HYPOTHESIS

Infectious Agents and Cancer

Open Access



Could *Mycobacterium avium* subspecies *paratuberculosis* cause Crohn's disease, ulcerative colitis...and colorectal cancer?

Ellen S. Pierce

Abstract: Infectious agents are known causes of human cancers. Schistosoma japonicum and Schistosoma mansoni cause a percentage of colorectal cancers in countries where the respective Schistosoma species are prevalent. Colorectal cancer is a complication of ulcerative colitis and colonic Crohn's disease, the two main forms of idiopathic inflammatory bowel disease (IIBD). Mycobacterium avium subspecies paratuberculosis (MAP), the cause of a chronic intestinal disease in domestic and wild ruminants, is one suspected cause of IIBD. MAP may therefore be involved in the pathogenesis of IIBD-associated colorectal cancer as well as colorectal cancer in individuals without IIBD (sporadic colorectal cancer) in countries where MAP infection of domestic livestock is prevalent and MAP's presence in soil and water is extensive. MAP organisms have been identified in the intestines of patients with sporadic colorectal cancer and IIBD when high magnification, oil immersion light microscopy (×1000 total magnification rather than the usual ×400 total magnification) is used. Research has demonstrated MAP's ability to invade intestinal goblet cells and cause acute and chronic goblet cell hyperplasia. Goblet cell hyperplasia is the little-recognized initial pathologic lesion of sporadic colorectal cancer, referred to as transitional mucosa, aberrant crypt foci, goblet cell hyperplastic polyps or transitional polyps. It is the even lesser-recognized initial pathologic feature of IIBD, referred to as hypermucinous mucosa, hyperplastic-like mucosal change, serrated epithelial changes, flat serrated changes, goblet cell rich mucosa or epithelial hyperplasia. Goblet cell hyperplasia is the precursor lesion of adenomas and dysplasia in the classical colorectal cancer pathway, of sessile serrated adenomas and serrated dysplasia in the serrated colorectal cancer pathway, and of flat and elevated dysplasia and dysplasia-associated lesions or masses in IIBD-associated intestinal cancers. MAP's invasion of intestinal goblet cells may result in the initial pathologic lesion of IIBD and sporadic colorectal cancer. MAP's persistence in infected intestines may result in the eventual development of both IIBDassociated and sporadic colorectal cancer.

Keywords: Goblet, Carcinomas, Adenomas, Infection, Cancerization, Serrated, Transitional mucosa, Aberrant foci, Inflammatory bowel disease

Introduction

Infectious agents are known causes of human cancers [1– 3]. *Mycobacterium avium* subspecies *paratuberculosis* (MAP), the cause of a chronic intestinal disease in domestic and wild ruminants called Johne's disease [4], is a long suspected cause of Crohn's disease [5, 6] and a recently proposed cause of ulcerative colitis [7], the other main form of idiopathic inflammatory bowel disease (IIBD). If MAP causes IIBD, it may be one cause of the colorectal cancers that are a complication of IIBD [8, 9]. MAP may also be one cause of colorectal cancer in patients without

Correspondence: ellenpiercemd@gmail.com Spokane Valley, Washington, USA IIBD (sporadic colorectal cancer) in countries where MAP infection of domestic livestock is endemic [10] and MAP's contamination of soil [11] and water [12] is extensive.

The possibility that MAP is involved in the pathogenesis of colorectal cancer, in a patient with or without IIBD [13], is based on the following observations.

Other microorganisms are known causes of colorectal cancer

Schistosoma mansoni and *Schistosoma japonicum* cause a percentage of colorectal cancers in countries where the respective *Schistosoma* species are endemic [14–16].



© The Author(s). 2018 **Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

A particular lesion, goblet cell hyperplasia, is the littlerecognized initial pathologic lesion of sporadic colorectal cancer, ulcerative colitis and Crohn's disease

In 1969, Filipe and colleagues first described the histopathologic components of transitional mucosa [17–19], which will subsequently be referred to as "goblet cell hyperplasia" or the "goblet cell hyperplasia lesion" (see Additional file 1):

- 1. The actual goblet cell hyperplasia, simply an increase in the number of goblet cells lining the colonic crypts.
- 2. The hyperplastic goblet cells are hypertrophic, longer and plumber than normal.
- 3. The crypts lined by hyperplastic goblet cells are either longer and wider or shorter and wider than normal.

Other authors emphasized one additional feature of transitional mucosa, the greatly increased amount of extracellular mucus coating the lesion produced by the hypertrophic and hyperplastic goblet cells [20, 21].

Beginning in 1991, two groups published their gross and histologic tangential (parallel to the mucosal surface) visualization of transitional mucosa, noticing the crypts were wider than normal but not that they were lined predominantly or exclusively by goblet cells, and called their lesion "aberrant crypt foci," which is merely the goblet cell hyperplasia lesion in cross section [22–25].

In 2003, Torlakovic and colleagues [26] redefined the "hyperplastic" polyp as a serrated polyp and split the former hyperplastic polyp into two categories, the microvesicular type serrated polyp and the goblet cell type serrated polyp. They recognized that their goblet cell type serrated polyp is the precursor of the microvesicular type serrated polyp and noted its similarity to transitional mucosa, but they did not realize that it is the identical lesion as transitional mucosa [26].

Goblet cell hyperplasia is the rarely recognized initial pathologic lesion of Crohn's disease and therefore of Crohn's disease-associated intestinal cancers. Van Patter and colleagues' 1954 treatise on regional enteritis [27] described goblet cell hyperplasia as follows:

The epithelium of the small bowel normally contains a variable number of secreting units – the goblet cells. In the vicinity of the lesions, the number of goblet cells was increased enormously, frequently to the point of complete replacement of other epithelial elements [27].

They speculated that whatever caused Crohn's disease was the cause of the observed goblet cell hyperplasia: There is some evidence to suggest that the etiologic agent is to be found in the fecal stream and that it makes its first appearance in the proximal portion of the small bowel...If this agent resides in the fecal stream it may exert its influence on the normal epithelial cells in the region of the future lesion, causing them to be replaced by goblet cells [27].

A sparse literature discusses goblet cell hyperplasia and its prominent extracellular mucus component as major pathologic features of Crohn's disease [28, 29] and as the precursor lesion of epithelial dysplasia and therefore of Crohn's disease-associated intestinal cancers, calling the lesion hyperplastic-like mucosal change [30].

Described as "epithelial hyperplasia," "metaplastic changes," "goblet cell rich epithelium" or "hypermucinous mucosa," more subtle but more extensive goblet cell hyperplasia has occasionally [31–35] been recognized as the precursor of dysplasia and colorectal cancer in ulcerative colitis. A single article describes goblet cell hyperplasia in ulcerative colitis as such and documents its uniform presence in ulcerative colitis-affected colons with dysplasia [32].

Known as "transitional mucosa," goblet cell hyperplasia is the precursor of dysplasia and adenomas [36] in the classical colorectal cancer pathway [37]. Transitional mucosa lines the stalks of pedunculated polyps [38, 39], forms the bases of tubular and villous adenomas [38, 39] and surrounds colorectal carcinomas [18, 19, 40, 41]. Transitional mucosa is a major component of the field cancerization theory in colorectal cancer [42].

Known as the "goblet cell type serrated polyp" [26, 43], goblet cell hyperplasia is the precursor lesion of the microvesicular type serrated polyp [26] and therefore of the sessile serrated adenoma [43] – serrated dysplasia [44] – serrated carcinoma [45] serrated colorectal cancer pathway [46]. The "transitional polyp" [21, 47] has rarely been recognized as the precursor lesion in both classical and serrated colorectal cancer pathways [48].

Of course, dysplasia and colorectal cancer develop from the goblet cell hyperplasia lesion seen in cross section, aberrant crypt foci, by either [49] the classical [22–25, 36, 50–52] or serrated [49] pathways.

Known by its alternative names, including the recently rediscovered "flat serrated change" [53] or "serrated epithelial changes" [54–56], goblet cell hyperplasia is the precursor of flat and elevated dysplasia [57] and dysplasiaassociated lesions or masses [58] in IIBD-associated intestinal cancers as well as of classical adenomas in IIBD patients [59–62]. Like sporadic colorectal cancer patients, IIBD patients develop colorectal cancer by the classical or serrated pathways [63, 64]. Like in IIBD patients, the flat dysplasia ("flat adenoma") – flat carcinoma pathway occurs in sporadic colorectal cancer patients [52, 65–67].

Pathogenic microorganisms are the only natural cause of intestinal goblet cell hyperplasia

While small intestinal goblet cell hyperplasia results from azoxymethane administration [68] and massive small intestinal resection [69], pathogenic bacteria and parasites are the only natural causes of intestinal goblet cell hyperplasia [70, 71], including the protozoan parasite *Giardia lamblia/intestinalis* [72], the helminthes *Trichinella spiralis* [73] and *Nippostrongylis brasiliensis* [74, 75], the bacteria *Yersinia enterocolitica* [76] and various *Shigella* species [77].

Goblet cell hyperplasia results from infection with the human pathogenic helminths *Schistosoma mansoni* and *Schistosoma japonicum* [78, 79], where it has been specifically referred to as "transitional mucosa" [14] and is the precursor lesion of dysplasia and colorectal carcinoma in infected patients [14–16].

Since colonic type goblet cell hyperplasia caused by the human pathogenic bacterium *Helicobacter pylori* occurs in the stomach, where colonic type goblet cells are not normally present, it is called incomplete intestinal (colonic) metaplasia and is the immediate precursor lesion of gastric cancer [80, 81].

Goblet cell hyperplasia is the rarely recognized histopathologic feature of the resolving phase of the murine pathogenic bacterium *Citrobacter rodentium* (Fig. 1b) [82, 83], which is an animal model of IIBD [84], epithelial-mesenchymal transition and tumorigenesis [85, 86]. *Citrobacter rodentium*'s effects on and interactions with goblet cells have been documented to cause the more well-known pathologic features of transmissible murine colonic hyperplasia, including the elongation of crypts, "depletion" of the mucinogen granule compartment and variable shapes of the goblet cells (Fig. 1a) [87, 88]. A single article demonstrates MAP flooding into and hovering in clouds above human intestinal goblet cells [89]. MAP attaches to and invades bovine intestinal goblet cells [90, 91] and causes acute [91] and chronic [92] goblet cell hyperplasia.

The persistence of a microorganism within infected tissues is one way that microorganism causes cancer, with proposed carcinogenic mechanisms including cycles of chronic inflammation and repair, chronic hyperplasia ('proliferation') which destabilizes DNA and suppression of apoptosis [2, 3].

MAP has been accidentally discovered in the intestines of patients with sporadic colorectal cancer

A follow-up to an article demonstrating that MAP organisms are small and require oil immersion ($\times 100$ oil immersion objective or $\times 1000$ total magnification) to be identified by light microscopy [93] identified *Mycobacterium avium* organisms (of which MAP is a subspecies) in two of three control patients with sporadic colorectal cancer [94].

Conclusion: The possibility that MAP causes colorectal cancer is a testable hypothesis

MAP organisms may be concentrated [95] in the following locations:

1. in the extracellular mucus that is a prominent component of the goblet cell hyperplasia lesion and mucinous and serrated carcinomas, and comprises the "mucus cap" [96, 97] or "coat" [98] of sessile serrated adenomas, contravening current recommendations [43, 98] to carefully wash off this prominent histopathologic feature.





- 2. within the hypertrophic apical granule compartment of the hyperplastic goblet cells lining the goblet cell hyperplasia lesion.
- 3. in the lamina propria and submucosa of the goblet cell hyperplasia lesion and adenomas.
- 4. within the tumor stroma of colorectal cancers.

MAP can also be identified in humans by culture, polymerase chain reaction and antibody evaluations of tissue, blood and stool [99–107].

Additional file

Additional file 1: Descriptions and illustrations of the goblet cell hyperplasia lesion. The supplementary file discusses the descriptions and illustrations of the goblet cell hyperplasia lesion found in some of the references in the main text. (DOC 200 kb)

Abbreviations

IIBD: Idiopathic inflammatory bowel disease; H&E: Hematoxylin and eosin; MAP: *Mycobacterium avium* subspecies *paratuberculosis*; PAS: Periodic Acid-Schiff

Acknowledgements

My research would not be possible without the assistance of librarians past (Sandy Keno, Gail Leong and Kathryn Kane) and present (Dr. Beth Hill) at the Providence Sacred Heart Medical Center and Children's Hospital's Health Sciences Library in Spokane, Washington, now part of the Providence Library system, as well as the other libraries that participate in the FreeShare Library group within the Docline National Network of Libraries of Medicine. Dr. Bruce Vallance very kindly provided the photomicrographs for Fig. 1. Thank you to Judi Heidel of Perfectly Clear Copyediting Services for editing this paper.

Dedicated to the memory of Cyrus E. Rubin, M.D., mentor and friend.

Funding

None

Availability of data and materials

Not applicable

Authors' contributions

Not applicable

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The author declares that no competing interests exist.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 29 June 2017 Accepted: 12 December 2017 Published online: 04 January 2018

References

- Kuper H, Adami HO, Trichopoulos D. Infections as a major preventable cause of human cancer. J Intern Med. 2000;248:171–83.
- Lax AJ, Thomas W. How bacteria could cause cancer: one step at a time. Trends Microbiol. 2002;10:293–9.
- Vennervald BJ, Polman K. Helminths and malignancy. Parasite Immunol. 2009;31:686–96.

- 4. Clarke CJ. The pathology and pathogenesis of paratuberculosis in ruminants and other species. J Comp Pathol. 1997;116:217–61.
- Kuenstner JT, Naser S, Chamberlin W, Borody T, Graham DY, McNees A, Hermon-Taylor J, Hermon-Taylor A, Dow CT, Thayer W, et al: The consensus from the Mycobacterium Avium Ssp. paratuberculosis (MAP) conference 2017. Front Public Health 2017;5:208.
- Davis WC, Kuenstner JT, Singh SV: Resolution of Crohn's (Johne's) disease with antibiotics: what are the next steps? Expert Rev Gastroenterol Hepatol 2017;11:393-396.
- Pierce ES. Ulcerative colitis and Crohn's disease: is Mycobacterium Avium subspecies paratuberculosis the common villain? Gut Pathog. 2010;2:21.
- Kim ER, Chang DK. Colorectal cancer in inflammatory bowel disease: the risk, pathogenesis, prevention and diagnosis. World J Gastroenterol. 2014;20: 9872–81.
- Choi PM, Zelig MP. Similarity of colorectal cancer in Crohn's disease and ulcerative colitis: implications for carcinogenesis and prevention. Gut. 1994; 35:950–4.
- Lombard JE, Gardner IA, Jafarzadeh SR, Fossler CP, Harris B, Capsel RT, Wagner BA, Johnson WO. Herd-level prevalence of Mycobacterium Avium subsp. paratuberculosis infection in United States dairy herds in 2007. Prev Vet Med. 2013;108:234–8.
- Rhodes G, Henrys P, Thomson BC, Pickup RW. Mycobacterium Avium subspecies paratuberculosis is widely distributed in British soils and waters: implications for animal and human health. Environ Microbiol. 2013;15:2761–74.
- King DN, Donohue MJ, Vesper SJ, Villegas EN, Ware MW, Vogel ME, Furlong EF, Kolpin DW, Glassmeyer ST, Pfaller S. Microbial pathogens in source and treated waters from drinking water treatment plants in the United States and implications for human health. Sci Total Environ. 2016;562:987–95.
- Rhodes JM. Unifying hypothesis for inflammatory bowel disease and associated colon cancer: sticking the pieces together with sugar. Lancet. 1996;347:40–4.
- Ming-Chai C, Chi-Yuan C, Pei-Yu C, Jen-Chun H. Evolution of colorectal cancer in schistsosomiasis: transitional mucosal changes adjacent to large intestinal carcinoma in colectomy specimens. Cancer. 1980;46:1661–75.
- Madbouly KM, Senagore AJ, Mukerjee A, Hussien AM, Shehata MA, Navine P, Delaney CP, Fazio VW. Colorectal cancer in a population with endemic Schistosoma Mansoni: is this an at-risk population? Int J Color Dis. 2007;22:175–81.
- Wang M, QB W, He WB, Wang ZQ. Clinicopathological characteristics and prognosis of schistosomal colorectal cancer. Color Dis. 2016;18:1005–9.
- Filipe MI. Value of histochemical reactions for mucosubstances in the diagnosis of certain pathological conditions of the colon and rectum. Gut. 1969;10:577–86.
- Filipe MI, Branfoot AC. Abnormal patterns of mucus secretion in apparently normal mucosa of large intestine with carcinoma. Cancer. 1974;34:282–90.
- 19. Greaves P, Filipe MI, Branfoot AC. Transitional mucosa and survival in human colorectal cancer. Cancer. 1980;46:764–70.
- 20. Sundblad AS, Paz RA. Mucinous carcinomas of the colon and rectum and their relation to polyps. Cancer. 1982;50:2504–9.
- 21. Heilmann KL, Schmidbauer G, Schyma G. The transitional polyp of the colorectal mucosa. Pathol Res Pract. 1987;182:690–3.
- Pretlow TP, Barrow BJ, Ashton WS, O'Riordan MA, Pretlow TG, Jurcisek JA, Stellato TA. Aberrant crypts: putative preneoplastic foci in human colonic mucosa. Cancer Res. 1991;51:1564–7.
- Pretlow TP, O'Riordan MA, Pretlow TG, Stellato TA. Aberrant crypts in human colonic mucosa: putative preneoplastic lesions. J Cell Biochem Suppl. 1992; 16G:55–62.
- 24. Roncucci L, Stamp D, Medline A, Cullen JB, Bruce WR. Identification and quantification of aberrant crypt foci and microadenomas in the human colon. Hum Pathol. 1991;22:287–94.
- Roncucci L, Medline A, Bruce WR. Classification of aberrant crypt foci and microadenomas in human colon. Cancer Epidemiol Biomarkers Prev. 1991;1:57–60.
- Torlakovic E, Skovlund E, Snover DC, Torlakovic G, Nesland JM. Morphologic reappraisal of serrated colorectal polyps. Am J Surg Pathol. 2003;27:65–81.
- 27. Van Patter WN, Bargen JA, Dockerty MB, Feldman WH, Mayo CW, Waugh JM. Regional enteritis. Gastroenterology. 1954;26:347–450.
- Dvorak AM, Connell AB, Dickersin GR. Crohn's disease: a scanning electron microscopic study. Hum Pathol. 1979;10:165–77.

- Nagel E, Bartels M, Pichlmayr R. Scanning electron-microscopic lesions in Crohn's disease: relevance for the interpretation of postoperative recurrence. Gastroenterology. 1995;108:376–82.
- Kilgore SP, Sigel JE, Goldblum JR. Hyperplastic-like mucosal change in Crohn's disease: an unusual form of dysplasia? Modern Pathol. 2000;13:797–801.
- Shnitka TK. Current concepts of the pathogenesis and pathology of inflammatory lesions of the intestine. Can Med Assoc J. 1964;91:7–22.
- Vaiphei K, Saha M, Sharma BC, Bhasin DK, Singh K. Goblet cell status in idiopathic ulcerative colitis–implication in surveillance program. Indian J Pathol Microbiol. 2004;47:16–21.
- Andersen SN, Lovig T, Clausen OP, Bakka A, Fausa O, Rognum TO. Villous, hypermucinous mucosa in long standing ulcerative colitis shows high frequency of K-ras mutations. Gut. 1999;45:686–92.
- Jass JR, Sugihara K, Love SB. Basis of sialic acid heterogeneity in ulcerative colitis. J Clin Pathol. 1988;41:388–92.
- Jass JR, England J, Miller K. Value of mucin histochemistry in follow up surveillance of patients with long standing ulcerative colitis. J Clin Pathol. 1986;39:393–8.
- Takayama T, Katsuki S, Takahashi Y, Ohi M, Nojiri S, Sakamaki S, Kato J, Kogawa K, Miyake H, Niitsu Y. Aberrant crypt foci of the colon as precursors of adenoma and cancer. N Engl J Med. 1998;339:1277–84.
- Leslie A, Carey FA, Pratt NR, Steele RJ. The colorectal adenoma-carcinoma sequence. Br J Surg. 2002;89:845–60.
- Lanza G Jr, Altavilla G, Cavazzini L, Negrini R. Colonic mucosa adjacent to adenomas and hyperplastic polyps–a morphological and histochemical study. Histopathology. 1985;9:857–73.
- 39. Fenoglio-Preiser CM, Hutter RV. Colorectal polyps: pathologic diagnosis and clinical significance. CA Cancer J Clin. 1985;35:322–44.
- Dawson PA, Filipe MI. An ultrastructural and histochemical study of the mucous membrane adjacent to and remote from carcinoma of the colon. Cancer. 1976;37:2388–98.
- Shamsuddin AK, Weiss L, Phelps PC, Trump BF. Colon epithelium. IV. Human colon carcinogenesis. Changes in human colon mucosa adjacent to and remote from carcinomas of the colon. J Natl Cancer Inst. 1981;66:413–9.
- Patel A, Tripathi G, Gopalakrishnan K, Williams N, Arasaradnam RP. Field cancerisation in colorectal cancer: a new frontier or pastures past? World J Gastroenterol. 2015;21:3763–72.
- Rex DK, Ahnen DJ, Baron JA, Batts KP, Burke CA, Burt RW, Goldblum JR, Guillem JG, Kahi CJ, Kalady MF, et al. Serrated lesions of the colorectum: review and recommendations from an expert panel. The American journal of gastroenterology. 2012;107:1315–29. quiz 1314, 1330
- Batts KP. The pathology of serrated colorectal neoplasia: practical answers for common questions. Modern Pathol. 2015;28(Suppl 1):S80–7.
- Garcia-Solano J, Perez-Guillermo M, Conesa-Zamora P, Acosta-Ortega J, Trujillo-Santos J, Cerezuela-Fuentes P, Makinen MJ. Clinicopathologic study of 85 colorectal serrated adenocarcinomas: further insights into the full recognition of a new subset of colorectal carcinoma. Hum Pathol. 2010;41:1359–68.
- 46. Yamane L, Scapulatempo-Neto C, Reis RM, Guimaraes DP. Serrated pathway in colorectal carcinogenesis. World J Gastroenterol. 2014;20:2634–40.
- 47. Franzin G, Scarpa A, Dina R, Zamboni G, Fratton A. Transitional polyps of the colon. Endoscopy. 1982;14:174–5.
- Ansher AF, Lewis JH, Fleischer DE, Cattau EL Jr, Collen MJ, O'Kieffe DA, Korman LY, Benjamin SB. Hyperplastic colonic polyps as a marker for adenomatous colonic polyps. Am J Gastroenterol. 1989;84:113–7.
- Rosenberg DW, Yang S, Pleau DC, Greenspan EJ, Stevens RG, Rajan TV, Heinen CD, Levine J, Zhou Y, O'Brien MJ. Mutations in BRAF and KRAS differentially distinguish serrated versus non-serrated hyperplastic aberrant crypt foci in humans. Cancer Res. 2007;67:3551–4.
- 50. Cheng L, Lai MD. Aberrant crypt foci as microscopic precursors of colorectal cancer. World J Gastroenterol. 2003;9:2642–9.
- 51. Siu IM, Pretlow TG, Amini SB, Pretlow TP. Identification of dysplasia in human colonic aberrant crypt foci. Am J Pathol. 1997;150:1805–13.
- 52. Kuramoto S, Oohara T. Minute cancers arising de novo in the human large intestine. Cancer. 1988;61:829–34.
- Atwaibi M, Batts KP, Weinberg DI, McCabe RP. Mo1705 Flat Serrated Change: Does it Predict the Development of Colonic Mucosal Dysplasia in Inflammatory Bowel Disease? Gastroenterology. 2012;142:S-665.
- 54. Johnson DH, Khanna S, Smyrk TC, Loftus EV Jr, Anderson KS, Mahoney DW, Ahlquist DA, Kisiel JB. Detection rate and outcome of colonic serrated

epithelial changes in patients with ulcerative colitis or Crohn's colitis. Aliment Pharmacol Ther. 2014;39:1408–17.

- Parian AM, Koh JM, Badamas J, Giardiello FM, Montgomery EA, Lazarev M. 42 Serrated Epithelial Changes Are Associated With Colorectal Dysplasia in Inflammatory Bowel Disease. Gastroenterology. 2013;144:S-11.
- Parian A, Koh J, Limketkai BN, Eluri S, Rubin DT, Brant SR, Ha CY, Bayless TM, Giardiello F, Hart J, et al: Association between serrated epithelial changes and colorectal dysplasia in inflammatory bowel disease. Gastrointest Endosc 2016;84:87-95 e81.
- Matkowskyj KA, Chen ZE, Rao MS, Yang GY. Dysplastic lesions in inflammatory bowel disease: molecular pathogenesis to morphology. Arch Pathol Lab Med. 2013;137:338–50.
- Blackstone MO, Riddell RH, Rogers BH, Levin B. Dysplasia-associated lesion or mass (DALM) detected by colonoscopy in long-standing ulcerative colitis: an indication for colectomy. Gastroenterology. 1981;80:366–74.
- Torres C, Antonioli D, Odze RD. Polypoid dysplasia and adenomas in inflammatory bowel disease: a clinical, pathologic, and follow-up study of 89 polyps from 59 patients. Am J Surg Pathol. 1998;22:275–84.
- Engelsgjerd M, Farraye FA, Odze RD. Polypectomy may be adequate treatment for adenoma-like dysplastic lesions in chronic ulcerative colitis. Gastroenterology. 1999;117:1288–94. discussion 1488-1291
- Quinn AM, Farraye FA, Naini BV, Cerda S, Coukos J, Li Y, Khor T, Odze RD. Polypectomy is adequate treatment for adenoma-like dysplastic lesions (DALMs) in Crohn's disease. Inflamm Bowel Dis. 2013;19:1186–93.
- Neumann H, Vieth M, Langner C, Neurath MF, Mudter J. Cancer risk in IBD: how to diagnose and how to manage DALM and ALM. World J Gastroenterol. 2011;17:3184–91.
- Bossard C, Denis MG, Bezieau S, Bach-Ngohou K, Bourreille A, Laboisse CL, Mosnier JF. Involvement of the serrated neoplasia pathway in inflammatory bowel disease-related colorectal oncogenesis. Oncol Rep. 2007;18:1093–7.
- Shen J, Gibson JA, Schulte S, Khurana H, Farraye FA, Levine J, Burakoff R, Cerda S, Qazi T, Hamilton M, et al. Clinical, pathologic, and outcome study of hyperplastic and sessile serrated polyps in inflammatory bowel disease. Hum Pathol. 2015;46:1548–56.
- Geboes K, De Hertogh G, Bisschops R, Geboes K. Flat adenomas, significance, detection, treatment. Ann Gastroenterol. 2010;23:266–9.
- Naravadi V, Gupta N, Early D, Jonnalagadda S, Wani SB, Gaddam S, Sharma P, Edmundowicz SA, Bansal A, Rastogi A. Prevalence of advanced histological features and synchronous neoplasia in patients with flat adenomas. Gastrointest Endosc. 2016;83:795–9.
- 67. Zhan T, Hahn F, Hielscher T, Betge J, Kahler G, Ebert MP, Belle S. Frequent co-occurrence of high-grade dysplasia in large flat colonic polyps (>20 mm) and synchronous polyps. BMC Gastroenterol. 2015;15:82.
- Olubuyide IO, Bristol JB, Williamson RC. Goblet cell changes during intestinal adaptation to azoxymethane and enteric bypass in the rat. Br J Cancer. 1985;51:383–8.
- Haxhija EQ, Yang H, Spencer AU, Sun X, Teitelbaum DH. Intestinal epithelial cell proliferation is dependent on the site of massive small bowel resection. Pediatr Surg Int. 2007;23:379–90.
- Deplancke B, Gaskins HR. Microbial modulation of innate defense: goblet cells and the intestinal mucus layer. Am J Clin Nutr. 2001;73:11315–415.
- 71. Kim YS, Ho SB. Intestinal goblet cells and mucins in health and disease: recent insights and progress. Curr Gastroenterol Rep. 2010;12:319–30.
- Ponce-Macotela M, Gonzalez-Maciel A, Reynoso-Robles R, Martinez-Gordillo MN. Goblet cells: are they an unspecific barrier against Giardia Intestinalis or a gate? Parasitol Res. 2008;102:509–13.
- Knight PA, Brown JK, Pemberton AD. Innate immune response mechanisms in the intestinal epithelium: potential roles for mast cells and goblet cells in the expulsion of adult Trichinella spiralis. Parasitology. 2008;135:655–70.
- Soga K, Yamauchi J, Kawai Y, Yamada M, Uchikawa R, Tegoshi T, Mitsufuji S, Yoshikawa T, Arizono N. Alteration of the expression profiles of acidic mucin, sialytransferase, and sulfotransferases in the intestinal epithelium of rats infected with the nematode Nippostrongylus brasiliensis. Parasitol Res. 2008;103:1427–34.
- Moro K, Yamada T, Tanabe M, Takeuchi T, Ikawa T, Kawamoto H, Furusawa J, Ohtani M, Fujii H, Koyasu S. Innate production of T(H)2 cytokines by adipose tissue-associated c-kit(+)Sca-1(+) lymphoid cells. Nature. 2010;463:540–4.
- 76. Mantle M, Atkins E, Kelly J, Thakore E, Buret A, Gall DG. Effects of Yersinia enterocolitica infection on rabbit intestinal and colonic goblet

cells and mucin: morphometrics, histochemistry, and biochemistry. Gut. 1991;32:1131–8.

- Yang JY, Lee SN, Chang SY, Ko HJ, Ryu S, Kweon MN. A mouse model of shigellosis by intraperitoneal infection. J Infect Dis. 2014;209:203–15.
- Marillier RG, Michels C, Smith EM, Fick LC, Leeto M, Dewals B, Horsnell WG, Brombacher F. IL-4/IL-13 independent goblet cell hyperplasia in experimental helminth infections. BMC Immunol. 2008;9:11.
- Couto JL, Ferreira Hda S, da Rocha DB, Duarte ME, Assuncao ML, Coutinho Ede M. Structural changes in the jejunal mucosa of mice infected with Schistosoma Mansoni, fed low or high protein diets. Rev Soc Bras Med Trop. 2002;35:601–7.
- Correa P, Houghton J. Carcinogenesis of helicobacter pylori. Gastroenterology. 2007;133:659–72.
- Semino-Mora C, Doi SQ, Marty A, Simko V, Carlstedt I, Dubois A. Intracellular and interstitial expression of helicobacter pylori virulence genes in gastric precancerous intestinal metaplasia and adenocarcinoma. J Infect Dis. 2003; 187:1165–77.
- Barthold SW, Coleman GL, Jacoby RO, Livestone EM, Jonas AM. Transmissible murine colonic hyperplasia. Vet Pathol. 1978;15:223–36.
- Bergstrom KS, Morampudi V, Chan JM, Bhinder G, Lau J, Yang H, Ma C, Huang T, Ryz N, Sham HP, et al. Goblet cell derived RELM-beta recruits CD4 + T cells during infectious colitis to promote protective intestinal epithelial cell proliferation. PLoS Pathog. 2015;11:e1005108.
- Higgins LM, Frankel G, Douce G, Dougan G, MacDonald TT. Citrobacter rodentium infection in mice elicits a mucosal Th1 cytokine response and lesions similar to those in murine inflammatory bowel disease. Infect Immun. 1999;67:3031–9.
- Chandrakesan P, Roy B, Jakkula LU, Ahmed I, Ramamoorthy P, Tawfik O, Papineni R, Houchen C, Anant S, Umar S. Utility of a bacterial infection model to study epithelial-mesenchymal transition, mesenchymalepithelial transition or tumorigenesis. Oncogene. 2014;33:2639–54.
- Newman JV, Kosaka T, Sheppard BJ, Fox JG, Schauer DB. Bacterial infection promotes colon tumorigenesis in Apc(min/+) mice. J Infect Dis. 2001;184:227–30.
- Bergstrom KS, Guttman JA, Rumi M, Ma C, Bouzari S, Khan MA, Gibson DL, Vogl AW, Vallance BA. Modulation of intestinal goblet cell function during infection by an attaching and effacing bacterial pathogen. Infect Immun. 2008;76:796–811.
- Chan JM, Bhinder G, Sham HP, Ryz N, Huang T, Bergstrom KS, Vallance BA. CD4+ T cells drive goblet cell depletion during Citrobacter rodentium infection. Infect Immun. 2013;81:4649–58.
- Golan L, Livneh-Kol A, Gonen E, Yagel S, Rosenshine I, Shpigel NY. Mycobacterium Avium paratuberculosis invades human small-intestinal goblet cells and elicits inflammation. J Infect Dis. 2009;199:350–4.
- Schleig PM, Buergelt CD, Davis JK, Williams E, Monif GR, Davidson MK. Attachment of Mycobacterium Avium subspecies paratuberculosis to bovine intestinal organ cultures: method development and strain differences. Vet Microbiol. 2005;108:271–9.
- 91. Khare S, Nunes JS, Figueiredo JF, Lawhon SD, Rossetti CA, Gull T, Rice-Ficht AC, Adams LG. Early phase morphological lesions and transcriptional responses of bovine ileum infected with Mycobacterium Avium subsp. paratuberculosis. Vet Pathol. 2009;46:717–28.
- Charavaryamath C, Gonzalez-Cano P, Fries P, Gomis S, Doig K, Scruten E, Potter A, Napper S, Griebel PJ. Host responses to persistent Mycobacterium Avium subspecies paratuberculosis infection in surgically isolated bovine ileal segments. Clin Vac Immunol. 2013;20:156–65.
- Jeyanathan M, Alexander DC, Turenne CY, Girard C, Behr MA. Evaluation of in situ methods used to detect Mycobacterium Avium subsp. paratuberculosis in samples from patients with Crohn's disease. J Clin Microbiol. 2006;44:2942–50.
- Jeyanathan M, Boutros-Tadros O, Radhi J, Semret M, Bitton A, Behr MA. Visualization of Mycobacterium Avium in Crohn's tissue by oilimmersion microscopy. Microbes Infect. 2007;9:1567–73.
- 95. Pierce ES. Where are all the Mycobacterium Avium subspecies paratuberculosis in patients with Crohn's disease? PLoS Pathog. 2009;5:e1000234.
- Lee EJ, Kim MJ, Chun SM, Jang SJ, Kim DS, Lee DH, Youk EG. Sessile serrated adenoma/polyps with a depressed surface: a rare form of sessile serrated adenoma/polyp. Diagn Pathol. 2015;10:75.
- 97. Pereyra L, Gomez EJ, Gonzalez R, Fischer C, Erana GB, Torres AG, Correa L, Mella JM, Panigadi GN, Luna P, et al. Finding sessile serrated adenomas: is it

possible to identify them during conventional colonoscopy? Dig Dis Sci. 2014;59:3021–6.

- Sweetser S, Smyrk TC, Sugumar A. Serrated polyps: critical precursors to colorectal cancer. Expert Rev Gastroenterol Hepatol. 2011;5:627–35.
- Naser SA, Ghobrial G, Romero C, Valentine JF. Culture of Mycobacterium Avium subspecies paratuberculosis from the blood of patients with Crohn's disease. Lancet. 2004;364:1039–44.
- Timms VJ, Daskalopoulos G, Mitchell HM, Neilan BA. The association of Mycobacterium Avium subsp. paratuberculosis with inflammatory bowel disease. PLoS One. 2016;11:e0148731.
- 101. Singh AV, Singh SV, Makharia GK, Singh PK, Sohal JS. Presence and characterization of Mycobacterium Avium subspecies paratuberculosis from clinical and suspected cases of Crohn's disease and in the healthy human population in India. Int J Infect Dis. 2008;12:190–7.
- 102. Banche G, Allizond V, Sostegni R, Lavagna A, Bergallo M, Sidoti F, Daperno M, Rocca R, Cuffini AM. Application of multiple laboratory tests for Mycobacterium Avium Ssp. paratuberculosis detection in Crohn's disease patient specimens. New Microbiol. 2015;38:357–67.
- 103. Singh SV, Kumar N, Sohal JS, Singh AV, Singh PK, Agrawal ND, Gupta S, Chaubey KK, Kumar A, Rawat KD. First mass screening of the human population to estimate the bio-load of Mycobacterium Avium subspecies paratuberculosis in North India. J Biol Sci. 2014;14:237.
- 104. Singh SV, Kuenstner JT, Davis WC, Agarwal P, Kumar N, Singh D, Gupta S, Chaubey KK, Kumar A, Misri J, et al. Concurrent resolution of chronic diarrhea likely due to Crohn's disease and infection with Mycobacterium Avium paratuberculosis. Front Med. 2016;3:49.
- 105. Tuci A, Tonon F, Castellani L, Sartini A, Roda G, Marocchi M, Caponi A, Munarini A, Rosati G, Ugolini G, et al. Fecal detection of Mycobacterium Avium paratuberculosis using the IS900 DNA sequence in Crohn's disease and ulcerative colitis patients and healthy subjects. Dig Dis Sci. 2011;56: 2957–62.
- 106. Feller M, Huwiler K, Stephan R, Altpeter E, Shang A, Furrer H, Pfyffer GE, Jemmi T, Baumgartner A, Egger M. Mycobacterium Avium subspecies paratuberculosis and Crohn's disease: a systematic review and meta-analysis. Lancet Infect Dis. 2007;7:607–13.
- 107. Abubakar I, Myhill D, Aliyu SH, Hunter PR. Detection of Mycobacterium Avium subspecies paratuberculosis from patients with Crohn's disease using nucleic acid-based techniques: a systematic review and meta-analysis. Inflamm Bowel Dis. 2008;14:401–10.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at www.biomedcentral.com/submit

