



## OPEN ACCESS

EDITED BY  
Jian Li,  
Monash University, Australia

REVIEWED BY  
Jing Yang,  
First Affiliated Hospital of Zhengzhou  
University, China  
Yu-Wei Lin,  
Monash University, Australia  
Cornelia Landersdorfer,  
Monash University, Australia

\*CORRESPONDENCE  
Huadong Chen,  
chuadong666@163.com

SPECIALTY SECTION  
This article was submitted to  
Pharmacology of Infectious Diseases,  
a section of the journal  
Frontiers in Pharmacology

RECEIVED 12 July 2022  
ACCEPTED 17 August 2022  
PUBLISHED 09 September 2022

CITATION  
Chen H and Li P (2022), Commentary:  
Population pharmacokinetics of colistin  
sulfate in critically ill patients: Exposure  
and clinical efficacy.  
*Front. Pharmacol.* 13:992085.  
doi: 10.3389/fphar.2022.992085

COPYRIGHT  
© 2022 Chen and Li. This is an open-  
access article distributed under the  
terms of the [Creative Commons  
Attribution License \(CC BY\)](#). The use,  
distribution or reproduction in other  
forums is permitted, provided the  
original author(s) and the copyright  
owner(s) are credited and that the  
original publication in this journal is  
cited, in accordance with accepted  
academic practice. No use, distribution  
or reproduction is permitted which does  
not comply with these terms.

# Commentary: Population pharmacokinetics of colistin sulfate in critically ill patients: Exposure and clinical efficacy

Huadong Chen\* and Piaopiao Li

Department of Pharmacy, Affiliated Dongyang Hospital of Wenzhou Medical University, Dongyang, China

## KEYWORDS

polymyxin, colistin, pharmacokinetic, pharmacodynamic, dose optimization

## A Commentary on

### Population pharmacokinetics of colistin sulfate in critically ill patients: Exposure and clinical efficacy

by Yu X-b, Zhang X-S, Wang Y-X, Wang Y-Z, Zhou H-M, Xu F-M, Yu J-H, Zhang L-W, Dai Y, Zhou Z-Y, Zhang C-H, Lin G-Y and Pan J-Y (2022). *Front. Pharmacol.* 13:915958. doi: 10.3389/fphar.2022.915958

## Introduction

Recently, Yu et al., 2022 published a study of great interest “Population pharmacokinetics of colistin sulfate in critically ill patients: Exposure and clinical efficacy” in *Frontiers in Pharmacology*. Polymyxins are considered as last-resort for treatment of infection caused by carbapenem resistant organisms (Heavner et al., 2018). Usually, there are two available polymyxin products, polymyxin B sulfate and sodium colistin methanesulfonate. But a new form of polymyxin for intravenous use, colistin sulfate, came back to Chinese market recently. It has a similar chemical structure as polymyxin B but usually does not cause skin hyperpigmentation after intravenous administration (Nahhas et al., 2019). This population pharmacokinetic study of colistin sulfate is important for its optimal use. However, we believe that some weak points in the research should be addressed and this would benefit readers.

## Dose of colistin sulfate

Different conventions are used to define the dose of polymyxins, and this has caused significant confusions in polymyxin dosing (Li et al., 2006; Nation et al., 2015). The product of colistin sulfate is labeled as 500,000 IU per vial. But we used mg/L when we

determined the MIC of bacteria or drug concentration in plasma. The insert of colistin sulfate does not provide the convention of mg and IU, and there are few papers about the convention. Yu et al. claimed that according to an expert consensus by the Infectious Disease Society of China, 1 mg of pure colistin base equals 17,000 IU of colistin (Infectious Diseases Society of China, 2021). It is not correct. The expert consensus clearly indicated that 1 mg of colistin sulfate equals approximately 22,300 IU (Infectious Diseases Society of China, 2021). In the Chinese Pharmacopeia, it says that the titer of 1 mg colistin sulfate should be no less than 17,000 IU (Chinese Pharmacopeia Committee, 2015). But it is the low-limit of potency and should not be used for convention. The manufacture should improve the label and convention of colistin sulfate to facilitate its use.

## Pharmacokinetic/pharmacodynamic modeling

Yu et al. had developed a population pharmacokinetic model for colistin sulfate, and the model only accounts for intravenous administration. But 52.4% of the included patients received inhaled colistin sulfate. There is no data on how much of inhaled colistin can reach system circulation. If the amount reaching the systemic circulation was not negligible, then the simulations from the developed population PK model would probably overestimate the plasma concentration given that input from other routes of administration was ignored during model development.

The result of Monte Carlo simulation by Yu et al. indicated that the probability of target attainment (PTA) of PKPD target was satisfied when the minimum inhibitory concentration (MIC) of bacteria was 0.5 mg/L. But for pathogens with an MIC of 1 mg/L, only 3 off-label dosages (the maximum daily dose was 1.5 million IU) had PTA > 90% in patients with different renal function levels. However, pathogens with MICs of 0.5 mg/L or below only take a small part. EUCAST MIC distribution data indicated that the percentages of pathogens with MICs of 0.5 mg/L or below for common multidrug-resistant bacteria, *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, and *Klebsiella pneumoniae* were 8.3%, 27.9%, and 76.9%, respectively (European Committee on Antimicrobial Susceptibility Testing, 2022).

We have also run Monte Carlo simulation based on the published final model using NONMEM (version 7.5.0, ICON, Ellicott City, MD, United States) coupled with PDxPop (version 5.3, ICON, Gaithersburg, MD, United States). Virtual patients (10,000 patients per CLCR level) infected by bacteria with EUCAST MIC distribution were generated (European Committee on Antimicrobial Susceptibility Testing, 2022). The ratio of the unbound concentration–time curve to the MIC ( $fAUC/MIC$ ) was also used as the PK/PD index, and the

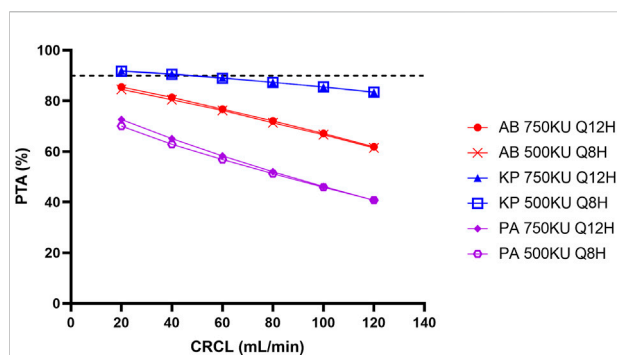


FIGURE 1

Simulated probability of achieving target attainment (PTA) of colistin sulfate at maximum daily dose for different micro-organisms. AB, *Acinetobacter baumannii*; KP, *Klebsiella pneumoniae*; PA, *Pseudomonas aeruginosa*; CLCR, creatinine clearance rate.

target  $fAUC/MIC$  was also set to 15 for *Pseudomonas aeruginosa* and *Acinetobacter baumannii*. The target  $fAUC/MIC$  for *Klebsiella pneumoniae* was set to 17.4, which was the target of polymyxin B for 1-log kill of bacteria in murine thigh infection model (Landersdorfer et al., 2018). AUCs at steady state of virtual patients were calculated using the pkr package (version 0.1.3) in R software (version 4.0.5) with a linear-up and linear-down method. The unbound fraction was the same as in the original study. The result is shown in Figure 1. The PTAs of the maximum daily dose (divided into 2 or 3 doses) were all below 90% for *Pseudomonas aeruginosa* and *Acinetobacter baumannii*. The MICs of *Klebsiella pneumoniae* were lower, but the PTAs achieved 90% only for patients with poor renal function. It should be noted that the PK/PD targets used in the simulation correspond to 1-log kill of bacteria in the animal model, but the targets for 2-log kill of bacteria are much higher (Tsuji et al., 2019). For some type of infection, such as lung infection, even the maximum tolerated dose cannot achieve bacterial stasis. Thus, from the perspective of current PK/PD data, intravenous colistin sulfate can only be used for infections caused by bacteria with very low MICs (e.g., 0.5 mg/L or below).

Moreover, Yu et al. stated that the PTA was higher for the dosing interval of 12 h than that of 8 h, at the same daily dose. This is not possible for an AUC-based PK/PD index and a PK model with linear elimination.

## Nephrotoxicity and clinical efficacy

Among the 42 included patients, only 2 patients developed acute kidney injury (AKI). Yu et al. claimed that the adverse events were not associated with colistin use but no detail was provided. Readers cannot judge whether those who experienced AKI were due to exposure to higher colistin concentration or due

to other confounding factors. The prevalence of AKI in this study is quite low, as the reported prevalence of AKI in ICU patients was around 30% in other literatures (Chiofolo et al., 2019). The sample size of Yu et al.'s report was too limited to make any meaningful conclusions.

The 30-day mortality rate was 23.8% in Yu et al.'s study, and this result was satisfied in patients with drug-resistant bacterial infection. However, there may be no relationship between systemic colistin exposure and clinical efficacy. We noticed that nearly 70% of patients had respiratory tract infections, however, both preclinical and clinical studies indicated that intravenous polymyxins were suboptimal for lung infection (Cheah et al., 2015; Tsuji et al., 2019). The included patients also received other antibiotics (52.4% carbapenems, 47.6% cefoperazone-sulbactam, and other antibiotics) and inhaled colistin (52.4% of the patients), which may contribute to the low mortality, rather than intravenous colistin sulfate.

## Conclusion

A clear convention for the dose of colistin sulfate is urgently needed. PK/PD modeling showed that the PTA of colistin sulfate was poor for common hospital-acquired infections caused by *P. aeruginosa*, *A. baumannii*, and *K. pneumoniae*, even at the highest daily dose. Based on the very limited literature, intravenous colistin sulfate can only be used when the MIC of

the pathogen is below 0.5 mg/L. The dose of colistin sulfate should be further optimized by balancing efficacy and toxicity.

## Author contributions

Conceptualization, HC; Formal analysis, PL; Writing—original draft, HC and PL; Writing—review and editing, HC.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## References

- Cheah, S. E., Wang, J., Nguyen, V. T. T., Turnidge, J. D., Li, J., and Nation, R. L. (2015). New pharmacokinetic/pharmacodynamic studies of systemically administered colistin against *Pseudomonas aeruginosa* and acinetobacter baumannii in mouse thigh and lung infection models: Smaller response in lung infection. *J. Antimicrob. Chemother.* 70, 3291–3297. doi:10.1093/jac/dkv267
- Chinese Pharmacopoeia Committee (2015). *Chinese Pharmacopoeia, section 2*. Beijing.
- Chiofolo, C., Chbat, N., Ghosh, E., Eshelman, L., and Kashani, K. (2019). Automated continuous acute kidney injury prediction and surveillance: A random forest model. *Mayo Clin. Proc.* 94, 783–792. doi:10.1016/j.mayocp.2019.02.009
- European Committee on Antimicrobial Susceptibility Testing Data from the EUCAST MIC distribution website. Available at: <https://mic.eucast.org/> (accessed Jul 9, 2022).
- Heavner, M. S., Claeys, K. C., Masich, A. M., and Gonzales, J. P. (2018). Pharmacokinetic and pharmacodynamic considerations of antibiotics of last resort in treating gram-negative infections in adult critically ill patients. *Curr. Infect. Dis. Rep.* 20, 10. doi:10.1007/s11908-018-0614-0
- Infectious Diseases Society of China (2021). Multi-disciplinary expert consensus on the optimal clinical use of the polymyxins in China. *Chin. J. Tubercul. Respir. Dis.* 44 (4), 292–310. doi:10.3760/cma.j.cn112147-20201109-01091
- Landersdorfer, C. B., Wang, J., Wirth, V., Chen, K., Kaye, K. S., Tsuji, B. T., et al. (2018). Pharmacokinetics/pharmacodynamics of systemically administered polymyxin B against *Klebsiella pneumoniae* in mouse thigh and lung infection models. *J. Antimicrob. Chemother.* 73, 462–468. doi:10.1093/jac/dkx409
- Li, J., Nation, R. L., Turnidge, J. D., Milne, R. W., Coulthard, K., Rayner, C. R., et al. (2006). Colistin: The re-emerging antibiotic for multidrug-resistant gram-negative bacterial infections. *Lancet. Infect. Dis.* 6 (9), 589–601. doi:10.1016/S1473-3099(06)70580-1
- Nahas, A. F., Braunberger, T. L., and Hamzavi, I. H. (2019). An update on drug-induced pigmentation. *Am. J. Clin. Dermatol.* 20, 75–96. doi:10.1007/s40257-018-0393-2
- Nation, R. L., Li, J., Cars, O., Couet, W., Dudley, M. N., Kaye, K. S., et al. (2015). Framework for optimisation of the clinical use of colistin and polymyxin B: The Prato polymyxin consensus. *Lancet. Infect. Dis.* 15, 225–234. doi:10.1016/S1473-3099(14)70850-3
- Tsuji, B. T., Pogue, J. M., Zavascki, A. P., Paul, M., Daikos, G. L., Forrest, A., et al. (2019). International consensus guidelines for the optimal use of the polymyxins: Endorsed by the American college of clinical pharmacy (ACCP), European society of clinical microbiology and infectious Diseases (ESCMID), infectious Diseases society of America (IDSA), international society for anti-infective Pharmacology (ISAP), society of critical care medicine (SCCM), and society of infectious Diseases pharmacists (SIDP). *Pharmacotherapy* 39, 10–39. doi:10.1002/phar.2209
- Yu, X., Zhang, X., Wang, Y.-X., Wang, Y., Zhou, H., Xu, F.-M., et al. (2022). Population pharmacokinetics of colistin sulfate in critically ill patients: Exposure and clinical efficacy. *Front. Pharmacol.* 13, 915958–915959. doi:10.3389/fphar.2022.915958