




Review

Immunomodulation by Inflammation during Liver and Gastrointestinal Tumorigenesis and Aging

Nao Nagai ^{1,†}, Yotaro Kudo ^{2,†} , Daisuke Aki ¹, Hayato Nakagawa ² and Koji Taniguchi ^{1,3,*}

¹ Department of Microbiology and Immunology, Keio University School of Medicine, 35 Shinanomachi, Shinjuku-ku, Tokyo 160-8582, Japan; nnagai@keio.jp (N.N.); adaisuke61@outlook.jp (D.A.)

² Department of Gastroenterology, Graduate School of Medicine, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8655, Japan; yotaro-ky@umin.ac.jp (Y.K.); hayaton0120@gmail.com (H.N.)

³ Department of Pathology, Faculty of Medicine and Graduate School of Medicine, Hokkaido University, North 15, West 7, Kita-ku, Sapporo, Hokkaido 060-8638, Japan

* Correspondence: kotaniguchi@keio.jp; Tel.: +81-3-5363-3483

† These authors are equally contributed.

Abstract: Chronic inflammation is thought to promote tumorigenesis and metastasis by several mechanisms, such as affecting tumor cells directly, establishing a tumor-supporting microenvironment, enhancing tumor angiogenesis, and suppressing antitumor immunity. In this review, we discuss the recent advances in our understanding of how inflammation induces the immunosuppressive tumor microenvironment, such as increasing the level of pro-inflammatory cytokines, chemokines, and immunosuppressive molecules, inducing immune checkpoint molecules and cytotoxic T-cell exhaustion, and accumulating regulatory T (Treg) cells and myeloid-derived suppressor cells (MDSCs). The suppression of antitumor immunity by inflammation is especially examined in the liver and colorectal cancer. In addition, chronic inflammation is induced during aging and causes age-related diseases, including cancer, by affecting immunity. Therefore, we also discuss the age-related diseases regulated by inflammation, especially in the liver and colon.

Keywords: inflammation; cancer; immunosuppression; tumor microenvironment; aging



Citation: Nagai, N.; Kudo, Y.; Aki, D.; Nakagawa, H.; Taniguchi, K. Immunomodulation by Inflammation during Liver and Gastrointestinal Tumorigenesis and Aging. *Int. J. Mol. Sci.* **2021**, *22*, 2238. <https://doi.org/10.3390/ijms22052238>

Academic Editors: Javier Conde Aranda and Morena Scotece

Received: 1 February 2021
Accepted: 19 February 2021
Published: 24 February 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Inflammation is divided into two types by duration, acute inflammation, and chronic inflammation. Acute inflammation is considered beneficial because of its roles in eliminating pathogens and necrotic cells and inducing tissue regeneration and repair [1]. In acute inflammation, immunosuppression occurs after tissue repair or the elimination of pathogens and necrotic cells to resolve inflammation. However, in chronic inflammation, immunosuppression permanently persists due to the lack of the normal negative feedback mechanism.

Chronic inflammation plays a vital role in tumorigenesis and metastasis. Growth factors and pro-inflammatory cytokines induced by chronic inflammation directly enhance cancer cell proliferation, suppress cell death, and induce angiogenesis. Meanwhile, chemokines produced in the tumor microenvironment recruit several types of immunosuppressive cells, including regulatory T (Treg) cells and myeloid-derived suppressor cells (MDSCs), and create an immunosuppressive environment, resulting in tumor initiation and progression. Prostaglandins, essential in inducing inflammatory responses, are also critical mediators in establishing the immunosuppressive microenvironment. Chronic inflammation is also closely related to aging and promotes age-related diseases by affecting immunity.

In this review, we will discuss the recent advances in the research on how inflammation affects antitumor immunity and causes age-related diseases, especially in the liver and colon.

2. Inflammation Suppresses Antitumor Immunity

Tumor-promoting inflammation is one of the recognized hallmarks of cancer [2]. Cancer-related inflammation is categorized into four types, which include 1. Chronic inflammation caused by infection and autoimmunity, 2. inflammation caused by environmental and dietary exposure, 3. inflammation caused by therapy-induced exposure, and 4. tumor-associated inflammation [3]. Inflammation, especially chronic inflammation, promotes tumorigenesis and metastasis through several mechanisms, including the suppression of antitumor immunity. Various molecules, cells, and processes, including immune checkpoint molecules, Treg cells and MDSCs, and angiogenesis, are involved in suppressing antitumor immunity mediated by inflammation.

The induction of immune checkpoint molecules in CD8⁺ T cells is crucial in establishing the immunosuppressive microenvironment [4]. The activation of immune checkpoint pathways in CD8⁺ T cells induces T cell exhaustion or dysfunction, resulting in decreased cell proliferation, the expression of effector molecules, such as interferon (IFN)- γ , tumor necrosis factor (TNF), and interleukin-2 (IL-2), immunosuppression, and immunotolerance. Therefore, immune checkpoint inhibitors can be used to treat cancer. The successful use of immune checkpoint inhibitors and chimeric antigen receptor-engineered T (CAR-T) therapy has dramatically changed the significance of immunotherapy in cancer treatment. However, the efficacy of immune checkpoint inhibitors remains unsatisfactory in some cancer types, including colorectal cancer (CRC) [4]. While various combination therapies with potentially improved efficacy are in clinical testing, it remains vital to understand why immune checkpoint inhibitors can only reactivate antitumor immunity in the immunosuppressive microenvironment in a portion of patients.

The limited efficacy of the immune checkpoint inhibitors is due to T cell exhaustion in the tumor microenvironment; therefore, it is crucial to delineate the mechanism of T cell exhaustion. The surface markers for exhausted or dysfunctional T cells are primarily inhibitory receptors, including PD-1, CTLA-4, TIM-3, LAG-3, and TIGIT [5–8]. The PD-1/PD-L1 axis likely plays the most crucial role in T cell exhaustion in mouse models and clinical outcomes. In addition, compared to the functional effector or memory T cells [9], the transcriptional factors encoded by exhausted or dysfunctional T cells are considerably different, including NR4A (NR4A1, NR4A2, NR4A3), TOX, Eomes, T-bet, Prdm1 (Blimp-1), NF-AT, and BATF [10–15]; all these proteins regulate the level of PD-1 [16]. NR4A1 increases the level of PD-1 and TIM-3, and NR4A1-deficient T cells produce more IFN- γ and TNF and mediate tumor regression more significantly. These data suggest that NR4A is an attractive novel target of cancer immunotherapy; for example, camptothecin, a topoisomerase inhibitor, is identified as an NR4A inhibitor and to enhance antitumor immunity [17].

Tregs and MDSCs are also crucial immune cells that induce the immunosuppressive microenvironment [16]. Tregs, which express an essential transcriptional factor, forkhead box P3 (FOXP3), suppress antitumor immunity by producing anti-inflammatory cytokines, such as transforming growth factor (TGF)- β and IL-10 [16]. Furthermore, Tregs repress cytotoxic lymphocytes that produce granzyme and perforin, suppress cell proliferation via IL-2 signaling, and interact with dendritic cells that regulate effector cell function and maturation [18]. CCL17 and CCL22, induced in an inflammatory condition, play an essential role in recruiting Tregs to the tumor microenvironment [19,20]. Once exposed to inflammatory conditions, Treg cells acquire a robust, long-lasting immunosuppressive function [21].

There may be two populations of functionally distinct Treg cells with differential levels of FOXP3. While the Treg population is increased in patients with esophageal cancer, gastric cancer, or CRC and the number of infiltrating Treg cells in a tumor is associated with poor prognosis in most cancer types, the infiltration of FOXP3-positive T cells in CRC is associated with a more favorable prognosis [22]. Therefore, there may be two functionally distinct populations of FOXP3-positive CD4⁺ T cells in CRC, the suppression-competent FOXP3^{high} Treg cells, and the non-suppressive FOXP3^{low} T cells that prevent tumor formation via

antitumor immunity [23]. While Tregs are immunosuppressive in inflammatory bowel disease (IBD) and gastritis, they are converted by chronic inflammation to IL-17-producing Tregs, which do not produce IL-10 and are found in colon adenomas [24].

While MDSCs can induce Tregs, they can also suppress effector T cells [25]. MDSCs are a heterogenic population of immature myeloid cells, including progenitors of macrophages, DCs, and granulocytes. MDSCs induce immunosuppression by producing TGF- β , IL-10, reactive oxygen species (ROS), nitric oxide (NO), arginase1, and indoleamine oxidase. MDSCs also promote tumorigenesis by protecting tumor cells from senescence, inhibiting CD8⁺ T cell and natural killer (NK) cell cytotoxicity, and promoting cancer stem cell expansion [26]. The expression of inflammatory mediators, such as IL-1 β , IL-6, prostaglandin E₂ (PGE₂), granulocyte macrophage colony-stimulating factor (GM-CSF), and vascular endothelial growth factor (VEGF), is increased in the tumor microenvironment, thus recruiting MDSCs in the plasma to the site of inflammation and inducing their activation. In addition, the CXCL8-CXCR2 axis plays a vital role in the recruitment of MDSCs [27].

3. Tumor Angiogenesis Develops a Tolerogenic Tumor Microenvironment

Abnormalities in blood vessels also help shape an immunosuppressive microenvironment that induces cancer cells to become aggressive and enable them to resist cytotoxic therapy. Angiogenesis is the development of new blood vessels. Sustained angiogenesis is one of the hallmarks of malignancies since the blood supply is essential for tumor development and growth [28]. Unlike normal blood vessels, the vessels in sustained angiogenesis are structurally and functionally abnormal, immature with abnormally high leakiness [29], and lack pericytes and basement membrane [30]. Additionally, in the tumor microenvironment, they increase the permeability coupled with the elevation in interstitial fluid pressure [30,31]. Angiogenic factors, including VEGF and angiopoietin family proteins, are produced by cancer cells and stromal cells, such as the endothelial and immune cells. VEGF also causes immune suppression by inhibiting the maturation of DCs from progenitors [32], attracting MDSCs in the bone marrow to the periphery [33,34], inhibiting cytotoxic CD8⁺ T cells by inducing the expression of TOX, and upregulating production of PD-1 and other inhibitory checkpoints molecules [35,36]. In addition, VEGF directly induces Treg proliferation in murine models [37]. Collectively, tumor angiogenesis and the tumor's abnormal vascular structure play critical roles in developing a tolerogenic tumor microenvironment.

Recently, epigenetic dysregulation, which plays a role in tumor angiogenesis and immune evasion, has been proposed as a novel hallmark of cancer [38]. MicroRNAs are also critically important players in remodeling the tumor microenvironment by affecting endothelial cells and several immune cells [39]. However, further studies are necessary to delineate the role of epigenetic dysregulation and microRNAs in the remodeling of the tumor microenvironment.

Next, we will discuss the immunosuppression in the liver and CRC, two cancers significantly driven by inflammation.

4. Inflammation and Antitumor Immunity in Hepatocellular Carcinoma

With 745,000 deaths annually, liver cancer is the second biggest cause of cancer deaths worldwide [40]. Hepatocellular carcinoma (HCC) is the predominant form of primary liver cancer. It has several known risk factors, such as chronic hepatitis virus infections, alcohol abuse, autoimmune hepatitis, and several metabolic diseases, including diabetes mellitus [41]. HCC is driven by chronic inflammation [42]. The underlying epidemiology of chronic liver diseases has shifted from being caused by the hepatitis B virus (HBV) or hepatitis C virus (HCV) infection to metabolic disorders, such as non-alcoholic steatohepatitis (NASH) [43]. Regardless of the etiology, most HCCs have underlying cirrhosis caused by chronic liver inflammation [44,45].

Chronic liver inflammation damages hepatocytes, resulting in cell death. This cell death induces compensatory cell proliferation in the liver, a highly regenerative organ.

Meanwhile, the inflammation-induced ROS cause DNA damage and consequently increase the frequency of oncogenic mutations. Thus, the compensatory cell proliferation coupled with the accumulated oncogenic mutations induces malignant transformations in the liver [46,47]. Additionally, chronic inflammation supports tumor development and progression by altering the liver immune environments with several mechanisms, allowing malignant cells to escape from immune surveillance.

The immune system eradicates emerging cancer cells. The innate immune cells, including macrophages, DCs, and NK cells, are involved in the first line of defense, i.e., monitoring the pathogens and tumor cells. On the other hand, adaptive immunity also contributes to antitumor immunity via the immune surveillance by the CD4⁺ T cells and CD8⁺ T cells (Figure 1).

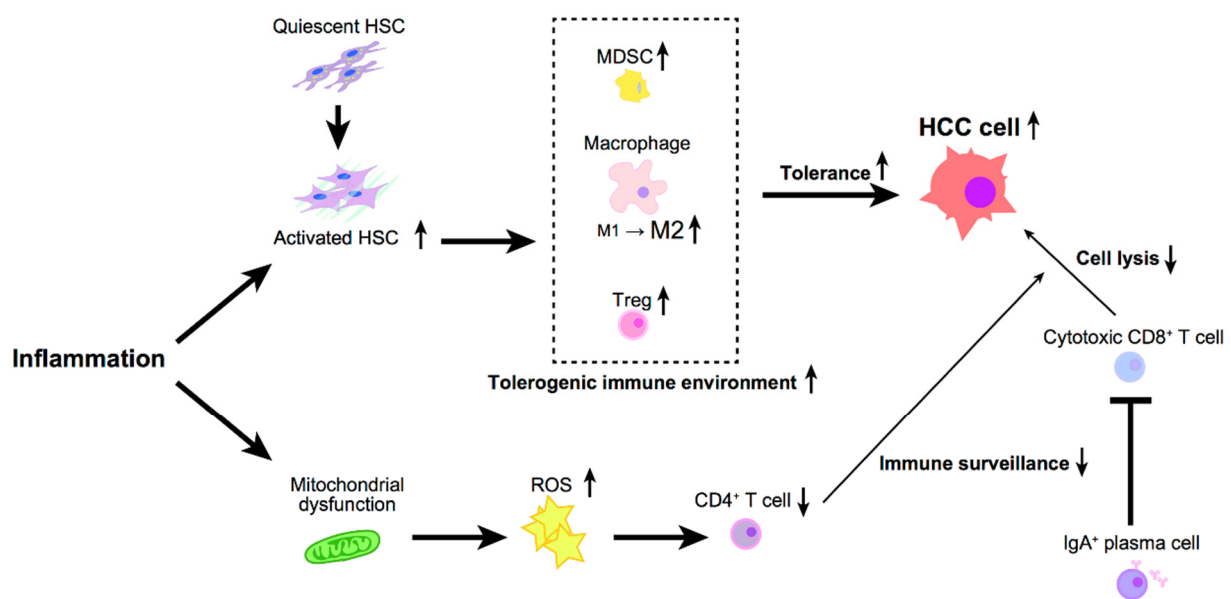


Figure 1. Inflammation oriented cancer-associated immune environment in hepatocellular carcinoma (HCC).

The activation of innate immune receptors, such as Toll-like receptors (TLRs), via IL-6 induction through a universal TLRs adaptor, MyD88, plays a considerable role in tumor development in the liver [48,49]. Because approximately 75% of the hepatic blood is supplied through the portal vein, the liver is exposed to the potentially immunogenic dietary and microbial products from the gut [50], such as lipopolysaccharide (LPS), a gram-negative bacterial cell component; LPS is also a ligand for TLR4. TLR4 plays a crucial role in hepatocarcinogenesis. TLR4 deficiency ameliorated HCC's burden in several HCC mouse models, including the diethylnitrosamine (DEN) and carbon tetrachloride (CCl₄) induction model, or the mice with knocked-out genes encoding the hepatocyte-specific transforming growth factor- β -activated kinase 1 (*TAK1*), phosphatase, or the tensin homolog (*PTEN*) [51–53]. The importance of gut-derived LPS has been well studied. LPS depletion by oral antibiotics administration decreased tumor burden in these HCC mice models, while continuous LPS administration enhanced hepatocarcinogenesis [51,53]. Collectively, these data strongly suggest that TLR4 and gut-derived LPS play a critical role in HCC development.

On the other hand, the repression of adaptive immunity also plays a role in HCC development. The number of intrahepatic CD4⁺ T cells is reduced in livers with non-alcoholic fatty liver disease (NAFLD), accelerating hepatocarcinogenesis [54]. Fatty acid accumulation in the NAFLD liver induced the production of mitochondrial ROS, promoting the loss of CD4⁺ T cells and impairing immune surveillance. Moreover, the role of immunoglobulin A (IgA)-producing plasma cells in NASH-associated HCC development was recently reported [55]. There was an accumulation of IgA⁺ cells in the liver of mice with NASH-

induced HCC. These IgA⁺ cells produce PD-L1 and IL-10 to suppress antitumor cytotoxic CD8⁺ T cells, thus promoting HCC. Genetic or pharmacological interference with IgA⁺ cells generation attenuated hepatocarcinogenesis, achieving cytotoxic T cell-mediated HCC regression. These data suggest that the inflammation-induced suppression of cytotoxic CD8⁺ T cells was achieved through IgA⁺ plasma cells, leading to the development of HCC.

Hepatic stellate cells (HSCs) significantly modulate the liver's immune microenvironment in collaboration with immune cells, despite their limited capacity as antigen-presenting cells (APCs) [56] (Figure 1). HSCs are extracellular matrix-producing stromal cells involved in liver fibrosis; the activation of HSCs promotes HCC development [57,58]. In addition, HSCs change macrophage polarization from an inflammatory (M1) to an immunosuppressive (M2) signature, promoting tumor progression [59,60].

Besides, HSCs contribute to the escape of HCC cells from immune surveillance by regulating the population of Treg cells and MDSCs. HSCs inhibit lymphocyte infiltration in tumors, induce apoptosis of infiltrating mononuclear cells, and enhance the expression of PD-L1 and the number of CD4⁺CD25⁺ Treg cells in immunocompetent mice [57]. HSCs store retinoids to produce retinoic acid during activation. It has been shown that isolated murine HSCs enhance the differentiation of isolated naive OT-II T cells co-cultured with splenic DCs in the presence of TGF- β 1 into functional Foxp3⁺ Treg cells in an all-trans-retinoic acid-dependent manner. This study indicates that HSCs influence DC-primed T cells to differentiate into functional Treg cells, resulting in the liver's tolerogenicity [61].

The capacity of HSCs to transform peripheral blood monocytes into MDSCs has been studied [62]. Isolated human monocytes from healthy donors co-cultured with human HSCs display downregulation of HLA-DR and develop a phenotype similar to that of MDSCs. This induction of MDSCs is only achieved upon co-culture with activated HSCs but not with quiescent HSCs, and is mediated by the cells' physical interaction via CD44 but not by soluble factors.

HSCs also suppress immunity by inhibiting cytotoxic CD8⁺ T cell activation via a CD54-mediated mechanism [63]. Isolated murine HSCs or human HSC cell line LX-2 prevent the priming of naive CD8⁺ T cells by APCs. The inhibition of T cell activation is associated with HSC activation. Mechanistically, the increased level of CD54 on HSCs restricts the production of IL-2 receptor and IL-2 in T cells, which attenuates T cell activation.

Most of the tolerogenic mechanisms of HSCs depend on the activated state of HSCs. HSCs are activated in the inflamed liver via several mechanisms. For example, inflammation activates caspase-1 and increases the release of IL-1 β , which facilitates parenchymal liver injury. Furthermore, danger-associated molecular patterns (DAMPs) released from dying hepatocytes promote HSCs activation via inflammasome signaling; NACHT, LRR, and PYD domains-containing protein 3 (NLRP3), a significant component of the inflammasome and a downstream of DAMPs, contribute to HSC activation [64]. In addition, IL-33, which is released from damaged hepatocytes, also contributes to HSC activation through the expansion of innate lymphoid cells [65]. Moreover, hepatic macrophages, recruited in the damaged liver, produce cytokines and chemokines, including TGF- β , PDGF, TNF, IL-1 β , CCL3, and CCL5, which lead to HSC activation [66–69]. As mentioned above, HSCs are activated during liver inflammation; inflammation plays a crucial role in immune suppression via HSCs activation.

5. Inflammation-Induced Immunoregulation in Colorectal Cancer

Chronic inflammation is associated with CRC, especially IBD-induced CRC. Chronic inflammation even plays a crucial role in the development and progression of sporadic CRC, also known as "tumor-elicited inflammation", by inducing the loss of the tumor suppressor adenomatous polyposis coli (APC) [70–72] (Figure 2). The loss of APC in colonic epithelial cells induces adenoma formation through β -catenin activation, resulting in a decrease in the mucus layer and barrier dysfunction [70]. Consequently, gut bacteria or bacterial products or both, such as LPS, translocate into the lamina propria and activate the TAMs. The TAMs produce pro-inflammatory cytokines, including IL-23, which activates IL-

IL-17-producing cells, such as Th17 cells. Finally, pro-inflammatory cytokines, including TNF, IL-6, and IL-17 produced by IL-17-producing cells, directly promote the proliferation and survival of adenoma cells via the activation of NF- κ B and STAT3 [70,71]. TNF and IL-6 are two major pro-inflammatory cytokines produced at high levels in human CRC; they mainly activate the IKK-NF- κ B and JAK-STAT3 pathways, respectively [73,74]. IL-6 also activates the Ras-MAPK, PI3K-Akt, and Src family kinase (SFK)-YAP pathways [74,75]. In addition to activating β -catenin, the loss of APC continuously activates the SFK-YAP pathway by establishing a positive feedback loop through the upregulation of gp130/IL6ST, a common receptor for the IL-6 family cytokines. The upregulation of gp130 on the tumor cell surface also leads to the response to a low concentration of IL-6 in low-grade inflammation [72]. IL-6 suppresses antitumor immunity by several mechanisms [76,77]. In a mouse experiment using the mouse colon cancer cell line CT-26, IL-6 deletion in mice enhances antitumor immunity by augmenting type-1 immunity, suggesting the immunosuppressive role of IL-6 in the tumor microenvironment [78]. On the other hand, it is unclear whether TNF is involved in immunosuppression.

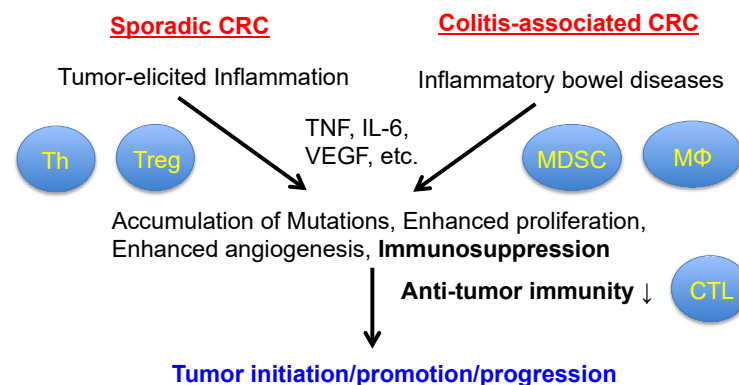


Figure 2. The role of inflammation in colorectal tumorigenesis.

6. Induction of Aging-Related Diseases by Inflammation

Aging is a complex and heterogeneous process characterized by the progressive loss of physiological integrity that leads to functional impairment and increased susceptibility to death [79]. Aging is affected by environmental, genetic, and epigenetic factors; it is a significant risk factor for cancer [80]. Recent studies revealed that chronic inflammation functions as a critical driver of aging and age-related diseases [81–86]. Inflammaging is one of the hallmarks of aging. It is indicated by the increased levels of circulating TNF, IL-6, and C-reactive protein and defined as an age-related chronic, systemic, and unresolved low-grade inflammation [87,88]. Inflammaging causes cytokine dysregulation and affects the immune system with the progressive activation of macrophages and related cells, potentially linking aging to immunosenescence and age-related diseases, such as cancer, Alzheimer’s disease, rheumatoid arthritis, cardiovascular diseases, and type II diabetes [89].

Various cells and molecules are involved in inflammaging. Although many cell types, including epithelial, fibroblast, and endothelial, are related to inflammation in age-related diseases, immune cells mainly induce chronic inflammation during aging.

It is thought that the continuous upregulation of pro-inflammatory mediators, such as TNF, IL-6, COX-2, and iNOS, is induced during aging due to the imbalance in redox status and oxidative stress that activate pro-inflammatory signaling pathways, including NF- κ B, and STAT3 [90,91]. A two to four-fold increase in the serum levels of inflammatory mediators is observed during aging [89]. Additionally, it is reported that coagulation factors activate inflammatory signaling beyond their original role in the coagulation system [82]. Moreover, microRNAs link inflammaging with cellular senescence and cancer [92].

Also, some cellular processes are involved in inflammaging. The upregulation of endoplasmic reticulum (ER) stress as well as the accumulation of inflammasome and lipid

are also observed during aging and might play an important role in age-related diseases. Lastly, cellular senescence and dysregulated innate immunity have been found to cause prolonged chronic inflammation even after the initial stimulus has been removed [82].

Aging is accompanied by the change of epigenetic information in both dividing and non-dividing cells. Epigenetic changes significantly influence the aging process, life span, and induction and progression of age-related diseases, including cancer [93–95]. Diet plays an essential role in inducing epigenetic changes; however, it is challenging to predict which epigenetic alterations induced by different diets are beneficial or detrimental for aging. Further studies are required.

7. Liver Diseases with Aging

Most human malignancies, including HCC, are considered age-related diseases because their incidences increase with age [42,96]. Aging is an independent risk factor, at a hazard ratio of 1.02 per year, for the overall survival of HCC patients with both hepatitis virus-related or non-viral etiologies [97,98].

An increased number of senescent hepatocytes are observed among chronic liver diseases of several etiologies. In the aged liver, the number of senescent cells is also increased in hepatocytes and stromal cells, including stellate and immune cells. Cellular senescence develops due to telomere shortening and accumulated DNA damage mediated by oxidative stress [99]. Morphological changes in hepatocytes in aged livers can be associated with increased polyploidy and mitochondrial dysfunction [100,101], which enhance the liver's susceptibility to damage and decline in regeneration [102].

Meanwhile, senescent cells also secrete pro-inflammatory cytokines to accelerate the development of chronic liver diseases [103]. In addition, aged hepatic stellate cells demonstrate enhanced activity [104]. The aged liver sinusoidal endothelial cells display dysregulated vasodilatory capacity, which leads to portal hypertension contributing to developing liver cirrhosis and liver dysfunction in chronic liver diseases [104]. On the other hand, the aging of Kupffer cells, the residual liver macrophages, has been reported to be heterogeneous and context-dependent [105].

Considering the importance of chronic inflammation and the dysregulated immune surveillance in the pathogenesis of chronic liver diseases and HCC, it is likely that inflammaging plays a specific role in these liver diseases. There is accumulating evidence demonstrating inflammaging's contribution to immune dysfunction [106]. However, it remains to be elucidated whether and how inflammaging contributes to the pathogenesis of liver diseases, including HCC.

8. Inflammation and Aging in the Gastrointestinal Tract

During aging, somatic clones with driver mutations expand in normal tissues, resulting in cancer development. During inflammaging, the pathophysiological alterations in the colon occur at molecular and cellular levels, and the regenerative ability of the epithelium is decreased by inflammation [86,107]. Recently, two Japanese groups have reported similar findings that such clonal expansion occurs in the inflamed colon epithelium of patients with ulcerative colitis [108,109]. The epithelium accumulates somatic mutations in several genes associated with IL-17 signaling, including *NFKBIZ*, *TRAF3IP2*, *ZC3H12A*, *PIGR*, and *HNRNPF*. These mutations lead to the downregulation of IL-17 and other pro-inflammatory signals. However, these somatic mutations are rarely found in sporadic and colitis-associated CRC, suggesting the negative selection for mutated epithelium during CRC development.

The development of esophageal cancer, especially esophageal squamous cell carcinoma, is highly associated with chronic inflammation induced by tobacco smoking and alcohol drinking and pro-inflammatory cytokines, including the IL-6 family cytokines [74,110]. The clonal expansion of epithelium in the inflamed colon is also observed in the esophageal epithelium during aging [111]. The progressive age-related expansion of the esophageal epithelial clones with driver mutations (predominantly *NOTCH1*) is substantially acceler-

ated by alcohol drinking and heavy smoking. A high frequency of *NOTCH1* and *PPM1D* mutations are observed in the physiologically normal esophageal epithelial cells, compared to the common mutations in esophageal carcinoma. Therefore, the observed remodeling of the esophageal epithelium by driver-mutated clones is an inevitable result of normal aging, leading to esophageal tumorigenesis.

9. Immune Signal Modulation as a Potential Therapeutic Target of HCC and CRC

An immunologically active “inflamed” tumor microenvironment has been associated with improved prognosis in HCC patients [112–114]. In one instance, the intratumor immune environment of 234 primary HCCs was assessed by examining their cytolytic activity (CYT), defined as the average production of granzyme A and perforin [115], and was found to be immunosuppressive, consistent with the discussion above [116]. The immune gene signatures of these primary HCCs were characterized to categorize the HCCs into four subclasses by different active immunosuppressive mechanisms: Tumor-associated macrophage (TAM), CTNNB1, CYT, and Treg. Interestingly, the Treg subgroup showed poorer prognosis, suggesting that the impact of Treg induction had a pivotal role in shaping the tolerogenic state of the HCCs.

Immune checkpoint molecules are critical in inhibiting antitumor immunity; among them, PD-1 and CTLA-4 have been studied extensively [117]. Therapeutic strategies targeting these molecules aim to block T cell-mediated tumor immunosuppression and Treg activity, consistent with a previous report that emphasizes the significance of Treg activity in a tumor immunosuppressive environment [116].

Clinical trials have been carried out to test monoclonal antibodies targeting immune checkpoint molecules; these therapeutics have been found efficacious in suppressing tumor growth, resulting in favorable patient prognosis. Thus, they have already been approved by the United States Food and Drug Administration (FDA) for unresectable or metastatic HCC [118–120] and CRC [121,122] (Tables 1 and 2). Immune checkpoint inhibitor therapy is only effective in CRC patients who have microsatellite instability-high (MSI-H) or mismatch repair-deficient (dMMR) tumors or both. Recently, pembrolizumab was approved by FDA as a first-line therapy for patients with unresectable or metastatic MSI-H or dMMR CRC.

An animal experiment using the transplantation model of CRC cells demonstrated that a combination of PD-1 antibodies with VEGF inhibition achieved a strong and synergic antitumor effect, consistent with the previous finding that VEGF produced by tumors shapes the immunosuppressive microenvironment. Lenvatinib, a multi-kinase inhibitor with anti-angiogenic potential and a clinically used agent in advanced HCC patients, induced more significant tumor regression combined with the PD-1 antibodies in the HCC mouse model [123]. Indeed, an emerging regimen involving a PD-L1 monoclonal antibody, atezolizumab, and the VEGF inhibitor bevacizumab has recently received FDA approval for its demonstrated improvement in the overall and progression-free survival in patients with unresectable HCC compared to that of the approved first-line treatment with a multi-kinase inhibitor, sorafenib [124].

Table 1. FDA-approved checkpoint molecules in HCC.

Drug	Target	Line of Therapy	Administration
Atezolizumab	Fully human, anti-PD-L1 IgG1 monoclonal antibody	1st	Used in combination with bevacizumab for patients with unresectable HCC
Nivolumab	Fully human, anti-PD-1 IgG4 monoclonal antibody	2nd	Used alone or with ipilimumab for patients who have already been treated with sorafenib
Pembrolizumab	Fully human, anti-PD-1 IgG4 monoclonal antibody	2nd	Used for patients who have already been treated with sorafenib
Ipilimumab	Fully human anti-CTLA-4 IgG1 monoclonal antibody	2nd	Used in combination with nivolumab for patients who already been treated with sorafenib

Table 2. FDA-approved checkpoint molecules in CRC.

Drug	Target	Line of Therapy	Administration
Pembrolizumab	Fully human, anti-PD-1 IgG4 monoclonal antibody	1st	Used alone for patients with unresectable or metastatic MSI-H/dMMR CRC
Nivolumab	Fully human, anti-PD-1 IgG4 monoclonal antibody	2nd	Used in combination with ipilimumab for patients with metastatic MSI-H/dMMR CRC that progressed following treatment with a fluoropyrimidine, oxaliplatin, and irinotecan
Ipilimumab	Fully human anti-CTLA-4 IgG1 monoclonal antibody	2nd	Used in combination with nivolumab for patients with metastatic MSI-H/dMMR CRC that progressed following treatment with a fluoropyrimidine, oxaliplatin, and irinotecan

Also, non-steroidal anti-inflammatory drugs (NSAIDs) are shown to have both protective and therapeutic effects against cancer [125]. The long-term use of NSAIDs reduces the risk of familial adenomatous polyposis as well as sporadic CRC. In addition, NSAIDs decrease the incident and recurrent risk of HCC. However, the NSAIDs' mechanisms of suppressing tumorigenesis and metastasis are still controversial.

10. Conclusions and Perspective

Chronic inflammation, which is closely related to establishing the immunosuppressive tumor microenvironment, dampens the effect of cancer immunotherapeutics, such as immune checkpoint inhibitors and CAR-T cells. Therefore, it is vital to develop a method to shift the immunosuppressive tumor microenvironment to an immunocompetent state. IL-6 might be one of the attractive targets for improving the efficacy of cancer immunotherapy. It might also delay aging and alleviate age-related diseases by modulating the pro-inflammatory environment and subsequently resolving inflammation.

Author Contributions: Writing—review and editing, N.N., Y.K., D.A., H.N. and K.T. All authors have read and agreed to the published version of the manuscript.

Funding: This work is supported by JSPS KAKENHI (JP 15K21775, JP 20H03758), AMED (PRIME) under Grant Number JP 18gm6210008/19gm6210008/20gm6210008, the “Kibou Projects” Startup Support for Young Researchers in Immunology, the Keio Gijuku Academic Development Funds, the Uehara Memorial Foundation, the Kanae Foundation for the Promotion of Medical Science, the Astellas Foundation for Research on Metabolic Disorders, the SENSHIN Medical Research Foundation, research grants from Bristol-Myers Squibb, the SGH foundation, the MSD Life Science Foundation, the Ichiro Kanehara Foundation for the Promotion of Medical Sciences and Medical Care, the Yasuda Medical Foundation, the Suzuken Memorial Foundation, the Pancreas Research Foundation of Japan, the Waksman Foundation of Japan Inc., the Japanese Foundation for Multidisciplinary Treatment of Cancer, Project Mirai Cancer Research Grants from the Japan Cancer Society, the Okinaka Memorial Institute for Medical Research, the Asahi Glass Foundation, the Foundation for Promotion of Cancer Research, the Kobayashi Foundation for Cancer Research, the Toray Science Foundation, the Vehicle Racing Commemorative Foundation, the JSR-Keio University Medical and Chemical Innovation Center (JKiC), a research grant from the Public Trust Surgery Research Fund, a Research Grant of the Princess Takamatsu Cancer Research Fund, the Tokyo Biomedical Foundation, the Daiichi Sankyo Foundation of Life Science, the Mochida Memorial Foundation for Medical and Pharmaceutical Research, the Medical Research Encouragement Prize of the Japan Medical Association, the Terumo Foundation for Life Sciences and Arts, the Yakult-Bioscience Foundation, the Novartis Foundation, the Mitsubishi Foundation, and the Takeda Science Foundation (all to K.T.). This work is also supported by Bristol-Myers Squibb Research Grant, Takeda Science Foundation, MSD Life Science Foundation, The Naito Foundation, Life Science Foundation of Japan, The Cell Science Research Foundation, KAKENHI Grant Number 18K07994, AMED under Grant Number JP19fk0210059, JP20fk0210059, JP18fk0210040, JP19fk0210040, and JP20fk0210040 (all to H.N.).

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Karin, M.; Clevers, H. Reparative inflammation takes charge of tissue regeneration. *Nature* **2016**, *529*, 307–315. [[CrossRef](#)]
2. Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. *Cell* **2011**, *144*, 646–674. [[CrossRef](#)]
3. Grivnenkov, S.I.; Greten, F.R.; Karin, M. Immunity, inflammation, and cancer. *Cell* **2010**, *140*, 883–899. [[CrossRef](#)]
4. Chamoto, K.; Hatae, R.; Honjo, T. Current issues and perspectives in PD-1 blockade cancer immunotherapy. *Int. J. Clin. Oncol.* **2020**, *25*, 790–800. [[CrossRef](#)]
5. Barber, D.L.; Wherry, E.J.; Masopust, D.; Zhu, B.; Allison, J.P.; Sharpe, A.H.; Freeman, G.J.; Ahmed, R. Restoring function in exhausted CD8 T cells during chronic viral infection. *Nature* **2006**, *439*, 682–687. [[CrossRef](#)]
6. Jones, R.B.; Ndhlovu, L.C.; Barbour, J.D.; Sheth, P.M.; Jha, A.R.; Long, B.R.; Wong, J.C.; Satkunarajah, M.; Schwenker, M.; Chapman, J.M.; et al. Tim-3 expression defines a novel population of dysfunctional T cells with highly elevated frequencies in progressive HIV-1 infection. *J. Exp. Med.* **2008**, *205*, 2763–2779. [[CrossRef](#)]
7. Blackburn, S.D.; Shin, H.; Haining, W.N.; Zou, T.; Workman, C.J.; Polley, A.; Betts, M.R.; Freeman, G.J.; Vignali, D.A.; Wherry, E.J. Coregulation of CD8+ T cell exhaustion by multiple inhibitory receptors during chronic viral infection. *Nat. Immunol.* **2009**, *10*, 29–37. [[CrossRef](#)]
8. Johnston, R.J.; Comps-Agrar, L.; Hackney, J.; Yu, X.; Huseni, M.; Yang, Y.; Park, S.; Javinal, V.; Chiu, H.; Irving, B.; et al. The immunoreceptor TIGIT regulates antitumor and antiviral CD8(+) T cell effector function. *Cancer Cell* **2014**, *26*, 923–937. [[CrossRef](#)]
9. Blank, C.U.; Haining, W.N.; Held, W.; Hogan, P.G.; Kallies, A.; Lugli, E.; Lynn, R.C.; Philip, M.; Rao, A.; Restifo, N.P.; et al. Defining ‘T cell exhaustion’. *Nat. Rev. Immunol.* **2019**, *19*, 665–674. [[CrossRef](#)]
10. Liu, X.; Wang, Y.; Lu, H.; Li, J.; Yan, X.; Xiao, M.; Hao, J.; Alekseev, A.; Khong, H.; Chen, T.; et al. Genome-wide analysis identifies NR4A1 as a key mediator of T cell dysfunction. *Nature* **2019**, *567*, 525–529. [[CrossRef](#)] [[PubMed](#)]
11. Scott, A.C.; Dündar, F.; Zumbo, P.; Chandran, S.S.; Klebanoff, C.A.; Shakiba, M.; Trivedi, P.; Menocal, L.; Appleby, H.; Camara, S.; et al. TOX is a critical regulator of tumour-specific T cell differentiation. *Nature* **2019**, *571*, 270–274. [[CrossRef](#)] [[PubMed](#)]
12. Khan, O.; Giles, J.R.; McDonald, S.; Manne, S.; Ngiow, S.F.; Patel, K.P.; Werner, M.T.; Huang, A.C.; Alexander, K.A.; Wu, J.E.; et al. TOX transcriptionally and epigenetically programs CD8(+) T cell exhaustion. *Nature* **2019**, *571*, 211–218. [[CrossRef](#)] [[PubMed](#)]
13. Alfei, F.; Kanev, K.; Hofmann, M.; Wu, M.; Ghoneim, H.E.; Roelli, P.; Utzschneider, D.T.; von Hoesslin, M.; Cullen, J.G.; Fan, Y.; et al. TOX reinforces the phenotype and longevity of exhausted T cells in chronic viral infection. *Nature* **2019**, *571*, 265–269. [[CrossRef](#)]
14. Shin, H.; Blackburn, S.D.; Intlekofer, A.M.; Kao, C.; Angelosanto, J.M.; Reiner, S.L.; Wherry, E.J. A role for the transcriptional repressor Blimp-1 in CD8(+) T cell exhaustion during chronic viral infection. *Immunity* **2009**, *31*, 309–320. [[CrossRef](#)]
15. Quigley, M.; Pereyra, F.; Nilsson, B.; Porichis, F.; Fonseca, C.; Eichbaum, Q.; Julg, B.; Jesneck, J.L.; Brosnahan, K.; Imam, S.; et al. Transcriptional analysis of HIV-specific CD8+ T cells shows that PD-1 inhibits T cell function by upregulating BATF. *Nat. Med.* **2010**, *16*, 1147–1151. [[CrossRef](#)]
16. Zhang, Z.; Liu, S.; Zhang, B.; Qiao, L.; Zhang, Y.; Zhang, Y. T Cell Dysfunction and Exhaustion in Cancer. *Front. Cell Dev. Biol.* **2020**, *8*, 17. [[CrossRef](#)]
17. Hibino, S.; Chikuma, S.; Kondo, T.; Ito, M.; Nakatsukasa, H.; Omata-Mise, S.; Yoshimura, A. Inhibition of Nr4a Receptors Enhances Antitumor Immunity by Breaking Treg-Mediated Immune Tolerance. *Cancer Res.* **2018**, *78*, 3027–3040. [[CrossRef](#)]
18. Arce-Sillas, A.; Álvarez-Luquín, D.D.; Tamaya-Domínguez, B.; Gomez-Fuentes, S.; Trejo-García, A.; Melo-Salas, M.; Cárdenas, G.; Rodríguez-Ramírez, J.; Adalid-Peralta, L. Regulatory T Cells: Molecular Actions on Effector Cells in Immune Regulation. *J. Immunol. Res.* **2016**, *2016*, 1720827. [[CrossRef](#)]
19. Curiel, T.J.; Coukos, G.; Zou, L.; Alvarez, X.; Cheng, P.; Mottram, P.; Evdemon-Hogan, M.; Conejo-Garcia, J.R.; Zhang, L.; Burow, M.; et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat. Med.* **2004**, *10*, 942–949. [[CrossRef](#)]
20. Mizukami, Y.; Kono, K.; Kawaguchi, Y.; Akaike, H.; Kamimura, K.; Sugai, H.; Fujii, H. CCL17 and CCL22 chemokines within tumor microenvironment are related to accumulation of Foxp3+ regulatory T cells in gastric cancer. *Int. J. Cancer* **2008**, *122*, 2286–2293. [[CrossRef](#)]
21. van der Veeken, J.; Gonzalez, A.J.; Cho, H.; Arvey, A.; Hemmers, S.; Leslie, C.S.; Rudensky, A.Y. Memory of Inflammation in Regulatory T Cells. *Cell* **2016**, *166*, 977–990. [[CrossRef](#)]
22. Saleh, R.; Elkord, E. FoxP3(+) T regulatory cells in cancer: Prognostic biomarkers and therapeutic targets. *Cancer Lett.* **2020**, *490*, 174–185. [[CrossRef](#)]
23. Saito, T.; Nishikawa, H.; Wada, H.; Nagano, Y.; Sugiyama, D.; Atarashi, K.; Maeda, Y.; Hamaguchi, M.; Ohkura, N.; Sato, E.; et al. Two FOXP3(+)CD4(+) T cell subpopulations distinctly control the prognosis of colorectal cancers. *Nat. Med.* **2016**, *22*, 679–684. [[CrossRef](#)]
24. Gounaris, E.; Blatner, N.R.; Dennis, K.; Magnusson, F.; Gurish, M.F.; Strom, T.B.; Beckhove, P.; Gounari, F.; Khazaie, K. T-regulatory cells shift from a protective anti-inflammatory to a cancer-promoting proinflammatory phenotype in polyposis. *Cancer Res.* **2009**, *69*, 5490–5497. [[CrossRef](#)]
25. Nakamura, T.; Ushigome, H. Myeloid-Derived Suppressor Cells as a Regulator of Immunity in Organ Transplantation. *Int. J. Mol. Sci.* **2018**, *19*, 2357. [[CrossRef](#)]
26. Wang, D.; DuBois, R.N. Immunosuppression associated with chronic inflammation in the tumor microenvironment. *Carcinogenesis* **2015**, *36*, 1085–1093. [[CrossRef](#)]

27. Ha, H.; Debnath, B.; Neamati, N. Role of the CXCL8-CXCR1/2 Axis in Cancer and Inflammatory Diseases. *Theranostics* **2017**, *7*, 1543–1588. [[CrossRef](#)] [[PubMed](#)]
28. Hanahan, D.; Weinberg, R.A. The hallmarks of cancer. *Cell* **2000**, *100*, 57–70. [[CrossRef](#)]
29. Tong, R.T.; Boucher, Y.; Kozin, S.V.; Winkler, F.; Hicklin, D.J.; Jain, R.K. Vascular normalization by vascular endothelial growth factor receptor 2 blockade induces a pressure gradient across the vasculature and improves drug penetration in tumors. *Cancer Res.* **2004**, *64*, 3731–3736. [[CrossRef](#)] [[PubMed](#)]
30. Winkler, F.; Kozin, S.V.; Tong, R.T.; Chae, S.S.; Booth, M.F.; Garkavtsev, I.; Xu, L.; Hicklin, D.J.; Fukumura, D.; di Tomaso, E.; et al. Kinetics of vascular normalization by VEGFR2 blockade governs brain tumor response to radiation: Role of oxygenation, angiopoietin-1, and matrix metalloproteinases. *Cancer Cell* **2004**, *6*, 553–563. [[CrossRef](#)]
31. Willett, C.G.; Boucher, Y.; di Tomaso, E.; Duda, D.G.; Munn, L.L.; Tong, R.T.; Chung, D.C.; Sahani, D.V.; Kalva, S.P.; Kozin, S.V.; et al. Direct evidence that the VEGF-specific antibody bevacizumab has antivascular effects in human rectal cancer. *Nat. Med.* **2004**, *10*, 145–147. [[CrossRef](#)] [[PubMed](#)]
32. Gabrilovich, D.I.; Chen, H.L.; Girgis, K.R.; Cunningham, H.T.; Meny, G.M.; Nadaf, S.; Kavanaugh, D.; Carbone, D.P. Production of vascular endothelial growth factor by human tumors inhibits the functional maturation of dendritic cells. *Nat. Med.* **1996**, *2*, 1096–1103. [[CrossRef](#)]
33. Horikawa, N.; Abiko, K.; Matsumura, N.; Hamanishi, J.; Baba, T.; Yamaguchi, K.; Yoshioka, Y.; Koshiyama, M.; Konishi, I. Expression of Vascular Endothelial Growth Factor in Ovarian Cancer Inhibits Tumor Immunity through the Accumulation of Myeloid-Derived Suppressor Cells. *Clin. Cancer Res.* **2017**, *23*, 587–599. [[CrossRef](#)]
34. Ohm, J.E.; Carbone, D.P. VEGF as a mediator of tumor-associated immunodeficiency. *Immunol. Res.* **2001**, *23*, 263–272. [[CrossRef](#)]
35. Kim, C.G.; Jang, M.; Kim, Y.; Leem, G.; Kim, K.H.; Lee, H.; Kim, T.S.; Choi, S.J.; Kim, H.D.; Han, J.W.; et al. VEGF-A drives TOX-dependent T cell exhaustion in anti-PD-1-resistant microsatellite stable colorectal cancers. *Sci. Immunol.* **2019**, *4*. [[CrossRef](#)] [[PubMed](#)]
36. Voron, T.; Colussi, O.; Marcheteau, E.; Pernot, S.; Nizard, M.; Pointet, A.L.; Latreche, S.; Bergaya, S.; Benhamouda, N.; Tanchot, C.; et al. VEGF-A modulates expression of inhibitory checkpoints on CD8+ T cells in tumors. *J. Exp. Med.* **2015**, *212*, 139–148. [[CrossRef](#)] [[PubMed](#)]
37. Terme, M.; Pernot, S.; Marcheteau, E.; Sandoval, F.; Benhamouda, N.; Colussi, O.; Dubreuil, O.; Carpentier, A.F.; Tartour, E.; Taieb, J. VEGFA-VEGFR pathway blockade inhibits tumor-induced regulatory T-cell proliferation in colorectal cancer. *Cancer Res.* **2013**, *73*, 539–549. [[CrossRef](#)]
38. Senga, S.S.; Grose, R.P. Hallmarks of cancer—the new testament. *Open Biol.* **2021**, *11*, 200358. [[CrossRef](#)]
39. Pan, Z.; Tian, Y.; Niu, G.; Cao, C. Role of microRNAs in remodeling the tumor microenvironment (Review). *Int. J. Oncol.* **2020**, *56*, 407–416. [[CrossRef](#)]
40. Ferlay, J.; Soerjomataram, I.; Dikshit, R.; Eser, S.; Mathers, C.; Rebelo, M.; Parkin, D.M.; Forman, D.; Bray, F. Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. *Int. J. Cancer* **2015**, *136*, E359–E386. [[CrossRef](#)]
41. El-Serag, H.B. Hepatocellular carcinoma. *N. Engl. J. Med.* **2011**, *365*, 1118–1127. [[CrossRef](#)]
42. Villanueva, A. Hepatocellular Carcinoma. *N. Engl. J. Med.* **2019**, *380*, 1450–1462. [[CrossRef](#)] [[PubMed](#)]
43. Kim, D.; Li, A.A.; Perumpail, B.J.; Gadiparthi, C.; Kim, W.; Cholankeril, G.; Glenn, J.S.; Harrison, S.A.; Younossi, Z.M.; Ahmed, A. Changing Trends in Etiology-Based and Ethnicity-Based Annual Mortality Rates of Cirrhosis and Hepatocellular Carcinoma in the United States. *Hepatology* **2019**, *69*, 1064–1074. [[CrossRef](#)]
44. Ringelhan, M.; Pfister, D.; O'Connor, T.; Pikarsky, E.; Heikenwalder, M. The immunology of hepatocellular carcinoma. *Nat. Immunol.* **2018**, *19*, 222–232. [[CrossRef](#)]
45. Nakagawa, H.; Maeda, S. Inflammation- and stress-related signaling pathways in hepatocarcinogenesis. *World J. Gastroenterol.* **2012**, *18*, 4071–4081. [[CrossRef](#)] [[PubMed](#)]
46. Maeda, S.; Kamata, H.; Luo, J.L.; Leffert, H.; Karin, M. IKKbeta couples hepatocyte death to cytokine-driven compensatory proliferation that promotes chemical hepatocarcinogenesis. *Cell* **2005**, *121*, 977–990. [[CrossRef](#)]
47. Nakagawa, H.; Umemura, A.; Taniguchi, K.; Font-Burgada, J.; Dhar, D.; Ogata, H.; Zhong, Z.; Valasek, M.A.; Seki, E.; Hidalgo, J.; et al. ER stress cooperates with hypernutrition to trigger TNF-dependent spontaneous HCC development. *Cancer Cell* **2014**, *26*, 331–343. [[CrossRef](#)]
48. Naugler, W.E.; Sakurai, T.; Kim, S.; Maeda, S.; Kim, K.; Elsharkawy, A.M.; Karin, M. Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. *Science* **2007**, *317*, 121–124. [[CrossRef](#)]
49. Roh, Y.S.; Seki, E. Toll-like receptors in alcoholic liver disease, non-alcoholic steatohepatitis and carcinogenesis. *J. Gastroenterol. Hepatol.* **2013**, *28* (Suppl. 1), 38–42. [[CrossRef](#)]
50. Guillot, A.; Tacke, F. Liver Macrophages: Old Dogmas and New Insights. *Hepatology. Commun.* **2019**, *3*, 730–743. [[CrossRef](#)]
51. Dapito, D.H.; Mencin, A.; Gwak, G.Y.; Pradere, J.P.; Jang, M.K.; Mederacke, I.; Caviglia, J.M.; Khiabanian, H.; Adeyemi, A.; Bataller, R.; et al. Promotion of hepatocellular carcinoma by the intestinal microbiota and TLR4. *Cancer Cell* **2012**, *21*, 504–516. [[CrossRef](#)] [[PubMed](#)]
52. Song, I.J.; Yang, Y.M.; Inokuchi-Shimizu, S.; Roh, Y.S.; Yang, L.; Seki, E. The contribution of toll-like receptor signaling to the development of liver fibrosis and cancer in hepatocyte-specific TAK1-deleted mice. *Int. J. Cancer* **2018**, *142*, 81–91. [[CrossRef](#)] [[PubMed](#)]

53. Miura, K.; Ishioka, M.; Minami, S.; Horie, Y.; Ohshima, S.; Goto, T.; Ohnishi, H. Toll-like Receptor 4 on Macrophage Promotes the Development of Steatohepatitis-related Hepatocellular Carcinoma in Mice. *J. Biol. Chem.* **2016**, *291*, 11504–11517. [[CrossRef](#)] [[PubMed](#)]
54. Ma, C.; Kesarwala, A.H.; Eggert, T.; Medina-Echeverz, J.; Kleiner, D.E.; Jin, P.; Stroncek, D.F.; Terabe, M.; Kapoor, V.; ElGindi, M.; et al. NAFLD causes selective CD4(+) T lymphocyte loss and promotes hepatocarcinogenesis. *Nature* **2016**, *531*, 253–257. [[CrossRef](#)]
55. Shalapour, S.; Lin, X.J.; Bastian, I.N.; Brain, J.; Burt, A.D.; Aksenov, A.A.; Vrbanac, A.F.; Li, W.; Perkins, A.; Matsutani, T.; et al. Inflammation-induced IgA+ cells dismantle anti-liver cancer immunity. *Nature* **2017**, *551*, 340–345. [[CrossRef](#)] [[PubMed](#)]
56. Schildberg, F.A.; Kurts, C.; Knolle, P.A. Prominent regulatory but weak antigen-presenting cell function of hepatic stellate cells. *Hepatology* **2011**, *54*, 1108. [[CrossRef](#)]
57. Zhao, W.; Zhang, L.; Yin, Z.; Su, W.; Ren, G.; Zhou, C.; You, J.; Fan, J.; Wang, X. Activated hepatic stellate cells promote hepatocellular carcinoma development in immunocompetent mice. *Int. J. Cancer* **2011**, *129*, 2651–2661. [[CrossRef](#)]
58. Duran, A.; Hernandez, E.D.; Reina-Campos, M.; Castilla, E.A.; Subramaniam, S.; Raghunandan, S.; Roberts, L.R.; Kisseleva, T.; Karin, M.; Diaz-Meco, M.T.; et al. p62/SQSTM1 by Binding to Vitamin D Receptor Inhibits Hepatic Stellate Cell Activity, Fibrosis, and Liver Cancer. *Cancer Cell* **2016**, *30*, 595–609. [[CrossRef](#)]
59. Ji, J.; Eggert, T.; Budhu, A.; Forgues, M.; Takai, A.; Dang, H.; Ye, Q.; Lee, J.S.; Kim, J.H.; Greten, T.F.; et al. Hepatic stellate cell and monocyte interaction contributes to poor prognosis in hepatocellular carcinoma. *Hepatology* **2015**, *62*, 481–495. [[CrossRef](#)]
60. Lujambio, A.; Akkari, L.; Simon, J.; Grace, D.; Tschaharganeh, D.F.; Bolden, J.E.; Zhao, Z.; Thapar, V.; Joyce, J.A.; Krizhanovsky, V.; et al. Non-cell-autonomous tumor suppression by p53. *Cell* **2013**, *153*, 449–460. [[CrossRef](#)]
61. Dunham, R.M.; Thapa, M.; Velazquez, V.M.; Elrod, E.J.; Denning, T.L.; Pulendran, B.; Grakoui, A. Hepatic stellate cells preferentially induce Foxp3+ regulatory T cells by production of retinoic acid. *J. Immunol.* **2013**, *190*, 2009–2016. [[CrossRef](#)] [[PubMed](#)]
62. Hochst, B.; Schildberg, F.A.; Sauerborn, P.; Gabel, Y.A.; Gevensleben, H.; Goltz, D.; Heukamp, L.C.; Turler, A.; Ballmaier, M.; Gieseke, F.; et al. Activated human hepatic stellate cells induce myeloid derived suppressor cells from peripheral blood monocytes in a CD44-dependent fashion. *J. Hepatol.* **2013**, *59*, 528–535. [[CrossRef](#)]
63. Schildberg, F.A.; Wojtalla, A.; Siegmund, S.V.; Endl, E.; Diehl, L.; Abdullah, Z.; Kurts, C.; Knolle, P.A. Murine hepatic stellate cells veto CD8 T cell activation by a CD54-dependent mechanism. *Hepatology* **2011**, *54*, 262–272. [[CrossRef](#)]
64. Wree, A.; Eguchi, A.; McGeough, M.D.; Pena, C.A.; Johnson, C.D.; Canbay, A.; Hoffman, H.M.; Feldstein, A.E. NLRP3 inflammasome activation results in hepatocyte pyroptosis, liver inflammation, and fibrosis in mice. *Hepatology* **2014**, *59*, 898–910. [[CrossRef](#)]
65. McHedlidze, T.; Waldner, M.; Zopf, S.; Walker, J.; Rankin, A.L.; Schuchmann, M.; Voehringer, D.; McKenzie, A.N.; Neurath, M.F.; Pflanz, S.; et al. Interleukin-33-dependent innate lymphoid cells mediate hepatic fibrosis. *Immunity* **2013**, *39*, 357–371. [[CrossRef](#)] [[PubMed](#)]
66. Pradere, J.P.; Kluwe, J.; De Minicis, S.; Jiao, J.J.; Gwak, G.Y.; Dapito, D.H.; Jang, M.K.; Guenther, N.D.; Mederacke, I.; Friedman, R.; et al. Hepatic macrophages but not dendritic cells contribute to liver fibrosis by promoting the survival of activated hepatic stellate cells in mice. *Hepatology* **2013**, *58*, 1461–1473. [[CrossRef](#)]
67. Seki, E.; De Minicis, S.; Gwak, G.Y.; Kluwe, J.; Inokuchi, S.; Bursill, C.A.; Llovet, J.M.; Brenner, D.A.; Schwabe, R.F. CCR1 and CCR5 promote hepatic fibrosis in mice. *J. Clin. Investig.* **2009**, *119*, 1858–1870. [[CrossRef](#)]
68. Heinrichs, D.; Berres, M.L.; Nellen, A.; Fischer, P.; Scholten, D.; Trautwein, C.; Wasmuth, H.E.; Sahin, H. The chemokine CCL3 promotes experimental liver fibrosis in mice. *PLoS ONE* **2013**, *8*, e66106. [[CrossRef](#)] [[PubMed](#)]
69. Berres, M.L.; Koenen, R.R.; Rueland, A.; Zaldivar, M.M.; Heinrichs, D.; Sahin, H.; Schmitz, P.; Streetz, K.L.; Berg, T.; Gassler, N.; et al. Antagonism of the chemokine Ccl5 ameliorates experimental liver fibrosis in mice. *J. Clin. Investig.* **2010**, *120*, 4129–4140. [[CrossRef](#)]
70. Grivennikov, S.I.; Wang, K.; Mucida, D.; Stewart, C.A.; Schnabl, B.; Jauch, D.; Taniguchi, K.; Yu, G.Y.; Osterreicher, C.H.; Hung, K.E.; et al. Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. *Nature* **2012**, *491*, 254–258. [[CrossRef](#)] [[PubMed](#)]
71. Wang, K.; Kim, M.K.; Di Caro, G.; Wong, J.; Shalapour, S.; Wan, J.; Zhang, W.; Zhong, Z.; Sanchez-Lopez, E.; Wu, L.W.; et al. Interleukin-17 receptor signaling in transformed enterocytes promotes early colorectal tumorigenesis. *Immunity* **2014**, *41*, 1052–1063. [[CrossRef](#)]
72. Taniguchi, K.; Moroishi, T.; de Jong, P.R.; Krawczyk, M.; Grebbin, B.M.; Luo, H.; Xu, R.H.; Golob-Schwarzl, N.; Schweiger, C.; Wang, K.; et al. YAP-IL-6ST autoregulatory loop activated on APC loss controls colonic tumorigenesis. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 1643–1648. [[CrossRef](#)]
73. Taniguchi, K.; Karin, M. NF- κ B, inflammation, immunity and cancer: Coming of age. *Nat. Rev. Immunol.* **2018**, *18*, 309–324. [[CrossRef](#)] [[PubMed](#)]
74. Taniguchi, K.; Karin, M. IL-6 and related cytokines as the critical lynchpins between inflammation and cancer. *Semin. Immunol.* **2014**, *26*, 54–74. [[CrossRef](#)]
75. Taniguchi, K.; Wu, L.W.; Grivennikov, S.I.; de Jong, P.R.; Lian, I.; Yu, F.X.; Wang, K.; Ho, S.B.; Boland, B.S.; Chang, J.T.; et al. A gp130-Src-YAP module links inflammation to epithelial regeneration. *Nature* **2015**, *519*, 57–62. [[CrossRef](#)]

76. Liu, Q.; Yu, S.; Li, A.; Xu, H.; Han, X.; Wu, K. Targeting interleukin-6 to relieve immunosuppression in tumor microenvironment. *Tumour Biol. J. Int. Soc. Oncodev. Biol. Med.* **2017**, *39*, 1010428317712445. [[CrossRef](#)]
77. Kitamura, H.; Ohno, Y.; Toyoshima, Y.; Ohtake, J.; Homma, S.; Kawamura, H.; Takahashi, N.; Taketomi, A. Interleukin-6/STAT3 signaling as a promising target to improve the efficacy of cancer immunotherapy. *Cancer Sci.* **2017**, *108*, 1947–1952. [[CrossRef](#)]
78. Ohno, Y.; Toyoshima, Y.; Yurino, H.; Monma, N.; Xiang, H.; Sumida, K.; Kaneumi, S.; Terada, S.; Hashimoto, S.; Ikeo, K.; et al. Lack of interleukin-6 in the tumor microenvironment augments type-1 immunity and increases the efficacy of cancer immunotherapy. *Cancer Sci.* **2017**, *108*, 1959–1966. [[CrossRef](#)]
79. Lopez-Otin, C.; Blasco, M.A.; Partridge, L.; Serrano, M.; Kroemer, G. The hallmarks of aging. *Cell* **2013**, *153*, 1194–1217. [[CrossRef](#)] [[PubMed](#)]
80. Leonardi, G.C.; Accardi, G.; Monastero, R.; Nicoletti, F.; Libra, M. Ageing: From inflammation to cancer. *Immun. Ageing I A* **2018**, *15*, 1. [[CrossRef](#)] [[PubMed](#)]
81. Goldberg, E.L.; Dixit, V.D. Drivers of age-related inflammation and strategies for healthspan extension. *Immunol. Rev.* **2015**, *265*, 63–74. [[CrossRef](#)] [[PubMed](#)]
82. Sanada, F.; Taniyama, Y.; Muratsu, J.; Otsu, R.; Shimizu, H.; Rakugi, H.; Morishita, R. Source of Chronic Inflammation in Aging. *Front. Cardiovasc. Med.* **2018**, *5*, 12. [[CrossRef](#)] [[PubMed](#)]
83. Rea, I.M.; Gibson, D.S.; McGilligan, V.; McNerlan, S.E.; Alexander, H.D.; Ross, O.A. Age and Age-Related Diseases: Role of Inflammation Triggers and Cytokines. *Front. Immunol.* **2018**, *9*, 586. [[CrossRef](#)]
84. Chung, H.Y.; Kim, D.H.; Lee, E.K.; Chung, K.W.; Chung, S.; Lee, B.; Seo, A.Y.; Chung, J.H.; Jung, Y.S.; Im, E.; et al. Redefining Chronic Inflammation in Aging and Age-Related Diseases: Proposal of the Senoinflammation Concept. *Aging Dis.* **2019**, *10*, 367–382. [[CrossRef](#)]
85. Li, P.H.; Zhang, R.; Cheng, L.Q.; Liu, J.J.; Chen, H.Z. Metabolic regulation of immune cells in proinflammatory microenvironments and diseases during ageing. *Ageing Res. Rev.* **2020**, 101165. [[CrossRef](#)]
86. Xia, S.; Zhang, X.; Zheng, S.; Khanabdali, R.; Kalionis, B.; Wu, J.; Wan, W.; Tai, X. An Update on Inflamm-Aging: Mechanisms, Prevention, and Treatment. *J. Immunol. Res.* **2016**, *2016*, 8426874. [[CrossRef](#)]
87. Franceschi, C.; Bonafe, M.; Valensin, S.; Olivieri, F.; De Luca, M.; Ottaviani, E.; De Benedictis, G. Inflamm-aging. An evolutionary perspective on immunosenescence. *Ann. N. Y. Acad. Sci.* **2000**, *908*, 244–254. [[CrossRef](#)] [[PubMed](#)]
88. Franceschi, C.; Campisi, J. Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases. *J. Gerontol. A Biol. Sci. Med. Sci.* **2014**, *69* (Suppl. 1), S4–S9. [[CrossRef](#)]
89. Vasto, S.; Candore, G.; Balistreri, C.R.; Caruso, M.; Colonna-Romano, G.; Grimaldi, M.P.; Listi, F.; Nuzzo, D.; Lio, D.; Caruso, C. Inflammatory networks in ageing, age-related diseases and longevity. *Mech. Ageing Dev.* **2007**, *128*, 83–91. [[CrossRef](#)]
90. Chung, H.Y.; Cesari, M.; Anton, S.; Marzetti, E.; Giovannini, S.; Seo, A.Y.; Carter, C.; Yu, B.P.; Leeuwenburgh, C. Molecular inflammation: Underpinnings of aging and age-related diseases. *Ageing Res. Rev.* **2009**, *8*, 18–30. [[CrossRef](#)]
91. Chung, H.Y.; Lee, E.K.; Choi, Y.J.; Kim, J.M.; Kim, D.H.; Zou, Y.; Kim, C.H.; Lee, J.; Kim, H.S.; Kim, N.D.; et al. Molecular inflammation as an underlying mechanism of the aging process and age-related diseases. *J. Dent. Res.* **2011**, *90*, 830–840. [[CrossRef](#)]
92. Olivieri, F.; Rippo, M.R.; Monsurrò, V.; Salvioli, S.; Capri, M.; Procopio, A.D.; Franceschi, C. MicroRNAs linking inflamm-aging, cellular senescence and cancer. *Ageing Res. Rev.* **2013**, *12*, 1056–1068. [[CrossRef](#)] [[PubMed](#)]
93. Tollefsbol, T.O. Dietary epigenetics in cancer and aging. *Cancer Treat. Res.* **2014**, *159*, 257–267. [[CrossRef](#)]
94. Pal, S.; Tyler, J.K. Epigenetics and aging. *Sci. Adv.* **2016**, *2*, e1600584. [[CrossRef](#)]
95. Sapienza, C.; Issa, J.P. Diet, Nutrition, and Cancer Epigenetics. *Annu. Rev. Nutr.* **2016**, *36*, 665–681. [[CrossRef](#)] [[PubMed](#)]
96. White, M.C.; Holman, D.M.; Boehm, J.E.; Peipins, L.A.; Grossman, M.; Henley, S.J. Age and cancer risk: A potentially modifiable relationship. *Am. J. Prev. Med.* **2014**, *46*, S7–S15. [[CrossRef](#)]
97. Lok, A.S.; Seeff, L.B.; Morgan, T.R.; di Bisceglie, A.M.; Sterling, R.K.; Curto, T.M.; Everson, G.T.; Lindsay, K.L.; Lee, W.M.; Bonkovsky, H.L.; et al. Incidence of hepatocellular carcinoma and associated risk factors in hepatitis C-related advanced liver disease. *Gastroenterology* **2009**, *136*, 138–148. [[CrossRef](#)]
98. Tateishi, R.; Uchino, K.; Fujiwara, N.; Takehara, T.; Okanoue, T.; Seike, M.; Yoshiji, H.; Yatsushashi, H.; Shimizu, M.; Torimura, T.; et al. A nationwide survey on non-B, non-C hepatocellular carcinoma in Japan: 2011–2015 update. *J. Gastroenterol.* **2019**, *54*, 367–376. [[CrossRef](#)] [[PubMed](#)]
99. Ben-Porath, I.; Weinberg, R.A. The signals and pathways activating cellular senescence. *Int. J. Biochem. Cell Biol.* **2005**, *37*, 961–976. [[CrossRef](#)] [[PubMed](#)]
100. Kim, I.H.; Kisseleva, T.; Brenner, D.A. Aging and liver disease. *Curr. Opin. Gastroenterol.* **2015**, *31*, 184–191. [[CrossRef](#)]
101. Sastre, J.; Pallardo, F.V.; Pla, R.; Pellin, A.; Juan, G.; O'Connor, J.E.; Estrela, J.M.; Miquel, J.; Vina, J. Aging of the liver: Age-associated mitochondrial damage in intact hepatocytes. *Hepatology* **1996**, *24*, 1199–1205. [[CrossRef](#)] [[PubMed](#)]
102. Poulouse, N.; Raju, R. Aging and injury: Alterations in cellular energetics and organ function. *Aging Dis.* **2014**, *5*, 101–108. [[CrossRef](#)] [[PubMed](#)]
103. Campisi, J. Replicative senescence: An old lives' tale? *Cell* **1996**, *84*, 497–500. [[CrossRef](#)]
104. Maeso-Diaz, R.; Ortega-Ribera, M.; Fernandez-Iglesias, A.; Hide, D.; Munoz, L.; Hessheimer, A.J.; Vila, S.; Frances, R.; Fondevila, C.; Albillos, A.; et al. Effects of aging on liver microcirculatory function and sinusoidal phenotype. *Aging Cell* **2018**, *17*, e12829. [[CrossRef](#)]

105. Kingham, T.P.; Chaudhry, U.I.; Plitas, G.; Katz, S.C.; Raab, J.; DeMatteo, R.P. Murine liver plasmacytoid dendritic cells become potent immunostimulatory cells after Flt-3 ligand expansion. *Hepatology* **2007**, *45*, 445–454. [[CrossRef](#)]
106. Frasca, D.; Blomberg, B.B. Inflammaging decreases adaptive and innate immune responses in mice and humans. *Biogerontology* **2016**, *17*, 7–19. [[CrossRef](#)] [[PubMed](#)]
107. Sipos, F.; Leiszter, K.; Tulassay, Z. Effect of ageing on colonic mucosal regeneration. *World J. Gastroenterol.* **2011**, *17*, 2981–2986. [[CrossRef](#)] [[PubMed](#)]
108. Kakiuchi, N.; Yoshida, K.; Uchino, M.; Kihara, T.; Akaki, K.; Inoue, Y.; Kawada, K.; Nagayama, S.; Yokoyama, A.; Yamamoto, S.; et al. Frequent mutations that converge on the NFKBIZ pathway in ulcerative colitis. *Nature* **2020**, *577*, 260–265. [[CrossRef](#)] [[PubMed](#)]
109. Nanki, K.; Fujii, M.; Shimokawa, M.; Matano, M.; Nishikori, S.; Date, S.; Takano, A.; Toshimitsu, K.; Ohta, Y.; Takahashi, S.; et al. Somatic inflammatory gene mutations in human ulcerative colitis epithelium. *Nature* **2020**, *577*, 254–259. [[CrossRef](#)]
110. Kawazoe, T.; Saeki, H.; Oki, E.; Oda, Y.; Meahara, Y.; Mori, M.; Taniguchi, K. Autocrine leukemia inhibitory factor promotes esophageal squamous cell carcinoma progression via Src family kinase-dependent Yes-associated protein activation. *Mol. Cancer Res. MCR* **2020**. [[CrossRef](#)]
111. Yokoyama, A.; Kakiuchi, N.; Yoshizato, T.; Nannya, Y.; Suzuki, H.; Takeuchi, Y.; Shiozawa, Y.; Sato, Y.; Aoki, K.; Kim, S.K.; et al. Age-related remodelling of oesophageal epithelia by mutated cancer drivers. *Nature* **2019**, *565*, 312–317. [[CrossRef](#)]
112. Chew, V.; Tow, C.; Teo, M.; Wong, H.L.; Chan, J.; Gehring, A.; Loh, M.; Bolze, A.; Quek, R.; Lee, V.K.; et al. Inflammatory tumour microenvironment is associated with superior survival in hepatocellular carcinoma patients. *J. Hepatol.* **2010**, *52*, 370–379. [[CrossRef](#)]
113. Sia, D.; Jiao, Y.; Martinez-Quetglas, I.; Kuchuk, O.; Villacorta-Martin, C.; Castro de Moura, M.; Putra, J.; Camprecios, G.; Bassaganyas, L.; Akers, N.; et al. Identification of an Immune-specific Class of Hepatocellular Carcinoma, Based on Molecular Features. *Gastroenterology* **2017**, *153*, 812–826. [[CrossRef](#)] [[PubMed](#)]
114. Kurebayashi, Y.; Ojima, H.; Tsujikawa, H.; Kubota, N.; Maehara, J.; Abe, Y.; Kitago, M.; Shinoda, M.; Kitagawa, Y.; Sakamoto, M. Landscape of immune microenvironment in hepatocellular carcinoma and its additional impact on histological and molecular classification. *Hepatology* **2018**, *68*, 1025–1041. [[CrossRef](#)] [[PubMed](#)]
115. Rooney, M.S.; Shukla, S.A.; Wu, C.J.; Getz, G.; Hacohen, N. Molecular and genetic properties of tumors associated with local immune cytolytic activity. *Cell* **2015**, *160*, 48–61. [[CrossRef](#)]
116. Fujita, M.; Yamaguchi, R.; Hasegawa, T.; Shimada, S.; Arihiro, K.; Hayashi, S.; Maejima, K.; Nakano, K.; Fujimoto, A.; Ono, A.; et al. Classification of primary liver cancer with immunosuppression mechanisms and correlation with genomic alterations. *EBioMedicine* **2020**, *53*, 102659. [[CrossRef](#)] [[PubMed](#)]
117. Inarrairaegui, M.; Melero, I.; Sangro, B. Immunotherapy of Hepatocellular Carcinoma: Facts and Hopes. *Clin. Cancer Res.* **2018**, *24*, 1518–1524. [[CrossRef](#)]
118. Sangro, B.; Gomez-Martin, C.; de la Mata, M.; Inarrairaegui, M.; Garralda, E.; Barrera, P.; Riezu-Boj, J.I.; Larrea, E.; Alfaro, C.; Sarobe, P.; et al. A clinical trial of CTLA-4 blockade with tremelimumab in patients with hepatocellular carcinoma and chronic hepatitis C. *J. Hepatol.* **2013**, *59*, 81–88. [[CrossRef](#)]
119. Duffy, A.G.; Ulahannan, S.V.; Makorova-Rusher, O.; Rahma, O.; Wedemeyer, H.; Pratt, D.; Davis, J.L.; Hughes, M.S.; Heller, T.; ElGindi, M.; et al. Tremelimumab in combination with ablation in patients with advanced hepatocellular carcinoma. *J. Hepatol.* **2017**, *66*, 545–551. [[CrossRef](#)]
120. El-Khoueiry, A.B.; Sangro, B.; Yau, T.; Crocenzi, T.S.; Kudo, M.; Hsu, C.; Kim, T.Y.; Choo, S.P.; Trojan, J.; Welling, T.H.R.; et al. Nivolumab in patients with advanced hepatocellular carcinoma (CheckMate 040): An open-label, non-comparative, phase 1/2 dose escalation and expansion trial. *Lancet* **2017**, *389*, 2492–2502. [[CrossRef](#)]
121. Lizardo, D.Y.; Kuang, C.; Hao, S.; Yu, J.; Huang, Y.; Zhang, L. Immunotherapy efficacy on mismatch repair-deficient colorectal cancer: From bench to bedside. *Biochim. Biophys. Acta Rev. Cancer* **2020**, *1874*, 188447. [[CrossRef](#)] [[PubMed](#)]
122. Huyghe, N.; Baldin, P.; Van den Eynde, M. Immunotherapy with immune checkpoint inhibitors in colorectal cancer: What is the future beyond deficient mismatch-repair tumours? *Gastroenterol. Rep.* **2020**, *8*, 11–24. [[CrossRef](#)] [[PubMed](#)]
123. Kimura, T.; Kato, Y.; Ozawa, Y.; Kodama, K.; Ito, J.; Ichikawa, K.; Yamada, K.; Hori, Y.; Tabata, K.; Takase, K.; et al. Immunomodulatory activity of lenvatinib contributes to antitumor activity in the Hepa1-6 hepatocellular carcinoma model. *Cancer Sci.* **2018**, *109*, 3993–4002. [[CrossRef](#)] [[PubMed](#)]
124. Finn, R.S.; Qin, S.; Ikeda, M.; Galle, P.R.; Ducreux, M.; Kim, T.Y.; Kudo, M.; Breder, V.; Merle, P.; Kaseb, A.O.; et al. Atezolizumab plus Bevacizumab in Unresectable Hepatocellular Carcinoma. *N. Engl. J. Med.* **2020**, *382*, 1894–1905. [[CrossRef](#)] [[PubMed](#)]
125. Wang, D.; DuBois, R.N. The role of anti-inflammatory drugs in colorectal cancer. *Annu. Rev. Med.* **2013**, *64*, 131–144. [[CrossRef](#)]