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Influence of angiotensin converting enzyme (*ACE*) gene rs4362 polymorphism on the progression of kidney failure in patients with autosomal dominant polycystic kidney disease (ADPKD)

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Background & objectives: Autosomal dominant polycystic kidney disease (ADPKD) is an inherited systemic disorder, characterized by the fluid filled cysts in the kidneys leading to end stage renal failure in later years of life. Hypertension is one of the major factors independently contributing to the chronic kidney disease (CKD) progression. The renin-angiotensin aldosterone system (RAAS) genes have been extensively studied as hypertension candidate genes. The aim of the present study was to investigate the role of angiotensin converting enzyme tagging - single nucleotide polymorphisms (*ACE* tag-SNPs) in progression of CKD in patients with ADPKD.

Methods: In the present study six *ACE* tagSNPs (angiotensin converting enzyme tag single nucleotide polymorphisms) and insertion/deletion (I/D) in 102 ADPKD patients and 106 control subjects were investigated. The tagSNPs were genotyped using FRET-based KASPar method and *ACE* ID by polymerase chain reaction (PCR) and electrophoresis. Genotypes and haplotypes were compared between ADPKD patients and controls. Univariate and multivariate logistic regression analyses were performed to assess the effect of genotypes and hypertension on CKD advancement. Mantel-Haenszel (M-H) stratified analysis was performed to study the relationship between different CKD stages and hypertension and their interaction.

Results: All loci were polymorphic and except rs4293 SNP the remaining loci followed Hardy-Weinberg equilibrium. Distribution of *ACE* genotypes and haplotypes in controls and ADPKD patients was not significant. A significant linkage disequilibrium (LD) was observed between SNPs forming two LD blocks. The univariate analysis revealed that the age, hypertension, family history of diabetes and *ACE* rs4362 contributed to the advancement of CKD.

Interpretation & conclusions: The results suggest that the *ACE* genotypes are effect modifiers of the relationship between hypertension and CKD advancement among the ADPKD patients.

Key words ACE gene - ADPKD - CKD - gene-environment interaction - tagSNP

Autosomal dominant polycystic kidney disease (ADPKD) is the most common inherited systemic disorder, characterized by the fluid filled cysts in the kidneys leading to end stage renal failure in later years of life¹. Although the exact genetic factors that favour renal progression have not been identified, studies on mouse and human models suggest that the genetic variations play a significant role in the differential renal progression that is observed in ADPKD^{2,3}. Intrafamilial heterogeneity that is found in the expression of ADPKD, suggests the involvement of environmental factors along with the genetic factors in the progression of ADPKD⁴. Polycystic kidney disease genes products share sequence homology and are thought to play an important role in cell-cell and cell-matrix interactions and in structure of a receptor-channel complex which regulates renal ion transport⁵. Patients with ADPKD exhibit non-uniformity of the renal disease progression. This variability was observed from both mouse² and human⁶ models, suggesting that disease-modifying loci may play a role in the variable renal progression found in ADPKD.

Hypertension is one of the clinical manifestations in majority of ADPKD cases and correlates with the progressive kidney enlargement with high prognostic values. The renin-angiotensin aldosterone system (RAAS) is known to regulate the blood pressure and fluid balance⁷. The activity of RAAS system is regulated by the rate of production of angiotensinogen from rennin that is mediated by angiotensin converting enzyme (ACE)⁸. In view of the role of hypertension on the progression of *chronic kidney disease* in ADPKD, the gene polymorphisms of *ACE* are of great interest.

The *ACE* gene spans over 24 kb with 26 exons and is located on chromosome 17. The presence or absence of a 287-bp repeat sequence [insertion/deletion (I/D) polymorphism] at intron 16 has been used as a common marker in the susceptibility of various disorders⁹. An initial study showed a deleterious effect of the DD genotype on renal survival in ADPKD¹⁰. The direct biological mechanism by which *ACE* ID polymorphism might influence serum ACE levels remains unclear. In the present study the role of *ACE* ID and six additional tagging-single nucleotide polymorphisms (tagSNPs) were investigated to unravel the ACE gene modifier effect for renal disease progression in patients with ADPKD.

Material & Methods

A total of 102 consecutive patients affected by ADPKD (55.88% men) and 106 controls (60.38% men) attending the outpatient department of Nephrology, SriRamachandraUniversity, Chennai, India, from March 2010 to December 2012 were consecutively selected. The diagnosis of ADPKD was based on previously described Ravine ultrasound criteria¹¹. Patients with other conditions that could influence renal function such as diabetes or pregnancy were excluded. Upon screening, a total of 106 healthy unrelated individuals without family history of polycystic kidney disease or any other kidney related complications (60.38% men) were included as a control. All control subjects were non-diabetic, normotensive and aged between 30-60 yr. The study was approved by the Institutional Ethics Committee of the Sri Ramachandra University, Chennai, India. A written informed consent was obtained from the study participants. The total number of cysts in the ADPKD patients was detected by using ultrasound imaging and the estimated glomerular filtration rate (eGFR) was assessed based on Modification of Diet in Renal Disease (MDRD) formula¹². Further, the stage of kidney disease was determined by using eGFR and all the ADPKD patients were divided into two groups: early stages (CKD stages 1-3; eGFR 30 to 90 ml/min/1.73 m²) and advanced (CKD stages 4; eGFR 15-29 ml/min/1.73 m² and 5; eGFR < 15 ml/min/1.73 m²) stages. Genomic DNA from the peripheral blood samples was extracted by phenol chloroform extraction and ethanol precipitation protocol¹³.

SNP selection and genotyping: All tagging single nucleotide polymorphisms (tagSNPs) covering ACE gene from the release 2.0 Phase II data of the HapMap Project (www.hapmap.org) were selected using pairwise tagging method¹⁴. The tagSNPs were chosen according to the following criterion: $r^2 \ge 0.8$ and minor allele frequency of ≥ 5 per cent in the Gujarati Indians in Houston (GIH) population^{14,15}. A common well studied ID polymorphism was also included⁸. Fluorescence resonance energy transfer (FRET)-based KASPar SNP genotyping assay method (KBioscience, Herts, UK) was used to genotype the tag-SNPs on an Applied Biosystems thermocycler (ABI Prism 9700, Foster City, CA, USA) using the fluorescent probe primers (designed by K Bioscience, UK using KrakenTM software system). The fluorescent endpoint readings were taken using the 7900 SDS software (ABI Prism 7900, Foster City, CA, USA). The genotyping success rate ranged from 99.3 to 100.0 per cent. The polymerase chain reaction (PCR) and electrophoresis were used to detect the ACE gene insertion deletion polymorphism⁸. All homozygous deletion samples were further subjected to a second PCR amplification with insertion-specific primers to prevent mistyping of heterozygous genotypes¹⁶.

Statistical analysis: Allele frequencies were calculated by the gene-counting method¹⁷. Genotypes at each SNP were tested for Hardy-Weinberg equilibrium by using χ^2 test with one degree of freedom. Chi square test was carried out to check the association between ADPKD and controls. Pairwise linkage disequilibrium (LD) measures (D' and r²) and haplotype blocks were assessed using the default setting of the Haploview software¹⁴. In ADPKD patients univariate and multivariate logistic regression analysis was performed to assess the effect of genotypes and hypertension on CKD progression. The influence of different genotypes on the relationship between different CKD stages and hypertension and their interaction was examined using the Mantel-Haenszel stratified analysis. All statistical analyses were performed using SPSS 16.0 (SPSS, Inc., Chicago, USA).

Results

ACE gene polymorphisms and ADPKD: Clinical characteristics of both controls and ADPKD patients are given in Table I. The patients had significantly higher (P<0.001) creatinine and blood urea nitrogen levels and significantly (P<0.001) lower eGFR and calcium levels as compared to controls. For ACE gene GIH population yielded six tagSNPs (rs4293, rs4309, rs4311, rs4325, rs4329 and rs4362)¹⁵. The allele and genotype frequencies of the six tagSNPs along with insertion deletion polymorphisms in patients and controls are shown in Table II. All polymorphisms except rs4293 followed Hardy-Weinberg equilibrium. Distribution of ACE genotypes between the control and ADPKD groups was not significant (Table II). Analysis of LD showed a strong and significant LD between the markers by forming two LD blocks (Figure). The first LD block included rs4311 and rs4325, which were separated by 2.4 kb in introns 9 and 13, respectively. The second LD block included rs4329, I/D and rs4362, which encompassed intron 13, intron 16 and

dominant polycystic kidney disease (ADPKD) and controls				
Variables	Controls (n=106)	ADPKD patients (n=102)		
Age (yr)	53.97 ± 12.80	$46.92 \pm 11.60^{***}$		
Haemoglobin (g/dl)	11.73 ± 2.56	$10.67 \pm 2.05^{***}$		
BUN (mg/dl)	13.35 ± 7.01	$28.41 \pm 24.79^{***}$		
Creatinine (mg/dl)	0.95 ± 0.21	$3.02\pm 2.58^{***}$		
Sodium (mmol/l)	136.45 ± 5.78	137.11 ± 8.74		
Potassium (mmol/l)	3.96 ± 0.88	$4.49\pm2.13^{\ast}$		
Chloride (mmol/l)	101.13 ± 6.09	101.26 ± 11.84		
Bicarbonate (mmol/l)	24.51 ± 3.69	$22.86 \pm 4.20^{**}$		
Calcium (mg/dl)	10.16 ± 1.40	$7.71 \pm 1.14^{***}$		
eGFR (ml/min/1.73 m ²)	82.57 ± 20.72	$43.79 \pm 34.65^{***}$		
BUN, blood urea nitrogen; eGFR, estimated glomerular filtration rate $P^*<0.05$, **<0.01, ***<0.001 compared to controls				

Table I. Clinical characteristics of patients with autosomal

exon 21, respectively. The rs4309 SNP located in exon 8 remained outside the LD blocks. Haplotypes constructed using the SNPs located in individual LD blocks are presented in the Figure. Comparison of haplotypes between ADPKD and control groups revealed absence of any significant association with ADPKD.

ACE gene polymorphisms CKD advancement: Among ADPKD patients, 49 (48%) showed advanced CKD stage with mean age of 50.73 ± 10.27 yr and 53 (52%)showed moderate progression with 43.4 ± 11.74 yr of age. Distribution of genotypes between these CKD groups revealed that only rs4362 was associated with the development of CKD stage (P=0.046). The TT genotype significantly increased the risk of advanced CKD in ADPKD (OR=4.97; 95% CI: 1.06-32.52) (Table III). The univariate analysis revealed that the age, hypertension and family history of diabetes contributed to the significant advancement of CKD, but the gender had no effect on CKD advancement (Table III). No evidence of heterogeneity of the effect of hypertension on CKD stages was observed among different genotypes of the studied polymorphisms. This indicated the absence of confounding effect of genotype on the relationship between CKD advancement and hypertension. For rs4362 genotypes the Mantel-Haenszel (M-H) combined OR for hypertension was

SNP	Genotype	Control (n=106)	ADPKD patients (n=102)	OR (95% CI)	P value
rs4293	AA	47 (44.3)	31 (30.3)	Reference	
	AG	37 (34.9)	45 (44.1)	1.84 (0.98-3.46)	
	GG	22 (20.7)	26 (25.5)	1.79 (0.87-3.71)	0.115
HWp		0.007	0.24		
rs4309	TT	40 (37.7)	29 (28.4)	Reference	
	TC	42 (39.6)	43 (42.1)	1.41 (0.75-2.68)	
	CC	24 (22.6)	30 (29.4)	1.72 (0.84-3.54)	0.307
HWp		0.052	0.11		
rs4311	CC	51 (48.1)	43 (42.1)	Reference	
	TC	39 (36.7)	41 (40.2)	1.25 (0.69-2.27)	
	TT	16 (15.0)	18 (17.6)	1.33 (0.61-2.93)	0.68
HWp		0.073	0.143		
rs4325	AA	38 (35.8)	25 (24.5)	Reference	
	CA	43 (40.5)	48 (47.0)	1.70 (0.89-3.25)	
	CC	25 (23.5)	29 (28.4)	1.76 (0.85-3.68)	0.203
HWp		0.069	0.56		
rs4329	GG	43 (40.5)	32 (31.3)	Reference	
	GA	42 (39.6)	50 (49.0)	1.60 (0.87-2.96)	
	AA	21(19.98)	20 (19.6)	1.28 (0.60-2.75)	0.323
HWp		0.076	0.952		
ACE I/D	II	43 (40.5)	32 (31.3)	Reference	
	ID	42 (39.6)	50 (49.0)	1.60 (0.87-2.96)	
	DD	21 (19.98)	20 (19.6)	1.28 (0.60-2.75)	0.323
HWp		0.076	0.952		
rs4362	CC	45 (42.4)	36 (35.2)	Reference	
	TC	47 (44.3)	52 (50.9)	1.38 (0.77-2.49)	
	TT	14 (13.2)	14 (13.7)	1.25 (0.53-2.96)	0.554
HWp		0.755	0.483		

found to be 4.81 and 95 per cent CI is 1.26-18.4 (Table IV). Both univariate (OR=4.97; 95% CI: 1.06-32.52) (Table III) and multivariate analyses (OR=17.04; 95% CI: 2.38-121.95) revealed that the TT genotype of rs4362 increased the risk of CKD advancement in ADPKD after controlling age, sex, hypertension and family history of diabetes (Table V). The adjusted effect

of hypertension on CKD advancement was found to be independent of other risk factors (OR=6.63; CI: 1.38-31.81).

Discussion

Analysis of tagSNPs within the *ACE* gene in ADPKD patients and control subjects did not show

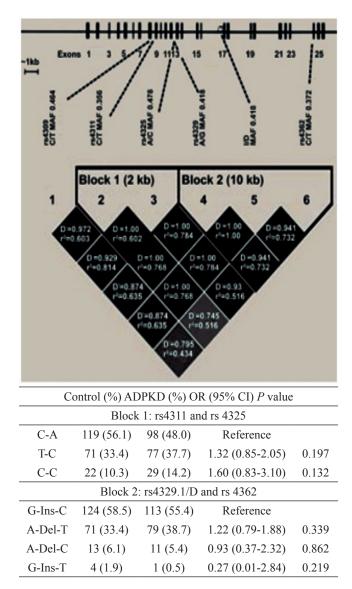


Figure. Linkage disequilibrium (LD) map and haplotype analysis of ACE. Relative positions of the tagSNPs and their MAFs were shown in the upper portion. Pairwise LD measures (D and r^2) were shown by the LD map. The *ACE* gene haplotypes distribution in control and ADPKD patients was shown in the bottom.

any significant association with ADPKD. Strong LD was found among all SNPs studied, covering a region of about 13.8 kb within the *ACE* gene. Comparison of haplotypes between ADPKD and control groups also revealed the absence of a significant association with ADPKD. The univariate analysis revealed that the age, hypertension, family history of diabetes and *ACE* rs4362 contributed to the advancement of CKD. The modifier effect of these factors remained even after controlling other variables in multivariate analysis,

indicating that the *ACE* genotypes are effect modifiers of the relationship between hypertension and CKD advancement among the ADPKD patients.

Our study had some limitations. First, it was a nested study, hence there was a possibility of selection bias. Second, was the limited sample size in case group, the estimated 95 per cent CI of the effect of TT genotype on the advancement of CKD stages was found to be wider. Despite these limitations, analysis of tagSNPs of *ACE* gene revealed that the *ACE* polymorphism had a significant modifier effect on the progression of renal disease in ADPKD.

In ADPKD, hypertension is an early sign that occurs even before the reduction in glomerular filtration rate. Hence hypertension is one of the most important factors that influences the CKD progression in ADPKD^{18,19}. Inappropriate activation of RAAS has been shown to be the major cause in the pathogenesis of hypertension in individuals with ADPKD²⁰. However, the MDRD (Modification of Diet in Renal Disease) study²¹ demonstrated that the ACE inhibitor therapy or blood pressure control did not reduce the rate of decline in GFR of ADPKD patients. In contrast, pharmacological blockade of the RAAS using ACE inhibitors significantly reduced CKD progression²². ACE is suspected to play a key role and is the first candidate modifier gene that has been investigated in ADPKD²³. An initial study of ACE gene found that the DD genotype increased the risk of developing end stage renal failure at an early age in PKD1 patients⁹. Subsequent studies on the influence of ACE I/D polymorphism on ADPKD progression have yielded conflicting results²³⁻²⁵. A comprehensive meta-analysis encompassing all relevant studies did not support the hypothesis that the DD genotype increased risk of developing end stage renal failure in patients with ADPKD²⁶.

The synonymous marker rs4362 (C19329T) located within the 5'end of exon 23 was considered the functional candidate. It has been shown that the rs4362 marks the angiotensin-converting enzyme gene insertion-deletion variant²⁷. A study conducted in Mexican individuals²⁸ revealed that the rs4362 polymorphisms were associated with an increased risk of hypertension and ACE serum levels. These results suggest that the rs4362 polymorphisms can modify the renal function and leads to CKD progression by altering the plasma ACE levels.

Variants/	Genotype	CKD stages			Р	
variables	-	Early stage N (%)	Advanced stage N (%)	Univariate OR (95% CI)	value	
rs4293	AA	19 (61.3)	12 (38.7)	1	0.443	
	GA	21 (46.7)	24 (53.3)	1.80 (0.65,5.11)		
	GG	13 (50)	13 (50)	1.57 (0.49,5.18)		
rs4309	TT	14 (46.7)	16 (53.3)	1	0.429	
	TC	21 (48.8)	22 (51.2)	0.92 (0.32,2.58)		
	CC	18 (62.1)	11 (37.9)	0.54 (0.17,1.70)		
rs4311	CC	27 (62.8)	16 (37.2)	1	0.151	
	TC	19 (46.3)	22 (53.7)	1.94 (0.75,5.12)		
	TT	7 (38.9)	11 (61.1)	2.61 (0.75,9.73)		
rs4325	AA	16 (64)	9 (36)	1	0.347	
	CA	24 (50)	24 (50)	1.76 (0.59,5.49)		
	CC	13 (44.8)	16 (55.2)	2.16 (0.64,7.59)		
rs4329	GG	19 (59.4)	13 (40.6)	1	0.213	
	GA	27 (54)	23 (46)	1.24 (0.46,3.39)		
	AA	7 (35)	13 (65)	2.66 (0.74,10.27)		
ACE I/D	II	19 (59.4)	13 (40.6)	1	0.213	
	ID	27 (54)	23 (46)	1.24 (0.46,3.39)		
	DD	7 (35)	13 (65)	2.66 (0.74,10.27)		
cs4362	CC	21 (58.3)	15 (41.7)	1	0.046	
	TC	29 (55.8)	23 (44.2)	1.11 (0.43,2.87)		
	TT	3 (21.4)	11 (78.6)	4.97 (1.06,32.52)		
Age	≤40	23 (71.9)	9 (28.1)	1	0.015	
	(40-60)	26 (45.6)	31 (54.4)	3.01 (1.11,8.77)		
	>60	4 (30.8)	9 (69.2)	5.50 (1.17,31.17)		
Sex	F	21 (46.7)	24 (53.3)	1	0.452	
	М	32 (56.1)	25 (43.9)	0.68 (0.29,1.61)		
HT	No	13 (76.5)	4 (23.5)	1	0.027	
	Yes	40 (47.1)	45 (52.9)	3.66 (1.01,16.45)		
FH-DM	No	36 (66.7)	18 (33.3)	1	0.002	
	yes	17 (35.4)	31 (64.6)	3.65 (1.49,8.98)		

A study which examined the detailed renal functions of young ADPKD patients showed abnormal kidney function at younger age group $(27 \pm 5 \text{ yr})$ and GFR decreased but was not significantly different from that of the normal healthy controls²⁹. Moreover, the measured eGFR in relation to age in the hypertensive

children with ADPKD was an observed risk for increased renal volume and decreased renal function as compared children with normal blood pressure³⁰. These results suggest that the renal compensation mechanism might terminate as the age increases and with the acceleration of associated risk factors.

stages and hyper Gene	rtension stra Genotype	$\frac{\text{trifted by } ACE \text{ genotyp}}{\text{OR (95\% CI)}}$	P value [*]
Gene	Genotype	for HT	1 value
rs4293	AA	3.78 (0.346,202.2)	0.624
	GA	6.9 (0.68,353.7)	
	GG	1.62 (0.151,23.2)	
M-H combined		3.55 (1.063,11.9)	
rs4309	TT	1.87 (0.18,26.1)	0.61
	ТС	8.03 (0.84,403.4)	
	CC	2.77 (0.226,154.4)	
M-H combined		3.65 (1.096,12.1)	
rs4311	TT	6.1 (0.69,298.1)	0.427
	TC	5.38 (0.47,288.1)	
	CC	0.76 (0.01,17.9)	
M-H combined		3.62 (1.05,12.5)	
rs4325	AA	1.8 (0.12,108.8)	0.514
	CA	9.08 (1.01,443.8)	
	CC	2.05 (0.2,28.8)	
M-H combined		3.68 (1.11,12.3)	
rs4329	GG	3.1 (0.26,170.3)	0.412
	GA	8.92 (1.04,428.3)	
	AA	0.92 (0.01,21.4)	
M-H combined		3.85 (1.11,13.3)	
ACE I/D	II	3.1 (0.26,170.3)	0.412
	ID	8.92 (1.04,428.3)	
	DD	0.92 (0.01,21.4)	
M-H combined		3.85 (1.11,13.3)	
rs4362	CC	3.2 (0.27,173.4)	0.717
	TC	8.1 (0.95,387.6)	
	TT	2.11 (0.03,66)	
M-H combined		4.81 (1.26,18.4)	
HT, hypertensio *Homogeneity to OR, odds ratio		ntel-Haenszel	

In conclusion, our study showed that rs4362 polymorphism in *ACE* gene had a significant modifier effect on the progression of renal disease in patients with ADPKD.

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Table V. Adjusted effects of risk factors or	i chronic kidney
disease (CKD) stages	

Factors	OR (95% CI)	P value*	
HT: Yes vs No	6.63 (1.38, 31.81)	0.018	
SEX: M vs F	0.45 (0.16, 1.24)	0.123	
Age: (40,60 yr) vs, ≤40 yr	6.71 (1.99, 22.62)	0.002	
Age: (60,90 yr) vs, ≤40 yr	15.3 (2.44, 96.06)	0.004	
FH-DM: Yes vs No	7.67 (2.54, 23.14)	0.001	
rs4362: TC vs CC	1.14 (0.41, 3.16)	0.801	
rs4362: TT vs CC	17.04 (2.38, 121.95)	0.005	
HT, hypertension; FH-DM, family history of diabetes mellitus; OR, odds ratio; CI, confidence interval *Wald test <i>P</i> value			

Conflicts of Interest: None.

References

- 1. Sun Y, Zhou H, Yang BX. Drug discovery for polycystic kidney disease. *Acta Pharmacol Sin* 2011; *32* : 805-16.
- Wu G, Tian X, Nishimura S, Markowitz GS, D'Agati V, Park JH, *et al.* Trans-heterozygous Pkd1 and Pkd2 mutations modify expression of polycystic kidney disease. *Hum Mol Genet* 2002; *11*: 1845-54.
- Hoefele J, Mayer K, Scholz M, Klein HG. Novel PKD1 and PKD2 mutations in autosomal dominant polycystic kidney disease (ADPKD). *Nephrol Dial Transplant* 2011; 26 : 2181-8.
- Devuyst O. Variable progression of autosomal dominant polycystic kidney disease: genetic and molecular counterparts. *Nephrol Ther* 2006; 2 (Suppl 2): S104-8.
- Koptides M, Deltas CC. Autosomal dominant polycystic kidney disease: molecular genetics and molecular pathogenesis. *Hum Genet* 2000; 107: 115-26.
- 6. Fain PR, McFann KK, Taylor MR, Tison M, Johnson AM, Reed B, *et al.* Modifier genes play a significant role in the phenotypic expression of *PKD1*. *Kidney Int* 2005; *67* : 1256-67.
- Grobe JL, Grobe CL, Beltz TG, Westphal SG, Morgan DA, Xu D, *et al.* The brain renin-angiotensin system controls divergent efferent mechanisms to regulate fluid and energy balance. *Cell Metab* 2010; *12*: 431-42.
- 8. Rigat B, Hubert C, Corvol P, Soubrier F. PCR detection of the insertion/deletion polymorphism of the human angiotensin converting enzyme gene (*DCP1*) (dipeptidyl carboxypeptidase 1). *Nucleic Acids Res* 1992; 20: 1433.
- Mizuiri S, Hemmi H, Kumanomidou H, Iwamoto M, Miyagi M, Sakai K, *et al.* Angiotensin-converting enzyme (ACE) I/D genotype and renal ACE gene expression. *Kidney Int* 2001; 60 : 1124-30.
- Baboolal K, Ravine D, Daniels J, Williams N, Holmans P, Coles GA, *et al.* Association of the angiotensin I converting enzyme gene deletion polymorphism with early onset of ESRF in PKD1 adult polycystic kidney disease. *Kidney Int* 1997; 52: 607-13.

- Ravine D, Gibson RN, Walker RG, Sheffield LJ, Kincaid-Smith P, Danks DM. Evaluation of ultrasonographic diagnostic criteria for autosomal dominant polycystic kidney disease 1. *Lancet* 1994; 343: 824-7.
- Levey AS, Bosch JP, Lewis JB, Greene T, Rogers N, Roth D. A more accurate method to estimate glomerular filtration rate from serum creatinine: a new prediction equation. Modification of Diet in Renal Disease Study Group. *Ann Intern Med* 1999; *130* : 461-70.
- Sambrook J, Russell DW. *Molecular cloning : a laboratory manual*. 3rd ed. Cold Spring Harbor, N.Y.: Cold Spring Harbor Laboratory Press; 2001.
- 14. Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 2005; *21* : 263-5.
- Thorisson GA, Smith AV, Krishnan L, Stein LD. The International Hap Map Project Web site. *Genome Res* 2005; 15: 1592-3.
- Shanmugam V, Sell KW, Saha BK. Mistyping ACE heterozygotes. PCR Methods Appl 1993; 3: 120-1.
- 17. Ceppellini BR, Siniscalco M, Smith CAB. The estimation of gene frequencies in a random-mating population. *Ann Hum Genet* 1955; 20: 97-115.
- Torres VE, Grantham JJ, Chapman AB, Mrug M, Bae KT, King BF Jr, *et al.* Potentially modifiable factors affecting the progression of autosomal dominant polycystic kidney disease. *Clin J Am Soc Nephrol* 2011; 6 : 640-7.
- 19. Schrier RW, Johnson AM, McFann K, Chapman AB. The role of parental hypertension in the frequency and age of diagnosis of hypertension in offspring with autosomal-dominant polycystic kidney disease. *Kidney Int* 2003; *64* : 1792-9.
- Ramanathan G, Elumalai R, Periyasamy S, Lakkakula B. Role of renin-angiotensin-aldosterone system gene polymorphisms and hypertension-induced end-stage renal disease in autosomal dominant polycystic kidney disease. *Iran J Kidney Dis* 2014; 8 : 265-77.
- Levey AS, Greene T, Beck GJ, Caggiula AW, Kusek JW, Hunsicker LG, *et al.* Dietary protein restriction and the progression of chronic renal disease: what have all of the results of the MDRD study shown? Modification of diet in

renal disease study group. *J Am Soc Nephrol* 1999; *10* : 2426-39.

- Lattanzio MR, Weir MR. Does blockade of the reninangiotensin-aldosterone system slow progression of all forms of kidney disease? *Curr Hypertens Rep* 2010; *12*: 369-77.
- Perez-Oller L, Torra R, Badenas C, Mila M, Darnell A. Influence of the *ACE* gene polymorphism in the progression of renal failure in autosomal dominant polycystic kidney disease. *Am J Kidney Dis* 1999; 34 : 273-8.
- Lee KB, Kim UK, Lee CC. Association of the ACE gene polymorphism with the progression of autosomal dominant polycystic kidney disease. J Korean Med Sci 2000; 15: 431-5.
- Gumprecht J, Zychma MJ, Karasek D, Grzeszczak W. ACE gene I/D polymorphism and the presence of renal failure or hypertension in autosomal dominant polycystic kidney disease (ADPKD). Nephrol Dial Transplant 2007; 22: 1483.
- Pereira TV, Nunes AC, Rudnicki M, Magistroni R, Albertazzi A, Pereira AC, et al. Influence of ACE I/D gene polymorphism in the progression of renal failure in autosomal dominant polycystic kidney disease: a meta-analysis. Nephrol Dial Transplant 2006; 21: 3155-63.
- Marciante KD, Bis JC, Rieder MJ, Reiner AP, Lumley T, Monks SA, *et al.* Renin-angiotensin system haplotypes and the risk of myocardial infarction and stroke in pharmacologically treated hypertensive patients. *Am J Epidemiol* 2007; *166* : 19-27.
- Martinez-Rodriguez N, Posadas-Romero C, Villarreal-Molina T, Vallejo M, Del-Valle-Mondragon L, Ramirez-Bello J, *et al.* Single nucleotide polymorphisms of the angiotensinconverting enzyme (ACE) gene are associated with essential hypertension and increased ACE enzyme levels in Mexican individuals. *PLoS One* 2013; 8 : e65700.
- Meijer E, Rook M, Tent H, Navis G, van der Jagt EJ, de Jong PE, *et al.* Early renal abnormalities in autosomal dominant polycystic kidney disease. *Clin J Am Soc Nephrol* 2010; 5 : 1091-8.
- Cadnapaphornchai MA, McFann K, Strain JD, Masoumi A, Schrier RW. Prospective change in renal volume and function in children with ADPKD. *Clin J Am Soc Nephrol* 2009; 4: 820-9.

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