



Complete Genome Sequences of *Arthrobacter* Phages Eraser, Kaylissa, and Phives

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ABSTRACT Bacteriophages Phives, Kaylissa, and Eraser are siphoviruses infecting *Arthrobacter globiformis* B-2880 that were isolated in fall 2019 in Long Island, New York, from soil samples collected in Old Westbury, New York. All three bacteriophages are assigned to phage cluster AZ based on gene content similarity. While many aspects of the genomes are similar across the three phages, the endolysin genes for the phages are different and are located in different locations within the genomes.

B acteriophages are the most abundant organisms in the biosphere (1). Recent efforts to isolate and characterize bacteriophages are providing valuable insights into host-pathogen evolutionary relationships and uncovering potential therapeutic and biotechnical applications (2–4). *Arthrobacter* phages Phives, Kaylissa, and Eraser were isolated from soil samples from the campus of the New York Institute of Technology in Old Westbury, New York (Table 1), as part of the Science Education Alliance-Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES) program (5). Phage isolation, plaque purification, and genome extraction were performed according to protocols described in the SEA-PHAGES Discovery Guide (https://seaphagesphagediscoveryguide.helpdocsonline.com/home). All three bacteriophages were isolated by enrichment on *Arthrobacter globiformis* B-2880 with peptone-yeast-calcium (PYCa) medium at 30°C and purified with three rounds of plaque purification. Plaques of Phives and Kaylissa have a bullseye morphology, while Eraser has larger (8- to 12-mm) round, clear plaques. Negative-stain transmission electron microscopy (TEM) analysis shows that all three phages have icosahedral heads and noncontractile tails (Fig. 1), reflecting siphoviral morphology (6).

Genome extraction was performed from high-titer lysates using the Wizard DNA cleanup kit (Promega), and sequencing was performed at the University of Pittsburgh. Libraries were constructed using the NEBNext Ultra II FS DNA library preparation kit and sequenced using an Illumina MiSeq v3 platform generating 150-bp unpaired ends. Raw reads were assembled using Newbler v2.9 with default settings, generating single contigs with coverage of approximately $2,651\times$ for Phives, $2,171\times$ for Kaylissa, and $2,081\times$ for Eraser (7). The phage contigs were checked using Consed v29 to evaluate completeness and determine genomic termini (8). The genome parameters (length, GC content, and termini) and accession numbers (GenBank and SRA) are shown in Table 1.

All bioinformatic analyses and software were used with default parameters. The three phages were assigned to cluster AZ based on shared gene content similarity (GCS) exceeding at least 35% using the online tool at the PhagesDB database (https://phagesdb.org/genecontent/) (9, 10). Coding regions were predicted using GeneMark v3.25 (11) and GLIMMER v3.02b (12) and subsequently manually curated using DNA Master v5.23.6 (13), Phamerator (14), BLAST (15), and Starterator v1.0.1 and v1.2 (http://phages.wustl.edu/starterator). No tRNA genes were identified with ARAGORN v1.2.41 (16). Functions for each coding sequence were evaluated using NCBI BLASTP v2.9 (15), HHpred v3.2 (17), and Phamerator (14). Membrane proteins were predicted using TMHMM v2.0 (18).

Editor Kenneth M. Stedman, Portland State University

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The authors declare no conflict of interest.

Received 24 February 2022 Accepted 17 March 2022 Published 7 April 2022

	GenBank accession	SRA accession	Sampling location		Avg coverage	No. of reads	Genome	GC content	Genome end	No. of
Phage	no.	no.	coordinates	Cluster	(×)	(thousands)	size (bp)	(%)	(3' overhang)	genes
Phives	MT889376	SRX12198771	40.813139N, 73.604667W	AZ	2,651	775.7	44,204	67.3	CGAAGGGGCAT	70
Kaylissa	MZ005682.1	SRX12198769	40.7887N, 73.5996W	AZ	2,171	669.9	44,124	67.6	CGAAGGGGCAT	71
Eraser	MZ747516	SRX12198766	40.81255N, 73.604333W	AZ	2,082	609.6	43,608	66	CGAAGGGGCAT	69

TABLE 1 Phage GenBank and SRA accession numbers and genome assembly results

Phives, Kaylissa, and Eraser have genome architectures consistent with *Arthrobacter* phage cluster AZ genomes. The genomes of all three phages have defined ends with 11-base, complementary, 3' single-stranded extensions. Predicted genes in the left halves of the genomes are well conserved among the three phages and code for virion structure and assembly proteins. The right halves of the genomes are less well conserved, and predicted genes with no known function are prevalent. Phives, Kaylissa, and Eraser contain a serine integrase gene and are predicted to be temperate, although no repressor gene has been identified. The putative endolysin genes for these phages are in different locations within the genomes (Phives, gene 1; Eraser, gene 25; Kaylissa, gene 59), and they share less than 31% amino acid identity with each other.

Data availability. GenBank and Sequence Read Archive (SRA) accession numbers for phages Phives, Kaylissa, and Eraser are provided in Table 1.

ACKNOWLEDGMENTS

This project was supported by the Howard Hughes Medical Institute SEA-PHAGES program and by the Department of Biological and Chemical Sciences at the New York Institute of Technology.

We thank Graham F. Hatfull, Deborah Jacobs-Sera, Daniel A. Russell, and Rebecca A. Garlena at the University of Pittsburgh for their technical support during the sequencing and annotation of these genomes. We also thank Susan Van Horn of the Cellular and Molecular Imaging (CMIC) TEM facility at Stony Brook University for assistance in collecting TEM images of bacteriophages Phives and Kaylissa and Jacqueline Keighron of the New York Institute of



FIG 1 Plaque morphology (A to C) and TEM images (D to F) of *Arthrobacter* phages Eraser (A and D), Phives (B and E), and Kaylissa (C and F). Phages were incubated at 30°C for 48 h prior to imaging. Phage lysates were negatively stained with 1% uranyl acetate for TEM.

Technology for collection of TEM images of bacteriophage Eraser at the Brookhaven National Laboratory Center for Functional Nanomaterials.

REFERENCES

- Hatfull GF. 2015. Dark matter of the biosphere: the amazing world of bacteriophage diversity. J Virol 89:8107–8110. https://doi.org/10.1128/JVI.01340-15.
- Hatfull GF. 2020. Actinobacteriophages: genomics, dynamics, and applications. Annu Rev Virol 7:37–61. https://doi.org/10.1146/annurev-virology-122019 -070009.
- 3. Jacobs-Sera D, Abad LA, Alvey RM, Anders KR, Aull HG, Bhalla SS, Blumer LS, Bollivar DW, Bonilla JA, Butela KA, Coomans RJ, Cresawn SG, D'Elia T, Diaz A, Divens AM, Edgington NP, Frederick GD, Gainey MD, Garlena RA, Grant KW, Gurney SMR, Hendrickson HL, Hughes LE, Kenna MA, Klyczek KK, Kotturi H, Mavrich TN, McKinney AL, Merkhofer EC, Moberg Parker J, Molloy SD, Monti DL, Pape-Zambito DA, Pollenz RS, Pope WH, Reyna NS, Rinehart CA, Russell DA, Shaffer CD, Sivanathan V, Stoner TH, Stukey J, Sunnen CN, Tolsma SS, Tsourkas PK, Wallen JR, Ware VC, Warner MH, Washington JM, Westover KM, Whitefleet-Smith JL, Wiersma-Koch HI, Williams DC, Zack KM, Hatfull GF. 2020. Genomic diversity of bacteriophages infecting *Microbacterium* spp. Plos One 15:e0234636. https://doi.org/10.1371/journal.pone.0234636.
- 4. Klyczek KK, Bonilla JA, Jacobs-Sera D, Adair TL, Afram P, Allen KG, Archambault ML, Aziz RM, Bagnasco FG, Ball SL, Barrett NA, Benjamin RC, Blasi CJ, Borst K, Braun MA, Broomell H, Brown CB, Brynell ZS, Bue AB, Burke SO, Casazza W, Cautela JA, Chen K, Chimalakonda NS, Chudoff D, Connor JA, Cross TS, Curtis KN, Dahlke JA, Deaton BM, Degroote SJ, DeNigris DM, DeRuff KC, Dolan M, Dunbar D, Egan MS, Evans DR, Fahnestock AK, Farooq A, Finn G, Fratus CR, Gaffney BL, Garlena RA, Garrigan KE, Gibbon BC, Goedde MA, Guerrero Bustamante CA, Harrison M, Hartwell MC, Heckman EL, Huang J, Hughes LE, Hyduchak KM, Jacob AE, Kaku M, Karstens AW, et al. 2017. Tales of diversity: genomic and morphological characteristics of forty-six *Arthrobacter* phages. PLoS One 12:e0180517. https://doi.org/10.1371/journal.pone.0180517.
- 5. Jordan TC, Burnett SH, Carson S, Caruso SM, Clase K, DeJong RJ, Dennehy JJ, Denver DR, Dunbar D, Elgin SCR, Findley AM, Gissendanner CR, Golebiewska UP, Guild N, Hartzog GA, Grillo WH, Hollowell GP, Hughes LE, Johnson A, King RA, Lewis LO, Li W, Rosenzweig F, Rubin MR, Saha MS, Sandoz J, Shaffer CD, Taylor B, Temple L, Vazquez E, Ware VC, Barker LP, Bradley KW, Jacobs-Sera D, Pope WH, Russell DA, Cresawn SG, Lopatto D, Bailey CP, Hatfull GF. 2014. A broadly implementable research course in phage discovery and genomics for first-year undergraduate students. mBio 5:e01051-13. https://doi.org/10.1128/mBio.01051-13.
- Ackermann HW. 1998. Tailed bacteriophages: the order Caudovirales. Adv Virus Res 51:135–201. https://doi.org/10.1016/s0065-3527(08)60785-x.

- Miller JR, Koren S, Sutton G. 2010. Assembly algorithms for next-generation sequencing data. Genomics 95:315–327. https://doi.org/10.1016/j.ygeno.2010 .03.001.
- Gordon D, Green P. 2013. Consed: a graphical editor for next-generation sequencing. Bioinformatics 29:2936–2937. https://doi.org/10.1093/bioinformatics/ btt515.
- 9. Russell DA, Hatfull GF. 2017. PhagesDB: the actinobacteriophage database. Bioinformatics 33:784–786. https://doi.org/10.1093/bioinformatics/btw711.
- Pope WH, Mavrich TN, Garlena RA, Guerrero-Bustamante CA, Jacobs-Sera D, Montgomery MT, Russell DA, Warner MH, Science Education Alliance-Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES), Hatfull GF. 2017. Bacteriophages of *Gordonia* spp. display a spectrum of diversity and genetic relationships. mBio 8:e01069-17. https://doi.org/10.1128/mBio.01069-17.
- Besemer J, Borodovsky M. 2005. GeneMark: web software for gene finding in prokaryotes, eukaryotes and viruses. Nucleic Acids Res 33:W451–W454. https:// doi.org/10.1093/nar/gki487.
- 12. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679. https://doi.org/10.1093/bioinformatics/btm009.
- Pope WH, Jacobs-Sera D. 2018. Annotation of bacteriophage genome sequences using DNA Master: an overview, p 217–229. *In* Clokie MRJ, Kropinski AM, Lavigne R (ed), Bacteriophages: methods and protocols, vol 3. Springer, New York, NY.
- Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. BMC Bioinformatics 12:395. https://doi.org/10.1186/1471-2105-12-395.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. https://doi.org/10.1093/nar/gkh152.
- Söding J, Biegert A, Lupas AN. 2005. The HHpred interactive server for protein homology detection and structure prediction. Nucleic Acids Res 33:W244–W248. https://doi.org/10.1093/nar/gki408.
- Krogh A, Larsson B, von Heijne G, Sonnhammer ELL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. J Mol Biol 305:567–580. https://doi.org/10.1006/jmbi .2000.4315.