



Complete Genome Sequence of a *Pseudomonas aeruginosa* Isolate from a Kidney Stone

Genevieve Johnson,^a Nicole Stark,^b Alan J. Wolfe,^c  Catherine Putonti^{a,b,c,d}

^aBioinformatics Program, Loyola University Chicago, Chicago, Illinois, USA

^bDepartment of Biology, Loyola University Chicago, Chicago, Illinois, USA

^cDepartment of Microbiology and Immunology, Stritch School of Medicine, Loyola University Chicago, Maywood, Illinois, USA

^dDepartment of Computer Science, Loyola University Chicago, Chicago, Illinois, USA

ABSTRACT Recently, we isolated a temperate bacteriophage, *Pseudomonas* phage Dobby, from a calcium oxalate kidney stone. Here, we present the complete genome of the bacterial host harboring this phage, *Pseudomonas aeruginosa* UMB2738. From the analysis of the genome sequence, five additional prophage sequences were identified.

While bacteria are the cause of magnesium-ammonium-phosphate (struvite) kidney stones (1), they may also contribute to calcium kidney stones (2–5). Studies have successfully cultured bacteria from kidney stones (2–8) and have most frequently isolated *Escherichia coli* and *Pseudomonas* species (9). In a recent study by Barr-Bear et al. (5), *E. coli* and *Pseudomonas aeruginosa* were cultured from two calcium oxalate (CaOx) stones. From one of these stones from a male patient, two *P. aeruginosa* colonies, identified via 16S rRNA gene sequencing, were isolated (5). In one of these isolates, UMB2738, the φ CTX-like temperate *Pseudomonas* phage Dobby was isolated and characterized (10). Similarly, a temperate phage was induced from the other *P. aeruginosa* isolate, UMB2744. This prompted us to sequence both of these bacterial isolates to ascertain first if they belonged to the same strain and, subsequently, to characterize *P. aeruginosa* from CaOx stones.

P. aeruginosa strains UMB2738 and UMB2744 were isolated and cultured using the expanded quantitative urinary culture (EQUC) protocol (11) as described by Barr-Bear et al. (5) and stored at -80°C . From this freezer stock, each isolate was streaked onto a 1.7% LB agar plate and grown overnight at 37°C . A single colony from each plate was selected and grown in LB liquid medium overnight at 37°C with shaking with 5-mm sterile glass beads to minimize biofilm formation. DNA was extracted using the Qiagen DNeasy UltraClean microbial kit and quantified using a Qubit fluorometer. DNA was sent to the Microbial Genomic Sequencing Center (MiGS) at the University of Pittsburgh for sequencing. Briefly, DNA was enzymatically fragmented using an Illumina tagmentation enzyme, and indices were attached using PCR. The barcoded samples were multiplexed on an Illumina NextSeq 500 flow cell, producing 1,053,541 and 943,073 pairs of 151-bp reads for UMB2738 and UMB2744, respectively. Default parameters were used for all software tools unless otherwise noted. Raw reads were trimmed using Sickle v1.33 (<https://github.com/najoshi/sickle>) and then assembled using SPAdes v3.13.0 (parameters, only-assembler; $k = 55,77,99,127$) (12). The assembled contigs were concatenated for each sample and compared via the progressiveMauve algorithm v1.1.1 (13) in Geneious Prime (Biomatters Ltd., Auckland, New Zealand). This comparison revealed that UMB2738 and UMB2744 had identical genomes (pairwise identity = 100%). Thus, the genome was reassembled with all 13,523,760 trimmed reads from both barcodes using SPAdes and the same parameters. Here, we refer to this strain as UMB2738.

Citation Johnson G, Stark N, Wolfe AJ, Putonti C. 2019. Complete genome sequence of a *Pseudomonas aeruginosa* isolate from a kidney stone. Microbiol Resour Announc 8:e01073-19. <https://doi.org/10.1128/MRA.01073-19>.

Editor David A. Baltrus, University of Arizona

Copyright © 2019 Johnson et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Catherine Putonti, cputonti@luc.edu.

G.J. and N.S. contributed equally to this work.

Received 28 August 2019

Accepted 1 September 2019

Published 19 September 2019

The genome assembly of *P. aeruginosa* UMB2738 includes 87 contigs and consists of 6,720,267 bp with a GC content of 66.15%. The genome coverage is 79.35 \times , calculated by BBMap v38.47 (<https://sourceforge.net/projects/bbmap/>), and the assembly has an N_{50} score of 135,797. Genome annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v4.8 (14). No CRISPR/Cas genes or arrays were found within the genome by CRISPRCasFinder (15). The genome encodes 6,298 protein coding genes, three rRNA operons, and 57 tRNAs. As this strain was the source of the temperate *Pseudomonas* phage Dobby (10), the genome assembly was queried via blastn (16) for the Dobby sequence (GenBank accession number [MK034952](#)) and located in the first, largest contig of the assembly. The *P. aeruginosa* UMB2738 genome was also screened for phage sequences using VirSorter v1.0.5 (17). Six prophage sequences were predicted, including the region encoding Dobby. VirSorter also identified *Pseudomonas* phage Pf1 and a relative of *Pseudomonas* phage vB_Pae_CF79a (GenBank accession number [MK510971](#); query coverage, 45%; identity, 97.18%). The other two phage sequences predicted do not exhibit any sequence homology to characterized phage sequences.

Sequencing of this bacterial isolate will inform our future work with *Pseudomonas* phage Dobby, which is just one of the phages harbored by this bacterium. Furthermore, the availability of this genome, representative of bacteria within kidney stones, can add insight into future studies of the urinary tract microbiota.

Data availability. This whole-genome shotgun (WGS) project has been deposited in GenBank under the accession number [VSIZ00000000](#). Raw sequence reads are deposited under accession numbers [SRR9992785](#) and [SRR9992786](#). The WGS and SRA records are associated with BioProject number [PRJNA316969](#).

ACKNOWLEDGMENTS

We thank the authors of the prior study that isolated the bacteria, Evan Barr-Bear, Vijay Saxena, Evann Hilt, Krystal Thomas-White, Megan Schober, Birong Li, Brian Becknell, David Hains, and Andrew Schwaderer. For prior patient recruitment, we acknowledge the Loyola Urinary Education and Research Collaborative (LUEREC), specifically Mary Tulke, Linda Brubaker, Elizabeth Mueller, Cynthia Brincat, Susanne Taege, and Tanaka Dune, and the patients who provided the samples for this study.

G.J. is funded through Loyola's Mulcahy Research Fellowship.

REFERENCES

- Flannigan R, Choy WH, Chew B, Lange D. 2014. Renal struvite stones—pathogenesis, microbiology, and management strategies. *Nat Rev Urol* 11:333–341. <https://doi.org/10.1038/nrurol.2014.99>.
- Thompson RB, Stamey TA. 1973. Bacteriology of infected stones. *Urology* 2:627–633. [https://doi.org/10.1016/0090-4295\(73\)90323-3](https://doi.org/10.1016/0090-4295(73)90323-3).
- Golechha S, Solanki A. 2001. Bacteriology and chemical composition of renal calculi accompanying urinary tract infection. *Indian J Urol* 17:111–117.
- Tavichakorntrakool R, Prasongwattana V, Sungkeeree S, Saisud P, Sribenjalux P, Pimratana C, Bovornpadungkitti S, Sriboonlue P, Thongboonkerd V. 2012. Extensive characterizations of bacteria isolated from catheterized urine and stone matrices in patients with nephrolithiasis. *Nephrol Dial Transplant* 27:4125–4130. <https://doi.org/10.1093/ndt/gfs057>.
- Barr-Bear E, Saxena V, Hilt EE, Thomas-White K, Schober M, Li B, Becknell B, Hains DS, Wolfe AJ, Schwaderer AL. 2015. The interaction between Enterobacteriaceae and calcium oxalate deposits. *PLoS One* 10: e0139575. <https://doi.org/10.1371/journal.pone.0139575>.
- Tavichakorntrakool R, Boonsiri P, Prasongwattana V, Lulitanond A, Wongkham C, Thongboonkerd V. 2017. Differential colony size, cell length, and cellular proteome of *Escherichia coli* isolated from urine vs. stone nidus of kidney stone patients. *Clin Chim Acta* 466:112–119. <https://doi.org/10.1016/j.cca.2016.12.018>.
- Wang X, Krambeck AE, Williams JC, Tang X, Rule AD, Zhao F, Bergstralh E, Haskic Z, Edeh S, Holmes DR, 3rd, Herrera Hernandez LP, Lieske JC. 2014. Distinguishing characteristics of idiopathic calcium oxalate kidney stone formers with low amounts of Randall's plaque. *Clin J Am Soc Nephrol* 9:1757–1763. <https://doi.org/10.2215/CJN.01490214>.
- Manzoor MAP, Singh B, Agrawal AK, Arun AB, Mujeeburahiman M, Rekha PD. 2018. Morphological and micro-tomographic study on evolution of struvite in synthetic urine infected with bacteria and investigation of its pathological biomineralization. *PLoS One* 13:e0202306. <https://doi.org/10.1371/journal.pone.0202306>.
- Schwaderer AL, Wolfe AJ. 2017. The association between bacteria and urinary stones. *Ann Transl Med* 5:32–32. <https://doi.org/10.21037/atm.2016.11.73>.
- Johnson G, Wolfe AJ, Putonti C. 2019. Characterization of the ϕ CTX-like *Pseudomonas aeruginosa* phage Dobby isolated from the kidney stone microbiota. *Access Microbiol* 1. <https://doi.org/10.1099/acmi.0.000002>.
- Hilt EE, McKinley K, Pearce MM, Rosenfeld AB, Zilliox MJ, Mueller ER, Brubaker L, Gai X, Wolfe AJ, Schreckenberger PC. 2014. Urine is not sterile: use of enhanced urine culture techniques to detect resident bacterial flora in the adult female bladder. *J Clin Microbiol* 52:871–876. <https://doi.org/10.1128/JCM.02876-13>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Darling ACE, Mau B, Blattner FR, Perna NT. 2004. Mauve: multiple alignment of conserved genomic sequence with rearrangements. *Genome Res* 14:1394–1403. <https://doi.org/10.1101/gr.2289704>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP,

- Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44: 6614–6624. <https://doi.org/10.1093/nar/gkw569>.
15. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a Web tool to identify clustered regularly interspaced short palindromic repeats. *Nucleic Acids Res* 35:W52–W57. <https://doi.org/10.1093/nar/gkm360>.
 16. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic Local Alignment Search Tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
 17. Roux S, Enault F, Hurwitz BL, Sullivan MB. 2015. VirSorter: mining viral signal from microbial genomic data. *PeerJ* 3:e985. <https://doi.org/10.7717/peerj.985>.