall survival of patients. Currently, the screening of alpha-fetoprotein (AFP) is the major detection of HCC, however, the sensitivity of AFP was not enough to distinguish HCC, especially for the patients with negative AFP or low AFP levels.^{3,4} Therefore, it is of an urgent need to explore accessory molecules that could indicate the onset and development of HCC.

Oleanolic acid (OA), a pentacyclic triterpenoid, has been demonstrated to be widely distributed in various herbs and possessed several pharmacological activities, including anti-inflammation and antitumor.⁵ For example, the inhibitory effect of OA on tumor progression of osteosarcoma was observed and was speculated to be a result of regulating mTOR signaling.⁶ In HCC, OA was previously reported to suppress epithelial-mesenchymal transition and induce cell apoptosis and autophagy.^{7,8} The mechanism investigation would benefit the clinical application of OA in tumor treatment.

Graphical Abstract

It has been reviewed that the development of HBV-associated HCC involved a variety of molecules, including differently expressed non-coding RNAs.⁹ Previously, regulating corresponding molecular biomarkers is always considered as the pathway of the different active ingredients in herbs during their antitumor effects. OA was reported to cause the elevated expression of miR-122 in non-small cell lung cancer, which enhanced the suppression of cell proliferation by OA.¹⁰ Increasing interests have been paid to long non-coding RNAs (lncRNAs) in recent tumor research, which illustrated its significant role in disease diagnosis, prognosis, and development of human cancers as well as the pharmacological effect of various drugs or herbs. For instance, the alleviated effect of OA on arteriosclerosis was revealed to result from the elevation of lincRNA-p21.¹¹ lncRNA SNHG18 (SNHG18) was reported to be a potential diagnosis indicator of HCC with a significant downregulation compared with normal samples.¹² The involvement of SNHG18 has been reported in various malignant tumors, such as glioma and myeloma.^{13,14} However, the specific function of SNHG18 in HBV-associated HCC and whether it mediated the antitumor effect of OA remains unclear.

Here, the function of SNHG18 in HBV-associated HCC was investigated and evaluated its role during the inhibition of tumor progression by OA.

Materials and Methods**Cell Culture**

Four HCC cell lines with an HBV transformant, HepG2/2.2.15, SNU182, MHCC97H, PLC8024, and a normal human liver cell L02 were obtained from ATCC. The DMEM culture medium (Invitrogen, USA) with 10% FBS was used for the incubation of HepG2.2.15, MHCC97H, PLC8024, and L02 cells, while the SNU182 cell was cultured in the RPMI 1640 culture medium (GIBCO, USA). Cell incubation was conducted at 37°C with 5% CO₂.

Cell Treatment and Cell Transfection

After 24 h of cell culture, the cultured cells were treated with 30 µg/mL OA or transfected with pcDNA3.1-SNHG18 with the employment of Lipofectamine 2000 (Invitrogen, USA). The transfection efficiency was evaluated by SNHG18 expression with quantitative PCR.

Cell Proliferation Assessment

The proliferation of treated cells (5×10^4 cells/well) was assessed in the 96-well plates supplied with a completed corresponding culture medium incubating at 37°C for 0, 24, 48, and 72 h followed by adding CCK8 (Dojindo, Japan) to each well. OD450 of each well was detected with a microplate reader as the indicator of cell proliferation.

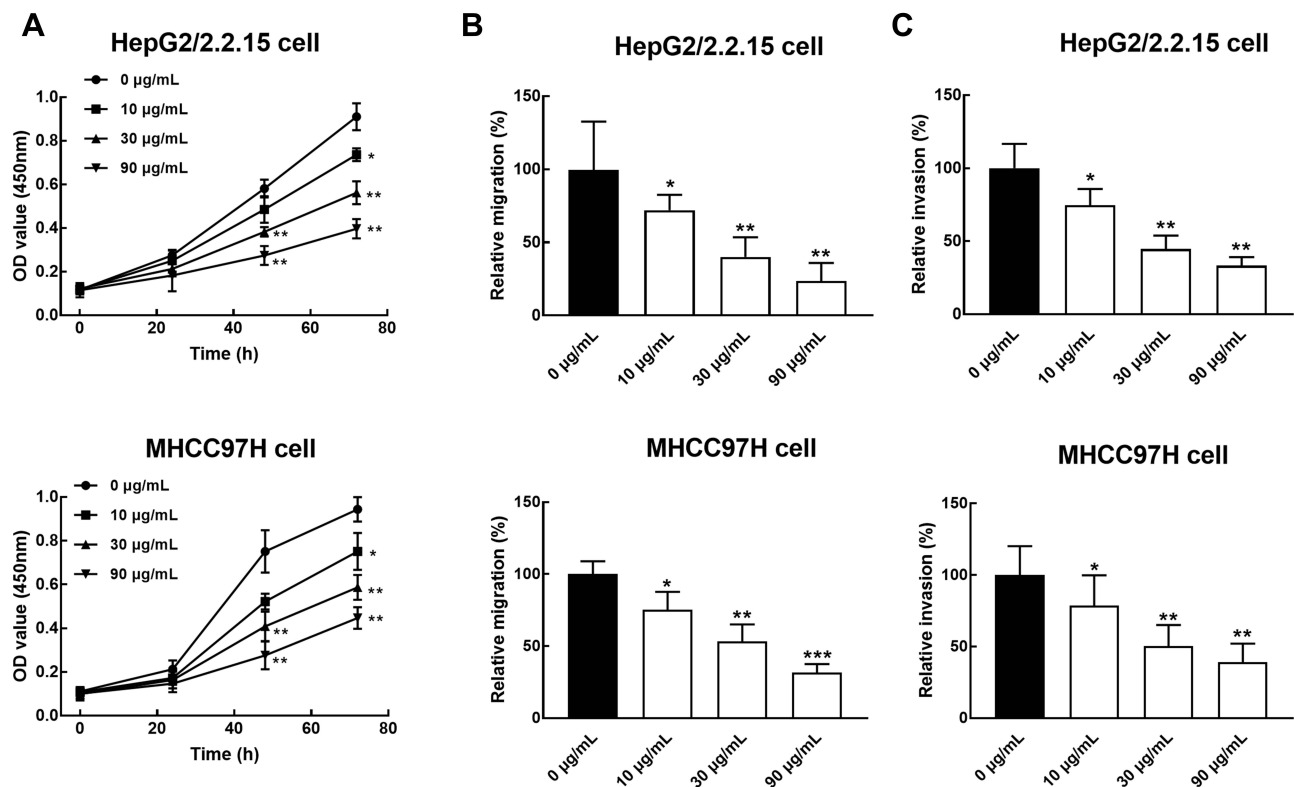


Figure 1 OA showed significant antitumor effects on HBV-associated HCC cells. The proliferation (A), migration (B), and invasion (C) of HepG2/2.2.15 and HCC97H cells were significantly suppressed by OA, and the inhibition was enhanced with the increase of OA concentration. **p* < 0.05, ***p* < 0.01, ****p* < 0.001.

Cell Migration and Invasion Evaluation

Treated cells (5×10^4 cells/well) were seeded into the upper chambers of the 24-well plates (Matrigel chambers for invasion evaluation, Corning, USA). The seeded cells were supplied with an FBS-free culture medium and maintained at 37°C for 24 h. Then, the cells on the bottom surface were fixed with and stained for 30 min. The number of cells was evaluated with a microscope.

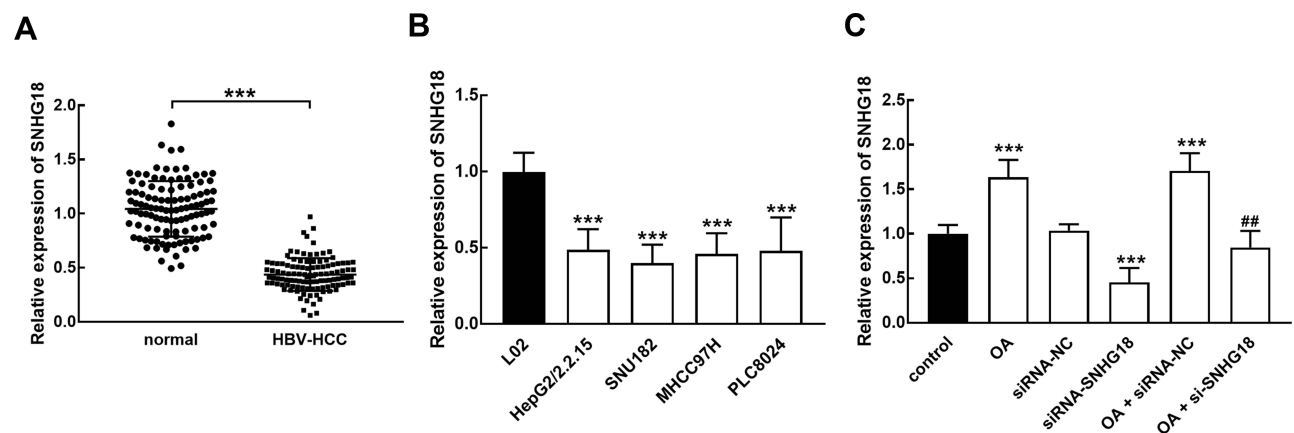


Figure 2 SNHG18 expression in HBV-associated HCC. (A and B) The significant downregulation of SNHG18 was observed in HBV-associated HCC tissues (A) and cells (B). (C) OA dramatically enhanced the expression of SNHG18 and reversed its knockdown induced by siRNA transfection. ****p* < 0.001 relative to the normal tissues and cells. ##*p* < 0.01 relative to the transfection of siRNA-SNHG18.

SNHG18 Expression Evaluation

The expression of SNHG18 was estimated by quantitative PCR with the help of Bio-Rad CFX96 (Bio-Rad Lab, USA) using SYBR Green. Total RNA was extracted and reversed to cDNA with Superscript II (Thermo Scientific, USA). The SNHG18 expression level was calculated by the $2^{-\Delta\Delta CT}$ method with GAPDH as the internal reference.

HBV-Associated HCC Tissues

One hundred and eleven pairs of tumor and matched normal tissues were obtained from HBV-infected HCC patients. All patients were diagnosed and received surgical treatment at The First Affiliated Hospital of Jiamusi University from 2013–2015. The included patients had never received any antitumor therapy before the enrollment. The collected tissues were stored in liquid nitrogen and frozen at -80°C until further analyses. Informed consent was obtained from all individual participants included in the study and all patients were followed up for 5 years to obtain their survival status. This study was performed according to the principles of the Declaration of Helsinki and approved by the Ethics Committee of The First Affiliated Hospital of Jiamusi University.

Table I Association Between lncRNA SNHG18 and Clinicopathological Features of HBV-Associated HCC Patients

	Cases (n = 111)	Expression of lncRNA SNHG18		P value
		High (n = 53)	Low (n = 58)	
Age (years)				0.644
≤ 55	54	27	27	
>55	57	26	31	
Gender				0.347
Male	74	33	41	
Female	37	20	17	
AFP (ng/mL)				0.012*
≤ 20	53	27	26	
>20	58	26	32	
Tumor size (cm)				0.155
≤ 5	55	30	25	
>5	56	23	33	
TNM stage				0.027*
I–II	79	43	36	
III	32	10	22	
Differentiation				0.182
Well + moderate	77	40	37	
Poor	34	13	21	
Liver cirrhosis				0.252
Negative	30	17	13	
Positive	81	36	45	

Notes: * $P < 0.05$ indicates the significant association.

Abbreviation: AFP, α -fetoprotein.

Statistical Analysis

All experiments were conducted in triplicate independent repeats. Obtained data were presented as mean value \pm SD. and analyzed by GraphPad Primer 7.0 (GraphPad, USA). The differences were evaluated with one-way ANOVA followed by Turkey's post-hoc test. Kaplan-Meier and multivariate Cox regression analysis were used to conduct the prognostic evaluation. $p < 0.05$ indicated a statistically significant difference.

Results

OA Inhibited Cellular Processes of HBV-HCC

In the typical HBV-associated HCC cells, the incubation with OA significantly suppressed HepG2/2.2.15 and MHCC97H cell proliferation ($p < 0.05$, $p < 0.01$, Figure 1A). Meanwhile, the significant inhibitory effect of OA was also exerted on HepG2/2.2.15 and MHCC97H cell migration (Figure 1B) and invasion (Figure 1C, $p < 0.05$, $p < 0.01$, $p < 0.001$), and the inhibitory effect was enhanced with the increasing concentration of OA.

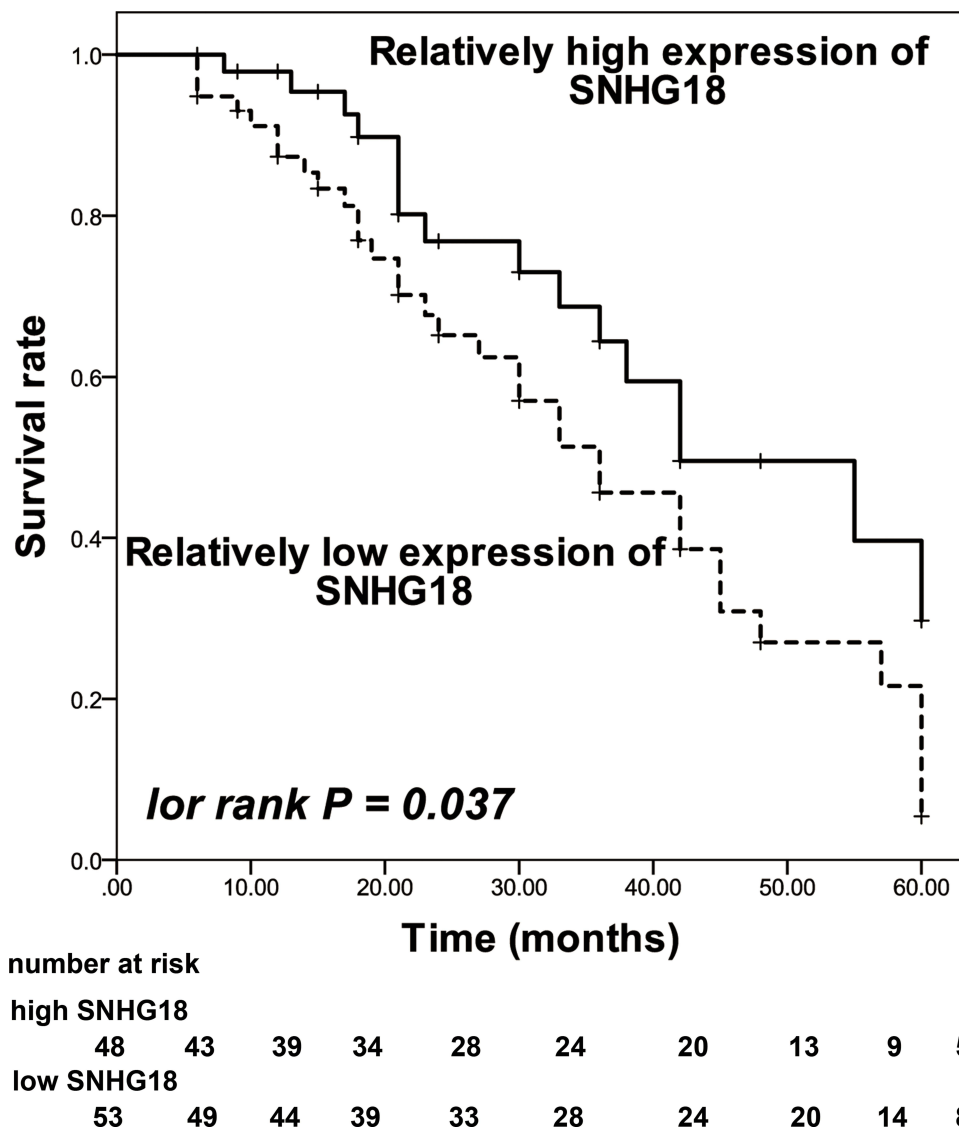


Figure 3 Kaplan-Meier curve of enrolled patients based on the average expression of SNHG18 in collected tumor tissues. Patients with a relatively low expression of SNHG18 showed a poorer survival rate in comparison with patients with relatively high SNHG18 expression. log rank $P = 0.037$.

Table 2 Prognostic Value of Clinicopathological Features Evaluated by Multiple Cox Regression Analysis

	HR Value	95% CI	P value
LncRNA SNHG18	2.701	1.306–5.585	0.007***
Age	1.259	0.705–2.248	0.437
Gender	1.426	0.760–2.675	0.269
AFP	1.662	0.854–3.236	0.135
Tumor size	1.726	0.904–3.294	0.098
TNM stage	2.123	1.148–3.926	0.016*
Differentiation	1.633	0.820–3.251	0.163
Liver cirrhosis	1.767	0.881–3.547	0.109

Notes: * $P < 0.05$ *** $P < 0.001$ indicates the significantly prognostic value.

Abbreviation: AFP, α -fetoprotein.

OA Elevated the Expression of SNHG18 in HBV-HCC

In collected HBV- HCC tissues, the level of SNHG18 was significantly lower than that in normal tissues ($p < 0.001$, Figure 2A). Consistently, the downregulation of SNHG18 was also found in HBV-associated HCC cells relative to normal L02 cells ($p < 0.001$, Figure 2B). In HepG2/2.2.15 cell, the transfection of siRNA-SNHG18 exerted an inhibition on SNHG18 expression, while the treatment with 30 $\mu\text{g}/\text{mL}$ OA dramatically enhanced SNHG18 expression compared with untreated cells, which was reversed by the transfection of si-SNHG18 ($p < 0.01$, $p < 0.001$, Figure 2C).

SNHG18 Serves as a Biomarker of HBV-HCC

The enrolled patients were grouped according to the average expression of SNHG18 in tumor tissues. The high SNHG18 group included 33 males and 20 females and the low SNHG18 group included 58 patients (41 males and 17 females). Among major clinicopathological characteristics of patients, the AFP levels ($p = 0.012$) and the TNM stage ($p = 0.027$) of patients were found to be closely associated with the expression of SNHG18 (Table 1).

Furthermore, the overall survival of patients with relatively low SNHG18 level was much poorer than patients with relatively high SNHG18 level (log rank $p = 0.037$, Figure 3). SNHG18 was also an independent prognostic biomarker of HBV-associated HCC with a HR value of 2.701 (95% CI = 1.306–5.585, $p = 0.007$) as well as TNM with the HR value of 2.123 (95% CI = 1.148–3.926, $p = 0.016$, Table 2).

SNHG18 Mediates the Antitumor Effect of OA in HCC

In HBV- HCC cells, the knockdown of SNHG18 showed a significant promoted effect on critical processes, including cell proliferation (Figure 4A), migration (Figure 4B), and invasion (Figure 4C) compared with untransfected cells ($p < 0.05$, $p < 0.01$). Meanwhile, the downregulation of SNHG18 was found to attenuate the antitumor effect of OA on HepG2/2.2.15 and MHCC97H cells ($p < 0.01$, Figure 4A–C).

Discussion

The antitumor effect of OA has been widely reported in colorectal cancer, non-small cell lung cancer, and breast cancer.^{10,15,16} In HCC, the inhibitory effect of OA on tumor progression has also been uncovered,^{7,8} but the molecular mechanism has not been explored yet. The suppressed effect of OA on tumor progression of HBV-associated HCC was observed, which is consistent with former studies.^{7,8} Meanwhile, the inhibitory effect of OA was found to perform in a concentration-dependent manner, which was enhanced with the increasing concentration.

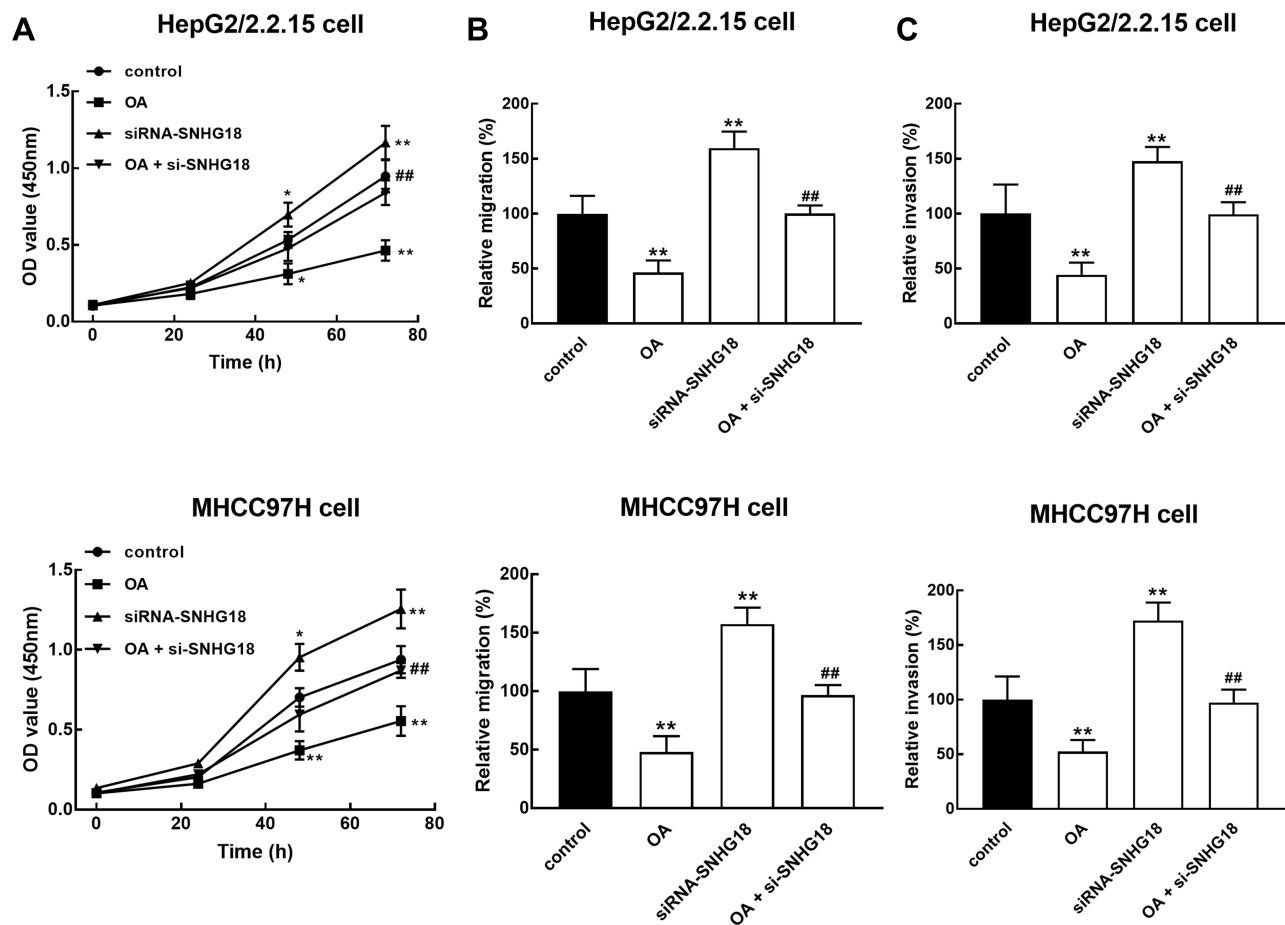


Figure 4 SNHG18 mediated the antitumor effect of OA in HBV-associated HCC. SNHG18 acted as a tumor suppressor of HBV-associated HCC, of which the downregulation exerted significantly promoted effect on cell proliferation (A), migration (B), and invasion (C). While the knockdown of SNHG18 was able to alleviate the inhibition of HBV-associated HCC by OA. * $p < 0.05$, ** $p < 0.01$ relative to the control group. ## $p < 0.01$ relative to the treatment of OA (30 $\mu\text{g/mL}$).

Previously, OA was suggested to enhance miR-421 in pancreatic cancer, which was considered as the molecular mechanism underlying the antitumor effect of OA.¹⁷ The JNK/mTOR signaling pathway was illustrated to participate in the protective effect of OA on autophagy in lung cancer, breast cancer, osteosarcoma, pancreatic cancer, and prostate cancer cells.¹⁸ Here, OA was revealed to regulate the expression of SNHG18 in HBV-associated HCC cells, which was hypothesized to be the potential mechanism of the inhibition of HCC development. Due to the high replication rates and viral mutations, HBV infection has been considered as a serious health problem and the postoperative survival rate of HBV-associated HCC patients is pessimistic.^{19,20} Improving the monitoring of the development of HBV-associated HCC remains a challenge in its clinical therapy. Recently, a number of biomarkers have been leaked out in tumor research. For example, lncRNA PCNAP1 promoted HBV replication via interacting with HBV ccDNA and therefore drive tumor progression and hepatocarcinogenesis.²¹ The involvement of SNHG18 was disclosed in glioma, due to the fact that SNHG18 could enhance the radioresistance of glioma cells and promote cell migration and invasion.^{14,22} Similarly, SNHG18 was not only associated with the severity and progression of osteosarcoma, but it was also able to predict the poor prognosis of patients.¹³ Herein, the significant downregulation of SNHG18 was observed in HBV-associated HCC, which is consistent with the previous assessment of SNHG18 expression in HCC.¹² The reduced expression of SNHG18 showed a close relationship with the high AFP level and advanced TNM stage of patients, which are an indicator of the malignant progression of HCC, indicating the potential involvement of HBV-associated HCC of SNHG18. Additionally, SNHG18 was also found to be significantly related to the overall survival and was appraised as an independent prognostic indicator of HBV-associated HCC together with the TNM stage.

Moreover, SNHG18 was also observed to exert inhibitory effects on major cellular processes of HBV-associated HCC, including cell growth, metastasis, and invasion, suggesting its tumor suppressor role. OA elevated the expression level of SNHG18, while SNHG18 could also alleviate the inhibition of HBV-associated HCC development by OA. Therefore, the antitumor effect of OA in HBV-associated HCC was speculated to be a result of the enhancement of SNHG18, which suppressed tumor progression.

However, the relatively small sample size of this study compared with previous data from the public databases might limit the results. The insignificant association between SNHG18 and some patients' critical clinicopathological features, such as tumor size, differentiation, and liver cirrhosis.^{23–29} These characteristics of patients are also crucial for the disease development and the clinical outcome of patients, the unobvious significance of these features might result from the limited sample size. On the other hand, the lack of in vivo validation is also a prospect of this study in further investigations.

Conclusion

Taken together, SNHG18 participated in the development of HBV-associated HCC and predicted the prognosis of patients. OA showed a dramatic antitumor effect on the cellular processes of HBV-associated HCC and increased the expression of SNHG18. SNHG18 was demonstrated as a tumor suppressor mediating the antitumor effect of OA in HBV-associated HCC.

Funding

The present study was supported by The Team of Jiamusi University Biology (Grant No.jdxktd2019003), The screening of new anti-tumor targets and drug designing (Grant No.jcyxhxt2021-01), Horizontal subject of Jiangsu Vocational College of Medicine, Key Laboratory of Microecology-immune Regulatory Network and Related Diseases Open Project (Grant No.2021-SZD-JC-001).

Disclosure

The authors report no conflicts of interest in this work.

References

1. Chen Y, Tian Z. HBV-induced immune imbalance in the development of HCC. *Front Immunol.* 2019;10:2048. doi:10.3389/fimmu.2019.02048
2. Xie Y. Hepatitis B virus-associated hepatocellular carcinoma. *Adv Exp Med Biol.* 2017;1018:11–21.
3. Luo P, Wu S, Yu Y, et al. Current status and perspective biomarkers in AFP negative HCC: towards screening for and diagnosing hepatocellular carcinoma at an earlier stage. *Pathol Oncol Res.* 2020;26(2):599–603. doi:10.1007/s12253-019-00585-5
4. Wang W, Wei C. Advances in the early diagnosis of hepatocellular carcinoma. *Genes Dis.* 2020;7(3):308–319. doi:10.1016/j.gendis.2020.01.014
5. Pollier J, Goossens A. Oleanolic acid. *Phytochemistry.* 2012;77:10–15. doi:10.1016/j.phytochem.2011.12.022
6. Zhou R, Zhang Z, Zhao L, et al. Inhibition of mTOR signaling by oleanolic acid contributes to its anti-tumor activity in osteosarcoma cells. *J Orthop Res.* 2011;29(6):846–852. doi:10.1002/jor.21311
7. Wang H, Zhong W, Zhao J, et al. Oleanolic acid inhibits epithelial-mesenchymal transition of hepatocellular carcinoma by promoting iNOS dimerization. *Mol Cancer Ther.* 2019;18(1):62–74. doi:10.1158/1535-7163.MCT-18-0448
8. Zhou W, Zeng X, Wu X. Effect of oleanolic acid on apoptosis and autophagy of SMMC-7721 hepatoma cells. *Med Sci Monit.* 2020;26:e921606. doi:10.12659/MSM.921606
9. Chaturvedi VK, Singh A, Dubey SK, Hetta HF, John J, Singh MP. Molecular mechanistic insight of hepatitis B virus mediated hepatocellular carcinoma. *Microb Pathog.* 2019;128:184–194. doi:10.1016/j.micpath.2019.01.004
10. Zhao X, Liu M, Li D. Oleanolic acid suppresses the proliferation of lung carcinoma cells by miR-122/Cyclin G1/MEF2D axis. *Mol Cell Biochem.* 2015;400(1–2):1–7. doi:10.1007/s11010-014-2228-7
11. Han D, Zhang X, Zhang J, et al. Oleanolic acid suppresses vascular smooth muscle cell proliferation by increasing lincRNA-p21 expression. *Oncol Lett.* 2016;12(5):3519–3522. doi:10.3892/ol.2016.5096
12. Liu XF, Thin KZ, Ming XL, et al. Small nucleolar RNA host gene 18 acts as a tumor suppressor and a diagnostic indicator in hepatocellular carcinoma. *Technol Cancer Res Treat.* 2018;17:1533033818794494. doi:10.1177/1533033818794494
13. Huang LJ, Shen Y, Bai J, et al. High expression levels of long noncoding RNA small nucleolar RNA host gene 18 and semaphorin 5A indicate poor prognosis in multiple myeloma. *Acta Haematol.* 2020;143(3):279–288. doi:10.1159/000502404
14. Zheng R, Yao Q, Ren C, et al. Upregulation of long noncoding RNA small nucleolar RNA host gene 18 promotes radioresistance of glioma by repressing semaphorin 5A. *Int J Radiat Oncol Biol Phys.* 2016;96(4):877–887. doi:10.1016/j.ijrobp.2016.07.036
15. Bao Y, Zhang S, Chen Z, et al. Synergistic chemotherapy for breast cancer and breast cancer brain metastases via paclitaxel-loaded oleanolic acid nanoparticles. *Mol Pharm.* 2020;17(4):1343–1351. doi:10.1021/acs.molpharmaceut.0c00044

16. Niu G, Sun L, Pei Y, Wang D. Oleanolic acid inhibits colorectal cancer angiogenesis by blocking the VEGFR2 signaling pathway. *Anticancer Agents Med Chem.* 2018;18(4):583–590. doi:10.2174/1871520617666171020124916
17. Shopit A, Li X, Tang Z, et al. miR-421 up-regulation by the oleanolic acid derivative K73-03 regulates epigenetically SPINK1 transcription in pancreatic cancer cells leading to metabolic changes and enhanced apoptosis. *Pharmacol Res.* 2020;161:105130. doi:10.1016/j.phrs.2020.105130
18. Liu J, Zheng L, Zhong J, Wu N, Liu G, Lin X. Oleanolic acid induces protective autophagy in cancer cells through the JNK and mTOR pathways. *Oncol Rep.* 2014;32(2):567–572. doi:10.3892/or.2014.3239
19. Li YW, Yang FC, Lu HQ, Zhang JS. Hepatocellular carcinoma and hepatitis B surface protein. *World J Gastroenterol.* 2016;22(6):1943–1952. doi:10.3748/wjg.v22.i6.1943
20. Tong S, Revill P. Overview of hepatitis B viral replication and genetic variability. *J Hepatol.* 2016;64(1 Suppl):S4–S16. doi:10.1016/j.jhep.2016.01.027
21. Feng J, Yang G, Liu Y, et al. LncRNA PCNAP1 modulates hepatitis B virus replication and enhances tumor growth of liver cancer. *Theranostics.* 2019;9(18):5227–5245. doi:10.7150/thno.34273
22. Zheng R, Yao Q, Li X, Xu B. Long noncoding ribonucleic acid SNHG18 promotes glioma cell motility via disruption of alpha-enolase nucleocytoplasmic transport. *Front Genet.* 2019;10:1140. doi:10.3389/fgene.2019.01140
23. Fattovich G, Stroffolini T, Zagni I, Donato F. Hepatocellular carcinoma in cirrhosis: incidence and risk factors. *Gastroenterology.* 2004;127(5 Suppl 1):S35–S50. doi:10.1053/j.gastro.2004.09.014
24. Kanda T, Goto T, Hirotsu Y, Moriyama M, Omata M. Molecular mechanisms driving progression of liver cirrhosis towards hepatocellular carcinoma in chronic hepatitis B and C infections: a review. *Int J Mol Sci.* 2019;20(6):1358. doi:10.3390/ijms20061358
25. Daoud A, Teeter L, Ghobrial RM, et al. Transplantation for hepatocellular carcinoma: is there a tumor size limit? *Transplant Proc.* 2018;50(10):3577–3581. doi:10.1016/j.transproceed.2018.04.038
26. Huang WJ, Jeng YM, Lai HS, Sheu FY, Lai PL, Yuan RH. Tumor size is a major determinant of prognosis of resected stage I hepatocellular carcinoma. *Langenbecks Arch Surg.* 2015;400(6):725–734. doi:10.1007/s00423-015-1329-4
27. Kim SJ, Lee KK, Kim DG. Tumor size predicts the biological behavior and influence of operative modalities in hepatocellular carcinoma. *Hepato-gastroenterology.* 2010;57(97):121–126.
28. Liu M, Yan Q, Sun Y, et al. A hepatocyte differentiation model reveals two subtypes of liver cancer with different oncofetal properties and therapeutic targets. *Proc Natl Acad Sci U S A.* 2020;117(11):6103–6113. doi:10.1073/pnas.1912146117
29. Zhang W, Mu D, Feng K. Hierarchical potential differentiation of liver cancer stem cells. *Adv Clin Exp Med.* 2017;26(7):1137–1141. doi:10.17219/acem/66343

Cancer Management and Research

Dovepress

Publish your work in this journal

Cancer Management and Research is an international, peer-reviewed open access journal focusing on cancer research and the optimal use of preventative and integrated treatment interventions to achieve improved outcomes, enhanced survival and quality of life for the cancer patient. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/cancer-management-and-research-journal>