

Regulatory roles of noncoding RNAs in intervertebral disc degeneration as potential therapeutic targets (Review)

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Abstract. Intervertebral disc degeneration (IDD) is the leading cause of lower back pain, which is one of the primary factors that lead to disability and pose a serious economic burden. The key pathological processes involved are extracellular matrix degradation, autophagy, apoptosis, and inflammation of nucleus pulposus cells. Non-coding RNAs (ncRNAs), including microRNAs, long ncRNAs and circular RNAs, are key regulators of the aforementioned processes. ncRNAs are differentially expressed in tissues of the intervertebral disc between healthy individuals and patients and participate in the pathological progression of IDD via a complex pattern of gene regulation. However, the regulatory mechanisms of ncRNAs in IDD remain unclear. The present review summarizes the latest insights into the regulatory role of ncRNAs in IDD and sheds light on potentially novel therapeutic strategies for IDD that may be implemented in the future.

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1. Introduction

Intervertebral disc degeneration (IDD) is one of the primary causes of lower back pain, which poses serious social and economic burdens (1). The global prevalence of IDD is >60%, which leads to high costs for society (2). The pathogenesis of IDD is complex and involves genetic, aging and environmental (3) and inflammatory factors (4). The gross anatomy of the ID primarily comprises interdependent tissue compartments: The nucleus pulposus (NP) is surrounded by a fibrocartilaginous annulus fibrosus (5). The NP serves an essential role in maintaining homeostasis of the ID and the key cellular change during the development of IDD concerns centrally located NP cells that undergo phenotypical transition, compromising the structural integrity of discs (5). Although numerous researchers have performed experimental studies on the potential mechanism of IDD in recent years (6,7), the underlying precise pathological mechanisms of IDD remain unclear. Therefore, a further investigation of the molecular regulatory mechanisms underlying the homeostasis of the ID may contribute to identifying novel potential therapeutic targets for IDD.

A series of genetic and biological regulators are associated with IDD pathogenesis and non-coding RNAs (ncRNAs) are the critical factors (8,9). ncRNAs, primarily consisting of microRNAs (miRNAs or miRs), long ncRNAs (lncRNAs) and circular RNAs (circRNAs), are critical regulators in various cellular processes, such as cell proliferation, autophagy and apoptosis. miRNAs, a class of ncRNAs ~22 nucleotides in length, can recognize the 3'-untranslated regions of target mRNAs via complementary base pairing and suppress gene expression at the post-transcriptional level (10). lncRNA is a type of ncRNA with a length >200 nucleotides that binds to proteins or mRNAs using its nucleotide sequence or folded secondary structure and regulates gene expression through multiple mechanisms at the transcriptional and post-transcriptional levels (11). circRNA, primarily formed through reverse splicing, is a type of endogenous covalently closed RNA molecule that serves as a miRNA sponge to regulate the miRNA-associated cell processes (12). It has been previously documented that the level of aberrant ncRNAs is involved in various aspects of IDD, including NP cell proliferation (13),

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autophagy (14), apoptosis (15), extracellular matrix (ECM) regeneration (16) and inflammatory response (17). Therefore, dysregulation of ncRNAs in NP cells may be a crucial pathological mechanism underlying the initiation and development of IDD. Lan *et al* (18) reported 76 pairs of differentially expressed circRNAs in IDD using microarray datasets. Liu *et al* (19) documented that 636 circRNAs are differentially expressed in IDD compared with normal controls. lncRNA human leukocyte antigen complex group 18 (HCG18) is upregulated in patients with IDD and luciferase reporter assays have indicated that HCG18 may act as an endogenous sponge to decrease expression of miR-146a-5p in NP cells, thus upregulating the target gene of miR-146a-5p, decreasing the proliferation of NP cells and finally resulting in cell apoptosis (20). Zhu *et al* (21) reported that circVMA21, serving as a sponge for miR-200c, downregulates expression of X-linked inhibitor of apoptosis (XIAP). Thus, decreased expression of XIAP in inflammatory cytokine-treated NP cells is directly associated with excessive cell apoptosis. Furthermore, intradiscal injection of circVMA21 may alleviate IDD in a rat model (21). Accumulating evidence has shown the key regulatory role of ncRNAs in IDD (22,23). In addition, it has been reported that ncRNAs, targeted to IDs using viruses and other vectors, can reverse the pathological process of IDD and rescue ID function at the genetic level (22). These findings indicate the therapeutic potential of ncRNAs in IDD.

The present review aimed to summarize the latest studies on ncRNAs involved in IDD pathology and clarify the key roles of ncRNAs in the IDD process through regulating ECM degradation, autophagy, apoptosis and inflammation, which could provide a novel biotherapeutic strategy for clinical treatment of IDD (Fig. 1).

2. ncRNAs in ECM degradation

The primary features of IDD are the decrease in NP cell number and ECM (23). The structural components of ECM primarily comprise type-II collagen and aggrecan, which play a critical role in the mechanical functionality of the ID (24). However, reduced synthesis or increased degradation of ECM, induced by dysregulation of NP cell metabolism, leads to a reduced water-binding capacity of the tissue and structural collapse (24). NP cells maintain homeostasis and compositional integrity of the ID by regulating the metabolism of the ECM (25). Dysregulation of gene expression in NP cells induces enhanced release of ECM-degrading enzymes and decreased production of ECM structural molecules during the development of IDD (25). ncRNAs, as regulators of epigenetic modification, are involved in metabolic dysregulation in NP cells (26) (Table I).

Zhou *et al* (27) reported that expression of miR-31-5p is downregulated in IDD. Moreover, miR-31-5p facilitates NP cell proliferation, inhibits apoptosis, promotes ECM formation and downregulates expression of matrix-degrading enzymes in NP cells (27). Mechanistically, miR-31-5p regulates the pathway of stromal cell-derived factor-1/chemokine receptor 7 in IDD and upregulation of miR-31-5p can suppress the development of IDD *in vitro* (27). Xie *et al* (28) documented that miR-31-5p can inhibit apoptosis in endplate chondrocytes via regulating activating transcription factor 6 (ATF6) and that mesenchymal

stem cell-derived exosomes containing miR-31-5p suppress apoptosis and calcification in endplate chondrocytes. miR-31-5p serves an important role in promoting tumorigenesis and metastasis in several types of cancer, such as colon adenocarcinoma and lung adenocarcinoma (29). miR-654-5p has been reported to be involved in various biological activities, such as cancer progression, liver regeneration and cardioprotection (30,31). Wang *et al* (32) showed that the expression of miR-654-5p is dysregulated in degenerated NP tissues obtained from patients with IDD. Further assays indicated that miR-654-5p promotes ECM degradation by upregulating expression levels of matrix metalloproteinase (MMP)-3, MMP-9 and MMP-13, while downregulating collagen I, collagen II, SRY-box transcription factor 9 (SOX9) and aggrecan via autophagy suppression by binding to autophagy-related (ATG)7 to activate the PI3K/AKT/mTOR pathway (32). Du *et al* (33) explored the role of miR-16 in the inflammatory response in NP cells stimulated by lipopolysaccharide (LPS); miR-16 upregulated the expression of ECM genes (aggrecan and collagen II) in NP cells, while it downregulated genes associated with ECM-degrading enzymes [MMP3, MMP13, A disintegrin and metalloproteinase (ADAM) with thrombospondin type 1 motif (ADAMTS)4 and ADAMTS5]. Mechanistically, they found that miR-16 suppressed nuclear factor-kappa β (NF- κ B) and mitogen-activated protein kinase (MAPK) signaling pathways by targeting their upstream TGF- β -activated kinase 1 and MAP3K7-binding protein 3 gene, which serves a protective role in LPS-induced IDD (33). Several studies have shown that miR-129-5p is involved in the process of IDD (34,35). He *et al* (34) analyzed microarray datasets from the Gene Expression Omnibus database and found that miR-129-5p was dysregulated in IDD specimens, indicating that it may participate in the development of IDD and thus may serve as a potential therapeutic target for IDD. Li *et al* (35) reported that miR-129-5p serves a protective role in IDD via inhibiting apoptosis and promoting the proliferation of NP cells. Yang and Sun (36) found that miR-129-5p is downregulated in patients with IDD and is involved in the development of IDD via inhibition of NP cell apoptosis through targeting bone morphogenetic protein 2. miR-129-5p is reported to be downregulated in the NP obtained from patients with IDD (37). Furthermore, enhancer of zeste homolog 2 (EZH2) suppresses miR-129-5p, a target of MAPK1, via H3K27me3 modification, thereby facilitating development of IDD (37). Cui and Zhang (38) found that miR-129-5p is downregulated in NP tissues obtained from patients with IDD and delivery of miR-129-5p to NP cells via extracellular vesicles causes NP cell apoptosis. Moreover, they revealed that miR-129-5p plays a protective role in IDD by inhibiting the p38/MAPK pathway via targeting leucine-rich α 2-glycoprotein1, thus providing a novel therapeutic target for IDD (38).

Tan *et al* (39) detected the expression of lncRNA growth arrest specific 5 (GAS5) in normal and degenerative NP cells and found that lncRNA GAS5 was significantly upregulated in degenerative NP cells. In addition, they found that the silencing of GAS5 promoted the viability of NP cells and improved ECM formation. Mechanistically, lncRNA GAS5, which is located in the cytoplasm, sponges miR-26a-5p to improve the expression of phosphatase and tensin homolog (PTEN) and suppress the PI3K/Akt signaling pathway, thus

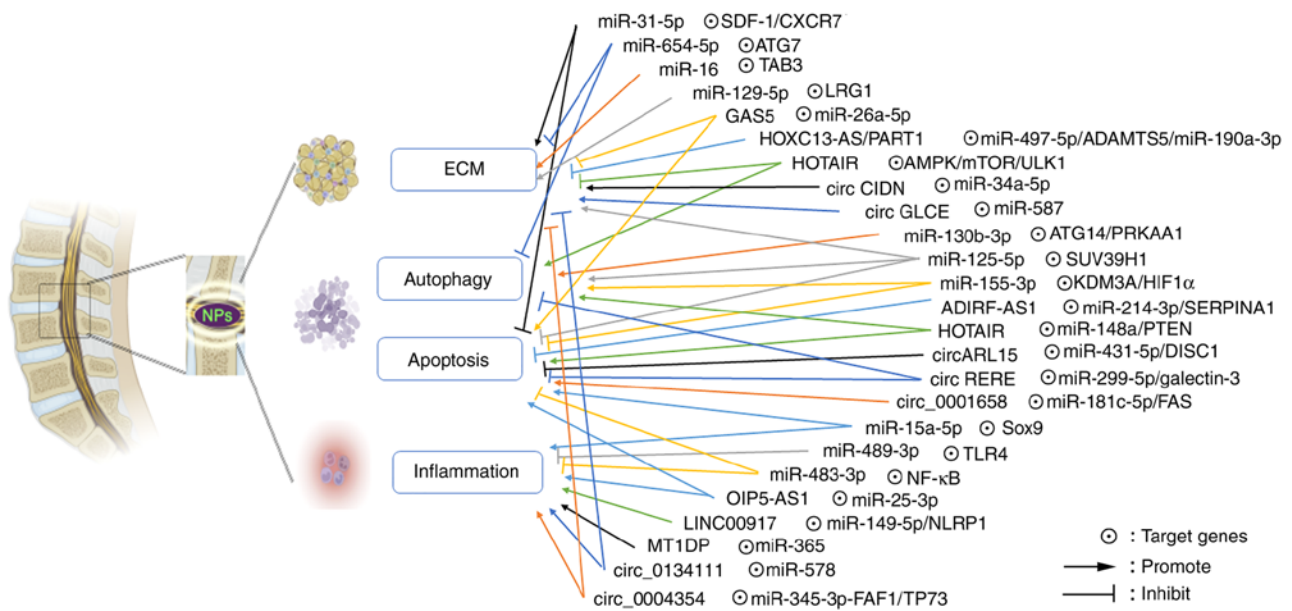


Figure 1. Regulatory role and the target genes of ncRNAs in the pathology of IDD. The ncRNAs have been investigated for their role in IDD via affecting ECM degradation, autophagy, apoptosis and inflammation. ncRNAs may simultaneously promote or inhibit one or multiple biological processes to participate in the progress of IDD, such as miR-125-5p, HOTAIR, circ_0004354 and others. IDD, intervertebral disc degeneration; ncRNA, non-coding RNA; ECM, extracellular matrix; HOTAIR, HOX transcript antisense intergenic RNA; NP, nucleus pulposus. Each color represents one ncRNA.

inhibiting ECM formation (39). Jing and Liu (40) found that lncRNA HOXC13-antisense (AS) is upregulated in IDD specimens and the aberrant expression of HOXC13-AS increases expression levels of MMP-3 and ADAMTS4 while suppressing the expression of aggrecan and collagen II in NP cells. In addition, the same study found that upregulated HOXC13-AS also promoted expression of inflammatory cytokines in NP cells. Mechanistically, HOXC13-AS promotes inflammatory cytokine production and ECM degradation via regulation of the miR-497-5p/ADAMTS5 pathway, which indicates that HOXC13-AS may be a potential therapeutic target for IDD (40). Prior studies have shown that lncRNA HOX transcript AS intergenic RNA (HOTAIR) is involved in several cellular processes, such as apoptosis, autophagy, proliferation and tumorigenesis (41-43). Zhan *et al* (44) reported that expression of HOTAIR in NP cells is associated with IDD grade and HOTAIR upregulation induces autophagy to improve ECM degradation via the AMP-activated protein kinase/mTOR/Unc-51 like autophagy activating kinase 1 pathway *in vitro*. A further *in vivo* study indicated that the suppression of HOTAIR alleviates IDD in rats, suggesting that the inhibition of HOTAIR has the potential to become a therapeutic target for IDD (44). In addition, Zhan *et al* (45) also demonstrated that HOTAIR promotes ECM degradation through the Wnt/ β -catenin signaling pathway. lncRNA prostate androgen-regulated transcript 1 (PART1) is involved in various types of cancer as an oncogene by regulating different signaling pathways (46,47). Zhao *et al* (48) found that lncRNA PART1 is upregulated in the NP tissue obtained from patients with IDD using genome-wide RNA sequencing. Recently, Zhang *et al* (49) showed that expression of PART1 was upregulated in LPS-treated NP cells while knockdown of PART1 ameliorated cell apoptosis and promoted ECM degradation in NP cells stimulated with LPS. Furthermore, the same study showed that PART1 participated in progression of IDD

by targeting miR-190a-3p to regulate cell viability and ECM degradation, indicating that PART1 may serve as a potential target for IDD treatment (49).

Xiang *et al* (50) examined the circRNA expression profile from compression-treated NP cells using circRNA microarray assays and found that circRNA-CIDN was downregulated and upregulated expression of circRNA-CIDN suppressed NP cell apoptosis and ECM degradation. circRNA-CIDN binds to miR-34a-5p and serves as a sponge to regulate silent mating type information regulation 2 homolog 1, which suggested the use of circRNA-CIDN may be a potential treatment strategy for IDD (50). Chen *et al* (51) performed microarray assays of IDD samples and found that circGLCE was downregulated. The same study confirmed that circGLCE exists in the cytoplasm of NP cells and knockdown of circGLCE induced apoptosis and improved expression of matrix-degrading enzymes in NP cells (51). Mechanistically, circGLCE alleviates IDD by suppressing the apoptosis of NP cells and ECM degradation via binding to miR-587 as a sponge to regulate signal transducing adaptor family member 1 expression, which demonstrates that circGLCE may serve as a potential therapeutic target for IDD (51).

3. ncRNAs in autophagy and apoptosis

Autophagy, a highly conserved biological phenomenon, is the primary intracellular degradation process by which cytoplasmic organelles, macromolecules and proteins are transported and degraded in lysosomes, so it plays a critical role in maintaining cellular homeostasis of NP in response to stressful stimulation (52). Dysregulation of autophagy may lead to human diseases such as obesity, atherosclerosis, metabolic disorder and osteoporosis (53). The latest research indicates that autophagy may serve a role in the pathophysiology of IDD (54). Emerging evidence has shown that ncRNAs

Table I. Roles of ncRNAs in ECM degradation.

A, miR				
First author/s, year	ncRNA	Target	Function	(Refs.)
Zhou <i>et al.</i> , 2021	miR-31-5p	Stromal cell-derived factor-1/ chemokine receptor 7	Promote proliferation and ECM formation; inhibit apoptosis	(27)
Wang <i>et al.</i> , 2021	miR-654-5p	Autophagy-related 7	Inhibit autophagy; promote ECM degradation	(32)
Du <i>et al.</i> , 2021	miR-16	TGF- β -activated kinase 1 and MAP3K7-binding protein 3 gene	Promote ECM formation	(33)
Cui and Zhang, 2021	miR-129-5p	Leucine-rich α 2-glycoprotein1	Inhibit ECM degradation	(38)
B, Long ncRNA				
First author/s, year	ncRNA	Target	Function	(Refs.)
Tan <i>et al.</i> , 2021	GAS5	miR-26a-5p	Inhibit ECM formation	(39)
Jing and Liu, 2021	HOXC13-AS	miR-497-5p/disintegrin and metallopeptidase with thrombospondin type 1 motif 5	Promote ECM degradation	(40)
Zhan <i>et al.</i> , 2020	HOTAIR	AMP-activated protein kinase/ mTOR/ Unc-51 like autophagy activating kinase 1	Promote autophagy and ECM degradation	(44)
Zhang <i>et al.</i> , 2021	PART1	miR-190a-3p	Promote ECM degradation	(49)
C, circRNA				
First author/s, year	ncRNA	Target	Function	(Refs.)
Xiang <i>et al.</i> , 2020	circCIDN	miR-34a-5p	Inhibit ECM degradation	(50)
Chen <i>et al.</i> , 2020	circGLCE	miR-587	Inhibit ECM degradation	(51)

ncRNA, non-coding RNA; ECM, extracellular matrix; circ, circular; miR, microRNA.

are essential for cellular functions of the NP, including autophagy (54).

Normal apoptosis, which is autonomous programmed cell death regulated by genes, can preserve the stability of the intracellular environment (55). The apoptosis of NP cells is a key pathological feature of IDD and high rates of apoptosis are associated with degenerative processes involving IDs, which reduce the number of cells in NP tissue, damage ID structure and function and induce a disturbance of tissue homeostasis (55). Degenerated IDs exhibit higher rates of apoptosis, but the underlying mechanisms remain unclear (55). Importantly, evidence indicates that ncRNAs are involved in abnormal apoptosis of NP cells during IDD (56). It is reported that numerous ncRNAs can simultaneously affect the process of apoptosis and autophagy during development of IDD (57,58). Several studies have indicated that autophagy alleviates IDD to protect NP cells from apoptosis and the imbalance between autophagy and apoptosis is involved in IDD progress (6,59) (Table II).

miR-130b is involved in various pathophysiological processes, such as regulation of cell proliferation and metastasis

in cancer as an oncogene (60,61), affecting macrophage polarization (62) and modulation of epithelial-mesenchymal crosstalk in pulmonary fibrosis (63). Recently, Wu *et al.* (57) discovered that miR-130b-3p is dysregulated in degenerative NP obtained from humans and rats and that miR-130b-3p significantly influences IDD progression both *in vivo* and *in vitro*. In addition, they found that inhibiting miR-130b-3p protects NP cells from oxidative stress-stimulated dysfunction and miR-130b-3p inhibition promotes autophagy of NP cells. Mechanistically, they illustrated that miR-130b-3p regulated autophagy-related 14 (ATG14) and protein kinase AMP-activated catalytic subunit α 1 (PRKAA1) directly and inhibited expression of ATG14 or PRKAA1 while the application of the autophagy inhibitor (3-Methyladenine) suppressed autophagic flux and attenuated the protective effects of miR-130b-3p inhibition in tertbutyl hydro-peroxide (TBHP)-stimulated NP cells. Notably, IDD was improved in a rat model by the injection of AAV-miR-130b-3p, which inhibits miR-130b-3p (57). These findings showed that by focusing on ATG14 and PRKAA1, miR-130b-3p inhibition may increase

Table II. Roles of ncRNAs in autophagy and apoptosis.

A, miR				
First author/s, year	ncRNA	Target	Function	(Refs.)
Wu <i>et al</i> , 2022	miR-130b-3p	Autophagy-related 14/protein kinase AMP-activated catalytic subunit α 1	Promote autophagy	(57)
Chen and Jiang, 2022	miR-125-5p	Suppressor of variegation 3-9 homolog 1	Promote autophagy; inhibit apoptosis and ECM degradation	(66)
Zhou <i>et al</i> , 2021	miR-155-3p	Lysine demethylase 3A/Hypoxia-inducible factor 1- α	Promote autophagy; inhibit apoptosis	(71)
Yan <i>et al</i> , 2022	miR-328-5p	WW domain containing E3 ubiquitin protein ligase 2	Promote apoptosis; inhibit proliferation	(73)
B, Long ncRNA				
First author/s, year	ncRNA	Target	Function	(Refs.)
Yu <i>et al</i> , 2022	Growth arrest specific 5	miR-17-3p/ Ang-2	Promote apoptosis; promote ECM degradation	(58)
Zhong <i>et al</i> , 2022	ADIRF-AS1	miR-214-3p/ SERPINA1	Inhibit apoptosis and senescence	(79)
Zhang <i>et al</i> , 2022	HOTAIR	miR-148a/PTEN	Promote apoptosis and autophagy	(84)
C, circRNA				
First author/s, year	ncRNA	Target	Function	(Refs.)
Wang <i>et al</i> , 2021	circARL15	miR-431-5p circRERE/DISC1	Inhibit apoptosis; promote proliferation	(85)
Wang <i>et al</i> , 2021	circRERE	miR-299-5p/galectin-3	Inhibit apoptosis and autophagy	(86)
Meng and Xu 2021	circ_0001658	miR-181c-5p/FAS	Promote apoptosis; inhibit proliferation	(89)

ncRNA, non-coding RNA; ECM, extracellular matrix; miR, microRNA; circ, circular.

autophagic flow and decrease IDD, which suggested that miR-130b-3p may serve as a useful treatment approach for IDD.

Exosomes are protein and nucleic acid-containing membrane vesicles with a diameter of 40-100 nm that are released from cells via exocytosis following fusion with the plasma membrane (64). A recent report indicated that exosomes serve a key role in NP apoptosis and autophagy (65). Recently, Chen and Jiang (66) found that cartilage endplate stem cell-derived exosomes induce NP cell autophagy but suppress apoptosis and ECM degradation. Furthermore, they discovered that expression of miR-125-5p is significantly upregulated in exosomes and miR-125-5p regulates suppressor of variegation 3-9 homolog 1 (SUV39H1) to improve autophagy and inhibit apoptosis and as ECM degradation in NP cells. They found that miR-125-5p derived from cartilage endplate stem cell-derived exosomes improves microtubule associated protein 1 light chain 3 (LC3)B expression in NP tissue of IDD rats, which suggested that miR-125-5p may serve as a therapeutic target for IDD. In addition, miR-125-5p could influence

the inflammatory response and be involved in the apoptosis of intestinal epithelial cells via the Janus kinase 1/STAT3 and NF- κ B pathways in ulcerative colitis (67). miR-125a-5p was also reported to downregulate TNF receptor superfamily member 1B gene (TNFRSF1B) expression and improve osteoclast differentiation, which indicated that miR-125a-5p is involved in osteoclast differentiation (68).

Studies have shown that miR-155-3p, derived from miR-155, is associated with IDD. Wang *et al* (69) documented that Fas-induced apoptosis is promoted when miR-155 is suppressed and inhibited when miR-155 is upregulated in human NP cells. Zhang *et al* (70) reported that upregulation of miR-155 decreases IDD, while its suppression worsens IDD. Recently, Zhou *et al* (71) revealed that miR-155-3p is downregulated in NP tissues and cells of IDD. In addition, upregulation of the expression of miR-155-3p or suppression of lysine demethylase 3A (KDM3A) contribute to autophagy and inhibit apoptosis of NP cells. Mechanistically, miR-155-3p suppresses expression of KDM3A and Hypoxia-inducible factor 1- α in NP cells from IDD samples, which may be a potential

approach for the treatment of IDD. According to a recent study, miR-328-5p is abnormally expressed in numerous types of cancer (colorectal cancer and lung cancer) and is associated with apoptosis of cancer cells (72). Yan *et al* (73) found that the expression of miR-328-5p is upregulated significantly, while the expression of its target gene WW domain containing E3 ubiquitin protein ligase 2 (WWP2) is downregulated in IDD tissues compared with normal tissues. miR-328-5p upregulation is positively, whereas WWP2 downregulation is inversely, linked with the degeneration grade of IDD. In addition, they identified that miR-328-5p suppresses proliferation and causes apoptosis of NP cells via regulation of the expression of Bcl-2, Bax and caspase-3. Mechanistically, miR-328-5p may affect NP cell apoptosis by targeting WWP2 and consequently be involved in the development of IDD, which may provide a novel potential target for IDD therapy.

lncRNAs are involved in the processes of apoptosis and autophagy of NP cells in IDD (74). Growth arrest-specific transcript 5 (GAS5), which has been documented to serve as a tumor suppressor in various types of cancer, is related to the process of osteoarthritis as a pathogenic lncRNA (75). A previous study revealed the ability of GAS5 to promote apoptosis of NP cells via targeting miR-26-5p and suppression of GAS5 expression is a potential therapeutic target for the treatment of IDD (39). In addition, GAS5 has also been shown to target other miRNAs, such as miR-18a-5p and miR-17-5p (39,76). Recently, Yu *et al* (58) found that lncRNA GAS5 is upregulated in the NP tissue obtained from patients with IDD and it could target miR-17-3p and affect Ang-2 expression. Furthermore, they reported that downregulation of GAS5 suppresses levels of activated caspase-3, activated caspase-7, activated caspase-9, MMP13 and ADAMTS4, whereas GAS5 upregulates collagen II expression. Importantly, they discovered that GAS5 suppression inhibits apoptosis and ECM degradation in NP cells while promoting proliferation via upregulating miR-17-3p. In addition, GAS5 inhibition or miR-17-3p upregulation relieves the degree of IDD in mouse models, which demonstrates that inhibition of GAS5 suppresses NP cell apoptosis and improves ECM remodeling, ultimately alleviating IDD via miR-17-3p-dependent inhibition of Ang-2. These results indicated that GAS5 could serve as a novel therapeutic target for IDD treatment.

lncRNA adipogenesis regulatory factor-antisense RNA 1 (ADIRF-AS1), located at 10q23.2, is broadly expressed in numerous types of cell, such as endothelial and epithelial cells. Previously, it has been reported that expression of ADIRF-AS1 is downregulated in more than 100 cancer cell lines (77). Xu *et al* (78) reported that ADIRF-AS1 is upregulated in osteosarcoma (OS) and the overall survival of patients with OS and high ADIRF-AS1 expression levels is shorter than that of those with low ADIRF-AS1 expression levels. Furthermore, ADIRF-AS1 suppression inhibits migration and invasiveness of OS cells and promotes apoptosis. Mechanistically, ADIRF-AS1 could target miR-761 and upregulate insulin receptor substrate 1. Recently, Zhong *et al* (79) showed that ADIRF-AS1 expression is inhibited in high-grade degenerated NP tissues and is positively correlated with expression of serpin family A member 1 (SERPINA1). In addition, ADIRF-AS1 upregulation decreases degenerative changes in NP cells. miR-214-3p directly binds to SERPINA1 and

ADIRF-AS1 and negatively regulates ADIRF-AS1 expression. Furthermore, upregulation of ADIRF-AS1 relieves IDD via targeting miR-214-3p to promote the expression of SERPINA1, which suggested that ADIRF-AS1 may be used as a potential target for IDD treatment.

HOTAIR has been reported to be involved in the development of numerous types of cancer, such as breast and lung cancer as well as hepatocellular carcinoma (80-82). Wang *et al* (83) reported that HOTAIR is involved in autophagy. Recently, HOTAIR was documented to participate in progression of IDD by regulating NP cell apoptosis, senescence and autophagy (44). Zhan *et al* (44) found that expression of HOTAIR is positively correlated with IDD grade and its upregulation improves autophagy. Further *in vivo* experiments demonstrated that downregulation of HOTAIR ameliorates IDD in rat model of IDD, which indicated that suppression of HOTAIR expression may serve as a treatment for IDD. Zhang *et al* (84) reported that HOTAIR is significantly upregulated in degenerative NP cells and its downregulation suppresses degenerative NP cell apoptosis and autophagy. In addition, they showed that HOTAIR increases expression of PTEN via sponging miR-148a. *In vivo* assay confirmed that HOTAIR suppression inhibits autophagy and apoptosis in ID tissues, which ameliorates pathological injury in the IDD model.

Studies have reported that circRNAs, acting as competing endogenous RNAs, are associated with pathogenesis of IDD by affecting NP cell apoptosis and autophagy. Recently, Wang *et al* (85) analyzed the dataset GSE67567 from the Gene Expression Omnibus database and found that the expression of circARL15 was significantly downregulated in IDD tissues. They also found that circARL15 expression was negatively associated with miR-431-5p and positively correlated with DISC1 scaffold protein (DISC1). In addition, they showed that circARL15 suppresses NP cell apoptosis but contributes to NP cell proliferation via regulating the miR-431-5p/DISC1 signaling axis, which suggested that circARL15 may serve as a biomarker and provide a promising therapeutic target for patients with IDD.

Wang *et al* (86) recently discovered that miR-299-5p is downregulated in IDD tissues and H₂O₂-induced NP cells, while the overexpression of miR-299-5p improves cell viability and suppresses apoptosis and autophagy under H₂O₂ stimulation. Moreover, circRERE expression is significantly upregulated in IDD tissues and H₂O₂-stimulated NP cells. Inhibition of circRERE reverses the effects of miR-299-5p upregulation on cell viability, apoptosis and autophagy in NP cells. Mechanistically, circRERE assists in H₂O₂-stimulated apoptosis and autophagy of NP cells via the miR-299-5p/galectin-3 pathway, which may provide a potential target for clinical therapy of IDD in the future (86). Liu *et al* (87) found that circRERE is downregulated in osteoarthritis cartilage and chondrocytes because of its increased N6-methyladenosine (m6A) modification. circRERE downregulation promotes the apoptosis of chondrocytes and upregulation of circRERE improves osteoarthritis via targeting the miR-195-5p/IRF2BPL/ β -catenin signaling pathway (87). These reports indicated that circRERE serves a critical role in bone-associated disease.

Wang *et al* (88) reported that circ_0001658 is significantly upregulated in osteosarcoma tissue and increases proliferation,

Table III. Role of ncRNAs in inflammation.

A, miR				
First author/s, year	ncRNA	Target	Function	(Refs.)
Zhang <i>et al</i> , 2021	miR-15a-5p	Sox9	Promote inflammation and apoptosis	(94)
Dong and Dong, 2021	miR-489-3p	Toll-like receptor 4	Inhibit inflammation	(95)
Ji <i>et al</i> , 2021	miR-483-3p	NF- κ B	Inhibit inflammation and apoptosis	(96)
B, Long ncRNA				
First author/s, year	ncRNA	Target	Function	(Refs.)
Che <i>et al</i> , 2021	OIP5-AS1	miR-25-3p	Promote apoptosis and inflammation; inhibit proliferation	(97)
Li <i>et al</i> , 2022	LINC00917	miR-149-5p/NLR Family Pyrin domain Containing 1	Promote inflammation, inhibit proliferation	(102)
Liao <i>et al</i> , 2021	MT1DP	miR-365	Promote inflammation	(103)
C, Circular RNA				
First author/s, year	ncRNA	Target	Function	(Refs.)
Yan <i>et al</i> , 2022	circ_0134111	miR-578	Promote inflammation and ECM degradation	(106)
Li <i>et al</i> , 2022	circ_0004354	miR-345-3p-FAF1/tumor protein P73	Promote inflammation and ECM degradation	(107)

ncRNA, non-coding RNA; ECM, extracellular matrix; miR, microRNA.

migration and invasion of osteosarcoma cells and suppresses apoptosis by targeting the miR-382-5p/YB-1 signaling pathway. Meng and Xu (89) found that circ_0001658 expression is significantly increased in the NP tissue obtained from patients with IDD and hsa-miR-181c-5p is downregulated. In addition, they revealed that circ_0001658 upregulation suppressed proliferation of NP cells and increased apoptosis. Mechanistically, circ_0001658 served as a sponge for miR-181c-5p and regulated expression of Fas in NP cells, which indicated that circ_0001658 may be a novel target for the treatment of IDD in the future. Recently, circ_0001658 was found to be upregulated in gastric cancer tissue (90). In addition, the same study showed that the inhibition of circ_0001658 expression resulted in decreased autophagy and apoptosis of gastric cancer cells via miR-182 targeting, which suggested that circ_0001658 may serve as a potential target for gastric cancer treatment. The aforementioned results indicated that circ_0001658 may serve a key role in a variety of diseases.

4. ncRNAs in inflammation

Aggravation of inflammatory cytokine levels contributes to IDD. Various studies have demonstrated upregulation of the proinflammatory cytokines, such as Tumor necrosis factor (TNF)- α , IL-6, IL-1 β and IL-1 α , in IDD (91,92). These

proinflammatory cytokines stimulate ECM degradation and reformation of the phenotype of NP cells, thus causing degeneration of the ID (93). Studies (Table III) have indicated that ncRNAs are associated with production of inflammatory cytokines in NP tissue.

Zhang *et al* (94) reported that miR-15a-5p is upregulated in ID tissues obtained from patients with IDD and mouse models and downregulating miR-15a-5p expression increases Sox9 to trigger phospho-p65 expression, inhibit NP cell apoptosis and inflammatory factor levels, facilitate proliferation of NP cells and arrest the NP cells at the S and G₂/M phase. Furthermore, they revealed that inhibition of miR-15a-5p promotes Sox9 expression to suppress the inflammatory response and apoptosis of NP cells in mice with IDD via the NF- κ B signaling pathway, which indicated that targeting miR-15a-5p may be helpful for IDD therapy. Dong and Dong (95) found that miR-489-3p is significantly downregulated in LPS-stimulated human NP cells. In addition, miR-489-3p inhibits LPS-stimulated suppression of cell viability and increases apoptosis as well as levels of inflammatory cytokines. miR-489-3p plays an important role in inhibiting LPS-induced suppression of ECM deposition through downregulation of the expression of aggrecan and collagen type II in human NP cells. Mechanistically, miR-489-3p is involved in suppressing LPS-induced activation of the NF- κ B pathway in human NP

cells, which indicates that miR-489-3p may serve as a potential treatment target for IDD. Arctigenin (ATG), an active ingredient of the Chinese herbal medicine *Arctium lappa*, serves a key role in anti-inflammatory and antioxidant effects. Recently, Ji *et al* (96) reported that ATG suppresses apoptosis and inflammation while improving miR-483-3p expression. Moreover, inhibition of miR-483-3p reverses these effects.

Previously, OIP5 antisense RNA 1 (OIP5-AS1) was reported to induce apoptosis of oxidized low-density lipoprotein (ox-LDL)-stimulated vascular endothelial cells by regulating glycogen synthase kinase 3 β (GSK-3 β) via recruiting EZH2 (97), while inhibition of OIP5-AS1 improves cell viability and suppresses apoptosis in ox-LDL-stimulated human endothelial cells (98). Moreover, it was demonstrated that lncRNA OIP5-AS1 serves an important role in suppressing osteoblast differentiation of valve interstitial cells via the miR-137/twist-related protein 11 signaling pathway (99). Further study indicated that lncRNA OIP5-AS1 is associated with occurrence and development of osteoarthritis and downregulation of lncRNA OIP5-AS1 worsens osteoarthritis by regulating the miR-29b-3p/progranulin axis (100). Recently, Che *et al* (97) found that OIP5-AS1 is upregulated in IDD tissues and silencing of OIP5-AS1 promotes cell proliferation but inhibits apoptosis and ECM degradation. In addition, OIP5-AS1 suppression inhibits the inflammatory response in LPS-induced NP cells. Mechanistically, OIP5-AS1 regulates proliferation, apoptosis, inflammation and ECM degradation via targeting miR-25-3p, which suggested that the OIP5-AS1/miR-25-3p axis may be a potential target for IDD treatment. Previously, it was reported that 135 lncRNAs are increased and 170 lncRNAs are decreased in IDD samples, among which LINC00917 is the most upregulated lncRNA and is hypothesized to regulate IDD progression (101). Recently, Li *et al* (102) found that LINC00917 is significantly increased in TBHP-stimulated NP cells and inhibition of LINC00917 improves proliferation and suppresses the inflammatory response and pyroptosis of NP cells. Furthermore, they revealed that LINC00917 silencing restores NP cellular function and inhibits IDD progression by regulating the miR-149-5p/NLR family pyrin domain containing 1 signaling pathway. Liao *et al* (103) reported that metallothionein 1D pseudogene (MT1DP) and miR-365 are significantly upregulated in human IDD NP tissue and NP cells, while the expression of nuclear factor erythroid 2-related factor 2 (NRF-2) is significantly downregulated. They found that upregulation of MT1DP and miR-365 and restriction of NRF-2 inhibit NP cell viability and cause apoptosis and inflammation. Furthermore, MT1DP and miR-365 induce inflammation in NP cells by damaging the mitochondrial membrane and mitochondrial function.

circRNAs have been suggested as treatment targets for several types of disease, such as cancer and metabolic and cardiovascular diseases (104). Liu and Zhang (105) reported that expression of circ_0134111 is increased in IL-1 β -activated chondrocytes and knockdown of circ_0134111 reverses IL-1 β -induced cell decreases by suppressing apoptosis. In addition, they reported that circ_0134111 causes osteoarthritis progression via targeting miR-224-5p and regulating C-C motif chemokine ligand 1. Recently, Yan *et al* (106) found that circ_0134111 expression is significantly upregulated in IDD tissue and the elevation of circ_0134111 is greater in severe

IDD cases. In addition, they discovered that IL-1 β and TNF- α significantly increase circ_0134111 expression in NP cells. Furthermore, circ_0134111 overexpression increases proliferation, inflammatory cytokine production and ECM degradation in NP cells. circ_0134111 promotes progression of IDD by improving NP cell inflammation and ECM degradation partly by regulating miR-578, which suggested that circ_0134111 may serve as a potential target for IDD treatment. Li *et al* (107) found that circ_0004354 competes with circ_0040039 to stimulate the progression of IDD by regulating miR-345-3p-FAF1/tumour protein P73 pathway-mediated apoptosis, inflammatory response and ECM degradation of NP cells. These results provide insight into the circ_0004354-mediated post-transcriptional regulation of IDD, thus contributing to the development of a promising treatment target for IDD in future.

5. Conclusion

Over the past few years, emerging studies have indicated that a series of ncRNAs are involved in the progression of IDD, particularly NP cell phenotypical regulation, which provides insight into the pathogenesis of IDD (8,9). Certain dysregulated ncRNAs may be useful diagnostic biomarkers and therapeutic targets in the future with continued research (108). The present review discussed progression of IDD, the role of miRNAs and lncRNAs and control of downstream target genes to regulate ECM degradation, inflammation, autophagy and apoptosis of NP cells. The present section discusses the key regulatory role of ncRNAs in the intervention and treatment of IDD from a macro perspective, as well as limitations of research in clinical practice and potential future research directions.

ncRNAs have potential for investigation as medications or biomarkers for the treatment of IDD according to the findings of prior studies (108,109). However, to the best of our knowledge, no clinical trials have been performed on ncRNAs in IDD and all discussed studies have only used cell/animal experiments. Nonetheless, the theoretical approach needs to translate into a real course of action. Based on animal models, it is possible to slow down or stop the progression of IDD by boosting, lowering or eliminating the expression of one or more particular miRNAs, lncRNAs and circRNAs *in vivo* (21,44). To date, most studies have primarily focused on disease samples, cells and small animals, which are dissimilar from the biomechanics of humans (106,107). Therefore, there is an urgent need for an improved animal model of the human spine. Additionally, the regulatory mechanism of humans is complex. Thus, delivering therapeutic ncRNAs to nearby degenerating IDs via a carrier appears to be a workable treatment plan for IDD in the future (108).

Numerous obstacles still need to be overcome, such as the fact that an ncRNA binding to its target gene is not entirely complementary (44,84), indicating that one ncRNA may control several targets or that one target can be controlled by multiple ncRNAs. The first challenge to be addressed in clinical application of ncRNAs will be improvement of the specificity of ncRNAs toward the targets. Therefore, highly selective ncRNAs that serve a role in numerous IDD-associated processes may provide opportunities to prevent IDD. Thus, ncRNAs, such as miRNAs, lncRNAs and circRNAs, as targets for the therapy of IDD may become a unique research

area with the development of nanoparticle technology which could delivery ncRNAs and a thorough understanding of the etiology of IDD. Several delivery methods have been created to minimize off-target effects, particularly concerning nanoparticles, which stand out due to their stability, small size, biocompatibility and self-assembly properties (110,111). Thus, the potential of nanoparticles as efficient ncRNA delivery systems is appealing and merits further research.

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Authors' contributions

CG and YC wrote the manuscript. YW and YH edited the manuscript. All authors have read and approved the final manuscript. Data authentication is not applicable.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

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Competing interests

The authors declare that they have no competing interests.

References

- Maher C, Underwood M and Buchbinder R: Non-specific low back pain. *Lancet* 389: 736-747, 2017.
- Gianola S, Castellini G, Andreano A, Corbetta D, Frigerio P, Pecoraro V, Redaelli V, Tettamanti A, Turolla A, Moja L and Valsecchi MG: Effectiveness of treatments for acute and sub-acute mechanical non-specific low back pain: Protocol for a systematic review and network meta-analysis. *Syst Rev* 8: 196, 2019.
- Tessier S and Risbud MV: Understanding embryonic development for cell-based therapies of intervertebral disc degeneration: Toward an effort to treat disc degeneration subphenotypes. *Dev Dyn* 250: 302-317, 2021.
- Lyu FJ, Cui H, Pan H, Mc Cheung K, Cao X, Iatridis JC and Zheng Z: Painful intervertebral disc degeneration and inflammation: From laboratory evidence to clinical interventions. *Bone Res* 9: 7, 2021.
- Ji ML, Jiang H, Zhang XJ, Shi PL, Li C, Wu H, Wu XT, Wang YT, Wang C and Lu J: Preclinical development of a microRNA-based therapy for intervertebral disc degeneration. *Nat Commun* 9: 5051, 2018.
- Xu G, Liu C, Jiang J, Liang T, Yu C, Qin Z, Zhang Z, Lu Z and Zhan X: A novel mechanism of intervertebral disc degeneration: Imbalance between autophagy and apoptosis. *Epigenomics* 12: 1095-1108, 2020.
- Dong W, Liu J, Lv Y, Wang F, Liu T, Sun S, Liao B, Shu Z and Qian J: miR-640 aggravates intervertebral disc degeneration via NF- κ B and WNT signalling pathway. *Cell Prolif* 52: e12664, 2019.
- Memczak S, Jens M, Elefsinioti A, Torti F, Krueger J, Rybak A, Maier L, Mackowiak SD, Gregersen LH, Munschauer M, *et al*: Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* 495: 333-338, 2013.
- Mitra A, Pfeifer K and Park KS: Circular RNAs and competing endogenous RNA (ceRNA) networks. *Transl Cancer Res* 7 (Suppl 5): S624-S628, 2018.
- Kabekkodu SP, Shukla V, Varghese VK, D' Souza J, Chakrabarty S and Satyamoorthy K: Clustered miRNAs and their role in biological functions and diseases. *Biol Rev Camb Philos Soc* 93: 1955-1986, 2018.
- Bridges MC, Daulagala AC and Kourtidis A: LNCcation: LncRNA localization and function. *J Cell Biol* 220: e202009045, 2021.
- Zheng S, Zhang X, Odame E, Xu X, Chen Y, Ye J, Zhou H, Dai D, Kyei B, Zhan S, *et al*: CircRNA-Protein interactions in muscle development and diseases. *Int J Mol Sci* 22: 3262, 2021.
- Liu H, Huang X, Liu X, Xiao S, Zhang Y, Xiang T, Shen X, Wang G and Sheng B: miR-21 promotes human nucleus pulposus cell proliferation through PTEN/AKT signaling. *Int J Mol Sci* 15: 4007-4018, 2014.
- Xie L, Huang W, Fang Z, Ding F, Zou F, Ma X, Tao J, Guo J, Xia X, Wang H, *et al*: CircERCC2 ameliorated intervertebral disc degeneration by regulating mitophagy and apoptosis through miR-182-5p/SIRT1 axis. *Cell Death Dis* 10: 751, 2019.
- Wang T, Li P, Ma X, Tian P, Han C, Zang J, Kong J and Yan H: MicroRNA-494 inhibition protects nucleus pulposus cells from TNF- α -induced apoptosis by targeting JunD. *Biochimie* 115: 1-7, 2015.
- Jing W and Jiang W: MicroRNA-93 regulates collagen loss by targeting MMP3 in human nucleus pulposus cells. *Cell Prolif* 48: 284-292, 2015.
- Gu SX, Li X, Hamilton JL, Chee A, Kc R, Chen D, An HS, Kim JS, Oh CD, Ma YZ, *et al*: MicroRNA-146a reduces IL-1 dependent inflammatory responses in the intervertebral disc. *Gene* 555: 80-87, 2015.
- Lan PH, Liu ZH, Pei YJ, Wu ZG, Yu Y, Yang YF, Liu X, Che L, Ma CJ, Xie YK, *et al*: Landscape of RNAs in human lumbar disc degeneration. *Oncotarget* 7: 63166-63176, 2016.
- Liu X, Che L, Xie YK, Hu QJ, Ma CJ, Pei YJ, Wu ZG, Liu ZH, Fan LY and Wang HQ: Noncoding RNAs in human intervertebral disc degeneration: An integrated microarray study. *Genom Data* 5: 80-81, 2015.
- Xi Y, Jiang T, Wang W, Yu J, Wang Y, Wu X and He Y: Long non-coding HCG18 promotes intervertebral disc degeneration by sponging miR-146a-5p and regulating TRAF6 expression. *Sci Rep* 7: 13234, 2017.
- Zhu J, Zhang X, Gao W, Hu H, Wang X and Hao D: lncRNA/circRNA-miRNA-mRNA ceRNA network in lumbar intervertebral disc degeneration. *Mol Med Rep* 20: 3160-3174, 2019.
- Fontana G, See E and Pandit A: Current trends in biologics delivery to restore intervertebral disc anabolism. *Adv Drug Deliv Rev* 84: 146-158, 2015.
- Xu D, Ma X, Sun C, Han J, Zhou C, Wong SH, Chan MTV and Wu WKK: Circular RNAs in intervertebral disc degeneration: An updated review. *Front Mol Biosci* 8: 781424, 2022.
- Cazzanelli P and Wuertz-Kozak K: MicroRNAs in intervertebral disc degeneration, apoptosis, inflammation, and mechanobiology. *Int J Mol Sci* 21: 3601, 2020.
- Dowdell J, Erwin M, Choma T, Vaccaro A, Iatridis J and Cho SK: Intervertebral disk degeneration and repair. *Neurosurgery* 80 (3S): S46-S54, 2017.
- Li G, Ma L, He S, Luo R, Wang B, Zhang W, Song Y, Liao Z, Ke W, Xiang Q, *et al*: WTAP-mediated m⁶A modification of lncRNA NORAD promotes intervertebral disc degeneration. *Nat Commun* 13: 1469, 2022.
- Zhou Y, Deng M, Su J, Zhang W, Liu D and Wang Z: The role of miR-31-5p in the development of intervertebral disc degeneration and its therapeutic potential. *Front Cell Dev Biol* 9: 633974, 2021.

28. Xie L, Chen Z, Liu M, Huang W, Zou F, Ma X, Tao J, Guo J, Xia X, Lyu F, *et al*: MSC-Derived exosomes protect vertebral endplate chondrocytes against apoptosis and calcification via the miR-31-5p/ATF6 Axis. *Mol Ther Nucleic Acids* 22: 601-614, 2020.
29. Mi B, Li Q, Li T, Liu G and Sai J: High miR-31-5p expression promotes colon adenocarcinoma progression by targeting TNS1. *Aging (Albany NY)* 12: 7480-7490, 2020.
30. Li P, Cai JX, Han F, Wang J, Zhou JJ, Shen KW and Wang LH: Expression and significance of miR-654-5p and miR-376b-3p in patients with colon cancer. *World J Gastrointest Oncol* 12: 492-502, 2020.
31. Lu M, Wang C, Chen W, Mao C and Wang J: miR-654-5p Targets GRAP to promote proliferation, metastasis, and chemoresistance of oral squamous cell carcinoma through Ras/MAPK signaling. *DNA Cell Biol* 37: 381-388, 2018.
32. Wang S, Guo Y, Zhang X and Wang C: miR-654-5p inhibits autophagy by targeting ATG7 via mTOR signaling in intervertebral disc degeneration. *Mol Med Rep* 23: 444, 2021.
33. Du K, He X and Deng J: MicroRNA-16 inhibits the lipopolysaccharide-induced inflammatory response in nucleus pulposus cells of the intervertebral disc by targeting TAB3. *Arch Med Sci* 17: 500-513, 2018.
34. He J, Xue R, Li S, Lv J, Zhang Y, Fan L, Teng Y and Wei H: Identification of the potential molecular targets for human intervertebral disc degeneration based on bioinformatic methods. *Int J Mol Med* 36: 1593-1600, 2015.
35. Li N, Gao Q, Zhou W, Lv X, Yang X and Liu X: MicroRNA-129-5p affects immune privilege and apoptosis of nucleus pulposus cells via regulating FADD in intervertebral disc degeneration. *Cell Cycle* 19: 933-948, 2020.
36. Yang W and Sun P: Downregulation of microRNA-129-5p increases the risk of intervertebral disc degeneration by promoting the apoptosis of nucleus pulposus cells via targeting BMP2. *J Cell Biochem* 120: 19684-19690, 2019.
37. Zhou M, He SJ, Liu W, Yang MJ, Hou ZY, Meng Q and Qian ZL: EZH2 upregulates the expression of MAPK1 to promote intervertebral disc degeneration via suppression of miR-129-5p. *J Gene Med* 24: e3395, 2022.
38. Cui S and Zhang L: microRNA-129-5p shuttled by mesenchymal stem cell-derived extracellular vesicles alleviates intervertebral disc degeneration via blockade of LRG1-mediated p38 MAPK activation. *J Tissue Eng* 12: 20417314211021679, 2021.
39. Tan L, Xie Y, Yuan Y and Hu K: LncRNA GAS5 as miR-26a-5p sponge regulates the PTEN/PI3K/Akt axis and affects extracellular matrix synthesis in degenerative nucleus pulposus cells in vitro. *Front Neurol* 12: 653341, 2021.
40. Jing W and Liu W: HOXC13-AS induced extracellular matrix loss via targeting miR-497-5p/ADAMT5 in intervertebral disc. *Front Mol Biosci* 8: 643997, 2021.
41. Bao X, Ren T, Huang Y, Sun K, Wang S, Liu K, Zheng B and Guo W: Knockdown of long non-coding RNA HOTAIR increases miR-454-3p by targeting Stat3 and Atg12 to inhibit chondrosarcoma growth. *Cell Death Dis* 8: e2605, 2017.
42. Li E, Zhao Z, Ma B and Zhang J: Long noncoding RNA HOTAIR promotes the proliferation and metastasis of osteosarcoma cells through the AKT/mTOR signaling pathway. *Exp Ther Med* 14: 5321-5328, 2017.
43. Mercer TR, Dinger ME and Mattick JS: Long non-coding RNAs: Insights into functions. *Nat Rev Genet* 10: 155-159, 2009.
44. Zhan S, Wang K, Xiang Q, Song Y, Li S, Liang H, Luo R, Wang B, Liao Z, Zhang Y and Yang C: lncRNA HOTAIR upregulates autophagy to promote apoptosis and senescence of nucleus pulposus cells. *J Cell Physiol* 235: 2195-2208, 2020.
45. Zhan S, Wang K, Song Y, Li S, Yin H, Luo R, Liao Z, Wu X, Zhang Y and Yang C: Long non-coding RNA HOTAIR modulates intervertebral disc degenerative changes via Wnt/ β -catenin pathway. *Arthritis Res Ther* 21: 201, 2019.
46. Zhu D, Yu Y, Wang W, Wu K, Liu D, Yang Y, Zhang C, Qi Y and Zhao S: Long noncoding RNA PART1 promotes progression of non-small cell lung cancer cells via JAK-STAT signaling pathway. *Cancer Med* 8: 6064-6081, 2019.
47. Zhou T, Wu L, Ma N, Tang F, Zong Z and Chen S: LncRNA PART1 regulates colorectal cancer via targeting miR-150-5p/miR-520h/CTNBN1 and activating Wnt/ β -catenin pathway. *Int J Biochem Cell Biol* 118: 105637, 2020.
48. Zhao B, Lu M, Wang D, Li H and He X: Genome-Wide identification of long noncoding RNAs in human intervertebral disc degeneration by RNA sequencing. *Biomed Res Int* 2016: 3684875, 2016.
49. Zhang Z, Huo Y, Zhou Z, Zhang P and Hu J: Role of lncRNA PART1 in intervertebral disc degeneration and associated underlying mechanism. *Exp Ther Med* 21: 131, 2021.
50. Xiang Q, Kang L, Wang J, Liao Z, Song Y, Zhao K, Wang K, Yang C and Zhang Y: CircRNA-CIDN mitigated compression loading-induced damage in human nucleus pulposus cells via miR-34a-5p/SIRT1 axis. *EBioMedicine* 53: 102679, 2020.
51. Chen Z, Zhang W, Deng M, Li Y and Zhou Y: CircGLCE alleviates intervertebral disc degeneration by regulating apoptosis and matrix degradation through the targeting of miR-587/STAP1. *Aging (Albany NY)* 12: 21971-21991, 2020.
52. Mizushima N and Komatsu M: Autophagy: Renovation of cells and tissues. *Cell* 147: 728-741, 2011.
53. Kim KH and Lee MS: Autophagy-a key player in cellular and body metabolism. *Nat Rev Endocrinol* 10: 322-337, 2014.
54. Kritschil R, Scott M, Sowa G and Vo N: Role of autophagy in intervertebral disc degeneration. *J Cell Physiol* 237: 1266-1284, 2022.
55. Wang F, Cai F, Shi R, Wang XH and Wu XT: Aging and age related stresses: A senescence mechanism of intervertebral disc degeneration. *Osteoarthritis Cartilage* 24: 398-408, 2016.
56. Li H, Tian L, Li J, Li Y, Du L, Huo Z and Xu B: The roles of circRNAs in intervertebral disc degeneration: Inflammation, extracellular matrix metabolism, and apoptosis. *Anal Cell Pathol (Amst)* 2022: 9550499, 2022.
57. Wu T, Jia X, Zhu Z, Guo K, Wang Q, Gao Z, Li X, Huang Y and Wu D: Inhibition of miR-130b-3p restores autophagy and attenuates intervertebral disc degeneration through mediating ATG14 and PRKAA1. *Apoptosis* 27: 409-425, 2022.
58. Yu X, Liu Q, Wang Y, Bao Y, Jiang Y, Li M, Li Z, Wang B, Yu L, Wang S, *et al*: Depleted Long Noncoding RNA GAS5 relieves intervertebral disc degeneration via microRNA-17-3p/Ang-2. *Oxid Med Cell Longev* 2022: 1792412, 2022.
59. Wen F, Yu J, He CJ, Zhang ZW and Yang AF: β -ecdysterone protects against apoptosis by promoting autophagy in nucleus pulposus cells and ameliorates disc degeneration. *Mol Med Rep* 19: 2440-2448, 2019.
60. Yu X, Wang ZL, Han CL, Wang MW, Jin Y, Jin XB and Xia QH: LncRNA CASC15 functions as an oncogene by sponging miR-130b-3p in bladder cancer. *Eur Rev Med Pharmacol Sci* 24: 7203, 2020.
61. Liao Y, Wang C, Yang Z, Liu W, Yuan Y, Li K, Zhang Y, Wang Y, Shi Y, Qiu Y, *et al*: Dysregulated Sp1/miR-130b-3p/HOXA5 axis contributes to tumor angiogenesis and progression of hepatocellular carcinoma. *Theranostics* 10: 5209-5224, 2020.
62. Guo Q, Zhu X, Wei R, Zhao L, Zhang Z, Yin X, Zhang Y, Chu C, Wang B and Li X: miR-130b-3p regulates M1 macrophage polarization via targeting IRF1. *J Cell Physiol* 236: 2008-2022, 2021.
63. Li S, Geng J, Xu X, Huang X, Leng D, Jiang D, Liang J, Wang C, Jiang D and Dai H: miR-130b-3p modulates epithelial-mesenchymal crosstalk in lung fibrosis by targeting IGF-1. *PLoS One* 11: e0150418, 2016.
64. Kalluri R and LeBleu VS: The biology, function, and biomedical applications of exosomes. *Science* 367: eaau6977, 2020.
65. Luo L, Jian X, Sun H, Qin J, Wang Y, Zhang J, Shen Z, Yang D, Li C, Zhao P, *et al*: Cartilage endplate stem cells inhibit intervertebral disc degeneration by releasing exosomes to nucleus pulposus cells to activate Akt/autophagy. *Stem Cells* 39: 467-481, 2021.
66. Chen D and Jiang X: Exosomes-derived miR-125-5p from cartilage endplate stem cells regulates autophagy and ECM metabolism in nucleus pulposus by targeting SUV38H1. *Exp Cell Res* 414: 113066, 2022.
67. Yao D, Zhou Z, Wang P, Zheng L, Huang Y, Duan Y, Liu B and Li Y: MiR-125-5p/IL-6R axis regulates macrophage inflammatory response and intestinal epithelial cell apoptosis in ulcerative colitis through JAK1/STAT3 and NF- κ B pathway. *Cell Cycle* 20: 2547-2564, 2021.
68. Sun L, Lian JX and Meng S: MiR-125a-5p promotes osteoclastogenesis by targeting TNFRSF1B. *Cell Mol Biol Lett* 24: 23, 2019.
69. Wang HQ, Yu XD, Liu ZH, Cheng X, Samartzis D, Jia LT, Wu SX, Huang J, Chen J and Luo ZJ: Deregulated miR-155 promotes Fas-mediated apoptosis in human intervertebral disc degeneration by targeting FADD and caspase-3. *J Pathol* 225: 232-242, 2011.
70. Zhang WL, Chen YF, Meng HZ, Du JJ, Luan GN, Wang HQ, Yang MW and Luo ZJ: Role of miR-155 in the regulation of MMP-16 expression in intervertebral disc degeneration. *J Orthop Res* 35: 1323-1334, 2017.

71. Zhou X, Li J, Teng J, Liu Y, Zhang D, Liu L and Zhang W: microRNA-155-3p attenuates intervertebral disc degeneration via inhibition of KDM3A and HIF1 α . *Inflamm Res* 70: 297-308, 2021.
72. Liu Z, Xu L, Zhang K, Guo B, Cui Z and Gao N: LINC00210 plays oncogenic roles in non-small cell lung cancer by sponging microRNA-328-5p. *Exp Ther Med* 19: 3325-3331, 2020.
73. Yan J, Wu LG, Zhang M, Fang T, Pan W, Zhao JL and Zhou Q: miR-328-5p induces human intervertebral disc degeneration by targeting WWP2. *Oxid Med Cell Longev* 2022: 3511967, 2022.
74. Li Z, Li X, Chen C, Li S, Shen J, Tse G, Chan MTV and Wu WKK: Long non-coding RNAs in nucleus pulposus cell function and intervertebral disc degeneration. *Cell Prolif* 51: e12483, 2018.
75. Mayama T, Marr AK and Kino T: Differential expression of glucocorticoid receptor noncoding RNA Repressor Gas5 in autoimmune and inflammatory diseases. *Horm Metab Res* 48: 550-557, 2016.
76. Kolenda T, Guglas K, Koczyńska M, Sobocińska J, Teresiak A, Bliźniak R and Lamperska K: Good or not good: Role of miR-18a in cancer biology. *Rep Pract Oncol Radiother* 25: 808-819, 2020.
77. Klijn C, Durinck S, Stawiski EW, Haverty PM, Jiang Z, Liu H, Degenhardt J, Mayba O, Gnad F, Liu J, *et al*: A comprehensive transcriptional portrait of human cancer cell lines. *Nat Biotechnol* 33: 306-312, 2015.
78. Xu L, Tan Y, Xu F and Zhang Y: Long noncoding RNA ADIRF antisense RNA 1 upregulates insulin receptor substrate 1 to decrease the aggressiveness of osteosarcoma by sponging microRNA-761. *Bioengineered* 13: 2028-2043, 2022.
79. Zhong H, Zhou Z, Guo L, Liu FS, Wang X, Li J, Lv GH and Zou MX: SERPINA1 is a hub gene associated with intervertebral disc degeneration grade and affects the nucleus pulposus cell phenotype through the ADIRF-AS1/miR-214-3p axis. *Transl Res* 245: 99-116, 2022.
80. Zhao W, Geng D, Li S, Chen Z and Sun M: LncRNA HOTAIR influences cell growth, migration, invasion, and apoptosis via the miR-20a-5p/HMGA2 axis in breast cancer. *Cancer Med* 7: 842-855, 2018.
81. Loewen G, Jayawickramarajah J, Zhuo Y and Shan B: Functions of lncRNA HOTAIR in lung cancer. *J Hematol Oncol* 7: 90, 2014.
82. Yang L, Peng X, Li Y, Zhang X, Ma Y, Wu C, Fan Q, Wei S, Li H and Liu J: Long non-coding RNA HOTAIR promotes exosome secretion by regulating RAB35 and SNAP23 in hepatocellular carcinoma. *Mol Cancer* 18: 78, 2019.
83. Wang X, Liu W, Wang P and Li S: RNA interference of long noncoding RNA HOTAIR suppresses autophagy and promotes apoptosis and sensitivity to cisplatin in oral squamous cell carcinoma. *J Oral Pathol Med* 47: 930-937, 2018.
84. Zhang S, Song S, Cui W, Liu X and Sun Z: Mechanism of long Noncoding RNA HOTAIR in nucleus pulposus cell autophagy and apoptosis in intervertebral disc degeneration. *Evid Based Complement Alternat Med* 2022: 8504601, 2022.
85. Wang H, Zhu Y, Cao L, Guo Z, Sun K, Qiu W and Fan H: circARL15 plays a critical role in intervertebral disc degeneration by modulating miR-431-5p/DISC1. *Front Genet* 12: 669598, 2021.
86. Wang R, Zhou X, Luo G, Zhang J, Yang M and Song C: CircRNA RERE Promotes the oxidative stress-induced apoptosis and autophagy of nucleus pulposus cells through the miR-299-5p/Galectin-3 Axis. *J Healthc Eng* 2021: 2771712, 2021.
87. Liu Y, Yang Y, Lin Y, Wei B, Hu X, Xu L, Zhang W and Lu J: N⁶-methyladenosine-modified circRNA RERE modulates osteoarthritis by regulating β -catenin ubiquitination and degradation. *Cell Prolif*: Jun 22, 2022 (Epub ahead of print).
88. Wang L, Wang P, Su X and Zhao B: Circ_0001658 promotes the proliferation and metastasis of osteosarcoma cells via regulating miR-382-5p/YB-1 axis. *Cell Biochem Funct* 38: 77-86, 2020.
89. Meng GD and Xu BS: Circular RNA hsa_circ_0001658 inhibits intervertebral disc degeneration development by regulating hsa-miR-181c-5p/FAS. *Comput Math Methods Med* 2021: 7853335, 2021.
90. Duan X, Yu X and Li Z: Circular RNA hsa_circ_0001658 regulates apoptosis and autophagy in gastric cancer through microRNA-182/Ras-related protein Rab-10 signaling axis. *Bioengineered* 13: 2387-2397, 2022.
91. Cosamalón-Gan I, Cosamalón-Gan T, Mattos-Piaggio G, Villar-Suárez V, García-Cosamalón J and Vega-Álvarez JA: Inflammation in the intervertebral disc herniation. *Neurocirugía (Astur: Engl Ed)* 32: 21-35, 2021.
92. Le Maitre CL, Hoyland JA and Freemont AJ: Catabolic cytokine expression in degenerate and herniated human intervertebral discs: IL-1beta and TNFalpha expression profile. *Arthritis Res Ther* 9: R77, 2007.
93. Risbud MV and Shapiro IM: Role of cytokines in intervertebral disc degeneration: Pain and disc content. *Nat Rev Rheumatol* 10: 44-56, 2014.
94. Zhang S, Song S, Zhuang Y, Hu J, Cui W, Wang X, Zhao Z, Liu X and Sun Z: Role of microRNA-15a-5p/Sox9/NF- κ B axis in inflammatory factors and apoptosis of murine nucleus pulposus cells in intervertebral disc degeneration. *Life Sci* 277: 119408, 2021.
95. Dong L and Dong B: miR-489-3p overexpression inhibits lipopolysaccharide-induced nucleus pulposus cell apoptosis, inflammation and extracellular matrix degradation via targeting Toll-like receptor 4. *Exp Ther Med* 22: 1323, 2021.
96. Ji Z, Guo R, Ma Z and Li H: Arctigenin inhibits apoptosis, extracellular matrix degradation, and inflammation in human nucleus pulposus cells by up-regulating miR-483-3p. *J Clin Lab Anal* 36: e24508, 2022.
97. Che Z, Xueqin J and Zhang Z: LncRNA OIP5-AS1 accelerates intervertebral disc degeneration by targeting miR-25-3p. *Bioengineered* 12: 11201-11212, 2021.
98. Zhang C, Yang H, Li Y, Huo P and Ma P: LncRNA OIP5-AS1 regulates oxidative low-density lipoprotein-mediated endothelial cell injury via miR-320a/LOX1 axis. *Mol Cell Biochem* 467: 15-25, 2020.
99. Zheng D, Wang B, Zhu X, Hu J, Sun J, Xuan J and Ge Z: LncRNA OIP5-AS1 inhibits osteoblast differentiation of valve interstitial cells via miR-137/TWIST11 axis. *Biochem Biophys Res Commun* 511: 826-832, 2019.
100. Zhi L, Zhao J, Zhao H, Qing Z, Liu H and Ma J: Downregulation of LncRNA OIP5-AS1 Induced by IL-1 β aggravates osteoarthritis via regulating miR-29b-3p/PGRN. *Cartilage* 13 (2_suppl): 1345S-1355S, 2021.
101. Chen Y, Ni H, Zhao Y, Chen K, Li M, Li C, Zhu X and Fu Q: Potential role of lncRNAs in contributing to pathogenesis of intervertebral disc degeneration based on microarray data. *Med Sci Monit* 21: 3449-3458, 2015.
102. Li T, Peng Y, Chen Y, Huang X, Li X, Zhang Z and Du J: Long intergenic non-coding RNA-00917 regulates the proliferation, inflammation, and pyroptosis of nucleus pulposus cells via targeting miR-149-5p/NOD-like receptor protein 1 axis. *Bioengineered* 13: 6036-6047, 2022.
103. Liao ZW, Fan ZW, Huang Y, Liang CY, Liu C, Huang S and Chen CW: Long non-coding RNA MT1DP interacts with miR-365 and induces apoptosis of nucleus pulposus cells by repressing NRF-2-induced anti-oxidation in lumbar disc herniation. *Ann Transl Med* 9: 151, 2021.
104. Huang JG, Tang X, Wang JJ, Liu J, Chen P and Sun Y: A circular RNA, circUSP36, accelerates endothelial cell dysfunction in atherosclerosis by adsorbing miR-637 to enhance WNT4 expression. *Bioengineered* 12: 6759-6770, 2021.
105. Liu Y and Zhang Y: Hsa_circ_0134111 promotes osteoarthritis progression by regulating miR-224-5p/CCL1 interaction. *Aging (Albany NY)* 13: 20383-20394, 2021.
106. Yan P, Sun C, Luan L, Han J, Qu Y, Zhou C and Xu D: Hsa_circ_0134111 promotes intervertebral disc degeneration via sponging miR-578. *Cell Death Discov* 8: 55, 2022.
107. Li Y, Wu X, Li J, Du L, Wang X, Cao J, Li H, Huo Z, Li G, Pan D, *et al*: Circ_0004354 might compete with circ_0040039 to induce NPCs death and inflammatory response by targeting miR-345-3p-FAF1/TP73 axis in intervertebral disc degeneration. *Oxid Med Cell Longev* 2022: 2776440, 2022.
108. Xin J, Wang Y, Zheng Z, Wang S, Na S and Zhang S: Treatment of intervertebral disc degeneration. *Orthop Surg* 14: 1271-1280, 2022.
109. Guo HY, Guo MK, Wan ZY, Song F and Wang HQ: Emerging evidence on noncoding-RNA regulatory machinery in intervertebral disc degeneration: A narrative review. *Arthritis Res Ther* 22: 270, 2020.
110. Fekrazad R, Naghdi N, Nokhbatolfighahaei H and Bagheri H: The combination of laser therapy and metal nanoparticles in cancer treatment originated from epithelial tissues: A literature review. *J Lasers Med Sci* 7: 62-75, 2016.
111. Zhao K, Li D, Shi C, Ma X, Rong G, Kang H, Wang X and Sun B: Biodegradable polymeric nanoparticles as the delivery carrier for drug. *Curr Drug Deliv* 13: 494-499, 2016.

