



## Draft Genome Sequences of Four Antibiotic-Resistant Salmonella Strains Isolated from Australian Red Meat Animal Species

Amreeta Sarjit,<sup>a,c</sup> DJoshua T. Ravensdale,<sup>a</sup> Ranil Coorey,<sup>b</sup> Narelle Fegan,<sup>c</sup> Gary A. Dykes<sup>a</sup>

<sup>a</sup>School of Public Health, Curtin University, Bentley, Western Australia, Australia <sup>b</sup>School of Molecular and Life Sciences, Curtin University, Bentley, Western Australia, Australia <sup>c</sup>CSIRO Agriculture and Food, Brisbane, Queensland, Australia

**ABSTRACT** The genome sequences of four antibiotic-resistant *Salmonella* strains isolated from red meat animals in Australia are presented. Multidrug-resistant *Salmonella enterica* serovar Heidelberg 329 and *Salmonella enterica* serovar Typhimurium 2470 harbored an IncHI2 plasmid similar to the multidrug-resistant *S*. Heidelberg strain N13-01290 plasmid pN13-01290\_23 previously isolated in Canada.

Salmonella enterica is a leading cause of human foodborne bacterial gastroenteritis worldwide (1). The widespread use of antibiotics in animal husbandry has resulted in multiantibiotic-resistant *Salmonella* strains in many countries, but their presence in Australia is generally low (2).

Forty-nine Salmonella strains consisting of Salmonella enterica serovars Anatum, Bovismorbificans, Heidelberg, Newport, Saintpaul, Typhimurium, and Virchow, isolated from different red meat animals consisting of cattle, sheep, and goats in Australia from 2001 to 2013, were isolated as described previously (3-5). All strains were grown in tryptone soy broth (TSB; Oxoid, UK) at 37°C for 24 h for genomic DNA extraction. Genomic DNA was extracted and purified using the Wizard genomic DNA purification kit (Promega, USA), according to the manufacturer's instructions. The purity and concentration of the genomic DNA were determined using agarose gel electrophoresis and by a Qubit 2.0 fluorometer. The genomic DNA was fragmented and tagged using a Nextera XT DNA library preparation version 3 600-cycle MiSeq kit, followed by whole-genome sequencing on an Illumina MiSeq V3 sequencer (2 imes 300-bp reads). The quality of the reads was determined using the BayesHammer error correction tool to improve the quality of the assembly. Genomes were assembled using SPAdes, and the genome qualities of the final assemblies were confirmed using the Quality Assessment Tool (QUAST) for genome assembly algorithm, followed by annotation using Rapid Annotations using Subsystems Technology (RAST) and the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (6-8). Antimicrobial resistance genes were determined using ResFinder (9). The genome sequences of three strains of S. Heidelberg (329 and 632 from goat rumen and 2581 from cattle feces) and one strain of S. Typhimurium (2470 from cattle feces), which were the only strains found to have antibiotic resistance genes, are presented here. Plasmid replicons were determined using PlasmidFinder (10). Plasmid contigs were identified using a local BLAST search against a database of known plasmids in PATRIC and mlplasmids (11, 12). Default parameters were used for all software, unless otherwise specified.

Genome features, including the total number of sequence reads, number of contigs,  $N_{50}$ ,  $L_{50}$ , assembly size, GC content, and numbers of coding sequences (CDS), tRNAs, rRNAs, and plasmids across the four *Salmonella* genomes are shown in Table 1. The

Citation Sarjit A, Ravensdale JT, Coorey R, Fegan N, Dykes GA. 2019. Draft genome sequences of four antibiotic-resistant *Salmonella* strains isolated from Australian red meat animal species. Microbiol Resour Announc 8:e00925-19. https://doi.org/10.1128/ MRA.00925-19.

**Editor** Steven R. Gill, University of Rochester School of Medicine and Dentistry

**Copyright** © 2019 Sarjit et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Gary A. Dykes, gary.dykes@curtin.edu.au.

Received 31 July 2019 Accepted 4 August 2019 Published 29 August 2019

Parameter	Data for strain:						
	S. Heidelberg 329	S. Heidelberg 632	S. Heidelberg 2581 <sup>a</sup>	S. Typhimurium 2470			
Source	Goat rumen	Goat rumen	Cattle feces	Cattle feces			
Date of isolation (day/mo/yr)	13/12/2001	27/2/2002	27/8/2013	26/2/2013			
Total no. of sequence reads	3,047,918	1,395,904	1,851,866	1,790,304			
No. of contigs	69	34	28	63			
N <sub>50</sub> (bp)	191,631	740,277	641,189	231,719			
L <sub>50</sub>	9	2	2	7			
Assembly length (bp)	5,069,302	4,780,505	4,685,585	5,071,508			
GC content (%)	51.9	52.1	52.2	51.9			
No. of CDS	5,172	4,773	4,654	5,181			
No. of tRNAs	81	79	79	82			
No. of rRNAs	12	13	13	11			
No. of plasmids	3	3	0	2			
Accession no.	SWJR0100000	SWMX0000000	SWMW0000000	SWMV0000000			
Contig accession no. (plasmid)	SWJR01000007 (p329_1), SWJR01000047 (p329_2), SWJR01000048 (p329_3)	SWMX01000015 (p632_1), SWMX01000017 (p632_2), SWMX01000020 (p632_3)	NA	SWMV01000009 (p2470_1), SWMV01000043 (p2470_2)			
Sequence Read Archive accession no.	PRJNA534061	PRJNA540014	PRJNA540012	PRJNA540010			

<b>TABLE 1</b> Summary	v of the whole-c	enome features of	S. Heidelberg	and S. T	vphimurium	strains
------------------------	------------------	-------------------	---------------	----------	------------	---------

<sup>a</sup> NA, not applicable.

sources and dates of isolation of the four *Salmonella* strains are also shown in Table 1. A 100% orthologous average nucleotide identity was identified between the genomes of *S*. Heidelberg 329 and *S*. Typhimurium 2470 and between *S*. Heidelberg 632 and 2581, using the OrthoANI algorithm (13). This suggests a close phylogenetic relationship between the two sets of strains.

The S. Heidelberg 329 and S. Typhimurium 2470 plasmids belong to the IncHI2 group and that of S. Heidelberg 632 to the IncII group. IncHI2 is a plasmid lineage associated with the spread of antibiotic-resistant Salmonella spp. (1). S. Heidelberg 329 and S. Typhimurium 2470 harbored a plasmid similar to S. Heidelberg N13-01290/pN13-01290\_23 (GenBank accession number CP012931), isolated from turkey meat in Canada (14). A nucleotide BLAST search of specific contigs in S. Heidelberg 329 (p329\_1) and S. Typhimurium 2470 (p2470\_1) against the pN13-01290\_23 sequence revealed 99% identity to both strain contigs with 81% coverage. S. Heidelberg pN13-01290\_23 belongs to the IncHI2 group and carries aph(3")-Ib and aph(6)-Id (aminoglycoside resistance), *bla*<sub>TEM-1B</sub> (beta-lactam resistance), *sul1* (sulfonamide resistance), and tet(B) (tetracycline resistance) genes. S. Heidelberg 329 and S. Typhimurium 2470 have several antibiotic resistance genes, including aac(6')-laa, aph(3")-lb, aph(6)-ld, aadA1, *bla*<sub>TEM-1B</sub>, *mph*(B) (macrolide resistance), *sul1* and *sul2* (sulfonamide resistance), *tet*(A) (tetracycline resistance), and dfrA1 (trimethoprim resistance). All of these antibiotic resistance genes were located on the matching contig of pN13-01290\_23, with the exception of the beta-lactam resistance gene, which was located in a different contig.

*S.* Heidelberg 632 and 2581 carry *aac(6')-laa* (aminoglycoside resistance gene) and *fosA7* (fosfomycin resistance gene) only on their chromosomes. A previous study reported that the *fosA7* gene was found on the chromosome in *S*. Heidelberg isolated from poultry in Canada, although this gene is rarely reported among *Salmonella* spp. (15). Multidrug resistance efflux pump genes, such as *acrA* and *mdfA*, were detected on the chromosome in all *Salmonella* strains used in this study. Further understanding of antibiotic-resistant *Salmonella* spp. is required.

**Data availability.** The whole-genome shotgun project sequences of *Salmonella* strains 329, 632, 2581, and 2470 have been deposited at DDBJ/ENA/GenBank under the accession numbers listed in Table 1.

## ACKNOWLEDGMENT

This work was funded by the Australian Meat Processors Corporation (AMPC). No specific grant number is associated with this funding.

## REFERENCES

- Chen W, Fang T, Zhou X, Zhang D, Shi X, Shi C. 2016. IncHI2 plasmids are predominant in antibiotic-resistant *Salmonella* isolates. Front Microbiol 7:1566. https://doi.org/10.3389/fmicb.2016.01566.
- Barlow RS, McMillan KE, Duffy LL, Fegan N, Jordan D, Mellor GE. 2015. Prevalence and antimicrobial resistance of *Salmonella* and *Escherichia coli* from Australian cattle populations at slaughter. J Food Prot 78: 912–920. https://doi.org/10.4315/0362-028X.JFP-14-476.
- Fegan N, Vanderlinde P, Higgs G, Desmarchelier P. 2004. Quantification and prevalence of *Salmonella* in beef cattle presenting at slaughter. J Appl Microbiol 97:892–898. https://doi.org/10.1111/j.1365-2672 .2004.02380.x.
- Duffy LL, Small A, Fegan N. 2010. Concentration and prevalence of *Escherichia coli* O157 and *Salmonella* serotypes in sheep during slaughter at two Australian abattoirs. Aust Vet J 88:399–404. https://doi.org/ 10.1111/j.1751-0813.2010.00623.x.
- Duffy L, Barlow R, Fegan N, Vanderlinde P. 2009. Prevalence and serotypes of *Salmonella* associated with goats at two Australian abattoirs. Lett Appl Microbiol 48:193–197. https://doi.org/10.1111/j.1472-765X.2008.02501.x.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. https:// doi.org/10.1038/srep08365.
- Klimke W, Agarwala R, Badretdin A, Chetvernin S, Ciufo S, Fedorov B, Kiryutin B, O'Neill K, Resch W, Resenchuk S, Schafer S, Tolstoy I, Tatusova T. 2009. The National Center for Biotechnology Information's protein clusters database. Nucleic Acids Res 37:D216–D223. https://doi.org/10 .1093/nar/gkn734.

- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https://doi .org/10.1093/jac/dks261.
- Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. PlasmidFinder and pMLST: in silico detection and typing of plasmids. Antimicrob Agents Chemother 58:3895–3903. https://doi.org/10.1128/AAC.02412-14.
- Wattam AR, Abraham D, Dalay O, Disz TL, Driscoll T, Gabbard JL, Gillespie JJ, Gough R, Hix D, Kenyon R, Machi D, Mao C, Nordberg EK, Olson R, Overbeek R, Pusch GD, Shukla M, Schulman J, Stevens RL, Sullivan DE, Vonstein V, Warren A, Will R, Wilson MJ, Yoo HS, Zhang C, Zhang Y, Sobral BW. 2014. PATRIC, the bacterial bioinformatics database and analysis resource. Nucleic Acids Res 42:D581–D591. https://doi.org/10 .1093/nar/gkt1099.
- Arredondo-Alonso S, Rogers MRC, Braat JC, Verschuuren TD, Top J, Corander J, Willems RJL, Schürch AC. 2018. mlplasmids: a user-friendly tool to predict plasmid- and chromosome-derived sequences for single species. Microb Genom 4:1–15. https://doi.org/10.1099/mgen.0.000224.
- Lee I, Kim YO, Park SC, Chun J. 2016. OrthoANI: an improved algorithm and software for calculating average nucleotide identity. Int J Syst Evol Microbiol 66:1100–1103. https://doi.org/10.1099/ijsem.0.000760.
- Labbé G, Edirmanasinghe R, Ziebell K, Nash JHE, Bekal S, Parmley EJ, Mulvey MR, Johnson RP. 2016. Complete genome and plasmid sequences of three Canadian isolates of Salmonella enterica subsp. enterica serovar Heidelberg from human and food sources. Genome Announc 4:e01526-15. https://doi.org/10.1128/genomeA.01526-15.
- Dhanani AS, Block G, Dewar K, Forgetta V, Topp E, Beiko RG, Diarra MS. 2015. Genomic comparison of non-typhoidal *Salmonella enterica* serovars Typhimurium, Enteritidis, Heidelberg, Hadar and Kentucky Isolates from broiler chickens. PLoS One 10:e0128773. https://doi.org/10.1371/ journal.pone.0128773.