Research Report

Characterization of the Contribution of Genetic Background and Gender to Disease Progression in the SOD1 G93A Mouse Model of Amyotrophic Lateral Sclerosis: A Meta-Analysis

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Abstract.

Background: The SOD1 G93A mouse model of amyotrophic lateral sclerosis (ALS) is the most frequently used model to examine ALS pathophysiology. There is a lack of homogeneity in usage of the SOD1 G93A mouse, including differences in genetic background and gender, which could confound the field's results.

Objective: In an analysis of 97 studies, we characterized the ALS progression for the high transgene copy control SOD1 G93A mouse on the basis of disease onset, overall lifespan, and disease duration for male and female mice on the B6SJL and C57BL/6J genetic backgrounds and quantified magnitudes of differences between groups.

Methods: Mean age at onset, onset assessment measure, disease duration, and overall lifespan data from each study were extracted and statistically modeled as the response of linear regression with the sex and genetic background factored as predictors. Additional examination was performed on differing experimental onset and endpoint assessment measures.

Results: C57BL/6 background mice show delayed onset of symptoms, increased lifespan, and an extended disease duration compared to their sex-matched B6SJL counterparts. Female B6SJL generally experience extended lifespan and delayed onset compared to their male counterparts, while female mice on the C57BL/6 background show delayed onset but no difference in survival compared to their male counterparts. Finally, different experimental protocols (tremor, rotarod, etc.) for onset determination result in notably different onset means.

Conclusions: Overall, the observed effect of sex on disease endpoints was smaller than that which can be attributed to the genetic background. The often-reported increase in lifespan for female mice was observed only for mice on the B6SJL background, implicating a strain-dependent effect of sex on disease progression that manifests despite identical mutant SOD1 expression.

Keywords: Lou Gehrig's disease, ALS, Amyotrophic Lateral Sclerosis, motoneuron disease, transgenic mice, rotarod, hindlimb tremor, forelimb tremor

INTRODUCTION

Amyotrophic Lateral Sclerosis (ALS) is a neurodegenerative disorder characterized by progressive loss of motor neurons, muscular atrophy, functional deficits, paralysis, and death. ALS is frequently studied with the aid of animal models. In particular, the super-oxide dismustase-1 glycine 93 to alanine mutation (SOD1 G93A) transgenic mouse model is widely utilized [36, 37] as it emulates the associated SOD1 mutations in human familial ALS [38] and exhibits pathological progression similar to that which is seen in human forms of the disease [39]. In fact, a PubMed

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search returns at the end of the 2014-year over 1,200 different articles containing "G93A" and "ALS". Despite all of the numerous publications in this popular ALS mouse model, there remains controversy as to how genetic background, sex, and different experimental measures of disease onset and end point may or may not affect the outcomes of studies examining the SOD1 G93A pathophysiology or potential ALS treatments within this model.

Experimentally, the efficacy of a potential therapeutic treatment for ALS is evaluated on the basis of its effect on the parameters governing disease progression in the SOD1 G93A transgenic mouse, including age at onset, lifespan, disease duration, and motor performance. However, these parameters have been found to differ on the basis of transgenic copy number [40-42], genetic background [26-28], and sex [25, 43, 44] of the mice, all of which tend to vary in usage between studies. Additionally, the disease onset may be qualified through a variety of measures including gait abnormalities [45], limb tremors [46], electromyography [47], body weight declines [48], or deficits in motor performance tests, such as the rotarod [49] or grip [50] tests, each of which may differ from the others due to some difference in sensitivity of the test or through a variation in the underlying physiological mechanism being measured.

Prior studies commonly report delayed onset and progression of disease in females in both human [51] and mouse [43, 44, 47] variants of ALS. However, there have been conflicting reports in the literature as to the effect of sex and genetic background on SOD1 G93A disease onset and lifespan. Some studies report that female mice experience longer lifespans [44] in addition to delayed onset, while other studies find no difference in disease endpoint [47]. Additionally, the magnitude of the female-associated neuroprotective effect may vary on the basis of genetic background. In particular, Mancuso et al. [26] observed the femaleassociated protective effect on lifespan in mice bred on the C57BL/6 background, but not in B6SJL mice, which conflicts with the results of Heiman-Patterson [27] that report a sex-related effect on lifespan for mice on the B6SJL mice but not for those on the C57BL/6 background. As there is an inconsistency in the results reported by individual studies, it is of interest to resolve the conflicting results so the SOD1 G93A mouse model is better understood and potential therapeutics may be better predicted.

The lack of homogeneity in methodology between studies complicates comparative analysis in the SOD1 G93A field's wealth of published data. The precise effect of genetic background, sex, and experimental measures of progression (rotarod, tremor, etc.) must be characterized to better assess the true impact of underlying disease mechanisms and potential therapeutics in both previous and future SOD1 G93A experimental studies. In this meta-analysis utilizing data from 97 studies, we characterize the disease progression parameters of onset, survival, and disease duration for the SOD1 G93A mouse and determine the effect of sex and genetic background on those parameters.

MATERIALS AND METHODS

Inclusion criteria

Data recapture was completed per previously published methodology [1]. Briefly, PubMed and Medline were searched in March 2013 for inclusion of articles based on the appearance of a combination of the following terms in the article title or abstract: "G93A", "Amyotrophic Lateral Sclerosis", "ALS", and "transgenic mouse". That set was constrained by searching for the appearance of one or more of the following terms in the title, abstract, or figure caption: "onset", "survival", "lifespan", "rotarod", "motor performance". The viability of each potential study was then manually determined. Included studies were those containing unique experimental data and citing the use of the high copy SOD1 G93A mouse from Jackson Laboratories (note that the overwhelming majority of studies do not quantitatively cite the actual number of transgene copies but rather only qualitatively state transgene copy number as being either "high" or "low"). Additional reported information required for study inclusion consisted of: mean mouse age at disease onset, mean mouse age at death, defined measures for calculation of disease progression, and clearly defined sample size information.

Data organization

Within each study of interest, independent groups of mice were defined as groups of mice used experimentally that exhibited a unique set of the following parameters: number of mice in the group, sex of the mice, genetic background, mean age at onset, and mean age at death. For each independent group of SOD1 G93A mice used experimentally within a study, the mean age at disease onset and death were tabulated. The disease duration was taken as the within-group difference between the mean survival and the mean onset. Additionally, the particular indicator used in the study for determining the time of disease onset was recorded and categorized.

Definitions of terms and categories

All included data are for the high copy number SOD1 G93A mouse model of ALS in the absence of treatment effects. Mice were categorized based on their genetic background as either originating from the B6SJL-Tg (SOD1-G93A)1Gur/J or the B6.Cg-Tg(SOD1-G93A)1Gur/J lines. For the ease of discussion, mice of B6SJL-Tg(SOD1-G93A)1Gur/J and B6.Cg-Tg(SOD1-G93A)1Gur/J lines will be referred to as "B6SJL" and "C57BL/6", respectively. A group of mice was considered to be either male or female if that qualification fit all mice in the group and was considered mixed otherwise.

A diverse set of onset determination methodologies were broadly grouped in to four categories. Studies that defined onset through any sort of deficit in the rotarod test were deemed "Rotarod Decline". Any mention of using tremors in the hindlimb as an indicator was deemed "Hindlimb Tremor" and references to tremors in the forelimb or non-specific locations were grouped as "Non-Hindlimb Tremor". Onsets qualified through declines in bodyweight, gait abnormalities, limb paralysis, deficits in grip strength, and were grouped with articles without a specific definition of onset in to a catch-all category called "General", due to the low overall usage of each of those methods.

Analytical methods

The disease progression parameters of age at onset, survival, and disease duration for the SOD1 G93A mouse were analyzed independently as the response variables of multiple ordinary least squares regression models in Stata (StataCorp, LP) with the genetic background and sex of the mice included as categorical predictors with full interaction. Each observation used in the calculation corresponds to an independent group of mice as defined by a unique combination of grouping factors and disease progression parameters with values for the disease progression parameters equal to the mean value of the parameter reported by the particular study. Observations were weighted in Stata on the basis of the group sample size and were factored as analytic weights [52], which are proportional to the inverse of the variance of the measure and normalized to the total number of points, giving results comparable to models weighted with the inverse variance directly [53]. Additional regression models were generated with the age at onset or survival included as an interacting continuous predictor when the response variable was survival or disease duration, as appropriate. The model with the largest R^2 value was used for analysis. Post-estimation for the predicted mean value for specific combinations of covariates was computed with the "margins" command in Stata. The computed means are taken as the estimation on the interaction terms for the included groups and the results are plotted as categorical bar charts with significance between categories of mice determined through multiple comparisons, with the Bonferroni correction, on the margins output. In cases where continuous covariates were included as an interaction term, the estimation uses the integrated mean value for that particular categorization.

Analogous Cox proportional hazard regression models [54] were generated for each of onset, survival, and disease duration with the mean value from the study entering the model as count-time data with a frequency corresponding to the mice used in that study. Like the linear regression models, the regression was performed with full interaction of the categorical predictors. Predicted hazard ratios were obtained from the model and groups compared through multiple comparisons on the hazard ratios.

All statistical tests and models were computed in Stata (StataCorp LP). Graphics and tables were created in IGOR Pro (WaveMetrics). The significance level was set at 0.05 for all statistical tests. Data are expressed as mean \pm SEM.

RESULTS

Data overview

The literature search returned 97 articles (Table 1) and 150 independent groups of mice with sufficient detail to fully characterize the data. The data is fully characterized if the article reports the mean onset, mean time of death, and sample size for each independent group and there must be enough information in the paper to categorize the groups in terms of genetic background, sex, and onset determination criteria. Of the articles surveyed, 85 groups of mice were classified as mixed sex, 37 groups were comprised solely of male mice, and 28 groups had only female mice. 91 groups were bred on the B6SJL background and 59 on the C57BL/6 (Table 1). Among mixed sex B6SJL articles, 12 report even distributions of males and females [2-13], 3 report uneven distributions without an overall bias [14-16], and the remainder report nothing concerning sex distribution. Of the mixed sex C57BL/6

Data source and sample information				
Category	Articles	Groups	Mice	References
B6SJL, Female	15	17	203	[11, 47, 56, 61–72]
B6SJL, Male	14	16	192	[11, 47, 56, 61, 62, 67, 69, 70, 72–77]
B6SJL, Mixed	43	58	909	[2–16, 46, 49, 71, 78–102]
C57BL/6, Female	9	11	173	[25, 44, 45, 103–108]
C57BL/6, Male	14	21	294	[22, 25, 44, 45, 50, 57, 58, 104, 105, 109–113]
C57BL/6, Mixed	22	27	472	[17-20, 22, 23, 25, 45, 48, 104, 114-125]
B6SJL, General/Other	30	45	628	[3-5, 10, 11, 13-16, 61, 66-69, 71, 73-79, 82, 84, 85, 87, 88, 95, 98, 99]
B6SJL, Hindlimb Tremor	5	9	163	[46, 70, 72, 92, 94]
B6SJL, Non-Hindlimb Tremor	6	8	171	[7, 56, 62, 81, 91, 102]
B6SJL, Rotarod Decline	20	29	342	[2, 8, 9, 12, 47, 49, 63–65, 80, 83, 86, 89, 90, 93, 96, 97, 100, 101, 126]
C57BL/6, General/Other	16	26	358	[19–21, 45, 48, 50, 104–106, 109, 113, 115, 118–120, 123]
C57BL/6, Hindlimb Tremor	3	8	93	[57, 58, 108]
C57BL/6, Non-Hindlimb Tremor	3	6	153	[17, 25, 110]
C57BL/6, Rotarod Decline	14	19	335	[18, 22, 23, 44, 103, 107, 111, 112, 114, 116, 117, 121, 122, 124]

Table 1 Data source and sample information

Number of articles, independent groups, and mice that relate to each category of interest. Categories are named first by the genetic background strain followed by either a sex group label or a label for the onset determination category. Articles may be counted in multiple categories if that article contains information for multiple groups.

articles, 5 report balanced sex distributions [17–21], 4 have unbalanced groups with a slight overall bias towards male mice among the studies [22–25], while the remainder of the articles report nothing concerning the number of males and females. To determine onset of the disease, 34 articles used deficits in the rotarod test, 8 articles used the onset of hindlimb tremors, 9 articles looked at forelimb or non-specific tremors, and 46 articles fell into the miscellaneous "General" category.

Onset

The raw data distributions show that the mean age at onset reported by the articles is most frequently in the range of 90 to 100 days (Fig. 1B) and the raw data plot qualitatively suggests that mice on the C57BL/6 background have later onsets than their B6SJL counterparts (Fig. 1A). When linear regression is run with onset as the response, genetic background strain is found to significantly impact the regression output (p=0.002)and the effect of sex is found to be insignificant as a regression term and in subsequent multiple comparisons. The mixed sex mice were the only sex category to significantly differ on the basis of the genetic background (p = 0.002). The predicted mean onset was 99.86 ± 2.161 days for mixed sex B6SJL groups and 111.4 ± 2.857 days for the mixed C57BL/6 mice (Fig. 2A). If strain is taken as the independent covariate, the main effect estimates onset as 99.27 ± 1.79 days for the B6SJL mice and 109.9 ± 2.063 days for the C57BL/6. It should be stated that although the mean onset is found to differ by the genetic background it is overall a poor sole predictor of onset as the adjusted \mathbf{R}^2 for this regression fit is low at 0.0776.

When the onset data are modeled as a Coxproportional hazards, the predicted survival curve for disease onset shows a difference (p < 0.001) between the mice on the B6SJL and C57BL/6 genetic backgrounds (Fig. 2B). In contrast to the linear regression, this model predicts some difference in onset due to sex. Within the B6SJL group, female mice showed later onsets than male mice (p = 0.001) and the male C57BL/6 mice showed onsets earlier than both the mixed and female C57BL/6 mice (p < 0.001). No other comparisons were statistically significant.

Onset category

Differences in reported mean onsets between articles that defined their onset determination criteria were evaluated through regression with genetic background strain and the onset category included as categorical predictors (Fig. 3). The adjusted R² for the model was 0.23. Within the B6SJL background, only hindlimb tremor with a mean onset of 85.93 ± 4.527 days was significantly different from the General category with a mean onset of 100.6 ± 2.330 days (p = 0.005). Within C57BL/6, the onsets determined by both hindlimb tremors and non-hindlimb tremors, with mean onsets of 103.7 ± 5.994 days and 94.06 ± 4.657 days, were found be significantly earlier than those of the General category with a mean onset of 118.9 ± 3.055 days (p = 0.025 and p < 0.001, respectively).

Survival

The histogram for the mean survival appears somewhat bimodal with one peak in the range of 120 to 130 days and another between 150 and 160 days (Fig. 1D). It follows from the raw data plot (Fig. 1C) that the



Fig. 1. Representation of raw data from literature. The (A) mean onset, (C) mean survival, (E), mean disease duration for each independent group of mice was plotted with a size proportional to the sample size of that group. Data was stratified by both the sex of the group and genetic background strain. The (B) mean onset, (D) mean survival, and (F) mean disease duration were aggregated across groups and the distributions weighted by group sample size and results accumulated in histograms.

first peak likely corresponds to the mean survival distribution for the B6SJL samples while the second peak fits more closely to the distribution of survival times for the C57BL/6, and the difference in magnitude is, in part, a product of the lower prevalence of C57BL/6 samples in the dataset. The regression output with survival as a response finds a significant difference between the two genetic background strains within each sex group (p < 0.05) with an adjusted R² of 0.4867 when onset is included as a predictor (Fig. 4A). The predicted mean lifespans for each category with a representative onset were as follows: female B6SJL 135.77 \pm 2.93 days, female C57BL/6 149.53 \pm 3.30 days, male B6SJL 129.01 \pm 3.1 days, male C57BL/6 145.95 \pm 2.33 days, mixed sex B6SJL 127.39 \pm 1.46 days, and mixed sex C57BL/6 144.4 \pm 1.97 days. As in the case of linear model for onset, there were no withinstrain differences due to sex after Bonferroni correction



Fig. 2. The effect of G93A mouse genetic background strain and sex on the age at disease onset. (A) Linear regression was performed with the mean age at onset for each group modeled as the response with mouse sex and genetic background factored as categorical predictors with full interaction. Data represents the predicted response \pm SEM computed at values of the interaction term corresponding to each category. Observations were weighted on the basis of sample size. *p < 0.05 after Bonferroni correction. (B) The time until onset was modeled as Cox proportional hazards with mouse sex and genetic background factored as categorical predictors with full interaction. Data represents the survival curve for onset predicted by the Cox model for combinations of categorical predictors. Observations were weighted as frequencies given by the group sample size.

100

120

Age (Davs)

140

160

80

for this model. However, the survival curves generated from the analogous Cox model show a sex difference within the B6SJL background with the females surviving longer than the males or mixed groups (p < 0.001), with no sex differences found within C57BL/6 background mice (p > 0.05) (Fig. 4B). Additionally, the survival curves clearly show prolonged lifespan for the C57BL/6 background mice compared to their B6SJL counterparts (p < 0.001).

The result that the B6SJL mixed group survival is lower than both the male-only and female-only group is peculiar, as the onset results for the same data show what one would expect with respect to mixed mice (Fig. 2). To investigate this result, we iterated the model with subsets of the original data and with altered



Fig. 3. The variation in onset times by onset determination criteria and genetic background strain. Linear regression was performed with the mean age at onset for each group modeled as the response and the onset definition category and genetic background factored as categorical predictors with full interaction. Data represents the predicted response \pm SEM computed at values of the categorical interaction term corresponding to each category. Observations were weighted on the basis of sample size. *p < 0.05 for the comparison between the marked column and the "General" group within the strain.

weights and found the separation of the mixed sex survival curve from the male and female curves to be a robust result. Given that greater than two-thirds of the mixed sex studies do not comment regarding gender bias or distribution (see Data Overview and Table 1), we cannot rule out unintended gender bias, which combined with statistical variation, could possibly account for the unexplained lower survival curve of the mixed sex groups, especially in B6SJL.

Disease duration

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The disease duration data are distributed roughly normally with the majority of the data falling between 10 and 40 days (Fig. 1E). Three options for linear regression are available when duration is the response: onset or survival can be included as continuous interacting covariates or duration can be treated solely as a function of the categorical predictors. In the case of only categorical predictors, the adjusted R^2 for the model is extremely low at 0.056. The addition of onset as a predictor in that model returns an adjusted R^2 of 0.642 and the replacement of the onset term with survival gives an R^2 of 0.225. The predicted mean outputs for the high fit model that includes onset as a predictor are shown in Fig. 5A. Within each sex group, the mice on a C57BL/6 background showed lengthened disease duration. There were no significant

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Fig. 4. The effect of G93A mouse genetic background strain and sex on the age at death. (A) Linear regression was performed with the mean age at death for each group modeled as the response with mouse sex and genetic background factored as categorical predictors and onset included as a continuous predictor with full interaction between the terms. Data represents the predicted response \pm SEM computed at values of the categorical interaction term corresponding to each category at the group-specific mean onset. Observations were weighted on the basis of sample size. p < 0.05 after Bonferroni correction. (B) The age at death was modeled as Cox proportional hazards with mouse sex and genetic background factored as categorical predictors with full interaction. Onset was included as an independent continuous predictor. Data represents the survival curve predicted by the Cox model for combinations of categorical predictors. Observations were weighted as frequencies given by the group sample size.

differences in disease duration due to sex within genetic background groups after Bonferroni correction. The predicted mean disease durations for the B6SJL and C57BL/6 mice were 25.49 ± 1.22 days and 41.95 ± 1.40 days, respectively. The Cox proportional hazards model with the probability of survival factored as the response at a particular day post-onset (Fig. 5B) shows a similar pattern to that which is seen when the age is used as the time point (Fig. 4B). Comparisons on the output imply that female mice in the B6SJL category exhibit longer disease duration than mixed mice (p = 0.009) and male groups (p = 0.054, Bonferroni corrected). Analogous to the



Fig. 5. The effect of G93A mouse genetic background strain and sex on the duration of disease. (A) Linear regression was performed with the disease duration for each group modeled as the response with mouse sex and genetic background factored as categorical predictors and onset included as a continuous predictor with full interaction between the terms. Data represents the predicted response \pm SEM computed at values of the categorical interaction term corresponding to each category at the group-specific mean onset. Observations were weighted on the basis of sample size. p < 0.05 after Bonferroni correction. (B) The disease duration was modeled as Cox proportional hazards with mouse sex and genetic background factored as categorical predictors with full interaction. Onset was included as an independent continuous predictor. Data represents the survival curve predicted by the Cox model for combinations of categorical predictors. Observations were weighted as frequencies given by the group sample size.

survival model, this sex difference is not apparent within the C57BL/6 background groups, with no statistically significant differences between any of the C57BL/6 sex groups (p > 0.05, Bonferroni corrected) while overall the C57BL/6 mice show extended disease duration compared to the B6SJL mice (p = 0.001).

DISCUSSION

As the SOD1 G93A mouse is the primary animal model for ALS research, it is important that the disease

progression characteristics of commonly used variants of the model are known so that researchers may be better equipped to interpret their results. The results of this analysis confirm the existence of sex and genetic background related effects on disease endpoints. We find that the mice bred on the C57BL/6 background mice show delayed onset of disease, extended lifespan, and extended disease duration compared to their B6SJL counterparts. The often reported female-related neuroprotective protective effect on lifespan was only observed for B6SJL mice and not for those on C57BL/6 background, in corroboration with the results of Heiman-Patterson [27, 28] who reports a similar background-dependent effect of sex on disease, but these results conflict with the results of Mancuso [26]. However, the noted difference in result is not troubling, as the Heiman-Patterson study reports a larger colony size than Mancuso study. Overall, genetic background and sex had a smaller effect on measures of disease onset than they did on the lifespan or disease duration. This warrants further investigation, as our preliminary analysis into onset qualification methodology (Fig. 3) indicates that the variation in methodology used to determine onset is a large contributor to numerical variation in disease onset. As the experimental record expands in size, future comprehensive aggregate analysis that integrates quantitative measures of body weight, gait, motor performance tests, muscular size, EMG, and neuron counts in conjunction with genetic background and sex information has the potential to give insight into the mechanisms through which genetics affect the various systems associated with disease progression.

Sex

Overall, the expected difference in sex groups was not observed to a large degree when the mean predicted values in onset, lifespan, and disease duration are compared for the various linear models in that the difference is not statistically significant (p > 0.05)after Bonferroni correction). However, the subsequent survival analysis with Cox proportional hazards regression reveals the linear modeling technique to be conservative, as the minute qualitative difference observed between sex groups (Figs. 2A, 4A, 5A) becomes statistically significant when the hazard functions are compared (Figs. 2B, 4B, 5B), suggesting that the sex difference does exist, but is overshadowed by the larger effect due to genetic background when the data are modeled linearly. That comparison of hazard ratios suggests a small but significant effect of the female sex compared to males in terms of the onset

for mice in both groups (p < 0.001) (Fig. 2B). Interestingly, the magnitude of the sex difference is larger for the survival data (Fig. 4B), but only within the B6SJL group. Additionally, there are no differences in lifespan due to sex within the C57BL/6 group. Female mice in the B6SJL category showed longer disease duration than their mixed (p=0.009) and male (p=0.054)after Bonferroni correction) counterparts, but the effect was smaller than when the overall lifespan was compared. The existence of a strain-dependent effect of sex on the progression of disease implies that the inherent genetic differences observed between backgrounds have some interactive effect on the manifestation of the female hormone-related protective effect [43] and further investigation of this relationship may provide insight into the underlying pathophysiological mechanisms of ALS. For example, given that ALS is considered a multi-factorial disease, sex-dependent differences in disease progression and overall duration may only be present with certain types of underlying ALS etiologies rather than homogenously across all ALS phenotypes.

Genetic background

The results of this analysis imply that mice bred on the C57BL/6 genetic background experience disease onset and death at a later age than B6SJL mice and that the duration of disease in C57BL/6 mice is significantly extended. Survival and disease duration varied on the basis of genetic background within each sex group independently, but onset varied on the basis of genetic background only within the mixed sex group and when the results were aggregated. Without regards to sex, the mean onset and lifespan was found to be 99.27 ± 1.79 days and 130.23 ± 1.22 days for B6SJL background mice and 109.9 ± 2.06 days and 146.67 ± 1.42 days for C57BL/6 mice. The estimate for the B6SJL survival corresponds fairly well to the 128.9 ± 9.1 days [30] reported by Jackson Laboratories; however, the estimate for C57BL/6 survival poorly aligns with the 157.1 ± 9.3 days reported by Jackson Laboratories [30]. Previously, it has been shown that C57BL/6 background mice show deficits in motor neuron counts, compound muscle action potentials, and motor potentials two to four weeks earlier than their B6SJL counterparts [26] without a difference in onset measured by rotarod performance. As our results show a later onset and longer lifespan for the C57BL/6 mice, this implies that those mice experience milder symptoms despite a potentially more severe underlying pathology.

The mechanism through which the genetic background affects disease progression is unclear, but previous studies have shown that various mouse models exhibit phenotype variation that may affect ALS progression. These differences may manifest as variation in microglia-mediated neuro-inflammation [28, 29] and/or the progression or affect of hyperactivity of metabolic processes [30] inherent in ALS progression [31]. In particular, Jackson Laboratories reports hyperactivity of metabolism in terms of increased oxygen consumption, decreased circulating insulin levels, abnormal energy expenditures, etc. in the B6SJL mice but not for C57BL/6 mice [30], potentially explaining the more rapid progression of disease in the B6SJL mice.

On a similar note, a recent study of non-familial clinical ALS patients showed that antecedent disease is substantially less prevalent among ALS patients, including metabolic disturbances like diabetes, obesity and thyroid disease, which suggests the possibility of neuroprotection-either "other disease is protective of ALS" or "ALS is protective of other disease" [32]. In fact, the pathophysiological process oscillations seen in models examining the SOD1 G93A underlying pathophysiology, suggest the possibility of "hypervigilant regulation" (in control theory a too-high feedback gain). Regulatory delays combined with the too-high feedback gain(s) of hypervigilant regulation, could ultimately result in system instabilities identified in the SOD1 G93A model that dramatically impact the onset and progression of ALS [32-34]. In short, genetic differences underlying cellular and systemic regulation could result in different ALS phenotypes, both in transgenic mouse models and in humans. Further assessment of pathology dynamics will help to distinguish phenotypic characteristics [32, 33, 35].

Onset category

It is of interest to the field to characterize the time course of the onset of various symptoms for each mouse variant so that individual results can be better understood and gauged in the context of other studies. There are differences in the sensitivities of various measures of onset to detect specific aspects of the underlying pathophysiology [25, 55]. Some researchers utilize a graded motor score that accounts for visualized gait impairments, limb tremors, and motor deficits [56, 57], but those systems are not standardized between studies. The linear modeling method used to investigate sex and genetic background is easily extended to allow for onset characterization to be analyzed as a categorical covariate. Within both the B6SJL and C57BL/6 background groups, onsets due to tremors were developed at an earlier age than the control grouping, which is consistent with reports that tremors develop prior to outright motor deficits [58]. Bodyweight declines are often cited as developing prior to or in sync with the development of tremors [59], but we were unable to test this directly due to the low prevalence of bodyweight onsets in the dataset.

CONCLUSION

Overall, the analysis of data from 97 articles confirmed that the SOD1 G93A mouse lifespan, disease duration, and onset vary largely between the two most widely used genetic backgrounds, with those bred on the C57BL/6 background showing a milder delayed onset of disease with increased lifespans and longer disease duration when compared to the B6SJL mouse. The expected delayed onset and extended lifespan typically reported in female mice was observed only in SOD1 G93A mice on the B6SJL background, and the effect was altogether small compared to the strain difference. The existence of a strain-dependent effect of sex on the progression of disease demonstrates the importance of the proper identification of groupspecific characteristics and their potential relationships to the underlying pathophysiology. Finally, different onset determination protocols (general, rotarod, tremor, etc.) can result in notably different mean onsets. Based on the initial analysis of this study, it appears that tremors or grasping protocols are more sensitive for onset determination whereas motor function protocols like rotarod are better for determining motor deficit progression and endpoint; however, further analysis is needed to confirm this finding. Like all fields, the SOD1 G93A transgenic mouse field would benefit from greater consistency and documentation in experimental methodology to assist in consolidating conflicting pathophysiology assessment and/or treatments and lessen the impact of confounding variables [33, 60].

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CONFLICTS OF INTEREST

The authors report no conflicts of interest.

REFERENCES

- [1] Mitchell, C. S., Halicek, M., Kim, R., Tilva, K., Lee, R. H. in Society for Neuroscience, San Diego, CA, 2013.
- [2] Turner, B. J., Lopes, E. C., Cheema, S. S. The serotonin precursor 5-hydroxytryptophan delays neuromuscular disease in murine familial amyotrophic lateral sclerosis. Amyotroph Lateral Scler Other Motor Neuron Disord. 2003; 4: 171-176.
- [3] Wu, A. S., Kiaei, M., Aguirre, N., Crow, J. P., Calingasan, N. Y., Browne, S. E., Beal, M. F. Iron porphyrin treatment extends survival in a transgenic animal model of amyotrophic lateral sclerosis. J Neurochem. 2003; 85: 142-150.
- [4] Fischer, L. R., Culver, D. G., Tennant, P., Davis, A. A., Wang, M., Castellano-Sanchez, A., Khan, J., Polak, M. A., Glass, J. D. Amyotrophic lateral sclerosis is a distal axonopathy: Evidence in mice and man. Exp Neurol. 2004; 185: 232-240.
- [5] Kaspar, B. K., Frost, L. M., Christian, L., Umapathi, P., Gage, F. H. Synergy of insulin-like growth factor-1 and exercise in amyotrophic lateral sclerosis. Ann Neurol. 2005; 57: 649-655.
- [6] Petri, S., Kiaei, M., Damiano, M., Hiller, A., Wille, E., Manfredi, G., Calingasan, N. Y., Szeto, H. H., Beal, M. F. Cell-permeable peptide antioxidants as a novel therapeutic approach in a mouse model of amyotrophic lateral sclerosis. J Neurochem. 2006; 98: 1141-1148.
- [7] Solomon, J. N., Lewis, C. A., Ajami, B., Corbel, S. Y., Rossi, F. M., Krieger, C. Origin and distribution of bone marrowderived cells in the central nervous system in a mouse model of amyotrophic lateral sclerosis. Glia. 2006; 53: 744-753.
- [8] Towne, C., Raoul, C., Schneider, B. L., Aebischer, P. Systemic AAV6 delivery mediating RNA interference against SOD1: Neuromuscular transduction does not alter disease progression in fALS mice. Mol Ther. 2008; 16: 1018-1025.
- [9] Neymotin, A., Petri, S., Calingasan, N. Y., Wille, E., Schafer, P., Stewart, C., Hensley, K., Beal, M. F., Kiaei, M. Lenalidomide (Revlimid) administration at symptom onset is neuroprotective in a mouse model of amyotrophic lateral sclerosis. Exp Neurol. 2009; 220: 191-197.
- [10] Sekiya, M., Ichiyanagi, T., Ikeshiro, Y., Yokozawa, T. The Chinese prescription Wen-Pi-Tang extract delays disease onset in amyotrophic lateral sclerosis model mice while attenuating the activation of glial cells in the spinal cord. Biol Pharm Bull. 2009; 32: 382-388.
- [11] Knippenberg, S., Thau, N., Dengler, R., Petri, S. Significance of behavioural tests in a transgenic mouse model of amyotrophic lateral sclerosis (ALS). Behav Brain Res. 2010; 213: 82-87.
- [12] Calvo, A. C., Moreno-Igoa, M., Mancuso, R., Manzano, R., Olivan, S., Munoz, M. J., Penas, C., Zaragoza, P., Navarro, X., Osta, R. Lack of a synergistic effect of a non-viral ALS gene therapy based on BDNF and a TTC fusion molecule. Orphanet J Rare Dis. 2011; 6: 10.
- [13] Genestine, M., Caricati, E., Fico, A., Richelme, S., Hassani, H., Sunyach, C., Lamballe, F., Panzica, G. C., Pettmann, B., Helmbacher, F., Raoul, C., Maina, F., Dono, R. Enhanced neuronal Met signalling levels in ALS mice delay disease onset. Cell Death Dis. 2011; 2: e130.
- [14] Liebetanz, D., Hagemann, K., von Lewinski, F., Kahler, E., Paulus, W. Extensive exercise is not harmful in amyotrophic lateral sclerosis. Eur J Neurosci. 2004; 20: 3115-3120.
- [15] West, M., Mhatre, M., Ceballos, A., Floyd, R. A., Grammas, P., Gabbita, S. P., Hamdheydari, L., Mai, T., Mou, S., Pye, Q. N., Stewart, C., West, S., Williamson, K. S., Zemlan, F., Hensley, K. The arachidonic acid 5-lipoxygenase inhibitor nordihydroguaiaretic acid inhibits tumor necrosis

factor alpha activation of microglia and extends survival of G93A-SOD1 transgenic mice. J Neurochem. 2004; 91: 133-143.

- [16] Hamadeh, M. J., Rodriguez, M. C., Kaczor, J. J., Tarnopolsky, M. A. Caloric restriction transiently improves motor performance but hastens clinical onset of disease in the Cu/Zn-superoxide dismutase mutant G93A mouse. Muscle Nerve. 2005; 31: 214-220.
- [17] Corti, S., Locatelli, F., Donadoni, C., Guglieri, M., Papadimitriou, D., Strazzer, S., Del Bo, R., Comi, G. P. Wild-type bone marrow cells ameliorate the phenotype of SOD1-G93A ALS mice and contribute to CNS, heart and skeletal muscle tissues. Brain. 2004; 127: 2518-2532.
- [18] Corti, S., Locatelli, F., Papadimitriou, D., Del Bo, R., Nizzardo, M., Nardini, M., Donadoni, C., Salani, S., Fortunato, F., Strazzer, S., Bresolin, N., Comi, G. P. Neural stem cells LewisX+ CXCR4+ modify disease progression in an amyotrophic lateral sclerosis model. Brain. 2007; 130: 1289-1305.
- [19] Turner, B. J., Ackerley, S., Davies, K. E., Talbot, K. Dismutase-competent SOD1 mutant accumulation in myelinating Schwann cells is not detrimental to normal or transgenic ALS model mice. Hum Mol Genet. 2010; 19: 815-824.
- [20] Dibaj, P., Zschuntzsch, J., Steffens, H., Scheffel, J., Goricke, B., Weishaupt, J. H., Le Meur, K., Kirchhoff, F., Hanisch, U. K., Schomburg, E. D., Neusch, C. Influence of methylene blue on microglia-induced inflammation and motor neuron degeneration in the SOD1(G93A) model for ALS. PLoS One. 2012; 7: e43963.
- [21] Ringer, C., Buning, L. S., Schafer, M. K., Eiden, L. E., Weihe, E., Schutz, B. PACAP signaling exerts opposing effects on neuroprotection and neuroinflammation during disease progression in the SOD1(G93A) mouse model of amyotrophic lateral sclerosis. Neurobiol Dis. 2013; 54: 32-42.
- [22] Chiba, T., Hashimoto, Y., Tajima, H., Yamada, M., Kato, R., Niikura, T., Terashita, K., Schulman, H., Aiso, S., Kita, Y., Matsuoka, M., Nishimoto, I. Neuroprotective effect of activity-dependent neurotrophic factor against toxicity from familial amyotrophic lateral sclerosis-linked mutant SOD1 *in vitro* and *in vivo*. J Neurosci Res. 2004; 78: 542-552.
- [23] Chiba, T., Yamada, M., Sasabe, J., Terashita, K., Aiso, S., Matsuoka, M., Nishimoto, I. Colivelin prolongs survival of an ALS model mouse. Biochem Biophys Res Commun. 2006; 343: 793-798.
- [24] Crosio, C., Casciati, A., Iaccarino, C., Rotilio, G., Carri, M. T. Bel2a1 serves as a switch in death of motor neurons in amyotrophic lateral sclerosis. Cell Death Differ. 2006; 13: 2150-2153.
- [25] Hayworth, C. R., Gonzalez-Lima, F. Pre-symptomatic detection of chronic motor deficits and genotype prediction in congenic B6.SOD1(G93A) ALS mouse model. Neuroscience. 2009; 164: 975-985.
- [26] Mancuso, R., Olivan, S., Mancera, P., Pasten-Zamorano, A., Manzano, R., Casas, C., Osta, R., Navarro, X. Effect of genetic background on onset and disease progression in the SOD1-G93A model of amyotrophic lateral sclerosis. Amyotroph Lateral Scler. 2012; 13: 302-310.
- [27] Heiman-Patterson, T. D., Deitch, J. S., Blankenhorn, E. P., Erwin, K. L., Perreault, M. J., Alexander, B. K., Byers, N., Toman, I., Alexander, G. M. Background and gender effects on survival in the TgN(SOD1-G93A)1Gur mouse model of ALS. J Neurol Sci. 2005; 236: 1-7.

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- [28] Heiman-Patterson, T. D., Sher, R. B., Blankenhorn, E. A., Alexander, G., Deitch, J. S., Kunst, C. B., Maragakis, N., Cox, G. Effect of genetic background on phenotype variability in transgenic mouse models of amyotrophic lateral sclerosis: A window of opportunity in the search for genetic modifiers. Amyotroph Lateral Scler. 2011; 12: 79-86.
- [29] Nikodemova, M., Watters, J. J. Outbred ICR/CD1 mice display more severe neuroinflammation mediated by microglial TLR4/CD14 activation than inbred C57Bl/6 mice. Neuroscience. 2011; 190: 67-74.
- [30] Mouse Genome Database (MGD) at the Mouse Genome Informatics website, The Jackson Laboratory, http://www.informatics.jax.org, Accessed December 7.
- [31] Dupuis, L., Pradat, P. -F., Ludolph, A. C., Loeffler, J. -P. Energy metabolism in amyotrophic lateral sclerosis. The Lancet. Neurology. 2011; 10: 75-82.
- [32] Mitchell, C. S., Hollinger, S. K., Goswami, S. D., Polak, M., Lee, R. H., Glass, J. D. Antecedent disease is less prevalent in Amyotrophic Lateral Sclerosis. Neurodegener Dis. 2015. (Accepted 11/11/14: Issue not yet assigned).
- [33] Mitchell, C. S., Lee, R. H. In Amyotrophic Lateral Sclerosis InTech, 2012. http://www.intechopen.com/books/amyo trophic-lateral-sclerosis/dynamic-meta-analysis-as-a-thera peutic-prediction-tool-for-amyotrophic-lateral-sclerosis.
- [34] Mitchell, C. S., Lee, R. H. Cargo distributions differentiate pathological axonal transport impairments. J Theor Biol. 2012; 300: 277-291.
- [35] Mitchell, C. S., Lee, R. H. Pathology dynamics predict spinal cord injury therapeutic success. J Neurotrauma. 2008; 25: 1483-1497.
- [36] Gurney, M. E. Transgenic animal models of familial amyotrophic lateral sclerosis. J Neurol. 1997; 244(Suppl 2): S15-S20.
- [37] Gurney, M. E. Transgenic-mouse model of amyotrophic lateral sclerosis. N Engl J Med. 1994; 331: 1721-1722.
- [38] Siddique, T., Deng, H. X. Genetics of amyotrophic lateral sclerosis. Hum Mol Genet. 1996; 5 Spec No: 1465-1470.
- [39] Synofzik, M., Fernandez-Santiago, R., Maetzler, W., Schols, L., Andersen, P. M. The human G93A SOD1 phenotype closely resembles sporadic amyotrophic lateral sclerosis. J Neurol Neurosurg Psychiatry. 2010; 81: 764-767.
- [40] Acevedo-Arozena, A., Kalmar, B., Essa, S., Ricketts, T., Joyce, P., Kent, R., Rowe, C., Parker, A., Gray, A., Hafezparast, M., Thorpe, J. R., Greensmith, L., Fisher, E. M. A comprehensive assessment of the SOD1G93A low-copy transgenic mouse, which models human amyotrophic lateral sclerosis. Dis Model Mech. 2011; 4: 686-700.
- [41] Ripps, M. E., Huntley, G. W., Hof, P. R., Morrison, J. H., Gordon, J. W. Transgenic mice expressing an altered murine superoxide dismutase gene provide an animal model of amyotrophic lateral sclerosis. Proc Natl Acad Sci U S A. 1995; 92: 689-693.
- [42] Wooley, C. M., Sher, R. B., Kale, A., Frankel, W. N., Cox, G. A., Seburn, K. L. Gait analysis detects early changes in transgenic SOD1(G93A) mice. Muscle Nerve. 2005; 32: 43-50.
- [43] Bame, M., Pentiak, P. A., Needleman, R., Brusilow, W. S. Effect of sex on lifespan, disease progression, and the response to methionine sulfoximine in the SOD1 G93A mouse model for ALS. Gend Med. 2012; 9: 524-535.
- [44] Choi, C. I., Lee, Y. D., Gwag, B. J., Cho, S. I., Kim, S. S., Suh-Kim, H. Effects of estrogen on lifespan and motor functions in female hSOD1 G93A transgenic mice. J Neurol Sci. 2008; 268: 40-47.

- [45] Steinacker, P., Hawlik, A., Lehnert, S., Jahn, O., Meier, S., Gorz, E., Braunstein, K. E., Krzovska, M., Schwalenstocker, B., Jesse, S., Propper, C., Bockers, T., Ludolph, A., Otto, M. Neuroprotective function of cellular prion protein in a mouse model of amyotrophic lateral sclerosis. Am J Pathol. 2010; 176: 1409-1420.
- [46] Li, W., Brakefield, D., Pan, Y., Hunter, D., Myckatyn, T. M., Parsadanian, A. Muscle-derived but not centrally derived transgene GDNF is neuroprotective in G93A-SOD1 mouse model of ALS. Exp Neurol. 2007; 203: 457-471.
- [47] Miana-Mena, F. J., Munoz, M. J., Yague, G., Mendez, M., Moreno, M., Ciriza, J., Zaragoza, P., Osta, R. Optimal methods to characterize the G93A mouse model of ALS. Amyotroph Lateral Scler Other Motor Neuron Disord. 2005; 6: 55-62.
- [48] Shimojo, Y., Kosaka, K., Noda, Y., Shimizu, T., Shirasawa, T. Effect of rosmarinic acid in motor dysfunction and life span in a mouse model of familial amyotrophic lateral sclerosis. J Neurosci Res. 2010; 88: 896-904.
- [49] Kiaei, M., Kipiani, K., Petri, S., Chen, J., Calingasan, N. Y., Beal, M. F. Celastrol blocks neuronal cell death and extends life in transgenic mouse model of amyotrophic lateral sclerosis. Neurodegener Dis. 2005; 2: 246-254.
- [50] Schutz, B., Reimann, J., Dumitrescu-Ozimek, L., Kappes-Horn, K., Landreth, G. E., Schurmann, B., Zimmer, A., Heneka, M. T. The oral antidiabetic pioglitazone protects from neurodegeneration and amyotrophic lateral sclerosislike symptoms in superoxide dismutase-G93A transgenic mice. J Neurosci. 2005; 25: 7805-7812.
- [51] McCombe, P. A., Henderson, R. D. Effects of gender in amyotrophic lateral sclerosis. Gend Med. 2010; 7: 557-570.
 [52] (2013) StataCorp LP, College Station, TX.
- 52] (2015) StataColp LF, Conege Station, 1A
- [53] Cruz-Sanchez, F. F., Moral, A., Rossi, M. L., Quinto, L., Castejon, C., Tolosa, E., de Belleroche, J. Synaptophysin in spinal anterior horn in aging and ALS: An immunohistological study. J Neural Transm. 1996; 103: 1317-1329.
- [54] Cox D. Regression Models and Life-Tables In Breakthroughs in Statistics, Kotz S, Johnson N, eds. Springer New York, 1992, pp. 527-541.
- [55] Lepore, A. C., O'Donnell, J., Kim, A. S., Williams, T., Tuteja, A., Rao, M. S., Kelley, L. L., Campanelli, J. T., Maragakis, N. J. Human glial-restricted progenitor transplantation into cervical spinal cord of the SOD1 mouse model of ALS. PLoS One. 2011; 6: e25968.
- [56] Alves, C. J., de Santana, L. P., dos Santos, A. J., de Oliveira, G. P., Duobles, T., Scorisa, J. M., Martins, R. S., Maximino, J. R., Chadi, G. Early motor and electrophysiological changes in transgenic mouse model of amyotrophic lateral sclerosis and gender differences on clinical outcome. Brain Res. 2011; 1394: 90-104.
- [57] Lerman, B. J., Hoffman, E. P., Sutherland, M. L., Bouri, K., Hsu, D. K., Liu, F. T., Rothstein, J. D., Knoblach, S. M. Deletion of galectin-3 exacerbates microglial activation and accelerates disease progression and demise in a SOD1(G93A) mouse model of amyotrophic lateral sclerosis. Brain Behav. 2012; 2: 563-575.
- [58] Kasai, A., Kinjo, T., Ishihara, R., Sakai, I., Ishimaru, Y., Yoshioka, Y., Yamamuro, A., Ishige, K., Ito, Y., Maeda, S. Apelin deficiency accelerates the progression of amyotrophic lateral sclerosis. PLoS One. 2011; 6: e23968.
- [59] Matsumoto, A., Okada, Y., Nakamichi, M., Nakamura, M., Toyama, Y., Sobue, G., Nagai, M., Aoki, M., Itoyama, Y., Okano, H. Disease progression of human SOD1 (G93A) transgenic ALS model rats. Journal Of Neuroscience Research. 2006; 83: 119-133.

- [60] Foley, A. M., Ammar, Z. M., Lee, R. H., Mitchell, C. S. Systematic review of the relationship between amyloid-beta levels and measures of transgenic mouse cognitive deficit in Alzheimer's disease. J Alzheimers Dis. 2015; 44: 787-795.
- [61] Ferrante, R. J., Klein, A. M., Dedeoglu, A., Beal, M. F. Therapeutic efficacy of EGb761 (Gingko biloba extract) in a transgenic mouse model of amyotrophic lateral sclerosis. J Mol Neurosci. 2001; 17: 89-96.
- [62] Azari, M. F., Lopes, E. C., Stubna, C., Turner, B. J., Zang, D., Nicola, N. A., Kurek, J. B., Cheema, S. S. Behavioural and anatomical effects of systemically administered leukemia inhibitory factor in the SOD1(G93A G1H) mouse model of familial amyotrophic lateral sclerosis. Brain Res. 2003; 982: 92-97.
- [63] Pompl, P. N., Ho, L., Bianchi, M., McManus, T., Qin, W., Pasinetti, G. M. A therapeutic role for cyclooxygenase-2 inhibitors in a transgenic mouse model of amyotrophic lateral sclerosis. FASEB J. 2003; 17: 725-727.
- [64] Zheng, C., Nennesmo, I., Fadeel, B., Henter, J. I. Vascular endothelial growth factor prolongs survival in a transgenic mouse model of ALS. Ann Neurol. 2004; 56: 564-567.
- [65] Schutz, B. Imbalanced excitatory to inhibitory synaptic input precedes motor neuron degeneration in an animal model of amyotrophic lateral sclerosis. Neurobiol Dis. 2005; 20: 131-140.
- [66] Ito, H., Wate, R., Zhang, J., Ohnishi, S., Kaneko, S., Ito, H., Nakano, S., Kusaka, H. Treatment with edaravone, initiated at symptom onset, slows motor decline and decreases SOD1 deposition in ALS mice. Exp Neurol. 2008; 213: 448-455.
- [67] Rohde, G., Kermer, P., Reed, J. C., Bahr, M., Weishaupt, J. H. Neuron-specific overexpression of the co-chaperone Bcl-2associated athanogene-1 in superoxide dismutase 1(G93A)transgenic mice. Neuroscience. 2008; 157: 844-849.
- [68] Smittkamp, S. E., Brown, J. W., Stanford, J. A. Time-course and characterization of orolingual motor deficits in B6SJL-Tg(SOD1-G93A)1Gur/J mice. Neuroscience. 2008; 151: 613-621.
- [69] Stam, N. C., Nithianantharajah, J., Howard, M. L., Atkin, J. D., Cheema, S. S., Hannan, A. J. Sex-specific behavioural effects of environmental enrichment in a transgenic mouse model of amyotrophic lateral sclerosis. Eur J Neurosci. 2008; 28: 717-723.
- [70] Gianforcaro, A., Hamadeh, M. J. Dietary vitamin D3 supplementation at 10x the adequate intake improves functional capacity in the G93A transgenic mouse model of ALS, a pilot study. CNS Neurosci Ther. 2012; 18: 547-557.
- [71] Knippenberg, S., Thau, N., Schwabe, K., Dengler, R., Schambach, A., Hass, R., Petri, S. Intraspinal injection of human umbilical cord blood-derived cells is neuroprotective in a transgenic mouse model of amyotrophic lateral sclerosis. Neurodegener Dis. 2012; 9: 107-120.
- [72] Gianforcaro, A., Solomon, J. A., Hamadeh, M. J. Vitamin D(3) at 50x AI attenuates the decline in paw grip endurance, but not disease outcomes, in the G93A mouse model of ALS, and is toxic in females. PLoS One. 2013; 8: e30243.
- [73] Chiu, A. Y., Zhai, P., Dal Canto, M. C., Peters, T. M., Kwon, Y. W., Prattis, S. M., Gurney, M. E. Age-dependent penetrance of disease in a transgenic mouse model of familial amyotrophic lateral sclerosis. Mol Cell Neurosci. 1995; 6: 349-362.
- [74] Hamadeh, M. J., Tarnopolsky, M. A. Transient caloric restriction in early adulthood hastens disease endpoint in male, but not female, Cu/Zn-SOD mutant G93A mice. Muscle Nerve. 2006; 34: 709-719.

- [75] Ohta, Y., Nagai, M., Nagata, T., Murakami, T., Nagano, I., Narai, H., Kurata, T., Shiote, M., Shoji, M., Abe, K. Intrathecal injection of epidermal growth factor and fibroblast growth factor 2 promotes proliferation of neural precursor cells in the spinal cords of mice with mutant human SOD1 gene. J Neurosci Res. 2006; 84: 980-992.
- [76] Marcuzzo, S., Zucca, I., Mastropietro, A., de Rosbo, N. K., Cavalcante, P., Tartari, S., Bonanno, S., Preite, L., Mantegazza, R., Bernasconi, P. Hind limb muscle atrophy precedes cerebral neuronal degeneration in G93A-SOD1 mouse model of amyotrophic lateral sclerosis: A longitudinal MRI study. Exp Neurol. 2011; 231: 30-37.
- [77] Zhao, Z., Lange, D. J., Voustianiouk, A., MacGrogan, D., Ho, L., Suh, J., Humala, N., Thiyagarajan, M., Wang, J., Pasinetti, G. M. A ketogenic diet as a potential novel therapeutic intervention in amyotrophic lateral sclerosis. BMC Neurosci. 2006; 7: 29.
- [78] Hottinger, A. F., Fine, E. G., Gurney, M. E., Zurn, A. D., Aebischer, P. The copper chelator d-penicillamine delays onset of disease and extends survival in a transgenic mouse model of familial amyotrophic lateral sclerosis. Eur J Neurosci. 1997; 9: 1548-1551.
- [79] Andreassen, O. A., Dedeoglu, A., Klivenyi, P., Beal, M. F., Bush, A. I. N-acetyl-L-cysteine improves survival and preserves motor performance in an animal model of familial amyotrophic lateral sclerosis. Neuroreport. 2000; 11: 2491-2493.
- [80] Li, M., Ona, V. O., Guegan, C., Chen, M., Jackson-Lewis, V., Andrews, L. J., Olszewski, A. J., Stieg, P. E., Lee, J. P., Przedborski, S., Friedlander, R. M. Functional role of caspase-1 and caspase-3 in an ALS transgenic mouse model. Science. 2000; 288: 335-339.
- [81] Acsadi, G., Anguelov, R. A., Yang, H., Toth, G., Thomas, R., Jani, A., Wang, Y., Ianakova, E., Mohammad, S., Lewis, R. A., Shy, M. E. Increased survival and function of SOD1 mice after glial cell-derived neurotrophic factor gene therapy. Hum Gene Ther. 2002; 13: 1047-1059.
- [82] Drachman, D. B., Frank, K., Dykes-Hoberg, M., Teismann, P., Almer, G., Przedborski, S., Rothstein, J. D. Cyclooxygenase 2 inhibition protects motor neurons and prolongs survival in a transgenic mouse model of ALS. Ann Neurol. 2002; 52: 771-778.
- [83] Wang, L. J., Lu, Y. Y., Muramatsu, S., Ikeguchi, K., Fujimoto, K., Okada, T., Mizukami, H., Matsushita, T., Hanazono, Y., Kume, A., Nagatsu, T., Ozawa, K., Nakano, I. Neuroprotective effects of glial cell line-derived neurotrophic factor mediated by an adeno-associated virus vector in a transgenic animal model of amyotrophic lateral sclerosis. J Neurosci. 2002; 22: 6920-6928.
- [84] Guo, H., Lai, L., Butchbach, M. E., Stockinger, M. P., Shan, X., Bishop, G. A., Lin, C. L. Increased expression of the glial glutamate transporter EAAT2 modulates excitotoxicity and delays the onset but not the outcome of ALS in mice. Hum Mol Genet. 2003; 12: 2519-2532.
- [85] Turner, B. J., Lopes, E. C., Cheema, S. S. Neuromuscular accumulation of mutant superoxide dismutase 1 aggregates in a transgenic mouse model of familial amyotrophic lateral sclerosis. Neurosci Lett. 2003; 350: 132-136.
- [86] Takahashi, S., Kulkarni, A. B. Mutant superoxide dismutase 1 causes motor neuron degeneration independent of cyclindependent kinase 5 activation by p35 or p25. J Neurochem. 2004; 88: 1295-1304.
- [87] Day, W. A., Koishi, K., Nukuda, H., McLennan, I. S. Transforming growth factor-beta 2 causes an acute improvement

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in the motor performance of transgenic ALS mice. Neurobiol Dis. 2005; 19: 323-330.

- [88] Gilchrist, C. A., Gray, D. A., Stieber, A., Gonatas, N. K., Kopito, R. R. Effect of ubiquitin expression on neuropathogenesis in a mouse model of familial amyotrophic lateral sclerosis. Neuropathol Appl Neurobiol. 2005; 31: 20-33.
- [89] Kiaei, M., Kipiani, K., Chen, J., Calingasan, N. Y., Beal, M. F. Peroxisome proliferator-activated receptor-gamma agonist extends survival in transgenic mouse model of amyotrophic lateral sclerosis. Exp Neurol. 2005; 191: 331-336.
- [90] Kiaei, M., Petri, S., Kipiani, K., Gardian, G., Choi, D. K., Chen, J., Calingasan, N. Y., Schafer, P., Muller, G. W., Stewart, C., Hensley, K., Beal, M. F. Thalidomide and lenalidomide extend survival in a transgenic mouse model of amyotrophic lateral sclerosis. J Neurosci. 2006; 26: 2467-2473.
- [91] Koh, S. H., Lee, S. M., Kim, H. Y., Lee, K. Y., Lee, Y. J., Kim, H. T., Kim, J., Kim, M. H., Hwang, M. S., Song, C., Yang, K. W., Lee, K. W., Kim, S. H., Kim, O. H. The effect of epigallocatechin gallate on suppressing disease progression of ALS model mice. Neurosci Lett. 2006; 395: 103-107.
- [92] Lorenzl, S., Narr, S., Angele, B., Krell, H. W., Gregorio, J., Kiaei, M., Pfister, H. W., Beal, M. F. The matrix metalloproteinases inhibitor Ro 28-2653 [correction of Ro 26-2853] extends survival in transgenic ALS mice. Exp Neurol. 2006; 200: 166-171.
- [93] Petri, S., Kiaei, M., Kipiani, K., Chen, J., Calingasan, N. Y., Crow, J. P., Beal, M. F. Additive neuroprotective effects of a histone deacetylase inhibitor and a catalytic antioxidant in a transgenic mouse model of amyotrophic lateral sclerosis. Neurobiol Dis. 2006; 22: 40-49.
- [94] Weishaupt, J. H., Bartels, C., Polking, E., Dietrich, J., Rohde, G., Poeggeler, B., Mertens, N., Sperling, S., Bohn, M., Huther, G., Schneider, A., Bach, A., Siren, A. L., Hardeland, R., Bahr, M., Nave, K. A., Ehrenreich, H. Reduced oxidative damage in ALS by high-dose enteral melatonin treatment. J Pineal Res. 2006; 41: 313-323.
- [95] Glas, M., Popp, B., Angele, B., Koedel, U., Chahli, C., Schmalix, W. A., Anneser, J. M., Pfister, H. W., Lorenzl, S. A role for the urokinase-type plasminogen activator system in amyotrophic lateral sclerosis. Exp Neurol. 2007; 207: 350-356.
- [96] Joo, I. S., Hwang, D. H., Seok, J. I., Shin, S. K., Kim, S. U. Oral administration of memantine prolongs survival in a transgenic mouse model of amyotrophic lateral sclerosis. J Clin Neurol. 2007; 3: 181-186.
- [97] Wang, H., Guan, Y., Wang, X., Smith, K., Cormier, K., Zhu, S., Stavrovskaya, I. G., Huo, C., Ferrante, R. J., Kristal, B. S., Friedlander, R. M. Nortriptyline delays disease onset in models of chronic neurodegeneration. Eur J Neurosci. 2007; 26: 633-641.
- [98] Garbuzova-Davis, S., Sanberg, C. D., Kuzmin-Nichols, N., Willing, A. E., Gemma, C., Bickford, P. C., Miller, C., Rossi, R., Sanberg, P. R. Human umbilical cord blood treatment in a mouse model of ALS: Optimization of cell dose. PLoS One. 2008; 3: e2494.
- [99] Lev, N., Ickowicz, D., Barhum, Y., Melamed, E., Offen, D. DJ-1 changes in G93A-SOD1 transgenic mice: Implications for oxidative stress in ALS. J Mol Neurosci. 2009; 38: 94-102.
- [100] Shimazawa, M., Tanaka, H., Ito, Y., Morimoto, N., Tsuruma, K., Kadokura, M., Tamura, S., Inoue, T., Yamada, M., Takahashi, H., Warita, H., Aoki, M., Hara, H. An inducer of VGF protects cells against ER stress-induced cell death and pro-

longs survival in the mutant SOD1 animal models of familial ALS. PLoS One. 2010; 5: e15307.

- [101] Moreno-Igoa, M., Calvo, A. C., Ciriza, J., Munoz, M. J., Zaragoza, P., Osta, R. Non-viral gene delivery of the GDNF, either alone or fused to the C-fragment of tetanus toxin protein, prolongs survival in a mouse ALS model. Restor Neurol Neurosci. 2012; 30: 69-80.
- [102] Zhou, C., Zhang, C., Zhao, R., Chi, S., Ge, P., Zhang, C. Human marrow stromal cells reduce microglial activation to protect motor neurons in a transgenic mouse model of amyotrophic lateral sclerosis. J Neuroinflammation. 2013; 10: 52.
- [103] Van Den Bosch, L., Tilkin, P., Lemmens, G., Robberecht, W. Minocycline delays disease onset and mortality in a transgenic model of ALS. Neuroreport. 2002; 13: 1067-1070.
- [104] Pardo, A. C., Wong, V., Benson, L. M., Dykes, M., Tanaka, K., Rothstein, J. D., Maragakis, N. J. Loss of the astrocyte glutamate transporter GLT1 modifies disease in SOD1(G93A) mice. Experimental Neurology. 2006; 201: 120-130.
- [105] Banks, G. T., Bros-Facer, V., Williams, H. P., Chia, R., Achilli, F., Bryson, J. B., Greensmith, L., Fisher, E. M. Mutant glycyl-tRNA synthetase (Gars) ameliorates SOD1(G93A) motor neuron degeneration phenotype but has little affect on Loa dynein heavy chain mutant mice. PLoS One. 2009; 4: e6218.
- [106] Ebert, S., Goos, M., Rollwagen, L., Baake, D., Zech, W. D., Esselmann, H., Wiltfang, J., Mollenhauer, B., Schliebs, R., Gerber, J., Nau, R. Recurrent systemic infections with Streptococcus pneumoniae do not aggravate the course of experimental neurodegenerative diseases. J Neurosci Res. 2010; 88: 1124-1136.
- [107] Hashimoto, K., Hayashi, Y., Watabe, K., Inuzuka, T., Hozumi, I. Metallothionein-III prevents neuronal death and prolongs life span in amyotrophic lateral sclerosis model mice. Neuroscience. 2011; 189: 293-298.
- [108] Feng, X., Peng, Y., Liu, M., Cui, L. DL-3-n-butylphthalide extends survival by attenuating glial activation in a mouse model of amyotrophic lateral sclerosis. Neuropharmacology. 2012; 62: 1004-1010.
- [109] Sharp, P. S., Akbar, M. T., Bouri, S., Senda, A., Joshi, K., Chen, H. J., Latchman, D. S., Wells, D. J., de Belleroche, J. Protective effects of heat shock protein 27 in a model of ALS occur in the early stages of disease progression. Neurobiol Dis. 2008; 30: 42-55.
- [110] Narai, H., Manabe, Y., Nagai, M., Nagano, I., Ohta, Y., Murakami, T., Takehisa, Y., Kamiya, T., Abe, K. Early detachment of neuromuscular junction proteins in ALS mice with SODG93A mutation. Neurol Int. 2009; 1: e16.
- [111] Vaknin, I., Kunis, G., Miller, O., Butovsky, O., Bukshpan, S., Beers, D. R., Henkel, J. S., Yoles, E., Appel, S. H., Schwartz, M. Excess circulating alternatively activated myeloid (M2) cells accelerate ALS progression while inhibiting experimental autoimmune encephalomyelitis. PLoS One. 2011; 6: e26921.
- [112] Yoo, Y. E., Ko, C. P. Treatment with trichostatin A initiated after disease onset delays disease progression and increases survival in a mouse model of amyotrophic lateral sclerosis. Exp Neurol. 2011; 231: 147-159.
- [113] Zhu, Y. B., Sheng, Z. H. Increased axonal mitochondrial mobility does not slow amyotrophic lateral sclerosis (ALS)like disease in mutant SOD1 mice. J Biol Chem. 2011; 286: 23432-23440.
- [114] Ralph, G. S., Radcliffe, P. A., Day, D. M., Carthy, J. M., Leroux, M. A., Lee, D. C., Wong, L. F., Bilsland, L. G.,

Greensmith, L., Kingsman, S. M., Mitrophanous, K. A., Mazarakis, N. D., Azzouz, M. Silencing mutant SOD1 using RNAi protects against neurodegeneration and extends survival in an ALS model. Nat Med. 2005; 11: 429-433.

- [115] Saito, Y., Yokota, T., Mitani, T., Ito, K., Anzai, M., Miyagishi, M., Taira, K., Mizusawa, H. Transgenic small interfering RNA halts amyotrophic lateral sclerosis in a mouse model. J Biol Chem. 2005; 280: 42826-42830.
- [116] Gould, T. W., Buss, R. R., Vinsant, S., Prevette, D., Sun, W., Knudson, C. M., Milligan, C. E., Oppenheim, R. W. Complete dissociation of motor neuron death from motor dysfunction by Bax deletion in a mouse model of ALS. J Neurosci. 2006; 26: 8774-8786.
- [117] Poesen, K., Lambrechts, D., Van Damme, P., Dhondt, J., Bender, F., Frank, N., Bogaert, E., Claes, B., Heylen, L., Verheyen, A., Raes, K., Tjwa, M., Eriksson, U., Shibuya, M., Nuydens, R., Van Den Bosch, L., Meert, T., D'Hooge, R., Sendtner, M., Robberecht, W., Carmeliet, P. Novel role for vascular endothelial growth factor (VEGF) receptor-1 and its ligand VEGF-B in motor neuron degeneration. J Neurosci. 2008; 28: 10451-10459.
- [118] Turner, B. J., Parkinson, N. J., Davies, K. E., Talbot, K. Survival motor neuron deficiency enhances progression in an amyotrophic lateral sclerosis mouse model. Neurobiol Dis. 2009; 34: 511-517.
- [119] Taes, I., Goris, A., Lemmens, R., van Es, M. A., van den Berg, L. H., Chio, A., Traynor, B. J., Birve, A., Andersen, P., Slowik, A., Tomik, B., Brown, R. H., Jr., Shaw, C. E., Al-Chalabi, A., Boonen, S., Van Den Bosch, L., Dubois, B., Van Damme, P., Robberecht, W. Tau levels do

not influence human ALS or motor neuron degeneration in the SOD1G93A mouse. Neurology. 2010; 74: 1687-1693.

- [120] Crosio, C., Valle, C., Casciati, A., Iaccarino, C., Carri, M. T. Astroglial inhibition of NF-kappaB does not ameliorate disease onset and progression in a mouse model for amyotrophic lateral sclerosis (ALS). PLoS One. 2011; 6: e17187.
- [121] Finkelstein, A., Kunis, G., Seksenyan, A., Ronen, A., Berkutzki, T., Azoulay, D., Koronyo-Hamaoui, M., Schwartz, M. Abnormal changes in NKT cells, the IGF-1 axis, and liver pathology in an animal model of ALS. PLoS One. 2011; 6: e22374.
- [122] Keller, A. F., Gravel, M., Kriz, J. Treatment with minocycline after disease onset alters astrocyte reactivity and increases microgliosis in SOD1 mutant mice. Exp Neurol. 2011; 228: 69-79.
- [123] Bhattacharya, A., Bokov, A., Muller, F. L., Jernigan, A. L., Maslin, K., Diaz, V., Richardson, A., Van Remmen, H. Dietary restriction but not rapamycin extends disease onset and survival of the H46R/H48Q mouse model of ALS. Neurobiol Aging. 2012; 33: 1829-1832.
- [124] Perez-Garcia, M. J., Burden, S. J. Increasing MuSK activity delays denervation and improves motor function in ALS mice. Cell Rep. 2012; 2: 497-502.
- [125] Ringer, C., Weihe, E., Schutz, B. Calcitonin gene-related peptide expression levels predict motor neuron vulnerability in the superoxide dismutase 1-G93A mouse model of amyotrophic lateral sclerosis. Neurobiol Dis. 2012; 45: 547-554.
- [126] Petri, S., Kiaei, M., Wille, E., Calingasan, N. Y., Flint Beal, M. Loss of Fas ligand-function improves survival in G93Atransgenic ALS mice. J Neurol Sci. 2006; 251: 44-49.