

Molecular insights into probiotic mechanisms of action employed against intestinal pathogenic bacteria

Winschau F. van Zyl, Shelly M. Deane, and Leon M.T. Dicks

Department of Microbiology, Stellenbosch University, Stellenbosch, South Africa

ABSTRACT

Gastrointestinal (GI) diseases, and in particular those caused by bacterial infections, are a major cause of morbidity and mortality worldwide. Treatment is becoming increasingly difficult due to the increase in number of species that have developed resistance to antibiotics. Probiotic lactic acid bacteria (LAB) have considerable potential as alternatives to antibiotics, both in prophylactic and therapeutic applications. Several studies have documented a reduction, or prevention, of GI diseases by probiotic bacteria. Since the activities of probiotic bacteria are closely linked with conditions in the host's GI-tract (GIT) and changes in the population of enteric microorganisms, a deeper understanding of gut-microbial interactions is required in the selection of the most suitable probiotic. This necessitates a deeper understanding of the molecular capabilities of probiotic bacteria. In this review, we explore how probiotic microorganisms interact with enteric pathogens in the GIT. The significance of probiotic colonization and persistence in the GIT is also addressed.

ARTICLE HISTORY

Received 1 July 2020
Revised 2 September 2020
Accepted 8 September 2020


Keywords

Probiotics; lactic acid bacteria; colonization; gastrointestinal tract; enteric pathogens; competitive exclusion; antimicrobial compounds; bacteriocins

Introduction

Lactic acid bacteria (LAB) play a major role in the preservation and organoleptic profile of fermented food products, but are equally important in affecting the composition and diversity of intestinal microbiota.^{1–3} Some of the most important beneficial effects include stimulation of the host's immune system, prevention of antibiotic-associated diarrhea, treatment of inflammatory bowel disease (IBD) and irritable bowel syndrome (IBS), alleviation of lactose intolerance, lowering of cholesterol levels and prevention of life-threatening GI infections such as *Clostridium difficile*-associated diarrhea.^{4–8} Renewed interest in probiotics initiated the launch of an increasing number of probiotic-containing supplements that claim to confer specific health benefits to the consumer.^{9,10} Many of these products are driven by aggressive marketing through pharmaceutical and nutritional companies, often without a clear understanding of the interactions between probiotic bacteria, normal commensal microorganisms, pathogens and the host.

Lactic acid bacteria are indigenous to the small and large intestine of humans and animals and exert a number of probiotic properties, such as binding to receptors and physically excluding pathogens, production of antimicrobial substances, strengthening of the gut mucosal barrier and modulation of the immune system.^{11–15} It is therefore important to have an in-depth understanding of the specific metabolic and genetic interactions between probiotic bacteria, the host intestinal mucosa and enteric pathogens in the GIT. Commensal bacteria also act as a protective barrier against pathogens by providing mucosal protection and stimulation of the immune system.^{10,15} The most predominant genera used in probiotic supplements are *Lactococcus*, *Lactobacillus* and *Bifidobacterium* spp. derived from humans and animals.¹¹ It is important that probiotic strains survive passage through the stomach, resist bile salts and digestive enzymes in the small intestinal tract and reach the colon in sufficient numbers.¹⁶ The number of viable cells surviving the journey through the GIT is, however, strain specific and depends on the dosage and duration of administration.^{10,16}

CONTACT Leon M.T. Dicks;  LMTD@sun.ac.za  Department of Microbiology; Stellenbosch University, Stellenbosch 7602, South Africa

Abbreviations: GI, gastrointestinal; GIT, gastrointestinal tract; LAB, lactic acid bacteria; IBD, inflammatory bowel disease; IBS, irritable bowel syndrome; IECs, intestinal epithelial cells; EHEC, enterohemorrhagic *E. coli*; Msa, mannose-specific adhesion protein; S-layer, surface layer; CWSS, cell wall sorting signal; VRE, vancomycin-resistant enterococci; EPEC, enteropathogenic *E. coli*; BLIS, bacteriocin-like inhibitory substances; LMW, low molecular weight; ETEC, enterotoxigenic *E. coli*; CFA, colonization factor antigen; DCs; dendritic cells; MAPK, mitogen-activated protein kinases; NF- κ B, nuclear factor kappa B; IgA, immunoglobulin A.

© 2020 The Author(s). Published with license by Taylor & Francis Group, LLC.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Due to the complex nature of the human GIT, only a few in-depth studies on interactions between probiotic bacteria and enteric pathogens have been published and many rely on *in vitro* data to decipher the mechanistic basis underlying a specific health benefit.¹⁷ A specific health benefit may also be attributed to a combination of mechanisms. This is important, as strains from the same species may evoke different responses in the host.¹³ Several probiotic strains secrete secondary metabolites and peptides with antimicrobial activity that may interact directly with the host or pathogens.^{15,18} Ultimately, a detailed characterization of these interactions will significantly improve the application of probiotics to support and enhance human health. In this review, the mechanisms probiotic bacteria use to interact with enteric pathogens, and the ability to colonize the GIT, are discussed.

Colonization and persistence of probiotic bacteria in the GIT

Strains that colonize the GIT have a greater beneficial effect on the host than strains passing through the GIT. This may be because adhesion to mucus and epithelial cells not only provides the strain with a competitive advantage, but forms a stronger interaction with the host that leads to recognition of the probiotic and stimulation of the host's immune response.^{15,17} Furthermore, colonization by probiotic strains prevents adhesion of enteric pathogens to intestinal cells.²² Several studies have shown how probiotics interfere with the ability of GI pathogens, such as *Salmonella typhimurium*, *Clostridium sporogenes* and *Enterococcus faecalis*, to adhere to Caco-2 cells.^{23–25} A diverse spectrum of pathogens targeted by probiotic bacteria and their reported health promoting effects is listed in Table 1.

Table 1. Health effects of probiotic bacteria and main pathogens targeted.

Probiotic strains	Pathogen(s) ^a	Reported effects
<i>Lactobacillus rhamnosus</i> GG ^{4,26,27}	<i>Helicobacter pylori</i> , <i>rotavirus</i> , <i>C. difficile</i>	Reduced diarrhea and nausea in a human trial. Immune enhancement. Used for alleviation of atopic dermatitis in children, stabilization of intestinal permeability
<i>L. johnsonii</i> La1 ²⁸	<i>H. pylori</i>	Regular ingestion modulated <i>H. pylori</i> colonization in children
<i>L. casei</i> DG ²⁹	<i>H. pylori</i>	Increased eradication rate of <i>H. pylori</i> infection when supplemented with first-line therapies
<i>L. casei</i> CRL431 ³⁰	<i>Salmonella enterica</i> serovar Typhimurium	Preventative administration protected mice against infection
<i>L. rhamnosus</i> HN001 ³¹	<i>Salmonella enterica</i> serovar Typhimurium	Conferred immune enhancement and protection against <i>Salmonella</i> infection in mice
<i>Bifidobacterium longum</i> Bb46 ³²	<i>Salmonella enterica</i> serovar Typhimurium	Protective effect against <i>Salmonella</i> challenge in gnotobiotic mice
<i>L. plantarum</i> 423 and <i>Enterococcus mundtii</i> ST4SA ³³	<i>Salmonella enterica</i> serovar Typhimurium	Alleviated symptoms of <i>Salmonella</i> infections in challenge study using rats
<i>L. casei</i> BL23 and <i>L. paracasei</i> CNCM I-3689 ³⁴	<i>Listeria monocytogenes</i>	Decreased pathogen systemic dissemination in orally infected mice
<i>L. salivarius</i> UC118 ³⁵	<i>L. monocytogenes</i>	Protected mice from pathogenic infection in liver and spleen
<i>L. plantarum</i> 423 and <i>E. mundtii</i> ST4SA ^{36,37}	<i>L. monocytogenes</i>	Excluded the pathogen from the intestinal tract of mice after daily administrations of probiotic strains
<i>Lactococcus lactis</i> MM19 and <i>Pediocin acidilactici</i> MM33 ³⁸	Vancomycin resistant enterococci (VRE)	Modulated intestinal microbiota and reduced pathogen intestinal colonization in mice.
<i>L. rhamnosus</i> R0011 and <i>L. acidophilus</i> R0052 ³⁹	<i>Citrobacter rodentium</i>	Pre-treatment with the probiotic strains attenuated pathogen infection in mice
<i>L. reuteri</i> ⁴⁰	<i>C. rodentium</i>	Attenuated <i>C. rodentium</i> -induced colitis in mice. Significantly decreased diarrhea symptoms in infants and children.
<i>B. breve</i> ⁴¹	<i>Escherichia coli</i> O157:H7	Protected mice from Shiga toxic-producing <i>E. coli</i> .
<i>Pediococcus pentasaceus</i> NB-17 ⁴²	n/a	Effectively stimulated immune cell activities and allergic inhibitory effects
<i>Oenococcus oeni</i> 9115 ⁴³	n/a	Significantly decreased acid-induced colitis in mice. Modulated the immune response of immunocompetent cells <i>in vitro</i> .
<i>B. infantis</i> UCC 36524 ^{4,10,42,43}	<i>Clostridium</i>	Reduced clostridia levels and increased lactobacilli and bifidobacteria. Increased blood phagocytic activity. Reduced inflammation in mice.
<i>Saccharomyces boulardii</i> ^{44,45}	<i>C. difficile</i>	Used for prevention and treatment of antibiotics associated and acute diarrhea in children, treatment of <i>C. difficile</i> colitis, prevention of diarrhea in critically ill tube-fed patients
<i>B. adolescentis</i> ⁴⁶	<i>Bacteroides thetaiotaomicron</i>	Significantly modulated both systemic and intestinal immune response in germ-free rats.
<i>L. acidophilus</i> ⁴⁷	n/a	Reduced the severity of Irritable Bowel Syndrome.

^aPathogen (s): n/a, not applicable

Earlier studies on the colonization of probiotics were based on *in vitro* studies demonstrating the ability of strains to adhere to cell lines such as Caco-2, HT-29 and HT29-MTX.^{48,49} Although these studies simulated GIT-models and have provided valuable insights into the adherence of probiotic cells, it remains an *in vitro* approach that is unable to recapitulate the complex multicellular nature of the GIT. Studies using cell lines require specialized equipment and facilities to keep the cells viable. Because of these reasons, studies on the survival and colonization of probiotic bacteria are mostly done by analyzing fecal samples.⁵⁰ From the recovery of cells in feces after probiotic intervention, the persistence of strains is calculated, providing that cell numbers in the dosage are known and all methods are standardized. Probiotic cells that persist in feces for the longest time and highest numbers indicate a higher colonization and persistence in the GIT. *In vivo* pharmacokinetics of probiotics can be studied by comparing cell numbers (in fecal material) between specific strains before and after ingestion.^{51,52} Other techniques used include intestinal intubation and pyxigraphy.⁵² Antibiotic resistance markers can be used to clearly identify probiotic cells in fecal samples.^{36,52} When no strain identification is used, results may be difficult to interpret, since endogenous probiotic cells can also be excreted in feces. The pharmacokinetics of different probiotic LAB using fecal recuperation are listed in Table 2. The survival and persistence of ingested probiotics differs greatly between genera and even between strains. *Lactobacillus* and *Bifidobacterium* spp. have been extensively explored as probiotics, since they form an integral part of the natural gut microbiome of humans and animals.^{11,64} *Bifidobacterium lactis* LAFTI B94, *B. longum* SB T2928, *Lactobacillus rhamnosus* DR20, *Lactobacillus gasseri* SBT2055 and *Enterococcus mundtii* ST4SA persisted in high numbers for the longest time Table 2. In comparison, fecal recuperation of *Lactococcus lactis* MG 1363 and *Lactobacillus fermentum* KLD was much lower. In a recent study, the probiotic strains *Lactobacillus plantarum* 423 and *E. mundtii* ST4SA were transformed with a plasmid containing the bioluminescence firefly luciferase gene (*fluc*) from *Photinus pyralis*.⁵⁹ This allowed monitoring of the migration of the strains through the GIT and

in mouse feces in real time and in a noninvasive manner. With the use of bioluminescent imaging (BLI), the authors detected cell numbers as low as 10^4 CFU/100 mg feces. Imaging revealed that *E. mundtii* ST4SA persisted in feces throughout the trial period (>20 days), whilst *L. plantarum* 423 persisted for 13 days after the last day (day 5) of intragastric administration.⁵⁹ BLI provides three-dimensional images of cells as they migrate through the GIT (65). Information is gathered in real-time, using an *in vivo* imaging system (IVIS). Only metabolically active cells are detected. The technique has been used in several studies.^{36,37,59,65,66} Van Zyl et al.⁵⁹ used BLI to study the transit of *L. plantarum* 423 and *E. mundtii* ST4SA in the digestive tract of mice for 9 consecutive days. Data generated using the technique correlated with viable cell counts. For a review on the application of optical imaging systems in *in vivo* tracking of LAB, the reader is referred to ref. 66.

Some reports have suggested that non-viable and non-colonizing probiotics may also confer certain health benefits to the host.^{11,18,67–69} During GI passage, non-colonizing, or transiently colonizing probiotic bacteria continue to be metabolically active, thus conferring beneficial health effects to their host.¹⁶ In a study by Kullen et al.⁷⁰ human

Table 2. Pharmacokinetics of probiotic strains measured using fecal recuperation.

Strain	Dosage	Fecal recuperation	Persistence (day)
<i>B. lactis</i> LAFTI B94 ⁵³	1×10^{11} CFU for 7 days	1.8×10^9 CFU/g	28
<i>B. lactis</i> Bb12 ⁵⁴	1×10^{11} CFU	8×10^7 CFU/g	14
<i>B. longum</i> SB T2928 ⁵⁵	7×10^{11} CFU for 7 days	1×10^9 CFU/g	>30
<i>L. rhamnosus</i> GG ⁵⁶	6×10^{10} CFU for 12 days	4×10^4 CFU/g	14
<i>L. rhamnosus</i> DR20 ⁵⁷	1.6×10^9 CFU for ± 182 days	6.3×10^5 CFU/g	60
<i>L. salivarius</i> UCC118 ⁵⁸	1×10^{10} CFU for 21 days	$1 \times 10^3 - 1 \times 10^7$ CFU/g	>21
<i>L. plantarum</i> 423 ⁵⁹	4×10^9 CFU for 5 days	1×10^5 CFU/g	18 days
<i>L. plantarum</i> 299 v ⁶⁰	2×10^{10} CFU for 21 days	1×10^7 CFU/g	>8
<i>L. plantarum</i> NCIMB 8826 ⁵¹	5×10^{10} CFU for 7 days	1×10^8 CFU/g	14
<i>L. fermentum</i> KLD ⁵¹	1.5×10^9 CFU	3.2×10^4 CFU/g	1
<i>L. gasseri</i> SBT2055 ⁶¹	1×10^{11} CFU for 7 days	1×10^7 CFU/g	>31
<i>Enterococcus mundtii</i> ST4SA ⁵⁹	4×10^9 CFU for 5 days	1×10^6 CFU/100 mg	>20
<i>S. thermophilus</i> ⁶²	1.2×10^{12} CFU	5×10^6 CFU/g	6
<i>Lc. lactis</i> MG 1363 ⁶³	1×10^{11} CFU for 4 days	1×10^4 CFU/g	3

volunteers were administered a probiotic strain of *Bifidobacterium* and the recovery of the strain in feces was monitored. The strain was detected in feces at increasing cell numbers during days (8 days) of administration, but could not be recovered in fecal material after the last oral administration. The authors concluded that although the administered strain did not colonize the human GIT, colonization and prolonged persistence may not be required to achieve a significant probiotic effect. Similar results were reported by Fujiwara and coworkers.^{71,72} The authors found that bifidobacteria produce a 100 kDa protein, which actively prevents the adherence of pathogenic *Escherichia coli* to intestinal mucosal cells by blocking their binding to the glycolipid binding receptor gangliotetraosylceramide. Therefore, the competitive exclusion of the pathogenic strain may not have been related to direct live cell-to-cell competition for intestinal adhesion sites.

Microorganisms found in fecal samples are usually inhabitants of the lower intestine, such as the colon.¹⁰ In humans, intubation at specific intestinal sites is used to determine probiotic colonization in the upper sections of the GIT. Biopsies can be taken of the portions of the intestinal tract

where probiotics are likely to colonize, proliferate and produce their metabolites.^{51,69,73,74} *Lactobacillus rhamnosus* GG is one of the best studied probiotic strains and plays a role in the prevention or treatment of antibiotic-associated diarrhea, flatulence, rotavirus gastro-enteritis, and stomach and abdominal pain.^{26,27,71,72} However, when *L. rhamnosus* GG in fermented milk was administered to human volunteers, the strain showed only limited persistence in feces and could not be recovered in 67% of the subjects after 7 days of the last dosage.⁷⁵ The same results were obtained when a milk formula containing the strain was fed to premature infants.⁷⁶ However, Alander et al.⁷⁴ did recover *L. rhamnosus* GG from colonic biopsies for lengthy periods after administration ceased. Human volunteers were administered with 6×10^{10} CFU of *L. rhamnosus* GG twice a day for 12 consecutive days. Cell numbers of *L. rhamnosus* GG in the feces decreased with time after the last bacterial dosage was administered. No cells of strain GG were detected in feces 14 days after the last dosage. However, *L. rhamnosus* GG persisted in biopsies taken from the colonic mucosa for up to 21 days at 7×10^4 CFU/biopsy sample after consumption ceased.⁷⁴ Concluded from these studies,

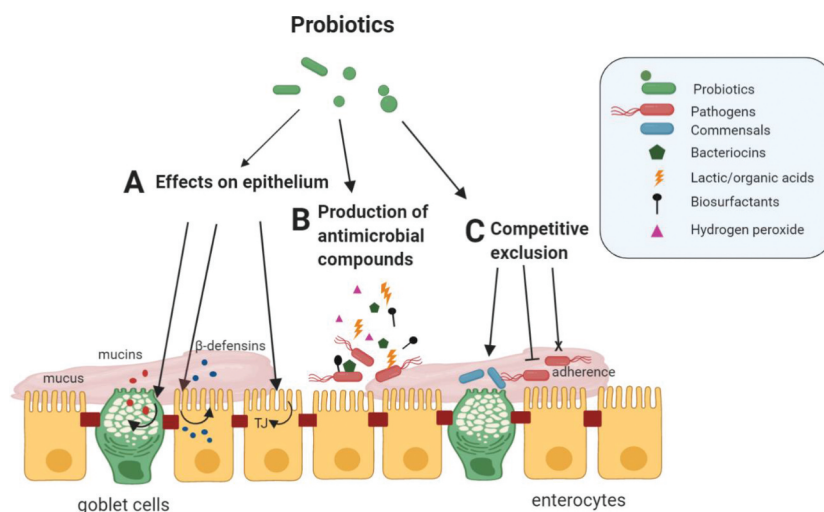


Figure 1. Probiotic mechanisms of action against enteric pathogens in the GIT. Probiotics can affect epithelial barrier integrity by numerous mechanisms. These include: **A.** direct effects on the intestinal epithelial cells (IECs). Probiotics can increase the secretion of mucin glycoproteins by goblet cells that assemble into a thick mucus layer. Probiotics can augment the secretion of antimicrobial proteins (defensins) by IECs that help to eliminate commensals or pathogens that penetrate the mucus layer. Probiotics can enhance the stability of intercellular junctional complexes (tight junctions (TJ)), which decreases the intercellular permeability of IECs to pathogens and other antigens. **B.** Most probiotics can inhibit enteric pathogens via the production of antimicrobial substances such as bacteriocins. **C.** Probiotics can compete with commensals and enteric pathogens for adhesion sites in the mucus layer or IECs, thereby preventing harmful colonization and enhancing barrier function. Probiotics can alter the natural gut microbiota composition and/or gene expression, enhancing barrier integrity through the commensal microbiota. Figure created in biorender (<http://biorender.io>).

fecal cell counts are not a true reflection of the number of viable cells in the GIT of humans and should be accompanied with intestinal biopsies at sites of colonization.

Competitive exclusion of enteric pathogens

The term competitive exclusion was first used by Greenberg (1969) to describe the exclusion of *S. typhimurium* from blowfly maggots.⁷⁷ This anti-pathogenic mechanism describes the scenario in which one bacterial species rigorously competes for adhesion to receptors in the GIT. The mechanisms of action used by one bacterial species to exclude another from the GIT differ and may include microbe-microbe interactions mediated by binding to the host mucosal interface at specific attachment sites, the secretion of antimicrobial substances and competition for available nutrients.^{14,15,17,18}

For enteropathogens to initiate infection, they have to cross the intestinal mucosal barrier before colonizing the GIT.⁷⁸ Once pathogens have penetrated the mucus layer overlying the intestinal epithelium, they attach to binding sites on epithelial cells.⁷⁹ Attachment is followed by intestinal colonization and infection.⁸⁰ Probiotics with adhesion capabilities protect the gut against enteric infections by preventing the attachment of pathogens **Figure 1**. Results from *in vitro* studies using human or animal mucosal material have demonstrated the effect of probiotic LAB on the competitive exclusion of pathogens.^{25,48,81–83} *Lactobacillus*

rhamnosus GG has excellent adhesion properties and prevented the internalization of enterohemorrhagic *E. coli* (EHEC) in human intestinal cell lines.⁸¹ Enteric pathogens, such as EHEC, use mannose-sensitive type 1 fimbriae to attach to oligosaccharide residues of glycoproteins or glycolipids on the surface of intestinal epithelial cells (IECs).⁸⁴ Probiotic strains of lactobacilli and bifidobacteria attach to the same receptor sites and exclude pathogens from binding to the GIT.^{85,86} Some probiotic strains have specific adhesion proteins on their cell surface that bind to carbohydrate moieties in the mucous layer, such as the mannose-specific adherence mechanism of *L. plantarum*.^{13,87,88} In some cases, competitive exclusion may be as simple as steric hindrance.⁸⁸ An overview of studies that analyzed the effect of probiotic LAB surface proteins on adhesion and competitive exclusion using mutant analysis is provided in **Table 3**. One example of a specific adhesion protein involved in competitive exclusion adhesion-receptor interactions in the GIT is the *L. plantarum* mannose-specific adhesion (Msa) protein.⁸⁷ A spontaneously mutated strain of the probiotic *L. plantarum* 299 v, thought to be affected in the *msa* gene, was unable to inhibit the attachment of EHEC to HT-29 epithelial cells compared to the wild-type.⁹¹ This suggested that Msa-containing probiotic strains could effectively exclude several other, if not all, type 1 fimbriated enteropathogens. Recently, Van Zyl et al.³⁷ demonstrated the involvement of the mucus-adhesion protein (*mapA*) of *L. plantarum* 423 in competitive exclusion of *Listeria monocytogenes* EGDe *in vivo*,

Table 3. Predicted function and mutant phenotypes of probiotic LAB cell surface adhesion genes.

Strain	Gene	Predicted function	Mutant phenotype
<i>L. plantarum</i> WCFS1 ⁸⁹	<i>srtA</i>	Sortase	Reduced mannose-specific binding; competitive ability in murine GIT not affected
<i>L. plantarum</i> WCFS1 ⁸⁹	<i>msa</i>	Mannose-specific adhesin	Reduced mannose-specific binding
<i>L. plantarum</i> WCFS1 ⁹⁰	<i>lp_2940</i>	Sortase-dependent cell wall protein	Reduced persistence in murine GIT
<i>L. plantarum</i> 299 v ⁹¹	<i>msa</i>	Mannose-specific adhesin	Reduced capability to prevent adherence of EHEC to HT-29 cells
<i>L. plantarum</i> 423 ³⁷	<i>mapA</i>	Mucus – adhesion protein (MapA)	Reduced capability to exclude <i>Listeria monocytogenes</i> EGDe from the GIT of mice
<i>L. acidophilus</i> NCFM ⁹²	<i>mub</i>	Mucus-binding protein (MUB)	Reduced binding to human Caco-2 cells
<i>L. acidophilus</i> NCFM ⁹²	<i>slpA</i>	S-layer protein	Reduced binding to human Caco-2 cells
<i>L. salivarius</i> UCC18 ⁹³	<i>srtA</i>	Sortase	Reduced binding to human Caco-2 and HT-29 cells
<i>L. salivarius</i> UCC18 ⁹³	<i>lspA</i>	Large surface protein (LSP), putative MUB	Reduced binding to human Caco-2 and HT-29 cells
<i>L. salivarius</i> UCC18 ⁹³	<i>lspB</i>	LSP, putative MUB	Binding to human Caco-2 and HT-29 cells not affected
<i>L. reuteri</i> 100–23 ⁹⁴	<i>lsp</i>	LSP	Reduced persistence in murine GIT
<i>L. johnsonii</i> NCC533 ²¹	<i>LJ1476</i>	Transpeptidase Sortase	Colonization dynamics similar to that of wild-type
<i>E. mundtii</i> ST4SA ³⁷	<i>srtA</i>	Sortase-dependent cell wall protein	Reduced capability to exclude <i>L. monocytogenes</i> EGDe from the GIT of mice
<i>E. mundtii</i> ST4SA ³⁷	<i>srtC</i>	Sortase-dependent cell wall protein	Reduced capability to exclude <i>L. monocytogenes</i> EGDe from the GIT of mice

using gene knockout analysis and BLI. The *mapA* negative mutant strain of *L. plantarum* 423 was unable to exclude *L. monocytogenes* EGDe.³⁷

Another example of a putative competitive exclusion factor is the collagen-binding protein of *L. fermentum*. Heinemann et al.⁹⁵ characterized the collagen surface-binding protein of *L. fermentum* RC-14, which inhibited the adherence of *E. faecalis* 1131. Other studies have demonstrated the role of surface layer (S-layer) extracts in the prevention of pathogens from attaching to, and thus colonizing, IECs.^{96,97} Chen et al.⁹⁶ showed that S-layer proteins anchored on the cell surface of *Lactobacillus crispatus* ZJ001 were responsible for competitive exclusion of *S. typhimurium* and EHEC. Similar results were recorded by Johnson-Henry et al.⁹⁷ The authors showed that S-layer protein extracts from *Lactobacillus helveticus* R0052 inhibited the adhesion of *E. coli* O157:H7 to Caco-2 cells. S-layer proteins are highly hydrophobic and it was suggested that pathogen adherence inhibition was mediated by hydrophobic group interactions as opposed to adhesion-receptor interactions.

Previous studies suggested that sortase-dependent cell surface proteins (SDPs) play a crucial role in probiotic-host interactions, adherence and colonization.^{98–102} Several SDPs have been identified with a role in *in vitro* and *in vivo* adhesion to intestinal cells, including mucus-binding cell surface proteins Table 3. In Gram-positive bacteria, sortases decorate the cell surface with a diverse array of proteins by covalently joining them to the cell wall (Sortase A) or by polymerizing proteins to construct complex multi-subunit pilin structures (Sortase C) on the cell surface.⁹⁹ Sortases are characterized as cysteine transpeptidases that join SDPs containing a specific cell wall sorting signal (CWSS) to an amino group located on the cell surface.¹⁰⁰ Sortase A enzymes anchor proteins that contain a CWSS with a LPXTG (where X donates any amino acid) C-terminal motif to the cell surface.⁹⁹ The LPXTG motif is recognized by the SrtA enzyme, which breaks the threonine and glycine peptide bond and then covalently links the threonine residue to the amino group of the pentaglycine bacterial cell wall cross bridge.^{93,101} Sortase C proteins catalyze a similar transpeptidation reaction, but recognize a QVPTGV sorting motif to construct pili that

promote microbial adhesion.¹⁰² Using mutant analysis coupled with *in vivo* BLI, a recent study showed that *E. mundtii* ST4SA sortase mutants (*srtA* and *srtC*) had a reduced ability to exclude *L. monocytogenes* EGDe from the GIT of mice compared to the wild-type derivative.³⁷

Several strains of lactobacilli and bifidobacteria inhibit, displace, and adhere to the same enterocyte layer as, enteropathogenic *Salmonella choleraesuis* serovar Typhimurium.¹⁰³ This indicates that the probiotic strains have the ability to effectively displace the pathogen after pathogenic colonization of the gut has occurred instead of being effective only when administered in a preventative manner. To gain a competitive advantage, the probiotics can thus modify the gut environment by producing inhibitory compounds, lowering pH levels and competing for nutrients.⁶⁴ *Lactobacillus* species such as *L. acidophilus* and *L. plantarum* have the ability to utilize complex carbohydrates such as fructans.^{104,105} Similarly, bifidobacteria are capable of metabolizing various plant dietary fibers using several depolymerizing enzymes.¹⁰⁵ Utilizing carbohydrate sources other than those used by enteropathogenic bacteria enable probiotic bacteria to widen their areas of colonization in the GIT and inhibit pathogens.

Production of antimicrobial compounds

Antimicrobial compounds, produced by probiotic bacteria, can exert direct antimicrobial action toward competing enteropathogens that may lead to the prevention of pathogenic colonization of the GIT Figure 1b.

Bacteriocins

Bacteriocins are ribosomally produced antimicrobial peptides that differ in terms of their size (2–10 kDa) and mechanisms of action (for a review, see ref. 108). The production of bacteriocins by probiotic bacteria (usually LAB) is a key mechanism of action used to inhibit pathogens in the GIT. Bacteriocins usually only inhibit specific species, often those closely related to the producer.^{106,107} Some bacteriocins are reported to have a much broader spectrum of antimicrobial activity.^{107–111} Bacteriocins such as nisin, produced by *Lc. lactis*,

plantaricin from *L. plantarum* and lacticin B from *L. acidophilus* are active against food-borne enteropathogens such as *Listeria*, *Clostridium*, *Bacillus*, methicillin-resistant *Staphylococcus aureus* and vancomycin-resistant enterococci (VRE).^{14–119} Bacteriocins may have a bacteriostatic or direct bactericidal effect on pathogens, thus limiting the ability of the cells to colonize the gut. The associated antimicrobial activities of bacteriocins allow bacteriocin-producing probiotic strains to gain a competitive advantage within the complex GI environment.¹²⁰

Bacteriocins adhere to microbial cells and penetrate phospholipid membranes due to their small size and variations in hydrophobic and hydrophilic properties.¹²¹ The general mechanisms of bacteriocin-mediated pathogen killing include the induction of cytoplasmic membrane permeabilization of sensitive bacteria that leads to cell leakages, inhibition of DNA and RNA synthesis and/or cell wall protein-synthesis.^{117,122,123} For instance, nisin acts by forming a complex with the cell membrane lipid II precursor, followed by the aggregation and incorporation of peptides to form discrete pores in the bacterial cell membrane.¹²⁴ A unique bacteriocin, bifidocin B, produced by *Bifidobacterium bifidum* NCFB, is active against several Gram-positive bacteria, including *Listeria*, *Enterococcus*, *Bacillus* and *Lactobacillus*, but shows no activity toward several other Gram-positive and Gram-negative bacteria.¹²⁵ The difference in activity between strains is related to the ability of Gram-negative bacteria to resist adsorption of bifidocin B, due to their cell wall composition.¹²⁶ However, several bacteriocins, such as mutacins (A–D), nisins (A and Z), lacticins (A164 and BH5), bacteriocins E 50–52 and OR7 are active against medically important Gram-negative organisms such as *Campylobacter*, *Helicobacter*, *Haemophilus*, *Neisseria* and *Salmonella* spp.^{127–132} In another study, a bacteriocin produced by *Bacillus amyloliquefaciens* RX7 showed broad-spectrum antibacterial, as well as antifungal, activity and inhibited the growth of *Candida albicans*, the causative agent of cutaneous candidiasis in humans.¹³³ While the mode of action of bacteriocins against Gram-positive bacteria has been studied in depth, the direct mechanism of action of bacteriocins against Gram-negative bacteria is poorly understood.^{132,134} Tiwari et al.¹³⁴ demonstrated the ability of bacteriocins enterocin E50–52,

pediocin PA-1 and its hybrid peptides, EP and PE, to induce the efflux of intracellular ATP and to dissipate the cellular transmembrane potential of *E. coli* O157:H7 and *S. enterica* serovar Enteritidis 20E1090. Bacteriocins are mostly cationic peptides, and this characteristic enables electrostatic interactions with the negatively charged head groups of bacterial phospholipid.¹³⁵ This is followed by insertion into the planar lipid bilayer or liposome membranes, leading to the formation of transient channels, leakage of cellular contents and subsequent cell death.¹³⁶

In addition to *in vitro* studies, several *in vivo* studies have demonstrated the inhibitory effect of purified bacteriocins and probiotic bacteriocin-producing strains in infectious animal models. Simonova et al.¹³⁷ observed that feeding rabbits with bacteriocin-producing *Enterococcus faecium* CCM7420 and its partially purified bacteriocin significantly reduced *Staphylococci* spp. cell numbers in the cecum thus protecting the animals against infection. Other studies found that the *E. faecium* EK13, enterocin A producing strain reduced *Salmonella* cell numbers in gnotobiotic Japanese quails and reduced the colonization of pathogenic *Staphylococcus* in the digestive tract of rabbits.^{138,139} The capacity of human-isolated nisin- and pediocin-producing LAB to reduce the intestinal colonization of VRE in mice was demonstrated for the first time by Millette et al.³⁸ Amyloliquecidin and penisin, produced by *B. amyloliquefaciens* and *Paenibacillus* sp. strain A3, respectively, significantly reduced methicillin resistant *S. aureus* (MRSA) infection levels in mice.^{140,141} Svetoch et al.¹⁴² reported a significant reduction of *Salmonella enteritidis* in broilers after oral administration of the *E. faecium* E 50–52 bacteriocin. Corr et al.³⁵ demonstrated that feeding mice with the *Lactobacillus salivarius* UCC11 bacteriocin Abp118-producing strain reduced *L. monocytogenes* cell numbers in the liver and spleen. A similar reduction in cell numbers of the same pathogen in the GIT of mice was observed when the animals were pre-treated with probiotic strains *L. plantarum* 423 and *E. mundtii* ST4SA, producing bacteriocins plantaricin 423 and mundticin ST, respectively.³⁶ Using gene knockout and reverse genetic analysis, the same authors confirmed bacteriocin production and adhesion proteins as mechanisms for the anti-listerial activity.³⁷

Several studies have demonstrated the topical application of bacteriocins to treat skin infections, mastitis and oral infections.^{140–147} Despite their powerful anti-infective therapeutic potential and a large selection of isolated and characterized bacteriocins, these peptides have not yet entered into clinical use.^{148–150} This is likely due to various production difficulties.^{149,151} However, progress in preclinical studies of several bacteriocins has proven promising. Several bacteriocins have been through different stages of preclinical development, targeting multi-drug resistant bacteria as well as cystic fibrosis.¹⁴⁹ These include the bacteriocins, NVB302 and NVB333 (both produced by *Actinoplanes liquoriae* NCIMB41362), mutacin 1140 (produced by *Streptococcus mutans* JH1000), NAI-107 (produced by *Microbispora corallina*) and Moli1901 (produced by *Streptomyces cinnamomeum*).¹⁴⁹

It is also important to consider that not all potential or developed probiotic strains that show *in vitro* antimicrobial activity against enteropathogens will be active *in vivo*. For example, despite the fact that a *Lactobacillus* sp. strain adhered to the jejunum and ileum of gnotobiotic pigs after oral administration and that the strain showed *in vitro* activity against enteropathogenic *E. coli* (EPEC), the strain failed to prevent EPEC colonization in the GIT of infected animals.¹⁵² Similar results were observed when *L. casei* subsp. *casei* failed to prevent the intestinal colonization of EPEC in the GIT of gnotobiotic or conventional piglets when the LAB strain was administered in a preventative setup.¹⁵³ Nevertheless, most probiotics use their bacteriocins to effectively interact with enteropathogens through either bacteriostatic or bactericidal activities. In doing so, they prevent pathogenic colonization of the host GIT and subsequent occurrence of disease.

Bacteriocin-like inhibitory substances

Bacteriocin-like inhibitory substances (BLIS) have a broader spectrum of antimicrobial activity. Many of these compounds are not fully characterized or do not share characteristics typical of bacteriocins.¹² The antimicrobial activities are not related to the production of lactic acid, other organic acids, or hydrogen peroxide.^{12,154} *Lactobacillus rhamnosus* GG secretes an antimicrobial substance with inhibitory activity against *Clostridium* spp., *Staphylococcus* spp.,

Enterobacteriaceae, *Streptococcus* spp., *Bacteriodes* spp., and *Pseudomonas* spp.¹⁵⁵ This low molecular weight (LMW) substance is characterized as heat-stable, distinct from lactic and acetic acids, and closely resembles a microcin that is normally produced by *Enterobacteriaceae* spp. These characteristics suggest that it could be a BLIS.¹² Similar substances with molecular weights and broad activity spectrums uncharacteristic of bacteriocins are produced by other lactobacilli, including strains of *L. acidophilus* and *L. delbrueckii*, their bactericidal effects are related to neither lactic acid nor hydrogen peroxide.^{156,157} Other studies have identified bacteriocin-like antimicrobial substances produced by several *Bifidobacterium* strains with broad spectrums of activity against both Gram-positive and Gram-negative pathogens such as *L. monocytogenes*, *Salmonella* spp. and *E. coli* spp.^{158–160}

Organic acids

An additional mechanism of pathogen displacement in the gut employed by probiotic bacteria is their ability to make the intestinal environment less suitable for pathogen growth. Probiotic LAB and commensal microbiota ferment carbohydrates in the GIT that lead to the production of metabolites such as acetic, formic, succinic and lactic acids, rendering the intestinal environment acidic and inhibiting the growth of bacterial pathogens.¹⁶¹ Organic acids, in particular lactic and acetic acid, repress the growth of many pathogenic bacteria in the GIT.^{12,15,64} The undissociated form of lactic acid functions as a permeabilizer of the Gram-negative bacterial outer cell membrane, after which it dissociates inside the bacterial cytoplasm following entry.¹⁶² The bacterial killing activity is exerted by lowering the intracellular pH level, through the accumulation of ionized forms of the organic acid and other antimicrobial compounds inside the cytoplasm.¹⁶³

De Keersmaecker et al.¹⁶⁴ demonstrated that the strong inhibitory effects of *L. rhamnosus* GG against *S. typhimurium* was due to lactic acid production. Lehto and Salminen¹⁶⁵ demonstrated the potential role of lactic acid in the ability of *Lactobacillus* strain GG to prevent the invasion of Caco-2 cells by *S. enterica* serovar Typhimurium. The authors suggested a pH-dependent mechanism after they observed that inhibition of the pathogen

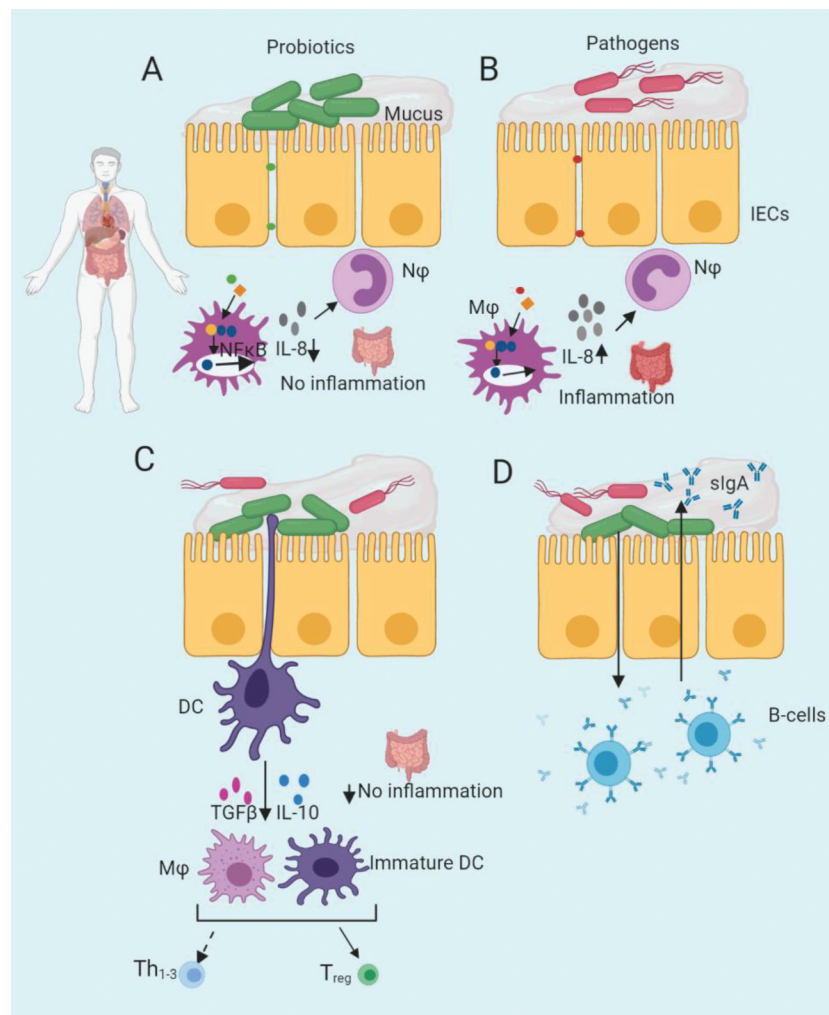


Figure 2. Mucosal immunomodulation by probiotics in the presence of enteric pathogens. **A.** Down-regulation by probiotic bacteria of pro-inflammatory cytokine (IL-8) secretion in the GIT. Probiotic bacteria (or their products) may dampen an innate immune response by inhibiting the NF- κ B inflammatory signaling pathway and influencing the production of IL-8 and subsequent recruitment of inflammatory immune cells to sites of intestinal injury. **B.** Activation of NF- κ B signaling pathway by enteric pathogens, resulting in severe inflammation of intestinal epithelium tissue. **C.** Probiotic signaling of dendritic cells (DCs) to stimulate the secretion of anti-inflammatory cytokines such as IL-10 in response to an intestinal infection. **D.** Probiotics can augment the levels of IgA-secreting plasma cells in the lamina propria and promote the transcytosis of secretory IgA (sIgA) across the epithelial cell layer and secretion into the luminal mucus layer, preventing and limiting bacterial penetration of host tissues. IECs, intestinal epithelial cells; IL-8, interleukin 8; IL-10, interleukin 10; M ϕ , macrophage; N ϕ , neutrophil; NF- κ B, nuclear factor-kappa B. TGF β , transforming growth factor- β ; Th₁₋₃, T helper cells; T_{reg}, regulatory T cells. Figure created in biorender (<http://biorender.io>).

was eliminated when the LAB culture was set to pH 7. In another study, the growth and expression of the HilA and InvF virulence factors by *Salmonella* were affected by lactic acid.¹⁶⁶

The inhibition of *E. coli* O157:H7 by different *Lactococcus* and *Lactobacillus* strains was attributed to the production of lactic acid and low pH.^{167,168} The growth of *Helicobacter pylori* was inhibited by different *Lactobacillus* and *Bifidobacterium* strains including *L. acidophilus*, *L. bulgaricus* and *Bifidobacterium bifidus*.^{169,170} These effects were linked to the

production of lactic, acetic and hydrochloric acid. In another study, the growth of four species of known enteropathogens, *H. pylori*, *Campylobacter jejuni*, *Campylobacter coli* and *C. difficile* was inhibited by *Lactobacillus* strains isolated from the human GIT, probably due to the production of organic acids.¹⁷¹ Based on these studies, it is reasonable to suggest that the production of organic acids by probiotics in the GIT makes the intestinal environment less favorable for their competitors and decreases the risk of enteric infections by pathogens.

Hydrogen peroxide

In addition to lactic acid and bacteriocin production, hydrogen peroxide (H_2O_2) production by commensal or probiotic LAB may be an important antimicrobial mechanism against pathogens.¹⁷² Hydrogen peroxide may cause reduced pathogen virulence, reduced pathogen invasion of epithelial cells or death of intestinal pathogens after epithelial intracellular diffusion which alters gene transcription and signal transduction.^{173,174} Several H_2O_2 -producing bacterial species with probiotic properties have been isolated, such as *B. bifidum*, the *Lactobacillus johnsonii* NCC 533 gut isolate, a *L. delbrueckii* subsp. *bulgaricus* yogurt isolate and normal microflora vaginal isolates such as *L. crispatus* and *L. gasseri*.^{174–181}

The ability of *L. johnsonii* NCC533 to generate up to millimolar quantities of H_2O_2 under aerobic conditions has been demonstrated.¹⁷⁶ The authors demonstrated the antimicrobial role of *L. johnsonii* NCC533 produced H_2O_2 against *S. enterica* serovar Typhimurium *in vitro*, and proposed that *L. johnsonii* NCC533 H_2O_2 -production could contribute to protection against the pathogen *in vivo*. Other studies have shown that H_2O_2 -producing *L. crispatus* F117 and *Lactobacillus paracasei* strains (F2 and F28) inhibited the growth of *S. aureus in vitro*.^{181,182} The beneficial role of H_2O_2 -producing probiotic LAB that form part of the vaginal microflora of healthy women has been studied extensively.^{180–183} Previous studies have reported that women carrying H_2O_2 -producing lactobacilli are less likely to develop bacterial vaginosis.^{180,183}

Siderophores

Iron is an essential micronutrient that plays a central role in the metabolism and proliferation of most gut microbes, including commensal bacteria and gut pathogens.¹⁸⁴ Siderophores are LMW, organic, high-affinity iron-chelating compounds produced by microorganisms such as bacteria and fungi.¹⁸⁵ These compounds inhibit the growth, proliferation and persistence of competing microbes by depriving them of iron. In doing so, siderophore-producing bacteria sequester free iron available in their environment that is essential to other microorganisms. For example, the growth of *Lc. lactis*, *C. difficile*

and *Clostridium perfringens* was inhibited in the GIT by iron-binding *Bifidobacterium* strains that produce siderophores.¹⁸⁶ Growth and adhesion of enteropathogenic *S. typhimurium* N15 and EHEC to IECs were inhibited by *B. pseudolongum* PV8-2 and *Bifidobacterium kashiwanohense* PV20-2 with high iron sequestration properties.¹⁸⁷

Biosurfactants

The production of biosurfactants by some LAB is another mechanism that can interfere with pathogen growth in the GIT. Biosurfactants are a group of compounds with surface and emulsifying activities used in many different biomedical applications.^{188,189} Several LAB strains have been isolated that produce either cell-bound or secreted biosurfactants with antibacterial, antiviral and antifungal properties.^{188–193} Biosurfactants cause permeabilization of cells by effecting changes that disrupt or lyse the physical cell membrane structure.¹⁹⁴ The use of biosurfactant-producing lactobacilli in the prevention of urogenital tract infections is of considerable interest.¹⁸⁸ These organisms are believed to compete with urogenital bacterial pathogens and yeast for adhesion sites on epithelial cells and control their growth by the production of biosurfactants.^{195–197} In another study, *L. casei* MRTL3 that produces a bacteriocin and a biosurfactant, inhibited a broad range of pathogens, including *L. monocytogenes*, *S. aureus*, *Shigella flexneri* and *Pseudomonas aeruginosa*.¹⁹⁸

Compounds inhibiting pathogen adhesion to intestinal cells

Adhesion to intestinal cells and subsequent colonization by enteropathogens is regarded a prerequisite for virulence.¹⁹⁹ Probiotic bacteria produce compounds that do not have a direct bactericidal effect, but contribute to the normal anti-infectious activities of the GIT by inhibiting the binding of pathogenic bacteria to the mucosal surface. Fujiwara et al.²⁰⁰ purified and characterized a novel proteinaceous compound in culture supernatants of *B. longum* SBT2928, termed BIF, that inhibits the adhesion of enterotoxigenic *E. coli* Pb176 (ETEC) to human HCT-8 IECs. The authors demonstrated that BIF blocks the binding of the ETEC Pb176 colonization

factor antigen (CFA) II adhesive factor to ganglioside-tetraosylceramide (bacterial binding structure) receptors on the intestinal cell surface, thereby preventing ETEC Pb176 colonization.²⁰⁰ Two *Bifidobacterium* strains, CA1 and F9, isolated from the GIT of infants produce a LMW, lipophilic, antibacterial compound that inhibits the adhesion of several pathogenic bacteria, including *S. typhimurium* SL1344 and *E. coli* C1845.²⁰¹

Stabilization of intestinal epithelial barrier

The GI epithelium consists of a uni-layer of cells covered by a mucus layer that is constantly exposed to the luminal contents and various enteric bacteria.^{78,202} The intestinal epithelial barrier consists of the mucus layer, the intestinal cells and the gut innate immune system.²⁰² This GI barrier functions as a key defense mechanism required to maintain epithelial integrity and to prevent infection by pathogens and excessive inflammation. Stabilization and maintenance of this barrier is thus of utmost importance to the host. Important defense mechanisms of the intestinal barrier against unwelcome intrusion of harmful antigens include the mucosal layer (mucin production), intercellular junctional complexes (tight and adherence junctions) and the secretion of antimicrobial peptides (such as defensins) and immunoglobulin A (IgA).^{202–205} Disruption of this barrier function can lead to inappropriate inflammatory responses due to invasion of the submucosa by bacteria or food antigens, which may result in intestinal disorders such as inflammatory bowel disease (IBD) and ulcerative colitis.^{203,206,207} Consumption of colonizing or non-colonizing probiotics can enhance barrier integrity which helps to protect the intestinal epithelium against enteric pathogens and chronic inflammation by direct effects on the epithelium (e.g. increasing mucin expression by goblet cells), modulation of the immune system and by direct effects on commensal and pathogenic bacteria (e.g. antimicrobial peptides and competition for adherence) **Figures 1 and 2.**

Intestinal epithelium cells are overlaid with a protective inner and outer mucus layer that limits bacterial movement and acts as a dynamic defense barrier against enteropathogens and other potentially harmful antigens.²⁰⁴ For enteropathogens to colonize the intestine, they have to penetrate the

mucus layer before they reach the intestinal epithelium.²⁰⁸ Mucins are the major macromolecular constituents of the epithelial mucus layer and are produced by specialized goblet cells in the intestinal tract.²⁰⁹ Probiotics are able to inhibit pathogen adherence to IECs by promoting the secretion of intestinal mucins and defensins **Figure 1a.** Several *Lactobacillus* species have been shown to increase the expression of specific mucin genes in human intestinal Caco-2 and HT29 cells, thus preventing the adherence and internalization of pathogenic *E. coli*.^{91,210,211} The adherence of EPEC was inhibited by *L. plantarum* 299 v-mediated increase in expression of the MUC2 and MUC3 mucins.^{91,211} Rats administered with VSL3 (pre- and probiotic mixture) for 7 consecutive days showed a 60-fold increase in MUC2 expression and an associated increase in mucin production.²¹² Therefore, increased mucus production mediated by probiotic bacteria *in vivo* may be a key mechanism in their interactions with enteropathogens to prevent infections and to improve intestinal barrier function.

Co-aggregation

Probiotic bacteria can prevent enteropathogenic adherence and intestinal colonization by co-aggregating with pathogens.^{213–215} In this process, probiotic bacteria interact closely with pathogens, allowing them the opportunity to release their anti-pathogenic substances in proximity to the pathogens. Probiotic LAB can form multi-cellular aggregates that are crucial for colonization of the oral cavity, the urogenital tract and the GIT.^{37,213,215–217} The ability of probiotic cells to co-aggregate is characterized by the clumping of cells that are genetically distinct, whereas auto-aggregation involves cells of the same strain.^{218,219} Auto- and co-aggregation have been reported for various *Lactobacillus* species, including *L. plantarum*, *L. reuteri*, *L. gasseri*, *L. crispatus* and *L. coryniformis*.^{219,220} Several studies have shown that the auto- and co-aggregation abilities of probiotic cells enhances their colonization and may enable the formation of a barrier to prevent colonization by pathogens.^{213–215,218} *Lactobacillus plantarum* strains (S1, A and B) co-aggregate with selected food-borne pathogens including *S. typhimurium* and *L. monocytogenes*.²¹⁸ *Lactobacillus plantarum* S1 co-aggregated best with EHEC at 41.5%, *L. plantarum*

A co-aggregated with *S. Typhimurium* at 40.5% and *L. plantarum* B co-aggregated with *L. monocytogenes* at 37.4%.²¹⁸ This is a clear indication that the ability of LAB strains to bind to the food-borne pathogens is not restricted to one species or a single strain. In another study, the adherence of ETEC to porcine enterocytes was affected by co-aggregation of the pathogen with selected *Lactobacillus* spp., including *L. fermentum*, *L. salivarius* and *L. delbrueckii*.²²¹

The co-aggregation ability of probiotic LAB is generally related to a great diversity of properties among cell surface adherence proteins.^{222,223} Kos et al.²¹⁵ demonstrated that differences in the hydrophobicity and hydrophilicity of the structural cell surface of *L. acidophilus* M9, *L. plantarum* L4 and *E. faecium* L3 may be responsible for the abilities of the strain to co-aggregate. In other examples, proteins involved in the maintenance of cell shape including the S-layer protein CbsA of *L. crispatus* JCM 5810, the DEAD box helicase AggH of *L. reuteri* 1063 and Apf of *L. gasseri* 4B2 were all responsible for the mediation of auto-aggregation.^{224–226} This suggests that the co-aggregation phenomenon of probiotic LAB may be a secondary activity of cell surface components involving random interactions with other surface components. Schachtsiek et al.²¹⁹ described the role of a *Lactobacillus coryniformis* DSM 20001 surface protein encoded by a *cpf* gene (co/aggregation-promoting factor) in the ability of the LAB strain to co-aggregate with *E. coli* K-88, *C. coli* and *C. jejuni*. The auto- and co-aggregation ability of *L. acidophilus* M92 was shown to be mediated by proteinaceous surface layer (S-layer) components, approximated at 45 kDa in size.²¹⁵

Co-aggregation of probiotic and pathogenic bacteria is also mediated via the attachment of probiotic cells to fimbriae expressed on the cell surface of pathogens.²¹³ This makes sense, as several studies have shown that probiotic LAB can prevent enteropathogenic binding to intestinal epithelial cells by attaching to the same carbohydrate receptor sites as the pathogens.^{14,18,85,86} The attachment of probiotic cells to the surface of pathogenic cells is dependent on the specific type of fimbriae expressed by the pathogen.²¹³ The expression of fimbriae by pathogens is important in colonization of the GIT, the vagina and perineum.^{84,196,213,227} For example, *E. coli* that express type I fimbriae are most commonly associated with urinary tract infections.²²⁷

Mizuno et al.²²⁰ presented *E. coli* fimbriae and lipopolysaccharide (LPS) as the essential mediators of the co-aggregation of *L. casei* NBRC 3831 with *E. coli* K-12. Spencer and Chesson²²¹ showed that selected strains of lactobacilli co-aggregate with enterotoxigenic *E. coli* expressing K88 fimbriae, but not with a K88-negative knockout mutant strain.

Inhibition of flagella motility

Flagella are known to play an important role as a virulence factor in many bacterial pathogens.²²⁸ Flagella allow pathogenic bacteria to respond to attractant and repellent gradients and are crucial for attachment to, and invasion of, eukaryotic cells.^{229–232} Foodborne pathogens such as *S. enterica* serovar Typhimurium require actively rotating flagella to rapidly contact and to efficiently penetrate GI epithelial cells. *Salmonella* Typhimurium remained noninvasive in infected mice when treated with a potent antibody that inhibits flagellum-based motility.²³³ Probiotic bacteria can impair the flagella motility of enteropathogens, thus preventing pathogenic colonization of the gut.

Lieven-Le Moal et al.²³⁴ demonstrated that anti-diarrhoeic *L. acidophilus* LB and its secreted products inhibited the entry of *S. enterica* serovar Typhimurium into human intestinal Caco-2 cells by disrupting the swimming motility of the diarrhea-associated enteropathogen. The authors showed that *L. acidophilus* LB secretes a heat stable LMW product that causes rapid depolarization of the *S. Typhimurium* SL1344 cytoplasmic membrane. The inhibitory activity did not affect bacterial viability or flagellum expression. The transient impairment of the swimming motility of *S. Typhimurium* SL1344 leads to a delay in the pathogen's capacity to induce F-actin membrane remodeling and thus entry into intestinal Caco-2 cells. In another study, levels of translocated *Salmonella* were dramatically lower in mice orally infected with *S. Typhimurium* when treated with the cell free supernatant of live probiotic lactobacilli, compared to that of untreated mice.²³⁵ It is possible that the difference between cell numbers of treated and untreated groups is due to a delay in pathogen translocation across the intestinal epithelial barrier

caused by the inhibition of *Salmonella* swimming motility. This in turn, exposed the pathogenic cells for extended periods to host defenses in the intestinal lumen that includes antimicrobial products from both the host cells and the microbiota.

Immune system modulation

It is well known that probiotic bacteria can exert regulatory effects on host innate and adaptive immune responses.²³⁶ These bacteria have the ability to modulate the functions of dendritic cells (DCs), monocytes/macrophages, and T and B lymphocytes, which enhances phagocytosis of invading gut pathogens.^{237,238} By stimulation of the host immune responses (specific and nonspecific), probiotic bacteria can displace pathogens in the GIT and prevent intestinal diseases.^{238–241}

Probiotic bacteria can interact with pathogens in the gut by antagonizing inflammatory responses induced by the gut pathogens.²³⁶ Inflammation allows pathogens to flourish at the expense of the natural microbiota and host intestinal health. Probiotic bacteria are able to trigger an anti-inflammatory response from the innate immune system by signaling DCs to secrete anti-inflammatory cytokines such as interleukin 10 (IL-10) **Figure 2c**.^{242,243} They can also elicit a decrease in pro-inflammatory cytokines during inflammation.²³⁷ Down-regulation of pro-inflammatory cytokine secretion from immune cells occurs as a result of probiotic bacterial interference with inflammatory signaling pathways such as nuclear factor-kappa B (NF- κ B) and mitogen-activated protein kinases (MAPK) **Figure 2a**.^{244,245} The activation of these signaling pathways leads to the secretion of pro-inflammatory cytokines that can severely damage the intestinal epithelial barrier. NF- κ B and MAPK signaling pathways are activated by enteric pathogens to stimulate the secretion of pro-inflammatory cytokines (e.g. IL-8), that lead to the recruitment of inflammatory immune cells (e.g. neutrophils) to the infected area resulting in severe inflammation, tissue damage and disease **Figure 2b**.^{244,245} Several studies have identified probiotic strains with the ability to suppress pro-inflammatory cytokine production to avoid pathogen-induced inflammation at infection sites.^{236,245–247} A study by Finamore et al.²⁴⁶ reported that *Lactobacillus amylovorus*

DSM 16698 protected IECs against the pro-inflammatory response induced by ETEC K88 through the suppression of pro-inflammatory cytokines IL-8 and IL-1 β . Another study demonstrated the ability of *L. casei* OLL2768 to suppress the ETEC-induced pro-inflammatory response by inhibition of NF- κ B and MAPK pathways that reduced pro-inflammatory cytokine levels.²⁴⁷

Probiotics also play a role in the stimulation and production of antibodies in the gut, particularly immunoglobulin A (IgA) **Figure 2d**.^{236,237} Antibodies released in the intestinal lumen can inhibit pathogen adherence to IECs by interfering with adhesive cell receptors on the pathogen's cell membrane. Previous studies have indicated that *Saccharomyces boulardii* and *L. rhamnosus* GG increased secretory IgA levels or immunoglobulin-secreting cell levels in the GIT.^{248,249} Other studies have reported that oral administration of probiotic lactobacilli increased IgA levels in children suffering from diarrhea, thereby shortening the duration of symptoms.^{237,250–252} Several probiotic strains can also modulate the host immune mechanisms by influencing phagocytosis of enteric pathogens by host immune phagocytic cells such as macrophages.^{14,18,64,248,251,253,254} The inhibition of enteropathogenic *P. aeruginosa* and *L. monocytogenes* in mice by a strain of *L. casei* correlates to an increase in abundance of macrophages.²⁵⁵ Furthermore, probiotic bacteria can affect phagocytotic cell activities not only in clinical situations but also in healthy subjects.^{256–258}

Conclusion

Elucidating the mechanisms of action of probiotic microorganisms is a difficult task given the complex nature of the human GI ecosystem. Probiotic mechanisms of action employed against enteric pathogens are diverse, heterogeneous and may be strain specific. This suggests that the mechanism(s) of action of one specific probiotic strain against a particular disease or pathogen cannot be generalized since different strains evoke different responses in the host. Thus, the health benefits conferred by one strain are not applicable to

another strain, even within the same species. Understanding the full potential of probiotics for therapeutic or prophylactic applications against GI diseases requires thorough investigations of probiotic-host and probiotic-pathogen interactions. Enhanced understanding of these interactions will enable the identification of true probiotics to target specific enteric diseases. While a number of recent *in vivo* studies have demonstrated the mechanistic basis behind observed probiotic effects at the molecular level, greater emphasis is warranted in this area of probiotic research. Unraveling the intricacies of probiotic-host and probiotic-pathogen interactions will improve the *in vitro* selection of the best probiotics based on key properties such as bacteriocin production and adhesion genes. Moreover, an efficient probiotic should exhibit stimulation of the hosts' immune system and ultimately, must have demonstrable beneficial health effects on the host. It is clear that the demonstration of key antimicrobial and protective probiotic mechanisms *in vivo* will allow for industry and consumers to choose scientifically validated probiotics for the prevention or treatment of various health problems.

Authors' contributions

All authors read and approved the final manuscript as submitted. This manuscript was mainly written by WVZ and edited by SD and LD.

Disclosure of potential conflicts of interest

The authors declare that they have no competing interests.

Funding

This work was supported by the National Research Foundation of South Africa (103843). WVZ received a post-doctoral fellowship from the National Research Foundation of South Africa.

References

- Gerritsen J, Smidt H, Rijkers GT, de Vos GM. Intestinal microbiota in human health and disease: the impact of probiotics. *Genes Nutr*. 2011;6:209–240.
- Bron PA, van Baarlen P, Kleerebezem M. Emerging molecular insights into the interaction between probiotics and the host intestinal mucosa. *Nat Revs*. 2012;10:66–78.
- Stanton C, Ross RP, Fitzgerald GF, van Sinderen D. Fermented functional foods based on probiotics and their biogenic metabolites. *Curr Opin Biotechnol*. 2005;16(2):198–203. doi:10.1016/j.copbio.2005.02.008.
- Varankovich NV, Nickerson MT, Korber DR. Probiotic based strategies for therapeutic and prophylactic use against multiple gastrointestinal diseases. *Front Microbiol*. 2015;6:685. doi:10.3389/fmicb.2015.00685.
- Kale-Pradhan PB, Jassal HK, Wilhelm SM. Role of *Lactobacillus* in the prevention of antibiotic-associated diarrhea: a meta-analysis. *Pharmacotherapy*. 2010;30(2):119–126. doi:10.1592/phco.30.2.119.
- Moayyedi P, Ford AC, Talley NJ, Cremonini F, Foxx-Orenstein AE, Brandt LJ, Quigley EM. The efficacy of probiotics in the treatment of irritable bowel syndrome: a systematic review. *Gut*. 2010;59(3):325–332. doi:10.1136/gut.2008.167270.
- Ishimwe N, Daliri EB, Lee BH, Fang F, Du G. The perspective on cholesterol-lowering mechanisms of probiotics. *Mol Nutr Food Res*. 2015;59:94–105.
- Andersson H, Asp NG, Bruce A, Roos A, Wadström T, Wold AE. Health effects of probiotics and prebiotics. A literature review on human studies. *Scandinavian J Nutr*. 2001;45:58–75.
- Culligan EP, Hill C, Sleator RD. Probiotics and gastrointestinal disease: successes, problems and future prospects. *Gut Pathog*. 2009;1(1):19. doi:10.1186/1757-4749-1-19.
- O'Sullivan GC, Kelly P, O'Halloran S, Collins C, Collins JK, Dunne C, Shanahan F. Probiotics: an emerging therapy. *Curr Pharm Des*. 2005;11(1):3–10. doi:10.2174/1381612053382368.
- Marco ML, Pavan S, Kleerebezem M. Towards understanding molecular modes of probiotic action. *Curr Opin Biotechnol*. 2006;17(2):204–210. doi:10.1016/j.copbio.2006.02.005.
- Servin AL. Antagonistic activities of lactobacilli and bifidobacteria against microbial pathogens. *FEMS Microbiol Revs*. 2004;28(4):405–440. doi:10.1016/j.femsre.2004.01.003.
- Lebeer S, Vanderleyden J, De Keersmaecker SJC. Genes and molecules of Lactobacilli supporting probiotic action. *Microbiol Mol Biol Rev*. 2008;72(4):728–764. doi:10.1128/MMBR.00017-08.
- Mathipa MG, Thantsha MS. Probiotic engineering: towards development of robust probiotic strains with enhanced functional properties and for targeted control of enteric pathogens. *Gut Pathog*. 2017;9(1):28. doi:10.1186/s13099-017-0178-9.
- Bermudez-Brito M, Plaza-Díaz J, Muñoz-Quezada S, Gómez-Llrente C, Gill A. Probiotic mechanisms of action. *Ann Nutr Metab*. 2012;61(2):160–174. doi:10.1159/000342079.

16. Bezkorovainy A. Probiotics: determinants of survival and growth in the gut. *Am J Clin Nutr.* 2001;73(2):S399–S405. doi:10.1093/ajcn/73.2.399s.
17. Corr SC, Hill C, Gahan CG. Understanding the mechanisms by which probiotics inhibit gastrointestinal pathogens. *Adv Food Nutr Res.* 2009;56:1–15.
18. Plaza-Diaz J, Rulz-Ojeda FJ, Gil-Campos M, Gil A. Mechanisms of action of probiotics. *Adv Nutr.* 2019;10(suppl_1):S49–S66. doi:10.1093/advances/nmy063.
19. Dunne C, O'Mahony L, Murphy L, Thornton G, Morrissey D, O'Halloran S, Feeney M, Flynn S, Fitzgerald G, Daly C, et al. In vitro selection criteria for probiotic bacteria of human origin: correlation with in vivo findings. *Am J Clin Nutr.* 2001;73(2):S386–S392. doi:10.1093/ajcn/73.2.386s.
20. Tuomola E, Crittenden R, Playne M, Isolauri E, Salminen S. Quality assurance criteria for probiotic bacteria. *Am J Clin Nutr.* 2001;73(2):S393–S398. doi:10.1093/ajcn/73.2.393s.
21. Denou E, Pridmore RD, Berger B, Panoff J, Arigoni F, Brüssow H. Identification of genes associated with the long-gut-persistence phenotype of the probiotic *Lactobacillus johnsonii* strain NCC533 using a combination of genomics and transcriptome analysis. *J Bacteriol.* 2007;190(9):3161–3168. doi:10.1128/JB.01637-07.
22. Hoepelman AIM, Tuomanen EI. Consequences of microbial attachment: directing host cell functions with adhesins. *Infect Immun.* 1992;60(5):1729–1733. doi:10.1128/IAI.60.5.1729-1733.1992.
23. Bernet MF, Brassart D, Neeser JR, Servin AL. Adhesion of human bifidobacterial strains to cultured human intestinal epithelial cells and inhibition of enteropathogen-cell interactions. *Appl Environ Microbiol.* 1992;59(12):4121–4218. doi:10.1128/AEM.59.12.4121-4128.1993.
24. Hudault S, Lievin V, Bernet-Camard MF, Servin AL. Antagonistic activity exerted in vitro and in vivo by *Lactobacillus casei* (strain GG) against *Salmonella typhimurium* C5 infection. *Appl Environ Microbiol.* 1993;63(2):513–518. doi:10.1128/AEM.63.2.513-518.1997.
25. Ramiah K, van Reenen C, Dicks LMT. Surface-bound proteins of *Lactobacillus plantarum* 423 that contribute to adhesion of Caco-2 cells and their role in competitive exclusion and displacement of *Clostridium sporogenes* and *Enterococcus faecalis*. *Res Microbiol.* 2008;159(6):470–475. doi:10.1016/j.resmic.2008.06.002.
26. Basu S, Paul DK, Ganguly S, Chatterjee M, Chandra PK. Efficacy of high-dose *Lactobacillus rhamnosus* GG in controlling acute watery diarrhea in Indian children: a randomized controlled trial. *J Clin Gastroenterol.* 2009;43(3):208–213. doi:10.1097/MCG.0b013e31815a5780.
27. Armuzzi A, Cremonini F, Bartolozzi F, Canducci F, Candelli M, Ojetti V, Cammarota V, Anti M, De Lorenzo A, Pola P. The effect of oral administration of *Lactobacillus* GG on antibiotic-associated gastrointestinal side-effects during *Helicobacter pylori* eradication therapy. *Aliment Pharmacol Ther.* 2001;15(2):163–169. doi:10.1046/j.1365-2036.2001.00923.x.
28. Cruchet S, Obregon MC, Salazar G, Diaz E, Gotteland M. Effect of the ingestion of a dietary product containing *Lactobacillus johnsonii* La1 on *Helicobacter pylori* colonization in children. *Nutr.* 2003;19(9):716–721. doi:10.1016/S0899-9007(03)00109-6.
29. Tursi A, Brandimarte G, Giorgetti GM, Modeo ME. Effect of *Lactobacillus casei* supplementation on the effectiveness and tolerability of a new second-line 10-day quadruple therapy after failure of a first attempt to cure *Helicobacter pylori* infection. *Med Sci Monit.* 2004;10:662–666.
30. de LeBlanc AD, Castillo NA, Perdigon G. Anti-infective mechanisms induced by a probiotic *Lactobacillus* strain against *salmonella enterica* serovar typhimurium infection. *Int J Food Microbiol.* 2010;138(3):223–231. doi:10.1016/j.ijfoodmicro.2010.01.020.
31. Gill HS, Shu Q, Lin H, Rutherford KJ, Cross ML. Protection against translocating *Salmonella typhimurium* infection in mice by feeding the immuno-enhancing probiotic *Lactobacillus rhamnosus* strain HN001. *Med Microbiol Immunol.* 2001;190(3):97–104. doi:10.1007/s004300100095.
32. Silva AM, Barbosa FHF, Duarte R, Vieira LQ, Arantes RME, Nicoli JR. Effect of *Bifidobacterium longum* ingestion on experimental salmonellosis in mice. *J Appl Microbiol.* 2004;97:29–37.
33. Dicks LMT, Ten Doeschate K. *Enterococcus mundtii* ST4SA and *Lactobacillus plantarum* 423 alleviated symptoms of *Salmonella* infection, as determined in Wistar rats challenged with *Salmonella enterica* serovar Typhimurium. *Curr Microbiol.* 2010;61(3):184–189. doi:10.1007/s00284-010-9594-5.
34. Archambaud C, Nahori M, Soubigou G, Becavin C, Laval L, Lechat P, Smokvina T, Langella P, Lecuit M, Cossart P. Impact of lactobacilli on orally acquired listeriosis. *Proc Natl Acad Sci USA.* 2012;109(41):16684–16689. doi:10.1073/pnas.1212809109.
35. Corr SC, Li Y, Riedel CU, O'Toole PW, Hill C, Gahan CGM. Bacteriocin production as a mechanism for the anti-infective activity of *Lactobacillus salivarius* UCC18. *Proc Natl Acad Sci USA.* 2007;104(18):7617–7621. doi:10.1073/pnas.0700440104.
36. Van Zyl WF, Deane SM, Dicks LMT. *Enterococcus mundtii* ST4SA and *Lactobacillus plantarum* 423 excludes *Listeria monocytogenes* from the GIT, as shown by bioluminescent studies in mice. *Benef Microbes.* 2015;7(2):227–235. doi:10.3920/BM2015.0082.
37. Van Zyl WF, Deane SM, Dicks LMT. Bacteriocin production and adhesion properties as mechanisms for the anti-listerial activity of *Lactobacillus plantarum* 423 and *Enterococcus mundtii* ST4SA. *Benef Microbes.* 2019;10(3):329–349. doi:10.3920/BM2018.0141.
38. Millette M, Cornut G, Dupont C, Shareck F, Archambault D, Lacroix M. Capacity of human nisin-

- and pediocin-producing lactic acid bacteria to reduce intestinal colonization by vancomycin-resistant enterococci. *Appl Environ Microbiol.* 2008;74(7):1997–2003. doi:10.1128/AEM.02150-07.
39. Johnson-Henry KC, Nadjafi M, Avitzur Y, Mitchell DJ, Ngan B, Galindo-Mata E, Jones NL, Sherman PM. Amelioration of the effects of *Citrobacter rodentium* infection in mice by pretreatment with probiotics. *J Infect Dis.* 2005;191(12):2106–2117. doi:10.1086/430318.
 40. Mackos AR, Eubank TD, Parry NMA, Bailey MT, McCormick BA. Probiotic *Lactobacillus reuteri* attenuates the stressor-enhanced severity of *Citrobacter rodentium* infection. *Infect Immun.* 2013;81(9):3253–3263. doi:10.1128/IAI.00278-13.
 41. Asahara T, Shimizu K, Nomoto K, Asahara T, Shimizu K, Nomoto K. probiotic bifidobacteria protect mice from lethal infection with shiga toxin-producing *escherichia coli* O157:H7. *Infect Immun.* 2004;72(4):2240–2247. doi:10.1128/IAI.72.4.2240-2247.2004.
 42. Jonganurakkun B, Wang Q, Xu SH, Tada Y, Minamida K, Yasokawa D, Sugi M, Hara H, Asano K. *Pediococcus pentosaceus* NB-17 for probiotic use. *J Biosci Bioeng.* 2008;106(1):69–73. doi:10.1263/jbb.106.69.
 43. Foligné B, Dewulf J, Breton J, Claisse O, Lonvaud-Funel A, Pot B. Probiotic properties of non-conventional lactic acid bacteria: immunomodulation by *Oenococcus oeni*. *Int J Food Microbiol.* 2010;140(2–3):136–145. doi:10.1016/j.ijfoodmicro.2010.04.007.
 44. Kurugol Z, Koturoglu G. Effects of *Saccharomyces boulardii* in children with acute diarrhoea. *Acta Paediatr.* 2005;94(1):44–47. doi:10.1080/08035250410022521.
 45. Kelesidis T, Pothoulakis C. Efficacy and safety of the probiotic *Saccharomyces boulardii* for the prevention and therapy of gastrointestinal disorders. *Therapeu Adv Gastroenterol.* 2012;5(2):111–125. doi:10.1177/1756283X11428502.
 46. Scharek L, Hartmann L, Heinevetter L, Blaut M. *Bifidobacterium adolescentis* modulates the specific immune response to another human gut bacterium, *Bacteroides thetaiotaomicron*, in gnotobiotic rats. *Immunobiol.* 2000;202(5):429–441. doi:10.1016/S0171-2985(00)80102-3.
 47. Sinn DH, Song JH, Kim HJ, Lee JH, Son HJ, Chang DK, Kim YH, Kim JJ, Rhee JC, Rhee PL. Therapeutic Effect of *Lactobacillus acidophilus*-SDC 2012, 2013 in patients with Irritable bowel syndrome. *Digest Dis Sci.* 2008;53(10):2714–2718. doi:10.1007/s10620-007-0196-4.
 48. Sambuy Y, De Angelis I, Ranaldi G, Scarino ML, Stammati A, Zucco F. The Caco-2 cell line as a model of the intestinal barrier: influence of cell and culture related factors on Caco-2 cell functional characteristics. *Cell Bio Toxicol.* 2005;21(1):1–26. doi:10.1007/s10565-005-0085-6.
 49. Coconier MH, Klaenhammer TR, Kernels S, Bernet MF, Servin AL. Protein-mediated adhesion of *Lactobacillus acidophilus* BG2FO4 on human enterocyte and mucus-secreting cell lines in culture.. *Appl Environ Microbiol.* 1992;58(6):2034–2039. doi:10.1128/AEM.58.6.2034-2039.1992.
 50. Pavan S, Desreumaux P, Mercenier A. Use of mouse models to evaluate the persistence, safety, and immune modulation capacities of lactic acid bacteria. *Clin Diagn Lab Immunol.* 2003;10(4):696–701. doi:10.1128/CDLI.10.4.696-701.2003.
 51. Vesa T, Pochart P, Marteau P. Pharmacokinetics of *Lactobacillus plantarum* NCIMB 8826, *Lactobacillus fermentum* KLD, and *Lactococcus lactis* MG 1363 in the human gastrointestinal tract. *Aliment Pharmacol Ther.* 2000;14(6):823–828. doi:10.1046/j.1365-2036.2000.00763.x.
 52. Marteau P, Vesa T. Pharmacokinetics of probiotics and biotherapeutic agents in humans. *Biosci Microflora.* 1998;17(1):1–6. doi:10.12938/bifidus1996.17.1.
 53. Su P, Henriksson A, Tandianus JE, Park JH, Foong F, Dunn NW. Detection and quantification of *Bifidobacterium lactis* LAFTI Å-Å½ B94 in human faecal samples from a consumption trial. *FEMS Microbiol Lett.* 2005;244(1):99–103. doi:10.1016/j.femsle.2005.01.022.
 54. Larsen CN, Nielsen S, Kaestel P, Brockmann E, Bennedsen M, Christensen HR, Eskesen DC, Jacobsen BL, Michaelsen KF. Dose-response study of probiotic bacteria *Bifidobacterium animalis* subsp. *lactis* BB-12 and *Lactobacillus paracasei* subsp. *paracasei* CRL-341 in healthy young adults. *Eur J Clin Nutr.* 2006;60(11):1284–1293. doi:10.1038/sj.ejcn.1602450.
 55. Fujiwara S, Seto Y, Kimura A, Hashiba H. Intestinal transit of an orally administered streptomycin-rifampicin-resistant variant of *Bifidobacterium longum* SBT2928: its long-term survival and effect on the intestinal microflora and metabolism. *J Appl Microbiol.* 2001;90(1):43–52. doi:10.1046/j.1365-2672.2001.01205.x.
 56. Alander M, De Smet I, Nollet L, Verstraete W, von Wright A, Mattila-Sandholm T. The effect of probiotic strains on the microbiota of the simulator of the human intestinal microbial ecosystem (SHIME). *Int J Food Microbiol.* 1999;46(1):71–79. doi:10.1016/S0168-1605(98)00182-2.
 57. Tannock GW, Munro K, Harmsen HJM, Welling GW, mart J, Gopal PK. Analysis of the fecal microflora of human subjects consuming a probiotic product containing *lactobacillus rhamnosus*DR20. *Appl Environ Microbiol.* 2000;66(6):2578–2588. doi:10.1128/AEM.66.6.2578-2588.2000.
 58. Collins J, Dunne C, Murphy L, Morrissey D, O'Mahony L, O'Sullivan E, Fitzgerald G, Kiely B, O'Sullivan G, Daly C, et al. A randomised controlled trial of a probiotic *Lactobacillus* strain in healthy adults: assessment of its delivery, transit and influence on microbial flora and enteric immunity. *Microbial Ecol Health Dis.* 2002;14(2):81–89. doi:10.1080/08910600260081720.
 59. Van Zyl WF, Deane SM, Dicks LMT. In vivo bioluminescence imaging of the spatial and temporal colonization of *Lactobacillus plantarum* 423 and *Enterococcus*

- mundtii* ST4SA in the intestinal tract of mice. BMC Microbiol. 2018;18(1):171. doi:10.1186/s12866-018-1315-4.
60. Johansson ML, Nobaek S, Berggren A, Nyman M, Bjorck I, Ahrne A, Jeppsson B, Molin G. Survival of *Lactobacillus plantarum* DSM 9843 (299v), and effect on the short-chain fatty acid content of faeces after ingestion of a rose-hip drink with fermented oats. Int J Food Microbiol. 1998;42(1-2):29-38. doi:10.1016/S0168-1605(98)00055-5.
 61. Fujiwara S, Seto Y, Kimura A, Hashiba H. Establishment of orally-administered *Lactobacillus gasei* SBT2055SR in the gastrointestinal tract of humans and its influence on intestinal microflora and metabolism. J Appl Microbiol. 2001;90(3):343-352. doi:10.1046/j.1365-2672.2001.01251.x.
 62. Brigidi P, Swennen E, Vitali B, Rossi M, Matteuzzi D. PCR detection of *Bifidobacterium* strains and *Streptococcus thermophilus* in faeces of human subjects after oral bacteriotherapy and yogurt consumption. Int J Food Microbiol. 2003;81(3):203-209. doi:10.1016/S0168-1605(02)00245-3.
 63. Klijin N, Weerkamp AH, de Vos WM. Genetic marking of *Lactococcus lactis* shows its survival in the human gastrointestinal tract.. Appl Environ Microbiol. 1995;61(7):2771-2774. doi:10.1128/AEM.61.7.2771-2774.1995.
 64. Dicks LMT, Botes M. Probiotic lactic acid bacteria in the gastrointestinal tract: health benefits, safety and mode of action. Benef Microbes. 2010;1:11-29.
 65. Andrue N, Zelmer A, Wiles S. Noninvasive biophotonic imaging for studies of infectious disease. FEMS Microbiol Rev. 2011;35(2):360-394. doi:10.1111/j.1574-6976.2010.00252.x.
 66. Van Zyl WF, Deane SM, Dicks LMT. Reporter systems for in vivo tracking of lactic acid bacteria in animal model studies. Gut Microbes. 2015;6(5):291-299. doi:10.1080/19490976.2015.1086058.
 67. Ouwehand AC, Salminen S. The health effects of cultured milk products with viable and non-viable bacteria. Int Dairy J. 1998;8(9):749-758. doi:10.1016/S0958-6946(98)00114-9.
 68. Xiao SD, Zhang DZ, Lu H, Jiang SH, Liu HY, Wang GS, Xu GM, Zhang ZB, Lin GJ, Wang GL. Multicenter, randomized, controlled trial of heat-killed *Lactobacillus acidophilus* LB in patients with chronic diarrhea. Adv Ther. 2003;20(5):253-260. doi:10.1007/BF02849854.
 69. Roy D, Delcenserie V. The protective role of probiotics in disturbed enteric microbiota. In: Malago JJ, Koninkx JFJG, Marinsek-Logar R, editors. Probiotic bacteria and enteric infections. Dordrecht (Netherlands): Springer; 2011. p. 221-261.
 70. Kullen MJ, Amann MM, O'Shaughnessy W, O'Sullivan DJ, Busta FF, Brady U. Differentiation of ingested bifidobacteria by DNA fingerprinting demonstrates the survival of an unmodified strain in the gastrointestinal tract of humans. J Nutr. 1997;127(1):89-94. doi:10.1093/jn/127.1.89.
 71. Siitonen S, Vapautalo H, Salminen S, Gordin A, Saxelin M, Wikberg R, Kirkola AL. Effect of *Lactobacillus* GG yoghurt in prevention of antibiotic associated diarrhoea. Ann Med. 1990;22(1):57-59. doi:10.3109/07853899009147243.
 72. Goldin BR, Gorbach SL, Saxelin M, Barakat S, Gualtieri L, Salminen S. Survival of *Lactobacillus* species (strain GG) in human gastrointestinal tract. Dig Dis Sci. 1992;37(1):121-128. doi:10.1007/BF01308354.
 73. Mättö J, Fondèn R, Tolvanen T, von Wright A, Vilpponen-Salmela T, Satokari R, Saarela M. Intestinal survival and persistence of probiotic *Lactobacillus* and *Bifidobacterium* strains administered in triple-strain yoghurt. Int Dairy J. 2006;16(10):1174-1180. doi:10.1016/j.idairyj.2005.10.007.
 74. Alander M, Korpela R, Saxelin M, Vilpponen-Salmela T, Mattila-Sandholm T, von Wright A. Recovery of *Lactobacillus rhamnosus* GG from human colonic biopsies. Lett Appl Microbiol. 1997;24(5):361-364. doi:10.1046/j.1472-765X.1997.00140.x.
 75. Guarino A, Berni Canani R, Immacolata Spagnuolo M, Albano F, Di Benedetto L. Oral bacterial therapy reduces the duration of symptoms and of viral excretion in children with mild diarrhoea. J Paed Gastro. 1998;25:516-519.
 76. Millar MR, Bacon C, Smith SL, Walke V, Hall MA. Enteral feeding of premature infants with *Lactobacillus* GG.. Arch Dis Child. 1993;69(5 Spec No):483-487. doi:10.1136/adc.69.5_Spec_No.483.
 77. Greenberg B. *Salmonella* suppression by known populations of bacteria in flies. J Bacteriol. 1969;99(3):629-635. doi:10.1128/JB.99.3.629-635.1969.
 78. Lièven-Le Moal V, Servin AL. The front line of enteric host defence against unwelcome intrusion of harmful microorganisms: mucins, antimicrobial peptides, and microbiota. Clin Microbiol Revs. 2006;19:315-337.
 79. Cossart P. Bacterial invasion: the paradigms of enteroinvasive pathogens. Sci. 2004;304(5668):242-248. doi:10.1126/science.1090124.
 80. Bibiloni R, Pérez PF, de Antoni GL. Will a high adhering capacity in a probiotic strain guarantee exclusion of pathogens from intestinal epithelia? Anaerobe. 1999;5(5):519-524. doi:10.1006/anae.1999.0198.
 81. Hirano J, Yoshida T, Sugiyama T, Koide N, Mori I, Yokochi T. The effect of *Lactobacillus rhamnosus* on enterohemorrhagic *Escherichia coli* infection of human intestinal cells in vitro. Microbiol Immunol. 2003;47(6):405-409. doi:10.1111/j.1348-0421.2003.tb03377.x.
 82. Tuomola EM, Ouwehand AC, Salminen S. The effect of probiotic bacteria on the adhesion of pathogens to human intestinal mucus. FEMS Immunol Med Microbiol. 1999;26(2):137-142. doi:10.1111/j.1574-695X.1999.tb01381.x.
 83. Hirn J, Nurmi E, Johansson T, Nuotio L. Long-term experience with competitive exclusion and salmonellas in Finland. Int J Food Microbiol. 1992;15(3-4):281-285. doi:10.1016/0168-1605(92)90059-C.

84. Le Bouguenec C. Adhesins and invasins of pathogenic *Escherichia coli*. *Int J Med Microbiol*. 2005;295(6--7):471–478. doi:10.1016/j.ijmm.2005.07.001.
85. Nesser JR, Granato D, Rouvet M, Servin A, Teneberg S, Karlsson KA. *Lactobacillus johnsonii* La1 shares carbohydrate-binding specificities with several enteropathogenic bacteria. *Glycobiol*. 2000;10(11):1193–1199. doi:10.1093/glycob/10.11.1193.
86. Mukai T, Asasaka T, Sato E, Mori K, Matsumoto M, Ohori H. Inhibition of binding of *Helicobacter pylori* to the glycolipid receptors by probiotic *Lactobacillus reuteri*. *FEMS Immunol Med Microbiol*. 2002;32:105–110.
87. Ofek I, Doyle RJ. Bacterial adhesion to cells and tissues. New York (USA): Chapman and Hall; 1994.
88. Coconnier MH, Bernet MF, Chauviere G, Servin AL. Adhering heat-killed human *Lactobacillus acidophilus* strain LB inhibits the process of pathogenicity of diarrhoeagenic bacteria in cultured human intestinal cells. *J Diarrhoeal Dis Res*. 1993;11:235–242.
89. Pretzer G, Snel J, Molenaar D, Wiersma A, Bron PA, Lambert J, de Vos WM, van der Meer R, Smits MA, Kleerebezem M. Biodiversity-based identification and functional characterization of the mannose-specific adhesin of *Lactobacillus plantarum*. *J Bacteriol*. 2005;187(17):6128–6136. doi:10.1128/JB.187.17.6128-6136.2005.
90. Bron PA, Meijer M, Bongers RS, de Vos WM, Kleerebezem M. Dynamics of competitive population abundance of *Lactobacillus plantarum* *ivi* gene mutants in faecal samples after passage through the gastrointestinal tract of mice. *J Appl Microbiol*. 2007;103:1424–1434.
91. Mack DR, Ahrne S, Hyde L, Wei S, Hollingsworth MA. Extracellular MUC3 mucin secretion follows adherence of *Lactobacillus* strains to intestinal epithelial cells in vitro. *Gut*. 2003;52(6):827–833. doi:10.1136/gut.52.6.827.
92. Buck BL, Altermann E, Svingerud T, Klaenhammer TR. Functional analysis of putative adhesion factors in *Lactobacillus acidophilus* NCFM. *Appl Environ Microbiol*. 2005;71(12):8344–8351. doi:10.1128/AEM.71.12.8344-8351.2005.
93. van Pijkeren JP, Canchaya C, Ryan KA, Li Y, Claesson MJ, Sheil B, Steidler L, O'Mahony L, Fitzgerald GF, van Sinderen D, et al. Comparative and functional analysis of sortase-dependent proteins in the predicted secretome of *Lactobacillus salivarius* UCC118. *Appl Environ Microbiol*. 2006;72(6):4143–4153. doi:10.1128/AEM.03023-05.
94. Walter J, Chagnaud P, Tannock GW, Loach DM, Dal Bello F, Jenkinson HF, Hammes WP, Hertel C. A high-molecular-mass surface protein (Lsp) and methionine sulfoxide reductase B (MsrB) contribute to the ecological performance of *Lactobacillus reuteri* in the murine gut. *Appl Environ Microbiol*. 2005;71(2):979–986. doi:10.1128/AEM.71.2.979-986.2005.
95. Heinemann C, Vlieg JETV, Janssen DB, Busscher HJ, van der Mei HC, Reid G. Purification and characterization of a surface-binding protein from *Lactobacillus fermentum* RC-14 that inhibits adhesion of *Enterococcus faecalis* 1131. *FEMS Microbiol Lett*. 2000;190(1):177–180. doi:10.1111/j.1574-6968.2000.tb09282.x.
96. Chen XY, Xu JJ, Shuai JB, Chen JS, Zhang ZF, Fang WH. The S-layer proteins of *Lactobacillus crispatus* strain ZJ001 is responsible for competitive exclusion against *Escherichia coli* O157: H7 and *Salmonella typhimurium*. *Int J Food Microbiol*. 2007;115(3):307–312. doi:10.1016/j.ijfoodmicro.2006.11.007.
97. Johnson-Henry KC, Hagen KE, Gordonpour M, Tompkins TA, Sherman PM. Surface-layer protein extracts from *Lactobacillus helveticus* inhibit enterohaemorrhagic *Escherichia coli* O157: h7 adhesion to epithelial cells. *Cell Microbiol*. 2007;9(2):356–367. doi:10.1111/j.1462-5822.2006.00791.x.
98. Boekhorst J, De Been MWHJ, Kleerebezem M, Siezen RJ. Genome-wide detection and analysis of cell wall-bound proteins with LPxTG-like sorting motifs. *J Bacteriol*. 2005;187(14):4928–4934. doi:10.1128/JB.187.14.4928-4934.2005.
99. Spirig T, Weine EM, Clubb RT. Sortase enzymes in Gram-positive bacteria. *Mol Microbiol*. 2011;82(5):1044–1059. doi:10.1111/j.1365-2958.2011.07887.x.
100. Clancy KW, Melvin JA, McCafferty DG. Sortase transpeptidases: insights into mechanism, substrate specificity, and inhibition. *Biopolymers*. 2010;94(4):385–396. doi:10.1002/bip.21472.
101. Navarre WW, Schneewind O. Surface proteins of Gram-positive bacteria and mechanisms of their targeting to the cell wall envelope. *Microbiol Mol Biol Rev*. 1999;63:174–229.
102. Barnett TC, Patel AR, Scott JR. A novel sortase, SrtC2, from *Streptococcus pyogenes* anchors a surface protein containing a QVPTGV motif to the cell wall. *J Bacteriol*. 2004;186(17):5865–5875. doi:10.1128/JB.186.17.5865-5875.2004.
103. Candela M, Perna F, Carnevali P, Vitali B, Ciati R, Gionchetti P, Rizzello F, Campieri M, Brigidi P. Interaction of probiotic *Lactobacillus* and *Bifidobacterium* strains with human intestinal epithelial cells: adhesion properties, competition against enteropathogens and modulation of IL-8 productions. *Int J Food Microbiol*. 2008;125(3):286–292. doi:10.1016/j.ijfoodmicro.2008.04.012.
104. Walter J. Ecological role of lactobacilli in the gastrointestinal tract: implications for fundamental and biomedical research. *Appl Environ Microbiol*. 2008;74(16):4985–4996. doi:10.1128/AEM.00753-08.
105. Rossi M, Corradini C, Amaretti A, Nicolini M, Pompei A, Zanoni S, Matteuzi D. Fermentation of fructooligosaccharides and inulin by bifidobacteria: a comparative study in pure and faecal cultures. *Appl Environ Microbiol*. 2005;71(10):6150–6158. doi:10.1128/AEM.71.10.6150-6158.2005.
106. Lajis AFB. Biomanufacturing process for the production of bacteriocins from Bacillaceae family. *Bioresour Bioprocess*. 2020;7(1):8. doi:10.1186/s40643-020-0295-z.

107. van der Donk WA, Nair SK. Structure and mechanism of lanthipeptide biosynthetic enzymes. *Curr Opin Struct Biol*. 2014;29:58–66. doi:10.1016/j.sbi.2014.09.006.
108. Dicks LMT, Dreyer L, Smith C, van Staden AD. A review: the fate of bacteriocins in the human gastro-intestinal tract: do they cross the gut-blood barrier. *Front Microbiol*. 2018;9:2297.
109. Smaoui S, Elleuch L, Bejar W, Karray-Rebai I, Ayadi I, Jaouadi B, Mathieu F, Chouayekh H, Bejar S, Mellouli L. Inhibition of fungi and Gram-negative bacteria by bacteriocin BacTN635 produced by *Lactobacillus plantarum* sp. TN635. *Reproduction*. 2009;128:117–126.
110. Drider D, Bendali F, Naghmouchi K, Chikindas M. Bacteriocins: not only antimicrobial agents. *Probiot Antimicrob Prot*. 2016;8(4):177–182. doi:10.1007/s12602-016-9223-0.
111. Chikindas M, Weeks R, Drider D, Chistyakov V, Dicks LMT. Functions and emerging applications of bacteriocins. *Curr Opin Biotechnol*. 2017;49:23–28. doi:10.1016/j.copbio.2017.07.011.
112. Fujiwara S, Hashiba H, Hirota T, Forstner JF. Proteinaceous factor(s) in culture supernatant fluids of bifidobacteria which prevents the binding of enterotoxigenic *Escherichia coli* to gangliosylceramide. *Appl Environ Microbiol*. 1997;63(2):506–512. doi:10.1128/AEM.63.2.506-512.1997.
113. Fujiwara S, Hashiba H, Hirota T, Forstner JF. Inhibition of the binding of enterotoxigenic *Escherichia coli* Pb176 to human intestinal epithelial cell line HCT-8 by an extracellular protein fraction containing BIF of *Bifidobacterium longum* SBT2928: suggestive evidence of blocking of the binding receptor gangliosylceramide on the cell surface. *Int J Food Microbiol*. 2001;67:97–106.
114. Singh VP. Recent approaches in food bio-preservation. *Open Vet J*. 2018;8(1):104–111. doi:10.4314/ovj.v8i1.16.
115. De Kwaadsteniet M, Van Reenen C, Dicks LMT. Evaluation of nisin F in the treatment of subcutaneous skin infections, as monitored by using a bioluminescent strain of *Staphylococcus aureus*. *Prob Antimicrob Prot*. 2010;2(2):61–65. doi:10.1007/s12602-009-9017-8.
116. Van Reenen C, Chikindas M, Van Zyl W, Dicks LMT. Characterization and heterologous expression of a class IIa bacteriocin, plantaricin 423 from *Lactobacillus plantarum* 423, in *Saccharomyces cerevisiae*. *Int J Food Microbiol*. 2003;81(1):29–40. doi:10.1016/S0168-1605(02)00164-2.
117. Cotter PD, Ross RP, Hill C. Bacteriocins—a viable alternative to antibiotics? *Nat Rev Microbiol*. 2012;11(2):95–105. doi:10.1038/nrmicro2937.
118. Ratchaneewan A, Pipatsatitpong D. Isolation and characterization of a bacteriocin with anti-MRSA activity from *Bacillus* sp. strain WASM9-25M. *Thai J Pharmacol*. 2014;36:19–28.
119. Garg N, Oman TJ, Andrew Wang TS, De Gonzalo CV, Walker S, van der Donk WA. Mode of action and structure-activity relationship studies of geobacillin I. *J Antibiot*. 2014;67(1):133–136. doi:10.1038/ja.2013.112.
120. O’Shea EF, Cotter PD, Stanton C, Ross RP, Hill C. Production of bioactive substances by intestinal bacteria as a basis for explaining probiotic mechanisms: bacteriocins and conjugated linoleic acid. *Int J Food Microbiol*. 2012;152(3):189–205. doi:10.1016/j.ijfoodmicro.2011.05.025.
121. Izadpanah A, Gallo RL. Antimicrobial peptides. *J Acad Dermatol*. 2005;52(3):381–390. doi:10.1016/j.jaad.2004.08.026.
122. Islam MR, Nagao JI, Zendo T, Sonomoto K. Antimicrobial mechanism of lantibiotics. *Biochem Soc Trans*. 2012;40(6):1528–1533. doi:10.1042/BST20120190.
123. Nishie M, Nagao JI, Sonomoto K. Antibacterial peptides “bacteriocins”: an overview of their diverse characteristics and applications. *Biocontrol Sci*. 2012;17(1):1–16. doi:10.4265/bio.17.1.
124. Bierbaum G, Sahl HG. Lantibiotics: mode of action, biosynthesis and bioengineering. *Curr Pharm Biotechnol*. 2009;10(1):2–18. doi:10.2174/138920109787048616.
125. Yildirim Z, Johnson MG. Characterization and antimicrobial spectrum of bifidocin B, a bacteriocin produced by *Bifidobacterium bifidum* NCFB 1454. *J Food Protect*. 1998;61:47–51.
126. Yildirim Z, Winters DK, Johnson MG. Purification, amino acid sequence and mode of action of bifidocin B produced by *Bifidobacterium bifidum* NCFB 1454. *J Appl Microbiol*. 1999;86(1):45–54. doi:10.1046/j.1365-2672.1999.00629.x.
127. Kim TS, Hur JW, Yu MA, Cheigh CI, Kim KN, Hwang JK, Pyun YR. Antagonism of *Helicobacter pylori* by bacteriocins of lactic acid bacteria. *J Food Protect*. 2003;66(1):3–12. doi:10.4315/0362-028X-66.1.3.
128. Stevens KA, Sheldon BW, Klapes NA, Klaenhammer TR. Nisin treatment for inactivation of *Salmonella* species and other gram-negative bacteria. *Appl Environ Microbiol*. 1991;57(12):3613–3615. doi:10.1128/AEM.57.12.3613-3615.1991.
129. Svetoch EA, Levchuk VP, Pokhilenko VD, Eruslanov BV, Mitsevich EV, Mitsevich IP, Perelygin VV, Stephanshin YG, Stern NJ. Inactivating methicillin-resistant *Staphylococcus aureus* and other pathogens by use of bacteriocins OR-7 and E 50-52. *J Clin Microbiol*. 2008;46(11):3863–3865. doi:10.1128/JCM.01081-08.
130. Morency H, Mota-Meira M, Lapointe G, Lacroix C, Lavoie MC. Comparison of the activity spectra against pathogens of bacterial strains producing a mutacin or a lantibiotic. *Can J Microbiol*. 2001;47(4):322–331. doi:10.1139/w01-013.
131. Ghodbane H, Elaidi S, Sabatie J, Achour S, Regaya JI. Bacteriocins active against multi-resistant Gram-negative bacteria implicated in nosocomial infections. *Infec Disorders-Drug Targets*. 2015;15(1):2–12. doi:10.2174/1871526514666140522113337.
132. Prudêncio CV, Dos Santos MT, Vanetti MCD. Strategies for the use of bacteriocins in Gram-negative bacteria: relevance in food microbiology. *J Food Sci Technol*. 2015;52:5408–5417.

133. Lim KB, Balolong MP, Kim SH, Oh JK, Lee JY, Kang DK. Isolation and characterization of a broad spectrum bacteriocin from *Bacillus amyloliquefaciens* RX7. *Biomed Res Int*. 2016;2016:8521476.
134. Tiwari SK, Noll KS, Cavera VL, Chikindas ML, Macfarlane GT. Improved antimicrobial activities of synthetic-hybrid bacteriocins designed from enterocin E50-52 and pediocin PA-1. *Appl Environ Microbiol*. 2014;81(5):1661–1667. doi:10.1128/AEM.03477-14.
135. Duclohier H, Molle SG. Antimicrobial peptide magainin I from *Xenopus* skin forms anion-permeable channels in planar lipid bilayers. *Biophys J*. 1989;56(5):1017–1021. doi:10.1016/S0006-3495(89)82746-8.
136. Kagan BL, Selsted ME, Ganz T, Lehrer RI. Antimicrobial defensin peptides form voltage-dependent ion-permeable channels in planar lipid bilayer membranes. *Proc Natl Acad Sci USA*. 1990;87(1):210–216. doi:10.1073/pnas.87.1.210.
137. Simanova MP, Laukova A, Chrastinova L, Stropfova V, Faix S, Vasilkova Z, Ondruska L, Jurcik R, Rafay J. *Enterococcus faecium* CCM7420, bacteriocin PPB CCM7420 and their effect in the digestive tract of rabbits. *Czech J Anim Sci*. 2009;54(No. 8):376–386. doi:10.17221/1659-CJAS.
138. Laukova A, Guba P, Nemcova R, Vasilkova Z. Reduction of *Salmonella* in gnotobiotic Japanese quails caused by the enterocin A-producing EK13 strain of *Enterococcus faecium*. *Vet Res Commun*. 2003;27(4):275–280. doi:10.1023/A:1024027923824.
139. Laukova A, Stropfova V, Skrikvanova V, Volek Z, Jindrichova E, Marounek M. Bacteriocin-producing strain of *Enterococcus faecium* EK13 with probiotic characteristics and its application in the digestive tract of rabbits. *Biol Bratisl*. 2006;61(6):779–782. doi:10.2478/s11756-006-0191-9.
140. Van Staden AD, Heunis T, Smith C, Deane SM, Dicks LMT. Efficacy of lantibiotic treatment of *staphylococcus aureus*-induced skin infections, monitored by in vivo bioluminescent imaging. *Antimicrob Agents Chemother*. 2016;60(7):3948–3955. doi:10.1128/AAC.02938-15.
141. Baindara P, Chaudhry V, Mittal G, Liao LM, Matos CO, Franco OL, Patil PB, Korpole S. Characterization of the antimicrobial peptide penisin, a class Ia novel lantibiotic from *Paenibacillus* sp. strain A3. *Antimicrob Agents Chemother*. 2015;60(1):580–591. doi:10.1128/AAC.01813-15.
142. Svetoch AE, Eruslanov BV, Pereglygin VV, Mitsevich EV, Mitsevich IP, Borzenkov VN, Levchuk VP, Svetoch OE, Kovalev YN, Stephanshin YG, et al. Diverse antimicrobial killing by *enterococcus faecium* E 50-52 bacteriocin. *J Agric Food Chem*. 2008;6(6):1942–1948. doi:10.1021/jf073284g.
143. Heunis TD, Smith C, Dicks LMT. Evaluation of a nisin-eluting nanofiber scaffold to treat *staphylococcus aureus*-induced skin infections in mice. *Antimicrob Agents Chemother*. 2013;57(8):3928–3935. doi:10.1128/AAC.00622-13.
144. Fernández L, Delgado S, Herrero H, Maldonado A, Rodríguez JM. The bacteriocin nisin, an effective agent for the treatment of staphylococcal mastitis during lactation. *J Hum Lact*. 2008;24(3):311–316. doi:10.1177/0890334408317435.
145. Turner S, Loveand R, Lyons K. An in vitro investigation of the antibacterial effect of nisin in root canals and canal wall radicular dentine. *Int Endod J*. 2004;37(10):664–671. doi:10.1111/j.1365-2591.2004.00846.x.
146. Hillman J, Mo J, McDonell E, Cvitkovitch D, Hillman C. Modification of an effector strain for replacement therapy of dental caries to enable clinical safety trials. *J Appl Microbiol*. 2007;102(5):1209–1219. doi:10.1111/j.1365-2672.2007.03316.x.
147. Dobson A, O'Connor P, Cotter P, Ross R, Hill C. Impact of the broad-spectrum antimicrobial peptide, lacticin 3147, on *Streptococcus mutans* growing in a biofilm and in human saliva. *J Appl Microbiol*. 2011;111(6):515–523. doi:10.1111/j.1365-2672.2011.05153.x.
148. Legala OE, Yassi H, Pflugmacher S, Neubauer P. Pharmacological and pharmacokinetic properties of lanthipeptides undergoing clinical studies. *Biotechnol Lett*. 2017;39(4):473–482. doi:10.1007/s10529-016-2279-9.
149. Legala OE, Neubauer P. Lanthipeptides: chemical synthesis versus in vivo biosynthesis as tools for pharmaceutical production. *Microb Cell Fact*. 2016;15(1):97. doi:10.1186/s12934-016-0502-y.
150. Lopetuso LR, Giorgio ME, Saviano A, Scalfaferrri F, Gasbarrini A, Cammarota G. Bacteriocins and bacteriophages: therapeutic weapons for gastrointestinal diseases? *Int J Molec Sci*. 2019;20(1):183. doi:10.3390/ijms20010183.
151. Sandiford SK. Perspectives on lantibiotic discovery-Where have we failed and what improvements are required? *Exp Opin Drug Disc*. 2015;10(4):315–320. doi:10.1517/17460441.2015.1016496.
152. Bomba A, Nemcova R, Kastel R, Herich R, Pataky J, Cizek M. Interactions of *Lactobacillus* spp. and enteropathogenic *Escherichia coli* under in vitro and in vivo conditions. *Vet Med*. 1996;41:155–158.
153. Bomba A, Nemcova R, Gancarcikova S, Herich R, Kastel R. Potentiation of the effectiveness of *Lactobacillus casei* in the prevention of *E. coli* induced diarrhoea in conventional and gnotobiotic pigs. *Adv Exp Med Biol*. 1999;473:185–190.
154. Malago JJ, Koninkx JFJG. Probiotic-pathogen interactions and enteric cytoprotection. In: Malago JJ, Koninkx JFJG, Marinsek-Logar R, editors. *Probiotic bacteria and enteric infections*. Dordrecht (Netherlands): Springer; 2011. p. 289–311.
155. Silva M, Jacobus N, Deneke C, Gorbach SL. Antimicrobial substance from a human *Lactobacillus* strain.. *Antimicrob Agents Chemother*. 1987;31(8):1231–1233. doi:10.1128/AAC.31.8.1231.

156. Van De Guchte M, Ehrlich SD, Maguin E. Production of growth-inhibiting factors by *Lactobacillus delbrueckii*. *J Appl Microbiol.* 2001;91(1):147–153. doi:10.1046/j.1365-2672.2001.01369.x.
157. Bernet-Camard MF, Lievin V, Brassart D, Neeser JR, Servin AL, Hudault S. The human *Lactobacillus acidophilus* strain LA1 secretes a nonbacteriocin antibacterial substance(s) active in vitro and in vivo.. *Appl Environ Microbiol.* 1997;63(7):2747–2753. doi:10.1128/AEM.63.7.2747-2753.1997.
158. O’Riordan K, Fitzferald GF. Evaluation of Bifidobacteria for the evaluation of antimicrobial compounds and assessment of performance in cottage cheese at refrigeration temperature. *J Appl Microbiol.* 1998;85(1):103–114. doi:10.1046/j.1365-2672.1998.00474.x.
159. Bielecka M, Biedrycka E, Biedrycka E, Smoragiewicz W, Smieszek M. Interaction of *Bifidobacterium* and *Salmonella* during associated growth. *Int J Food Microbiol.* 1998;45(2):151–155. doi:10.1016/S0168-1605(98)00150-0.
160. Lee YJ, Yu WK, Heo TR. Identification and screening for antimicrobial activity against *clostridium difficile* of *bifidobacterium* and *lactobacillus* species isolated from healthy infant faeces. *Int J Antimicrob Agents.* 2003;21(4):340–346. doi:10.1016/S0924-8579(02)00389-8.
161. Van den Bergh PA. Lactic acid bacteria, their metabolic products and interference with microbial growth. *FEMS Microbiol Rev.* 1993;12(1–3):221–238. doi:10.1111/j.1574-6976.1993.tb00020.x.
162. Alakomi HL, Skytta E, Saarela M, Mattila-Sandholm T, Latva-Kala K, Helander IM. Lactic acid permeabilizes gram-negative bacteria by disrupting the outer membrane. *Appl Environ Microbiol.* 2000;66(5):2001–2005. doi:10.1128/AEM.66.5.2001-2005.2000.
163. Russell JB, Diez-Gonzalez F. The effects of fermentation acids on bacterial growth. *Adv Microb Physiol.* 1998;39:205–234.
164. De Keersmaecker SC, Verhoeven TL, Desair J, Marchal K, Vanderleyden J, Nagy I. Strong antimicrobial activity of *Lactobacillus rhamnosus* GG against *Salmonella typhimurium* is due to accumulation of lactic acid. *FEMS Microbiol Lett.* 2006;259(1):89–96. doi:10.1111/j.1574-6968.2006.00250.x.
165. Lehto EM, Salminen SJ. Inhibition of *Salmonella typhimurium* adhesion to Caco-2 cell cultures by *Lactobacillus* strain GG spent culture supernate: only a pH effect? *FEMS Immunol Med Microbiol.* 1997;18(2):125–132. doi:10.1111/j.1574-695X.1997.tb01037.x.
166. Durant JA, Corrier DE, Ricke SC. Short-chain volatile fatty acids modulate the expression of the *hilA* and *invF* genes of *Salmonella typhimurium*. *J Food Prot.* 2000;63(5):573–578. doi:10.4315/0362-028X-63.5.573.
167. Brashears MM, Reilly SS, Gilliland SE. Antagonistic action of cells of *Lactobacillus lactis* toward *Escherichia coli* O157:h7on refrigerated raw chicken meat. *J Food Prot.* 1998;61(2):166–170. doi:10.4315/0362-028X-61.2.166.
168. Ogawa M, Shimizu K, Nomoto K, Tanaka R, Hamabata T, Yamasaki S, Takeda T, Takeda Y. Inhibition of in vitro growth of Shiga toxin-producing *Escherichia coli* O157: h7by probiotic *Lactobacillus* strains due to production of lactic acid. *Int J Food Microbiol.* 2001;68(1–2):135–140. doi:10.1016/S0168-1605(01)00465-2.
169. Midolo PD, Lambert JR, Hull R, Luo F. In vitro inhibition of *Helicobacter pylori* NCTC 11637 by organic acids and lactic acid bacteria. *J Appl Bacteriol.* 1995;79(4):475–479. doi:10.1111/j.1365-2672.1995.tb03164.x.
170. Bhatia SJ, Kochar N, Abraham P, Nair NG, Mehta AP. *Lactobacillus acidophilus* inhibits growth of *Campylobacter pylori* in vitro.. *J Clin Microbiol.* 1989;27(10):2328–2330. doi:10.1128/JCM.27.10.2328-2330.1989.
171. Strus M, Pakosz K, Gosciniak H, Przondo-Mordarska A, Rozynek E, Pituch H, Meisel-Mikolajczyk F, Heczko PB. Antagonistic activity of *Lactobacillus* bacteria strains against anaerobic gastrointestinal tract pathogens (*Helicobacter pylori*, *Campylobacter coli*, *Campylobacter jejuni*, *Clostridium difficile*). *Med Dosw Mikrobiol.* 2001;53:133–142.
172. Fayol-Messaoudi D, Berger CN, Coconnier-Polter MH, Lievin-Le Moal V, Servin AL. pH-, lactic acid-, and non-lactic acid-dependent activities of probiotic lactobacilli against *Salmonella enterica* serovar Typhimurium. *Appl Environ Microbiol.* 2005;71(10):6008–6013. doi:10.1128/AEM.71.10.6008-6013.2005.
173. Botteaux A, Hoste C, Dumont JE, Van Sande J, Allaoui A. Potential role of Noxes in the protection of mucosae: H₂O₂ as a bacterial repellent. *Microbes Infect.* 2009;11(5):537–544. doi:10.1016/j.micinf.2009.02.009.
174. Knaus UG, Hertzberger R, Pircalabioru GG, Yousefi SPM, Dos Santos FB. Pathogen control at the intestinal mucosa - H₂O₂ to the rescue. *Gut Microbes.* 2017;8(1):67–74. doi:10.1080/19490976.2017.1279378.
175. Alvarez LA, Kovacic L, Rodriguez J, Gosemann JH, Kubica M, Pircalabioru GG, Friedmacher F, Cean A, Ghişu A, Sărăndan MB, et al. NADPH oxidase-derived H₂O₂ subverts pathogen signalling by oxidative phosphorylation conversion to PB-DOPA. *Proc Natl Acad Sci USA.* 2016;113(37):10406–10411. doi:10.1073/pnas.1605443113.
176. Pridmore RD, Pittet AC, Praplan F, Cavadini C. Hydrogen peroxide production by *Lactobacillus johnsonii* NCC 533 and its role in anti-*Salmonella* activity. *FEMS Microbiol Lett.* 2008;283(2):210–215. doi:10.1111/j.1574-6968.2008.01176.x.
177. Kawasaki S, Satoh T, Todorok M, Niimura Y. b-type dihydroorotate dehydrogenase is purified as a H₂O₂-forming NADH oxidase from *Bifidobacterium bifidum*. *Appl Environ Microbiol.* 2009;75(3):629–636. doi:10.1128/AEM.02111-08.
178. Hertzberger R, Arents J, Dekker HL, Pridmore RD, Gysler C, Kleerebezem M, de Mattos MJT. H₂O₂ production in species of the *Lactobacillus acidophilus* group: a central role for a novel NADH-Dependent

- Flavin reductase. *Appl Environ Microbiol.* 2014;80:2229–2239.
179. Marty-Teyssset C, de la Torre F, Garel J. Increased production of hydrogen peroxide by *Lactobacillus delbrueckii* subsp. *bulgaricus* upon aeration: involvement of an NADH oxidase in oxidative stress. *Appl Environ Microbiol.* 2000;66(1):262–267. doi:10.1128/AEM.66.1.262-267.2000.
 180. Eschenbach DA, Davick PR, Williams BL, Klebanoff SJ, Young-Smith K, Critchlow CM, Holmes KK. Prevalence of hydrogen peroxide-producing *Lactobacillus* species in normal women and women with bacterial vaginosis. *J Clin Microbiol.* 1989;27(2):251–256. doi:10.1128/JCM.27.2.251-256.1989.
 181. Ocana VS, Pesce De Ruiz Holgado AA, Nader-Macias ME. Selection of vaginal H₂O₂-generating *Lactobacillus* species for probiotic use. *Curr Microbiol.* 1999;38(5):279–284. doi:10.1007/PL00006802.
 182. Ocana VS, De Ruiz Holgado AA, Nader-Macias ME. Growth inhibition of *Staphylococcus aureus* by H₂O₂-producing *Lactobacillus paracasei* subsp. *paracasei* isolated from the human vagina. *FEMS Immunol Med Microbiol.* 1999;23(2):87–92. doi:10.1016/S0928-8244(98)00116-3.
 183. Hawes SE, Hillier SL, Benedetti J, Stevens CE, Koutsky LA, Wolner-Hanssen P, Holmes KK. Hydrogen peroxide-producing lactobacilli and acquisition of vaginal infections. *J Infect Dis.* 1996;174(5):1058–1063. doi:10.1093/infdis/174.5.1058.
 184. Turrone F, Ventura M, Butto LF, Duranti S, O'Toole PW, Motherway MO, Sinderen D. Molecular dialogue between the human gut microbiota and the host: a *Lactobacillus* and *Bifidobacterium* perspective. *Cell Mol Life Sci.* 2014;71:183–203.
 185. Ahmed E, Holmström SJM. Siderophores in environmental research: roles and applications. *Microb Biotechnol.* 2014;7(3):196–208. doi:10.1111/1751-7915.12117.
 186. O'Sullivan DJ Isolated bifidobacteria that produce siderophores which inhibit growth of *Lactococcus lactis*. 2014. US patent 6746672B2.
 187. Vazquez-Gutierrez P, de Wouters T, Werder J, Chassard C, Lacroix C. High iron-sequestering Bifidobacteria inhibit enteropathogen growth and adhesion to intestinal epithelial cells in vitro. *Front Microbiol.* 2016;7:1480. doi:10.3389/fmicb.2016.01480.
 188. Rodrigues L, Banat IM, Teixeira J, Oliveira R. Biosurfactants: potential applications in medicine. *J Antimicrob Chemother.* 2006;57(4):609–618. doi:10.1093/jac/dkl024.
 189. Saharan BS, Sahu RK, Sharma D. A review on biosurfactants: fermentation, current developments and perspectives. *Genet Eng Biotechnol J.* 2012;1:1–14.
 190. Rodrigues LR, Teixeira JA, Van der Mei HC, Oliveira R. Physicochemical and functional characterization of a biosurfactant produced by *Lactococcus lactis* 53. *Colloids and Surf B Biointerfaces.* 2006;49(1):79–86. doi:10.1016/j.colsurfb.2006.03.003.
 191. Ciandrini E, Campana R, Casettari L, Perinelli DR, Faqioli L, Manti A, Palmieri GF, Papa S, Baffone W. Characterization of biosurfactants produced by *Lactobacillus* spp. and their activity against oral streptococci. *Appl Microbiol Biotechnol.* 2016;100(15):6767–6777. doi:10.1007/s00253-016-7531-7.
 192. Rodrigues LR, Teixeira JA, van der Mei HC, Oliveira R. Isolation and partial characterization of a biosurfactant produced by *Streptococcus thermophilus* A. *Colloids Surf B Biointerfaces.* 2006;53(1):105–112. doi:10.1016/j.colsurfb.2006.08.009.
 193. Saravanakumari P, Mani K. Structural characterization of a novel xylolipid biosurfactant from *Lactococcus lactis* and analysis of antibacterial activity against multi-drug resistant pathogens. *Bioresour Technol.* 2010;101(22):8851–8854. doi:10.1016/j.biortech.2010.06.104.
 194. Duarte C, Gudiña EJ, Lima CF, Rodrigues LR. Effects of biosurfactants on the viability and proliferation of human breast cancer cells. *AMB Express.* 2014;4(1):40. doi:10.1186/s13568-014-0040-0.
 195. Velraeds MM, Van Der Mei HC, Reid G, Busscher HJ. Inhibition of initial adhesion of uropathogenic *Enterococcus faecalis* by biosurfactants from *Lactobacillus* isolates. *Appl Environ Microbiol.* 1996;62(6):1958–1963. doi:10.1128/AEM.62.6.1958-1963.1996.
 196. Reid G, Cook RL, Bruce AW. Examination of strains of lactobacilli for properties that may influence bacterial interference in the urinary tract. *J Urol.* 1987;138(2):330–335. doi:10.1016/S0022-5347(17)43137-5.
 197. Goma E. Antimicrobial and anti-adhesive properties of biosurfactant produced by lactobacilli isolates, biofilm formation and aggregation ability. *J Gen Appl Microbiol.* 2013;59(6):425–436. doi:10.2323/jgam.59.425.
 198. Sharma D, Saharan BS. Simultaneous production of biosurfactants and bacteriocins by probiotic *Lactobacillus casei* MRTL3. *Int J Microbiol.* 2014;2014:698713.
 199. Weinstein DL, O'Neill BL, Hone DM, Metcalf ES. Differential early interactions between *Salmonella enterica* serovar Typhimurium and two other pathogenic *Salmonella* serovars with intestinal epithelial cells. *Infect Immun.* 1998;66(5):2310–2318. doi:10.1128/IAI.66.5.2310-2318.1998.
 200. Fujiwara S, Hashiba H, Hirota T, Forstner JF. Purification and characterization of a novel protein produced by *Bifidobacterium longum* SBT2928 that inhibits the binding of enterotoxigenic *Escherichia coli* Pb176 (CFA/II) to gangliosylceramide. *Appl Microbiol.* 1999;86(4):615–621. doi:10.1046/j.1365-2672.1999.00705.x.
 201. Lievin V, Peiffer I, Hudault S, Rochat F, Brassart D, Neeser JR, Servin AL. *Bifidobacterium* strains from resident infant human gastrointestinal microflora exert antimicrobial activity. *Gut.* 2000;47(5):646–652. doi:10.1136/gut.47.5.646.

202. Ohland CL, MacNaughton WK. Probiotic bacteria and intestinal epithelial barrier function. *Am J Physiol Gastrointest Liver Physiol.* 2010;298(6):G807–G819. doi:10.1152/ajpgi.00243.2009.
203. McGuckin MA, Eri R, Simms LA, Florin TH, Radford-Smith G. Intestinal barrier dysfunction in inflammatory bowel diseases. *Inflam Bowel Dis.* 2009;15(1):100–113. doi:10.1002/ibd.20539.
204. Hooper LV, Macpherson AJ. Immune adaptations that maintain homeostasis with the intestinal microbiota. *Nat Rev Immunol.* 2010;10(3):159–169. doi:10.1038/nri2710.
205. Schneeberger EE, Lynch RD. The tight junction: a multifunctional complex. *Am J Physiol Cell Physiol.* 2004;286(6):C1213–C1228. doi:10.1152/ajpcell.00558.2003.
206. Chichlowski M, Hale LP. Bacterial-mucosal interactions in inflammatory bowel disease - an alliance gone bad. *Am J Physiol Gastrointest Liver Physiol.* 2008;295(6):G1139–G1149. doi:10.1152/ajpgi.90516.2008.
207. Sartor RB. Mechanisms of disease: pathogenesis of Crohn's disease and ulcerative colitis. *Nat Clin Pract Gastroenterol Hepatol.* 2006;3(7):390–407. doi:10.1038/ncpgasthep0528.
208. Moncada DM, Kammanadiminti SJ, Chadee K. Mucin and Toll-like receptors in host defence against intestinal parasites. *Trends Parasitol.* 2003;19(7):305–311. doi:10.1016/S1471-4922(03)00122-3.
209. Corfield AP, Myerscough N, Longman R, Sylvester P, Arul S, Pignatelli M. Mucins and mucosal protection in the gastrointestinal tract: new prospects for mucins in the pathology of gastrointestinal disease. *Gut.* 2000;47(4):589–594. doi:10.1136/gut.47.4.589.
210. Mattar AF, Teitelbaum DH, Drongowski RA, Yongyi F, Harmon CM, Coran AG. Probiotics up-regulate MUC-2 mucin gene expression in a Caco-2 cell-culture model. *Pediatr Surg Int.* 2002;18(7):586–590. doi:10.1007/s00383-002-0855-7.
211. Mack DR, Michail S, Wei S, McDougall L, Hollingsworth MA. Probiotics inhibit enteropathogenic *E. coli* adherence in vitro by inducing intestinal mucin gene expression. *Am J Physiol.* 1999;276:G941–G950.
212. Caballero-Franco C, Keller K, Simone C, Chadee K. The VSL#3 probiotic formula induces mucin gene expression and secretion in colonic epithelial cells. *Am J Physiol Gastrointest Liver Physiol.* 2007;292(1):G315–G322. doi:10.1152/ajpgi.00265.2006.
213. Reid G, McGroarty JA, Angotti R, Cook RL. Lactobacillus inhibitor production against *Escherichia coli* and coaggregation ability with uropathogens. *Canadian J Microbiol.* 1988;34(3):344–351. doi:10.1139/m88-063.
214. Callado MC, Meriluoto J, Salminen S. Role of commercial probiotic strains against human pathogen adhesion to intestinal mucus. *Lett Appl Microbiol.* 2007;45:454–460.
215. Kos B, Susković J, Vuković S, Simpraga M, Frece J, Matosić S. Adhesion and aggregation ability of probiotic strain *Lactobacillus acidophilus* M92. *J Appl Microbiol.* 2003;94(6):981–987. doi:10.1046/j.1365-2672.2003.01915.x.
216. Kolenbrander PE. Oral microbial communities: biofilms, interactions, and genetic systems. *Annu Rev Microbiol.* 2000;54(1):413–439. doi:10.1146/annurev.micro.54.1.413.
217. Mastromarino P, Brigidi P, Macchia S, Maggi L, Pirovano F, Trinchieri V, Conte U, Matteuzzi D. Characterization and selection of vaginal *Lactobacillus* strains for the preparation of vaginal tablets. *J Appl Microbiol.* 2002;93(5):884–893. doi:10.1046/j.1365-2672.2002.01759.x.
218. Janković T, Frece J, Abram M, Gobin I. Aggregation ability of potential probiotic *Lactobacillus plantarum* strains. *Int J Sanitary Eng Res.* 2012;6:19–24.
219. Schachtsiek M, Hammes WP, Hertel C. Characterization of *Lactobacillus coryniformis* DSM 20001T surface protein Cpf mediating coaggregation with and aggregation among pathogens. *Appl Environ Microbiol.* 2004;70(12):7078–7085. doi:10.1128/AEM.70.12.7078-7085.2004.
220. Mizuno K, Mizuno M, Yamauchi M, Takemura AJ, Romero VM, Morikawa K. Adjacent-possible ecological niche: growth of *Lactobacillus* species co-cultured with *Escherichia coli* in a synthetic minimal medium. *Scient Rep.* 2017;7(1):12880. doi:10.1038/s41598-017-12894-3.
221. Spencer RJ, Chesson A. The effect of *Lactobacillus* spp. on the attachment of enterotoxigenic *Escherichia coli* to isolated porcine enterocytes. *J Appl Bacteriol.* 1994;77(2):215–220. doi:10.1111/j.1365-2672.1994.tb03066.x.
222. Del Re B, Sgorbati B, Miglioli M, Palenzona D. Adhesion, autoaggregation and hydrophobicity of 13 strains of *Bifidobacterium longum*. *Letters in Appl Microbiol.* 2000;31(6):438–442. doi:10.1046/j.1365-2672.2000.00845.x.
223. Boris S, Suárez JE, Barbés C. Characterization of the aggregation promoting factor from *Lactobacillus gasseri*, avaginal isolate. *J Appl Microbiol.* 1997;83(4):413–420. doi:10.1046/j.1365-2672.1997.00250.x.
224. Sillanpää J, Martinez B, Antikainen J, Toba T, Kalkkinen N, Tankka S, Lounatmaa K, Keränen L, Höök M, Westerlund-Wikström B, et al. Characterization of the collagen-binding S-layer protein CbsA of *Lactobacillus crispatus*. *J Bacteriol.* 2000;182(22):6440–6450. doi:10.1128/JB.182.22.6440-6450.2000.
225. Roos S, Lindgren S, Jonsson H. Autoaggregation of *Lactobacillus reuteri* is mediated by a putative DEAD-box helicase. *Mol Microbiol.* 1999;32(2):427–436. doi:10.1046/j.1365-2958.1999.01363.x.
226. Jankovic I, Ventura M, Meylan V, Rouvet M, Elli M, Zink R. Contribution of aggregation-promoting factor to maintenance of cell shape in *Lactobacillus gasseri* 4B2. *J Bacteriol.* 2003;185(11):3288–3296. doi:10.1128/JB.185.11.3288-3296.2003.
227. Reid G, Sobeli JD. Bacterial adherence in the pathogenesis of urinary tract infection: a review. *Rev Infect Dis.* 1987;9(3):470–487. doi:10.1093/clinids/9.3.470.
228. Josenhans C, Suerbaum S. The role of motility as a virulence factor in bacteria. *Int J Med Microbiol.* 2002;291(8):605–614. doi:10.1078/1438-4221-00173.

229. Stecher B, Hapfelmeier S, Müller C, Kremer M, Stallmach T, Hardt W. Flagella and chemotaxis are required for efficient induction of *Salmonella enterica* serovar Typhimurium colitis in streptomycin-pretreated mice. *Infect Immun*. 2004;72(7):4138–4150. doi:10.1128/IAI.72.7.4138-4150.2004.
230. Dibb-Fuller MP, Allen-Vercoe E, Thorns CJ, Woodward MJ. Fimbriae- and flagella-mediated association with and invasion of cultured epithelial cells by *Salmonella enteritidis*. *Microbiol*. 1999;145(5):1023–1031. doi:10.1099/13500872-145-5-1023.
231. Jones BD, Lee CA, Falkow S. Invasion by *Salmonella typhimurium* is affected by the direction of flagellar rotation.. *Infect Immun*. 1992;60(6):2475–2480. doi:10.1128/IAI.60.6.2475-2480.1992.
232. Schmitt CK, Ikeda JS, Darnell SC, Watson PR, Bispham J, Wallis TS, Weinstein DL, Metcalf ES, O'Brien AD. Absence of all components of the flagellar export and synthesis machinery differentially alters virulence of *Salmonella enterica* serovar Typhimurium in models of typhoid fever, survival in macrophages, tissue culture invasiveness, and calf enterocolitis. *Infect Immun*. 2001;69(9):5619–5625. doi:10.1128/IAI.69.9.5619-5625.2001.
233. Forbes SJ, Eschmann M, Mantis NJ. Inhibition of *Salmonella enterica* serovar Typhimurium motility and entry into epithelial cells by a protective antilipopolysaccharide monoclonal immunoglobulin a antibody. *Infect Immun*. 2008;76(9):4137–4144. doi:10.1128/IAI.00416-08.
234. Lièven-Le Moal V, Amsellem R, Servin AL. Impairment of swimming motility by anti-diarrheic *Lactobacillus acidophilus* strain LB retards internalization of *Salmonella enterica* serovar Typhimurium within human enterocyte-like cells. *Antimicrob Agents Chemother*. 2011;55(10):4810–4820. doi:10.1128/AAC.00418-11.
235. Fayol-Messaoudi D, Coconier-Polter MH, Moal VL, Atassi F, Berger CN, Servin AL. The *Lactobacillus plantarum* strain ACADC287 isolated from a Greek cheese demonstrates antagonistic activity in vitro and in vivo against *Salmonella enterica* serovar Typhimurium. *J Appl Microbiol*. 2007;103(3):657–665. doi:10.1111/j.1365-2672.2007.03293.x.
236. Yan F, Polk DB. Probiotics and immune health. *Curr Opin Gastroenterol*. 2011;27(6):496–501. doi:10.1097/MOG.0b013e32834baa4d.
237. Viașu-Bolocan LV, Popescu F, Bica C. Probiotics and their immunomodulatory potential. *Curr Health Sci J*. 2013;39:204–209.
238. Yan F, Polk DB. Probiotics: progress toward novel therapies for intestinal diseases. *Curr Opin Gastroenterol*. 2010;26(2):95–101. doi:10.1097/MOG.0b013e328335239a.
239. Meydani SN, Ha WK. Immunological effects of yoghurt. *Am J Clin Nutr*. 2000;71:861–872.
240. He F, Tuomola E, Arvilommi H, Salminen S. Modulation of humoral immune response through probiotic intake. *FEMS Immunol Med Microbiol*. 2000;29(1):47–52. doi:10.1111/j.1574-695X.2000.tb01504.x.
241. Malin M, Suomalainen H, Saxelin M, Isolauri E. Promotion of IgA immune response in patients with Crohn's Disease by oral bacteriotherapy with *Lactobacillus GG*. *Ann Nutr Metab*. 1996;40(3):137–145. doi:10.1159/000177907.
242. Hutchins AP, Diez D, Miranda-Saavedra D. The IL-10/STAT3-mediated anti-inflammatory response: recent developments and future challenges. *Brief Funct Genom*. 2013;12(6):489–498. doi:10.1093/bfgp/elt028.
243. Mirpuri J, Sotnikov I, Myers L, Denning TL, Yarovsky F, Parkos CA, Denning PW, Louis NA, Neu J. *Lactobacillus rhamnosus* (LGG) regulates IL-10 signaling in the developing murine colon through upregulation of the IL-10R2 receptor subunit. *PLoS ONE*. 2012;7(12):e51955. doi:10.1371/journal.pone.0051955.
244. Llewellyn A, Foey A. Probiotic modulation of innate cell pathogen sensing and signalling events. *Nutr*. 2017;9:1156.
245. Saxelin M, Tynkkynen S, Mattila-Sandholm T, de Vos WM. Probiotic and other functional microbes: from markets to mechanisms. *Curr Opin Biotechnol*. 2005;16(2):204–211. doi:10.1016/j.copbio.2005.02.003.
246. Finamore A, Roselli M, Imbinto A, Seeboth J, Oswald IP, Mengheri E, Heimesaat MM. *Lactobacillus amylovorus* inhibits the TLR4 inflammatory signalling triggered by enterotoxigenic *Escherichia coli* via modulation of the negative regulators and involvement of TLR2 in intestinal Caco-2 cells and pig explants. *PLoS ONE*. 2014;9(4):e94891. doi:10.1371/journal.pone.0094891.
247. Takanashi N, Tomosada Y, Villena J, Murata K, Takahashi T, Chiba E, Tohno M, Shimazu T, Aso H, Suda Y, et al. Advanced application of bovine intestinal epithelial cell line for evaluating regulatory effect of lactobacilli against heat-killed enterotoxigenic *Escherichia coli*-mediated inflammation. *BMC Microbiol*. 2013;13(1):54. doi:10.1186/1471-2180-13-54.
248. Kaur IP, Chopra K, Saini A. Probiotics: potential pharmaceutical applications. *Eur J Pharmacol*. 2002;15(1):1–9. doi:10.1016/S0928-0987(01)00209-3.
249. Parvez S, Malik KA, Khang KHY. Probiotics and their fermented food products are beneficial for health. *J Appl Microbiol*. 2006;100(6):1171–1185. doi:10.1111/j.1365-2672.2006.02963.x.
250. Roberfroid MB. Prebiotics and probiotics: are they functional goods? *Am J Clin Nutr*. 2000;71(6S):1682S–16827S. doi:10.1093/ajcn/71.6.1682S.
251. Isolauri E, Sütas Y, Kankaanpää P, Arvilommi H, Salminen S. Probiotics: effects on immunity. *Am J Clin Nutr*. 2001;73(2):444S–450S. doi:10.1093/ajcn/73.2.444s.
252. Guarino A, Canani RB, Spagnuolo MI, Albano F, Di Benedetto L. Oral bacterial therapy reduces the duration of symptoms and of viral excretion in children with mild diarrhoea. *J Ped Gastro Nutr*. 1997;25(5):516–519. doi:10.1097/00005176-199711000-00005.
253. Cross ML. Microbes versus microbes: immune signals generated by probiotic lactobacilli and their role in protection against microbial pathogens. *FEMS Immunol*

- Med Microbiol. 2002;34(4):245–253. doi:10.1111/j.1574-695X.2002.tb00632.x.
254. Tien MT, Girardin SE, Regnault B, Le Bourhis L, Dillies MA, Coppee JY, Bourdet-Sicard R, Sansonetti PJ, Pedron T. Anti-inflammatory effect of *Lactobacillus casei* on shigella-infected human intestinal epithelial cells. *J Immunol*. 2006;176(2):1228–1237. doi:10.4049/jimmunol.176.2.1228.
255. Driessen FM, De Boer R. Fermented milks with selected intestinal bacteria: a healthy trend in new products. *Neth Milk Dairy J*. 1989;43:367–382.
256. Roessler A, Friedrich U, Vogelsang H, Bauer A, Kaatz M, Hipler UC, Schmidt I, Jahreis G. The immune system in healthy adults and patients with atopic dermatitis seems to be affected differently by a probiotic intervention. *Clin Exp Allergy*. 2008;38:93–102.
257. Gill HS, Rutherford KJ, Cross ML, Gopal PK. Enhancement of immunity in the elderly by dietary supplementation with the probiotic *Bifidobacterium lactis* HN019. *Am J Clin Nutr*. 2001;74(6):833–839. doi:10.1093/ajcn/74.6.833.
258. Gill HS, Rutherford KJ, Cross ML. Dietary probiotic supplementation enhances natural killer cell activity in the elderly: an investigation of age-related immunological changes. *J Clin Immunol*. 2001;21(4):264–271. doi:10.1023/A:1010979225018.