Research Article

Effects of Antioxidant Supplementation on Metabolic Disorders in Obese Patients from Randomized Clinical Controls: A Meta-Analysis and Systematic Review

Jinyuan Wang,^{1,2} Biyun Liao,³ Changsheng Wang,² Ou Zhong,² Xiaocan Lei⁽¹⁾,^{1,2} and Yuli Yang¹

¹Hengyang Maternal and Childe Health Hospital, Hengyang, Hunan 421001, China

²*Clinical Antatomy & Reproductive Medicine Application Institute, Hengyang Medical School, University of South China, Hengyan, Hunan 421001, China*

³Reproductive Medicine Center, The Affiliated Hospital of Youjiang Medical University for Nationalities, Baise 533000, China

Correspondence should be addressed to Xiaocan Lei; 2019000013@usc.edu.cn and Yuli Yang; 4051827128@qq.com

Jinyuan Wang, Biyun Liao, and Changsheng Wang contributed equally to this work.

Received 9 February 2022; Revised 27 July 2022; Accepted 28 July 2022; Published 1 September 2022

Academic Editor: Sonia Medina

Copyright © 2022 Jinyuan Wang et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Purpose. This systematic review and meta-analysis aim at elucidating the heterogeneity in beneficial effects of antioxidant supplementation in obese adults by exploring the differential effects of antioxidant supplementation on basic indicators of obesity, lipid metabolism, systemic antioxidant capacity, inflammatory biomarkers, and liver function. Methods. The inclusion criteria specified randomized controlled trials with antioxidant intervention for adults (mean body mass index (BMI) > 30), from inception to Aug. 8, 2021, in the PubMed, Embase, The Cochrane Library, Web of Science, and Scopus databases. Meta-analysis and publication bias were performed using RevMan 5.4 software. Stata16 software was used to detect publication bias with Egger's and Begg's methods being mainly used. The data of basic indicators of obesity, lipid metabolism index, oxidative stress index, inflammatory biomarkers, and liver function index were collected to analyze the beneficial effects of antioxidant supplementation in obese patients. Results. A total of 30 studies were included in this study with a sample of 845 obese patients from the antioxidant supplementation group and 766 obese patients from the placebo control group. The metaanalysis showed that obese patients with antioxidant supplementation had lower BMI (mean difference (MD): -0.44[95% confidence interval (CI): -0.84, -0.04], p = 0.03), waist circumference (MD : -0.78 [95% CI : -1.45, -0.11], p = 0.02),fasting blood glucose (FBG) level (standardized mean difference (SMD): -4.92 [95%CI: -6.87, -2.98], p < 0.001) and homeostasis model assessment of insulin resistance (MD : -0.45 [95%CI : -0.61,-0.3], p < 0.001) when compared to the placebo group. Obese patients on antioxidant supplementation had lower levels of total cholesterol (SMD : -0.43 [95%CI : -0.84, -0.02], p = 0.04), triglycerides (SMD : -0.17 [95%CI : -0.31, -0.04], p = 0.01), low-density lipoprotein (SMD : -0.15 [95%CI : -0.29, -0.01], p = 0.03), malondialdehyde (SMD : -1.67 [95%CI : -2.69, -0.65], p = 0.001), and tumor necrosis factor-alpha (SMD : -2.69, -0.65], p = 0.001), and tumor necrosis factor-alpha (SMD : -2.69, -0.65], p = 0.001), and tumor necrosis factor-alpha (SMD : -2.69, -0.65], p = 0.001), and tumor necrosis factor-alpha (SMD : -2.69, -0.65], p = 0.001), and tumor necrosis factor-alpha (SMD : -2.69, -0.65], p = 0.001), and tumor necrosis factor-alpha (SMD : -2.69, -0.65], p = 0.001), and tumor necrosis factor-alpha (SMD : -2.69, -0.65], p = 0.001), p = 0.001, p-0.29 [95%CI: -0.56, -0.02], p = 0.03), respectively, when compared to the placebo group. In addition, obese patients with antioxidant supplementation had higher levels of high-density lipoprotein (SMD : 0.25 [95%CI : 0.03, 0.46], p = 0.03) and superoxide dismutase (SMD : 1.09 [95% CI : 0.52, 1.65], p < 0.001) when compared to the placebo group. Antioxidant supplementation had no effects on other analyzed parameters including waist-hip ratio, leptin, fat mass, interleukin-6, C-reactive protein, alanine transaminase, and aspartate transaminase in obese patients. Conclusion. The meta-analysis results indicated that antioxidant supplementation exerted potential beneficial effects in obese patients by regulating FBG, oxidative stress, and inflammation, whilst more high-quality studies are required to confirm these effects. The present study may provide important insights for the treatment of clinical obesity and obesity-associated complications.

1. Introduction

With the advance of people's living standard, the global obesity epidemic has become a focus and affects health of >2 billion people [1]. Nowadays, the prevalence of obesity has doubled in about 70 countries and continues to rise [2, 3]. The occurrence of obesity impacting the body's metabolic processes, following induce serious diseases, has seriously endangering people's health [3]. Recent studies have reported a high prevalence of overweight and obesity in patients experiencing a severe COVID-19 course, with serious complications requiring hospitalization and admission to intensive care units [4]. Studies have shown that the disorder of glucose metabolism, insulin resistance and elevated blood glucose in obese patients increase the susceptibility to diabetes [5]. Dyslipidemia, increased incidence of oxidative stress and excessive production of adipocyte derivatives have all been proposed to contribute to the cardiovascular diseases [6] and non-alcoholic fatty liver disease [7] in obese patients. There are several mechanisms by which obesity produces oxidative stress. The first of these is the mitochondrial and peroxisomal oxidation of fatty acids, which can produce reactive oxygen species (ROS) in oxidation reactions, while another mechanism is over-consumption of oxygen, which generates free radicals in the mitochondrial respiratory chain that is found coupled with oxidative phosphorylation in mitochondria [8]. Lipid-rich diets are also capable of generating ROS because they can alter oxygen metabolism. Finally, high ROS production leads to various abnormalities in obese patients [8]. Adipose tissue is not only a triglyceride storage organ, but studies have shown the role of white adipose tissue as a producer of certain bioactive substances called adipokines such as interleukin-6 (IL-6), which exerts many effects, ranging from defense to inflammation and tissue damage [9]. Inflammation is a manifestation of increased oxidative, which increases in subjects with obesity, which is related with insulin resistance and endothelial dysfunction. These changes may interact among themselves and amplify, producing, in this manner, the set of metabolic and vascular alterations [10]. Therefore, in order to modify the serious impact of obesity, it is imperative to reverse the effect of obesity on oxidative stress and inflammation.

Antioxidants, such as vitamins A, C, E, selenium, zinc, copper and manganese, is a class of substances that prevent the harmful effects of ROS in daily nutrition and human health, helping capture and neutralize free radicals [11]. Antioxidants are rich in a wide range of sources, for instance green tea, strawberries, eggplant, garlic, ginger and so on, being reported possessing multiple pharmacological functions including improving the lipid metabolic abnormalities [12], insulin sensitivity, total antioxidant status [13] and anti-inflammatory [14] associated with obesity. Studies found that polyphenols found in pigmented rice may play a key role in targeting specific therapeutic pathways in obesity-related oxidative stress and inflammation [15]. Quercetin, curcumin, and resveratrol possess the antioxidant and anti-inflammatory activities, and can exert beneficial effects in obesity [16]. However, the beneficial effects of antioxidants in obese patients are still a matter of debate. A

recent meta-analysis concludes that there is low-quality evidence of a beneficial effect of antioxidants to increase fertility [17]. In addition, clinical trials have reported contradictory results regarding the effect of vitamin E supplementation on weight status. Some studies reported an increase [18], some reported a decrease [19], and some reported no significant changes in body mass index (BMI) and weight following vitamin E supplementation [20]. Therefore, the effects of antioxidants on improving metabolic disorders in obese patients remain to be clarified.

In this study, we sorted out the data of basic indicators of obesity, lipid metabolism index, oxidative stress index, inflammatory biomarkers and liver function index, and aimed to clarify whether antioxidants supplementation could delay the progression of obesity and metabolic disorders in clinical application in the management of obese patients. The present study may provide important insights for the treatment of clinical obesity and obesity-associated complications.

2. Materials and Methods

2.1. Search Strategy. The electronic databases including PubMed, Embase, The Web of Science, The Cochrane library and Scopus databases were searched to collect clinical studies related to the efficacy of antioxidants supplementation in the treatment of obesity from inception to Aug 8, 2021. Articles published in peer-reviewed journals from 2006 to 2020, using a search strategy based on previous systematic reviews. Search is conducted by combining subjects and free words. The search term antioxidant* was added to the concept obesity. Detailed research strategy is shown in Appendix A.

2.2. Eligibility Criteria. Inclusion criteria: (1) Included articles were published in English, peer-reviewed, randomized and cluster randomized controlled trials (RCTs); (2) All patients had obesity (BMI≥30 kg/m2) who did not habitually use antioxidant supplements; (3) Intervention: The treatment group received antioxidant and the control group received placebo. (3) The primary outcomes were anthropometric measurements, and secondary outcome were metabolic index. (4) Antioxidants used in RCT are biological and chemical organic nutrients that include vitamins, minerals, and polyunsaturated fatty acids, for instance, vitamin E, vitamin C, carotenoids, carnitines, coenzyme Q10 (ubiquinol), cysteine, omega-3, omega-6, the micronutrients folate, selenium, zinc, resveratrol and melatonin derived from fruits, vegetables, medicine and compound nutrient and so on [21, 22].

Exclusion criteria: (1) review articles, expert opinions, case-series/reports, basic science and conference abstracts; (2) cell experiments, animal experiments and other literatures which have unavailable data; (3) literatures with obvious statistical errors and poor quality of information; (4) literature were too old to be traced or published in other language unidentified except English. (5) smoking; alcohol consumption; the presence of pregnancy or lactation; recent surgery, patients with coronary heart disease; known cases of

3

TABLE 1: Cohen's kappa statistics between the authors of clinical outcomes.

Oration	Т	he interrater reliability	The intrar	ater reliability
Outcomes	Pooled k	95% confidence interval (CI)	Pooled k	95% CI
BMI (kg/m ²)	0.62	(0.52, 0.70)	0.60	(0.55, 0.69)
WC (cm)	0.62	(0.54, 0.69)	0.59	(0.52, 0.69)
WHR (mmol/L)	0.65	(0.57, 0.73)	0.61	(0.53, 0.72)
Leptin (μ g/L)	0.66	(0.54, 0.72)	0.62	(0.51, 0.68)
FM (kg)	0.55	(0.42, 0.63)	0.58	(0.51, 0.65)
FBG (µg/dL)	0.60	(0.52, 0.67)	0.61	(0.53, 0.72)
HOMAir	0.57	(0.50, 0.66)	0.59	(0.51, 0.69)
TC (μ g/dL)	0.61	(0.54, 0.72)	0.59	(0.50, 0.72)
TG (μ g/dL)	0.59	(0.52, 0.69)	0.60	(0.52, 0.72)
LDL (μ g/dL)	0.63	(0.55, 0.70)	0.60	(0.54, 0.71)
HDL (μ g/dL)	0.65	(0.53, 0.67)	0.61	(0.51, 0.70)
MDA (mmol/L)	0.66	(0.60, 0.74)	0.65	(0.55, 0.76)
SOD (mmol/L)	0.60	(0.54, 0.69)	0.61	(0.52, 0.73)
TNF- α (pg/mL)	0.59	(0.52, 0.67)	0.59	(0.50, 0.72)
IL-6 (pg/mL)	0.60	(0.54, 0.69)	0.62	(0.50, 0.73)
CRP (mg/L)	0.64	(0.60, 0.72)	0.65	(0.53, 0.76)
ALT (U/L)	0.65	(0.60, 0.71)	0.62	(0.52, 0.72)
AST (U/L)	0.61	(0.56, 0.70)	0.60	(0.52, 0.68)

diabetes mellitus; patients with proven malignancy, chronic kidney or hepatic disease, asthma, chronic cough, chronic inflammatory disease and psychological problems; obesity due to endocrine disease (hypothyroidism or uncontrolled thyroid disease); and genetic obesity syndrome. or history of use of any dietary supplements in the 3 months prior to the study.

2.3. Literature Screening and Data Extraction. After literature retrieval, eliminating duplicate literatures, we carried out a relevance check of 10644 articles. By following thee inclusion and exclusion criteria, at least two reviewers independently screened the title, abstract and full text of the articles using a data collection form. After excluding the literatures with obviously irrelevant contents, we further read the full text. According to the inclusion and exclusion criteria, the included literatures were identified and then the data was extracted. The eligibility of the studies was assessed by two independent reviewers and discrepancy was resolved with double checking the data by either: (a) discussion and consensus or (b) by a third independent reviewer. Using Cohen's kappa statistic, the overall agreement rate prior to correcting discrepancies is shown in Table 1.

2.4. Statistical Analysis. Statistical meta-analyses of pooled effect and heterogeneity index (I^2) as well as forest and funnel plots were carried out using the RevMan 5.3 software. For assessing the reliability of the data extraction process, two reviewers independently screen and select the RCT, coding the characteristics of all studies that fulfilled the selection criteria. Continuous data were estimated with weighted mean difference (MD) and confidence intervals (CIs) were

set at 95%, and p < 0.05 was considered statistically significant. For the sake of eliminating the influence of different units, standardized mean difference (SMD) was used for assessment (Table 2). If $I^2 < 50\%$ or p > 0.1, which indicated that little heterogeneity among the included studies, the fixed effect model was used; on the contrary, the significant heterogeneity among the included studies result in the random effects model adopted. Egger's and Begg's tests were mainly performed by Stata16 software in order to detect publication bias, p > 0.05 indicates no significant publication bias (when the p values of Egger's and Begg's tests are contradictory, the Egger's examination results are more convincing).

3. Results

3.1. Study Selection. We identified 10,868 literatures in the initial retrieval, including PubMed (n = 465), Embase (n = 341), The Cochrane library (n = 4471), Web of Science (n = 4675), and Scopus (n = 921). After carefully reviewing the titles and abstracts of these literatures, 224 duplicate articles were excluded. After further screening, 30 studies were included in this meta-analysis, the literature screening process and results are shown in Figure 1. We evaluated RCTs from 30 studies and found a total of 27 trials reporting basic indicators of obesity, 17 trials reporting lipid metabolism, 4 trials reporting systemic antioxidant capacity, 9 trials reporting liver function (Figure 1).

3.2. Study Characteristics. A total of 30 individual RCTs were eventually included in this study, and 93.33% of RCTs ended the interventions by 6 to 15 weeks. The demographics of the

	Heter	rogeneity		0
Outcomes	I ² (%)	<i>p</i> value	Analysis	Summary statistic
BMI (kg/m ²)	0	0.03	Fixed	MD
WC (cm)	13	0.02	Fixed	MD
WHR (mmol/L)	100	0.44	Random	MD
Leptin (µg/L)	69	0.19	Random	SMD
FM (kg)	78	0.51	Random	SMD
FBG (µg/dL)	45	< 0.00001	Fixed	SMD
HOMAir	48	< 0.00001	Fixed	MD
TC (μ g/dL)	82	0.04	Random	SMD
TG (μg/dL)	0	0.01	Fixed	SMD
LDL (μ g/dL)	0	0.03	Fixed	SMD
HDL (µg/dL)	62	0.03	Random	SMD
MDA (mmol/L)	84	0.001	Random	SMD
SOD (mmol/L)	54	0.0002	Random	SMD
TNF-α (pg/mL)	31	0.03	Fixed	SMD
IL-6 (pg/mL)	0	0.05	Fixed	SMD
CRP (mg/L)	0	0.86	Random	SMD
ALT (U/L)	0	0.34	Fixed	MD
AST (U/L)	0	0.55	Random	MD

 TABLE 2: Statistical models of clinical outcomes.

BMI: body mass index; WC: waist circumference; WHR: waist-to-hip ratio; FM: fat mass; FBG: fasting plasma glucose; HOMA-ir: homeostasis model assessment of insulin resistance; TC: total cholesterol; TG: triglycerides; LDL: low-density lipoprotein; HDL: high-density lipoprotein; MDA: malondialdehyde; SOD: superoxide dismutase; TNF- α : tumor necrosis factor- α ; IL-6: interleukin-6; CRP: C-reactive protein; ALT: alanine transaminase; AST: aspartate transaminase.



FIGURE 1: Flowchart of study selection.

patients were summarized in Table 3 (placebo group: 845 patients; antioxidants supplementation group: 766 patients). The quality of the included articles was evaluated using the bias risk assessment tool for RCT in Cochrane Systematic

Review Manual 5.1.0, and all the included studies reached a medium to high level. However, some studies did not describe the method of random assignment of included cases, and did not describe whether the assignment was

studies.	
included	
cs of	
Characteristi	
3: 0	
TABLE	

Study	Country	Sample size (antioxidant/control)	Population characteristic (antioxidant/control)	Intervention (antioxidant/control)	Subgroup on antioxidants	Control group	Duration of intervention
Caballero [1]	UK	36/78	Age(years):42.9 \pm 7.1/42.8 \pm 7.2 BMI (kg/m ²): 30.0 \pm 4.4/30.3 \pm 3.5	The Ascophyllum (poly)phenol- rich blend 400 mg/day	Water-soluble antioxidants	Placebo	8 weeks
Collaborators et al. [2]	Poland	30/30	Age (years): 43.8 ± 8.2/41.0 ± 8.8 BMI (kg/m ²): 39.2 ± 6.0/37.5 ± 4.8	Average arginine 43.3 mg/kg/day bodyweight in women and 48.6 mg/kg/day in men	Water-soluble antioxidants	Placebo	3 months
Bazrafshani et al. [3]	Iran	16/19	Age (years): $38 \pm 10.9/42 \pm 14.4$ BMI (kg/m ²): $37.1 \pm 8.941.8 \pm 8.5$	L-selenomethionine (S) 240 µg/day	Water-soluble antioxidants	Placebo	12 weeks
Valerio et al. [4]	Iran	27/28	Age (years): $44.07 \pm 7.82/42.39 \pm 7.21$ BMI (kg/m ²): $33.16 \pm 1.72/35.03 \pm 3.48$	L-arginine 6 g/d	Water-soluble antioxidants	Placebo	8 weeks
Malone and Hansen [5]	Brazil	24/22	BMI (kg/m ²): $32.5 \pm 4.3/33.3 \pm 4.6$	Roasted baru almonds 20 g/day	Mixed	Placebo	8 weeks
Nikolopoulou and Kadoglou [6]	Brazil	24/22	BMI (kg/m ²): $32.54 \pm 4.35/33.34 \pm 4.69$	Roasted baru almonds 20 g/day	Mixed	Placebo	8 weeks
Milic et al. [7]	USA	19/19	BMI (kg/m ²): 92.22 \pm 3.52/92.75 \pm 4.13	Dried apple 240 kcal/day	Mixed	Placebo	8 weeks
Fernández-Sánchez et al. [8]	Iran	25/25	Age (years): 39.44 \pm 10.54/40.68 \pm 9.87 BMI (kg/m ²): 31.23 \pm 3.03/31.47 \pm 3.85	Zinc gluconate (contained 30 mg/d elemental zinc)	Water-soluble antioxidants	Placebo	12 weeks
Fonseca-Alaniz et al. [9]	Iran	23/22	Age (years): $40.6 \pm 5.6/38.8 \pm 6.5$ BMI (kg/m ²): $33.8 \pm 3.7/34.1 \pm 4.5$	ALA 1200 mg/day plus vitamin E 400 mg/day	Fat-soluble antioxidants	Placebo	12 weeks
Dludla et al. [10]	Iran	30/30	Age (years): 39.16 \pm 9.59/36.36 \pm 9.9 BMI (kg/m ²): 37.14 \pm 5.40/36.29 \pm 4.66	Majoun 10 g/day	Water-soluble antioxidants	Placebo	12 weeks
Bjorklund and Chirumbolo [11]	Iran	35/35	Age (years): $37.23 \pm 9.34/37.00 \pm 7.90$ BMI (kg/m ²) $29.24 \pm 3.36/30.39 \pm 4.69$	Caraway seed extract 30 ml/day	Water-soluble antioxidants	Placebo	12 weeks
Ohishi et al. [12]	Iran	18/22	Age (years): $35.63 \pm 3.2/32.95 \pm 1.7$ BMI (kg/m ²): $33.17 \pm 6.34/32.64 \pm 2.37$	Zinc 30 mg/day	Water-soluble antioxidants	Placebo	15 weeks
Szulinska et al. [13]	Germany	23/23	BMI (kg/m ²): 32.8 \pm 0.8/32.8 \pm 0.8	Epicatechin 25 mg/day	Water-soluble antioxidants	Placebo	2 weeks
Hosseinpour-Arjmand [14]	China	30/29	Age (years): $41.2 \pm 6.8/42.8 \pm 6.9$ BMI (kg/m ²): $31.1 \pm 2.7/30.8 \pm 2.5$	29 multivitamins and minerals one tablet/day	Mixed	Placebo	26 weeks
Callcott et al. [15]	Brazil	30/39	Age (years): $42.3 \pm 9.1/40.4 \pm 10.2$ BMI (kg/m ²): $34.2 \pm 5.1/37.1 \pm 7.2$	HD+frozen açaí (Euterpe oleracea Mart) 200 g/day	Water-soluble antioxidants	HD+placebo	60 days
Zhao et al. [16]	Mexico	15/24	Age (years): $33.7 \pm 11.9/38.8 \pm 9.59$ BMI (kg/m ²): $35.6 \pm 2.71/34.7 \pm 2.89$	Resveratrol 100 mg/day	Fat-soluble antioxidants	Placebo	24 weeks
Showell et al. [17]	Brazil	28/28	Age (years): 35.5 ± 6.5/33.9 ± 5.4 BMI (kg/m ²): 35.8 ± 2.2/36.5 ± 2.5	Zinc aminochelate 30 mg/day	Water-soluble antioxidants	Placebo	30 days

Study	Country	Sample size (antioxidant/control)	Population characteristic (antioxidant/control)	Intervention (antioxidant/control)	Subgroup on antioxidants	Control group	Duration of intervention
Emami et al. [18]	Iran	29/29	Age (years): $36.0 \pm 11.9/33.6 \pm 4.8$ BMI (kg/m ²): $33.6 \pm 4.8/32.7 \pm 3.7$	Dried licorice extract 0.5 g/day	Water-soluble antioxidants	Placebo	8 weeks
Ekhlasi et al. [19]	Netherlands	11/14	Age (years): $36 \pm 3/40 \pm 3$ BMI (kg/m ²): $30.5 \pm 0.7/29.7 \pm 1.1$	Polyphenols epigallocatechin- gallate 282 mg/d and resveratrol 80 mg/d	Water-soluble antioxidants	Placebo	12 weeks
Shadman et al. [20]	Poland	28/28	Age (years): $49.2 \pm 8.8/51.5 \pm 7.4$ BMI (kg/m ²): $32.5 \pm 3.3/33.9 \pm 2.3$	Green tea extract 1 capsule/day	Water-soluble Antioxidants	Placebo	8 weeks
Showell et al. [21]	Iran	30/30	Age (years): $31 \pm 8/33 \pm 8$ BMI (kg/m ²): $34.7 \pm 4.3/33.3 \pm 5.7$	Zinc 30 mg/kg	Water-soluble antioxidants	Placebo	4 weeks
Smits et al. [22]	Poland	24/35	Age (years): 47.31 \pm 12.04/45.92 \pm 9.33 BMI (kg/m ²): 34.44 \pm 2.69/34.77 \pm 3.00	DHA and EPA given in 3 capsules/day	Fat-soluble antioxidants	Placebo	12 weeks
Piche et al. [23]	Brazil	13/14	Age (years): 45.76 ± 2.58/45.07 ± 3.42 BMI (kg/m ²): 34.63 ± 1.20/33.82 ± 0.71	Juçara berry (Euterpe edulis Mart.) freeze-dried pulp 5 g/day	Mixed	Placebo	6 weeks
Leisegang et al. [24]	USA	15/17	Age (years): $54 \pm 3/49 \pm 3$ BMI (kg/m ²): $36.8 \pm 0.9/38.0 \pm 0.9$	Blueberry powder 45 g/day	Mixed	Placebo	8 weeks
Park et al. [25]	Poland	44/44	Age (years): $43.1 \pm 8.6/41.5 \pm 9.1$ BMI (kg/m ²): $36.8 \pm 6.3/36.1 \pm 4.9$	The average arginine 43.3 mg/kg/day	Water-soluble antioxidants	Placebo	6 months
Dostal et al. [26]	Poland	15/15	Age (years): $37.7 \pm 3.40/36.3 \pm 4.18$ BMI (kg/m ²): $37.8 \pm 1.51/38.2 \pm 1.94$	Melatonin 10 mg/day	Fat-soluble antioxidants	Placebo	30 days
Zhang et al. [27]	Poland	25/25	Age (years): $49.3 \pm 8.7/50.2 \pm 7.2$ BMI (kg/m ²): $33.5 \pm 6.7/33.3 \pm 6.2$	Spirulina 0.5 g/day	Water-soluble antioxidants	Placebo	12 weeks
Farr et al. [28]	Poland	46/46	Age (years): $53.0 \pm 5.8/53.6 \pm 5.5$ BMI (kg/m ²): $30.3 (26.7 - 38.3)/33.0 (29.2 - 36.1)$	Extract of garlic (2% allicin) 400 mg/d	Mixed	Placebo	12 weeks
Yadav et al. [29]	Iran	25/25	Age (years): 32.2 \pm 6.9/35.1 \pm 7.2 BMI (kg/m ²): 32.3 \pm 4.2/32.4 \pm 5.9	Green tea 1 g/day, capsaicin 100 mg/day, and ginger 200 mg/day	Water-soluble antioxidant	Placebo	8 weeks
Balsan et al. [30]	Australia	28/28	Age (years): 61.4 ± 1.5/57.9 ± 1.4 BMI (kg/m ²): 34.6 ± 0.7/37.0 ± 1.3	A fruit and vegetable concentrate supplement	Mixed	Placebo	8 weeks

TABLE 3: Continued.

hidden (Table 4). Thus, we assessed the study quality using the bias risk plots, and evaluation results of bias risk are shown in Supplemental Figure S1.

3.3. Effects of Antioxidant Supplementation on Basic Indicators of Obesity in Obese Patients. The forest plots from stratified meta-analyses in Figure 2 depicted a total of 27 study outcomes, which reported the effects of antioxidants supplementation on basic indicators of obesity. The results showed that obese patients with antioxidants supplementation had lower BMI (p = 0.03; Figure 2(a)), waist circumference (WC; p = 0.02; Figure 2(b)), fasting blood glucose (FBG) level (p < 0.001; Figure 2(f)) and homeostatic model assessment for insulin resistance (HOMA-IR; p < 0.001; Figure 2(g), respectively, when compared to the placebo group. While there was no statistical significance in waisthip ratio (WHR; p = 0.44; Figure 2(c)), leptin level (p =0.19; Figure 2(d)) and fat mass (FM; p = 0.51; Figure 2(e)) between placebo control and antioxidants treatment group. Begg's test and Egger's test showed no significant publication bias for BMI ($p_{(B)} = 1.4943$, $p_{(E)} = 0.8978$), WC ($p_{(B)} =$ 1.7225, $p_{(E)} = 0.7412$), FBG ($p_{(B)} = 1.5476$, $p_{(E)} = 0.9121$) and HOMA-IR ($p_{(B)} = 0.9015$, $p_{(E)} = 0.0629$). The corresponding funnel plots for evaluating the publication bias are shown in Supplemental Figure S2. The subgroup analysis showed that obese patients with water-soluble antioxidants supplementation had lower WC (p < 0.01), FBG (p < 0.001) and HOMA-IR (p < 0.001) when compared to the placebo group (Supplementary Table S1). Obese patients with fat-soluble antioxidants supplementation had higher leptin level when compared to the placebo group (p < 0.05; Supplementary Table S1). However, no significant difference was detected in the basic indicators of obesity between mixed antioxidants treatment group and placebo control group (p > 0.05; Supplementary Table S1).

3.4. Effects of Antioxidant Supplementation on Regulating Lipid Metabolism in Obese Patients. The forest plots from stratified meta-analyses in Figure 3 depicted a total of 17 study outcomes, which reported the effects of antioxidants on regulating lipid metabolism in obese patients. The results showed that obese patients on antioxidants supplementation had lower total cholesterol (TC) level (p = 0.04; Figure 3(a)), triglycerides (TG) level (p = 0.01; Figure 3(b)) and lowdensity lipoprotein (LDL) level (p = 0.03; Figure 3(c)), respectively, when compared to the placebo group. The high-density lipoprotein (HDL) level in antioxidants supplementation group was higher than placebo group (p = 0.03; Figure 3(d)). Begg's test and Egger's test showed no significant publication bias for TC ($p_{(B)} = 1.2455$, $p_{(E)} = 0.7151$), TG $(p_{(B)} = 1.6996, p_{(E)} = 0.3787),$ LDL $(p_{(B)} = 1.8747,$ $p_{(E)} = 0.3505$), and HDL ($p_{(B)} = 0.0377$, $p_{(E)} = 0.1254$). The corresponding funnel plots for evaluating the publication bias are shown in Supplemental Figure S3. In the subgroup analysis, no significant difference was detected in the TC, TG, LDL and HDC between water-soluble antioxidants treatment group and placebo control group (p > 0.05; Supplementary Table S2), and similar results were also detected in the fat-soluble antioxidants subgroup (p > 0.05; Supplementary Table S2). Furthermore, obese patients with mixed antioxidants supplementation had higher HDL when compared to the placebo group (p < 0.001; Supplementary Table S2).

3.5. Effects of Antioxidant Supplementation on Systemic Antioxidant Capacity in Obese Patients. The forest plots from stratified meta-analyses in Figure 4 depicted a total of 4 study outcomes, which reported the effects of antioxidants on regulating systemic antioxidant capacity in obese patients. The results showed that obese patients with antioxidants supplementation had lower malondialdehyde (MDA) level (p = 0.001; Figure 4(a)) when compared to placebo group. The superoxide dismutase (SOD) level in antioxidants supplementation group was higher than in the placebo group (p < 0.001; Figure 4(b)). Begg's test and Egger's test showed no significant publication bias for MDA ($p_{(B)} = 1.9633$, $p_{(E)} = 0.2680$), and SOD ($p_{(B)} = 1.7037$, $p_{(E)} = 0.5561$). The corresponding funnel plots for evaluating the publication bias are shown in Supplemental Figure S4. The subgroup analysis showed that obese patients with water-soluble antioxidants supplementation had lower MDA level (p < 0.01) and higher SOD level (p < 0.001) when compared to the placebo group (Supplementary Table S3). Obese patients with fat-soluble antioxidants supplementation had lower MDA level when compared to the placebo group (p < 0.001; Supplementary Table S3). In addition, obese patients with mixed antioxidants supplementation had higher SOD level when compared to the placebo group (p < 0.05; Supplementary Table S3).

3.6. Effects of Antioxidant Supplementation on Levels of Inflammatory Markers in Obese Patients. The forest plots from stratified meta-analyses in Figure 4 depicted a total of 4 study outcomes, which reported the effects of antioxidants on systemic antioxidant capacity in obese patients. The results showed that obese patients with antioxidants supplementation had lower tumor necrosis factor-alpha (TNF- α) level (p = 0.03; Figure 5(a)) when compared to the placebo group. However, antioxidants supplementation in the obese patients did not affect the levels of IL-6 (p = 0.05; Figure 5(b)) and C-reactive protein (CRP; p = 0.86; Figure 5(c)) when compared to the placebo group. Begg's test and Egger's test showed no significant publication bias for TNF- α ($p_{(B)} = 1.5558$), $p_{(E)} = 0.1752$). The corresponding funnel plots for evaluating the publication bias are shown in Supplemental Figure S5. The subgroup analysis showed that obese patients with water-soluble antioxidants supplementation had lower TNF- α level (p < 0.05) when compared to the placebo group (Supplementary Table S4). Obese patients with fat-soluble antioxidants supplementation had lower IL-6 level when compared to the placebo group (p < 0.05; Supplementary Table S4). However, no significant difference was detected in inflammatory markers between mixed antioxidants treatment group and placebo control group (p > 0.05; Supplementary Table S4).

Study	Randomization		Blinding of participants and (performance bia	d personnel s)	Allocation concealment (selection bias)	Integrality of date outcome (attrition bias)	Selective reporting (reporting bias)	Other bias
Caballero [1]	Computer simulation R	andom	Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Collaborators et al. [2]	Unclear		Unclear		Unclear	Unclear	Unclear	Unclear
Bazrafshani et al. [3]	Unclear		Single-blind simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Valerio et al. [4]	Unclear		Unclear		Unclear	Unclear	Unclear	Unclear
Malone and Hansen [5]	Computer simulation Ra	andom	Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Nikolopoulou and Kadoglou [6]	Computer simulation Ra	andom	Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Milic et al. [7]	Randomization list		Unclear		Unclear	Unclear	Unclear	Unclear
Fernández-Sánchez et al. [8]	Block size of 4 subjects' schedule		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Fonseca-Alaniz et al. [9]	Computer simulation R	andom	Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Dludla et al. [10]	Unclear		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Bjorklund and Chirumbolo [11]	Computer simulation Ra	andom	Triple-blind simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Ohishi et al. [12]	Blocked size of 4 number tables		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Szulinska et al. [13]	Blocked size of 4 number tables		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Hosseinpour-Arjmand [14]	Computer simulation Ra	andom	Unclear		Unclear	Unclear	Unclear	Unclear
Callcott et al. [15]	Simulation blocked R _i size of 4	andom	Double-blinded	Randomized	Unclear	Unclear	Unclear	Unclear
Zhao et al. [16]	Random number table		Simulation		Unclear	Unclear	Unclear	Unclear
Showell et al. [17]	Unclear		Unclear		Unclear	Unclear	Unclear	Unclear
Emami et al. [18]	Random number table with a permuted block size of two		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Ekhlasi et al. [19]	Unclear		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Shadman et al. [20]	Unclear		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Showell et al. [21]	Unclear		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Smits et al. [22]	Computer simulation		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Piche et al. [23]	Unclear R:	andom	Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Leisegang et al. [24]	Unclear		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Park et al. [25]	Unclear		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Dostal et al. [26]	Unclear		Unclear		Unclear	Unclear	Unclear	Unclear
Zhang et al. [27]	Unclear		Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear

TABLE 4: Risk of bias analysis of included trials.

		TABLE 4: Contir	nued.				
Study	Randomization	Blinding of participants and (performance bias)	l personnel)	Allocation concealment (selection bias)	Integrality of date outcome (attrition bias)	Selective reporting (reporting bias)	Other bias
Farr et al. [28]	Randomization list	Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Yadav et al. [29]	Computer simulation Random	Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear
Balsan et al. [30]	Unclear	Double-blinded simulation	Randomized	Unclear	Unclear	Unclear	Unclear

Studu on sub-moun		Antioxidan	s		Control		Mainht	Mean difference	Mean difference
	Mean	SD	Total	Mean	SD	Total	weight	IV, fixed, 95% Cl	IV, fixed, 95% Cl
Bogdanski, P 2012	-0.3	5.9025418	30	-0.2	4.850773	30	2.1%	-0.10 [-2.83, 2.63]	
Cavedon, E., J 2020	-0.2	9.0537285	16	-1.2	8.129576	19	0.5%	1.00 [-4.75, 6.75]	· · · · · · · · · · · · · · · · · · ·
Dashtabi, A 2016	-2.92	2.0648729	27	-0.13	3.455271	28	7.2%	-2.79 [-4.29, -1.29]	
de Souza, R. G. M 2018	-0.8	4.2260028	24	-0.72	28.24852	22	0.1%	-0.08 [-12.00, 11.84]	· · · · · · · · · · · · · · · · · · ·
Eisner, A 2020	-0.56	3.4710805	19	0	4.13	19	2.7%	-0.56 [-2.99, 1.87]	
Hosseinpour-Arjmand, S 2019	1.3	3.6510273	23	1.2	4.357752	22	2.9%	0.10 [-2.25, 2.45]	
Kazemipoor, M 2013	0	0	0	0	0	0		Not estimable	
Khorsandi, H 2019	1.67	5.8133295	18	0.55	2.340577	22	2.0%	1.12 [-1.74, 3.98]	
Kirch, N 2018	0	0	0	0	0	0		Not estimable	
Li, Y 2010	-1.4	2.3259407	30	-0.1	2.805352	29	9.3%	-1.30 [-2.62, 0.02]	
Luciana Nicolau Aranha 2020	-0.2	5.0029991	30	0.3	7.250517	39	1.9%	-0.50 [-3.40, 2.40]	
Maria Angelica Arzola-Paniagua 2016	-1.4	3.3867388	15	-1	3.082613	24	3.6%	-0.40 [-2.51, 1.71]	
Marreiro, D. D 2006	-0.2	2.3065125	28	-0.1	2.5	28	10.1%	-0.10 [-1.36, 1.16]	
Mohammad Alizadeh 2018	-0.8	4.8	28	-0.4	3.604164	28	3.2%	-0.40 [-2.62, 1.82]	
Pawel Bogdanski 2012	-0.4	3.2511536	28	-0.3	2.351595	28	7.3%	-0.10 [-1.59, 1.39]	
Polus, A 2016	-1.36	2.6509055	24	-1.54	2.956028	35	7.7%	0.18 [-1.26, 1.62]	
Santamarina, A.B 2020	0.13	1.2529964	13	0.24	0.764918	14	25.7%	-0.11 [-0.90, 0.68]	_
Stull, A. J 2010	0	0	0	0	0	0		Not estimable	
Suliburska, J 2014	-0.2	6.2024189	44	-0.3	4.950758	44	2.9%	0.10 [-2.24, 2.44]	
Szewczyk-Golec, K 2017	-2.3	1.5150248	15	-1.9	2.094087	15	9.4%	-0.40 [-1.71, 0.91]	-
Szulinska, M 2017	-1.8	6.2984125	25	0	6.009992	25	1.4%	-1.80 [-5.21, 1.61]	• • • • • • • • • • • • • • • • • • • •
Williams, E. J 2017	0	0	0	0	0	0		Not estimable	
Total (95% Cl)			437			471	100.0%	-0.44 [-0.84, -0.04]	◆
Heterogeneity: $v^2 = 15.43$ df = 17 (P	- 0 56).	$I^2 - 0\%$							r
Test for overall effect: $7 = 2.13$ ($P = 0.0$	- 0.50), 13)	1 = 0 /0							-4 -2 0 2 4
101 over all effect: Z = 2.13 (F = 0.0)	,,,,								Favours [antioxidants] Favours [control]

						(a))						
	А	ntioxidants			Control			Mean difference		М	ean differ	ence	
Study or subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, fixed, 95% Cl		IV	, fixed, 95	% Cl	
Bogdanski, P 2012	-0.9	10.2235	30	-1.1	8.428523	30	2.0%	0.20 [-4.54, 4.94]					
Dashtabi, A 2016	-7.16	5.798733	27	-0.29	10.75827	28	2.2%	-6.87 [-11.42, -2.32]		•	-		
de Souza, R. G. M 2018	-5.78	12.08754	24	-3.33	10.32093	22	1.1%	-2.45 [-8.93, 4.03]		-			
Eisner, A 2020	1.3	16.14645	19	0.87	14.55808	19	0.5%	0.43 [-9.35, 10.21]					
Hosseinpour-Arjmand, S 2019	-2.5	24.61524	23	-2.2	10.52283	22	0.4%	-0.30 [-11.28, 10.68]					
Kazemipoor, M 2013	-6.42	9.516386	31	-0.13	7.630983	29	2.4%	-6.29 [-10.64, -1.94]			-		
Khorsandi, H 2019	-5.12	10.25053	18	-1.5	9.66894	22	1.2%	-3.62 [-9.84, 2.60]					
Kirch, N 2018	-0.4	2.3	23	0.7	2.3	23	25.4%	-1.10 [-2.43, 0.23]					
Li, Y 2010	-2.4	6.954854	30	-0.3	8.770975	29	2.7%	-2.10 [-6.15, 1.95]				-	
Luciana Nicolau Aranha 2020	-1	13.2842	30	-1.4	17.10351	39	0.9%	0.40 [-6.77, 7.57]					-
Maria Angelica Arzola-Paniagua 2016	-4.2	8.954893	15	-2.1	8.305221	24	1.4%	-2.10 [-7.72, 3.52]	-				
Marreiro, D. D 2006	-0.9	5.802586	28	0.2	8.450444	28	3.1%	-1.10 [-4.90, 2.70]					
Mohammad Alizadeh 2018	-5.6	12.34139	28	-6.5	17.41063	28	0.7%	0.90 [-7.00, 8.80]					
Pawel Bogdanski 2012	-0.5	7.102112	28	-0.3	5.4111	28	4.1%	-0.20 [-3.51, 3.11]					
Polus, A 2016	0.1	2.505993	24	-0.5	3.104835	35	21.8%	0.60 [-0.84, 2.04]				_	
Szewczyk-Golec, K 2017	-5.3	4.160529	15	-5.4	4.466352	15	4.7%	0.10 [-2.99, 3.19]		-			
Szulinska, M 2017	-1.8	14.73669	25	-0.7	12.83121	25	0.8%	-1.10 [-8.76, 6.56]					
Williams, E. J 2017	-1.2	2.251666	28	-0.6	2.851315	28	24.8%	-0.60 [-1.95, 0.75]					
Total (95% Cl)			446			474	100.0%	-0.78 [-1.45, -0.11]			•		
Heterogeneity: $chi^2 = 19.54$, $df = 17$ (<i>I</i>	P = 0.30); $I^2 = 13\%$										1	
Test for overall effect: $Z = 2.29$ ($P = 0.0$	02)								-10	-5	0	5	10
									Favou	rs [antioxic	dants] Fa	vours [contr	ol]

(b)

Cturber and an and	A	ntioxidants	s T-+-1	Maria	Control	T. (.)	147.1.1.4	Mean difference		N	lean differen	ce	
Study or subgroup	Mean	SD	Iotal	Mean	SD	Total	weight	IV, random, 95% Cl		1V,	random, 959	6 CI	
Kazemipoor, M 2013	-4	0.05	31	-0.01	0.052915	29	20.0%	-3.99 [-4.02, -3.96]					
Khorsandi, H 2019	-0.01	0.101489	18	-0.01	0.065574	22	20.0%	0.00 [-0.05, 0.05]			•		
Luciana Nicolau Aranha 2020	0	0.1	30	0	0.1	39	20.0%	0.00 [-0.05, 0.05]			•		
Marreiro, D. D 2006	0.01	0.47697	28	0	0.045826	28	20.0%	0.01 [-0.17, 0.19]			•		
Santamarina, A.B 2020	0	0.017321	13	-0.01	0.017321	14	20.0%	0.01 [-0.00, 0.02]			- † -		
Total (95% Cl)			120		2	132	100.0%	-0.79 [-2.82, -1.23]					
Heterogeneity: $tau^2 = 5.32$; chi ² Test for overall effect: $Z = 0.77$	$P^2 = 73933$ (P = 0.44	5.97, df = 4 4)	(<i>P</i> < 0.	.00001);	$I^2 = 100\%$				-10	-5	0	5	10

Favours [antioxidants] Favours [control]

1	- 1
	C
۰.	\sim

FIGURE 2: Continued.

Oxidative Medicine and Cellular Longevity

	Aı	ntioxidants			Control			Std. Mean difference		Std. 1	Mean d	lifferenc	e	
Study or subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, random, 95% Cl		IV, ra	andom	, 95% С	1	
Cavedon, E., J 2020	-5	15.86915	16	0.8	17.31387	19	16.5%	-0.34 [-1.01, 0.33]			-	_		
Maria Angelica Arzola-Paniagua 2016	1.2	14.76516	15	0.8	9.882687	24	16.9%	0.03 [-0.61, 0.68]			-			
Marreiro, D. D 2006	4.1	13.53477	28	1.8	14.30524	28	18.9%	0.16 [-0.36, 0.69]						
Most, J 2018	-1.7	5.1	11	-1.2	4.5	14	14.6%	-0.10 [-0.89, 0.69]						
Payahoo, L 2014	12.4	17.94101	30	-2.2	21.12037	30	18.9%	0.74 [0.21, 1.26]						
Szewczyk-Golec, K 2017	3.5	4.762552	15	-4.8	5.941347	15	14.1%	1.50 [0.68, 2.32]						·
Total (95% Cl)			115			130	100.0%	0.32 [-0.16, 0.79]			-			
Heterogeneity: $tau^2 = 0.24$; $chi^2 = 16.2$	2, df =	5(P = 0.00))6); I ² =	= 69%						1				
Test for overall effect: $Z = 1.30$ ($P = 0.1$.9)								-2	-1	0		1	2
									Eavour	e [antiovid	antel I	Favoure	[contro]	d

(d)

Study or subgroup	A: Mean	ntioxidants SD	s Total	Mean	Control SD	Total	Weight	Std. Mean difference IV, random, 95% Cl	Std. Mean IV, randor	difference n, 95% Cl
Cavedon, E., J 2020	-9.7	3.538361	16	1.7	3.850974	19	7.1%	-3.00 [-4.00, -2.00]	•	
de Souza, R. G. M 2018	1.5	9.360791	24	0.69	8.060701	22	10.2%	0.09 [-0.49, 0.67]		•
Eisner, A 2020	0.7	12.7738	19	0.5	12.15031	19	9.8%	0.02 [-0.62, 0.65]		
Hosseinpour-Arjmand, S 2019	2.4	8.301807	23	2.8	9.250405	22	10.2%	-0.04 [-0.63, 0.54]		
Li, Y 2010	2.8	4.869292	30	0.4	5.965735	29	10.7%	0.44 [-0.08, 0.95]	-	
Maria Angelica Arzola-Paniagua 2016	0.9	7.044792	15	0.2	5.597508	24	9.7%	0.11 [-0.53, 0.76]		•
Marreiro, D. D 2006	-0.3	4.596738	28	-2	2.920616	28	10.6%	0.44 [-0.10, 0.97]	-	
Mohammad Alizadeh 2018	2.3	9.657639	28	2	6.509224	28	10.6%	0.04 [-0.49, 0.56]		
Stull, A. J 2010	0	2	25	0.5	2.3	25	10.4%	-0.23 [-0.78, 0.33]		
Williams, E. J 2017	0.1	1.7	28	0.3	2.45153	28	10.6%	-0.09 [-0.62, 0.43]		
Total (95% Cl)			236			244	100.0%	-0.13 [-0.53, 0.26]		
Heterogeneity: $tau^2 = 0.31$; $chi^2 = 40.9$	4, df =	9 (P < 0.00)	0001);	$I^2 = 789$	6					
	-1 -0.5 0	0.5 1								

Test for overall effect: Z = 0.66 (P = 0.51)

(e) Mean difference Mean difference Antioxidants Control Study or subgroup SD Total SD Total Weight IV, fixed, 95% Cl IV, fixed, 95% Cl Mean Mean Dashtabi, A 2016 7.566789 27 1.07 5.076918 32.3% 8.18 [-11.60, -4.76] -7.11 28 -3.2 Hosseinpour-Arjmand, S 2019 13.42796 10.35036 27.7% -3.20 [-10.19, 3.79] -6.4 28 22 Kamali, S. H 2012 -2.66 11.16195 39 4.80 9.851705 30 15.3% 7.54 [-12.51, -2.57] Khorsandi, H 2019 -3.33 10.43335 18 0 12.05988 22 7.8% -3.33 [-10.30, 3.64] Mohammad Alizadeh 2018 $^{-1}$ 7.562407 28 0.3 6.819824 28 26.5% -1.30 [-5.07, 2.47] Taghizadeh, M 2017 3.915354 25 14.80912 25 10.5% -2.70 [-8.70, 3.30] 0.3 3 160 155 100.0% -4.92 [-6.87, -2.98] Total (95% Cl) Heterogeneity: $chi^2 = 9.06$, df = 5 (P = 0.11); $I^2 = 45\%$ -10 -5 0 5 10

Test for overall effect: Z = 4.97 (P < 0.00001)

(f) Mean difference Mean difference Antioxidants Control Weight Study or subgroup SD Total Mean SD Total IV, fixed, 95% Cl IV, fixed, 95% Cl Mean Bogdanski, P 2012 -14 2.066398 30 0.2 3.351119 30 1.2% -1.60 [-3.01, -0.19] Eisner, A 2020 0.5 2.338803 19 0.1 2.457641 19 1.0% 0.40 [-1.13, 1.93] Fathi, M 2020 -0.59 0.545619 25 -0.50.517301 25 27.3% -0.09 [-0.38, 0.20] Marreiro, D. D 2006 -1.52.286919 28 -0.22.402082 28 1.6% -1.30 [-2.53, -0.07] Most, J 2018 -0.2 0.280535 110.36 0.250599 1453.0% -0.56 [-0.77, -0.35] Pawel Bogdanski 2012 1.216553 28 1.252996 28 5.7% -0.70 [-1.35, -0.05] $^{-1}$ -0.3 Payahoo, L 2014 -0.7 1.276715 30 0.1 2.70555 30 2.1% -0.80 [-1.87, 0.27] -0.50 [-1.04, 0.04] Taghizadeh, M 2017 -0.51.2 25 25 8.2% 0 0.655744 Total (95% Cl) 196 199 100.0% -0.45 [-0.61, -0.30] Heterogeneity: $chi^2 = 13.38$, df = 7 (P = 0.06); $I^2 = 48\%$ 0 Test for overall effect: Z = 5.78 (P = 0.00001) -2 -1 1 Favours [antioxidants] Favours [control]

(g)

FIGURE 2: Forest plot evaluating the effects of antioxidants on basic indicators of obesity: BMI (a), WC (b), WHR (c), leptin (d), FM (e), FBG (f), and HOMA-ir (g) in obesity patients and compared with the control group.

3.7. Effects of Antioxidant Supplementation on Liver Function in Obese Patients. The forest plots from stratified meta-analyses in Figure 5 depicted a total of 5 study outcomes, which reported the effects of antioxidants on liver function in obese patients. The results showed that antioxidants supplementation in obese patients did not affect the levels of alanine transaminase (ALT; p = 0.34; Figure 6(a)) and aspartate transaminase (AST; p = 0.55; Figure 6(b)) when compared to the placebo group. Begg's test and Egger's test showed no significant publication bias for AST ($p_{(B)}$ = 1.6918, $p_{(E)} = 0.9968$) and ALT ($p_{(B)} = 1.2659$, $p_{(E)} = 0.4782$). The corresponding funnel plots for evaluating the publication

Favours [antioxidants] Favours [control]

Favours [antioxidants] Favours [control]

	Antioxidants Control							Std. Mean difference	Std. Mean difference
Study or subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, random, 95% Cl	IV, random, 95% Cl
Dashtabi, A 2016	-16.25	32.17467	27	-5.18	29.70627	28	11.1%	-0.35 [-0.89, 0.18]	
de Souza, R. G. M 2019	-12.81	33.50791	24	-8.05	47.04823	22	10.7%	-0.12 [-0.69, 0.46]	
Fathi, M 2020	-19.92	31.28379	25	-18.28	35.34085	25	10.9%	-0.05 [-0.60, 0.51]	
Li, Y 2010	-0.6	0.754983	30	0.1	0.8	29	11.0%	-0.89 [-1.43, -0.35]	
Polus, A 2016	-0.11	0.871091	24	-0.01	0.977548	35	11.2%	-0.11 [-0.63, 0.41]	
Suliburska, J 2014	-0.1	1.113553	44	-0.1	0.888819	44	11.9%	0.00 [-0.42, 0.42]	
Szulinska, M 2017	-0.3	1.014889	25	0.2	0.8544	25	10.8%	-0.52 [-1.09, 0.04]	
Szulinska, M 2018	-0.17	0.783071	46	0.0.84	0.12	46	11.4%	-1.79 [-2.27, -1.30]	
Taghizadeh, M 2017	-2.3	26.10192	25	-1.7	37.5485	25	10.9%	-0.02 [-0.57, 0.54]	
Total (95% Cl)			270			279	100.0%	-0.43 [-0.84, -0.02]	•
Heterogeneity: $tau^2 = 0.31$	l; $chi^2 = 4$	3.55, df = 8	B (P < 0)	00001);	$I^2 = 82\%$				
Test for overall effect: $Z =$	2.07 (P =	-2 -1 0 1 2							

(a)

Favours [antioxidants] Favours [control]

	Aı	ntioxidants			Control			Std. Mean difference	Std. Mean difference
Study or subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, fixed, 95% Cl	IV, fixed, 95% Cl
Baldrick, F. R 2018	-0.02	1.34	36	-0.06	0.85	78	11.7%	0.04 [-0.36, 0.43]	
Bogdanski, P 2012	0	1.352775	30	-0.1	0.8544	30	7.1%	0.09 [-0.42, 0.59]	
Dashtabi, A 2016	-13.5	50.66012	27	-0.22	43.91519	28	6.5%	-0.29 [-0.82, 0.25]	
de Souza, R. G. M 2019	-15	53.00576	24	-10.05	75.13531	22	5.5%	-0.08 [-0.65, 0.50]	
Eisner, A 2020	9.3	32.6694	19	2.1	59.45158	19	4.5%	0.15 [-0.49, 0.78]	
Fathi, M 2020	-13.96	45.4974	25	-24.76	52.31725	25		Not estimable	
Kamali, S. H 2012	-4.76	53.41246	39	-0.24	160.2543	30	8.1%	-0.04 [-0.52, 0.44]	
Kazemipoor, M 2013	11.62	39.40324	31	23.14	46.58845	29	7.1%	-0.26 [-0.77, 0.24]	
Kirch, N 2018	0.011	0.240208	23	0.13	0.180831	23	5.2%	-0.55 [-1.14, 0.04]	
Li, Y 2010	-0.1	0.916515	30	0.2	1.178983	29	6.9%	-0.28 [-0.79, 0.23]	
Most, J 2018	0.18	0.260576	11	0.25	0.230651	14	2.9%	-0.28 [-1.07, 0.52]	
Polus, A 2016	-0.26	0.547814	24	-0.02	0.371618	35	6.5%	-0.53 [-1.05, 0.00]	
Stull, A. J 2010	0.13	0.175214	25	0.23	0.238956	25	5.8%	-0.47 [-1.03, 0.09]	
Suliburska, J 2014	-0.1	1.473092	44	0	0.818535	44	10.4%	-0.08 [-0.50, 0.33]	
Szulinska, M 2017	-0.1	0.953939	25	0.1	1.153256	25	5.9%	-0.19 [-0.74, 0.37]	
Taghizadeh, M 2017	-2.5	70.76129	25	8.2	53.20592	25	5.9%	-0.17 [-0.72, 0.39]	
Total (95% Cl)			413			456	100.0%	-0.17 [-0.31, -0.04]	•
Heterogeneity: $chi^2 = 8.5^{\circ}$	5. df = 14	(P = 0.086)	$I^{2} = 0$	%		100	2221070		· · · · · · · · · · · · · · · · · · ·
Test for overall effect: 7 =	253(D - 1)	- 0.01)	,. = 0						-1 -0.5 0 0.5 1
rest for overall effect: Z -	- 2.33 (F -	- 0.01)							Essence [see anim antal] Essence [sector]

Favours [experimental] Favours [control]



(b)

(c)

FIGURE 3: Continued.

Oxidative Medicine and Cellular Longevity

	А	ntioxidants			Control			Std. Mean difference	Std. Mean difference
Study or subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, random, 95% Cl	IV, random, 95% Cl
Baldrick, F. R 2018	-0.04	0.14	36	-0.04	0.11	78	8.0%	0.00 [-0.39, 0.39]	
Bogdanski, P 2012	0	0.3	30	0.1	0.3	30	6.8%	-0.33 [-0.84, 0.18]	
Dashtabi, A 2016	3.71	2.750873	27	-0.46	2.773878	28	5.9%	1.49 [0.89, 2.09]	
de Souza, R. G. M 2019	3.63	9.197195	24	-1.19	9.653947	22	6.1%	0.50 [-0.09, 1.09]	
Eisner, A 2020	3.1	4.293018	19	1.6	7.686352	19	5.6%	0.24 [-0.40, 0.87]	
Fathi, M 2020	2.24	9.140279	25	3.8	8.267787	25	6.4%	-0.18 [-0.73, 0.38]	
Kamali, S. H 2012	-6.17	9.92456	39	-2.35	9.977369	30	7.1%	-0.38 [-0.86, 0.10]	
Kazemipoor, M 2013	0.81	9.845705	31	-1.24	8.983758	29	6.8%	0.21 [-0.29, 0.72]	- +-
Li, Y 2010	0.4	0.360555	30	0.1	0.34641	29	6.6%	0.84 [0.30, 1.37]	
Luciana Nicolau Aranha 2020	1	14.20106	30	-1.7	14.02747	39	7.1%	0.19 [-0.29, 0.67]	- -
Pawel Bogdanski 2012	0.2	0.264575	28	0.1	0.2	28	6.6%	0.42 [-0.11, 0.95]	+
Stull, A. J 2010	-0.23	0.072111	25	-0.25	0.065574	25	6.4%	0.29 [-0.27, 0.84]	
Szulinska, M 2017	0	0.3	25	-0.1	0.4	25	6.4%	0.28 [-0.28, 0.84]	
Szulinska, M 2018	0.01	0.262298	46	-0.08	0.564712	46	7.8%	0.20 [-0.21, 0.61]	
Taghizadeh, M 2017	0.7	7.053368	25	-0.6	7.607891	25	6.4%	0.17 [-0.38, 0.73]	
Total (95% Cl)			440			478	100.0%	0.25 [0.03, 0.46]	•
Heterogeneity: tau ² = 0.11; chi	$^{2} = 37.33$	8, df = 14 (<i>H</i>	P = 0.00	$007); I^2$	= 62%				
Test for overall effect: $Z = 2.20$	P = 0.0	03)							-2 -1 0 1 2 Eavours [antioxidants] Eavours [control]
									Pavours [annoxidants] Favours [control]

(d)

FIGURE 3: Forest plot evaluating the effects of antioxidants on lipid metabolism indexes: TC (a), TG (b), LDL (c), and HDL (d) in obesity patients and compared with the control group.



(b)

FIGURE 4: Forest plot evaluating the effects of antioxidants on systemic antioxidant capacity indexes: MDA (a) and SOD (b) in obesity patients and compared with the control group.

bias are shown in Supplemental Figure S6. The subgroup analysis further revealed that obese patients with watersoluble, fat-soluble, or mixed antioxidants supplementation did not exhibit any significant changes in the levels of ALT (p > 0.05) and AST (p > 0.05) when compared to the placebo group (Supplementary Table S5).

4. Discussion

Obesity, which considered a global health problem, has met the medical definition of disease. Complications from obesity affect almost every tissue in the body, which is a major cause of cardiovascular events, diabetes [23] as well as infertility [24] and so on. Obesity triggers a host of metabolic disorders, while there are no effective treatments for it. Dietderived antioxidant, a class of substances that prevent the harmful effects of ROS in daily nutrition and human health, helping capture and neutralize free radicals, has plentiful positive effects on human metabolism [11]. Moreover, several different kinds of antioxidants are reported to improve the metabolic abnormalities associated with obesity [12–14]. However, the effects of antioxidants on improving metabolic disorders in obese patients are controversial. In this meta-analysis, we further explored the effects of antioxidants on obesity in terms of basic indicators of obesity, lipid metabolism, systemic antioxidant capacity, inflammatory biomarkers and liver function, and further examined whether antioxidants can effectively improve the metabolic function in obese patients, which may provide robust evidence for clinical obesity management.



FIGURE 5: Forest plot evaluating the effects of antioxidants on inflammatory biomarkers: TNF- α (a), IL-6 (b), and CRP (c) in obesity patients and compared with the control group.

Study or subgroup	Aı Mean	ntioxidants SD	Total	Mean	Control SD	Total	Weight	Mean difference IV, fixed, 95% Cl	Mean difference IV, fixed, 95% Cl		
Bogdanski, P 2012 Hosseinpour-Arjmand, S 2019 Kamali, S. H 2012 Mohammad Alizadeh 2018	-2.5 -8.5 -5.2 -0.5	14.81621 23.47105 8.678404 5.56507	24 23 30 28	-1.4 -6.6 0.38 -0.6	10.62968 16.46117 16.37316 7.65441	22 22 39 28	34.3% 13.5% 52.2%	-1.10 [-8.51, 6.31] -1.90 [-13.70, 9.90] -5.58 [-11.58, 0.42] Not estimable			
Total (95% Cl) 77 Heterogeneity: $chi^2 = 0.93$, $df = 2$ ($P = 0.63$); $I^2 = 0\%$ Test for overall effect: $Z = 1.60$ ($P = 0.11$)						83	100.0%	-3.55 [-7.88, 0.79]	-10 -5 0 5 10 Favours [antioxidants] Favours [control]		
(a)											
Study or subgroup	A Mean	ntioxidants SD	Total	Mean	Control SD	Total	Weight	Mean difference IV, fixed, 95% Cl	Mean difference IV, fixed, 95% Cl		

Study or subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, fixed, 95% Cl	IV, fixed, 95% Cl	
de Souza, R. G. M 2018	1.7	16.39909	24	2.9	9.071384	22	13.0%	-1.20 [-8.78, 6.38]		
Hosseinpour-Arjmand, S 2019	-6.2	13.95242	23	-3.3	9.18858	22	15.8%	-2.90 [-9.77, 3.97]		
Kamali, S. H 2012	-1.23	5.456409	30	0.24	8.215114	39	71.2%	-1.47 [-4.70, 1.76]		
Mohammad Alizadeh 2018	-0.6	3.407345	28	-0.8	4.784349	28		Not estimable		
Total (95% Cl) Heterogeneity: chi ² = 0.15, df = Test for overall effect: <i>Z</i> = 1.19 (77 %			83	100.0%	-0.66 [-4.39, 1.07]	-10 -5 0 5 10 Favours [antioxidants] Favours [control]			

(b)

FIGURE 6: Forest plot evaluating the effects of antioxidants on liver function indexes: ALT (a) and AST (b) obesity patients and compared with the control group.

Firstly, we collected data from 27 studies to assess the effects of antioxidants supplementation on improving physiological dimension in obese patients. The results of metaanalysis showed that antioxidants supplementation could significantly decrease BMI and WC level when compared to the placebo group. Previous studies also showed that

anthocyanin (possessing antioxidant properties) supplementation was sufficient to reduce the BMI and body weight in obese patients [25]. However, green tea extract as antioxidants supplementation was not associated with reductions in BMI or WC in obese women [26]. However, our studies failed to prove that antioxidants have a beneficial effect on WHR and FM levels in patients. Leptin as major adipokine and the product of obese gene, synthesized by the white adipocyte tissue, mastering feeding and metabolism acting at central level in the brain [27]. Increased leptin levels were found in obese animal [28]. Leptin could improve insulin sensitivity and reduce insulin resistance [29]. Our result showed that antioxidants supplementation did not significantly affect leptin level in the obese patient, which was consistent with previous findings showing that neither green tea intake nor resveratrol intake had significant effects on serum levels of leptin [30, 31]. On the other hand, antioxidants supplementation significantly decreased HOMA-IR and FBG value in obesity patients. Studies have shown that 60-day saccharomyces boulardii and superoxide Dismutase supplementation could decrease HOMA-IR in obese adults [32]. Ellulu et al. demonstrate that Vitamin C (500 mg twice daily) could reduce FBG in hypertensive and/or diabetic obese patients [33]. Collectively, the effect of antioxidants on basic indicators and glucose metabolic function with obesity was positive.

Obesity is a common disease, which is the manifestation of excess body fat and is closely associated with the disorder of lipid metabolism, which driven by the effects of insulin resistance and pro-inflammatory adipokines [34]. The disorder of lipid metabolism in obese patients is manifested as high TC, TG and LDL levels and low HDL levels [35]. High-fat diet causes the imbalance between lipid absorption and metabolism, resulting in lipid metabolism disorders, which can cause a variety of metabolic diseases such as cardiovascular and cerebrovascular events and non-alcoholic fatty liver [36]. However, antioxidants do modulate lipid metabolism in the animal studies and RCTs [37, 38]. Therefore, we collected data from 17 existing studies to evaluate the role antioxidants played in the treatment of dyslipidemia with obesity. The meta-analysis results revealed that that antioxidants supplementation could reduce TC, TG and LDL levels and increased HDL levels in obese patients when compared to the placebo group, which was consistent with previous findings [39, 40], However, the Begg's test suggests the publication bias in these included studies, thus, the effects of antioxidants on regulating lipid metabolism disorder require further confirmation by more studies.

The high fat diet and carbohydrates can cause a significant increase in oxidative stress and inflammation in obese patients [41]. MDA as a oxidative stress biomarker, is the products of the peroxidation of polyunsaturated fatty acids, and is elevated in the serum of obese human and animals [42, 43]. While with increasing of adipose tissue, the activity of antioxidant enzymes such as SOD diminished significantly [8]. Those above evidence suggests that systemic antioxidant capacity was impaired in obese patients, while, antioxidants supplementation has been found to alleviate the impairment in obese patients [44]. In this study, we collected data from 4 existing studies to evaluate the importance of antioxidants in the ameliorating systemic antioxidant capacity with obesity. Results of meta-analysis revealed that antioxidants supplementation reduced MDA level and increased SOD level in obese patients when compared to the placebo group. However, existing RCTs rarely mention changes in other oxidative stress markers such as catalase, glutathione, peroxidase, total antioxidant capacity and so on, therefore, these effects of antioxidant supplementation on these parameters have not been examined in this study. The above evidence indicated the beneficial effects of antioxidants on oxidative stress in obese patients; whilst more high-quality studies are still required to confirm the results from this meta-analysis.

Inflammation is also a common pathological process in obese patients [41]. Dysfunctional adipocytes can secrete inflammatory adipokines such as such as TNF- α and IL-6, which can initiate adipose tissue inflammation [45]. The occurrence of adipose tissue inflammation in different tissues can negatively impacts organ function, for example reduced oocyte quality [46] and cancer [47]. Besides, CRP is also increased in obese patients as proinflammatory biomarkers. In this study, results of meta-analysis indicated that antioxidants supplementation in obese patients significantly reduced TNF- α level when compared to the placebo group, while the IL-6 and CRP levels were not significantly affected by antioxidants supplementation. These results suggest that antioxidants supplementation may attenuated inflammation in obese patients, which may be confirmed by more highquality studies.

Accumulation of body fat and abnormal lipid metabolism in obese patients can seriously affect liver function [48, 49]. AST and ALT are mainly synthesized in the mitochondria of liver cells, and elevation of AST and ALT levels are closely correlated with impaired liver function. Animal studies have shown that antioxidants can improve nonalcoholic fatty liver disease in mice with high fat diet-induced obesity [50]. In this study, we collected data from 5 existing studies to evaluate the importance of antioxidants in liver function of obese patients. Our meta-analysis results showed that antioxidants supplementation in obese patients had no effect on AST and ALT levels; while the effects of antioxidants on the liver function of obese patients still require further investigation.

Antioxidants have different chemical structures and can be classified into two gross divisions, depending on their solubility in water (hydrophilic) or fat (hydrophobic). Generally, water soluble antioxidants react with ROS within cells or body fluids (blood serum, extracellular fluid, seminal plasma) while fat-soluble antioxidants are more prone to protect cell membranes from ROS-mediated lipid peroxidation [51, 52]. Lazzarino et al., showed that fat-soluble antioxidants in seminal plasma was much lower than water-soluble antioxidants, suggesting that their administration to treat male infertility characterized by excess ROS production should be performed for a prolonged period of time [51]. Our subgroup analysis showed that different types of antioxidants had differential effects on metabolic disorders, while the specific roles of water- and fat-soluble antioxidants in obese patients may still require further studies.

The strengthen of the study is that the meta-analysis is the first article to study the effects of clinical antioxidants on the metabolism of obesity, providing a therapeutic basis for clinical intervention in obesity-induced metabolic disorders. This systematic review has the following limitations: (1) studies, study populations, and main results in cumulative analyses were heterogeneous. Due to the relatively small number of studies, our analyses had limited power. (2) Only English paper was took into account in this study, which may affect the accuracy of the results. (3) The types of antioxidants included in the study were diverse. Thus, it was not clear which antioxidant was the most effective. In addition, for more precise findings and accurate conclusions, more high-quality trials are needed to assess the beneficial effects of antioxidants on obesity and its metabolism. (4) The protocol of this study has not been pre-registered, which may induce potential bias to the review.

5. Conclusions

The meta-analysis results indicated that antioxidants supplementation exerted potential beneficial effects in obese patients by regulating FBG, oxidative stress and inflammation, whilst more high-quality studies are required to confirm these effects. The present study may provide important insights for the treatment of clinical obesity and obesity-associated complications.

Appendix

A. Pubmed Search Strategy

Searched August 8, 2021.

#1 "Obesity"[MeSH Terms] OR "Overweight"[MeSH Terms] OR "Weight Gain"[MeSH Terms] OR "Body Mass Index"[MeSH Terms] (338,808)

#2 "obes*"[Title/Abstract] OR "adipos*"[Title/Abstract] OR "Overweight"[Title/Abstract] OR "Overweight"[Title/ Abstract] OR "weight gain"[Title/Abstract] OR "Body Mass Index"[Title/Abstract] OR "BMI"[Title/Abstract] (632,372)

#3 #1 OR #2 (703609)

#4 "Antioxidants" [MeSH Terms] OR "Vitamins" [MeSH Terms] OR "Vitamin E" [MeSH Terms] OR "Ascorbic Acid" [MeSH Terms] OR "Tocopherols" [MeSH Terms] OR "Selenium" [MeSH Terms] OR "Zinc" [MeSH Terms] OR "Ubiquinone" [MeSH Terms] OR "Acetylcysteine" [MeSH Terms] OR "Carnitine" [MeSH Terms] OR "Melatonin" [MeSH Terms] OR "Glutathione" [MeSH Terms] OR "Carotenoids" [MeSH Terms] OR "Arginine" [MeSH Terms] OR "Resveratrol" [MeSH Terms] (528,347)

#5 "Antioxidants"[Title/Abstract] OR "Vitamin"[Title/ Abstract] OR "vitamin E"[Title/Abstract] OR "tocopherol*" [Title/Abstract] OR "alpha tocopherol*"[Title/Abstract] OR "tocotrienol"[Title/Abstract] OR "vitamin C"[Title/ Abstract] OR "ascorbic acid"[Title/Abstract] OR "ascorb*" [Title/Abstract] OR "selenium"[Title/Abstract] OR "selen* "[Title/Abstract] OR "selenium"[Title/Abstract] OR "selen* "[Title/Abstract] OR "zinc"[Title/Abstract] OR "zinc*"[Title/ Abstract] OR "ubiquinone"[Title/Abstract] OR "ubiquinol" [Title/Abstract] OR "coenzyme Q10"[Title/Abstract] OR "CoQ10"[Title/Abstract] OR "Acetylcysteine"[Title/Abstract] OR "Carnitine"[Title/Abstract] OR "carnitene"[Title/Abstract] OR "melatonin"[Title/Abstract] OR "Glutathione"[Title/ Abstract] OR "GSH"[Title/Abstract] OR "carotene"[Title/ Abstract] OR "betacarotene"[Title/Abstract] OR "Arginine" [Title/Abstract] OR "resveratrol*"[Title/Abstract] (753,601)

#6 #4 OR #5 (955,211)

#7 #3 AND #6 (33091)

#8 ("Randomized Controlled Trial" [Publication Type] OR "Controlled Clinical Trial" [Publication Type] OR "Clinical Trials as Topic" [MeSH Terms:noexp] OR "randomized" [Title/Abstract] OR "placebo" [Title/Abstract] OR "randomly" [Title/Abstract] OR "trial" [Title/Abstract]) NOT ("Animals" [MeSH Terms] OR "mice" [Title/Abstract] OR "mouse" [Title/Abstract] OR "review" [Title/Abstract] OR "rats" [Title/Abstract] OR "fish" [Title/Abstract]) (169,491)

#9 #7 AND #8 (465)

B. Embase Search Strategy

Searched August 3, 2021.

#1 "obesity'/exp OR "overweight"/exp OR "weight gain"/ exp OR "body mass index"/exp (1110754)

#2 obes*:ab,ti OR adipos*:ab,ti OR overweight:ab,ti OR "over weight":ab,ti OR "weight gain":ab,ti OR "body mass index":ab,ti OR bmi:ab,ti (971,155)

#3 #1 OR #2 (1,263,198)

#4 "antioxidants"/exp OR "vitamins"/exp OR "vitamin e"/exp OR "ascorbic acid"/exp OR "Tocopherols"/exp OR "Selenium"/exp OR "Zinc"/exp OR "Ubiquinone"/exp OR "Acetylcysteine"/exp OR "Carnitine"/exp OR "Melatonin"/ exp OR "Glutathione"/exp OR "Carotenoids"/exp OR "Arginine"/exp OR "Resveratrol"/exp (1,278,012)

#5 Antioxidants:ab,ti OR Vitamin:ab,ti ORVitamin E:ab,ti OR tocopherol*:ab,ti OR alpha tocopherol*:ab,ti OR tocotrienol:ab,ti OR vitamin C:ab,ti OR ascorbic acid: ab,ti OR ascorb*:ab,ti OR selenium:ab,ti OR selen*:ab,ti OR zinc C:ab,ti OR zinc*:ab,ti OR ubiquinone:ab,ti OR ubiquinol:ab,ti OR coenzyme Q10:ab,ti OR CoQ10:ab,ti OR Acetylcysteine:ab,ti OR Carnitine:ab,ti OR carnitene:ab,ti OR melatonin:ab,ti OR Glutathione:ab,ti OR GSH:ab,ti OR carotene:ab,ti OR betacarotene:ab,ti OR Arginine:ab,ti OR resveratrol*:ab,ti (381,624)

#6 #4 OR #5 (1,418,211)

#7 ("randomized controlled trial"/exp OR "controlled clinical trial"/exp OR "clinical trials as topic"/exp OR "randomized":ab,ti OR "placebo":ab,ti OR "randomly":ab,ti OR "trial":ab,ti) NOT "animals"/exp NOT "mice":ab,ti NOT "mouse":ab,ti NOT "fish":ab,ti NOT "review":ab,ti (137,022)

#8 #6 AND #7 (4732)

#9 #3 AND #8(341)

(("obesity"/exp OR "overweight"/exp OR "weight gain"/ exp OR "body mass index"/exp) OR (obes*:ab,ti OR adipos*:ab,ti OR overweight:ab,ti OR "over weight":ab,ti OR "weight gain":ab,ti OR "body mass index":ab,ti OR bmi:ab,ti OR "HFD":ab,ti)) AND (("antioxidants"/exp OR "vitamins"/exp OR "vitamin e"/exp OR "ascorbic acid"/exp OR "Tocopherols"/exp OR "Selenium"/exp OR "Zinc"/exp OR "Ubiquinone"/exp OR "Acetylcysteine"/exp OR "Carotenoids"/exp OR "Arginine"/exp OR "Resveratrol"/ exp) OR (Antioxidants:ab,ti OR Vitamin:ab,ti ORVitamin E:ab,ti OR tocopherol*:ab,ti OR alpha tocopherol*:ab,ti OR tocotrienol:ab,ti OR vitamin C:ab,ti OR ascorbic acid: ab,ti OR ascorb*:ab,ti OR selenium:ab,ti OR selen*:ab,ti OR zinc C:ab,ti OR zinc*:ab,ti OR ubiquinone:ab,ti OR ubiquinol:ab,ti OR coenzyme Q10:ab,ti OR CoQ10:ab,ti OR Acetylcysteine:ab,ti OR Carnitine:ab,ti OR carnitene:ab,ti OR melatonin:ab,ti OR Glutathione:ab,ti OR GSH:ab,ti OR carotene:ab,ti OR betacarotene:ab,ti OR Arginine:ab,ti OR resveratrol*:ab,ti)) AND (("randomized controlled trial"/ exp OR "controlled clinical trial"/exp OR "clinical trials as topic"/exp OR "randomized":ab,ti OR "placebo":ab,ti OR "randomly":ab,ti OR "trial":ab,ti) NOT "animals"/exp NOT "mice":ab,ti NOT "mouse":ab,ti NOT "fish":ab,ti NOT "review":ab.ti)

C. The Cochrane Library Search Strategy

Searched August 8, 2021.

#1 MeSH descriptor:[Obesity] explode all trees(14700)

#2 MeSH descriptor:[Overweight] explode all trees(17455)

#3 MeSH descriptor:[Weight Gain] explode all trees(2668)

#4 MeSH descriptor:[Body Mass Index] explode all trees(10459)

#5 (obes*):ti,ab,kw(45850)

#6 (adipos*):ti,ab,kw(8227)

#7 (Overweight):ti,ab,kw(17932)

#8 (Over weight):ti,ab,kw(27326)

#9 (weight gain):ti,ab,kw(13878)

#10 (Body Mass Index):ti,ab,kw(44398)

#11 (BMI):ti,ab,kw(43349)

#12 #1 OR #2 OR #3 OR #4 OR #5 OR #6 OR #7 OR #8 OR #9 OR #10 OR #11 (122448)

#13 (Resveratrol*):ti,ab,kw(676)

#14 (betacarotene):ti,ab,kw(1226)

#15 MeSH descriptor: [Antioxidants] explode all trees(4999)

#16 (Antioxidants):ti,ab,kw (6575)

#17 MeSH descriptor:[Vitamins] explode all trees(4863)

#18 (Vitamin):ti,ab,kw(30878)

#19 MeSH descriptor:[Vitamin E] explode all trees(2572)

#20 ("Vitamin E"):ti,ab,kw(4792)

#21 MeSH descriptor:[Tocopherols] explode all trees(752)

#22 (Tocopherol*):ti,ab,kw(2989)

#23 (alpha tocopherol*):ti,ab,kw(2456)

#24 (tocotrienol):ti,ab,kw(121)

#25 MeSH descriptor:[Ascorbic Acid] explode all trees(2291)

#26 ("Vitamin C"):ti,ab,kw (3631)

#27 (ascorbic acid):ti,ab,kw(3915)

#28 (ascorb*):ti,ab,kw(4153)

#29 MeSH descriptor:[Selenium] explode all trees(736)

#30 (Selenium):ti,ab,kw (2163)

#31 (selen*):ti,ab,kw(2459)

#32 MeSH descriptor:[Zinc] explode all trees(1677)

#33 (Zinc):ti,ab,kw(8083)

#34 MeSH descriptor:[Ubiquinone] explode all trees(576)

#35 (ubiquinone):ti,ab,kw (673)

#36 (ubiquinol):ti,ab,kw(121)

#37 ("coenzyme Q10"):ti,ab,kw(1016)

#38 (CoQ10):ti,ab,kw(506)

#39 MeSH descriptor:[Acetylcysteine] explode all trees(1155)

#40 (Acetylcysteine):ti,ab,kw(2337)

#41 MeSH descriptor:[Carnitine] explode all trees(649)

#42 (Carnitine):ti,ab,kw (1825)

#43 (carnitene):ti,ab,kw(3)

#44 MeSH descriptor:[Melatonin] explode all trees(1255)

#45 (melatonin):ti,ab,kw (3058)

#46 MeSH descriptor:[Glutathione] explode all trees(692)

#47 (Glutathione):ti,ab,kw (3873)

#48 (GSH):ti,ab,kw(1286)

#49 MeSH descriptor:[Carotenoids] explode all trees(3764)

#50 (carotene):ti,ab,kw (2013)

#51 MeSH descriptor:[Arginine] explode all trees(1494) #52 (Arginine):ti,ab,kw (4569)

#53 MeSH descriptor:[Resveratrol] explode all trees(308) #54 #13 OR #14 OR #15 OR #16 OR #17 OR #18 OR #19

OR #20 OR #21 OR #22 OR #23 OR #24 OR #25 OR #26 OR #27 OR #28 OR #29 OR #30 OR #31 OR #32 OR #33 OR #34 OR #35 OR #36 OR #37 OR #38 OR #39 OR #40 OR #41 OR #42 OR #43 OR #44 OR #45 OR #46 OR #47 OR #48 OR #49

OR #50 OR #51 OR #52 OR #53 (61638)

#55 #12 AND #54 (7782)

#56 MeSH descriptor:[fish] explode all trees (316)

#57 (animals):ti,ab,kw(14781)

#58 MeSH descriptor:[animals] explode all trees(614391)

#59 MeSH descriptor:[mice] explode all trees (1065)

#60 (mice):ti,ab,kw(4831)

#61 (mouse):ti,ab,kw(4831)

#62 MeSH descriptor:[mouse] explode all trees (1065)

#63 MeSH descriptor:[rats] explode all trees (1008)

#64 (rats):ti,ab,kw(2816)

#65 (fish):ti,ab,kw(6542)

#66 #56 OR #57 OR #58 OR #59 OR #60 OR #61 OR #62

OR #63 OR #64 OR #65 (625298)

#67 #55 NOT #66(4475) (Trials4471)

D. Web of Science Search Strategy

Searched August 3, 2021.

#1 TS=(Obesity OR Overweight OR Weight Gain OR Body Mass Index) OR AB=(obes* OR adipos* OR Overweight OR Over weight OR weight gain OR Body Mass Index OR BMI) (657762)

#2 TS=(Antioxidants OR Vitamins OR Vitamin E OR Ascorbic Acid OR Tocopherols OR Selenium OR Zinc OR Ubiquinone OR Acetylcysteine OR Carnitine OR Melatonin OR Glutathione OR Carotenoids] OR Arginine OR Resveratrol) OR AB=(Antioxidants OR Vitamin OR vitamin E OR tocopherol* OR alpha tocopherol* OR tocotrienol OR vitamin C OR ascorbic acid OR ascorb* OR selenium OR selen* OR zinc OR zinc* OR ubiquinone OR ubiquinol OR coenzyme Q10 OR CoQ10 OR Acetylcysteine OR Carnitine OR carnitene OR melatonin OR Glutathione OR GSH OR carotene OR betacarotene OR Arginine OR resveratrol*) (926136) #3 (TS=(Randomized Controlled Trial OR Controlled Clinical Trial OR Clinical Trials as Topic) OR AB=(randomized OR placebo OR randomly OR trial)) NOT TS=(Animals) NOT AB=(mice) NOT AB=(mouse) NOT AB=(fish) NOT AB=(review) NOT TS=(rats) NOT TS=(rabbit) (911,209)

#4 #1 AND #2 AND #3 (4670)

(TS=(Randomized Controlled Trial OR Controlled Clinical Trial OR Clinical Trials as Topic) OR AB=(randomized OR placebo OR randomly OR trial)) NOT TS=(Animals) NOT AB=(mice) NOT AB=(mouse) NOT AB=(fish) NOT TS=(rats) NOT TS=(rabbit) NOT AB=(review) AND (TS=(Antioxidants OR Vitamins OR Vitamin E OR Ascorbic Acid OR Tocopherols OR Selenium OR Zinc OR Ubiquinone OR Acetylcysteine OR Carnitine OR Melatonin OR Glutathione OR Carotenoids] OR Arginine OR Resveratrol) OR AB=(Antioxidants OR Vitamin OR vitamin E OR tocopherol* OR alpha tocopherol* OR tocotrienol OR vitamin C OR ascorbic acid OR ascorb* OR selenium OR selen* OR zinc OR zinc* OR ubiquinone OR ubiquinol OR coenzyme Q10 OR CoQ10 OR Acetylcysteine OR Carnitine OR carnitene OR melatonin OR Glutathione OR GSH OR carotene OR betacarotene OR Arginine OR resveratrol*)) AND (TS=(Obesity OR Overweight OR Weight Gain OR Body Mass Index) OR AB=(obes* OR adipos* OR Overweight OR Over weight OR weight gain OR Body Mass Index OR BMI OR HFD))

E. Scopus Search Strategy

Searched August 3, 2021.

#1 TITLE-ABS (obes*) OR TITLE-ABS (obesity) OR TITLE-ABS (overweight) OR TITLE-ABS ("weight gain") OR TITLE-ABS ("body mass index") OR TITLE-ABS (adipos*) OR TITLE-ABS ("Over weight") OR TITLE-ABS (BMI) (756,855)

#2 TITLE-ABS (Antioxidants) OR TITLE-ABS (Vitamin) OR TITLE-ABS ("vitamin E") OR TITLE-ABS (tocopherol*) OR TITLE-ABS ("alpha tocopherol*") OR TITLE-ABS (tocotrienol) OR TITLE-ABS ("vitamin C") OR TITLE-ABS ("ascorbic acid") OR TITLE-ABS (ascorb*) OR TITLE-ABS (selenium) OR TITLE-ABS (ascorb*) OR TITLE-ABS (selenium) OR TITLE-ABS (selen*) OR TITLE-ABS (zinc) OR TITLE-ABS (ubiquinone) OR TITLE-ABS (ubiquinol) OR TITLE-ABS (ubiquinone) OR TITLE-ABS (coQ10) OR TITLE-ABS (coenzyme Q10") OR TITLE-ABS (CoQ10) OR TITLE-ABS (Acetylcysteine) OR TITLE-ABS (Carnitine) OR TITLE-ABS (carnitene) OR TITLE-ABS (melatonin) OR TITLE-ABS (Glutathione) OR TITLE-ABS (GSH) OR TITLE-ABS (carotene) OR TITLE-ABS (betacarotene) OR TITLE-ABS (Arginine) OR TITLE-ABS (resveratrol*) (1,413,400)

#3 (TITLE-ABS (randomized) OR TITLE-ABS (placebo) OR TITLE-ABS (randomly) OR TITLE-ABS (trial)) NOT TITLE-ABS (animals) OR TITLE-ABS (mice) OR TITLE-ABS (mouse) OR TITLE-ABS (review) (228386)

#4 #1 AND #2 AND 3 (921)

(TITLE-ABS (obes*) OR TITLE-ABS (obesity) OR TITLE-ABS (overweight) OR TITLE-ABS ("weight gain") OR TITLE-ABS ("body mass index") OR TITLE-ABS (adipos*) OR TITLE-ABS ("Over weight") OR TITLE-ABS ("BMI")) AND ((TITLE-ABS (randomized) OR TITLE-ABS (placebo) OR TITLE-ABS (randomly) OR TITLE-ABS (trial)) NOT TITLE-ABS (animals) OR TITLE-ABS (mice) OR TITLE-ABS (mouse) OR TITLE-ABS (review)) AND (TITLE-ABS (Antioxidants) OR TITLE-ABS (Vitamin) OR TITLE-ABS ("vitamin E") OR TITLE-ABS (tocopherol*) OR TITLE-ABS ("alpha tocopherol*") OR TITLE-ABS (tocotrienol) OR TITLE-ABS ("vitamin C") OR TITLE-ABS ("ascorbic acid") OR TITLE-ABS (ascorb*) OR TITLE-ABS (selenium) OR TITLE-ABS (selen*) OR TITLE-ABS (zinc) OR TITLE-ABS (ubiquinone) OR TITLE-ABS (ubiquinol) OR TITLE-ABS ("coenzyme Q10") OR TITLE-ABS (CoQ10) OR TITLE-ABS (Acetylcysteine) OR TITLE-ABS (Carnitine) OR TITLE-ABS (carnitene) OR TITLE-ABS (melatonin) OR TITLE-ABS (Glutathione) OR TITLE-ABS (GSH) OR TITLE-ABS (carotene) OR TITLE-ABS (betacarotene) OR TITLE-ABS (Arginine) OR TITLE-ABS (resveratrol*))

Data Availability

All the data are available upon reasonable request from the corresponding authors.

Conflicts of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Authors' Contributions

JYW, XCL, and YLY designed the whole study. JYW, CSW, and BYL collected and analyzed the data. JYW and OZ performed the statistical analysis. JYW wrote the manuscript. All authors approved the manuscript for submission. Jinyuan Wang, Biyun Liao and Changsheng Wang equally contributed to this work.

Acknowledgments

This work was supported by the National Natural Science Foundation of China (No. 82101720), the Scientific Research Elevation Project of Young Faculty from Guangxi Universities (2019KY0567), and the Scientific Research and Technological of Baise in China (20183331).

Supplementary Materials

Supplementary 1. Figure S1: risk of bias plot of included studies. Figure S2: funnel plot evaluating study bias of antioxidants on basic indicators of obesity: BMI (A), WC (B), WHR (C), leptin (D), FM (E), FBG (F), and HOMA-ir (G) in obesity patients and compared with the control group. Figure S3: funnel plot evaluating study bias for the effects of antioxidants on lipid metabolism indexes: TC (A), TG (B), LDL (C), and HDL (D) in obesity patients and compared with the control group. Figure S4: funnel plot evaluating study bias for the effects of antioxidants on systemic antioxidant capacity indexes MDA (A) and SOD (B) in obesity patients and compared with the control group. Figure S5: funnel plot evaluating study bias for the effects of antioxidants on inflammatory biomarkers: TNF- α (a), IL-6 (b), and CRP (c) in obesity patients and compared with the control group. Figure S6: funnel plot evaluating study bias for the effects of antioxidants on liver function indexes: ALT (A) and AST (B) in obesity patients and compared with the control group.

Supplementary 2. Table S1: subgroup analysis of the effects of antioxidants on BMI, WC, WHR, leptin, FM, FBG, and HOMA-ir in obesity patients and compared with the control group. Table S2: subgroup analysis of the effects of antioxidants on TC, TG, LDL, and HDL in obesity patients and compared with the control group. Table S3: subgroup analysis of the effects of antioxidants on MDA and SOD in obesity patients and compared with the compared with the control group. Table S4: subgroup analysis of the effects of antioxidants on TNF- α , IL-6, and CRP in obesity patients and compared with the control group. Table S5: subgroup analysis of the effects of antioxidants on the effects of antioxidants on the control group. Table S5: subgroup analysis of the effects of antioxidants on the effects of antioxidants on ALT and AST in obesity patients and compared with the control group.

References

- B. Caballero, "Humans against obesity: who will win?," Advances in Nutrition, vol. 10, Supplement_1, pp. S4–S9, 2019.
- [2] The GBD 2015 Obesity Collaborators, "Health effects of overweight and obesity in 195 countries over 25 years," *The New England Journal of Medicine*, vol. 377, no. 1, pp. 13–27, 2017.
- [3] S. Bazrafshani, H. Randhawa, Y. Ghaedi, S. Khan, and S. Sharbatti, "The prevalence of overweight and obesity among health care providers in the emirate of Ajman, UAE," *Journal* of Complementary Medicine Research, vol. 11, no. 3, pp. 40– 50, 2020.
- [4] A. Valerio, E. Nisoli, A. P. Rossi, M. Pellegrini, T. Todesco, and M. E. Ghoch, "Obesity and higher risk for severe complications of COVID-19: what to do when the two pandemics meet," *Journal of Population Therapeutics and Clinical Pharmacology*, vol. 27, Supplement Part 1, pp. e31–e36, 2020.
- [5] J. I. Malone and B. C. Hansen, "Does obesity cause type 2 diabetes mellitus (T2DM)? Or is it the opposite?," *Pediatric Diabetes*, vol. 20, no. 1, pp. 5–9, 2019.
- [6] A. Nikolopoulou and N. P. Kadoglou, "Obesity and metabolic syndrome as related to cardiovascular disease," *Expert Review* of Cardiovascular Therapy, vol. 10, no. 7, pp. 933–939, 2012.
- [7] S. Milić, D. Lulić, and D. Štimac, "Non-alcoholic fatty liver disease and obesity: biochemical, metabolic and clinical presentations," *World Journal of Gastroenterology*, vol. 20, no. 28, pp. 9330–9337, 2014.
- [8] A. Fernández-Sánchez, E. Madrigal-Santillán, M. Bautista et al., "Inflammation, oxidative stress, and obesity," *International Journal of Molecular Sciences*, vol. 12, no. 5, pp. 3117– 3132, 2011.
- [9] M. H. Fonseca-Alaniz, J. Takada, M. I. C. Alonso-Vale, and F. B. Lima, "O tecido adiposo como órgão endócrino: da teoria à prática," *Jornal de Pediatria*, vol. 83, no. 5, pp. S192–S203, 2007.
- [10] P. Dludla, B. Nkambule, B. Jack et al., "Inflammation and oxidative stress in an obese state and the protective effects of gallic acid," *Nutrients*, vol. 11, no. 1, p. 23, 2019.

- [11] G. Bjørklund and S. Chirumbolo, "Role of oxidative stress and antioxidants in daily nutrition and human health," *Nutrition*, vol. 33, pp. 311–321, 2017.
- [12] T. Ohishi, R. Fukutomi, Y. Shoji, S. Goto, and M. Isemura, "The beneficial effects of principal polyphenols from green tea, coffee, wine, and curry on obesity," *Molecules*, vol. 26, no. 2, p. 453, 2021.
- [13] M. Szulinska, M. Gibas-Dorna, E. Miller-Kasprzak et al., "Spirulina maxima improves insulin sensitivity, lipid profile, and total antioxidant status in obese patients with welltreated hypertension: a randomized double-blind placebocontrolled study," *European Review for Medical and Pharmacological Sciences*, vol. 21, no. 10, pp. 2473–2481, 2017.
- [14] S. Hosseinpour-Arjmand, F. Amirkhizi, and M. Ebrahimi-Mameghani, "The effect of alpha-lipoic acid on inflammatory markers and body composition in obese patients with nonalcoholic fatty liver disease: a randomized, double-blind, placebo-controlled trial," *Journal of Clinical Pharmacy and Therapeutics*, vol. 44, no. 2, pp. 258–267, 2019.
- [15] E. T. Callcott, C. L. Blanchard, P. Snell, and A. B. Santhakumar, "The anti-inflammatory and antioxidant effects of pigmented rice consumption in an obese cohort," *Food & Function*, vol. 10, no. 12, pp. 8016–8025, 2019.
- [16] Y. Zhao, B. Chen, J. Shen et al., "The beneficial effects of quercetin, curcumin, and resveratrol in obesity," Oxidative Medicine and Cellular Longevity, vol. 2017, Article ID 1459497, 8 pages, 2017.
- [17] M. G. Showell, R. Mackenzie-Proctor, V. Jordan, R. J. Hart, and Cochrane Gynaecology and Fertility Group, "Antioxidants for female subfertility," *Cochrane Database of Systematic Reviews*, vol. 7, no. 7, article Cd007807, 2017.
- [18] M. R. Emami, S. Jamshidi, M. Zarezadeh et al., "Can vitamin E supplementation affect obesity indices? A systematic review and meta-analysis of twenty-four randomized controlled trials," *Clinical Nutrition*, vol. 40, no. 5, pp. 3201–3209, 2021.
- [19] G. Ekhlasi, M. Zarrati, S. Agah et al., "Effects of symbiotic and vitamin E supplementation on blood pressure, nitric oxide and inflammatory factors in non-alcoholic fatty liver disease," *EXCLI Journal*, vol. 16, pp. 278–290, 2017.
- [20] Z. Shadman, F. A. Taleban, N. Saadat, and M. Hedayati, "Effect of conjugated linoleic acid and vitamin E on glycemic control, body composition, and inflammatory markers in overweight type 2 diabetics," *Journal of Diabetes and Metabolic Disorders*, vol. 12, no. 1, p. 42, 2013.
- [21] M. G. Showell, R. Mackenzie-Proctor, V. Jordan, and R. J. Hart, "Antioxidants for female subfertility," *Cochrane Database of Systematic Reviews*, vol. 2020, no. 11, article CD007807, 2020.
- [22] R. M. Smits, R. Mackenzie-Proctor, A. Yazdani et al., "Antioxidants for male subfertility," *Cochrane Database of Systematic Reviews*, vol. 2019, no. 3, 2019.
- [23] M.-E. Piché, A. Tchernof, and J.-P. Després, "Obesity phenotypes, diabetes, and cardiovascular diseases," *Circulation Research*, vol. 126, no. 11, pp. 1477–1500, 2020.
- [24] K. Leisegang, P. Sengupta, A. Agarwal, and R. Henkel, "Obesity and male infertility: mechanisms and management," *Andrologia*, vol. 53, no. 1, article e13617, 2021.
- [25] S. Park, M. Choi, and M. Lee, "Effects of anthocyanin supplementation on reduction of obesity criteria: a systematic review and meta-analysis of randomized controlled trials," *Nutrients*, vol. 13, no. 6, p. 2121, 2021.

- [26] A. M. Dostal, H. Samavat, L. Espejo, A. Y. Arikawa, N. R. Stendell-Hollis, and M. S. Kurzer, "Green tea extract and catechol-O-methyltransferase genotype modify fasting serum insulin and plasma adiponectin concentrations in a randomized controlled trial of overweight and obese postmenopausal women," *The Journal of Nutrition*, vol. 146, no. 1, pp. 38–45, 2016.
- [27] Y. Zhang, R. Proenca, M. Maffei, M. Barone, L. Leopold, and J. M. Friedman, "Positional cloning of the mouse obese gene and its human homologue," *Nature*, vol. 372, no. 6505, pp. 425–432, 1994.
- [28] O. M. Farr, A. Gavrieli, and C. S. Mantzoros, "Leptin applications in 2015: what have we learned about leptin and obesity?," *Current Opinion in Endocrinology, Diabetes, and Obesity*, vol. 22, no. 5, pp. 353–359, 2015.
- [29] A. Yadav, M. A. Kataria, V. Saini, and A. Yadav, "Role of leptin and adiponectin in insulin resistance," *Clinica Chimica Acta*, vol. 417, pp. 80–84, 2013.
- [30] G. Balsan, L. C. Pellanda, G. Sausen et al., "Effect of yerba mate and green tea on paraoxonase and leptin levels in patients affected by overweight or obesity and dyslipidemia: a randomized clinical trial," *Nutrition Journal*, vol. 18, no. 1, p. 5, 2019.
- [31] R. Tabrizi, O. R. Tamtaji, K. B. Lankarani et al., "The effects of resveratrol intake on weight loss: a systematic review and meta-analysis of randomized controlled trials," *Critical Reviews in Food Science and Nutrition*, vol. 60, no. 3, pp. 375–390, 2020.
- [32] M. Rondanelli, N. Miraglia, P. Putignano et al., "Effects of 60-day Saccharomyces boulardii and superoxide dismutase supplementation on body composition, hunger sensation, pro/antioxidant ratio, inflammation and hormonal lipometabolic biomarkers in obese adults: a double-blind, placebo-controlled trial," *Nutrients*, vol. 13, no. 8, p. 2512, 2021.
- [33] M. S. Ellulu, A. Rahmat, I. Patimah, H. Khaza'ai, and Y. Abed, "Effect of vitamin C on inflammation and metabolic markers in hypertensive and/or diabetic obese adults: a randomized controlled trial," *Drug Design, Development and Therapy*, vol. 9, pp. 3405–3412, 2015.
- [34] J. Vekic, A. Zeljkovic, A. Stefanovic, Z. Jelic-Ivanovic, and V. Spasojevic-Kalimanovska, "Obesity and dyslipidemia," *Metabolism*, vol. 92, pp. 71–81, 2019.
- [35] J. P. Després, S. Moorjani, P. J. Lupien, A. Tremblay, A. Nadeau, and C. Bouchard, "Regional distribution of body fat, plasma lipoproteins, and cardiovascular disease," *Arterio-sclerosis*, vol. 10, no. 4, pp. 497–511, 1990.
- [36] B. Klop, J. Elte, and M. Cabezas, "Dyslipidemia in obesity: mechanisms and potential targets," *Nutrients*, vol. 5, no. 4, pp. 1218–1240, 2013.
- [37] Y. Wu, H. Sun, R. Yi, F. Tan, and X. Zhao, "Anti-obesity effect of Liupao tea extract by modulating lipid metabolism and oxidative stress in high-fat-diet-induced obese mice," *Journal of Food Science*, vol. 86, no. 1, pp. 215–227, 2021.
- [38] A. F. G. Cicero, F. Fogacci, M. Bove, M. Giovannini, and C. Borghi, "Three-arm, placebo-controlled, randomized clinical trial evaluating the metabolic effect of a combined nutraceutical containing a bergamot standardized flavonoid extract in dyslipidemic overweight subjects," *Phytotherapy Research*, vol. 33, no. 8, pp. 2094–2101, 2019.
- [39] P. Bogdanski, J. Suliburska, M. Szulinska, M. Stepien, D. Pupek-Musialik, and A. Jablecka, "Green tea extract reduces blood pressure, inflammatory biomarkers, and oxidative stress

and improves parameters associated with insulin resistance in obese, hypertensive patients," *Nutrition Research*, vol. 32, no. 6, pp. 421–427, 2012.

- [40] S. H. Kamali, A. R. Khalaj, S. Hasani-Ranjbar et al., "Efficacy of 'Itrifal Saghir', a combination of three medicinal plants in the treatment of obesity; a randomized controlled trial," *DARU Journal of Pharmaceutical Sciences*, vol. 20, no. 1, 2012.
- [41] C. Patel, H. Ghanim, S. Ravishankar et al., "Prolonged reactive oxygen species generation and nuclear factor-kappaB activation after a high-fat, high-carbohydrate meal in the obese," *The Journal of Clinical Endocrinology and Metabolism*, vol. 92, no. 11, pp. 4476–4479, 2007.
- [42] M. T. Adnan, M. N. Amin, M. G. Uddin et al., "Increased concentration of serum MDA, decreased antioxidants and altered trace elements and macro-minerals are linked to obesity among Bangladeshi population," *Diabetes and Metabolic Syndrome: Clinical Research and Reviews*, vol. 13, no. 2, pp. 933– 938, 2019.
- [43] P. Wu, F. Zhang, Y. Dai, L. Han, and S. Chen, "Serum TNF-α, GTH and MDA of high-fat diet-induced obesity and obesity resistant rats," *Saudi Pharmaceutical Journal*, vol. 24, no. 3, pp. 333–336, 2016.
- [44] B. H. Halima, G. Sonia, K. Sarra, B. J. Houda, B. S. Fethi, and A. Abdallah, "Apple cider vinegar attenuates oxidative stress and reduces the risk of obesity in high-fat-fed male Wistar rats," *Journal of Medicinal Food*, vol. 21, no. 1, pp. 70–80, 2018.
- [45] P. Laharrague, A. M. Fontanilles, J. Tkaczuk, J. X. Corberand, L. Pénicaud, and L. Casteilla, "Inflammatory/haematopoietic cytokine production by human bone marrow adipocytes," *European Cytokine Network*, vol. 11, no. 4, pp. 634–639, 2000.
- [46] A. P. Snider and J. R. Wood, "Obesity induces ovarian inflammation and reduces oocyte quality," *Reproduction*, vol. 158, no. 3, pp. R79–R90, 2019.
- [47] R. Kolb, F. S. Sutterwala, and W. Zhang, "Obesity and cancer: inflammation bridges the two," *Current Opinion in Pharmacology*, vol. 29, pp. 77–89, 2016.
- [48] Z. D. Goodman, "The impact of obesity on liver histology," *Clinics in Liver Disease*, vol. 18, no. 1, pp. 33–40, 2014.
- [49] C. Y. Lian, Z. Z. Zhai, Z. F. Li, and L. Wang, "High fat diettriggered non-alcoholic fatty liver disease: a review of proposed mechanisms," *Chemico-Biological Interactions*, vol. 330, article 109199, 2020.
- [50] E. S. Lee, M. H. Kwon, H. M. Kim, H. B. Woo, C. M. Ahn, and C. H. Chung, "Curcumin analog CUR5-8 ameliorates nonalcoholic fatty liver disease in mice with high-fat diet-induced obesity," *Metabolism*, vol. 103, article 154015, 2020.
- [51] G. Lazzarino, I. Listorti, G. Bilotta et al., "Water- and fatsoluble antioxidants in human seminal plasma and serum of fertile males," *Antioxidants (Basel, Switzerland)*, vol. 8, no. 4, p. 96, 2019.
- [52] F. Lombardo, A. Sansone, F. Romanelli, D. Paoli, L. Gandini, and A. Lenzi, "The role of antioxidant therapy in the treatment of male infertility: an overview," *Asian Journal of Andrology*, vol. 13, no. 5, pp. 690–697, 2011.