



ORIGINAL ARTICLE

An ionic liquid supported CeO₂ nanoparticles–carbon nanotubes composite-enhanced electrochemical DNA-based sensor for the detection of Pb²⁺

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Abstract An electrochemical sensor incorporating a signal enhancement for the determination of lead (II) ions (Pb²⁺) was designed on the basis of the thrombin-binding aptamer (TBA) as a molecular recognition element and ionic liquid supported cerium oxide (CeO₂) nanoparticles–carbon nanotubes composite modification. The composite comprises nanoparticles CeO₂, multi-wall carbon nanotubes (MWNTs) and hydrophobic room temperature ionic liquid (RTIL) 1-ethyl-3-methylimidazolium tetrafluoroborate (EMIMBF₄). The electrochemical sensors were fabricated by immersing the CeO₂–MWNTs–EMIMBF₄ modified glassy carbon electrode (GCE) into the solution of TBA probe. In the presence of Pb²⁺, the TBA probe could form stable G-quartet structure by the specific binding interactions between Pb²⁺ and TBA. The TBA-bound Pb²⁺ can be electrochemically reduced, which provides a readout signal for quantitative detection of Pb²⁺. The reduction peak current is linearly related to the concentration of Pb²⁺ from 1.0 × 10⁻⁸ M to 1.0 × 10⁻⁵ M with a detection limit of 5 × 10⁻⁹ M. This work demonstrates that the CeO₂–MWNTs–EMIMBF₄ nanocomposite modified GCE provides a promising platform for immobilizing the TBA probe and enhancing the sensitivity of the DNA-based sensors.

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1. Introduction

Detection and quantification of metal ions and organic molecules found in biological systems and in the environment remains an active area of research, as these molecules are either quite beneficial or toxic to human health. Lead is one of the most toxic metallic pollutants, for example, lead can cause renal malfunction and inhibit brain development [1–3]. Although conventional detection methods, including atomic absorption spectrometry [4], inductively coupled plasma (ICP)-mass spectrometry [5], anodic stripping voltammetry [6], and atomic

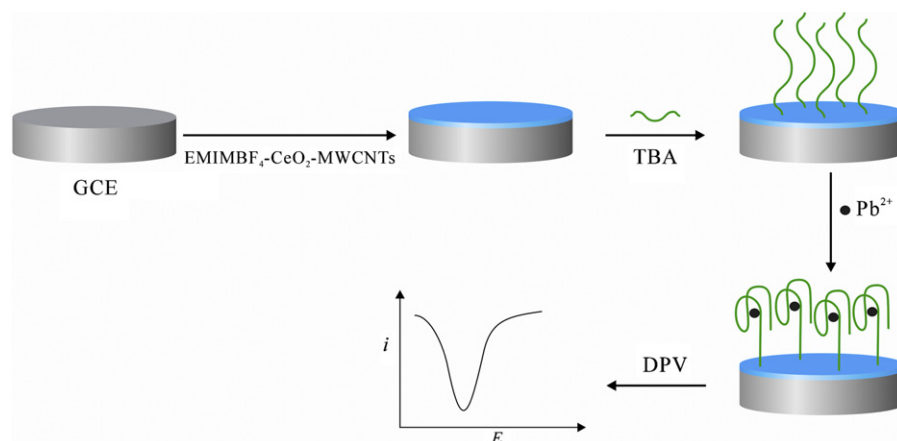


Figure 1 Schematic description of the electrochemical DNA-based sensor for the detection of Pb²⁺ ion.

fluorescence spectrometry [7], provide high sensitivity, they are not suitable for on-site detection because of their sophisticated equipments and sample treatments. In recent years, much optical and electrochemical techniques have been developed for the selective detection of Pb²⁺ based on small molecules [8], DNAzymes [9–13], oligonucleotides [14], polymers [15], and functional nanoparticles [16]. For example, Li and Lu [9] and Liu and Lu [10] developed a fluorescence resonance energy transfer (FRET)-based DNAzyme system for Pb²⁺ sensing. The sensor was made of FRET between fluorophore and quencher labeled on the DNAzyme (17E) and its substrate, respectively. In the presence of Pb²⁺, the 17E catalyzes hydrolytic cleavage of substrate and it turned on the fluorescence for sensing. Xiao et al. [11] developed an electrochemical DNAzyme-based Pb²⁺ biosensor fabricated by thiol-assembling a methylene blue-tagged thiol containing DNAzyme with a detection limit of 0.3 μM. However, many of these systems have limited practical use because of, for example, poor aqueous solubility, cross-sensitivity toward other metal ions, matrix interference, high cost (e.g., enzymes), complicated processing, the use of unstable molecules (e.g., RNA), or poor sensitivity [14]. Recently, Liu et al. [14] reported a technique for the highly selective and sensitive detection of Pb²⁺ using a thrombin-binding aptamer (TBA) probe labeled with the donor carboxyfluorescein and the quencher 4-[(4-(dimethylamino) phenyl] azo] benzoic acid at its 5' and 3' termini. Owing to the high sensitivity, inherent simplicity, low cost, and excellent compatibility to miniaturization technology for electrochemical techniques, the development of electrochemical sensors for Pb²⁺ on the basis of TBA as a molecular recognition might hold great potential for decentralized studies and on-site monitoring.

Room temperature ionic liquid (RTIL), composed of organic cations and various anions, represents a kind of novel nonaqueous but polar solvent [17]. It exhibits many unique advantages such as high chemical and thermal stabilities, negligible vapor pressure, high ionic conductivity, wide electrochemical windows, low toxicity, and ability to dissolve a wide range of organic and inorganic compounds [18]. The attractive properties of RTIL make them promising candidates for electrochemical DNA-based sensor [19–21]. Increasing attention has been paid to the modified electrodes with ionic liquid and nanomaterial composite in hopes of combining their unique properties. In recent years, that inorganic oxide nanoparticles are utilized to be the immobilizing carriers of ssDNA probe is becoming the focus of

research due to unique properties derived from their low dimensionality and possible quantum-confinement effects [22–24]. Among the inorganic oxide nanoparticles, cerium oxide (CeO₂) has been exploited as a promising material for biosensing owing to its unusual properties including large surface area, excellent biocompatibility, nontoxicity, high chemical stability, and strong adsorption ability (high isoelectric point ~9.2) [25,26]. It may be noted that positively charged surface of CeO₂ nanoparticles could be utilized for binding of negatively charged biomolecules. Recently, a novel nanocomposite membrane, comprising nanosized shuttle-shaped CeO₂, single-wall carbon nanotubes (SWNTs) and hydrophobic RTIL, was developed on the glassy carbon electrode for electrochemical sensing of the immobilization and hybridization of DNA [27]. However, to our best knowledge, the application of ionic liquid supported CeO₂ nanoparticles–multi-wall carbon nanotubes (MWNTs) composite for the determination of Pb²⁺ has not been reported on the basis of the TBA as a molecular recognition element.

The aim of this work was to improve the sensitivity of electrochemical DNA-based sensor for the detection of Pb²⁺ utilizing TBA as a molecular recognition element and ionic liquid supported CeO₂ nanoparticles–MWNTs composite of the electrochemical DNA-based sensor for the detection of Pb²⁺ ion is shown in Fig. 1. In this paper, an electrochemical sensor for the determination of Pb²⁺ was designed and the electrochemical characteristics of the electrochemical sensor fabricated were investigated. To our best knowledge, it is new example of electrochemical sensor for the determination of Pb²⁺ based on signal enhancement.

2. Experimental

2.1. Reagents and apparatus

MWNTs were obtained from Shenzhen Nanotech Port Co. Ltd. (Shenzhen, China). CeO₂ nanoparticle was obtained from Beijing Nachen Science and Technology Ltd. Company (Beijing, China). 1-Ethyl-3-methylimidazolium tetrafluoroborate (EMIMBF₄) was obtained from Lanzhou Institute of Chemical Physics of the Chinese Academy of Sciences. Pb²⁺ stock solution was prepared by dissolving Pb(NO₃)₂ with Millipore Milli-Q water. Thrombin-binding aptamers were synthesized by Shengggong Bioengineering

Ltd. Company (Shanghai, China). A 21-mer target aptamer probe adopted from literature [28] was used to bind with thrombin: 5'-CAC TGT GGT TGG TGT GGT TGG-(CH₂)₆-NH₂-3'. MgCl₂, CaCl₂, Hg (NO₃)₂, CuCl₂, ZnCl₂, and all other reagents were of analytical grade. A 0.10 M phosphate buffer solution (PBS, pH 7.40, 0.10 M NaCl+10 mM NaH₂PO₄/Na₂HPO₄) was used as hybridization buffer, washing solution and electrolyte. All reagents were of analytical grade. Millipore Milli-Q water (18 MΩ cm) was used throughout.

The experimental set-up for electrochemical measurement was the same as the previous paper [29].

2.2. Preparation of the CeO₂-MWNTs-EMIMBF₄/GCE

The purified MWCNTs were prepared as reported in the literature [29–31]. 10 mg of purified MWNTs were dispersed in 10 mL of dimethylformamide (DMF) with the aid of ultrasonication for 2 h to give a 1.0 mg/mL homogeneous black suspension. Then 0.2 mg of CeO₂ nanoparticles were added to the MWNTs suspension and the resulted suspension was ultrasonicated for 2 h. After that, EMIMBF₄ (final concentration (v/v) 50%) was dispersed in the CeO₂-MWNTs composite with the aid of ultrasonication [32]. Finally, a uniform CeO₂-MWNTs-EMIMBF₄ nanocomposite suspension was obtained. Before modification, the GCE was pretreated as previously described [33]. 5 μL of the CeO₂-MWNTs-EMIMBF₄ suspension was dropped on the GCE and let it dry at room temperature for 2 h, thus a uniform membrane coated electrode (CeO₂-MWNTs-EMIMBF₄/GCE) was obtained. The CeO₂/GCE, CeO₂-EMIMBF₄/GCE and CeO₂-MWNTs/GCE were fabricated through similar procedure.

2.3. DNA probe immobilization

Immobilization of TBAs was performed by immersing the CeO₂-MWNTs-EMIMBF₄/GCE into 2.0 mL PBS (pH 7.0) solution containing 5.0 × 10⁻⁶ M TBAs probe for 4 h at room temperature, followed by washing the electrode with 0.10 M PBS and then rinsing it with ultrapure water to remove the unimmobilized TBA probe, and this TBA probe-captured electrode was denoted as TBA/CeO₂-MWNTs-EMIMBF₄/GCE.

2.4. Electrochemical measurement

A sensor fabricated was immersed into different concentrations of Pb²⁺ solution for 60 min at 37 °C, followed by thoroughly washing with 0.10 M PBS to remove unbound Pb²⁺. After that, the electrode was transferred into electrochemical cell. The electrochemical reduction of Pb²⁺ to Pb⁰ was recorded by a cathodic scan of DPV. The DPV measurement was performed in the potential range from -0.5 to 0 V in 0.10 M PBS (pH 7.4, containing 0.1 M NaNO₃) with pulse amplitude of 50 ms and pulse width of 50 mV.

3. Results and discussion

3.1. Electrochemical characteristics of the CeO₂-MWNTs-EMIMBF₄ nanocomposite membrane

CV and EIS were used for monitoring the process of the fabrication of electrochemical DNA-based sensor for the

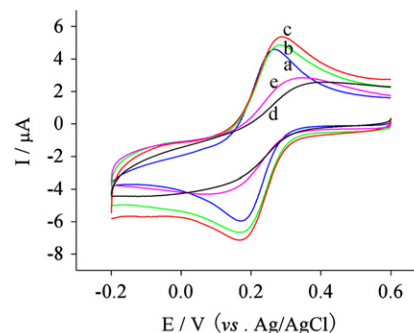


Figure 2 CV of obtained in 1 mL PBS (0.1 M, pH 7.40) containing 1 mM Fe(CN)₆³⁻/Fe(CN)₆⁴⁻ and 0.1 M KCl at a scan rate of 50 mV/s at (a) bare GCE, (b) CeO₂-MWNTs/GCE, (c) CeO₂-MWNTs-EMIMBF₄/GCE, (d) TBA/CeO₂-MWNTs-EMIMBF₄/GCE, and (e) TBA/CeO₂-MWNTs-EMIMBF₄/GCE incubate in Pb²⁺ solution for 60 min.

detection of Pb²⁺ in each step. As can be seen from Fig. 2, the CeO₂-MWNTs/GCE (Fig. 2, curve b) and the CeO₂-MWNTs-EMIMBF₄/GCE (Fig. 2, curve c) had larger CV current than bare GCE (Fig. 2, curve a), which ascribed to the fact that modification of the electrode with CeO₂-MWNTs-EMIMBF₄ could significantly enhance the effective electrode surface area and ionic conductivity. A current decrease (Fig. 2, curve d) appeared after exposing to TBA probe. It is well-known that an immobilized TBA probe as an electron-transfer blocking layer can hinder the diffusion of ferricyanide toward the electrode surface [34]. After incubation with 1.0 × 10⁻⁷ M Pb²⁺, the formation of the Pb²⁺/TBA/CeO₂-MWNTs-EMIMBF₄/GCE contributed to a significant increase in redox current (Fig. 2, curve e). This is probably attributed to the fact that the positive charge of Pb²⁺ serves to reduce the repulsion of [Fe(CN)₆]^{3-/4-} to the surface, promoting the interfacial electron transfer between the redox anions in solution and the electrode [35]. The results showed that the TBA probe is immobilized on the CeO₂-MWNTs-EMIMBF₄/GCE and the interaction between the electrochemical DNA-based sensor and Pb²⁺ arises.

Fig. 3 shows Nyquist plots of impedance spectra obtained at different electrodes. The change in semicircle diameter is a result in the change in the interfacial resistance R_{et} to electron transfer from their modified electrode to ferricyanide in solution. The electrochemical response was a nearly straight line (curve a), which is characteristic for a limiting step of the electrochemical process at a bare GCE. After CeO₂-MWNTs-EMIMBF₄ nanocomposite was modified (curve b), the electrochemical response was a nearly straight line. After TBA probe was immobilized onto the surface of CeO₂-MWNTs-EMIMBF₄/GCE, the R_{et} markedly increased to 96,370 Ω (curve c). This is attributed to the fact that the redox couple of K₃[Fe(CN)₆]/K₄[Fe(CN)₆] is suffered electrostatic repulsive forces from the immobilized TBA probe. In the presence of Pb²⁺, the R_{et} significantly decreased to 44,040 Ω (curve d). This is probably attributed to the fact that the conformation variation enhances the steric and coulombic force between adjacent DNA sequences [36], resulting in a decrease of R_{et} . The results extracted from EIS measurements (Fig. 3) are in good agreement with the results obtained from CV (Fig. 2).

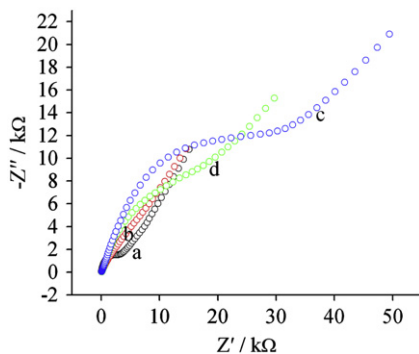


Figure 3 Nyquist plots of impedance spectra obtained in 1 mL PBS (0.1 M, pH 7.40) containing 1 mM Fe(CN)₆³⁻/Fe(CN)₆⁴⁻ and 0.1 M KCl at (a) bare GCE, (b) CeO₂-MWNTs-EMIMBF₄/GCE, (c) TBA/CeO₂-MWNTs-EMIMBF₄/GCE, and (d) TBA/CeO₂-MWNTs-EMIMBF₄/GCE incubated in Pb²⁺ solution for 60 min. The biased potential was 0.172 V. The frequency was 0.1–100 kHz and the amplitude was 5.0 mV.

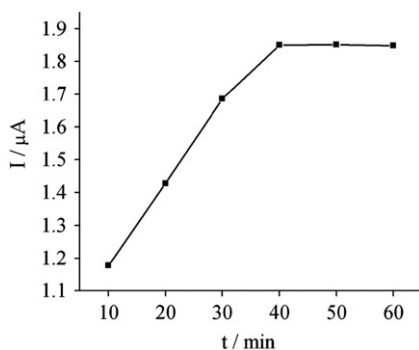


Figure 4 Dependence of peak current of the DNA-based sensor on incubation time in PBS (0.1 M, pH 7.40) solution containing 1.0 × 10⁻⁷ M Pb²⁺ by the CeO₂-MWNTs-EMIMBF₄ nanocomposite amplified strategy. DPV parameters: pulse amplitude, 50 ms; pulse width, 50 mV.

3.2. Optimization of incubation time

It was found that a different incubation time of Pb²⁺ caused a visible difference in the increase of peak current. Therefore, the dependence of Pb²⁺ incubation time on the increase of the peak current was studied to determine the optimum incubation time of Pb²⁺. As shown in Fig. 4, the reduction peak current increased immediately when 1.0 × 10⁻⁷ M Pb²⁺ was introduced and then tended to reach the maximum after more than 40 min. To ensure that sufficient amount of Pb²⁺ can be gathered on the electrode, 60 min was chosen as the Pb²⁺ incubation time. This incubation time is longer than that required in homogenous fluorescent methods (15 min) [14], indicating that the reaction of the surface-confined TBA probe with Pb²⁺ is much slower than that of the TBA probe with Pb²⁺ in solution. The incubation time of this electrochemical sensor for a real sample should be reduced.

3.3. Performance of the electrochemical sensor

The quantitative behavior of the electrochemical sensor fabricated was assessed under optimized analytical condition.

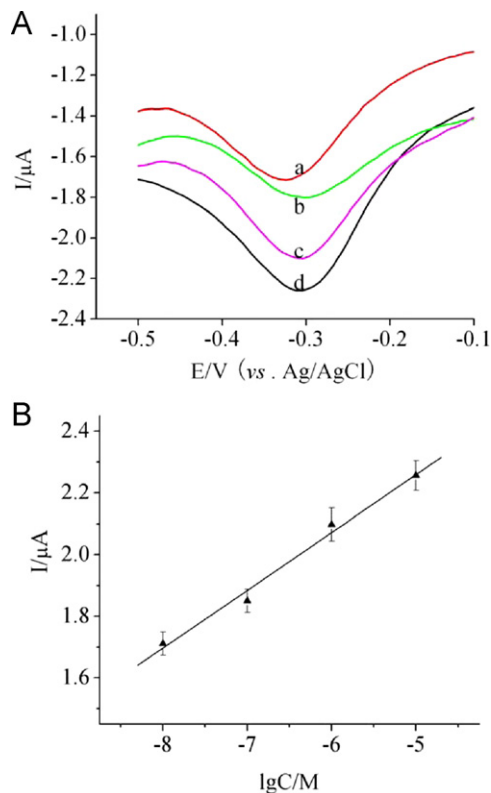


Figure 5 (A) DPV of the electrochemical sensor for the detection of different concentrations of Pb²⁺ by the CeO₂-MWNTs-EMIMBF₄ nanocomposite amplified strategy: (a) 1.0 × 10⁻⁸ M, (b) 1.0 × 10⁻⁷ M, (c) 1.0 × 10⁻⁶ M, and (d) 1.0 × 10⁻⁵ M. (B) The linear relationship between the peak current and the concentration of Pb²⁺. The DPV conditions are the same as Fig. 4.

Fig. 5 shows the DPV profiles of the sensor at different concentrations of Pb²⁺. The reduction peaks of Pb²⁺ appeared at ~0.3 V by DPV detection. From Fig. 5A, it can be seen that the peak current increases with an increase of the concentration of Pb²⁺. The peak current had a linear relationship with the concentration of Pb²⁺ in the range from 1.0 × 10⁻⁸ to 1.0 × 10⁻⁵ M (Fig. 5B). The linear regression equation was $I = 3.202 + 0.1882 \lg C$, (unit of C is M) and the correlation coefficient was 0.9943. The detection limit was 5.0 × 10⁻⁹ M Pb²⁺ ($S/N=3$). The detection limit is considerably lower than 72 nM, the EPA-defined maximal contamination level for Pb²⁺ in drinking water. The relative standard derivation for 5.0 × 10⁻⁷ M Pb²⁺ was 3.6% ($n=7$). The storage stability of the electrochemical DNA-based sensors fabricated was also checked. The results showed that the current response of the electrochemical DNA-based sensor to 1.0 × 10⁻⁷ M Pb²⁺ decreased about 4.5% after the sensor was stored in air at room temperature for 7 days. This suggests that the sensor has good stability. A satisfactory detection sensitivity, reproducibility and stability of the proposed sensor are, therefore, verified.

The electrochemical DNA-based Pb²⁺ sensor was also highly selective. Fig. 6 depicts the electrochemical response of Pb²⁺ in a mixture of five different interference metal ions. As shown in Fig. 6, the sensor hardly exhibited substantial responses to a mixture of another five different metal ions (1 mM of Ca²⁺, Mg²⁺, Hg²⁺, Cu²⁺, and Zn²⁺) (curve a). The

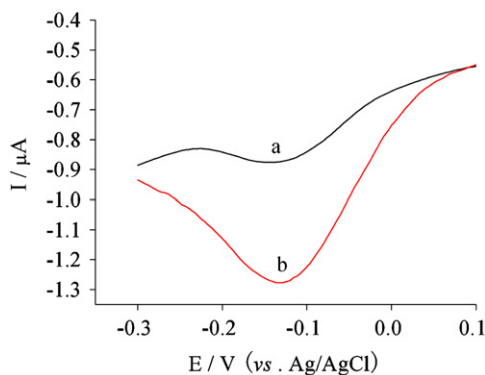


Figure 6 DPV corresponding to detection of (a) 0 and (b) 1.0×10^{-7} M of Pb^{2+} in a mixture of metal ions containing Ca^{2+} , Mg^{2+} , Hg^{2+} , Cu^{2+} , and Zn^{2+} (1 mM, each). The DPV conditions are the same as Fig. 4.

Table 1 The concentration of Pb^{2+} in water samples detected using the proposed electrochemical DNA-based sensor.

Samples	Added Pb^{2+} concentration (M)	Concentration found (M)	Recovery (%)	RSD (%), $n=3$
1	1.0×10^{-8}	9.52×10^{-7}	95.2	4.2
2	1.0×10^{-7}	9.68×10^{-6}	96.8	3.8
3	1.0×10^{-6}	9.76×10^{-5}	97.6	3.2

cross sensitivity of the sensor in a mixture of five different metal ions containing 1.0×10^{-7} M Pb^{2+} was also examined (curve b). The signal obtained from the mixture was slight lower than that obtained from pure Pb^{2+} solution. These results indicate that the electrochemical DNA-based sensor for the detection of Pb^{2+} exhibits high specificity. The result was in agreement with that the unique Pb^{2+} conformation formed by the T-base of DNA sequence, which is quite specific to Pb^{2+} [14,37].

The practical application of the designed sensor was evaluated by determination of the recovery of spiked Pb^{2+} in tap water samples. The analytical results are shown in Table 1. The analytical results show the acceptable relative standard deviation and quantitative recoveries, implying that the proposed sensor was applicable for practical Pb^{2+} detection.

4. Conclusions

An electrochemical DNA-based sensor for the detection of Pb^{2+} was developed based on the TBA as a molecular recognition element and ionic liquid supported CeO_2 nanoparticles-carbon nanotubes composite modification. The developed sensor exhibited a high sensitivity and selectivity. It demonstrated that the sensitivity of electrochemical DNA-based sensor could be greatly improved by using the ionic liquid supported nanoparticles-carbon nanotubes composite modified the electrode. Also because of the high sensitivity, low cost, miniaturization, and simple operation of electrochemical methods, we expect that the CeO_2 -MWNTs-

EMIMBF₄ nanocomposite amplified electrochemical DNA-based sensor could be effective in detecting Pb^{2+} in the actual environment, and also provide a reference value for the detection of other heavy metals ions.

Acknowledgments

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References

- [1] R. Levin, M.J. Brown, M.E. Kashtock, et al., Lead exposures in U.S. children, 2008: implications for prevention, *Environ. Health Perspect.* 116 (2008) 1285–1293.
- [2] H.A. Schroeder, I.H. Tipton, The human body burden of lead, *Arch. Environ. Health* 17 (1968) 965–978.
- [3] S. Araki, H. Sato, K. Yokoyama, et al., Subclinical neurophysiological effects of lead: a review on peripheral, central, and autonomic nervous system effects in lead workers, *Am. J. Ind. Med.* 37 (2000) 193–204.
- [4] L. Yang, S.S. Saavedra, Chemical sensing using sol-gel derived planar waveguides and indicator phases, *Anal. Chem.* 67 (1995) 1307–1314.
- [5] E. Webb, D. Amarasiriwardena, Inductively coupled plasma-mass (ICP-MS) and atomic emission spectrometry (ICP-AES): versatile analytical techniques to identify the archived elemental information in human teeth, *Microchem. J.* 81 (2005) 201–208.
- [6] A.T. Maghasi, S.D. Conklin, T. Shtoyko, et al., Spectroelectrochemical sensing based on attenuated total internal reflectance stripping voltammetry. 2. Determination of mercury and lead, *Anal. Chem.* 76 (2004) 1458–1465.
- [7] E. Wagner, W. Smith, J.D. Winegardner, Ultratrace determination of lead in whole blood using electrothermal atomization laser-excited atomic fluorescence spectrometry, *Anal. Chem.* 68 (1996) 3199–3203.
- [8] S. Deo, H.A. Godwin, A selective, ratiometric fluorescent sensor for Pb^{2+} , *J. Am. Chem. Soc.* 122 (2000) 174–175.
- [9] J. Li, Y. Lu, A highly sensitive and selective catalytic DNA biosensor for lead ions, *J. Am. Chem. Soc.* 122 (2000) 10466–10467.
- [10] J. Liu, Y. Lu, A colorimetric lead biosensor using DNAzyme-directed assembly of gold nanoparticles, *J. Am. Chem. Soc.* 125 (2003) 6642–6643.
- [11] Y. Xiao, A.A. Rowe, K.W. Plaxco, Electrochemical detection of parts-per-billion lead via an electrode-bound DNAzyme assembly, *J. Am. Chem. Soc.* 129 (2007) 262–263.
- [12] Y. Xiang, A. Tong, Y. Lu, Abasic site-containing DNAzyme and aptamer for label-free fluorescent detection of Pb^{2+} and adenosine with high sensitivity, selectivity, and tunable dynamic range, *J. Am. Chem. Soc.* 131 (2009) 15352–15357.
- [13] X. Yang, J. Xu, X. Tang, et al., A novel electrochemical DNAzyme sensor for the amplified detection of Pb^{2+} ions, *Chem. Commun.* 46 (2010) 3107–3109.
- [14] C.W. Liu, C.C. Huang, H.T. Chang, Highly selective DNA-based sensor for lead (II) and mercury (II) ions, *Anal. Chem.* 81 (2009) 2383–2387.
- [15] C.D. Geary, I. Zudans, A.V. Goponenko, et al., Electrochemical investigation of Pb^{2+} binding and transport through a polymerized

- crystalline colloidal array hydrogel containing benzo-18-crown-6, *Anal. Chem.* 77 (2005) 185–192.
- [16] J.H. Kim, S.H. Han, B.H. Chung, Improving Pb²⁺ detection using DNazyme-based fluorescence sensors by pairing fluorescence donors with gold nanoparticles, *Biosensors Bioelectron.* 26 (2011) 2125–2129.
- [17] T. Welton, Room-temperature ionic liquids. Solvents for synthesis and catalysis, *Chem. Rev.* 99 (1999) 2071–2084.
- [18] F. Zhao, X.E. Wu, M.K. Wang, et al., Electrochemical and bioelectrochemistry properties of room-temperature ionic liquids and carbon composite materials, *Anal. Chem.* 76 (2004) 4960–4967.
- [19] D. Wei, A. Ivaska, Applications of ionic liquids in electrochemical sensors, *Anal. Chim. Acta* 607 (2008) 125–126.
- [20] M.J.A. Shiddiky, A.J. Torriero, Application of ionic liquids in electrochemical sensing systems, *Biosensors Bioelectron.* 26 (2011) 1775–1787.
- [21] H. Liu, Y. Liu, J. Li, Ionic liquids in surface electrochemistry, *Phys. Chem. Chem. Phys.* 12 (2010) 1685–1697.
- [22] Y.H. Yang, Z.J. Wang, M.H. Yang, et al., Electrical detection of deoxyribonucleic acid hybridization based on carbon-nanotubes/nano zirconium dioxide/chitosan-modified electrodes, *Anal. Chim. Acta* 584 (2007) 268–274.
- [23] Y. Ma, K. Jiao, T. Yang, et al., Sensitive PAT gene sequence detection by nano-SiO₂/p-aminothiophenol self-assembled films DNA electrochemical biosensor based on impedance measurement, *Sensors Actuators B* 131 (2008) 565–571.
- [24] K.J. Feng, Y.H. Yang, Z.J. Wang, et al., A nano-porous CeO₂/chitosan composite film as the immobilization matrix for colorectal cancer DNA sequence-selective electrochemical biosensor, *Talanta* 70 (2006) 561–565.
- [25] Y. Wei, M.G. Li, S.F. Jiao, et al., On the complex formation of CdCl₂ with 1-furoylthioureas: preconcentration and voltammetric behavior of Cd(II) at carbon paste electrodes modified with 3-mono-substituted and 3,3-disubstituted derivatives, *Electrochim. Acta* 122 (2006) 766–772.
- [26] A.A. Ansari, A. Kaushik, P.R. Solanki, et al., Chitosan-iron oxide nanobiocomposite based immunosensor for ochratoxin-A, *Electrochem. Commun.* 10 (2008) 1364–1368.
- [27] W. Zhang, T. Yang, X. Zhuang, et al., An ionic liquid supported CeO₂ nanoshuttles–carbon nanotubes composite as a platform for impedance DNA hybridization sensing, *Biosensors Bioelectron.* 24 (2009) 2417–2422.
- [28] Z.S. Wu, M.M. Guo, S.B. Zhang, et al., Reusable electrochemical sensing platform for highly sensitive detection of small molecules based on structure-switching signaling aptamers, *Anal. Chem.* 79 (2007) 2933–2939.
- [29] X. Liu, Y. Li, J. Zheng, et al., Carbon nanotube-enhanced electrochemical aptasensor for the detection of thrombin, *Talanta* 81 (2010) 1619–1624.
- [30] H. Cai, X. Cao, Y. Jiang, et al., Carbon nanotube-enhanced electrochemical DNA biosensor for DNA hybridization detection, *Anal. Bioanal. Chem.* 375 (2003) 287–293.
- [31] Q.L. Sheng, J.B. Zheng, Enzymatically induced formation of neodymium hexacyanoferrate nanoparticles on the glucose oxidase/chitosan modified glass carbon electrode for the detection of glucose, *Biosensors Bioelectron.* 24 (2008) 429–434.
- [32] F. Xiao, F.Q. Zhao, J.W. Li, et al., Sensitive voltammetric determination of chloramphenicol by using single-wall carbon nanotube–gold nanoparticle–ionic liquid composite film modified glassy carbon electrodes, *Anal. Chim. Acta* 596 (2007) 79–85.
- [33] S. Niu, M. Zhao, L. Hu, et al., Carbon nanotube-enhanced DNA biosensor for DNA hybridization detection using rutin-Mn as electrochemical indicator, *Sensors Actuators B: Chem.* 135 (2008) 200–205.
- [34] M. Cho, S. Lee, S.Y. Han, et al., Electrochemical detection of mismatched DNA using a MutS probe, *Nucleic Acids Res.* 34 (2006) e75/1–e75/10.
- [35] A.K.H. Cheng, B. Ge, H.Z. Yu, Aptamer-based biosensors for label-free voltammetric detection of lysozyme, *Anal. Chem.* 79 (2007) 5158–5164.
- [36] C.W. Liu, C.C. Huang, H.T. Chang, Control over surface DNA density on gold nanoparticles allows selective and sensitive detection of mercury(II), *Langmuir* 24 (2008) 8346–8350.
- [37] I. Smirnov, R.H. Shafer, Lead is unusually effective in sequence-specific folding of DNA, *J. Mol. Bio.* 296 (2000) 1–5.