

Digital phenotyping from wearables using AI characterizes psychiatric disorders and identifies genetic associations

Jason J. Liu^{1,2,*}, Beatrice Borsari^{1,2,*}, Yunyang Li³, Susanna Liu^{1,2}, Yuan Gao⁴, Xin Xin^{1,2}, Shaoke Lou^{1,2}, Matthew Jensen^{1,2}, Diego Garrido-Martín⁵, Terril Verplaetse⁶, Garrett Ash^{7,8,9}, Jing Zhang¹⁰, Matthew J. Girgenti⁶, Walter Roberts^{6,9,#}, Mark Gerstein^{1,2,3,9,11,#}

¹ Program in Computational Biology and Bioinformatics, Yale University, New Haven, CT, 06511, USA

² Department of Molecular Biophysics and Biochemistry, Yale University, New Haven, CT, 06511, USA

³ Department of Computer Science, Yale University, New Haven, CT, 06511, USA

⁴ Yale School of Public Health, Division of Health Informatics, Yale University, New Haven, CT, 06511, USA

⁵ Department of Genetics, Microbiology and Statistics, Universitat de Barcelona (UB), Barcelona, 08028, Spain

⁶ Department of Psychiatry, Yale University School of Medicine, New Haven, CT, 06511, USA

⁷ Section of General Internal Medicine, Yale University School of Medicine, New Haven, CT, 06511, USA

⁸ Center for Pain, Research, Informatics, Medical Comorbidities and Education Center (PRIME), VA Connecticut Healthcare System, West Haven, CT, 06516, USA

⁹ Department of Biomedical Informatics and Data Science, Yale University, New Haven, CT, 06511, USA

¹⁰ Department of Computer Science, University of California, Irvine, CA, 92697, USA

¹¹ Department of Statistics and Data Science, Yale University, New Haven, CT, 06511, USA

* Co-first authors

Co-corresponding authors (pi@gersteinlab.org)

Abstract

Psychiatric disorders are complex and influenced by both genetic and environmental factors. However, studying the full spectrum of these disorders is hindered by practical limitations on measuring human behavior. This highlights the need for novel technologies that can measure behavioral changes at an intermediate level between diagnosis and genotype. Wearable devices are a promising tool in precision medicine, since they can record physiological measurements over time in response to environmental stimuli and do so at low cost and minimal invasiveness. Here we analyzed wearable and genetic data from a cohort of the Adolescent Brain Cognitive Development study. We generated >250 wearable-derived features and used them as intermediate phenotypes in an interpretable AI modeling framework to assign risk scores and classify adolescents with psychiatric disorders. Our model identifies key physiological processes and leverages their temporal patterns to achieve a higher performance than has been previously possible. To investigate how these physiological processes relate to the underlying genetic architecture of psychiatric disorders, we also utilized these intermediate phenotypes in univariate and multivariate GWAS. We identified a total of 29 significant genetic loci and 52 psychiatric-associated genes, including *ELFNI* and *ADORA3*. These results show that wearable-derived continuous features enable a more precise representation of psychiatric disorders and exhibit greater detection power compared to categorical diagnostic labels. In summary, we demonstrate how consumer wearable technology can facilitate dimensional approaches in precision psychiatry and uncover etiological linkages between behavior and genetics.

45 Introduction

46 Psychiatric disorders of childhood and adolescence currently affect 1 in 7 youths in the United
47 States and globally^{1,2}. Externalizing disorders such as attention-deficit/hyperactivity disorder, and
48 internalizing disorders such as anxiety, are among the most prevalent and represent a wide
49 spectrum of dysfunctional behavior patterns³. Treatment barriers are complex and multifaceted but
50 major contributors include our limited understanding of psychiatric phenotypes and difficulty
51 identifying youth individuals that experience these disorders.

52
53 Traditionally, psychiatric disorders have been conceptualized as categorical macrophenotypes,
54 based on clinical manifestations of a disease which are defined according to the number and type
55 of symptoms, and the presence of distress or impairment⁴⁻⁶. While this has practical benefits in
56 terms of reliability and ease of diagnosis, it poses several challenges to the research of these
57 disorders, and consequently to the development of treatments. In fact, psychiatric disorders are
58 complex and often comorbid, and this high degree of heterogeneity is not always accurately
59 translated into categorical diagnosis labels, which may be defined by arbitrary cut-offs. Instead,
60 intermediate phenotypes (i.e., quantitative traits that are positioned between genotype and
61 macrophenotype) may better capture the heterogeneity potentially missed by existing diagnostic
62 categories⁷⁻⁹. Additionally, genetic penetrance is expected to be higher for these intermediate
63 phenotypes compared to macrophenotypes, enabling improved dissection of the genetic
64 architecture underlying psychiatric disorders¹⁰. Nevertheless, many genome-wide association
65 studies (GWAS) aimed at identifying genetic variants or biomarkers for psychiatric disorders do
66 not consider these intermediate phenotypes and instead rely on dichotomised (i.e., binary) traits.
67 In fact, identifying intermediate phenotypes with clinical and biological relevance remains a
68 challenge¹¹.

69
70 Therefore, to improve our understanding of psychiatric disorders it is important that we identify
71 intermediate phenotypes that not only offer a more comprehensive representation of an
72 individual's behavior with respect to their environment, but also relate well with existing clinical
73 definitions and aid in diagnosis. Once identified, these intermediate phenotypes can then be also
74 used to guide more comprehensive studies to identify genetic associations and biomarkers that
75 may ultimately improve precision treatments.

76
77 To achieve this goal, it is important to leverage new emerging technologies that can quantitatively
78 assess an individual's behavioral patterns¹². Wearable sensors such as smartwatches collect data
79 that reflect physical and physiological processes (e.g., movement, pulse, metabolic intake), and
80 can be used to infer higher-order behavioral events (e.g., sleep, exercise) and their temporal
81 dynamics. Because of the documented relationship between such higher-order behavioral events
82 and mental health, and given their low cost and minimal invasiveness, wearable devices have
83 emerged as promising tools for mental health monitoring and psychiatric evaluation¹³⁻¹⁵.

84
85 Therefore, wearable sensors have the potential for capturing intermediate phenotypes relevant to
86 behavior and psychiatric disorders, ultimately enabling improved GWAS. However, significant
87 computational challenges remain in generating intermediate phenotypes from wearable-derived
88 data that describe the full spectrum of a given psychiatric disorder. Moreover, further curation of
89 these intermediate phenotypes is necessary to identify genetic associations that have clinical and
90 biological relevance.

91 To address these limitations, we developed an AI modeling framework that flexibly leverages data
92 from wearable devices to generate intermediate phenotypes in the form of static and dynamic
93 digital features. We establish the validity of these digital features as intermediate phenotypes by
94 classifying externalizing and internalizing disorders with an accuracy beyond baseline expectation,
95 and even surpassing the performance of some other gold-standard intermediate phenotypes such
96 as fMRI measurements¹⁶⁻¹⁹. Interpretability modules in our AI framework enable us to identify
97 key temporal and physiological insights between clinical diagnosis and digital features, further
98 supporting the validity of using these wearable-derived features as intermediate phenotypes. We
99 further curate these intermediate phenotypes and employ them in GWAS models to identify
100 genetic associations and biomarkers that capture the continuous spectrum of psychiatric disorders
101 and behavioral patterns. Finally, we identify 29 significant loci, several of which overlap
102 previously reported genetic variants associated with behavioral traits and mental illnesses and are
103 proximal to genes with a documented role in neurodevelopmental and psychiatric disorders.

104

105 In sum, this work shows how wearable devices can advance our understanding of psychiatric
106 disorders by establishing a more objective and dimensional approach that can ultimately lead to
107 improved treatments in precision psychiatry.

108 **Results**

109 **Leveraging the Adolescent Brain Cognitive Development cohort**

110 To improve our understanding of psychiatric disorders, we leveraged and analyzed a dataset from
111 a cohort of US adolescents recruited by the NIH Adolescent Brain Cognitive Development
112 Consortium (ABCD) project, consisting of clinical, wearable, and genetic data (**Fig. 1**). The ABCD
113 cohort consists of a total of 11,878 adolescents (5682 males and 6196 females), of age between
114 nine and fourteen years and belonging to four different ethnicities (**Suppl. Fig. S4.1**). We
115 identified nine categories of psychiatric phenotypes (**Suppl. Table S1.1**), which were established
116 using a gold standard parent diagnostic semi-structured interview (Kiddie Schedule for Affective
117 Disorders and Schizophrenia-5)²⁰. The healthy controls represented adolescents who did not meet
118 the criteria for any of those nine psychiatric disorders. We defined these clinical labels as the
119 categorical macrophenotypes in the study (**Fig. 1A-B**). Our modeling framework also utilized data
120 from cognitive tests (e.g., NIH Toolbox) and behavioral checklists (**Fig. 2A, Suppl. Fig. S2.1**).

121

122 **Generating intermediate phenotypes from wearable-derived data**

123 We processed data obtained from FitBit smartwatches, which comprise measurements of heart
124 rate, calories, activity intensity, steps, metabolic equivalents, sleep level and sleep intensity (**Fig.**
125 **1C, Suppl. Table S1.2**)²¹. These measurements quantify an individual's physiological processes
126 and their real-time changes in response to environmental stimuli, and can thus provide key
127 information about an individual's behavior.

128

129 To reconstruct the full spectrum of an individual's behavioral functioning from these data, we
130 applied two different feature engineering techniques, allowing us to generate wearable-derived
131 dynamic and static features, which we consider as intermediate phenotypes. The dynamic features
132 preserve the time-varying nature of the original data as a time series, enabling sequential and
133 temporal patterns of the data to be retained. In contrast, the static features summarize patterns of

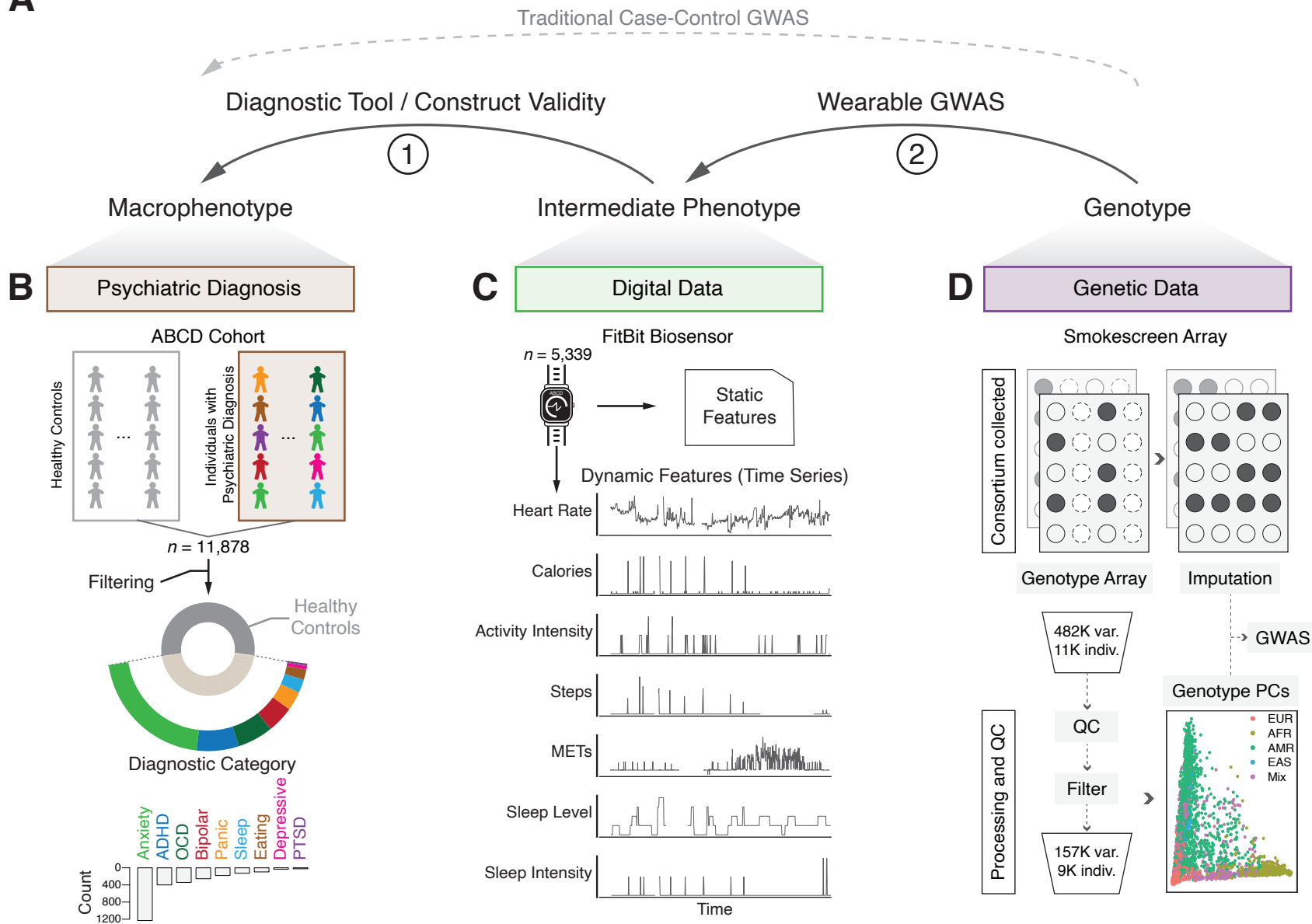
A

Figure 1. Leveraging clinical, digital, and genetic data of the ABCD cohort to improve characterization of psychiatric disorders.

A) Framework schematic describing how intermediate phenotypes from wearable-derived data are leveraged to better understand the association between macrophenotype and genotype. The link between intermediate phenotype and macrophenotype serves as construct validity and aid in diagnostics. Wearable GWAS is performed through genotype-to-intermediate-phenotype association studies. **B)** The Adolescent Brain Cognitive Development (ABCD) cohort contains 11,878 individuals spanning nine different categorical macrophenotypes based on clinical diagnosis from the Kiddie Schedule for Affective Disorders and Schizophrenia-5. A breakdown of the counts of each disorder is shown in the bottom bar graph, with anxiety disorder and ADHD being the most prevalent. “Bipolar” refers to bipolar or psychotic disorders. **C)** Digital data from FitBit biosensors are collected for 5,339 individuals. The collected time series data are then processed into dynamic and static features, with information spanning various physiological and higher order processes. **D)** Genetic data are collected by the ABCD consortium through Smokescreen genotyping array. Imputed genotypes are used for downstream GWAS analyses. The genotype arrays are subjected to best-practice processing and QC to ensure included individuals and SNPs are of high quality. PCA performed on 8,791 individuals and 157,556 genotyped SNPs reveals distinct ancestral clusters across the cohort and the inferred genotype principal components (PCs) are used as covariates in downstream analyses.

134 the digital data to produce time-invariant, quantitative features that are commonly used in
135 downstream modeling^{14,22}.

136
137 To generate dynamic features, we performed signal imputation and processing after filtering the
138 individuals with sparse data, and obtained 48 channels of time series (**Figure 2B, Suppl. Fig.**
139 **S2.3**). Compared to the static features, this further processing allowed us to preserve both local
140 and global temporal patterns potentially relevant to characterizing behavior and neurological
141 response to stimuli^{8,23}.

142
143 To generate static features, we first collected a total of 49 FitBit summary-based features (**Suppl.**
144 **Table S2.1a-b**). We next applied descriptive statistics (e.g. mean, median, etc.) to each of these
145 features and generated a total of 258 static features for each individual (**Fig. 2C, Suppl. Table**
146 **S2.2**)^{14,24}. We then grouped these static features into seven main clusters, each of which
147 summarizes different aspects of physiological and behavioral processes, such as heart rate, sleep
148 duration and quality, metabolic intake or physical activity (**Fig. 2D**).

149
150 Altogether, static and dynamic features represent the physiological and behavioral profiles of the
151 adolescents, and can be leveraged as intermediate phenotypes in a wide range of analyses to better
152 characterize psychiatric phenotypes, such as generating disorder-specific probability risk scores,
153 macrophenotype classification, model interpretability, and biomarker identification via wearable
154 GWAS.

155 156 **Predicting psychiatric phenotypes from wearable-derived intermediate phenotypes**

157 To demonstrate the validity of static and dynamic features as clinically relevant intermediate
158 phenotypes and to evaluate their utility as a diagnostic tool, we employed these features in an array
159 of classification tasks to identify individuals with either an externalizing (ADHD) or internalizing
160 (anxiety) disorder from their typically developing peers. We selected ADHD and anxiety due to
161 their high prevalence in adolescents, which is mirrored in the cohort (**Fig. 1B**)²⁵.

162
163 We applied a gradient boosting machine learning algorithm, XGBoost, for classification tasks
164 using static features (**Fig. 2C-D**)²⁶. On the other hand, to fully leverage the time series nature of
165 the dynamic features, we used a convolutional neural network for time series, featuring depthwise
166 separable convolution, called Xception (**Fig. 2E**)²⁷. Variable convolutional filters and residual
167 (skip) connections, coupled with efficient parametrization, allow information encoded in both
168 small and large receptive fields to be more optimally leveraged. Practically, this framework takes
169 into account local and global patterns of physiology and behavior when performing downstream
170 classification of psychiatric disorders. In both modeling approaches we included covariates that
171 accounted for demographic features, family history of disorders, and other clinical information
172 (**Fig. 2A, Suppl. Table S1.2**). To assess the benefit, in terms of model performance, of including
173 wearable-derived data, we also trained a baseline model using just the covariates, which served as
174 a comparison to the models including static or dynamic features. In practice, this comparison
175 allowed us to determine whether wearable-derived features can improve diagnostic accuracy
176 relative to that achievable using only a widely used broadband behavior rating scale.

177
178 After data filtering, we first used static features to classify 216 individuals with ADHD (an
179 externalizing disorder) versus 1,737 of their typically developing peers (healthy controls) (**Fig. 3A**

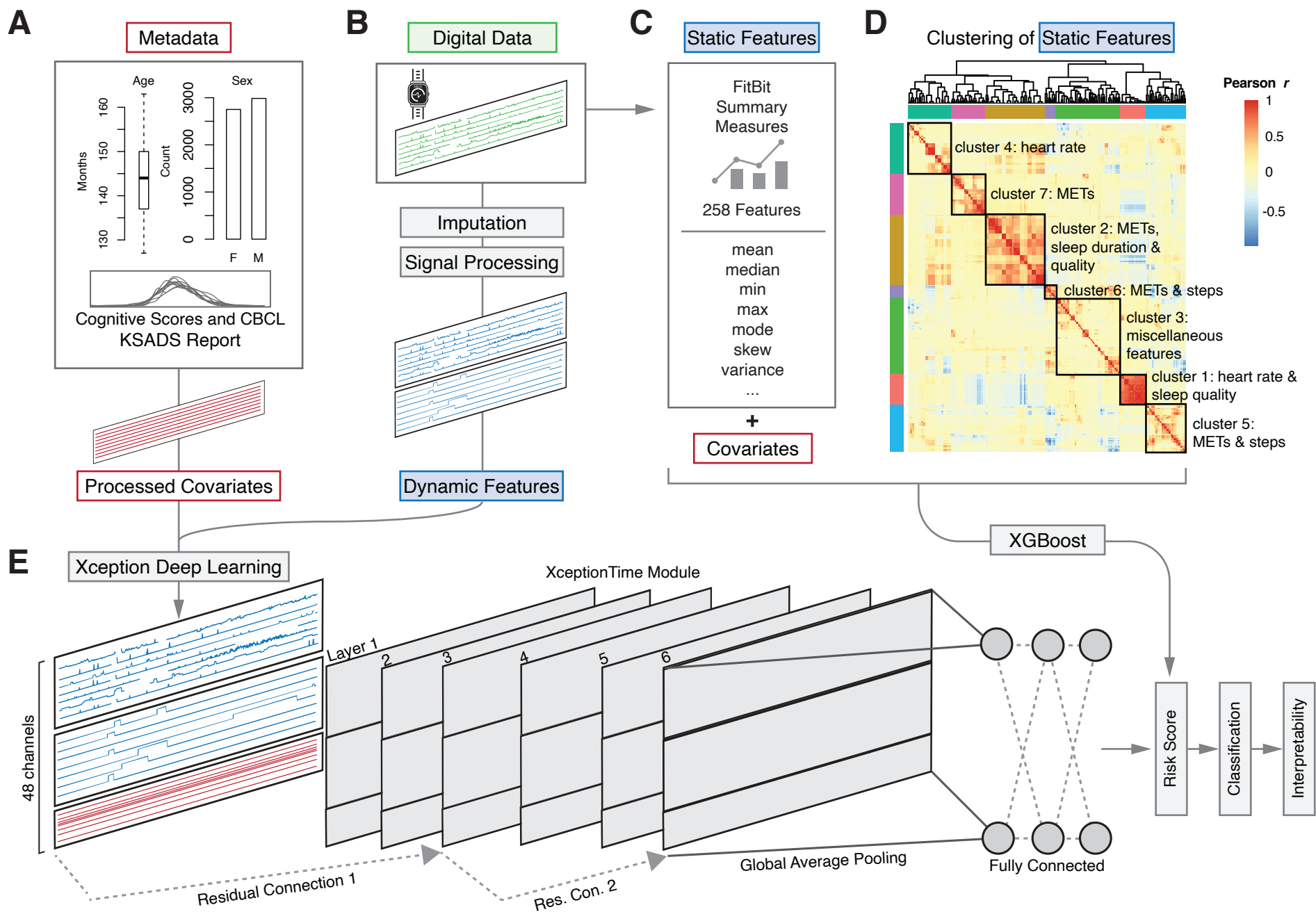


Figure 2. Workflow for data processing, feature engineering, and model architecture.

A) ABCD cohort metadata including various demographic features, cognitive test scores, and clinical characteristics are used as covariates and represent the input features used in our baseline comparison model. Features shown in this plot correspond to the filtered set of individuals with wearable data. **B)** Digital data collected by wearable biosensors are used to generate dynamic features after signal processing and imputation steps. Together with the processed covariates, these time series features represent the input features for the dynamic model. **C)** Summary statistics applied to digital data collected by wearables are used to generate a total of 258 static features. In addition to the covariates, these are the input features used in the static model. The static model leverages the machine learning framework, XGBoost, for downstream tasks such as risk score generation, classification, model interpretability, and wearable GWAS. **D)** Hierarchical clustering of the static features yields seven distinct physiological clusters of wearable data. **E)** The dynamic model is based on the Xception deep learning framework, and uses the generated 48 channels from the dynamic features and covariates as input into a convolution-like model. The architecture consists of six inception layers and residual connections. Global average pooling and a fully connected layer allow for similar downstream tasks as mentioned in C).

180 **and Suppl. Fig. 3.1).** Using static features with XGBoost, we achieved an average area under the
181 receiver operating characteristic curve (AUROC) of 0.87 and precision of 0.79. When using the
182 dynamic features and Xception, we were able to achieve an average AUROC of 0.89 and precision
183 of 0.83. The baseline model consisting of only the covariates achieved an average AUROC of
184 0.83, suggesting that the inclusion of wearable-derived features facilitates a clinically meaningful
185 improvement in diagnostic accuracy. This improvement between the baseline and dynamic
186 features model demonstrates statistical significance (one-sided t-test between baseline and
187 dynamic features model, p value = 0.0022).

188
189 Second, we evaluated the performance of our model using static or dynamic features in the
190 classification of 666 individuals diagnosed with anxiety disorder (internalizing disorder) versus
191 1,737 of their typically developing peers (healthy controls) (**Fig. 3B and Suppl. Fig. 3.2**). Here,
192 we again repeated the use of the same modeling framework, i.e., static features with XGBoost and
193 dynamic features with Xception, and compared it to the baseline covariate model. We found that
194 static and dynamic features achieve an average AUROC of 0.69 and 0.71 and precision of 0.64
195 and 0.68, respectively. In both models, the performance was greater than that of the baseline model
196 (average AUROC of 0.67), with the dynamic features model showing the most significant
197 performance improvement (one-sided t-test between baseline and dynamic feature model, p value
198 = 0.00016). Overall, the fact that the models using dynamic features achieved the highest
199 performance suggests the usefulness of the temporal patterns intrinsic to wearable-derived data
200 towards understanding behavior.

201 202 **Interpreting wearable features prioritized by the deep learning model**

203 Deep learning methods are typically characterized by complex internal structures that cannot be
204 easily interpreted by humans. While maximizing the classification accuracy is one crucial aspect
205 for characterizing complex phenotypes, understanding which features are most important in terms
206 of their individual contribution to performance is also critical. To this end, we utilized ablation
207 techniques to determine the relative contribution of each individual feature to model performance.
208 For the ADHD classification task, heart rate was the most important feature (largest change in
209 AUROC), followed by other dynamic features (i.e., sleep, steps, METs) as well as covariates such
210 as demographics, family history, and cognitive scores from picture memory and stop-signal
211 reaction time tests (**Fig. 3C, Suppl. Fig. S3.3**).

212
213 On the other hand, the ablation study for the anxiety classification task revealed a different set of
214 important features. In this case, sleep quality and stage, calories, and step count were the most
215 important dynamic features, whereas heart rate features, which were extremely important for
216 classifying ADHD, were not prioritized in the anxiety model (**Fig. 3D, Suppl. Fig. S3.5**).
217 Additionally, while the anxiety model prioritized some covariates that were relevant also for the
218 ADHD model (e.g., sex, family history, and family divorce), cognitive scores from tests such as
219 picture memory did not appear to be important for the identification of individuals diagnosed with
220 anxiety, consistent with theory-driven accounts of neurocognitive aspects of anxiety disorders²⁸.

221
222 Additionally, we assessed the importance for model performance of dynamic features at various
223 times throughout the day by adapting gradient-weighted class activation mapping (Grad-CAM)
224 strategies²⁹. We calculated the relative importance of each time point in our dynamic features. For
225 ADHD, we observed enriched significance of the heart rate dynamic feature around the early

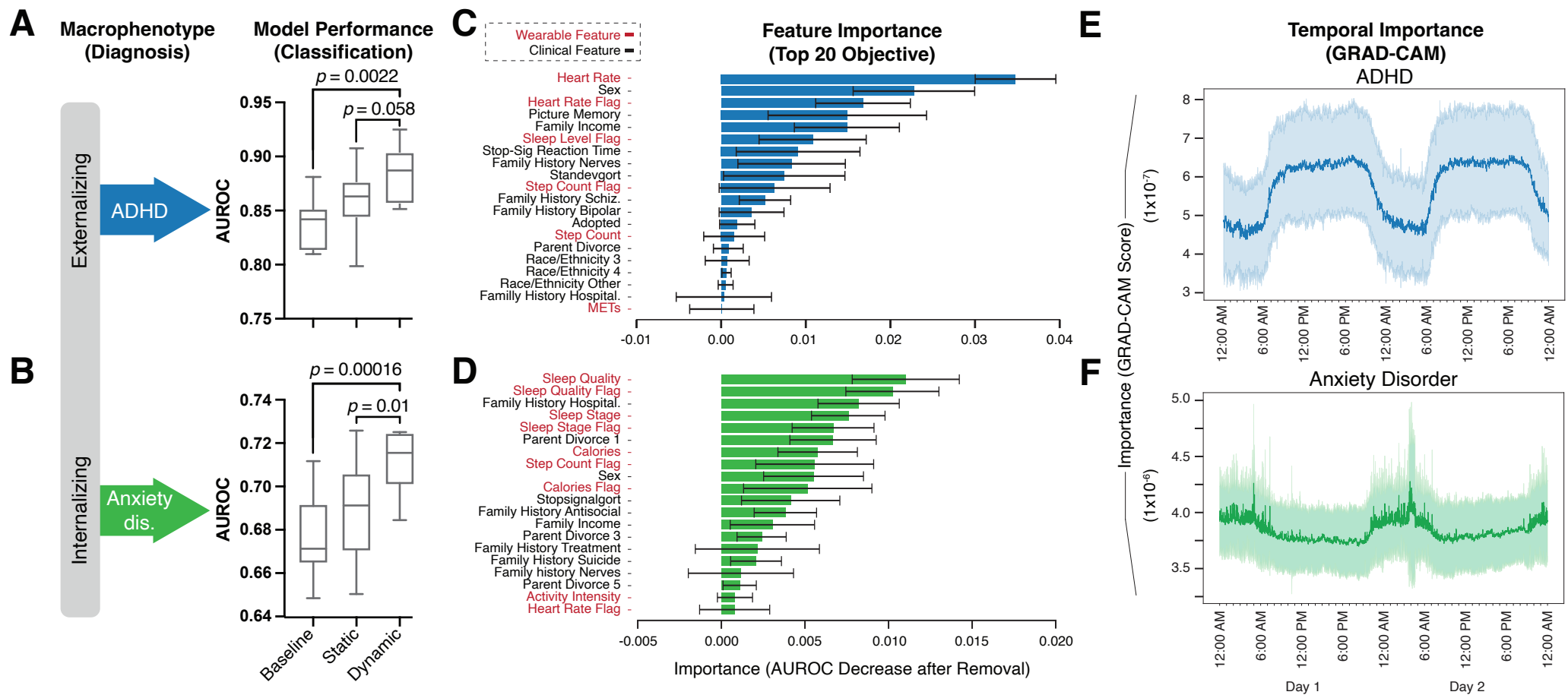


Figure 3. Performance and interpretability of psychiatric phenotype classification models.

A-B) Model performance for baseline, static, and dynamic models employed for classifying individuals with ADHD (blue, top) or individuals with anxiety disorder (green, bottom) versus healthy controls. *P* values were calculated using one-sided t-test. **C-D)** Feature importance based on ablation studies for the dynamic model for ADHD (blue, top) and anxiety disorder (green, bottom) classification. Wearable-derived dynamic features are shown in red font and clinical features (covariates) are shown in black font. Feature importance is equivalent to the decrease in model performance (AUROC) after removal of the given feature. **E-F)** Temporal importance during a 48-hour period for dynamic features in ADHD (blue, top) or anxiety disorder (green, bottom) classification based on the GRAD-CAM interpretability module. Importance is represented as the GRAD-CAM score, based on each time points contribution towards model performance.

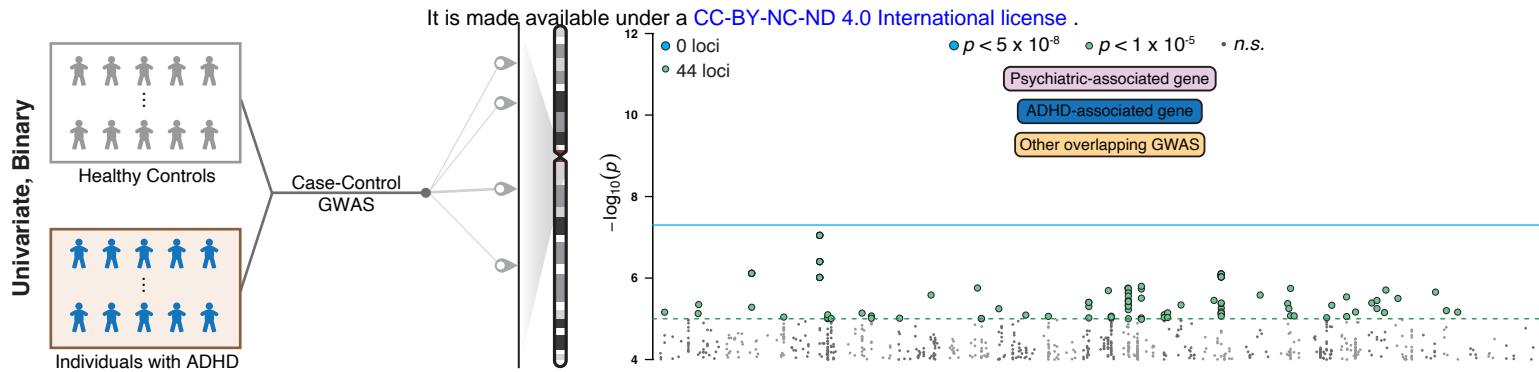
226 afternoon, potentially suggesting stronger behavioral differences between adolescents with ADHD
227 and their typically developing peers (healthy controls) during this time of day (**Fig. 3E, Suppl.**
228 **Fig. S3.4**). This is consistent with clinical research demonstrating time-of-day effects on ADHD
229 symptom expression³⁰. In contrast, sleep-related dynamic features during the night are much more
230 informative in classifying anxiety, consistent with clinical expectations (**Fig. 3F, Suppl. Fig.**
231 **S3.6**)³¹. Together, the ablation studies suggest a role for wearable-derived features to not only
232 serve as quantitative intermediate phenotypes, but also to more closely reveal insights into the
233 behavioral and physiological temporal patterns related to categorical macrophenotypes.

234 235 **Using wearable-derived features as intermediate phenotypes for wearable GWAS**

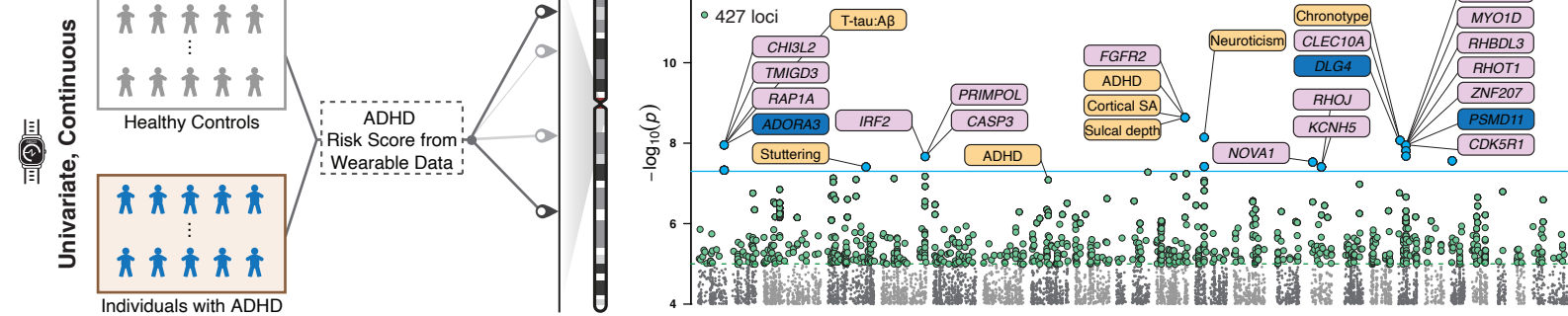
236 Our accurate classification of both internalizing and externalizing psychiatric phenotypes based
237 on wearable-derived features suggests that these features can serve as useful intermediate
238 phenotypes and may be leveraged to identify genetic associations with psychiatric conditions (**Fig.**
239 **4, Suppl. Fig. S4.1-S4.6**). To this end, we first focused specifically on ADHD, given the higher
240 predictive power observed with our models (**Fig. 3A-B**) and its higher estimated heritability
241 compared to anxiety (77-88% vs. 30-60%)^{32,33}. We selected 1,191 individuals (137 individuals
242 with ADHD and 1,054 healthy control individuals) with genetic and wearable data available, and
243 performed a GWAS using the continuous prediction scores obtained from our wearable modeling
244 framework. In practice, these scores represent risk probabilities for ADHD^{34,35}. We identified 10
245 genome-wide (p value $< 5 \cdot 10^{-8}$) significant loci and 21 psychiatric or brain-related genes (**Fig. 4B,**
246 **Table 1, Suppl. Fig. S4.7-S4.9, Suppl. Table S4.1**). Three of the identified genes (*ADORA3,*
247 *PSMD11* and *DLG4*) have been previously associated with ADHD, bolstering the overall
248 functional significance of the results³⁶⁻³⁸. Furthermore, several of these loci overlap with
249 previously reported GWAS SNPs related to ADHD, neuroticism, sleep disruption and other
250 clinically relevant traits (**Fig. 4B, Suppl. Fig. S4.10, Suppl. Table S4.2**). Note that here we used
251 a continuous risk score as a univariate response variable for the GWAS. In comparison, when
252 performing a traditional case-control GWAS for ADHD on the same set of individuals using the
253 binary diagnostic label (presence/absence of disorder) as response variable, we did not identify
254 any significant loci (**Fig. 1A, Fig. 4A, Suppl. Fig. S4.7-S4.9**). This result is consistent with the
255 higher statistical power of continuous measurements over dichotomized (i.e., binary) traits (**Suppl.**
256 **Fig. S4.11**), and with the findings that intermediate phenotypes show higher genetic penetrance
257 compared to macrophenotypes^{7,39-41}.

258
259 While the analysis above collapses wearable-derived features into a single continuous variable that
260 summarizes the risk score for a particular disorder (i.e., GWAS with a univariate response), it is
261 also possible to directly use the full set of wearable-derived features as a more comprehensive and
262 richer phenotype to represent the continuum of psychiatric disorders and their latent manifestations
263 (i.e., GWAS with a multivariate response). In fact, these features can collectively capture
264 behavioral patterns by measuring physiological processes and their real-time changes in response
265 to environmental stimuli, and unlike disease risk scores, are not restricted to a specific cohort of
266 individuals^{42,43}. In what follows, we employed a multivariate nonparametric test of association to
267 regress the vector of wearable-derived features on the genotype of each genetic variant, employing
268 a larger cohort that spans healthy controls and individuals with any psychiatric disorder ($n =$
269 $2,410$)⁴⁴. From this novel type of GWAS we identified 19 significant loci and 31 genes with a
270 documented role in neurodevelopmental and psychiatric disorders (**Fig. 4C, Table 1, Suppl. Fig.**
271 **S4.12-S4.14, and Suppl. Table S4.3**). Many of these loci overlap previously identified GWAS

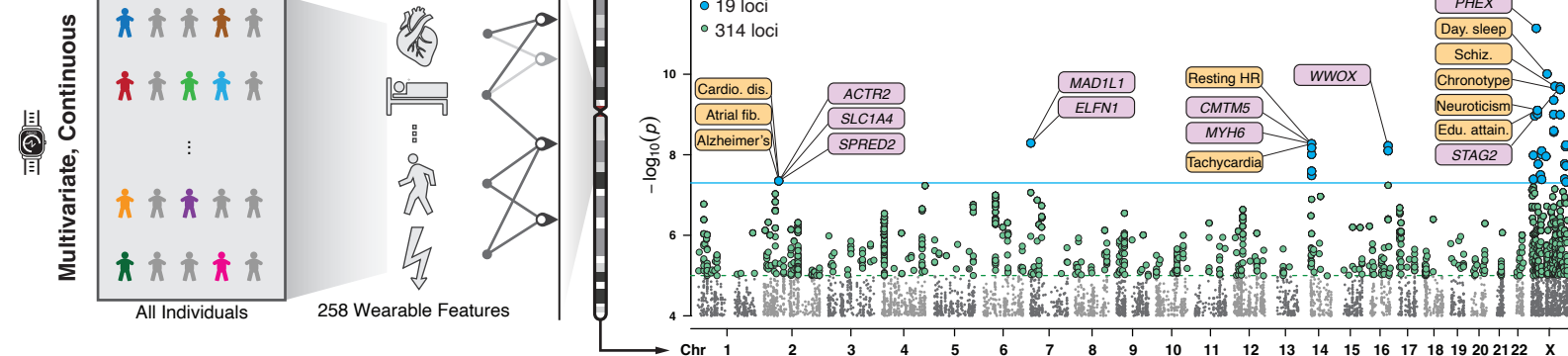
A



B



C



= Wearable GWAS

Figure 4. Manhattan plots summarizing the results of the univariate and multivariate wearable GWAS.

A) Left panel: Case-control GWAS on 1,191 individuals from the ABCD cohort. We employed the clinical diagnosis label as the univariate binary response variable for the GWAS (1 = “individual with ADHD”, $n = 137$; 0 = “healthy control individual”, $n = 1,054$). Right panel: Manhattan plot showing the $-\log_{10} p$ value of association between the genetic variants and the univariate binary response variable. No genetic variants passed the genome-wide significance threshold (p value $< 5 \cdot 10^{-8}$; blue line). Genetic variants with a suggestive p value ($< 10^{-5}$) are represented as green dots.

B) Analogous representation to panel A) using the wearable-derived risk scores for ADHD as univariate continuous response variable. The GWAS was performed on the same set of 1,191 individuals and using the same set of covariates as in panel A). 10 and 427 loci passed the p value thresholds of $5 \cdot 10^{-8}$ and $1 \cdot 10^{-5}$, respectively. A detailed list of genome-wide significant loci is provided in **Table 1** and **Suppl. Table S4.1**. Loci chr1:111,372,165-111,482,359, chr17:7,101,607-7,101,608, and chr17:32,256,997-32,283,356 are proximal to genes *ADORA3* (72 Kb), *DLG4* (86 Kb) and *PSMD11* (174 Kb) (highlighted in dark blue) respectively, which have been previously associated with ADHD. Other proximal genes related to other psychiatric disorders are highlighted in pink (evidence obtained from OpenTargets). Brain-related traits associated with genetic variants overlapping the ten genome-wide significant loci are highlighted in orange. GWAS associations were obtained from the EBI-NHGRI GWAS catalog.

C) Analogous representation to panel A) using the 258 wearable-derived static features as multivariate continuous response variable. The GWAS was performed on a set of 2,410 individuals (both healthy controls and individuals with any disorder). 19 and 314 loci passed the p value thresholds of $5 \cdot 10^{-8}$ and $1 \cdot 10^{-5}$, respectively. A detailed list of genome-wide significant loci is provided in **Table 1** and **Suppl. Table S4.3**. Neuropsychiatric-related genes proximal to the identified loci are shown in pink. Brain-, heart-, and sleep-related traits with associated variants overlapping the 19 loci are highlighted in orange.

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GWAS Method	Locus	Chr	Start	End	Lead Variant	Position	p value	Genes
Univariate continuous	1	1	111,372,165	111,482,359	rs114081965	111,372,166	1.11E-08	<i>TMIGD3, ADORA3,</i> <i>IRF1A, C17orf12</i>
Univariate continuous	2	3	161,873,055	161,927,820	rs79203233	161,909,261	3.89E-08	-
Univariate continuous	3	4	184,417,766	184,424,056	rs1425551	184,421,904	2.15E-08	<i>IRF2, CASP3,</i> <i>PRIMPOL</i>
Univariate continuous	4	10	121,524,611	121,582,200	rs140794722	121,524,612	2.31E-09	<i>FGFR2</i>
Univariate continuous	5	11	38,982,793	39,384,610	rs151239852	39,273,497	3.80E-08	-
Univariate continuous	6	14	26,214,683	26,216,532	rs149074469	26,216,532	2.95E-08	<i>NOVA1</i>
Univariate continuous	7	14	62,903,677	63,014,798	rs143225169	63,014,798	3.89E-08	<i>KCNH5, RHOJ</i>
Univariate continuous	8	17	7,101,607	7,101,608	rs11653054	7,101,608	8.51E-09	<i>CLEC10A, DLG4</i>
Univariate continuous	9	17	32,256,997	32,283,356	rs6505293	32,270,863	1.12E-08	<i>RHBDL3, RHOT1,</i> <i>C17orf75, ZNF207,</i> <i>PSMD11, LRRC37B,</i> <i>CDK5R1, MYO1D</i>
Univariate continuous	10	19	4,495,610	4,495,611	rs150855276	rs150855276	2.75E-08	-
Multivariate continuous	11	2	65,047,524	65,158,021	rs147959551	65,140,366	4.47E-08	<i>RAB1A, ACTR2,</i> <i>SLC1A4, SPRED2</i>
Multivariate continuous	12	7	1,789,321	1,791,353	rs113525298	1,791,353	5.10E-09	<i>MAD1L1, ELFN1,</i> <i>PSMG3, MAFK</i>
Multivariate continuous	13	14	23,392,601	23,418,974	rs365990	23,392,602	5.33E-09	<i>MYH6, CMTM5,</i> <i>IL25, BCL2L2-PABPN1,</i> <i>BCL2L2</i>
Multivariate continuous	14	16	79,283,253	79,302,474	rs8051625	79,288,217	5.98E-09	<i>WWOX</i>
Multivariate continuous	15	X	10,444,818	10,481,837	rs73492938	10,467,098	1.03E-08	<i>MID1, CLCN4</i>
Multivariate continuous	16	X	16,350,347	16,647,557	rs149504239	16,454,383	1.09E-09	-
Multivariate continuous	17	X	22,606,616	22,606,617	-	22,606,617	7.23E-12	<i>PHEX</i>
Multivariate continuous	18	X	27,164,449	28,601,484	rs200021485	27,782,503	7.89E-10	-
Multivariate continuous	19	X	30,757,116	30,896,124	rs150685307	30,831,630	1.70E-08	<i>TAB3, GK</i>
Multivariate continuous	20	X	40,530,738	40,530,739	rs149441354	40,530,739	3.17E-08	<i>ATP6AP2</i>
Multivariate continuous	21	X	45,687,487	45,858,851	-	45,819,973	8.04E-09	-
Multivariate continuous	22	X	62,901,143	64,518,530	rs7883352	63,935,445	1.09E-08	<i>ARHGEF9</i>
Multivariate continuous	23	X	67,188,492	68,693,105	rs180773472	68,693,105	9.82E-11	<i>OPHN1, AR</i>
Multivariate continuous	24	X	96,759,291	96,961,667	-	96,759,292	4.40E-10	-
Multivariate continuous	25	X	102,017,055	103,738,764	-	102,017,056	1.99E-10	<i>RAB40AL, GPRASP2</i>
Multivariate continuous	26	X	124,254,095	124,260,611	rs12012355	124,254,420	2.08E-10	<i>TEX13D, SH2D1A,</i> <i>TENM1, STAG2</i>
Multivariate continuous	27	X	143,176,627	143,212,922	rs12011450	143,190,440	1.69E-08	<i>SPANX4</i>
Multivariate continuous	28	X	146,722,378	146,739,375	rs5966302	146,729,545	6.26E-09	-
Multivariate continuous	29	X	148,716,235	148,915,910	rs57168704	148,907,585	5.71E-09	<i>AFF2</i>

Table 1. Results for the 29 genetic loci identified by the univariate and multivariate continuous wearable GWAS. For each locus we report the GWAS Method (univariate or multivariate continuous) that identifies the locus, the genomic coordinates in human assembly GRCh38, and the lead variant rsID with corresponding genomic position and p value. Brain-related or neuropsychiatric genes proximal to the locus are also listed (Suppl. Tables S4.1 and S4.3).

272 SNPs related to heart, sleep, metabolism, and brain traits (**Fig. 4C, Suppl. Fig. S4.15**). This aligns
273 with the close association between physiological functions, the central nervous system, and
274 individual behavior.

275

276 **Functionally dissecting and interpreting novel wearable GWAS loci**

277 To further investigate the loci identified by the behavioral GWAS, we dissected the variants using
278 a battery of publicly available genomic resources^{45,46}. Many of these loci overlap either GTEx
279 expression quantitative trait loci (eQTLs) or ENCODE candidate cis-regulatory elements (cCREs),
280 suggesting a link between the biochemical activity of these variants and their functional impact on
281 macrophenotype (**Suppl. Table S4.3**). We also explored the impact of these loci beyond
282 behavioral traits and their relationship with clinical psychopathology. For example, behavioral
283 traits significantly associated with a specific genetic variant may correlate with clinical symptoms
284 of a specific psychiatric cohort. Indeed, in some cases we show that the genetic variant in question
285 is also differentially enriched between that specific psychiatric cohort and healthy individuals.

286

287 For instance, we found the minor allele (G) at rs365990 to be significantly associated with an
288 increase in mean heart rate and a decrease in interday heart rate variation (**Fig. 5A, left**). The
289 variant, missense for *MYH6*, had been previously linked to atrial fibrillation, ventricular
290 tachycardia and resting heart rate, and the entire locus shows a significant enrichment of chromatin
291 features in heart samples compared to other tissues and organs (**Suppl. Fig. S5.1**)⁴⁷⁻⁵⁰. We also
292 found the same allele to be enriched in the bipolar/psychotic disorder cohort compared to healthy
293 controls (**Fig. 5A, right**). This cohort included youth meeting criteria for bipolar or unspecified
294 psychotic spectrum disorder, and such severe pathology is known to be associated with
295 characteristic irregularities in heart activity⁵¹⁻⁵³. SNP rs365990 is also a GTEx eQTL for the
296 *CMTM5* gene (**Suppl. Fig. S5.2**), which is highly expressed in brain subregions and has been
297 implicated in stress response and childhood adversity, further supporting the relevance of this locus
298 for psychiatric conditions in addition to heart pathophysiology^{46,54}.

299

300 In a similar fashion, we explored variants rs113525298 and rs147959551. The minor allele at
301 rs113525298 is associated with prolonged periods in bed and shorter vigorously active time during
302 the day, and appears at a lower frequency in the ADHD cohort compared to healthy individuals
303 (**Fig. 5B**). This suggests a potential protective role of the allele against hyperactivity disorders,
304 further supported by the proximity of the SNP to *ELFNI*, previously implicated in the
305 pathophysiology of ADHD^{55,56}. On the other hand, we found the minor allele at rs147959551
306 associated with shorter sedentary time at night and a shorter period of time identified as sleep based
307 on heart rate, two features suggestive of sleep disruption (**Fig. 5C**). The same allele is also enriched
308 in individuals with depression disorder, consistent with growing evidence implicating sleep
309 impairment as a transdiagnostic feature of many forms of adolescent psychopathology (**Fig. 5C,**
310 **right**)^{57,58}.

311

312 Overall, these results highlight how wearable-derived features can be leveraged as intermediate
313 phenotypes in GWAS and enable the identification of genetic variants relevant to clinical
314 psychiatry with significant effects on exhibited behavior in adolescents.

315

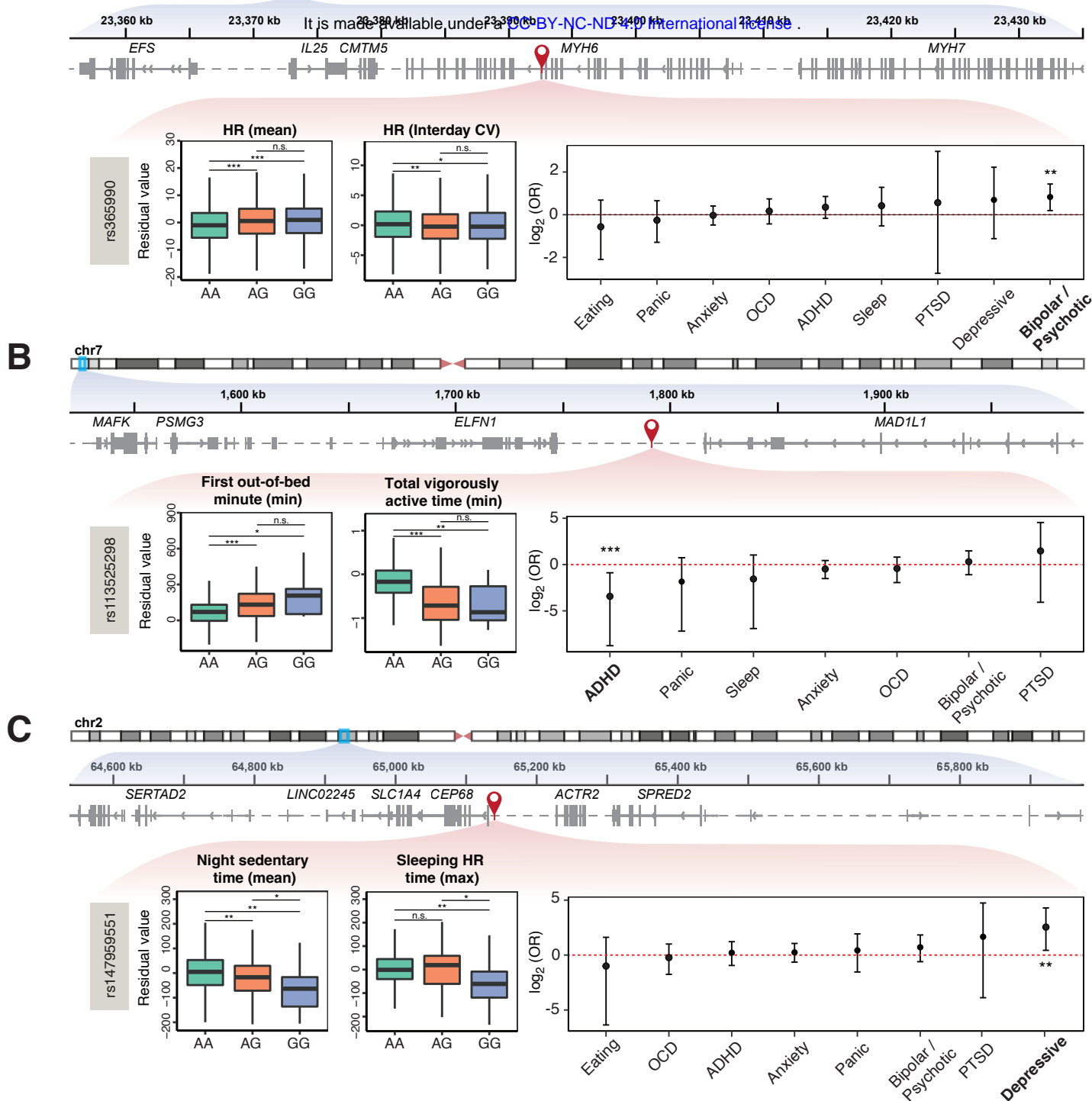


Figure 5. Exploring the genetic-physiological-psychiatric axis with wearable GWAS.

A) Left panel: rs365990 (chr14:23,392,602, A/G) is located in exon 25 of *MYH6*, and is associated with changes in wearable-derived heart rate features (multivariate GWAS p value = $5.33 \cdot 10^{-9}$). The boxplots show distributions of covariate-adjusted mean and interday coefficient of variation (CV) for heart rate across genotype groups at rs365990 (AA n individuals = 1,228; AG n individuals = 1,509; GG n individuals = 519). P values (two-sided Wilcoxon Rank-Sum test) for each pairwise comparison are also displayed, encoded as follows: *** ($p \leq 0.001$), ** ($0.001 < p \leq 0.01$), * ($0.01 < p \leq 0.05$), n.s. ($p > 0.05$). For visualization purposes, outliers are not shown. Right panel: enrichment, displayed as odds-ratio (\log_2 OR; y axis) of the minor allele (G) in individuals with different psychiatric disorders (x axis) compared to healthy controls. OR estimates and 95% confidence interval (error bar) are displayed. The red horizontal dashed line indicates no enrichment. The G allele is significantly more enriched in individuals with bipolar/psychotic disorder compared to healthy controls (two-sided Fisher test p value: $8.00 \cdot 10^{-3}$; FDR-adjusted p value: $7.00 \cdot 10^{-2}$). **B)** Similar representation for rs113525298 (chr7:1,791,353; AA n individuals = 2,294; AG n individuals = 101; GG n individuals = 15). rs113525298 is located 125 Kb from *ELFN1*, a gene that encodes a postsynaptic protein involved in the temporal dynamics of interneuron recruitment^{65,66}. *Elfn1* mutant mice exhibit hyperactivity that is treatable by psychostimulant medication^{55,56}. The G allele at rs113525298 is associated with increased minimum number of first-out-of-bed minutes and decreased minimum number of total-vigorously-active minutes (multivariate GWAS p value = $5.10 \cdot 10^{-9}$), and is significantly more enriched in healthy controls compared to individuals with ADHD (two-sided Fisher test p value: $9.00 \cdot 10^{-4}$; FDR-adjusted p value: $6.00 \cdot 10^{-3}$). **C)** Similar representation for rs147959551 (chr2:65,140,366; AA n individuals = 2,279; AG n individuals = 117; GG n individuals = 14), located near a cluster of genes relevant for several psychiatric disorders, such as *ACTR2* (schizophrenia; 87 Kb), *SLC1A4* (schizophrenia, bipolar disorder, major depressive disorder; 117 Kb) and *SPRED2* (schizophrenia, OCD; 170 Kb)⁶⁷⁻⁷⁷. The G allele of rs147959551 is associated with decreased mean number of sedentary-time-at-night minutes and decreased maximum number of sleep-based-on-heart-rate minutes (multivariate GWAS p value = $4.47 \cdot 10^{-8}$), and is significantly more enriched in individuals with depressive disorder compared to healthy controls (two-sided Fisher test p value: $9.74 \cdot 10^{-3}$; FDR-adjusted p value: $7.80 \cdot 10^{-2}$).

316 Discussion

317 Psychiatric disorders have been traditionally described with diagnostic categories based on
318 retrospective self-report of symptom sets. However, current efforts in the field are increasingly
319 leveraging novel technologies to transition from retrospective self-reporting and fixed symptom
320 sets to more dimensional conceptualizations, aiming to capture the complex and heterogeneous
321 nature of psychiatric disorders for more accurate research into their underlying structure⁶. One
322 approach to enhancing dimensional models is the use of intermediate phenotypes—quantitative
323 traits linked more closely to a disorder’s underlying molecular pathways. Although intermediate
324 phenotypes have been derived from cellular, tissue, and organ levels of information, computational
325 strategies that generate useful intermediate phenotypes in the behavioral domain are currently
326 limited. Wearable biosensors such as smartwatches offer a unique opportunity to objectively study
327 psychiatric disorders in a non-invasive way by measuring their underlying physiological
328 foundations of behavior over time.

329
330 Towards this end, we used wearable data to generate static and dynamic features that were
331 employed by our AI modeling framework as intermediate phenotypes to distinguish between
332 adolescents with and without psychiatric disorders. Models utilizing these wearable-derived
333 intermediate phenotypes performed comparably to those based on more expensive data sources
334 such as fMRI measurements^{18,59}. To gain critical theoretical insights and inform treatment
335 development efforts, we augmented the modeling framework with interpretability modules,
336 allowing us to pinpoint temporal and functional regions of the time series that were highly
337 correlated with overall disease state⁶⁰. These interpretability modules have the potential to
338 facilitate mechanistic studies that offer deeper insight into the underlying complexities of these
339 disorders. For example, our interpretability modules revealed that heart rate time series held high
340 importance in predicting ADHD. This finding aligns with the clinical manifestation of ADHD –
341 affected children are characterized by episodes of heightened arousal that are often incongruent
342 with environmental demands⁶¹. Conversely, the interpretability modules identified sleep intensity
343 and quality as key predictors in our anxiety disorder models, in line with known disruptions in
344 sleep patterns and circadian rhythms commonly seen in youth with anxiety disorders⁶².

345
346 Wearable-derived intermediate phenotypes are not just effective for detecting the presence of
347 psychiatric disorders in individuals; they also serve as a valuable research tool for understanding
348 the correspondence between behavior patterns and molecular attributes. This comprehensive
349 approach helps to uncover the foundational elements of pathological behavior patterns. In this
350 context, we focused on establishing links with genetics. Specifically, we showed that these
351 intermediate phenotypes can serve as response variables in GWAS models. Their continuous
352 nature enhances statistical power compared to categorical diagnostic labels. Furthermore, we took
353 advantage of the features’ correlated structure to create multivariate response variables for GWAS.
354 This strategy is statistically advantageous because it mitigates the multiple testing burden
355 associated with evaluating the numerous (>250) independent features. Conversely, from a
356 biological standpoint, these wearable GWAS allowed us to explore triaxial associations
357 encompassing genetic, physiological, and psychiatric factors. Utilizing our framework, we
358 successfully identified a significant association between a missense variant of the *MYH6* gene,
359 which encodes the cardiac muscle myosin, and heart rate patterns. Heart activity receives complex
360 inputs from the CNS, which implies behavioral influence and, in combination with our GWAS,
361 supports the notion of a gene-behavior-disorder pathway⁶³. Building on this finding, we discovered

362 enrichment of the same genetic variant among individuals with bipolar/psychotic disorders,
363 psychiatric conditions known to be associated with characteristic irregularities in heart activity⁵¹.
364 While additional research is needed to confirm such associations, our findings resonate with the
365 objectives of the RDoC initiative⁶. Specifically, wearable-derived intermediate phenotypes serve
366 as objective markers of behavior, bridging lower-level biological systems like genetics to broader
367 psychiatric disorders.

368
369 While we have employed these wearable-derived intermediate phenotypes in a targeted research
370 context (i.e., to enhance a psychiatric GWAS), their broad applicability make them promising for
371 other domains of health research. For example, the risk scores generated by our AI-modeling
372 framework could be used to assess disorder severity, and the genetic variants identified by our
373 wearable GWAS could be employed to construct more comprehensive polygenic risk scores for
374 behavioral and psychiatric disorders. Unlike other diseases (e.g., cancer) where objective
375 biomarkers are more common, psychiatry faces a significant barrier in treatment due to the lack of
376 objective and sensitive screening methods⁶⁴. Therefore, these physiological and genetic features
377 could be leveraged as objective biomarkers to more accurately subtype patients within diagnostic
378 categories, which in turn could help move towards precision treatment delivery in psychiatry.

379
380 Although the results presented in this study require further experimental validation, they illuminate
381 the transformative potential of wearable devices combined with AI modeling frameworks for
382 deepening our understanding of complex behavioral and psychiatric traits. We anticipate that
383 further development of our AI modeling framework, coupled with an expanded array of wearable
384 devices, could fundamentally transform how psychiatric disorders are measured and understood
385 in both research and clinical settings. This could lead to more nuanced digital intermediate
386 phenotypes and open new avenues for the study of human behavior.

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390
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401 <https://abcdstudy.org/federal-partners.html>. A listing of participating sites and a complete listing
402 of the study investigators can be found at https://abcdstudy.org/consortium_members/. ABCD
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405 of the authors and may not reflect the opinions or views of the NIH or ABCD consortium
406 investigators.

407 **Author Contributions**

408 Conceptualization: JL, BB, WR, MG

409 Methodology: JL, BB, YL, SL, YG, XX, SKL, MJ, DGM, TV, GA, JZ, MJG, WR, MG

410 Investigation: JL, BB, YL, SL, YG

411 Visualization: JL, BB, YL, SL, YG

412 Funding acquisition: MG

413 Data curation: TV, WR, MG

414 Supervision: WR, MG

415 Writing – original draft: JL, BB

416 Writing – review & editing: JL, BB, YL, SL, YG, XX, SKL, MJ, DGM, TV, GA, JZ, MJG, WR,
417 MG

418 **Competing interests**

419 The authors declare they have no competing interests.

420 **Methods**

421 **Dataset Description**

422

423 **ABCD Dataset**

424 The Adolescent Brain Cognitive Development (ABCD) study is a comprehensive longitudinal
425 project initiated in 2015 with the purpose of characterizing the neural, cognitive, and behavioral
426 aspects of adolescent development. Commissioned by a consortium of U.S. federal agencies,
427 ABCD investigators deeply phenotyped a large and representative sample of children aged 9-14
428 with plans to track their development into early adulthood. The ABCD dataset incorporated
429 multimodal brain imaging data, substance use history, behavioral and psychological measures,
430 genetic data, and an all-encompassing collection of demographic, physical health and activity,
431 mental health, and environmental information, including data derived from wearable devices.

432

433 **Clinical Diagnoses**

434 Clinical diagnoses were operationalized using the parent report version of the Kiddie Schedule for
435 Affective Disorders and Schizophrenia (KSADS). The KSADS is a gold standard semi-structured
436 diagnostic interview that is used to establish a broad range of clinical diagnoses in children and
437 adolescents. It has been used previously to define clinical groups in case/control studies conducted
438 with data from the ABCD study⁵⁹.

439

440 **Cohort Definitions**

441 We identified several clinical groups of interest in order to evaluate our framework across different
442 forms of psychopathology. The *nonclinical comparison* cohort was composed of youth who did
443 not meet any current diagnostic criteria for any disorder on their most recent administration of the
444 parent report KSADS. Similar diagnostic categories, based on ICD10 diagnostic codes, were
445 combined to create cohorts with sufficient sample sizes for our modeling framework. Each clinical
446 cohort was composed of the following diagnostic groups, based on the current reported symptom
447 sets (**Suppl. Table S1.1**). Some codes were included in multiple categories to balance the need for
448 sufficient sample size and homogenized cohort definitions.

449

450 **Preprocessing and Quality Control of Wearable Device Data**

451 We commenced by combining data from seven distinct wearable-derived modalities (heart rate,
452 calories, intensity, steps, METs, sleep level, and sleep intensity) for 5,339 individuals into a single
453 dataframe, resulting in highly sparse data structures. We excluded individuals with at least one
454 missing wearable modality, leaving us with 3,538 participants. To address the impact of missing
455 values on further analysis, we implemented a rigorous quality control procedure. In the initial
456 phase, we examined all potential time windows for two selected days each week per data modality.
457 Our objective was to balance the maximization of data inclusion with the assurance of its quality.
458 We pinpointed the time window that offered the best alignment - that is, the period which had the
459 highest number of valid measurements across all modalities. This procedure enabled us to
460 determine the most suitable time window for downstream analysis, taking into account both the
461 richness of the data and the necessity for top-quality inputs. In the next stage, we established a
462 criterion that each day must have at least 60% valid measurements within the identified optimal
463 window for an individual. Participants who did not meet this standard were removed from our

464 dataset. We provided a visual representation of the data processing and QC steps in **Suppl. Fig.**
465 **S2.3.**

466 **Imputation**

468 Missing values are still existent in the resulting QC-controlled time windows. To approach the
469 data missingness, we devised an imputation strategy for categorical and quantitative data
470 modalities, respectively. For the categorical data, we introduced a '*Not Recorded*' category into the
471 frame for imputation and subsequently applied label encoding. For the quantitative data, we used
472 the '*drift*' method from the sktime package (v0.19.1) with its default settings⁷⁸. Recognizing that
473 these imputation strategies may not be adept at capturing non-polynomial dynamics, we further
474 included an indicator time series for each data modality:

$$475 T_{\text{indicator}}(i) = \mathbf{1}(T(i) = \text{Na})$$

476 where $\mathbf{1}(\cdot)$ is the indicator function and $T(i) = \text{Na}$ indicates the data at time step i is missed.

478 We concatenated the indicator time series with the imputed time series along the channel
479 dimension. The indicator time series serves as a mask that shows where imputations have been
480 made, while the imputed time series contains both the actual and imputed data. By including this
481 additional indicator time series, we are effectively providing the model with the flexibility to learn
482 an adaptive imputation strategy, where the model can learn how to treat imputed data points based
483 on the surrounding, non-imputed data.

484 **Machine Learning Classifier**

485 **Problem Formulation**

488 We first formulated the phenotype classification as a canonical machine learning task with
489 *manually engineered features*, which is outlined as follows. Given an input for a set of features,
490 X , *machine learning classification* (MLC) targets an output value y which represents the
491 macrophenotype of the subject:

$$492 X \in \mathcal{R}^{N \times d} \mapsto y$$

494 Here, N is the number of individuals and d is the number of features. Specifically, we chose the
495 curated *XGBoostRegressor* model implemented in xgboost package (v1.7.5) as our backbone ML
496 models, i.e.:

$$497 \text{XGBOOSTREGRESSOR}(X) \mapsto y$$

499 **XGBoost** (eXtreme Gradient Boosting) has emerged as an effective machine learning framework,
500 noted for its optimized speed, scalability, and robustness²⁶. As a variant of gradient boosted
501 decision trees, XGBoost is tailored for efficiency and demonstrates consistent performance across
502 diverse machine learning applications. Central to XGBoost is its adeptness at engineering trees
503 which pinpoint and rectify residuals from prior iterations, continually refining model accuracy. In
504 this work, we take advantage of the strengths that XGBoost offers, guided by a carefully crafted
505 set of features.

506 **Feature Engineering**

508 Our feature engineering for the XGBoost model is elaborated below. Specifically, the time-
509 invariant wearable features X_w were primarily derived from summary statistics of the time-series

510 wearable. We identified seven clusters of time-invariant wearable features from a total of 258
511 features. We further included curated covariates X_{cov} as additional features to supplement the time-
512 invariant wearables features. Covariates used for machine learning model include demographic
513 background (sex, race, age at second-year follow-up, divorced parents, parents' level of education,
514 parent income, adoption) family history of psychiatric illness (bipolar disorder, schizophrenia,
515 antisocial behavior, nervous breakdown, psychiatric treatment, hospital admission, suicide),
516 cognitive scores (flanker test, picmemory, process speed, reading score, stop reaction time, etc.),
517 and child behavioral checklist (CBCL internalizing and externalizing scores). Our complete
518 features set encompasses both static wearable features and covariates:

$$519 \quad X = \text{Concat}(\{X_{\text{cov}}, X_w^1, X_w^2, \dots, X_w^7\})$$

520 where $\text{Concat}(\cdot)$ denotes concatenation on the feature dimension. This enabled us to characterize
521 a nuanced interplay of wearable features with individual covariates, substantially accentuating the
522 power of our model.
523

524

525 **Clustering of Wearable Static Features**

526 We considered the 258 wearable static features in a subset of 2,410 ABCD individuals with
527 complete genetic, wearable and covariate information (see Methods section “Multivariate
528 wearable GWAS”). We computed Pearson’s r correlation coefficients between all possible pairs
529 of features, and used these correlation values as distance measures to perform hierarchical
530 clustering (R function “hclust” & clustering method “Complete”). We also performed k-means
531 clustering of the correlation matrix by varying the number of clusters from two to twenty (R
532 function “kmeans”, with $nstart = 10$), and chose an optimal number of seven clusters based on the
533 elbow curve of the total within-cluster sum of squares. A heatmap representation of the seven
534 clusters is shown in **Figure 2D**, and the list of static features for each cluster is provided in **Suppl.**
535 **Tables S2.3-S2.4.**

536

537 **Class Balancing**

538 Imbalanced training labels, where one class substantially outnumbered the other (e.g. 1,737 control
539 individuals versus 216 ADHD individuals), pose a substantial impact on the model performance.
540 To address this issue and ensure a more robust model, we implemented stochastic downsampling
541 techniques on classes with higher representation in each run of the model. To formalize this, we
542 assume two classes, A and B , where $|A|$ and $|B|$ represent the number of instances in each class.
543 Assuming $|A| \geq |B|$, we calculate the ratio r :

544

$$545 \quad r = \frac{|B|}{|A|}$$

546 We then randomly select a subset A' from A such that:

547

$$548 \quad |A'| = r \times |A|$$

549 The downsampled dataset will then consist of A' and B :

550

$$551 \quad \text{Downsampled Dataset} = A' \cup B$$

552

553

554 Multichannel Time Series Classifier

555

556 Problem Formulation

557 We formulate the phenotype classification as a multichannel time series classification problem
558 which is described as follows. Given an input multichannel time series X :

559

$$560 X = \text{CONCAT}\{X_W, X_W^{\text{indicator}}, X_{\text{cov}}\},$$

561

562 where $X_W \in \mathcal{R}^{N \times c_w \times L}$, $X_W^{\text{indicator}} \in \{0, 1\}^{N \times c_w \times L}$, $X_{\text{cov}} \in \mathcal{R}^{N \times c_{\text{cov}} \times L}$. Here, N is the
563 number of samples, c_w the number of wearable modalities, L the number of measurements, X_W
564 the multichannel wearables data, $X_W^{\text{indicator}}$ the multichannel indicator data (See section
565 Imputation), and X_{cov} covariates (detail in next section). The *multi-channel time series*
566 *classification* (MCTSC) targets an output value y which represents the macrophenotype of the
567 subject:

568

$$569 X \in \mathcal{R}^{N \times C \times L} \mapsto y$$

570 $C = 2 \times c_w + c_{\text{cov}}$ is the number of input time-series channels. We further define a
571 parameterized model which maps X to the output y :

572

$$573 f_{\theta}(X) \rightarrow y$$

574 f represents the mapping function, which is parameterized by θ . To optimize θ , we employed *cross-*
575 *entropy loss with label smoothing* as the objective function, which is defined as:

576

$$577 \text{CE}(y, \hat{y}) := - \sum_{k=1}^K y_k \log(\hat{y}_k)$$

578 where K denotes the number of classes. We employed a label smoothing regularizer to the ground
579 truth label:

580

$$581 y_k^{\text{ls}} = y_k(1 - \alpha) + \frac{\alpha}{K}$$

582 Here, α is a smoothing parameter (we chose 0.1). This label smoothing technique helps to prevent
583 the model from becoming too confident about the class labels, which could potentially bolster its
584 generalization ability.

585

585 Covariate Integration

586 In order to integrate both covariates and time-series data for classification, we adapted the same
587 covariates described in XGBoost feature engineering into a time-series format. Essentially, we
588 transformed these variables into time-invariant sequences, where the value for each covariate
589 remains the same at every time step. The transformed time-series covariates were then merged
590 with wearable sensor data along the channel dimension. This approach allows the model to capture
591 potential interactions between covariates and wearable measures, wherein the model can adjust its
592 weights accordingly if a certain covariate influences the interpretation of the wearable data.

593

594 Xception Encoder

595 The XceptionTime encoder harnesses the power of one-dimensional convolutional neural
596 networks (1d-CNNs) as its underlying architecture²⁷. The model is structured with convolutional

597 filters of various sizes, which are sequentially followed by MaxPooling, Batch Normalization, and
 598 ReLU activation functions, which form residual connections. Formally:

599

$$\begin{aligned}
 600 \quad & H_{\text{bottleneck}}^l = \text{CONV1D}(H^{l-1}) \\
 601 \quad & H_{\text{MaxConvPool}}^l = \text{CONV1D}(\text{MAXPOOL}(H^{l-1})) \\
 602 \quad & \Delta H^l = \oplus \left\{ \oplus_k \left\{ \text{XCEPTIONCONV}_k(H_{\text{bottleneck}}^l) \right\}, H_{\text{MaxConvPool}}^l \right\} \\
 603 \quad & \Delta H^l = \text{BATCHNORM}(\Delta H^l) \\
 604 \quad & \Delta H^l = \text{RELU}(\Delta H^l) \\
 605 \quad & H^l = H^{l-1} + \Delta H^l \\
 606
 \end{aligned}$$

607 Here, $\text{CONV1D}(\cdot)$ denotes 1d convolution, $\text{XCEPTIONCONV}(\cdot)$ represents depthwise
 608 separable convolution, $\text{BATCHNORM}(\cdot)$ represents Batch Normalization, $\text{RELU}(\cdot)$ represents
 609 ReLU activation function, and \oplus aggregates feature maps from convolution filters of different
 610 sizes. A visual representation of the model could be found in **Suppl. Fig. S2.4**. In summary, the
 611 input feature maps are first projected to a bottleneck features map where the number of input
 612 channels is much larger than the number of output channels. A sequential operation of max pooling
 613 and 1d convolution is then performed on the input features maps to increase the expressivity of the
 614 model. The variation in the size of Xception convolutional filters gives rise to multi-level receptive
 615 fields, allowing the model to aggregate and process information at different levels of granularity
 616 or resolution. Such a property is particularly advantageous when dealing with data from wearable
 617 devices, as wearable data often exhibits both local patterns (i.e., minute-by-minute changes) and
 618 global trends (i.e., hourly or daily rhythms).

619

620 The XceptionTime encoder introduces a modification to the vanilla 1d convolution model by
 621 substituting the 1d convolution with a 1d depth-wise separable convolution. The operation can be
 622 broken down into two steps:

623

$$624 \quad \text{XCEPTIONCONV}(H^{l-1}) := \text{POINTWISECONV}(\text{DEPTHWISECONV}(H^{l-1}))$$

625

626 In contrast to the traditional convolution operation, the depth-wise separable convolution first
 627 applies a convolutional filter to each channel individually. This is followed by a 1x1 pointwise
 628 convolution module, which performs a linear combination of the outputs across channels. This
 629 process reduces the computational complexity of the model while still allowing for complex
 630 feature extraction. These steps are described in detail below:

631

632 *Depthwise Convolution:* This applies a single filter to each input channel which can be expressed
 633 as:

$$634 \quad F_c^d = H_c * K_c^d$$

635 where F_c^d is the output feature map for channel c after the depthwise convolution, H_c is the input
 636 feature map for channel c , and K_c^d is the depthwise filter (or kernel) for channel c . $*$ denotes the
 637 convolution operation.

638

639 *Pointwise Convolution:* This operation combines the outputs from the depthwise convolution
 640 across channels:

$$641 \quad F^p = \oplus_c Y_c^d * K^p$$

642 where F^p is the output feature map after the pointwise convolution, Y_c^d is the input feature map
643 for channel c , K^p is the pointwise filter, which has a spatial dimension of 1x1 and operates across
644 all channels, and \oplus is used to denote the aggregation of feature maps from all channels.

645

646 **Model Training and Evaluation**

647

648 **Training Details**

649 We split the dataset into 70% training set and 30% test set. We ran different experiments with 10
650 random seeds, and the final results were calculated as the mean of the 10 runs. This helps to
651 mitigate the risk of overfitting on a specific split, and provides a more robust estimate of the model
652 performance. We used ADAM as the optimizer for training, with 1×10^{-3} as the initial learning rate.
653 The neural network model is trained on an NVIDIA V100 32GB graphical processing unit using
654 the PyTorch and tsai deep learning libraries^{79,80}.

655

656 **Risk Scores**

657 In our study, we computed risk scores $RS \in \mathcal{R}^{N \times |\mathcal{K}|}$ by extracting the final layer of the deep
658 learning model, specifically the softmax probability. For the XGBoost model, we leverage the
659 `pred_prob` method implemented in the XGBoost library. Specifically:

660

$$RS(j) := P(y = j|x) = \frac{e^{f_j}}{\sum_{k \in \mathcal{K}} e^{f_k}}$$

661

662 where $f \in \mathcal{R}^{N \times K}$ is either the XceptionTime logits in the XceptionTime model, or the sum of
663 outputs from all trees in the XGBoostRegressor model. The softmax function, used in the final
664 layer of the deep learning model, returns probabilities for each category in a multi-class problem
665 that sum up to 1. Similarly, XGBoost's `predict_proba` method generates class probabilities as
666 output. These scores can serve as a measure of the 'risk' or likelihood associated with each class or
667 outcome. We utilized these risk scores as the response variable in our subsequent GWAS study
668 (see Methods section "Univariate Wearable GWAS"). This approach not only bridged the gap
669 between deep learning modeling and GWAS but also significantly enhanced the power of our
670 GWAS studies.

671

672 **Model Interpretability**

673

674 **Ablation Method for Step and Feature Importance**

675 The ablation method we present was used to measure the importance of features in a dataset.
676 Ablation methods are based on randomly rearranging the values of a feature or a group of features
677 across all subjects in the dataset, and then calculating an importance score based on the decrease
678 in a chosen metric. In our case, we utilized the *Area Under the Receiver Operating Characteristic*
679 *curve* (AUROC) as the metric to calculate this score. The rationale behind this method is that if a
680 feature is important for model predictions, shuffling the values of that feature should disrupt the
681 model's ability to make accurate predictions, leading to a decrease in the chosen performance
682 metric. The larger the decrease, the more important the feature is considered to be.

683

684 The ablation importance score can be applied to calculate both feature importance and step
685 importance. For step importance, the implementation is slightly different. Instead of shuffling
686 individual features, we shuffled the values within selected windows of the time series. The time

687 series was divided into windows of a chosen length (in our case, 1 hour), and these windows were
688 then shuffled across all subjects, allowing us to assess the importance of information at different
689 time steps or periods. If the model performance significantly decreases when the values within a
690 certain time window are shuffled, the information within that time window is important for the
691 model predictions.

692

693 **Grad-CAM**

694 The weighted class activation mapping (CAM) method is a well-established technique for
695 examining how a trained model makes its predictions²⁹. In the context of time-series data, it can
696 highlight which time steps are particularly influential in the model's decision-making process.

697

698 We first computed the gradient of the score for the predicted class y with respect to the feature

699 map of the first layer activations $A_0^{c,i}$ of a convolutional layer. This gradient, denoted as $\frac{\partial y}{\partial A_0^{c,i}}$,
700 provides a measure of how a small change in the activation of the convolutional layer could affect
701 the final prediction of the model. To convert these gradients into a measure of importance for each
702 channel (indexed by c), we employed a global average pooling, which calculates the average of all
703 gradients across the sequence length (indexed by i). This resulted in a set of channel-wise gradient
704 averages, denoted as α_c . Mathematically, this is expressed as:

705

$$\alpha_c := \frac{1}{Z} \sum_{i=1}^L \frac{\partial y}{\partial A_0^{c,i}}$$

706

707 where Z is a normalization constant, typically the total number of elements in the layer, and L is
708 the length of the sequence.

709

710 We next generated the Gradient-weighted Class Activation Mapping (Grad-CAM). This is a visual
711 representation of the importance of each time step for the model's predictions. The Grad-CAM,
712 denoted as L_{GM}^i , is defined as:

713

$$L_{GM}^i = \text{RELU}\left(\sum_c \alpha_c A_0^{c,i}\right)$$

714

715 where the ReLU (Rectified Linear Unit) function is used to ensure that only features with a positive
716 influence on the class of interest result in high activation. Essentially, this means that only the time
717 steps that positively contribute to the model's decision will have high importance scores.

718

719 Finally, for each time step, we computed the average Grad-CAM scores across the entire test set.
720 This allowed us to determine which time steps in the input data were most influential in the model
721 predictions.

722

723 **Genome-wide Association Studies (GWAS)**

724

725 **Quality Control of Genetic Data**

726 We obtained genotyped and imputed genetic data for 11,099 individuals as part of the ABCD Data
727 Release 3 (<https://abcdstudy.org/scientists/data-sharing/>). We used the genotyped data to infer

728 population stratification and the imputed data to perform the different GWAS described below
729 (see Methods sections “Univariate wearable GWAS” and “Multivariate wearable GWAS”).
730 We applied a quality control (QC) protocol on the genotyped data⁸¹. Specifically, we performed
731 the QC steps described in
732 https://github.com/MareesAT/GWA_tutorial/blob/master/1_QC_GWAS.zip (file
733 “1_Main_script_QC_GWAS.txt”) using PLINK v1.90b6.21⁸². Briefly, of the initial set of 516,598
734 variants, we kept those with a missingness rate across individuals < 0.02 ($n = 481,920$). Of the
735 initial set of 11,099 individuals, we kept those with a missingness rate across variants < 0.02 ($n =$
736 $10,660$). Next, we considered only variants located on autosomal chromosomes ($n = 470,076$),
737 those with Minor Allele Frequency (MAF) > 0.01 ($n = 427,704$), and those that did not deviate
738 from Hardy-Weinberg equilibrium (p value $\geq 10^{-10}$; $n = 370,002$). These variants were pruned to
739 a final set of 156,556 variants (window size = 50; number of variants to shift the window at each
740 step = 5; multiple correlation coefficient 0.2). We computed the heterozygosity rate for each
741 individual using the pruned set of variants, and kept individuals with a heterozygosity rate
742 deviating less than 3 standard deviations from the mean ($n = 10,467$). We also used pruned variants
743 to assess cryptic relatedness by identifying groups of individuals with Proportion Identity-By-
744 Descent ($\pi_{\hat{}}$) > 0.2 . For every group of related individuals, we then selected the individual with
745 the lowest variant missingness rate, leaving a total of 8,816 individuals. We used PLINK2 to
746 perform a Principal Component Analysis (PCA) on the 156,556 pruned genotyped variants from
747 the 8,816 selected individuals. We integrated the PCA results with the ethnicity score group
748 information provided by the ABCD metadata, which was available for 8,791 individuals (**Fig. 1D**
749 and **Suppl. Fig. S4.1-S4.4**).

750
751 We filtered the imputed genetic variants for MAF > 0.01 and estimated imputation accuracy (R^2) $>$
752 0.3 , and obtained a final set of 11,954,686 variants for the GWAS analysis (**Suppl. Fig. S4.5**). We
753 also computed distributions of R^2 for all (genotyped and imputed) variants, and of empirical leave-
754 one-out R^2 (ER^2) for genotyped variants (**Suppl. Fig. S4.6**).

756 **Covariates included in the GWAS**

757 We considered five different groups of covariates: basic (sex, age at second-year follow-up, first
758 five genotype PCs), behavioral (CBCL internalizing and externalizing scores, DSM internalizing
759 and externalizing scores), family history of psychiatric illness (bipolar disorder, schizophrenia,
760 antisocial behavior, nervous breakdown, psychiatric treatment, hospital admission, suicide),
761 family situation (divorced parents, parents’ level of education, family income, adoption), and other
762 (ACS raked propensity score, DNA extraction batch). 3,579 of the previously selected 8,791
763 individuals reported complete information for these 24 covariates.

765 **Univariate Wearable GWAS**

766 For this analysis, we focused on a subset of 1,191 individuals that were either diagnosed with
767 ADHD ($n = 137$) or belonged to the non-clinical control group ($n = 1,054$). We performed a GWAS
768 testing for association between genetic variants and ADHD diagnosis, encoded as a binary
769 outcome (ADHD = 1, control = 0; univariate binary GWAS; **Figure 4A**). We also obtained, for
770 each individual, ten different ADHD risk scores based on the XGBoost and Xception predictive
771 models (see Methods section “Risk Scores”). Specifically, we used risk scores from the following
772 six models: baseline model using CBCL externalizing score (“CBCL ext.”); baseline model using
773 CBCL internalizing score (“CBCL int.”); XGBoost model using wearable features (“XGB”);

774 XGBoost model using wearable features and CBCL scores (“XGB + CBCL”); Xception model
775 using wearable features (“Xception”); and Xception model using wearable features and CBCL
776 scores (“Xception + CBCL”). For models “XGB”, “XGB + CBCL”, “Xception” and “Xception +
777 CBCL”, we also implemented the “liability-CC” trait methodology³⁵. This methodology consists
778 of converting the predictive modeling risk score of the cases (individuals with ADHD) to a value
779 of 1, while keeping the original risk scores for the controls. These four additional types of scores
780 are labeled as “XGB v2”, “XGB + CBCL v2”, “Xception v2” and “Xception + CBCL v2”.

781
782 We performed ten different GWAS to test for associations between genetic variants and each of
783 these ten scores (univariate continuous GWAS; **Figure 4B**). We used PLINK2 to perform both
784 binary and continuous univariate GWAS, and the FUMA platform for loci definition (reference
785 panel population: “1000G Phase3 ALL”)^{83,84}. We first ran all GWASs using only the set of basic
786 covariates (sex, age, first five population structure PCs), as these were also used in previous GWAS
787 for ADHD^{85,86}. These results and the corresponding *p* value quantile-quantile plots are shown in
788 **Figure 4A-B, Table 1** (GWAS Method: “univariate continuous”), **Suppl. Fig. S4.7-S4.8** (“Basic
789 covariates”), and **Suppl. Table S4.1**. We also repeated both continuous and binary GWAS to
790 include all covariates described in Methods section “Covariates included in the GWAS”, apart
791 from the CBCL and DSM behavioral scores, which were employed as features for the predictive
792 models that generated the risk scores (**Suppl. Fig. S4.7 and S4.9** - “All covariates”).

793

794 **Statistical Power of Binary vs. Continuous Traits**

795 To compare the statistical power of genetic association testing using binary and continuous traits,
796 we simulated a cohort of 1,500 individuals. In each individual *i*, we generated biallelic SNPs with
797 a binomial model (i.e., the genotype at each SNP followed a binomial distribution, with the number
798 of trials equal to 2 and probability of success on each trial equal to a given MAF). We chose the
799 cohort size to approximate the number of individuals (*n* = 1,191) in the univariate GWAS for
800 ADHD described above (Methods section “Univariate wearable GWAS”). For each individual *i*,
801 we then simulated a continuous trait (*C_i*) as the sum of the genotype effect (*b*) at a given SNP with
802 genotype *x_i* (0, 1, or 2) and random noise (*e_i*):

803

$$804 C_i = x_i \cdot b + e_i$$

805 where

$$806 \mathbf{b} \sim U(0,1)$$

$$807 \mathbf{e} \sim N(0,1)$$

808

809

810 We also simulated a binary trait for each individual *i* (*B_i*), following

811

$$812 B_i = 1 \text{ if } C_i > \text{median}(C), \text{ otherwise } 0$$

813

814 where *C* is the vector of simulated continuous traits for the entire cohort.

815

816 For a particular genotype effect *b*, we ran 10,000 simulations. Under this scenario, we estimated
817 the power of the simulated continuous and binary traits as the fraction of significant (i.e.
818 Benjamini-Hochberg adjusted *p* value < 0.05) linear and logistic regression tests, respectively. We
819 employed linear and logistic regression as implemented in the R functions “lm” (library “stats”)

820 and “glm” (family = binomial; library “MASS”), respectively. Overall, we simulated 50 different
821 values of b across nine different MAFs (**Suppl. Fig. S4.11**).

822

823 **Multivariate Wearable GWAS**

824 This second type of GWAS consists in testing the association between genetic variants and a multi-
825 trait (multivariate) phenotype. In this case, we define the multivariate phenotype as the vector of
826 static summary features obtained from an individual’s wearable device (see Methods section
827 “Clustering of Wearable Static Features”). For this analysis, we considered all individuals with
828 complete genetic, wearable, and covariate data independently of their diagnosis.

829

830 We first conducted a wearable GWAS using the 14 static features related to heart rate as the
831 multivariate response variable, which were available for 3,256 individuals (features: InterdayCV,
832 InterdaySD, IntradayCV_mean, IntradayCV_median, IntradayCV_sd, IntradayMean_mean,
833 IntradayMean_median, IntradayMean_sd, IntradaySD_mean, IntradaySD_median,
834 IntradaySD_sd, Mean, Median, STD). We next aimed to include all 258 static features, which were
835 available for 2,410 individuals, and applied two different strategies to reduce the dimensionality
836 of the multivariate response. In one case, we performed a PCA of the individuals based on their
837 values for the 258 features, and used the first five PCs as the multivariate response. In the second
838 case, we considered each of the seven clusters of static features as a separate multivariate response,
839 and performed a GWAS for each cluster (see also Methods section “Clusters of static summary
840 features” below). Therefore, we ran a total of nine multivariate GWAS (one for heart rate features,
841 one for the first five PCs of all features, and one for each of the seven clusters of features). For all
842 multivariate GWAS, we defined a model that included the genotype and 24 covariates as
843 independent variables (see Methods section “Covariates included in the GWAS”).

844

845 We used the Multivariate Asymptotic Non-parametric Test of Association R package (MANTA,
846 <https://github.com/dgarrimar/manta>) to test for association between genetic variants and the
847 multivariate wearable trait, and performed all the analyses within a containerized Nextflow
848 pipeline, available at <https://github.com/dgarrimar/mvgwas-nf>^{44,87}. Since MANTA is a non-
849 parametric method, normalization of the GWAS traits was not required. After performing the
850 different GWAS runs, we used FUMA for loci definition (reference panel population: “1000G
851 Phase3 ALL”)⁸⁴. These results and the corresponding p value quantile-quantile plots are shown in
852 **Figure 4C, Table 1** (GWAS Method: “multivariate continuous”), **Suppl. Fig. S4.12**, and **Suppl.**
853 **Table S4.3**. As MANTA p values do not come from a normal distribution, we employed λ_X
854 (instead of the commonly used λ_G) to estimate the genomic inflation factor⁸⁸.

855

856 **Genome-wide vs. Study-wide Significance**

857 We selected the conventional genome-wide significant p value threshold of $5 \cdot 10^{-8}$ to identify
858 significant loci from all GWAS runs. However, in line with previous GWAS studies, we also
859 considered a study-wide significance threshold to account for the fact that multiple GWAS were
860 performed⁸⁹. In our case, the study-wide significant thresholds are $5 \cdot 10^{-9}$ ($5 \cdot 10^{-8} / 10$ GWAS runs)
861 for the univariate continuous GWAS for ADHD, and $5.56 \cdot 10^{-9}$ ($5 \cdot 10^{-8} / 9$ GWAS runs) for the
862 multivariate wearable GWAS. Based on these thresholds, one locus from the ADHD GWAS and
863 nine loci from the wearable GWAS would pass the study-wide significance threshold. Similar to
864 other GWAS, we also considered a suggestive p value threshold of $1 \cdot 10^{-5}$ (**Figure 4** and **Suppl.**
865 **Fig. S4.7**)⁸⁹.

866 **Chromosome X**

867 Because the multivariate GWAS (specifically in runs corresponding to feature clusters 2 and 7)
868 reported a large number of significant loci on chromosome X (**Table 1** and **Suppl. Table S4.3**),
869 we implemented additional quality controls to account for potential bias in chromosome X
870 variants. First, we tested whether chromosome X variants showed systematically lower p values
871 compared to variants located on autosomal chromosomes. To do this, we computed the proportion
872 of variants with p value $< 10^{-4}$ from each autosomal chromosome, and performed a one-sided
873 Fisher's exact test to evaluate whether this proportion was significantly lower compared to variants
874 on chromosome X for the same GWAS run. We found that this was the case only in GWAS runs
875 for clusters 2, 4, and 7 (**Suppl. Fig. S4.13**). We reasoned that if an unknown systematic bias related
876 to chromosome X was truly present (e.g., genotyping issues), we would observe the same situation
877 for every cluster of features. Given that we did not identify significant loci on chromosome X for
878 cluster 4, this analysis ruled out any unaccounted systematic bias related to chromosome X, and
879 confirms that wearable features in clusters 2 and 7 indeed show stronger association with variants
880 located on chromosome X. We also checked for imputation bias in chromosome X variants that
881 could systematically differentiate female and male individuals. To do this, we performed a PCA
882 of the 2,410 individuals based on their imputed genotypes at chromosome X variants, and did not
883 observe a separation between female and male individuals (**Suppl. Fig. S4.14**).

884

885 **Neuropsychiatry-related Proximal Genes and eGenes**

886 For each genome-wide significant locus, we retrieved the ten closest genes when considering a
887 window of ± 250 Kb from the center of the locus, using the GENCODE human genome annotation
888 version 41⁹⁰. Next, we labeled as "neuropsychiatric-related" those proximal genes that are
889 associated with psychiatric disorders according to OpenTargets (<https://platform.opentargets.org/>)
890 (**Suppl. Tables S4.1** and **S4.3**)⁹¹. We further intersected our catalog of genome-wide significant
891 loci with previous eQTL catalogs using BEDTools intersect (v2.30.0), and identified a subset of
892 proximal neuropsychiatric-related genes with eQTLs overlapping our list of loci^{46,92-95}. We labeled
893 these genes as "neuropsychiatric-related proximal eGenes" (**Suppl. Tables S4.1** and **S4.3**).

894

895 **Chromatin Dissection of locus chr14:23392601-23418974**

896 We first performed an exploratory analysis by intersecting our two lists of significant loci with the
897 ENCODE4 registry of candidate cis-regulatory elements
898 (cCREs)([https://www.encodeproject.org/search/?type=Annotation&encyclopedia_version=curre
899 nt&annotation_type=candidate+Cis-
900 Regulatory+Elements&annotation_type=chromatin+state&annotation_type=representative+DNA
901 se+hypersensitivity+sites&status=released&encyclopedia_version=ENCODE+v4](https://www.encodeproject.org/search/?type=Annotation&encyclopedia_version=current&annotation_type=candidate+Cis-Regulatory+Elements&annotation_type=chromatin+state&annotation_type=representative+DNAse+hypersensitivity+sites&status=released&encyclopedia_version=ENCODE+v4)) (**Suppl.
902 Tables S4.1** and **S4.3**)⁴⁵. Given the documented role of locus chr14:23392601-23418974 in heart-
903 related traits and diseases, we next evaluated the enrichment of heart-specific epigenetic features
904 (nucleosome positioning, histone modifications, and transcription factor (TF) binding) at this
905 locus. We downloaded peak calling files for DNase-seq, ATAC-seq, ChIP-seq (histone marks &
906 TFs) and Mint-ChIP-seq for histone marks available for human biosamples from the ENCODE
907 portal
908 ([https://www.encodeproject.org/metadata/?control_type%21=%2A&status=released&perturbed=
909 false&assay_title=Histone+ChIP-seq&assay_title=TF+ChIP-seq&assay_title=DNase-
910 seq&assay_title=ATAC-seq&assay_title=Mint-ChIP-
911 seq&files.file_type=bigBed+narrowPeak&replicates.library.biosample.donor.organism.scientific](https://www.encodeproject.org/metadata/?control_type%21=%2A&status=released&perturbed=false&assay_title=Histone+ChIP-seq&assay_title=TF+ChIP-seq&assay_title=DNase-seq&assay_title=ATAC-seq&assay_title=Mint-ChIP-seq&files.file_type=bigBed+narrowPeak&replicates.library.biosample.donor.organism.scientific)

912 [_name=Homo+sapiens&type=Experiment&files_analyses.status=released&files_preferred_default=true](https://www.encodeproject.org/report/?type=Experiment&files_analyses_status=released&files_preferred_default=true);
913 [t=true](https://www.encodeproject.org/report/?type=Experiment&control_type!=*&status=released&pe); access date: 09/27/2022)^{45,96}. We then grouped human biosamples based on their
914 “biosample ontology organ slim”
915 (https://www.encodeproject.org/report/?type=Experiment&control_type!=*&status=released&pe
916 [rturbed=false&assay_title=TF+ChIP-seq&assay_title=Histone+ChIP-seq&assay_title=DNase-](https://www.encodeproject.org/report/?type=Experiment&control_type!=*&status=released&pe)
917 [seq&assay_title=ATAC-seq&assay_title=Mint-ChIP-](https://www.encodeproject.org/report/?type=Experiment&control_type!=*&status=released&pe)
918 [seq&replicates.library.biosample.donor.organism.scientific_name=Homo+sapiens&field=biosam](https://www.encodeproject.org/report/?type=Experiment&control_type!=*&status=released&pe)
919 [ple_ontology.organ_slims&field=biosample_ontology.cell_slims&field=biosample_ontology.sys](https://www.encodeproject.org/report/?type=Experiment&control_type!=*&status=released&pe)
920 [tem_slims&field=%40id&field=biosample_ontology.term_name](https://www.encodeproject.org/report/?type=Experiment&control_type!=*&status=released&pe)). To test the tissue-specific
921 enrichment of chromatin features in a particular organ, we computed the number of times any of
922 the five significant variants at the locus overlapped a peak from experiments in that organ
923 compared to all other organs (two-sided Fisher’s exact test, Benjamini-Hochberg adjusted p value
924 < 0.1). For this analysis, we counted only once those overlaps involving variants that are < 100 bp
925 apart.

926

927 **Exploring the Genetic-behavioral-psychiatric Axis**

928 The multivariate wearable GWAS allowed us to first perform an exploratory analysis to identify
929 genetic variants associated with any of the seven clusters of wearable-derived features (genome-
930 wide significant p value $< 5 \cdot 10^{-8}$). To identify the specific features that are driving the significant
931 association between the cluster and the variant, we next performed unpaired Wilcoxon rank tests
932 between all three groups of genotype individuals (i.e., AA vs. AG, AA vs. GG, and AG vs. GG)
933 for each feature within a particular cluster. We then selected features where at least one of the three
934 Wilcoxon tests reported a Benjamini-Hochberg-adjusted p value < 0.1 , and showed examples for
935 three SNPs in **Figure 5A-C** (left panel). For each of these three examples, we next evaluated the
936 enrichment of the minor allele (in all three cases the G allele) in individuals within a specific
937 psychiatric cohort vs. non-clinical control individuals (two-sided Fisher exact test; **Figure 5A-C**
938 **right panel**). Given the reduced number of individuals with GG genotype for SNPs rs113525298
939 and rs147959551 (15 and 14, respectively), in these two cases the enrichment of the minor allele
940 was tested by merging individuals with AG and GG genotypes. For SNP rs365990, the enrichment
941 was computed only on individuals with GG genotype. For all tests, we required at least one
942 individual to be present in every cell of the 2×2 matrix employed for the Fisher’s exact test ($a =$
943 n individuals with minor allele AND part of the psychiatric cohort; $b = n$ individuals without minor
944 allele AND part of the psychiatric cohort; $c = n$ individuals with minor allele AND part of the
945 healthy controls; $d = n$ individuals without minor allele AND not part of the healthy controls).

946

947 **Intersection of genome-wide significant loci with the GWAS Catalog**

948 To assess the clinical relevance of our GWAS loci, we intersected them with variants from the
949 NHGRI-EBI GWAS catalog (<https://www.ebi.ac.uk/gwas/>; access date: 05/16/2023). For the loci
950 identified by our ADHD GWAS, we considered overlaps with brain- or neuropsychiatric-related
951 GWAS hits (**Figure 4B**). Because our wearable-derived features are mostly related to heart, sleep,
952 metabolism and physical activity, for the wearable GWAS loci we considered any overlaps with
953 heart-, sleep-, metabolism- and physical activity-related GWAS hits. Additionally, given the
954 presence of individuals with psychiatric disorders in the wearable GWAS cohort, we also
955 considered intersections with brain- or neuropsychiatric-related GWAS hits (**Figure 4C**). We
956 acknowledge that colocalization analysis would be the most appropriate way to compute these
957 intersections, and we performed this analysis for ADHD GWAS loci (see Methods section

958 “Colocalization analysis”). However, MANTA does not provide estimates of variant effect sizes
959 that can be directly employed in co-localization analysis. For this reason, we evaluated the strength
960 of these intersections against a null distribution. Specifically, we computed the proportion of
961 GWAS variants associated with a particular trait that overlap our significant loci, and compared it
962 to the proportions observed across 10,000 random sets of genomic loci with the same size and
963 chromosome location. We report the percentile of our GWAS enrichments compared to the null
964 distribution in **Suppl. Fig. S4.10** (univariate GWAS for ADHD) and **S4.15** (multivariate GWAS).
965

966 **Colocalization Analysis**

967 We performed colocalization analysis using the R package *coloc* (function *coloc.abf*, default
968 parameters) on the results obtained from the univariate continuous GWAS⁹⁷. Specifically, we
969 focused on two of the seven overlapping brain-related traits with available GWAS summary
970 statistics (**Figure 4B** and **Suppl. Table S4.2**), and tested the hypothesis of signal co-localization
971 between our ADHD risk scores at the intersecting loci. Locus chr17:32256997:32283356 reported
972 a posterior probability of 0.99 for signal co-localization with a locus previously associated with
973 chronotype measurement⁹⁸. We also tested locus chr7:68219282:68338849 (suggestive
974 association at p value $< 10^{-5}$) for co-localization with a previously reported locus for ADHD⁸⁵. In
975 this case, given that the two traits being tested are the same, we set all three parameters p_1 , p_2 and
976 p_{12} equal to $1 \cdot 10^{-5}$, and reported a posterior probability of 0.25.
977

978 **Code Availability**

979 The code for the paper is publicly available at <https://github.com/gersteinlab/ABCD>.
980

981 **Data and Materials Availability**

982 The data used in this study is available through the NIMH ABCD NDA portal
983 ([https://nda.nih.gov/general-query.html?q=query=featured-](https://nda.nih.gov/general-query.html?q=query=featured-datasets:Adolescent%20Brain%20Cognitive%20Development%20Study%20(ABCD))
984 [datasets:Adolescent%20Brain%20Cognitive%20Development%20Study%20\(ABCD\)](https://nda.nih.gov/general-query.html?q=query=featured-datasets:Adolescent%20Brain%20Cognitive%20Development%20Study%20(ABCD))). Data
985 used in the preparation of this article were obtained from the Adolescent Brain Cognitive
986 DevelopmentSM (ABCD) Study (<https://abcdstudy.org>), held in the NIMH Data Archive (NDA).
987 This is a multisite, longitudinal study designed to recruit more than 10,000 children age 9-10 and
988 follow them over 10 years into early adulthood. The ABCD Study® is supported by the National
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