

Supporting Information

Single Walled Carbon Nanotube Probes for Protease Characterization Directly in Cell Free Expression Reactions

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Supplement Section 1: DNA sequences used in this study

Below are the gene templates used for the CFE expressed proteins. All sequences were optimized for *Escherichia coli* using IDT's codon optimization tool. Each color represents the region according to Addgene pJ11-sfGFP gene for the Plasmid vector and our added sites for making circularized templates:

T7 Promoter – T7 Promoter

RBS – Ribosome binding site

Start – Start codon

Protein Sequence – Gene coding for protein

Stop – Stop codon

T7 Terminator – T7 Terminator

Circularization site – HindIII Digest

Primer Sequences – Primer sequences

Ori – Ori

KanR – KanR

Supplement Sequence 1: sfGFP LET (999 bp)

```
GTAAAACGACGGCCAGT AGCGCTATTA AAGCTT CGAAAT TAATACGACTCACTATAGG GAGACCACAA
CGGTTTCCCTCTAGAAATAAT TTTGTTTAACTTTAAGAAGGAGA TATACAT ATG AGCAAAGGTGAAGAAC
TGTTTACCGGCGTTGTGCCGATTCTGGTGGAACTGGATGGCGATGTGAACGGTCACAAATTCAGCGT
GCGTGGTGAAGGTGAAGGCGATGCCACGATTGGCAAACCTGACGCTGAAATTTATCTGCACCACCGGC
AAACTGCCGGTGCCGTGGCCGACGCTGGTGACCACCCTGACCTATGGCGTTCAGTGTTTTAGTCGC
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TACGATTAGCTTTAAAGATGATGGCAAATATAAACGCGCGCCGTTGTGAAATTTGAAGGCGATACCCT
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TGGTGATGGTCCGGTGCTGCTGCCGATAATCATTATCTGAGCACGCAGACCGTTCTGTCTAAAGATC
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TCCGCAGTTCGAAAAA TAATAA GTCGAC CGGCTGCTAACAAAGCCCGAAAGGAAGCTGAGTTGGCTG
CTGCCACCGCTGAGCAATAACTAGCATAACCCCTTGGGGCCTCTAAACGGGTCTTGAGGGGTTTTTT
GCTGAAAGCGAGACT AAGCTT TAAACTTCGG GTCATA GCTGTTTCCTG
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Supplement Sequence 2 : Subtilisin BPN' (1344 bp)

```
GTAAAACGACGGCCAGT AGCGCTATTA AAGCTT CGAAAT TAATACGACTCACTATAG GAGACCACA
ACGTTTCCCTCTAGAAATAATTTTGTCTTAACTTTAAG AAGGAGA TATACAT ATG GCAGGTAAAAGTAA
CGGCGAGAAAAAATACATCGTTGGCTTCAAACAAACGATGTCGACCATGAGCGCAGCGAAAAAGAAA
```

GATGTCATCAGCGAAAAAGGCGGTAAAGTGCAGAAACAATTCAAATACGTTGACGCGGCCAGTGCC
 ACCCTGAATGAAAAAGCAGTGAAAGAAGTGAAGAAAGATCCGTCCGTGGCGTACGTTGAAGAAGAC
 CATGTTGCTCACGCGTATGCCAGTCCGTTCCGTACGGTGTCTCACAAATTAAGCACCGGCTCTGC
 ATTCGCAGGGCTATACCGGTAGCAACGTTAAAGTCGCGGTGATTGATAGCGGCATCGACAGTTCCC
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 GCGGTCCGTCCGTTTCAGCCGCACTGAAAGCAGCTGTCGACAAAGCAGTTGCTTCCGGTGTGGTTG
 TTGTGGCCGCAGCCGGTAACGAAGGCACGTGAGGCTCATCGAGCACCCTGGGTTATCCGGGCAAA
 ACCCGTCGGTTATTGCGGTGCGGTGCCGTGGATTCTAGTAATCAGCGTGCGAGCTTTTCTCAGTTGG
 CCCGGAACCTGGACGTTATGGCCCCGGGTGTCTCTATTCAAAGTACGCTGCCGGGTAACAAATATGG
 CGCGTACAATGGTACCAGCATGGCATACCGCATGTGGCTGGTGCTGCGGCCCTGATCCTGAGCAA
 ACACCCGAACCTGGACGAATACCCAGGTTGCTCGAGCCTGGAAAACACCACGACCAAACTGGGCGA
 TTCTTTCTATTACGGCAAAGGTCTGATCAATGTTGAGGCAGCTGCGCAA **TAATAA** **GTCGAC** **CGG** **CTG**
CTAACAAGCCCCGAAAGGAAGCTGAGTTGGCTGCTGCCACCGCTGAGCAATAACTAGCATAACCCC
TTGGGGCCTCTAAACGGGTCTTGAGGGGTTTTTTGCTGAAAGC **GAGACT** **AAGCTT** **TAAACTTCGG** **GT**
CATAGCTGTTTCTG

Supplement Sequence 3 : Bovine serum Trypsin LET (EC:3.4.21.4) (1271 bp)

GTAAAACGACGGCCAGT **AGCGCTATTA** **AAGCTT** **CGAAAT** **TAATACGACTCACTATAGG** **GAGACCACAA**
CGGTTTCCCTCTAGAAATAAT **TTTGTTTAACTTTAAGAAGGAGA** **TATACAT** **ATG** **AAGACGTTTATTTTCT**
TGCGTTACTGGGGGCCCGCCGTAGCGTTC **CCAGTTGATGATGATGACAAAATCGTCGGCGGTTACACA**
TGCGGCGCGAATACGGTGCCCTACCAGGTGTCCCTGAACTCGGGCTACCATTTCTGCGGAGGCTCG
TTGATTAACAGCCAATGGGTGGTGTCGGCGGCTCATTGTTACAAATCGGGCATCCAAGTCCGTCTTG
GAGAGGACAACATAAACGTGTCGAAGGGAACGAACAGTTTATCTCCGCCAGCAAGAGCATCGTTCA
TCCTTCATACAACAGCAATACCTTGAACAACGATATCATGTTAATTAAATTGAAGAGCGCGGCTTCTTTA
AATCCCGCGTCGCCTCCATTTGCTGCCAATTCATGTGCTTCGGCGGGCACGCAAGTGTTAATTAG
CGGATGGGGTAATACGAAATCAAGTGGCACCAGCTACCCCGACGATTGAAATGCTTGAAAGCTCCTA
TTTTGTCTGACAGCTCCTGCAAGAGCGCATATCCAGGGCAGATTACTTCTAATATGTTCTGTGCAGGTT
ATCTGGAGGGAGGGAAGGATTATGTGAGGGTGATTGAGGGGGCCTGTCGATGCAGCGGTAAGC
TGCAAGGAATAGTCAGTTGGGGCTCGGGTTGCGCGCAAAAGAATAAACCAGGGGTCTACACGAAAGT
CTGCAACTACGTTTCGTGGATTAAGCAGACCATCGCTAGCAAC **TAATAA** **GTCGAC** **CGGCTGCTAACAA**
AGCCCGAAAGGAAGCTGAGTTGGCTGCTGCCACCGCTGAGCA **ATAACTAGCATAACCCCTTGGGGC**
CTCTAAACGGGTCTTGAGGGGTTTTTG **CTGAAAGCGAGACT** **AAGCTT** **TAAACTTCGG** **GT** **CATAGCTG**
TTTCCTG

Supplement Sequence 4 : Myoicolsin from *Myroides profundus*, H2BKX5 · H2BKX5_MYRPR (2322 bp)

GTAAAACGACGGCCAGT **AGCGCTATTA** **AAGCTT** **CGAAAT** **TAATACGACTCACTATAGG** **GAGACCACAA**
CGGTTTCCCTCTAGAAATAAT **TTTGTTTAACTTTAAGAAGGAGA** **TATACAT** **ATG** **CGGAAAATAATCGTTAC**
TGGGTTTGCTTTTTTATGCTTGGGAGTGTATGACAATTATGGTCAAATATCCACTCGCTCGGCCGT
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 ATGGGACAGATGGCATCGGGGGGTATGCTGGTGTCCAATCATAGCTATGGGTATAACTACTTCGATGA
 TTACGGGTATTTGAACGAGCCTAGTTTGATACGCAATTTTGGAGCATATTCAGAGCACTCTCGGGAGTT
 TGATAGAGTGGCATACTTATTTGCGTACTATCAGCCAGTTATAGCAGCCGGCAACGACGGCGAGTACC
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 AACCAAACAAGAAATACGGCTTTGGCCGAATCCTGTAGAAGATCATTTATATGTTAGTTTGGATAAGAC
 GTACAACGGAAAGGTAATTGACATGAAAGTCTACGATTAATGGACGGCTTGTCTGTCTTCGTCTGA
 AACCGTACAGCAAGAAAAGGTTAGCATAAATATGGCATCCTTAAATTCGAATATCTACGTTGTGGAGGT
 GAAAGGTGACAACCTGTCTAAACTGTACGCATCGCGAAGCGGTAATAAGTCGACCGGCTGCTAACA
 AAGCCCGAAAGGAAGCTGAGTTGGCTGCTGCCACCGCTGAGCAATAACTAGCATAACCCCTTGGGG
 CCTCTAAACGGGTCTTGAGGGGTTTTTCTGAAAGCGAGACTAAGCTTAAACTTCGGGTCATAGCT
 GTTTCCTG

Supplement Sequence 5: Trypsin from *Streptomyces* sp. CB00072 (1068 bp)

GTAAAACGACGGCCAGTAGCGCTATTAAGCTTCGAAATTAATACGACTCACTATAGGAGAGACCACAA
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 CGCCGAATGCGGTTAAGGTCAGATCGACCAAAGTACTTCAGGCTCCGGGATACAACGGGACGGGCA
 AGGACTGGGCTCTTATCAAATTGGCCAGCCTATTAACCAACCCACATTGAAGATTGCGACTACGACG
 GCGCATAACCAAGGGACCTTCACGGTGGCGGGCTGGGGAGCTAATCGTGAAGGGGGTCCCAACA
 ACGCCATTTACTGAAAGCTAATGTACCTTTCGTAAGTGATGCTGCTTGTGCTTCGGCCTACGGGAACG
 AACTGGTGGCAACGAGGAAATATGCGCTGGTTATCCAGATACTGGAGGGGTTGACACTTGTCAAGG
 AGACAGTGGAGGTCCAATGTTCCGAAAGACAACGCTGATGAGTGGGTTGAGGTAGGCATAGTAAGT
 TGGGGTTACGGGTGCGCACGTCCAGGCTATCCAGGCGGTGACGCAGAGGTGAGCACGTTTGCAGT
 GCTATCGCCTCCGCTGCTCGCACGTTGTAATAAGTCGACCGGCTGCTAACAAGCCCGAAAGGAAGC
 TGAGTTGGCTGCTGCCACCGCTGAGCAATAACTAGCATAACCCCTTGGGGCCTCTAAACGGGTCTTG
 AGGGGTTTTTCTGAAAGCGAGACTAAGCTTAAACTTCGGGTCATAGCTGTTTCCTG

Supplement Sequence 6: Chymotrypsin-C (human) EC:3.4.21.2 (1089 bp)

GTAAAACGACGGCCAGTAGCGCTATTAAGCTTCGAAATTAATACGACTCACTATAGGAGAGACCACAA
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 GTATCTCCAACACGCGTACCTACCGTGTAGCGGTGGGTAAGAATAACCTTGAAGTCGAAGACGAAGA
 GGGCAGCCTGTTTGTCGGAGTCGATACTATACACGTTTATAAGCGCTGGAATGCTTTGCTGTTGCGTA
 ATGATATTGCCTTAATAAAGTTAGCAGAGCATGTGCAATTGTGCGACACTATACAGGTAGCATGCCTTC
 CAGAGAAGGATTCTTACTTCCCAAGGACTATCCCTGCTATGTTACAGGGTGGGGTCGGCTGTGGAC
 TAACGGCCCAATCGCAGATAAGTTGCAGCAAGGATTACAACCTGTGGTAGATCACGCGACTTGCTCTC
 GTATAGACTGGTGGGGCTTCAGAGTAAAAAAGACCATGGTTTGTGCTGGCGGGGATGGGGTTATTTT
 TGCTTGTAATGGAGATTCGGGTGGACCGTTGAATTGTCAACTTGAGAATGGCTCATGGGAGGTTTTTG
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 TTCCGCGTATATAGATTGGATCAACGAAAAAATGCAACTGTAATAAGTCGACCGGCTGCTAACAAGCC
 CGAAAGGAAGCTGAGTTGGCTGCTGCCACCGCTGAGCAATAACTAGCATAACCCCTTGGGGCCTCTA
 AACGGGTCTTGAGGGGTTTTTCTGAAAGCGAGACTAAGCTTAAACTTCGGGTCATAGCTGTTTCC
 TG

Supplement Section 2: Supplement figures used in this study

Schematic and details of the custom built NIR fluorometer used in this study to measure SWCNT emission intensity

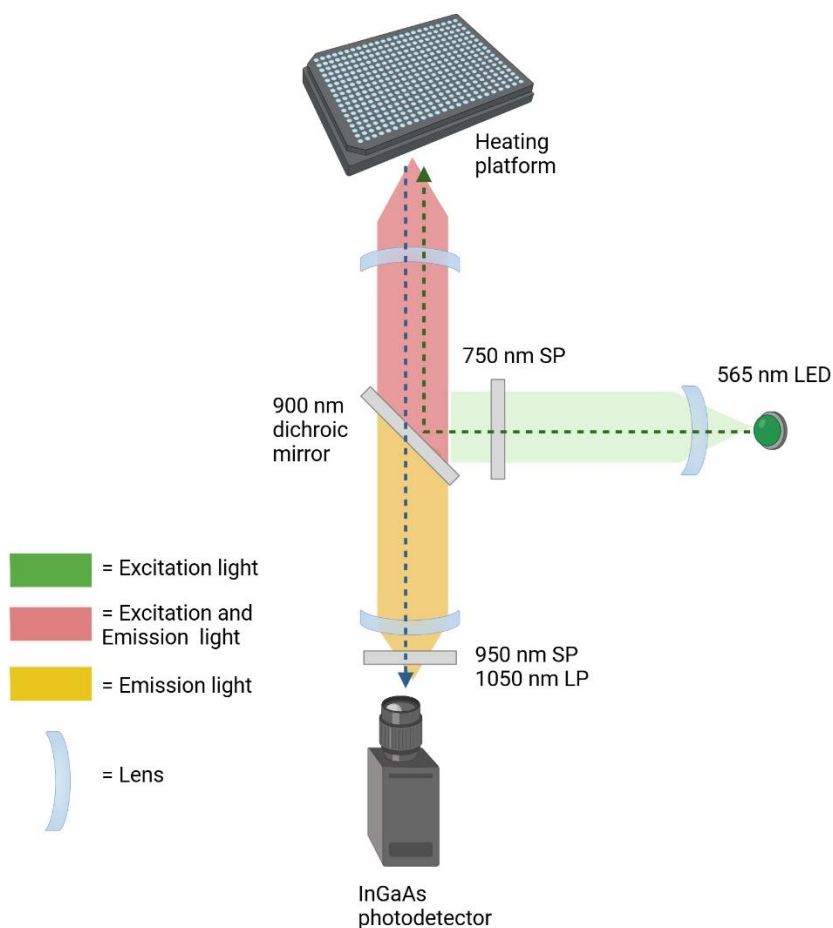


Figure S1. Schematic of the NIR fluorometer used in this study to measure SWCNT emission intensity.

Probe response to CFE proteins with direct sonicated protein wrapped SWCNTs.

The following figures illustrate the response of protein-wrapped SWCNT probes to end-products of cell-free expression (CFE) (Figure S2). After establishing a baseline, analytes were added to the probe solution at a 5% of the total volume (95% SWCNT solution). The initial signal drop can be attributed to three factors: dilution of the SWCNT concentration due to the addition of the analyte buffer, intrinsic signal instability of the SWCNTs to the CFE reaction buffers, and the unrecorded fluorescence data during sample addition on the fluorometer.

Figure S3 shows the response of directly sonicated BSA-SWCNT probes within the cell-free reaction. These probes experienced a signal loss of more than 50% and were unable to distinguish between subtilisin and control samples.

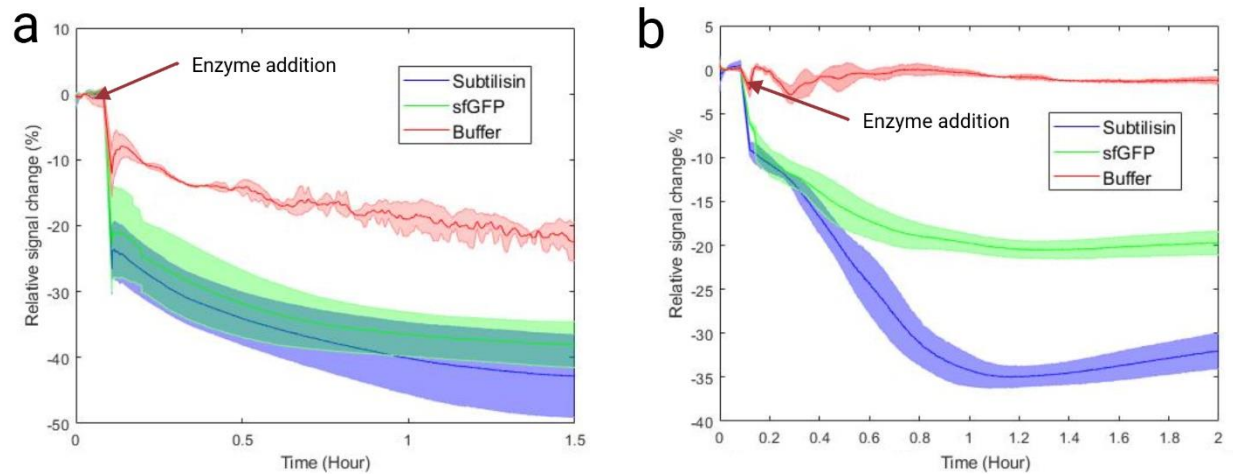


Figure S2. Probe response to CFE proteins with direct sonicated protein wrapped SWCNTs. a) lysozyme-wrapped SWCNT respond to different CFE products of subtilisin and sfGFP as control. Buffer added to test sensor stability upon small volume addition. b) BSA-wrapped SWCNT sensor response to different CFE products of subtilisin and sfGFP as control

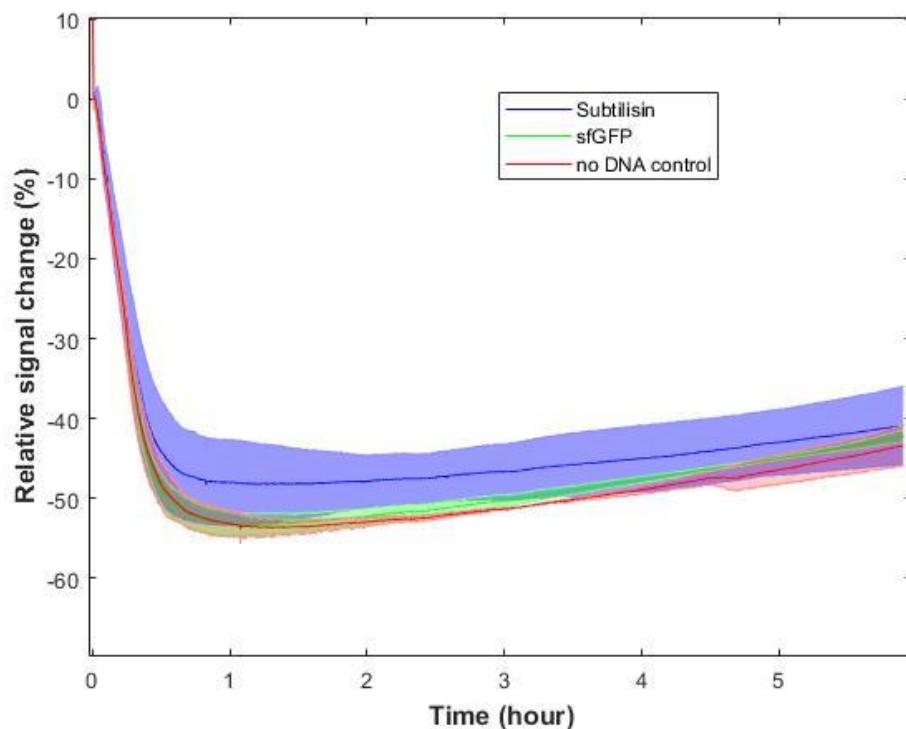


Figure S3. Direct sonicated BSA-wrapped sensor response inside CFE with different genes. Subtilisin as target, sfGFP to check the expression and no DNA control.

Characterization of conjugated protein SWCNT probes

Figure S4 presents the Fourier Transform Infrared Spectroscopy (FTIR) analysis of CMC-SWCNT, BSA-conjugated, and casein-conjugated probes. Key peaks are labeled, including the amide bond characteristic of protein samples (1647 cm^{-1}), highlighting the successful conjugation of proteins to the SWCNTs.

Figure S6 depicts the emission spectral changes of SWCNTs before and after 30 minutes of reaction with bacterial protease. These changes provide insights into the interaction between the SWCNT probes and the protease, demonstrating their potential for detecting enzymatic activity.

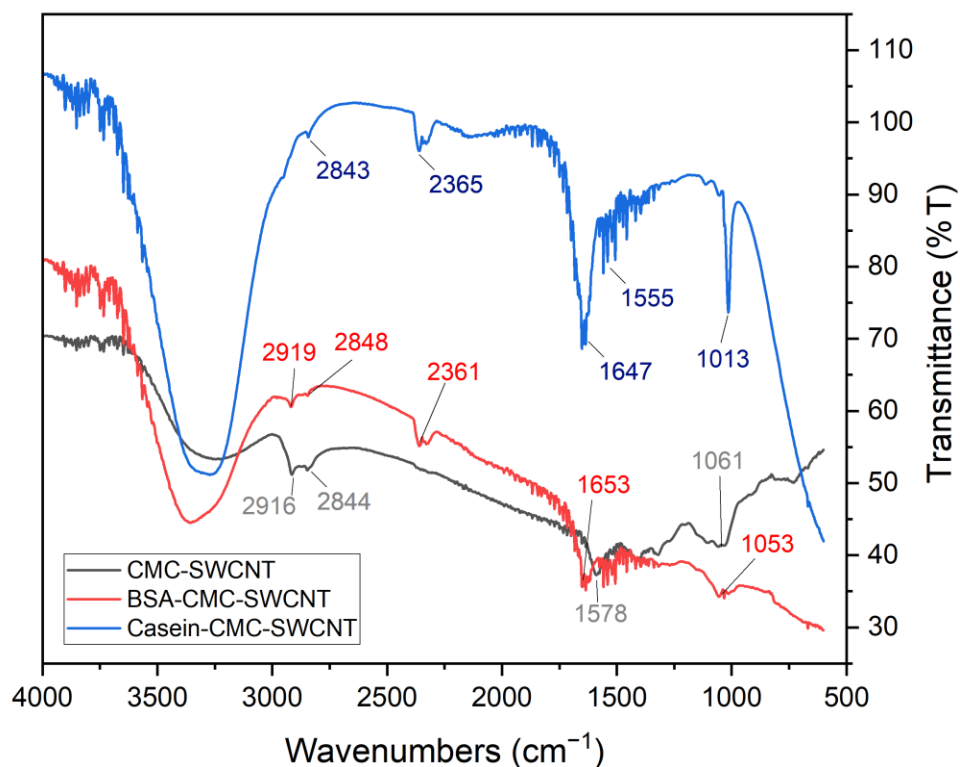


Figure S4. FTIR plot for the different SWCNT probes with CMC wrapping (black), BSA conjugated CMC-SWCNT (red), and casein conjugated CMC-SWCNT (blue)

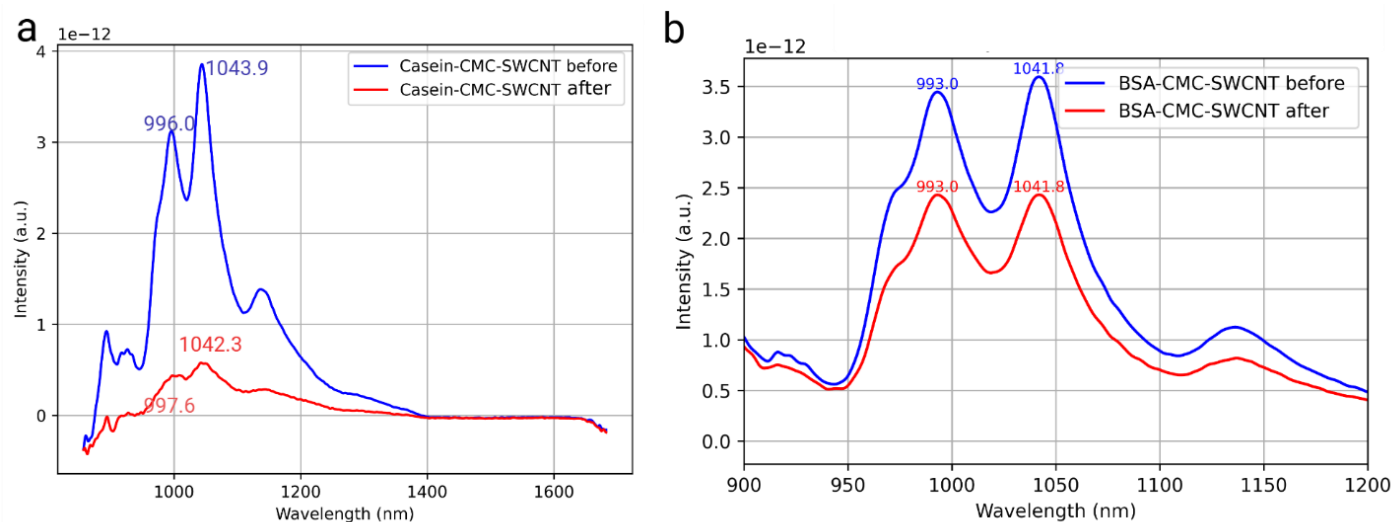


Figure S5 Emission spectra of a) casein conjugated and b) BSA conjugated probes before and after 30 minutes of incubation with 100 $\mu\text{g/ml}$ of bacterial protease.

Conjugated protein SWCNT probes response to proteases

The following figures (S7,S8) describe the responses of conjugated protein probes to addition of bacterial protease or CFE end products added to the probes. Figure S7 is the BSA conjugated probe response to

different levels of bacterial protease. Figure S8 describes the response of Bsa conjugated probes to different expression levels of subtilisin end-products. Figure S9,S10 show the response of casein and BSA conjugated probes inside the lysate based CFE reaction with the genes added from the beginning.

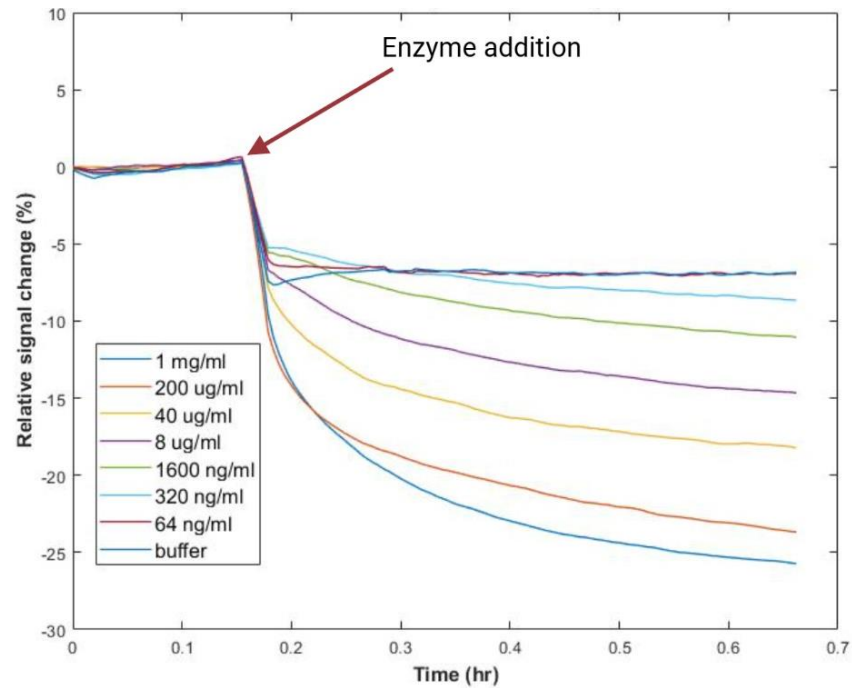


Figure S7. BSA conjugated sensor response to different concentrations of bacterial protease

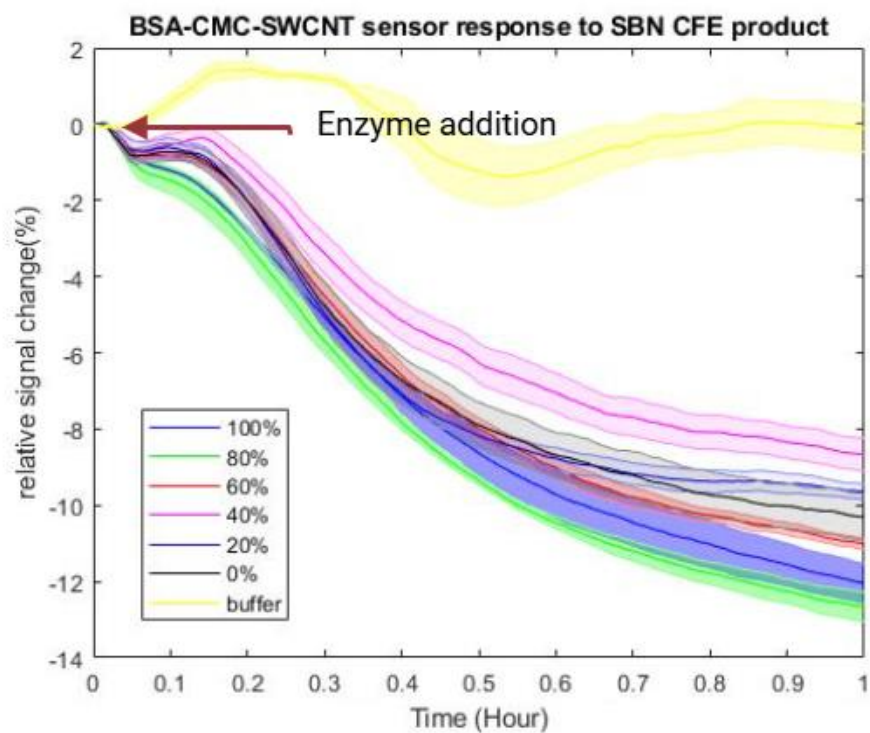


Figure S8. BSA conjugated sensor response to different mixtures of subtilisin and sfGFP CFE end product mixtures.

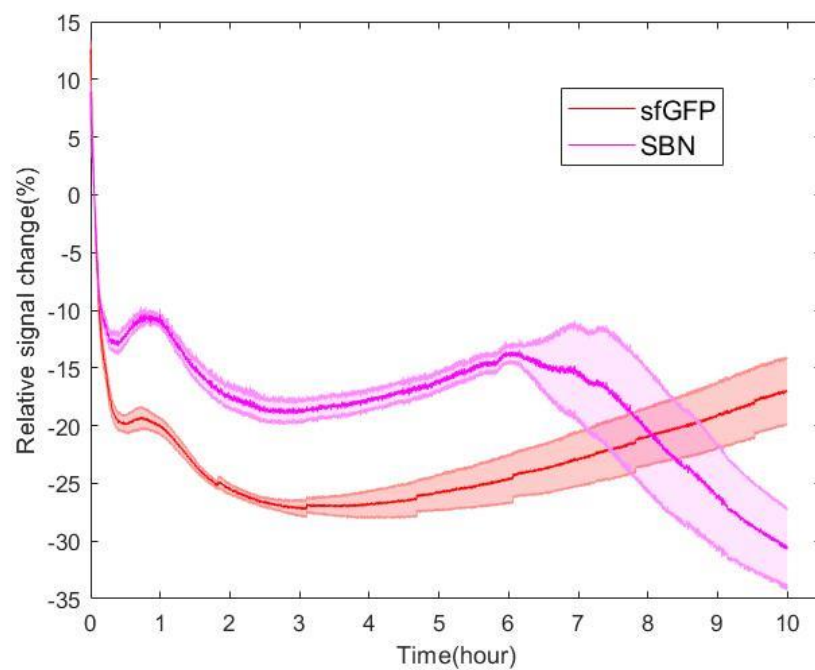
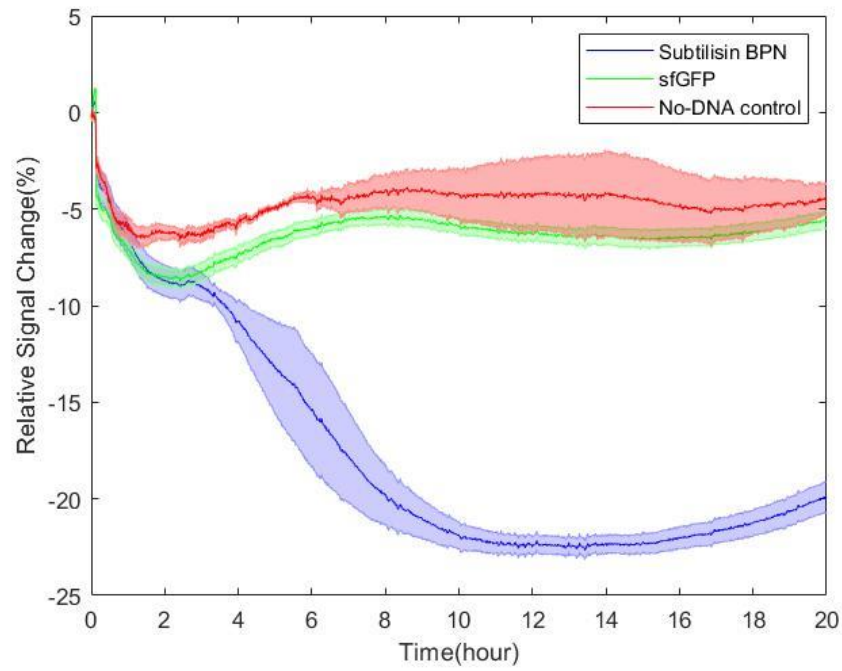


Figure S9. Casein conjugated probe response in the lysate-based CFE system with genes templates added at time = 0 h, showing the average of three replicates as solid lines with standard deviation as



shaded region.

Figure S10. BSA conjugated probe response in the lysate-based CFE system with genes templates added at time = 0 h, showing the average of three replicates as solid lines with standard deviation as shaded region.