

Inhibitor of RAGE and glucose-induced inflammation in bone marrow mesenchymal stem cells: Effect and mechanism of action

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Abstract. The occurrence and development of hyperglycemia-induced inflammation is associated with increased expression of receptor for advanced glycation end products (RAGE) and inflammatory factors, including IL-1 β , TNF- α and IL-6. Previous studies have reported that the nucleotide-binding oligomerization domain-like receptor protein 3 (NLRP3) inflammasome interacts with thioredoxin-interacting protein (TXNIP) and serves a crucial role in inflammation. FPS-ZM1 has been identified as target inhibitor of RAGE and has been shown to exert an anti-inflammatory effect *in vitro*. However, the underlying mechanism by which FPS-ZM1 impacts high glucose (HG)-induced inflammation in bone marrow mesenchymal stem cells (BMSCs) remains unclear. The present study explored the regulatory effect of FPS-ZM1 on HG-induced inflammation in BMSCs. Furthermore, the role of the TXNIP/NLRP3 inflammasome signaling pathway in the regulatory effects of FPS-ZM1 on HG-induced inflammation was studied. Cell viability was determined using Cell Counting Kit-8 and western blotting was used to assess the protein expression levels of RAGE. ELISA was used to determine the levels of inflammatory markers. Reverse transcription-quantitative PCR and western blotting were used to measure the mRNA and protein expression levels of TXNIP, caspase-1, thioredoxin (TRX), NLRP3 and apoptosis-related speck-like protein containing CARD (ASC). The results revealed that in BMSCs, RAGE expression was stimulated by HG, an effect which was reversed by treatment with FPS-ZM1. In addition, HG activated inflammatory factors, such as TNF- α , IL-1 β and IL-6; however, their levels were suppressed

when cells were treated with FPS-ZM1 or the TXNIP/NLRP3 pathway inhibitor, resveratrol (Res). Furthermore, FPS-ZM1 inhibited the mRNA and protein expression levels of TXNIP, caspase-1, NLRP3 and ASC, and promoted TRX expression, which was consistent with the effects of Res. These findings indicated that FPS-ZM1 may attenuate HG-induced inflammation in BMSCs. Furthermore, the TXNIP/NLRP3 inflammasome signaling pathway mediated the molecular mechanism underlying this effect.

Introduction

Diabetes mellitus is a disease associated with numerous complications (1). Hyperglycemia affects patients with diabetes by damaging macro- and microvessels, thus resulting in retinopathy, neuropathy and nephropathy, and cardiovascular, cerebrovascular and other serious complications (2,3). Although multiple mechanisms cause diabetic complications, growing evidence has suggested that immoderate aggregation of advanced glycation end products (AGEs) may be a causative factor (4). Excessive AGEs activate and interact with the receptor for AGEs (RAGE), leading to the activation of inflammatory factors (5,6). FPS-ZM1 is a specific and high-affinity inhibitor of RAGE (7). Numerous studies have reported that FPS-ZM1 exerts an anti-inflammatory effect on various cells, and on human periodontal ligament fibroblasts and human gingival fibroblasts (8-10). These findings suggested a promising application of FPS-ZM1 in preventing periodontal diseases. However, further studies are required to explore its application and importance in the medical field.

It has been reported that expression of RAGE is high in gingival fibroblasts, periodontal ligament fibroblasts and periodontal ligament stem cells in a high glucose (HG) environment (11). Under HG conditions, various inflammatory factors, including but not limited to IL-1 β , high mobility group box-1, IL-6, intercellular adhesion molecule-1 and TNF- α , can be activated through the combination of RAGE and AGEs (12,13). These inflammatory cytokines have been demonstrated to cause inflammation in periodontal tissues, including the alveolar bone and surrounding soft tissues (14,15). As observed previously, IL-6, IL-1 β and TNF- α largely accumulate in the periodontal tissues of patients with diabetes (16). Bone marrow mesenchymal stem cells (BMSCs) are present in the alveolar bone and possess multi-directional differentiation

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potential; these cells have an essential role in bone formation (17). It has previously been reported that the release of inflammatory cytokines promotes the proliferation of BMSCs in a HG environment, thus suggesting its application in the pathological process of periodontal diseases (18). Therefore, this study aimed to assess how HG activates inflammatory cytokines in BMSCs.

Several intracellular signaling pathways involved in the activation of inflammatory cytokines are currently being studied. Notably, to the best of our knowledge, the infection-associated proinflammatory function of NF- κ B is the most widely studied (19). The thioredoxin-interacting protein (TXNIP)/nucleotide-binding oligomerization domain-like receptor protein 3 (NLRP3) inflammasome pathway has been reported to be associated with diabetes-associated inflammation (20). The NLRP3 inflammasome has been well studied and is comprised of three components: NLRP3 protein, apoptosis-related speck-like protein containing CARD (ASC) and caspase-1 (21). The NLRP3 inflammasome can detect danger signals, known as danger-associated molecular patterns (22). NLRP3 agonists can activate caspase-1, which in turn releases the proinflammatory cytokines IL-18 and IL-1 β (23,24). The role of the NLRP3 inflammasome has been demonstrated in the pathogenic process of various diseases including, but not limited to, Alzheimer's disease, osteoarthritis and type 2 diabetes (25-27).

However, few studies have described the mechanism through which the NLRP3 inflammasome is activated. Functionally, TXNIP dissociates from thioredoxin (TRX), combines directly with NLRP3 and activates it (28). In addition, recent studies have demonstrated that TXNIP is upregulated in diabetes-related inflammation (20,29). In patients with diabetes, hyperglycemia has been suggested to stimulate the expression of TXNIP (30). Therefore, it may be hypothesized that HG activates the NLRP3 inflammasome by promoting TXNIP expression.

The aim of the present study was to assess the expression of RAGE in BMSCs under HG stimulation, and to investigate the expression of related inflammatory cytokines and the potential molecular mechanisms in response to a RAGE-specific inhibitor. Consequently, the feasibility of using RAGE as a target for drug therapy to prevent periodontal inflammation and promote the healing process of bone tissue injury in patients with diabetes was considered.

Materials and methods

Cell culture. BeNa Culture Collection; Beijing Beina Chunglian Biotechnology Research Institute provided the rat BMSCs. Cells were cultured under normal glucose (NG; 5 mM) or HG (25 mM) conditions in DMEM (HyClone; Cytiva) supplemented with 10% FBS (Gibco; Thermo Fisher Scientific, Inc.) and 1% penicillin-streptomycin solution (Beyotime Institute of Biotechnology). Cells were cultured in a humidified atmosphere containing 5% CO₂ at 37°C. Through the cell viability assay, an optimal concentration of FPS-ZM1 (500 nM; Beyotime Institute of Biotechnology) was selected to treat the cells, with or without HG stimulation for 48 h. The TXNIP inhibitor resveratrol (Res; 50 μ mol/l; Beyotime Institute of Biotechnology) was used to inhibit the

TXNIP/NLRP3 pathway. Cells were stimulated with Res for 2 h at 37°C in the dark and were then incubated under HG conditions for 48 h. BMSCs were cultured to 4-7 generations for subsequent experiments.

Cell viability assay. Cell viability was determined using the Cell Counting Kit (CCK)-8 assay (Dojindo Molecular Technologies, Inc.). Firstly, cells were seeded in a 96-well plate at a density of 5x10³ cells/well. The cells were grown to ~90% confluence and were then co-cultured with various concentrations of FPS-ZM1 (0, 250, 500 or 750 nM) and HG (25 mM) at 37°C. Subsequently, the viability of the cultured cells was detected after 24, 48 or 72 h. The original medium was removed and 100 μ l serum-free DMEM containing 10 μ l CCK-8 solution was added to each well. Cells were subsequently incubated for 2 h at 37°C with 5% CO₂. Finally, a Varioskan Flash microplate reader (Thermo Fisher Scientific, Inc.) was used to detect the absorbance value at 450 nm. The mean absorbance values of the control group obtained from the three different time-points were set as 100%. Experiments were conducted three times.

RNA isolation and reverse transcription-quantitative PCR (RT-qPCR). The expression of specific genes (NLRP3, ASC, caspase-1, TXNIP and TRX) were quantitatively assessed by RT-qPCR. RNA was isolated from cultured BMSCs using TRIzol[®] reagent (Invitrogen; Thermo Fisher Scientific, Inc.). Subsequently, RT of total RNA into cDNA was conducted using the Prime Script RT reagent kit (Takara Bio, Inc.), according to the manufacturer's protocol. SYBR FAST qPCR Master Mix (Kapa Biosystems; Roche Diagnostics) and a CFX-Connect 96 RT-qPCR system (Bio-Rad Laboratories, Inc.) were used to carry out the amplification of target genes. The PCR protocol was as follows: 95°C for 3 min, followed by 40 cycles at 95°C for 5 sec and 56°C for 10 sec, and a final extension step at 72°C for 25 sec. The mRNA levels of the specific genes were normalized to β -actin and expressed as a ratio to the internal reference. The primer sequences used for PCR are listed in Table I. The relative mRNA expression levels were calculated using the 2^{- $\Delta\Delta$ C_q} method and are presented as calculated values (31).

ELISA. Cell supernatants were obtained by centrifugation (1,000 x g; room temperature; 10 min) after treatment at 48 h and stored at -80°C for follow-up experiments. Activation of inflammatory factors was quantified using rat IL-1 β (cat. no. RLB00), TNF- α (cat. no. RTA00) and IL-6 (cat. no. R6000B) ELISA kits (all R&D Systems, Inc.), according to the manufacturer's protocols. Experiments were carried out three times.

Protein isolation and western blot analysis. The expression levels of specific proteins (NLRP3, ASC, caspase-1, TXNIP, TRX and RAGE) were detected by western blotting. Proteins were extracted from cultured BMSCs using RIPA buffer (Thermo Fisher Scientific, Inc.) at 4°C and total protein was boiled at 95°C for 10 min and centrifuged at room temperature at 12,000 x g for 10 min. The bicinchoninic acid protein assay kit (Beijing Solarbio Science & Technology, Co., Ltd.) was used for protein quantification. Protein samples (20 μ g) were

Table I. Primer sequences used for reverse transcription-quantitative PCR.

Gene	Primer sequence (5'-3')
NLRP3	F-CATCTTAGTCCTGCCAA
	R-CAACAGACGCTACACCC
ASC	F-AGCATCCAGCAAACCA
	R-GGACCCCATAGACCTCA
Caspase-1	F-TTGAAGAGCAGAAAGCA
	R-CAGTAGGAAACTCCGAAG
TXNIP	F-CAAGGTAAGTGTGCCG
	R-GATTCTGTGAAGGTGATGA
TRX	F-CCAACCTTTTGACCCTTT
	R-CCCTTCTTTCATTCCCTC
β -actin	F-TAGGAGCCAGGGCAGTA
	R-CGTTGACATCCGTAAAGAC

F, forward; R, reverse; NLRP3, nucleotide-binding oligomerization domain-like receptor protein 3; ASC, apoptosis-related speck-like protein containing CARD; TXNIP, thioredoxin-interacting protein; TRX, thioredoxin.

separated by 12% SDS-PAGE (Sigma-Aldrich; Merck KGaA) and were blotted onto polyvinylidene fluoride membranes (EMD Millipore). The membranes were then blocked in 5% skim milk in PBS-0.05% Tween-20 at room temperature for 1 h. The membranes were then incubated with primary antibodies against RAGE (rabbit; 1:1,000; cat. no. PAB32996), NLRP3 (rabbit; 1:2,000; cat. no. PAB37930), ASC (rabbit; 1:1,000; cat. no. PAB30696), caspase-1 (rabbit; 1:1,000; cat. no. PAB36756), TXNIP (rabbit; 1:1,000; cat. no. PAB43948), TRX (rabbit; 1:1,000; cat. no. PAB32168) and β -actin (rabbit; 1:1,000; cat. no. PAB36265) (all Bioswamp) at 4°C overnight. β -actin was used as the loading control. Subsequently, membranes were incubated with horseradish peroxidase-conjugated goat anti-rabbit antibodies (1:20,000; cat. no. SAB43714; Bioswamp) for 1 h at room temperature. An enhanced chemiluminescence kit (Analytik Jena AG) was used to measure reactivity, and the target bands were detected, and protein expression was semi-quantified using TANON GIS 4.2 software (Tanon Science & Technology Co., Ltd.). The detected protein expression levels were normalized to those of β -actin.

Statistical analysis. All data obtained from three experimental repeats are presented as the mean \pm standard deviation, and statistical analyses were conducted using SPSS 19.0 software (IBM Corp.). One-way analysis of variance was used for multiple group comparisons, and Tukey's post hoc test was used for intergroup comparisons. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

FPS-ZM1 inhibits HG-induced cell viability. To verify suitable time-points and the optimal concentration of FPS-ZM1 for follow-up experiments, BMSCs were treated with FPS-ZM1 (0, 250, 500 and 750 nM) for 24, 48 and 72 h. Alterations in

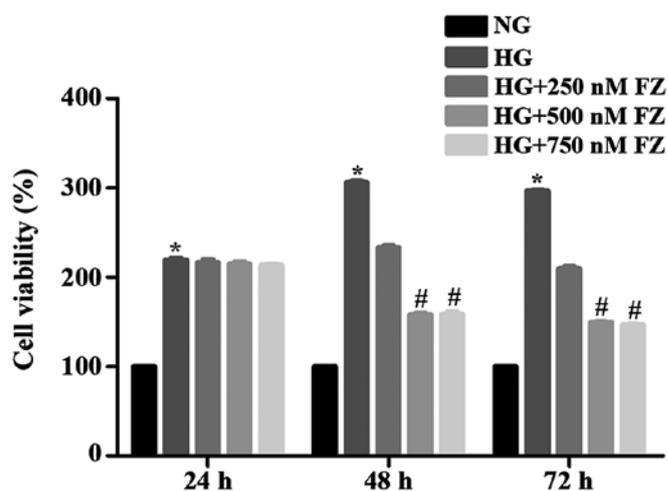


Figure 1. Effects of FZ (0, 250, 500 and 750 nM) on bone marrow mesenchymal stem cell viability at 24, 48 and 72 h. Cell Counting Kit-8 assay was performed after co-culturing cells with FPS-ZM1 and HG for 48 h. Data are presented as the mean \pm standard deviation. * $P < 0.05$ vs. the NG group; # $P < 0.05$ vs. the HG group. FZ, FPS-ZM1; HG, high glucose; NG, normal glucose.

BMSC viability are presented in Fig. 1. Compared with in the NG group, HG stimulation significantly enhanced the viability of BMSCs at all selected time points ($P < 0.05$). Notably, no significant alterations in BMSC viability were detected following treatment with 250, 500 or 750 nM FPS-ZM1 for 24 h. Conversely, treatment with 500 and 750 nM FPS-ZM1 for 48 or 72 h significantly alleviated HG-induced cell viability compared with the HG group ($P < 0.05$). No significant differences in viability were observed between cells treated with 500 or 750 nM FPS-ZM1 for 48 and 72 h ($P > 0.05$). These results indicated that HG may promote the viability of BMSCs at 48 h, but this effect may gradually decrease as the culture time increases.

FPS-ZM1 alleviates HG-induced inflammatory factor activation in BMSCs. ELISA kits were used to detect the levels of IL-6, TNF- α and IL-1 β in BMSCs under different treatments. The impact of FPS-ZM1 on HG-induced intracellular inflammation was also assessed. As presented in Fig. 2, the concentrations of TNF- α , IL-1 β and IL-6 were elevated under HG conditions compared with in the NG group ($P < 0.05$). However, FPS-ZM1 significantly reduced the effects of HG on inflammatory marker levels ($P < 0.05$). Similarly, the concentrations of TNF- α , IL-1 β and IL-6 were decreased in BMSCs after pretreatment with Res ($P < 0.05$).

FPS-ZM1 inhibits RAGE expression in HG-induced BMSCs. To explore the impact of FPS-ZM1 on RAGE expression in BMSCs under HG conditions, western blotting was used to assess the protein expression of RAGE. RAGE expression was increased in BMSCs under HG conditions, whereas treatment with FPS-ZM1 inhibited the increase in RAGE expression in HG-induced BMSCs ($P < 0.05$) (Fig. 3).

FPS-ZM1 inhibits HG-induced TXNIP/NLRP3 inflammatory activation. The present study aimed to determine whether FPS-ZM1 inhibited inflammation through the

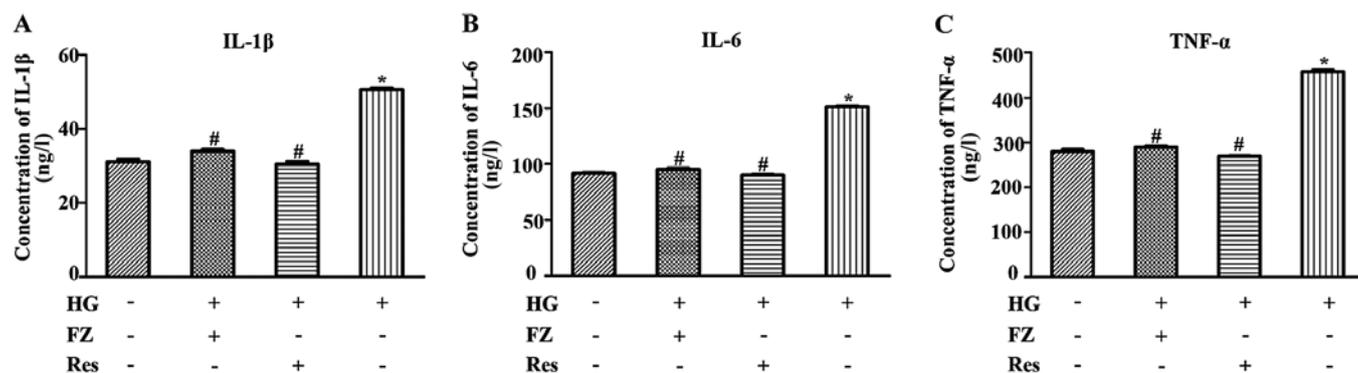


Figure 2. Effects of FZ treatment on the concentration of inflammatory factors in bone marrow mesenchymal stem cells. ELISA was performed after cells were pretreated with Res for 2 h or co-cultured with FZ and HG for 48 h. Levels of (A) IL-1 β , (B) IL-6 and (C) TNF- α were analyzed. Data are presented as the mean \pm standard deviation. *P<0.05 vs. the normal glucose group; #P<0.05 vs. the HG group. FZ, FPS-ZM1; Res, resveratrol; HG, high glucose.

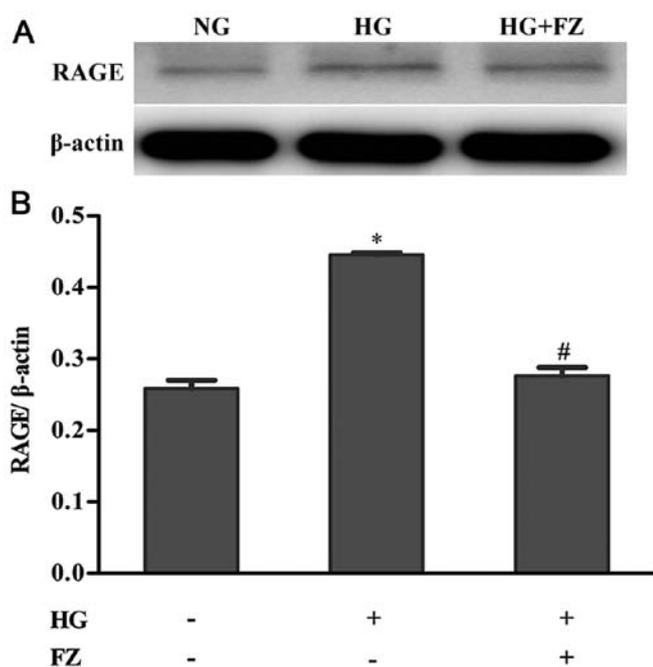


Figure 3. Effects of FZ on RAGE protein expression. Western blotting was performed after cells were co-cultured with FZ and HG for 48 h. β -actin was used as the loading control. (A) RAGE protein was measured by western blotting. (B) The RAGE/ β -actin ratio was calculated to perform densitometric analysis of band intensity. Data are presented as the mean \pm standard deviation. *P<0.05 vs. the NG group; #P<0.05 vs. the HG group. FZ, FPS-ZM1; RAGE, receptor for advanced glycation end products; NG, normal glucose; HG, high glucose.

TXNIP/TRX/NLRP3 inflammasome signaling pathway. As presented in Fig. 4, the mRNA expression of TXNIP, NLRP3, ASC and caspase-1 were increased, whereas TRX expression levels were decreased in HG-induced cells (P<0.05). Treatment with FPS-ZM1 or pretreatment with Res reduced the expression of TXNIP, caspase-1, NLRP3 and ASC, and enhanced TRX expression (P<0.05). Similar results were determined by western blotting; TXNIP, NLRP3, ASC and caspase-1 levels were increased, whereas TRX expression was decreased in HG-induced BMSCs (P<0.05). However, these effects were significantly reversed by FPS-ZM1 or Res (P<0.05) (Fig. 5). These results indicated that FPS-ZM1 may inhibit TXNIP/NLRP3 activation under a HG environment.

Discussion

AGEs have been reported to aggregate within the oral periodontal tissues of patients with diabetes, contributing to the inflammatory process in the surrounding soft and hard tissues (32,33). Inflammation may be enhanced when AGEs bind to RAGE, thus resulting in upregulation of various proinflammatory factors (34). IL-1 β is considered the most important cytokine, which serves a crucial role in the inflammatory process (35). IL-6 is a proinflammatory cytokine that is closely related to periodontitis and rheumatoid arthritis (36). TNF- α promotes T cells by secreting various activated cytokines, which are closely associated with autoimmune diseases, inflammation and diabetes (37).

RAGE is highly expressed in numerous cell types in periodontal tissues under HG stimulation, including BMSCs (18,38). It is well known that BMSCs are characterized by multidirectional differentiation and low immunity, and the potential immune-regulating effect of BMSCs has garnered increasing attention. BMSCs are functional and pivotal cells that repair damaged tissues and organs, including bone tissue, cartilage and joint injury (39). However, the differentiation of BMSCs into osteoblasts, lipoblasts or chondroblasts can be inhibited by inflammation, thus affecting the tissue repair process (40). BMSC inflammation has been reported to respond to HG; a previous study revealed that BMSCs isolated from diabetic rats exhibited stronger expression of NF- κ B and IL-18 compared with normal rats (41). The present study assessed the inflammatory response of BMSCs induced by HG *in vitro*. Subsequently, the effects of FPS-ZM1 on regulating HG-induced inflammation and the underlying potential mechanism were determined.

In the present study, the effects of HG were detected on RAGE expression in BMSCs; the results indicated that HG exposure stimulated the increased expression of RAGE in BMSCs. However, FPS-ZM1, a RAGE-specific inhibitor, reversed the enhanced expression of RAGE induced by HG. In addition, the effects of various concentrations of FPS-ZM1 (0, 250, 500 and 750 nM) for different durations on BMSCs were examined. The viability of BMSCs was enhanced under HG conditions. Conversely, FPS-ZM1 alleviated HG-induced viability. The present study identified significant differences between the NG group and the HG-treated groups regardless

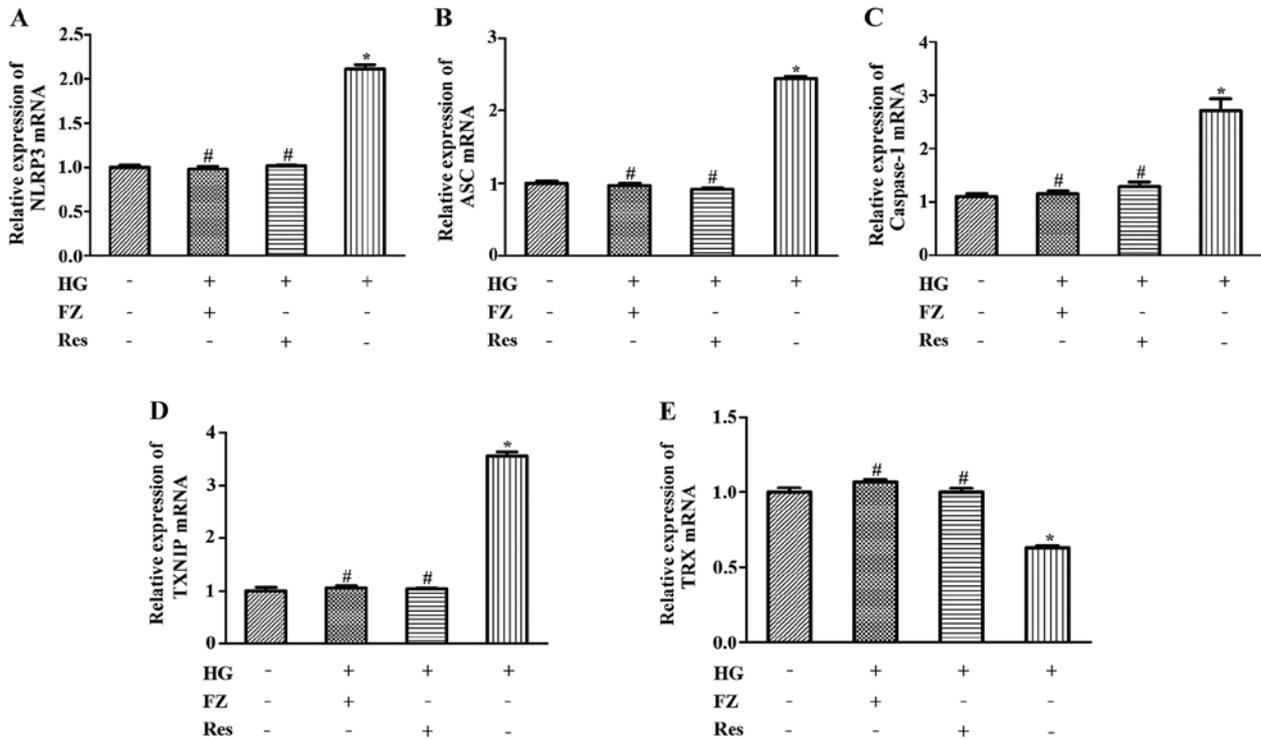


Figure 4. Effects of FZ on the mRNA expression levels of ASC, TXNIP, NLRP3, caspase-1 and TRX. Reverse transcription-quantitative PCR was performed after cells were pretreated with Res for 2 h or co-cultured with FZ and HG for 48 h. (A) NLRP3, (B) ASC, (C) caspase-1, (D) TXNIP and (E) TRX mRNA expression. Data are presented as the mean \pm standard deviation. * P <0.05 vs. the normal glucose group; # P <0.05 vs. the HG group. FZ, FPS-ZM1; ASC, apoptosis-related speck-like protein containing CARD; TXNIP, thioredoxin-interacting protein; NLRP3, nucleotide-binding oligomerization domain-like receptor protein 3; TRX, thioredoxin; HG, high glucose; Res, resveratrol.

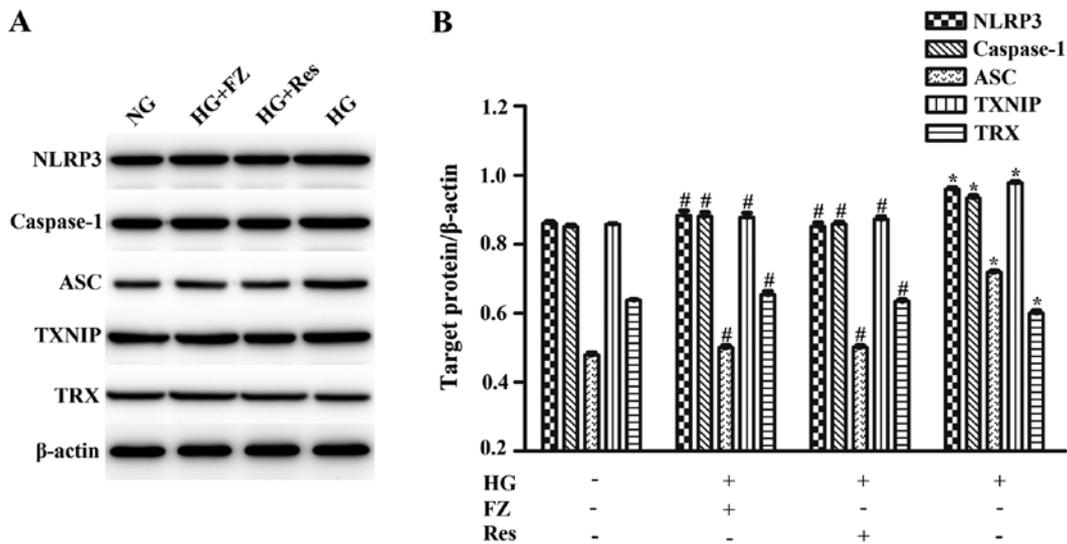


Figure 5. Effect of FZ on target protein expression levels, including ASC, TXNIP, NLRP3, caspase-1 and TRX. Western blot analysis was performed after cells were pretreated with Res for 2 h or co-cultured with FPS-ZM1 and HG for 48 h. β -actin was used as a loading control. (A) Protein expression levels of NLRP3, caspase-1, ASC, TXNIP and TRX were measured by western blotting. (B) NLRP3/ β -actin, caspase-1/ β -actin, ASC/ β -actin, TXNIP/ β -actin and TRX/ β -actin ratios were calculated to perform densitometric analysis of band intensity. Data are presented as the mean \pm standard deviation. * P <0.05 vs. the NG group. # P <0.05 vs. the HG group. FZ, FPS-ZM1; ASC, apoptosis-related speck-like protein containing CARD; TXNIP, thioredoxin-interacting protein; NLRP3, nucleotide-binding oligomerization domain-like receptor protein 3; TRX, thioredoxin; NG, normal glucose; HG, high glucose; Res, resveratrol.

of treatment duration. Cell viability of the HG group was significantly increased compared with NG group, and peaked at 48 h; however, the cell viability began to decrease from 72 h, which might be due to time-dependent HG aggravation of oxidative stress in BMSCs (42,43). It has been reported

that AGE expression can significantly inhibit the viability of cells, such as fibroblasts, in a time-dependent manner (44). AGEs may destroy stability of the internal environment by enhancing intracellular oxidative stress, thus inhibiting cell viability (45).

The present study assessed the effects of FPS-ZM1 on HG-stimulated inflammation in BMSCs. It was previously observed that HG conditions promoted inflammatory factors and inflammatory cytokines, such as TNF- α , IL-1 β and IL-6, in periodontal tissues (46,47). The present study on BMSCs presented similar findings; TNF- α , IL-1 β and IL-6 expression was enhanced after HG stimulation, whereas these effects were inhibited by FPS-ZM1. FPS-ZM1 exerted a protective effect against HG conditions *in vitro*; however, the mechanism underlying the anti-inflammatory effects of FPS-ZM1 has yet to be determined.

The present study indicated that the TXNIP/NLRP3 pathway may serve a role as the intracellular transduction pathway underlying the regulatory effects of FPS-ZM1 on HG-induced inflammation. The NLRP3 inflammasome is a protein complex that regulates inflammation and cell death (48). Previous studies have reported that NLRP3 can be overexpressed in epithelial cells or fibroblasts by glucose, oxidative stress and other types of stimulation, and it has been suggested to affect diabetes-related periodontal diseases (49,50). TXNIP has been recognized as an early mediator associated with diabetic inflammation (51). TXNIP binds to TRX and inhibits its activity; therefore, TXNIP is also called the TRX-binding protein (52). Hyperglycemia may upregulate TXNIP by directly activating the NLRP3 inflammasome, followed by an enhancement of inflammation-activated factors (53). In the present study, FPS-ZM1 downregulated HG-induced expression of TXNIP and NLRP3. Furthermore, the downregulation of TRX expression stimulated by HG was reversed by FPS-ZM1. These findings confirmed the role of the TXNIP/NLRP3 inflammasome signaling pathway in the regulatory effects of FPS-ZM1 on HG-induced BMSC. This pathway was also inactivated by Res.

A previous study reported that Res significantly inhibited the activity of the TXNIP/NLRP3 inflammasome pathway to achieve the pharmacological effect of inhibiting inflammation (54). In the present study, pretreatment with Res disrupted the activity of the TXNIP/NLRP3 inflammasome pathway. Furthermore, Res downregulated HG-induced expression of TNF- α , IL-1 β and IL-6. These findings further support the hypothesis that the TXNIP/NLRP3 inflammasome signaling pathway is involved in HG-stimulated inflammation. Therefore, inhibition of the TXNIP/NLRP3 inflammasome signaling pathway may be considered one of the mechanisms underlying the anti-inflammatory effects of FPS-ZM1.

The present study also revealed that although the mRNA and protein expression levels of inflammation-associated molecules, such as NLRP3, caspase-1, ASC and TXNIP, were higher in the HG group than in the NG group, there was a marked difference between mRNA and protein expression levels. There exists a linear correlation between mRNA and protein expression, and the change ratio should be the same; however, the results of other experimental studies have demonstrated that the linear correlation is not very high (55,56). This phenomenon has been reported by numerous studies and may be explained as follows: Gene expression is regulated in numerous ways, not just by transcriptional regulation (e.g., histone modification, DNA methylation and transcription factor regulation), but also by post-transcriptional alterations, such as microRNA targeting,

RNA-binding proteins or RNA modification (57-59). In addition, various factors, including mRNA degradation, protein degradation and protein folding may lead to differences between mRNA and protein expression levels (60,61). How the reasons for the differences observed between the mRNA and protein expression remain unknown and should be further investigated in the future.

In conclusion, the regulatory effects of FPS-ZM1 on HG-induced inflammation and NLRP3 accumulation in BMSCs were investigated. HG stimulated RAGE expression in BMSCs and mediated a series of inflammatory responses, whereas treatment with FPS-ZM1, a RAGE-specific inhibitor, protected BMSCs by exerting anti-inflammatory effects, specifically reducing HG-induced cell viability, inflammatory factor production and TXNIP/NLRP3 inflammasome signaling pathway activation. In addition, inactivation of the TXNIP/NLRP3 inflammasome by Res exhibited similar effects as FPS-ZM1, which indicated that the molecular mechanism underlying the anti-inflammatory effect of FPS-ZM1 may be associated with the TXNIP/NLRP3 inflammasome signaling pathway. Based on the anti-inflammatory effects, the results of the present study provided a novel therapeutic approach and target drug therapy for patients with diabetes with accelerated occurrence and development of periodontal diseases. Nonetheless, these studies were limited to *in vitro* experiments on BMSCs. Further studies regarding the role of FPS-ZM1 on peri-implantitis or periodontal tissues in animal models are required, as the human microenvironment is more complicated, and an assessment of the response *in vivo* is necessary.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

MJ and LG mainly contributed to experimental conception and design. MJ, XW and PW performed the experiments and collected the data. MJ, WP and BZ conducted the statistical analysis. MJ wrote the original draft. XW, PW and LG revised the completed draft. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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