

Capillaries as a Therapeutic Target for Heart Failure

Yohko Yoshida^{1,2}, Ippei Shimizu¹ and Tohru Minamino^{1,3}

Yohko Yoshida, Ippei Shimizu and Tohru Minamino are joint senior authors.

¹Department of Cardiovascular Biology and Medicine, Juntendo University Graduate School of Medicine, Tokyo, Japan

²Department of Advanced Senotherapeutics, Juntendo University Graduate School of Medicine, Tokyo, Japan

³Japan Agency for Medical Research and Development-Core Research for Evolutionary Medical Science and Technology (AMED-CREST), Japan Agency for Medical Research and Development, Tokyo, Japan.

Prognosis of heart failure remains poor, and it is urgent to find new therapies for this critical condition. Oxygen and metabolites are delivered through capillaries; therefore, they have critical roles in the maintenance of cardiac function. With aging or age-related disorders, capillary density is reduced in the heart, and the mechanisms involved in these processes were reported to suppress capillarization in this organ. Studies with rodents showed capillary rarefaction has causal roles for promoting pathologies in failing hearts. Drugs used as first-line therapies for heart failure were also shown to enhance the capillary network in the heart. Recently, the approach with senolysis is attracting enthusiasm in aging research. Genetic or pharmacological approaches concluded that the specific depletion of senescent cells, senolysis, led to reverse aging phenotype. Reagents mediating senolysis are described to be senolytics, and these compounds were shown to ameliorate cardiac dysfunction together with enhancement of capillarization in heart failure models. Studies indicate maintenance of the capillary network as critical for inhibition of pathologies in heart failure.

Key words: Heart failure, Capillaries, Aging, Senolysis

Introduction

Oxygen and metabolites are delivered through capillaries, and this platform has a critical role in maintaining organ homeostasis. With aging, capillary density was reported to diminish in heart, skeletal muscle, kidney, brain, liver, subdermal or abdominal white adipose tissues, pancreas, testis, thyroid gland, and brown adipose tissue¹⁻⁵). Aging associates with inhibition in vascular endothelial growth factor-A (VEGF-A) mediated angiogenic signaling, and this was reported to enhance aging phenotype in multiple organs¹). Accumulation of evidence indicates a decrease in capillary density accelerates undesirable aspects of aging.

Incidence of heart failure increases with age, and the prognosis of this disorder remains poor. Aged

hearts are characterized by fibrosis, inflammation, mitochondrial dysfunction, apoptosis⁶), and capillary density was reported to decline with aging in rodents and humans^{2, 7}). Studies indicate capillary rarefaction in the left ventricle (LV) has a close connection with functional decline in cardiac tissue⁸⁻¹⁰). Depending on the systolic or diastolic function of the LV, heart failure can be categorized into two groups. One is described as heart failure with reduced ejection fraction (HFrEF) and the other as heart failure with preserved ejection fraction (HFpEF). Recent studies showed sodium glucose co-transporter2 (SGLT2) inhibitors contributed to the suppression of hospitalization for heart failure both in patients with HFrEF and those with HFpEF¹¹⁻¹³). Compared with HFrEF, medication for HFpEF is limited¹⁴), and next-generation therapies for this critical disorder should be

Address for correspondence: Ippei Shimizu, Department of Cardiovascular Biology and Medicine, Juntendo University Graduate School of Medicine, 2-1-1 Hongo, Bunkyo-ku, Tokyo 113-8421, Japan E-mail: s.shimizu@juntendo.ac.jp, ippeishimizu@yahoo.co.jp

Received: January 28, 2022 Accepted for publication: February 21, 2022

Copyright©2022 Japan Atherosclerosis Society

This article is distributed under the terms of the latest version of CC BY-NC-SA defined by the Creative Commons Attribution License.

urgently established. HFpEF in patients increases with age, and this is associated with capillary rarefaction in LV¹⁵). Failing hearts based on several etiologies exhibit reduced capillary density. In this review article, we would like to focus on capillarization in cardiac tissues and discuss potential therapies for heart failure, targeting the enhancement of the capillary network in this organ.

1. Capillaries in the Heart with Various Conditions

In humans and other mammalian species, the heart is perfused through coronary circulation. Numerous bifurcations and anastomoses between capillaries create a dense vascular network, and this enables the delivery of nutrients and oxygen into cardiac tissues. Capillary rarefaction develops in hearts with aging or age-related cardiovascular-metabolic disorders, and these would be described in this chapter.

1-1. Aging

Aging is a nonmodifiable risk factor for cardiovascular diseases (CVDs). Aged heart develops functional and structural alterations^{16, 17}). Vasculature is also affected with age, and rodent hearts exhibited capillary rarefaction together with diminished oxygenation capacity^{7, 18, 19}). In C57BL/6J mice, capillary density was reduced in mice aged 18 months compared with those aged 2 months¹⁹). Young spontaneously hypertensive rats (SHRs) aged 2.5 months had a normal capillary density in their hypertrophied LV, whereas this declined in older animals aged 7 months²⁰). Rakusan *et al.* compared capillary density in patients with congenital aortic stenosis (AS) among infants, children (aged 9–14 years), or adults²). They concluded that capillary density diminished with age in humans under LV pressure overload²); however, underlying mechanisms are not fully understood. Senescent endothelial cells (ECs) exhibited reduced proliferative capacity together with diminished VEGF-A production²¹). At a certain level, reactive oxygen species (ROS) is considered to mediate homeostatic effects but, beyond the physiological threshold, initiate to promote unfavorable aspects of aging²²). ROS was reported to induce EC apoptosis and capillary rarefaction in the heart²³). Aging leads to an increase in decoy receptors for VEGF-A in circulation and suppresses VEGF-A-mediated angiogenic signaling; this was reported to enhance physiological aging in multiple organs, but cardiac tissue was not characterized in this paper¹). Recently, Kivela *et al.* reported transforming growth

factor- β (TGF- β)/ROS/Serpinh1 axis in ECs enhanced mesenchymal features in these cells, promoting cardiac fibrosis and capillary rarefaction¹⁹). These studies indicate that mechanisms associated with aging have causal roles for diminished capillary density in the heart, letting this tissue become prone to develop heart failure under stressed condition.

1-2. Obesity/Diabetes and Heart Failure

Obese individuals have a higher risk for developing HFpEF than lean subjects, and obese patients with HFpEF were reported to show worse exercise capacity than non-obese patients with HFpEF²⁴). Patients with HFpEF were shown to exhibit capillary rarefaction¹⁵), and metabolic stress is reported to reduce capillary density in humans and rodents²⁵⁻²⁷). In obese humans exhibiting BMI >30, coronary microvascular density was lower than in non-obese individuals²⁵). Wistar-Kyoto rats or C57BL/6J mice fed with high-fat diets developed capillary rarefaction in their respective hearts^{26, 27}). Mechanistically, Suda *et al.* showed metabolic stress reduced fibroblast growth factor 2 (FGF-2)/early growth response protein 1 (EGR-1)/VEGF-A signaling, and dipeptidyl peptidase 4 had causal roles for the suppression of this pathway²⁷). Dietary obesity also enhanced capillary rarefaction in C57BL/6J mice, and this was mediated through the activation of TGF- β /ROS/Serpinh1 signaling in ECs, sharing similar pathogenic mechanisms with their chronologically aged mice model¹⁹). Individuals with unhealthy obesity, generally characterized by enrichment in visceral adiposity and systemic insulin resistance, predisposes to type 2 diabetes. Approximately 45% of patients with HFpEF were reported to have type 2 diabetes²⁸), and overlapping pathologies are considered to exist between diabetes and HFpEF²⁹). At end-stage heart failure, cardiac capillary density became lower in patients with diabetes than in patients without diabetes³⁰). The same group also showed transgenic diabetic pig developed capillary rarefaction, and this was ameliorated with the introduction of adeno-associated virus encoding *Vegfa* (AAV-*Vegfa*). Compared with control diabetic pigs, ejection fraction did not improve in the AAV-*Vegfa* group³⁰). Interestingly, LV end-diastolic pressure and LV fibrosis were suppressed with the reintroduction of *Vegfa*, indicating the link between diminished capillary density and enhancement of fibrotic process³⁰). Streptozotocin-induced type 1 diabetic Wistar rats showed that the duration of hyperglycemia negatively correlated with capillary density in the heart³¹), suggesting glycemic overload as one of the mechanisms for promoting capillary

rarefaction under diabetic conditions.

1-3. Hypertension

In skeletal muscle, it was previously reported that an increase in mean arterial pressure is associated with a decrease in capillary density³². Few studies characterized cardiac capillary density in patients with hypertension. Biopsy samples obtained from patients with hypertensive heart disease (HHD), dilated cardiomyopathy (DCM), and renal failure with hemodialysis treatment (HD) showed capillary density as 1162 ± 189 , 1238 ± 261 , and 997 ± 183 (/mm²), respectively³³. In this paper, the percentage of hypertension was 100% in the HHD group, 8.9% in the DCM group, and 89.3% in the HD group. Studies analyzing capillary density in human hearts with AS showed that the capillary density of control adult hearts aged 15–30 years was 2249 ± 85 (/mm²), which became 2102 ± 103 (/mm²) in patients with congenital AS at similar age². The capillary density of patients with acquired AS aged 51–86 years was 1671 ± 66 (/mm²)². Through these papers published from different groups, it can be speculated that hypertension is associated with capillary rarefaction in the human heart. Further studies would show direct evidence of capillarization in patients with arterial hypertension. Rodent models with hypertension including Dahl salt-sensitive hypertensive rats^{34, 35} or high-salt diet³⁶ exhibited diminished capillary density in LV. High-salt diet, combined with Ang II administration in C57BL/6J mice, resulted in a decrease in capillary density³⁷. Capillary rarefaction also developed in SHR at 24 weeks of age, but this was comparable at 12 weeks of age³⁸. SHR were shown to exhibit reduced capillary density, but the number of smooth muscle α -actin positive arterioles increased in SHR than in control Wistar–Kyoto rats³⁹. Recent findings from Olianti *et al.* challenged previous reports by concluding that capillary rarefaction develops in an SHR model⁴⁰. They performed 3D imaging analyses in cardiac tissues and concluded that capillary density increased in SHR aged 4, 8, 18, and 24 weeks⁴⁰. Microvasculature is constituted with arterioles, capillaries, and venules, and characterization of these with specific cell markers would help us comprehensively understand capillarization in LV under aortic hypertension^{39, 41}. Activation of the renin–angiotensin–aldosterone system (RAAS) is one of the chief mechanisms that promote hypertension, and accumulated evidence indicates that suppression of RAAS enhances capillarization in the heart^{34, 42, 43}. LV pressure overload initially increased capillarization at compensated phase, and this was followed by capillary

rarefaction at decompensated phase heart failure⁹. Controlling blood pressure and suppressing afterload continues to be an important concept to suppress pathologies in the heart.

1-4. Aortic Valve Stenosis

Studies indicate the potential role of capillary rarefaction in the progression of pathologies in AS. LV outflow tract–capillary density of AS patients correlated with aortic valve area⁴⁴. Basal anteroseptal myocardium showed reduced capillarization in severe AS patients⁴⁵. Diminished capillary density associated with female gender, diabetes, obesity, heart failure symptoms, and low LV ejection fraction⁴⁴. As already described, patients with congenital AS exhibited reduced capillary density with chronological aging, and adults with acquired AS had lower capillary density compared with congenital AS children². The thoracic aortic constriction (TAC) model induces LV pressure overload and thereby mimics cardiac stress introduced with AS. In mice, TAC was shown to induce capillary rarefaction in LV^{8-10, 23, 44}. Accumulation of p53 in cardiac tissues⁹ or ECs¹⁰ suppressed angiogenic response in LV. ROS enhanced EC apoptosis during LV pressure overload and contributed to the progression of reduced systolic function²³. These results indicate enhancement of the capillary network in LV would become a therapy for heart failure associated with LV pressure overload.

1-5. Chronic Kidney Disease

Chronic kidney disease (CKD) is an independent risk for CVD⁴⁶. The hearts of experimental uremic Sprague–Dawley rats (5/6 nephrectomy (5/6 NX)) exhibited cardiac hypertrophy, interstitial fibrosis, and reduced capillary density⁴⁷. Nephrectomy-induced capillary rarefaction in the heart was completely inhibited by the administration of moxonidine, a central sympatholytic agent, but not by the calcium antagonist nifedipine⁴⁸. Another report showed transcript *Vegfa* reduced in the heart with 5/6 NX, and this was ameliorated with selective renal sympathetic denervation⁴⁹. These findings indicate that renal dysfunction accelerates capillary rarefaction through sympathetic nervous activation. Another study showed that treatment with an endothelin receptor antagonist normalized microvascular density⁵⁰. Amann *et al.* tested two angiotensin-converting enzyme (ACE) inhibitors and concluded that ramipril⁵¹, but not trandolapril⁵⁰, inhibited a decrease in capillary density in a uremic rat model. Additionally, the combination of erythropoietin and ACE inhibitor (enalapril) increased microvascular density in 5/6 NX rats⁵². Mechanistically, a decrease

in oxidative stress and apoptotic signaling contributed to maintaining LV capillary network under uremic condition⁵²). Di Marco *et al.* demonstrated the level of calcineurin increased in the heart with nephrectomy, and an inhibitor for this molecule normalized microvascular density in 5/6 NX rats⁵³). They showed that calcineurin inhibitor increased angiogenic or stem cell-related molecules including vascular endothelial growth factor 2 (VEGFR2), survivin, cKit-1, and stem cell factor⁵³). In CKD patients, the estimated glomerular filtration rate showed a negative correlation with soluble Flt-1 (sFlt-1) in plasma, and the level of sFlt-1 correlated with heart failure symptoms and early mortality⁵⁴). Rats administered with sFlt-1 had reduced LV capillary density, lower systolic and diastolic function, increase in TUNEL positive cells, and fibrosis in the heart compared with controls⁵⁴). In this paper, inhibition of sFlt-1 increased capillary density, myocardial blood volume, and ameliorated diastolic dysfunction in the 5/6 NX model. Several complex mechanisms have roles in reducing capillaries in the heart under renal dysfunction, and further studies are needed to understand pathologies of cardiorenal syndrome.

1-6. Amyloidosis

Cardiac amyloidosis is characterized by the extracellular deposition of insoluble fibrils composed of misfolded proteins called amyloid. The layers of the walls of intramural vessels are progressively infiltrated by the fibrils, eventually leading to microvascular obstruction⁵⁵). The amyloid deposits surrounding myocytes induce oxidative stress and affect the contractile function of these cells⁵⁶). Reduced stroke volume and compliance of the LV wall contribute to decreasing diastolic flow in the intramural vessels. In symptomatic patients with AL or transthyretin amyloidosis, myocyte blood flow became lower than in hypertensive patients exhibiting LV hypertrophy⁵⁷). Recently, Kim *et al.* showed direct evidence of reduced capillary density in AL amyloidosis patients⁵⁸). They analyzed cardiac biopsy samples and showed capillary density had a negative correlation with NT-proBNP level, amyloid load, LV diastolic, and systolic function⁵⁸). In this paper, the patients exhibiting capillary density of $>220/\text{mm}^2$ were shown to have a better prognosis than those exhibiting $<220/\text{mm}^2$ ⁵⁸). Together with therapy targeting removal of amyloid deposition, whether maintaining capillarization becomes a therapy for amyloidosis remains an open question to be explored.

1-7. Hypertrophic Cardiomyopathy

Myectomy samples showed that capillary density

decreased in patients with hypertrophic cardiomyopathy (HCM) compared with control individuals, and interestingly, this showed a negative correlation with left ventricular outflow tract (LVOT) pressure gradient⁵⁹). HCM patients with moderate hypertrophy and LVOT obstruction also showed reduced capillaries in LV compared with control subjects⁶⁰). DBA/2J mouse carrying variants in the two most susceptible genes—Mybpc3 and Myh7—demonstrates the features of HCM, and vessel density was shown to diminish in this mouse model compared with C57BL/6J wild type mice⁶¹). The fundamental question that remains to be answered is whether enhancing the capillary network reverses or slows down the pathologies in HCM.

2. Role of Senescence-Related Molecules in the Cardiac Capillary Network

Aging or age-related disorders are associated with capillary rarefaction in the heart. Studies indicate age-related mechanisms have causal roles for reducing capillarization in the heart, and these are described in this chapter.

2-1. p53

Chronological aging is associated with a higher prevalence of age-related diseases including heart failure, diabetes, and atherosclerotic disorders. Mechanisms of aging and age-related disorders are complex; however, studies indicate the crucial role of cellular senescence in the progression of these disorders^{9, 10, 22, 62-68}). The p53 protein, which is often described as the “guardian of the genome,” is a transcriptional factor involved in genomic stability. This molecule mediates the protective effect through coordinating DNA repair, cell-cycle regulation, and apoptosis. p53 also has a central role in inducing cellular senescence, and this promotes pathogenesis in age-related disorders. Previous reports showed p53 increased in the aged⁶⁹) or failing heart⁹). In mice, LV pressure overload induced capillary rarefaction together with enhanced p53 level in cardiac tissue^{9, 21}). Mechanistically, p53 enhanced ubiquitination and proteasomal degradation of hypoxia-inducible factor 1 α (HIF-1 α), thereby suppressing VEGF-A mediated angiogenesis^{9, 70}). Another study showed EC-p53 enhanced capillary rarefaction, tissue fibrosis, and inflammation in the heart during LV pressure overload^{10, 66}). Impairment of cardiac angiogenesis by p53 was also observed in other models such as angiotensin II (Ang II)-infused mice⁷¹). In diabetic rodents, pharmacological inhibition of p53 led to stabilization of HIF-1 α , enhanced capillarization, and

ameliorated cardiac dysfunction⁷²). Inhibition of p53 would contribute to increasing the capillary network in the heart and suppress pathogenesis in the failing heart.

2-2. p21

Protein p21, coded by cyclin-dependent kinase inhibitor 1a (*Cdkn1a*), is a cell-cycle regulator and locates downstream of p53⁷³. Activation of p21 triggers cellular senescence and apoptosis⁷⁴. C57BL/6 mice aged 24 months had a higher level of p21 in cardiac tissues compared with 4-month-old mice⁶⁹. Controversies exist regarding the role of p21 in capillarization. Systemic depletion of p21 diminished capillary density together with reduced cardiac systolic function⁷⁵. Another report showed radiation increased breast cancer type 1 susceptibility protein homolog (BRCA1) and p21 level in heart⁷⁶. EC-specific *Brca1* depletion resulted in reduced p21 and capillary density, indicating the protective effect of EC-BRCA1/p21 mediated DNA repair response in heart⁷⁶. In the dental pulp, the knockdown of p53 or p21 has led to an increase in capillary density, and studies showed p53/p21 signaling reduced Bmi-1-mediated vasculogenic differentiation of dental pulp stem cells⁷⁷. It was also reported that p21 negatively regulated the proliferation of old endothelial progenitor cells (EPCs) *in vitro* and *in vivo* settings⁷⁸. Another study showed that starvation-induced NADPH oxidase (Nox2) upregulation and ROS production promoted EC cycle arrest and apoptosis via the p21 pathway⁷⁹. The role of p21 in capillarization may be context dependent and different among organs, cells, and disorders.

2-3. p16

p16-INK4a is a cell-cycle regulator encoded by the *Cdkn2a* gene. p16 is widely used as a marker for senescent cells and has been reported to increase with age in cardiomyocytes⁸⁰, ECs⁸¹, and cardiac progenitor cells⁸². The level of p16 in the heart increased in aged mice⁶⁹. In humans, the protein levels of p53, p21, and p16 increased in ECs from older individuals (~60 years old), and those were shown to reduce in older exercising adults (~57 years old)⁸¹. Protein canopy2 (CNPY2) stimulates cell proliferation, and a previous report showed CNPY2-mediated p16 inhibition enhanced capillarization and tissue repair in the heart after myocardial infarction⁸³. Another study showed p16 deletion enhanced capillarization and promoted kidney regeneration after kidney ischemia–reperfusion injury⁸⁴. Further studies are needed to show the role of p16 in cardiac capillarization.

2-4. Sirtuins

Sirtuins (Sirt1-7) are a family of nicotinamide adenine dinucleotide (NAD) dependent histone deacetylases. NAD reduces with age and NAD administration is considered as a promising target to reverse aging phenotype⁸⁵. It is well known that sirtuins get activated by NAD, and sirtuins are known to have roles in aging and capillarization⁸⁶. SIRT1 inhibition induced premature cell senescence in human umbilical vein ECs (HUVECs), whereas overexpression of this molecule prevented them from premature senescence-like phenotype⁸⁷. The level of NAD was reported to decline with age in skeletal muscle ECs, and this had a causal role in reducing SIRT1 and capillary rarefaction in this organ⁸⁸. EC-specific Sirt1 knockout mice exhibited capillary rarefaction in the heart and developed diastolic dysfunction⁸⁹. EC-specific Sirt3 depletion also resulted in diminished capillarization in the heart together with diastolic dysfunction⁹⁰. Administration of Ang II-induced fibrosis diminished vascular density in the heart, and this was further enhanced with depletion of Sirt3⁹¹. The role of other sirtuins in cardiac capillarization continues to be an interesting topic to be explored.

2-5. AKT

The serine/threonine protein kinase AKT-mediated signaling has critical roles in the regulation of cardiac hypertrophy, contractile function, and coronary angiogenesis^{8, 92}. Mice aged 24–26 months showed high AKT levels in the heart compared with mice aged 3–4 months⁹³. In cardiac tissues, the role of AKT is bidirectional, and a summary of previous papers indicates that the level of AKT signaling should be kept under a certain physiological level for maintaining homeostasis of the heart⁹⁴. Short-term AKT activation induced physiological hypertrophy with maintained vascular density, suggesting that coronary angiogenesis is enhanced to keep pace with the growth of the myocardium⁹⁵. By contrast, prolonged AKT activation introduced pathological cardiac hypertrophy, characterized by enhanced fibrosis and capillary rarefaction⁹⁵. LV pressure overload upregulated AKT signaling, and heterozygous depletion of AKT ameliorated cardiac dysfunction by inhibiting excessive hypertrophic response in the heart together with an increase in vascular density⁸. In ECs, AKT promotes cell proliferation, survival, permeability, release of nitric oxide (NO), and cell migration⁹⁶ and is essential for VEGF-mediated angiogenesis⁹⁷. Gain of AKT signaling in ECs contributed to an increase in endothelial nitric oxide synthase (eNOS)/NO

pathway, and this contributed to the suppression of EC apoptosis together with enhanced capillarization in LV²³). By contrast, EC-AKT is also reported to enhance senescent phenotype. Senescent primary cultured human ECs exhibited a high level of phospho-AKT, and overexpression of this molecule suppressed population doubling and increased p53/p21 levels in these cells⁹⁸). These studies indicate AKT activation must be cell-targeted and within a certain physiological level.

2-6. Inflammation

Levels of circulating pro-inflammatory molecules, such as interleukin 6 (IL-6), tumor necrosis factor- α (TNF- α), and C-reactive protein increase with aging⁹⁹⁻¹⁰³) or in patients with HFpEF¹⁰⁴). Endothelial activating markers including ICAM-1 and E-selectin became higher in myocardial samples of a ZSF-1 HFpEF rat model or patients with HFpEF¹⁰⁵). Murine LV pressure overload model showed that ICAM-1 has a critical role in the infiltration of inflammatory macrophages into cardiac tissues⁶⁶). On a cellular level, TNF- α and IL-6 activate NADPH oxidase (NOX) in the coronary arteriolar wall, and this enhances the production of superoxide anion, decreases NO availability, and inhibits vasodilation¹⁰⁶).

Studies analyzing type 1 diabetic mice models concluded that hyperglycemia increased TNF- α in cardiac tissues, and this promoted capillary rarefaction by enhancing cell death in ECs¹⁰⁷). IL-10 was reported to suppress inflammatory response and promote capillary density through the activation of STAT3¹⁰⁸) or via the upregulation of heme clearance pathway¹⁰⁹) in a murine myocardial infarction model. Controversies remain for the role of inflammation in cardiac capillarization. In an Ang II high salt-induced hypertension model, IL-6 knockout mice had better cardiac ejection fraction, reduced macrophage infiltration, and fibrosis in LV, but capillary density was comparable between the genotypes³⁷). IL-6 knockout mice had reduced capillary density in the heart, together with fibrosis and LV dilatation¹¹⁰). In patients with myocardial infarction, circulating monocytes had a high level of semaphorin3A at day 30 after the onset of this disease¹¹¹). Heterozygous depletion of semaphorin3A suppressed cardiac inflammation and improved cardiac function; however, no change in capillary density was observed in the knockout mice¹¹¹). The role of inflammation in the capillary network seems complex and mediates a bidirectional or neutral effect for capillarization in the heart.

2-7. ROS

ROS has central roles in the progression of

pathologies in aging or age-related disorders. ROS including superoxide ($O_2^{\cdot-}$), hydrogen peroxide (H_2O_2), peroxyxynitrite ($OONO^{\cdot-}$), and the hydroxyl radical (HO^{\cdot}) are produced through physiological biological activities. At certain levels, they have roles as signaling mediators, however, become toxic in excess and thereby described as oxidative stress. Under physiological conditions, oxidative balance is tightly regulated by prooxidant and antioxidant systems. ROS has been shown to regulate multiple cellular functions in vessels including angiogenesis, EC apoptosis, and vascular tone^{112, 113}). ROS was reported to activate VEGFR2 in ECs and promote angiogenesis¹¹⁴). Low-density lipoprotein (LDL) induced endothelial dysfunction through the activation of lectin-like-oxidized LDL receptor-1 (LOX-1) and excessive ROS, and these led to a decrease in eNOS uncoupling¹¹⁵). ROS derives from several sources including mitochondrial respiratory chain enzymes, xanthine oxidases, lipoxygenases, and NOX proteins. With aging, complex I activity in mitochondria decreased in rats, and this was associated with the capacity of this organelle to produce H_2O_2 ¹¹⁶). NOX2 and NOX4 are predominant forms of NOXs expressed in the heart¹¹⁷). NOX2-derived ROS mediates the antiangiogenic effect, whereas NOX4-derived ROS was reported to have an angiogenic effect. In the brain, Nox2 activation was shown to enhance age-related cerebral capillary rarefaction¹¹⁸). In the hind limb ischemia model, Nox2 depletion resulted in a decrease in ROS level together with an increase in VEGF-A expression¹¹⁹). Studies using endothelial-specific Nox2 overexpression showed NOX2 activation promoted superoxide-driven cardiovascular dysfunction, macrophage recruitment, and adverse remodeling^{120, 121}). By contrast, NOX4 is constitutively active at a low level and induces protective effects in the heart under chronic stress¹²²). Systemic Nox4 knockout mice showed impaired angiogenesis in a hind limb ischemia model¹²³). The Nox4 knockout model exhibited cardiac capillary rarefaction and LV systolic dysfunction during LV pressure overload¹²²). Cardiomyocyte-specific Nox4 overexpression enhanced capillarization and improved LV systolic function¹²²). Mechanistically, Nox4 was shown to increase HIF-1 α /VEGF-A under hypoxic condition¹²²).

3. Pharmaceutical Approach for Enhancing Cardiac Capillary Network

Several types of drugs are reported to suppress pathologies in heart failure, contributing to the inhibition of hospitalization for heart failure, amelioration of symptoms, and improvement in LV

ejection fraction^{124, 125}). In this chapter, we would mainly focus on cardioprotective drugs and show their effects against the capillary network in LV.

3-1. ACEi/ARB/ARNI

Activation of RAAS mediated mainly through Ang II/Ang II receptor type 1 (AT1R) signaling induces myocardial hypertrophy and fibrosis and contributes to the progression of heart failure. RAAS inhibitors reduce HFrEF-related morbidity and mortality¹²⁶⁻¹²⁸). Accumulation of evidence indicates suppression of RAAS leads to enhancement of capillarization in the heart. In type 2 diabetic Otsuka Long-Evans Tokushima Fatty (OLETF) rat model, an AT1R blocker candesartan ameliorated LV capillary rarefaction and improved diastolic function at 36 weeks of age¹²⁹). Another study by de Boer *et al.* showed that microvascular density after myocardial infarction (MI) was decreased by cardiomyocyte-specific AT1R overexpression, and one of the AT1 receptor blockers losartan increased LV capillarization without an increase of VEGF¹³⁰). Another AT1 receptor blocker irbesartan suppressed oxidative stress and EC apoptosis, ameliorated cardiac hypertrophy, and increased capillary density in Dahl salt-sensitive (DS) rats¹³¹). An ACE inhibitor ramipril enhanced LV capillarization in an SHR⁴³). Another ACE inhibitor perindopril increased LV capillary density in DS rats³⁴). Perindopril was also reported to enhance LV capillarization in obese Zucker rats⁴²). The proangiogenic effect of ACE inhibition is considered partly mediated via the activation of bradykinin receptor/eNOS signaling in ECs^{43, 132, 133}). Interestingly, several studies indicate the proangiogenic effect of RAAS.

Subhypertensive dose Ang II/AT1R signaling increased VEGF-A/eNOS pathway and mediated angiogenic response in ischemic hindlimb model¹³⁴). Ang II/VEGF-A axis was also shown to induce the proliferation of EPCs¹³⁵). Enalapril or candesartan both inhibited VEGF-A-induced myocardial angiogenesis and capillarization in the normal heart of C57BL/6 male mice, but functional studies were not conducted to test LV function¹³⁶). LV capillary density was increased at 20 weeks of age in the OLETF diabetic rat model, and candesartan reduced vascular expression of VEGF-A, HIF-1 α , and capillary density in these rats¹³⁷). Exercise increased capillarization in the heart, but Ang II infusion showed a trend to reduce this in LV of Sprague–Dawley rats; however, this did not show statistical significance¹³⁸). The roles of RAAS in angiogenic response seems context dependent and are different among models, organs, and time frame of analyses.

Neprilysin (neutral endopeptidase, NEP) cleaves and inactivates peptide hormones such as glucagon, bradykinin, angiotensin, endothelin-1, and natriuretic peptides¹³⁹). PARADIGM-HF trial-tested angiotensin receptor–neprilysin inhibitor (ARNI) against enalapril in HFrEF patients, and the ARNI group showed a significant decrease in the risk of death and heart failure hospitalization¹⁴⁰). In an experimental MI rat model, ARNI increased transcript *Vegfa* in LV, but this did not associate with an increase in capillary density¹⁴¹). Suematsu *et al.* showed ARNI ameliorated cardiac dysfunction together with an increase in transcript *Vegfa* in hearts in ApoE KO mice, but capillary density was not characterized in this paper¹⁴²). Further studies are needed to show the role of NEP inhibitors in cardiac capillarization.

3-2. Beta-Blockers and Ivabradine

β -blockers continue to be the first-line therapy for heart failure^{124, 125}). Studies indicate the proangiogenic effect of this type of drug in cardiac tissues. A negative correlation between heart rate (HR) and capillary density indicates bradycardia promotes angiogenic response¹⁴³⁻¹⁴⁵). Bradycardia stimulated VEGF production and facilitated angiogenesis in heart¹⁴⁶). Reduced HR prolongs diastolic filling, and this triggers an increase in the stretch-associated release of VEGF¹⁴⁷). Cardiac capillary density was reduced in the renal failure model generated in Sprague–Dawley rats, and metoprolol was shown to ameliorate this¹⁴⁸). Either ivabradine or metoprolol enhanced LV capillary density in MI rats¹⁴⁵). Alinidine, another bradycardiac drug, increased VEGF and capillary density in hearts subjected to myocardial infarction¹⁴⁴). Ivabradine inhibits pacemaker current and reduces HR. Ivabradine was reported to reduce hospitalization of heart failure¹⁴⁹). In the MI rats, ivabradine administration enhanced capillary density in LV and improved cardiac systolic function^{145, 150}).

3-3. SGLT2 Inhibitor

Sodium glucose cotransporter 2 inhibitors (SGLT2i) were initially generated as a therapy for type 2 diabetes, and this class of drugs was shown to significantly reduce the risk of hospitalization for heart failure^{11, 12, 151, 152}). SGLT2i also reduced the risk of cardiovascular death or hospitalizations for heart failure in patients without diabetes^{153, 154}). Biological effects of SGLT2i are considered multifactorial, and these include a decrease in ROS and inflammation, increase in ketone production, and inhibition of sodium–hydrogen exchange¹⁵⁵). Several reports focused on the role of SGLT2i in cardiac capillaries. SGLT2 inhibition with empagliflozin (EMPA)

improved coronary microvascular function in prediabetic ob/ob mice¹⁵⁶. Another study revealed that EMPA inhibited the mitochondrial fission via AMPK signaling and rescued cardiac microvascular EC injury via ROS inhibition¹⁵⁷. Recently, Nakao *et al.* demonstrated that EMPA maintained LV capillarization and improved cardiac function in a murine LV pressure overload model²³. EMPA administration reduced ROS-mediated apoptosis in ECs, and together with the activation of AKT/eNOS/NO pathway, EMPA enhanced capillarization in LV under pressure overload²³. Whether SGLT2i also enhances capillary density in the human heart continues to be an interesting topic to be explored.

3-4. MRA

The Randomized Aldactone Evaluation Study demonstrated inhibition of mineralocorticoid receptor (MR) led to a 30% decrease in mortality rates in patients with heart failure after myocardial infarction¹⁵⁸. Aldosterone is the principal ligand for MR and is known to associate with increased risk for cardiovascular events¹⁵⁹. MR activation induces cardiac hypertrophy, inflammation, and fibrosis, and these were blocked via MR inhibition¹⁶⁰⁻¹⁶³. In a murine MI model, cardiomyocyte-specific MR depletion increased LV capillary density, reduced interstitial fibrosis and inflammation, and improved systolic function¹⁶¹. Deoxycorticosterone acetate administration induced hypertension and LV capillary rarefaction in WT mice, and this was ameliorated in EC-specific MR KO mice¹⁶². Another report showed EC-specific MR depletion improved systolic function, but capillary density was comparable between the genotype¹⁶⁴. Smooth muscle cell-specific MR knockout mice showed increased capillary density, diminished fibrosis in the heart, and improved cardiac function during LV pressure overload¹⁶³. Obesity and aging are associated with a dysregulation in MR signaling^{165, 166}. Together with evidence from preclinical studies, MR signaling continues to be an important therapeutic target for heart failure.

3-5. DPP4 Inhibitor

Dipeptidyl peptidase-4 (DPP4) inhibitor is a class of oral antidiabetic agents. The extracellular catalytic domain of DPP4 is responsible for the enzymatic degradation of several peptides, including incretins GLP-1 and GIP, neuropeptides, chemokines, and endogenous growth factors¹⁶⁷. Studies indicated DPP4 inhibitors cannot suppress pathologies of heart failure in humans¹⁶⁸⁻¹⁷⁰; however, preclinical studies suggest this type of drug enhances cardiac capillary density. In a dietary obese mice model, Suda *et al.*

showed linagliptin ameliorated capillary rarefaction in the heart through the activation of the FGF-2/EGR-1 pathway²⁷. The membrane-bound form of DPP4 was activated in diabetic rats, and this led to a decrease of myocardial stromal cell-derived factor-1 α (SDF-1 α) concentrations and angiogenesis in hearts¹⁷¹. In these rats, DPP4 suppression reversed cardiac fibrosis, diastolic dysfunction, and increased capillary density together with a restoration of SDF-1 α ¹⁷¹. Another study also demonstrated that DPP4 inhibitor (PKF275-055) suppressed the biological inactivation of SDF-1, increased mobilization of EPCs, and regenerated cardiac capillaries in ob/ob mice¹⁷². Studies analyzing rodents showed DPP4 inhibitors as promising therapies to combat heart failure; however, clinical trials could not show the cardioprotective effect of these drugs¹⁶⁸⁻¹⁷⁰. This may be due to differences among humans and rodents, relative differences in the dose of DPP4 inhibitors showing beneficial biological effects among species, or other unknown factors.

3-6. Senolytics

Senescent cells increase in aged tissues, and this is considered to associate with the progression of age-related cardiovascular-metabolic diseases^{22, 62, 63, 173, 174}. Senescence-associated molecules such as p53, p21, and p16 increase in heart with aging, LV pressure overload, myocardial ischemia, and diabetes^{9, 21, 66, 69, 72}. Recent studies testing genetic, as well as pharmacological models, showed elimination of senescent cells, described as “Senolysis,” reverses aging phenotype^{80, 175-177}. This approach provides an attractive therapeutic option for age-related CVDs. Senolytics are agents that can selectively target prosurvival proteins in senescent cells and induce cell death. Dasatinib, Quercetin, and Navitoclax are three major senolytics widely studied *in vivo* and *in vitro*¹⁷⁸. Besides the genetic senolytic model, these reagents have been shown to improve vascular function¹⁷⁹, reduce detrimental features of cardiac aging⁸⁰ and restore the regenerative capacity of hearts in murine myocardial ischemia, Ang II-infused cardiac hypertrophy, diabetic, or aged model^{80, 82, 180, 181}. Dookun *et al.* showed that clearance of senescent cells with Navitoclax improved cardiac recovery including enhanced angiogenesis after cardiac ischemia–reperfusion injury¹⁷⁷. Yu *et al.* showed Quercetin suppressed cardiac vascular rarefaction and improved systolic dysfunction in high-fat diet-fed mice¹⁸². The therapeutic potential of senolytics is now tested in humans. Justice *et al.* showed administration of Dasatinib+Quercetin in patients with idiopathic pulmonary fibrosis can improve physical function¹⁸³.

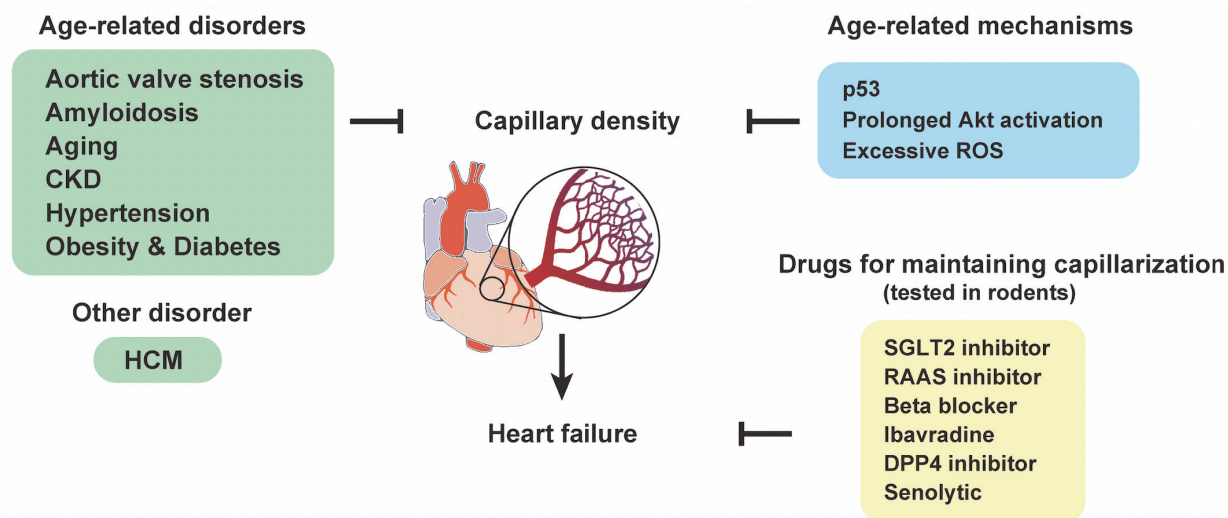


Fig. 1. Disorders and mechanisms for reduced capillary density in the heart

Age-related disorders including aortic valve stenosis, amyloidosis, aging, chronic kidney disease (CKD), hypertension, obesity, and diabetes diminish capillary density in the heart. Hypertrophic cardiomyopathy (HCM) is also associated with capillary rarefaction. Age-related mechanisms such as p53, prolonged AKT activation, and excessive ROS reduce capillary density. Drugs tested in rodents to enhance capillarization are also demonstrated. Class and name of drugs, tested species and models, analyzed organs, and effects are demonstrated.

Hickson *et al.* tested this combination of drugs in patients with diabetic kidney disease and found senescent cells reduced in subcutaneous adipose tissue¹⁸⁴. Recently, Suda *et al.* showed senolytic vaccination therapy reversed aging phenotype in obesity or atherosclerotic model¹⁸⁵. The senolytic approach would open new avenues for therapeutic options for age-related cardiovascular disorders.

4. Discussion

In this review article, we outlined capillarization in the heart under several pathogenic conditions. Capillary rarefaction develops with aging or age-related disorders, and studies indicate that this accelerates the functional decrease in cardiac tissues letting this organ prone to develop heart failure. Age-related mechanisms including p53-mediated cell senescence, sirtuins, AKT signaling, RAAS, chronic inflammation, and excessive ROS had critical roles for maintenance or suppression of capillarization in the heart. Drugs used as first-line therapies for patients with heart failure were reported to contribute to the maintenance of capillarization in the heart. Accumulating evidence indicates β -blockers, RAAS inhibitors, and SGLT2 inhibitors improve the prognosis of heart failure, and studies with rodents show these drugs also contribute to enhancing capillarization in the heart. Recently, approaches with senolysis are highlighted in aging and cardiovascular research. Studies showed specific depletion of

senescent cells ameliorated cardiac dysfunction in heart failure models, and interestingly, capillary density was also shown to improve with senolysis. Senolytics are now tested in humans, and results have shown a decrease in senescent cells and improvement in clinical symptoms. Many of the senolytics are classified as cancer drugs, and potential side effects are issues to be considered. Exploration of less toxic senolytics continues to be one of the interesting and important research topics. With chronological aging, capillary rarefaction develops in systemic organs, and this is considered to accelerate a functional decrease in these tissues. Exploration of reagents contributing to the enhancement of capillarization would help us find new drugs for age-related disorders including heart failure (**Fig. 1 and Table 1**).

COI

Yoshida Y, Shimizu I; None, Minamino T; Scholarship grants from Bourbon corporation.

References

- 1) Grunewald M, Kumar S, Sharife H, Volinsky E, Gileles-Hillel A, Licht T, Permyakova A, Hinden L, Azar S, Friedmann Y, Kupetz P, Tzuberi R, Anisimov A, Alitalo K, Horwitz M, Leebhoff S, Khoma OZ, Hlushchuk R, Djonov V, Abramovitch R, Tam J and Keshet E: Counteracting age-related VEGF signaling insufficiency promotes healthy aging and extends life span. *Science*, 2021; 373:

Table 1. Drugs, tested models, and their effects in cardiac capillarization

Class of drugs	Name of drugs	Tested species	Models	Organ	Effect	Ref
ACEi/ARB/ ARNI	Losartan	Rat	Myocardial infarction	Heart	Increased LV capillary density	de Boer <i>et al.</i> , 2003
	Irbesartan	Rat	Hypertension	Heart	Increased LV capillary density	Nako <i>et al.</i> , 2012
	Candesartan	Rat	Type 2 diabetes	Heart	Increased LV capillary density at 36 weeks of age	Hayashi <i>et al.</i> , 2003
	Candesartan	Rat	Type 2 diabetes	Heart	Reduced capillary density at 20 weeks of age	Jesmin <i>et al.</i> , 2002
	Candesartan or Enalapril	Mouse	Wild type mice injected with Vegfa plasmid	Heart	Reduced capillary density	Siddiqui <i>et al.</i> , 2005
	Ramipril	Rat	Hypertension	Heart	Increased LV capillary density	Zhu <i>et al.</i> , 1997
	Perindopril	Rat	Hypertension	Heart	Increased LV capillary density	Yazawa <i>et al.</i> , 2011
	Perindopril	Rat	Obesity	Heart	Increased LV capillary density	Toblli <i>et al.</i> , 2004
	Sacubitril/valsartan	Rat	Myocardial infarction	Heart	Increased transcript Vegfa, but no change in capillary density	Pfau <i>et al.</i> , 2019
	Sacubitril/valsartan	Mouse	Atherosclerosis	Heart	Increased transcript Vegfa, but capillary density was not characterized	Suematsu <i>et al.</i> , 2021
Beta blocker	Metoprolol	Rat	Myocardial infarction	Heart	Increased LV capillary density	Ulu <i>et al.</i> , 2009
	Metoprolol	Rat	Renal failure	Heart	Increased LV capillary density	Amann <i>et al.</i> , 2006)
Bradycardiac drug	Alinidine	Rat	Myocardial infarction	Heart	Increased LV capillary density	Lei <i>et al.</i> , 2004
	Ivabradine	Rat	Myocardial infarction	Heart	Increased LV capillary density	Ulu <i>et al.</i> , 2009
SGLT2 inhibitor	Empagliflozin	Mouse	LV pressure overload	Heart	Increased LV capillary density	Nakao <i>et al.</i> , 2021
DPP4 inhibitor	Linagliptin	Mouse	Obesity	Heart	Increased LV capillary density	Suda <i>et al.</i> , 2017
Senolytics	Navitoclax	Mouse	Myocardial infarction	Heart	Increased LV capillary density	Dookun <i>et al.</i> , 2020b
	Quercetin	Mouse	Obesity	Heart	Increased LV capillary density	Yu <i>et al.</i> , 2021

Class and name of drugs, tested species and models, analyzed organs, and effects are demonstrated.

- 2) Rakusan K, Flanagan MF, Geva T, Southern J and Van Praagh R: Morphometry of human coronary capillaries during normal growth and the effect of age in left ventricular pressure-overload hypertrophy. *Circulation*, 1992; 86: 38-46
- 3) Urbieto-Caceres VH, Syed FA, Lin J, Zhu XY, Jordan KL, Bell CC, Bentley MD, Lerman A, Khosla S and Lerman LO: Age-dependent renal cortical microvascular loss in female mice. *Am J Physiol Endocrinol Metab*, 2012; 302: E979-986
- 4) Hill LK, Hoang DM, Chiriboga LA, Wisniewski T, Sadowski MJ and Wadghiri YZ: Detection of

Cerebrovascular Loss in the Normal Aging C57BL/6 Mouse Brain Using in vivo Contrast-Enhanced Magnetic Resonance Angiography. *Front Aging Neurosci*, 2020; 12: 585218

- 5) Chen J, Lippo L, Labella R, Tan SL, Marsden BD, Dustin ML, Ramasamy SK and Kusumbe AP: Decreased blood vessel density and endothelial cell subset dynamics during ageing of the endocrine system. *EMBO J*, 2021; 40: e105242
- 6) Li H, Hastings MH, Rhee J, Trager LE, Roh JD and Rosenzweig A: Targeting Age-Related Pathways in Heart Failure. *Circ Res*, 2020; 126: 533-551

- 7) Iemitsu M, Maeda S, Jesmin S, Otsuki T and Miyauchi T: Exercise training improves aging-induced downregulation of VEGF angiogenic signaling cascade in hearts. *Am J Physiol Heart Circ Physiol*, 2006; 291: H1290-1298
- 8) Shimizu I, Minamino T, Toko H, Okada S, Ikeda H, Yasuda N, Tateno K, Moriya J, Yokoyama M, Nojima A, Koh GY, Akazawa H, Shiojima I, Kahn CR, Abel ED and Komuro I: Excessive cardiac insulin signaling exacerbates systolic dysfunction induced by pressure overload in rodents. *Journal of Clinical Investigation*, 2010; 120: 1506-1514
- 9) Sano M, Minamino T, Toko H, Miyauchi H, Orimo M, Qin Y, Akazawa H, Tateno K, Kayama Y, Harada M, Shimizu I, Asahara T, Hamada H, Tomita S, Molkenntin JD, Zou Y and Komuro I: p53-induced inhibition of Hif-1 causes cardiac dysfunction during pressure overload. *Nature*, 2007; 446: 444-448
- 10) Gogiraju R, Xu X, Bochenek ML, Steinbrecher JH, Lehnart SE, Wenzel P, Kessel M, Zeisberg EM, Dobbstein M and Schafer K: Endothelial p53 deletion improves angiogenesis and prevents cardiac fibrosis and heart failure induced by pressure overload in mice. *J Am Heart Assoc*, 2015; 4:
- 11) Wiviott SD, Raz I, Bonaca MP, Mosenzon O, Kato ET, Cahn A, Silverman MG, Zelniker TA, Kuder JF, Murphy SA, Bhatt DL, Leiter LA, McGuire DK, Wilding JPH, Ruff CT, Gause-Nilsson IAM, Fredriksson M, Johansson PA, Langkilde AM, Sabatine MS and Investigators D-T: Dapagliflozin and Cardiovascular Outcomes in Type 2 Diabetes. *N Engl J Med*, 2019; 380: 347-357
- 12) Zinman B, Wanner C, Lachin JM, Fitchett D, Bluhmki E, Hantel S, Mattheus M, Devins T, Johansen OE, Woerle HJ, Broedl UC, Inzucchi SE and Investigators E-RO: Empagliflozin, Cardiovascular Outcomes, and Mortality in Type 2 Diabetes. *N Engl J Med*, 2015; 373: 2117-2128
- 13) Anker SD, Butler J, Filippatos G, Ferreira JP, Bocchi E, Bohm M, Brunner-La Rocca HP, Choi DJ, Chopra V, Chuquiure-Valenzuela E, Giannetti N, Gomez-Mesa JE, Janssens S, Januzzi JL, Gonzalez-Juanatey JR, Merkely B, Nicholls SJ, Perrone SV, Pina IL, Ponikowski P, Senni M, Sim D, Spinar J, Squire I, Taddei S, Tsutsui H, Verma S, Vinereanu D, Zhang J, Carson P, Lam CSP, Marx N, Zeller C, Sattar N, Jamal W, Schnaidt S, Schnee JM, Brueckmann M, Pocock SJ, Zannad F, Packer M and Investigators EM-PT: Empagliflozin in Heart Failure with a Preserved Ejection Fraction. *N Engl J Med*, 2021; 385: 1451-1461
- 14) McDonagh TA, Metra M, Adamo M, Gardner RS, Baumbach A, Bohm M, Burri H, Butler J, Celutkiene J, Chioncel O, Cleland JGF, Coats AJS, Crespo-Leiro MG, Farmakis D, Gilard M, Heymans S, Hoes AW, Jaarsma T, Jankowska EA, Lainscak M, Lam CSP, Lyon AR, McMurray JJV, Mebazaa A, Mindham R, Muneretto C, Francesco Piepoli M, Price S, Rosano GMC, Ruschitzka F, Kathrine Skibelund A and Group ESCSD: 2021 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure. *Eur Heart J*, 2021; 42: 3599-3726
- 15) Mohammed SF, Hussain S, Mirzoyev SA, Edwards WD, Maleszewski JJ and Redfield MM: Coronary microvascular rarefaction and myocardial fibrosis in heart failure with preserved ejection fraction. *Circulation*, 2015; 131: 550-559
- 16) Loffredo FS, Steinhauser ML, Jay SM, Gannon J, Pancoast JR, Yalamanchi P, Sinha M, Dall'Osso C, Khong D, Shadrach JL, Miller CM, Singer BS, Stewart A, Psychogios N, Gerszten RE, Hartigan AJ, Kim MJ, Serwold T, Wagers AJ and Lee RT: Growth differentiation factor 11 is a circulating factor that reverses age-related cardiac hypertrophy. *Cell*, 2013; 153: 828-839
- 17) Biernacka A and Frangogiannis NG: Aging and Cardiac Fibrosis. *Aging Dis*, 2011; 2: 158-173
- 18) Roh JD, Houstis N, Yu A, Chang B, Yeri A, Li H, Hobson R, Lerchenmuller C, Vujic A, Chaudhari V, Damilano F, Platt C, Zlotoff D, Lee RT, Shah R, Jerosch-Herold M and Rosenzweig A: Exercise training reverses cardiac aging phenotypes associated with heart failure with preserved ejection fraction in male mice. *Aging Cell*, 2020; 19: e13159
- 19) Hemanthakumar KA, Fang S, Anisimov A, Mayranpaa MI, Mervaala E and Kivela R: Cardiovascular disease risk factors induce mesenchymal features and senescence in mouse cardiac endothelial cells. *Elife*, 2021; 10:
- 20) Tomanek RJ, Searls JC and Lachenbruch PA: Quantitative changes in the capillary bed during developing, peak, and stabilized cardiac hypertrophy in the spontaneously hypertensive rat. *Circ Res*, 1982; 51: 295-304
- 21) Li J, Zeng J, Wu L, Tao L, Liao Z, Chu M and Li L: Loss of P53 regresses cardiac remodeling induced by pressure overload partially through inhibiting HIF1alpha signaling in mice. *Biochem Biophys Res Commun*, 2018; 501: 394-399
- 22) Lopez-Otin C, Blasco MA, Partridge L, Serrano M and Kroemer G: The hallmarks of aging. *Cell*, 2013; 153: 1194-1217
- 23) Nakao M, Shimizu I, Katsuomi G, Yoshida Y, Suda M, Hayashi Y, Ikegami R, Hsiao YT, Okuda S, Soga T and Minamino T: Empagliflozin maintains capillarization and improves cardiac function in a murine model of left ventricular pressure overload. *Sci Rep*, 2021; 11: 18384
- 24) Obokata M, Reddy YNV, Pislaru SV, Melenovsky V and Borlaug BA: Evidence Supporting the Existence of a Distinct Obese Phenotype of Heart Failure With Preserved Ejection Fraction. *Circulation*, 2017; 136: 6-19
- 25) Campbell DJ, Somaratne JB, Prior DL, Yii M, Kenny JF, Newcomb AE, Kelly DJ and Black MJ: Obesity is associated with lower coronary microvascular density. *PLoS One*, 2013; 8: e81798
- 26) Machado MV, Vieira AB, da Conceicao FG, Nascimento AR, da Nobrega ACL and Tibirica E: Exercise training dose differentially alters muscle and heart capillary density and metabolic functions in an obese rat with metabolic syndrome. *Exp Physiol*, 2017; 102: 1716-1728
- 27) Suda M, Shimizu I, Yoshida Y, Hayashi Y, Ikegami R, Katsuomi G, Wakasugi T, Yoshida Y, Okuda S, Soga T and Minamino T: Inhibition of dipeptidyl peptidase-4 ameliorates cardiac ischemia and systolic dysfunction by

- up-regulating the FGF-2/EGR-1 pathway. *PLoS One*, 2017; 12: e0182422
- 28) Echouffo-Tcheugui JB, Xu H, DeVore AD, Schulte PJ, Butler J, Yancy CW, Bhatt DL, Hernandez AF, Heidenreich PA and Fonarow GC: Temporal trends and factors associated with diabetes mellitus among patients hospitalized with heart failure: Findings from Get With The Guidelines-Heart Failure registry. *Am Heart J*, 2016; 182: 9-20
 - 29) McHugh K, DeVore AD, Wu J, Matsouaka RA, Fonarow GC, Heidenreich PA, Yancy CW, Green JB, Altman N and Hernandez AF: Heart Failure With Preserved Ejection Fraction and Diabetes: JACC State-of-the-Art Review. *J Am Coll Cardiol*, 2019; 73: 602-611
 - 30) Hinkel R, Howe A, Renner S, Ng J, Lee S, Klett K, Kaczmarek V, Moretti A, Laugwitz KL, Skroblin P, Mayr M, Milting H, Dendorfer A, Reichart B, Wolf E and Kupatt C: Diabetes Mellitus-Induced Microvascular Destabilization in the Myocardium. *J Am Coll Cardiol*, 2017; 69: 131-143
 - 31) Ashoff A, Qadri F, Eggers R, Johren O, Raasch W and Dendorfer A: Pioglitazone prevents capillary rarefaction in streptozotocin-diabetic rats independently of glucose control and vascular endothelial growth factor expression. *J Vasc Res*, 2012; 49: 260-266
 - 32) Hedman A, Reneland R and Lithell HO: Alterations in skeletal muscle morphology in glucose-tolerant elderly hypertensive men: relationship to development of hypertension and heart rate. *J Hypertens*, 2000; 18: 559-565
 - 33) Yoshizawa S, Uto K, Nishikawa T, Hagiwara N and Oda H: Histological features of endomyocardial biopsies in patients undergoing hemodialysis: Comparison with dilated cardiomyopathy and hypertensive heart disease. *Cardiovasc Pathol*, 2020; 49: 107256
 - 34) Yazawa H, Miyachi M, Furukawa M, Takahashi K, Takatsu M, Tsuboi K, Ohtake M, Murase T, Hattori T, Kato Y, Murohara T and Nagata K: Angiotensin-converting enzyme inhibition promotes coronary angiogenesis in the failing heart of Dahl salt-sensitive hypertensive rats. *J Card Fail*, 2011; 17: 1041-1050
 - 35) Miyachi M, Yazawa H, Furukawa M, Tsuboi K, Ohtake M, Nishizawa T, Hashimoto K, Yokoi T, Kojima T, Murate T, Yokota M, Murohara T, Koike Y and Nagata K: Exercise training alters left ventricular geometry and attenuates heart failure in dahl salt-sensitive hypertensive rats. *Hypertension*, 2009; 53: 701-707
 - 36) Ihori H, Nozawa T, Sobajima M, Shida T, Fukui Y, Fujii N and Inoue H: Waon therapy attenuates cardiac hypertrophy and promotes myocardial capillary growth in hypertensive rats: a comparative study with fluvastatin. *Heart Vessels*, 2016; 31: 1361-1369
 - 37) Gonzalez GE, Rhaleb NE, D'Ambrosio MA, Nakagawa P, Liu Y, Leung P, Dai X, Yang XP, Peterson EL and Carretero OA: Deletion of interleukin-6 prevents cardiac inflammation, fibrosis and dysfunction without affecting blood pressure in angiotensin II-high salt-induced hypertension. *J Hypertens*, 2015; 33: 144-152
 - 38) Caudron J, Mulder P, Nicol L, Richard V, Thuillez C and Dacher JN: MR relaxometry and perfusion of the myocardium in spontaneously hypertensive rat: correlation with histopathology and effect of anti-hypertensive therapy. *Eur Radiol*, 2013; 23: 1871-1881
 - 39) Pu Q, Larouche I and Schiffrin EL: Effect of dual angiotensin converting enzyme/neutral endopeptidase inhibition, angiotensin converting enzyme inhibition, or AT1 antagonism on coronary microvasculature in spontaneously hypertensive rats. *Am J Hypertens*, 2003; 16: 931-937
 - 40) Olianti C, Costantini I, Giardini F, Lazzeri E, Crocini C, Ferrantini C, Pavone FS, Camici PG and Sacconi L: 3D imaging and morphometry of the heart capillary system in spontaneously hypertensive rats and normotensive controls. *Sci Rep*, 2020; 10: 14276
 - 41) Potente M and Makinen T: Vascular heterogeneity and specialization in development and disease. *Nat Rev Mol Cell Biol*, 2017; 18: 477-494
 - 42) Toblli JE, Cao G, DeRosa G, Di Gennaro F and Forcada P: Angiotensin-converting enzyme inhibition and angiogenesis in myocardium of obese Zucker rats. *Am J Hypertens*, 2004; 17: 172-180
 - 43) Zhu YC, Zhu YZ, Gohlke P, Stauss HM and Unger T: Effects of angiotensin-converting enzyme inhibition and angiotensin II AT1 receptor antagonism on cardiac parameters in left ventricular hypertrophy. *Am J Cardiol*, 1997; 80: 110A-117A
 - 44) Trenson S, Hermans H, Craps S, Pokreisz P, de Zeeuw P, Van Wauwe J, Gillijns H, Veltman D, Wei F, Caluwe E, Gijssbers R, Baatsen P, Staessen JA, Ghesquiere B, Carmeliet P, Rega F, Meuris B, Meyns B, Oosterlinck W, Duchenne J, Goetschalckx K, Voigt JU, Herregods MC, Herijgers P, Lutun A and Janssens S: Cardiac Microvascular Endothelial Cells in Pressure Overload-Induced Heart Disease. *Circ Heart Fail*, 2021; 14: e006979
 - 45) Mahmod M, Chan K, Raman B, Westaby J, Dass S, Petrou M, Sayeed R, Ashrafian H, Myerson SG, Karamitsos TD, Sheppard MN and Neubauer S: Histological Evidence for Impaired Myocardial Perfusion Reserve in Severe Aortic Stenosis. *JACC Cardiovasc Imaging*, 2019; 12: 2276-2278
 - 46) Go AS, Chertow GM, Fan D, McCulloch CE and Hsu CY: Chronic kidney disease and the risks of death, cardiovascular events, and hospitalization. *N Engl J Med*, 2004; 351: 1296-1305
 - 47) Amann K, Wiest G, Zimmer G, Gretz N, Ritz E and Mall G: Reduced capillary density in the myocardium of uremic rats--a stereological study. *Kidney Int*, 1992; 42: 1079-1085
 - 48) Tornig J, Amann K, Ritz E, Nichols C, Zeier M and Mall G: Arteriolar wall thickening, capillary rarefaction and interstitial fibrosis in the heart of rats with renal failure: the effects of ramipril, nifedipine and moxonidine. *J Am Soc Nephrol*, 1996; 7: 667-675
 - 49) Amann K, Odoni G, Benz K, Campean V, Jacobi J, Hilgers KF, Hartner A, Veelken R and Orth SR: Sympathetic blockade prevents the decrease in cardiac VEGF expression and capillary supply in experimental renal failure. *Am J Physiol Renal Physiol*, 2011; 300: F105-112
 - 50) Amann K, Munter K, Wessels S, Wagner J, Balajew V,

- Hergenroder S, Mall G and Ritz E: Endothelin A receptor blockade prevents capillary/myocyte mismatch in the heart of uremic animals. *J Am Soc Nephrol*, 2000; 11: 1702-1711
- 51) Amann K, Gassmann P, Buzello M, Orth SR, Tornig J, Gross ML, Magener A, Mall G and Ritz E: Effects of ACE inhibition and bradykinin antagonism on cardiovascular changes in uremic rats. *Kidney Int*, 2000; 58: 153-161
 - 52) Gut N, Piecha G, Aldebsi F, Schaefer S, Bekeredjian R, Schirmacher P, Ritz E and Gross-Weissmann ML: Erythropoietin combined with ACE inhibitor prevents heart remodeling in 5/6 nephrectomized rats independently of blood pressure and kidney function. *Am J Nephrol*, 2013; 38: 124-135
 - 53) Di Marco GS, Reuter S, Kentrup D, Ting L, Ting L, Grabner A, Jacobi AM, Pavenstadt H, Baba HA, Tiemann K and Brand M: Cardioprotective effect of calcineurin inhibition in an animal model of renal disease. *Eur Heart J*, 2011; 32: 1935-1945
 - 54) Di Marco GS, Kentrup D, Reuter S, Mayer AB, Golle L, Tiemann K, Fobker M, Engelbertz C, Breithardt G, Brand E, Reinecke H, Pavenstadt H and Brand M: Soluble Flt-1 links microvascular disease with heart failure in CKD. *Basic Res Cardiol*, 2015; 110: 30
 - 55) Neben-Wittich MA, Wittich CM, Mueller PS, Larson DR, Gertz MA and Edwards WD: Obstructive intramural coronary amyloidosis and myocardial ischemia are common in primary amyloidosis. *Am J Med*, 2005; 118: 1287
 - 56) Maleszewski JJ: Cardiac amyloidosis: pathology, nomenclature, and typing. *Cardiovasc Pathol*, 2015; 24: 343-350
 - 57) Dorbala S, Vangala D, Bruyere J, Jr., Quarta C, Kruger J, Padera R, Foster C, Hanley M, Di Carli MF and Falk R: Coronary microvascular dysfunction is related to abnormalities in myocardial structure and function in cardiac amyloidosis. *JACC Heart Fail*, 2014; 2: 358-367
 - 58) Kim D, Choi JO, Kim K, Kim SJ, Kim JS and Jeon ES: Clinical and prognostic implications of capillary density in patients with cardiac light chain amyloidosis. *ESC Heart Fail*, 2021; 8: 5594-5599
 - 59) Guclu A, Happe C, Eren S, Korkmaz IH, Niessen HW, Klein P, van Slegtenhorst M, Schinkel AF, Michels M, van Rossum AC, Germans T and van der Velden J: Left ventricular outflow tract gradient is associated with reduced capillary density in hypertrophic cardiomyopathy irrespective of genotype. *Eur J Clin Invest*, 2015; 45: 1252-1259
 - 60) Johansson B, Morner S, Waldenstrom A and Stal P: Myocardial capillary supply is limited in hypertrophic cardiomyopathy: a morphological analysis. *Int J Cardiol*, 2008; 126: 252-257
 - 61) Ku MC, Kober F, Lai YC, Pohlmann A, Qadri F, Bader M, Carrier L and Niendorf T: Cardiovascular magnetic resonance detects microvascular dysfunction in a mouse model of hypertrophic cardiomyopathy. *J Cardiovasc Magn Reson*, 2021; 23: 63
 - 62) Shimizu I, Yoshida Y, Suda M and Minamino T: DNA damage response and metabolic disease. *Cell Metab*, 2014; 20: 967-977
 - 63) Katsuumi G, Shimizu I, Yoshida Y and Minamino T: Vascular Senescence in Cardiovascular and Metabolic Diseases. *Front Cardiovasc Med*, 2018; 5: 18
 - 64) Shimizu I, Yoshida Y, Katsuno T, Tateno K, Okada S, Moriya J, Yokoyama M, Nojima A, Ito T, Zechner R, Komuro I, Kobayashi Y and Minamino T: p53-induced adipose tissue inflammation is critically involved in the development of insulin resistance in heart failure. *Cell Metab*, 2012; 15: 51-64
 - 65) Shimizu I, Yoshida Y, Moriya J, Nojima A, Uemura A, Kobayashi Y and Minamino T: Semaphorin3E-induced inflammation contributes to insulin resistance in dietary obesity. *Cell Metab*, 2013; 18: 491-504
 - 66) Yoshida Y, Shimizu I, Katsuumi G, Jiao S, Suda M, Hayashi Y and Minamino T: p53-Induced inflammation exacerbates cardiac dysfunction during pressure overload. *J Mol Cell Cardiol*, 2015; 85: 183-198
 - 67) Minamino T, Orimo M, Shimizu I, Kunieda T, Yokoyama M, Ito T, Nojima A, Nabetani A, Oike Y, Matsubara H, Ishikawa F and Komuro I: A crucial role for adipose tissue p53 in the regulation of insulin resistance. *Nat Med*, 2009; 15: 1082-1087
 - 68) Yokoyama M, Okada S, Nakagomi A, Moriya J, Shimizu I, Nojima A, Yoshida Y, Ichimiya H, Kamimura N, Kobayashi Y, Ohta S, Fruttiger M, Lozano G and Minamino T: Inhibition of endothelial p53 improves metabolic abnormalities related to dietary obesity. *Cell Rep*, 2014; 7: 1691-1703
 - 69) Wu L, Liu D, Wu Y, Wei X, Wang Z, Wang W, Zhang S, Yang H, Yi M and Liu H: p53 mediated transcription of Omi/HtrA2 in aging myocardium. *Biochem Biophys Res Commun*, 2019; 519: 734-739
 - 70) Ravi R, Mookerjee B, Bhujwala ZM, Sutter CH, Artemov D, Zeng Q, Dillehay LE, Madan A, Semenza GL and Bedi A: Regulation of tumor angiogenesis by p53-induced degradation of hypoxia-inducible factor 1alpha. *Genes Dev*, 2000; 14: 34-44
 - 71) Guan A, Gong H, Ye Y, Jia J, Zhang G, Li B, Yang C, Qian S, Sun A, Chen R, Ge J and Zou Y: Regulation of p53 by jagged1 contributes to angiotensin II-induced impairment of myocardial angiogenesis. *PLoS One*, 2013; 8: e76529
 - 72) Gu J, Wang S, Guo H, Tan Y, Liang Y, Feng A, Liu Q, Damodaran C, Zhang Z, Keller BB, Zhang C and Cai L: Inhibition of p53 prevents diabetic cardiomyopathy by preventing early-stage apoptosis and cell senescence, reduced glycolysis, and impaired angiogenesis. *Cell Death Dis*, 2018; 9: 82
 - 73) el-Deiry WS, Tokino T, Velculescu VE, Levy DB, Parsons R, Trent JM, Lin D, Mercer WE, Kinzler KW and Vogelstein B: WAF1, a potential mediator of p53 tumor suppression. *Cell*, 1993; 75: 817-825
 - 74) Spyridopoulos I, Isner JM and Losordo DW: Oncogenic ras induces premature senescence in endothelial cells: role of p21(Cip1/Waf1). *Basic Res Cardiol*, 2002; 97: 117-124
 - 75) Lee CL, Moding EJ, Cuneo KC, Li Y, Sullivan JM, Mao L, Washington I, Jeffords LB, Rodrigues RC, Ma Y, Das S, Kontos CD, Kim Y, Rockman HA and Kirsch DG: p53 functions in endothelial cells to prevent radiation-induced myocardial injury in mice. *Sci Signal*, 2012; 5:

- ra52
- 76) Zeng ZM, Du HY, Xiong L, Zeng XL, Zhang P, Cai J, Huang L and Liu AW: BRCA1 protects cardiac microvascular endothelial cells against irradiation by regulating p21-mediated cell cycle arrest. *Life Sci*, 2020; 244: 117342
 - 77) Zhang Z, Oh M, Sasaki JI and Nor JE: Inverse and reciprocal regulation of p53/p21 and Bmi-1 modulates vasculogenic differentiation of dental pulp stem cells. *Cell Death Dis*, 2021; 12: 644
 - 78) Lee SH, Lee JH, Yoo SY, Hur J, Kim HS and Kwon SM: Hypoxia inhibits cellular senescence to restore the therapeutic potential of old human endothelial progenitor cells via the hypoxia-inducible factor-1 α -TWIST-p21 axis. *Arterioscler Thromb Vasc Biol*, 2013; 33: 2407-2414
 - 79) Li JM, Fan LM, George VT and Brooks G: Nox2 regulates endothelial cell cycle arrest and apoptosis via p21cip1 and p53. *Free Radic Biol Med*, 2007; 43: 976-986
 - 80) Anderson R, Lagnado A, Maggiorani D, Walaszczyk A, Dookun E, Chapman J, Birch J, Salmonowicz H, Ogrodnik M, Jurk D, Proctor C, Correia-Melo C, Victorelli S, Fielder E, Berlinguer-Palmini R, Owens A, Greaves LC, Kolsky KL, Parini A, Douin-Echinard V, LeBrasseur NK, Arthur HM, Tual-Chalot S, Schafer MJ, Roos CM, Miller JD, Robertson N, Mann J, Adams PD, Tchkonina T, Kirkland JL, Mialet-Perez J, Richardson GD and Passos JF: Length-independent telomere damage drives post-mitotic cardiomyocyte senescence. *EMBO J*, 2019; 38:
 - 81) Rossman MJ, Kaplon RE, Hill SD, McNamara MN, Santos-Parker JR, Pierce GL, Seals DR and Donato AJ: Endothelial cell senescence with aging in healthy humans: prevention by habitual exercise and relation to vascular endothelial function. *Am J Physiol Heart Circ Physiol*, 2017; 313: H890-H895
 - 82) Lewis-McDougall FC, Ruchaya PJ, Domenjo-Vila E, Shin Teoh T, Prata L, Cottle BJ, Clark JE, Punjabi PP, Awad W, Torella D, Tchkonina T, Kirkland JL and Ellison-Hughes GM: Aged-senescent cells contribute to impaired heart regeneration. *Aging Cell*, 2019; 18: e12931
 - 83) Yin W, Guo J, Zhang C, Alibhai FJ, Li SH, Billia P, Wu J, Yau TM, Weisel RD and Li RK: Knockout of Canopy 2 activates p16(INK4a) pathway to impair cardiac repair. *J Mol Cell Cardiol*, 2019; 132: 36-48
 - 84) Lee DH, Wolstein JM, Pudasaini B and Plotkin M: INK4a deletion results in improved kidney regeneration and decreased capillary rarefaction after ischemia-reperfusion injury. *Am J Physiol Renal Physiol*, 2012; 302: F183-191
 - 85) Yoshino J, Baur JA and Imai SI: NAD(+) Intermediates: The Biology and Therapeutic Potential of NMN and NR. *Cell Metab*, 2018; 27: 513-528
 - 86) Zeng H and Chen JX: Microvascular Rarefaction and Heart Failure With Preserved Ejection Fraction. *Front Cardiovasc Med*, 2019; 6: 15
 - 87) Ota H, Akishita M, Eto M, Iijima K, Kaneki M and Ouchi Y: Sirt1 modulates premature senescence-like phenotype in human endothelial cells. *J Mol Cell Cardiol*, 2007; 43: 571-579
 - 88) Das A, Huang GX, Bonkowski MS, Longchamp A, Li C, Schultz MB, Kim LJ, Osborne B, Joshi S, Lu Y, Trevino-Villarreal JH, Kang MJ, Hung TT, Lee B, Williams EO, Igarashi M, Mitchell JR, Wu LE, Turner N, Arany Z, Guarente L and Sinclair DA: Impairment of an Endothelial NAD(+)-H2S Signaling Network Is a Reversible Cause of Vascular Aging. *Cell*, 2018; 173: 74-89 e20
 - 89) Maizel J, Xavier S, Chen J, Lin CH, Vasko R and Goligorsky MS: Sirtuin 1 ablation in endothelial cells is associated with impaired angiogenesis and diastolic dysfunction. *Am J Physiol Heart Circ Physiol*, 2014; 307: H1691-1704
 - 90) He X, Zeng H, Chen ST, Roman RJ, Aschner JL, Didion S and Chen JX: Endothelial specific SIRT3 deletion impairs glycolysis and angiogenesis and causes diastolic dysfunction. *J Mol Cell Cardiol*, 2017; 112: 104-113
 - 91) Su H, Zeng H, Liu B and Chen JX: Sirtuin 3 is essential for hypertension-induced cardiac fibrosis via mediating pericyte transition. *J Cell Mol Med*, 2020; 24: 8057-8068
 - 92) Shiojima I and Walsh K: Regulation of cardiac growth and coronary angiogenesis by the Akt/PKB signaling pathway. *Genes Dev*, 2006; 20: 3347-3365
 - 93) Hua YN, Zhang YM, Ceylan-Isik AF, Wold LE, Nunn JM and Ren J: Chronic akt activation accentuates aging-induced cardiac hypertrophy and myocardial contractile dysfunction: role of autophagy. *Basic Research in Cardiology*, 2011; 106: 1173-1191
 - 94) Shimizu I and Minamino T: Physiological and pathological cardiac hypertrophy. *Journal of Molecular and Cellular Cardiology*, 2016; 97: 245-262
 - 95) Shiojima I, Sato K, Izumiya Y, Schiekofer S, Ito M, Liao R, Colucci WS and Walsh K: Disruption of coordinated cardiac hypertrophy and angiogenesis contributes to the transition to heart failure. *J Clin Invest*, 2005; 115: 2108-2118
 - 96) Chaanine AH and Hajjar RJ: AKT signalling in the failing heart. *Eur J Heart Fail*, 2011; 13: 825-829
 - 97) Shiojima I and Walsh K: Role of Akt signaling in vascular homeostasis and angiogenesis. *Circ Res*, 2002; 90: 1243-1250
 - 98) Miyauchi H, Minamino T, Tateno K, Kunieda T, Toko H and Komuro I: Akt negatively regulates the in vitro lifespan of human endothelial cells via a p53/p21-dependent pathway. *EMBO J*, 2004; 23: 212-220
 - 99) Hsiao YT, Shimizu I, Yoshida Y and Minamino T: Role of circulating molecules in age-related cardiovascular and metabolic disorders. *Inflamm Regen*, 2022; 42: 2
 - 100) Alvarez-Rodriguez L, Lopez-Hoyos M, Munoz-Cacho P and Martinez-Taboada VM: Aging is associated with circulating cytokine dysregulation. *Cellular Immunology*, 2012; 273: 124-132
 - 101) Zhang B, Li XL, Zhao CR, Pan CL and Zhang Z: Interleukin-6 as a Predictor of the Risk of Cardiovascular Disease: A Meta-Analysis of Prospective Epidemiological Studies. *Immunological Investigations*, 2018; 47: 689-699
 - 102) Edsfeldt A, Grufman H, Ascitto G, Nitulescu M,

- Persson A, Nilsson M, Nilsson J and Goncalves I: Circulating cytokines reflect the expression of pro-inflammatory cytokines in atherosclerotic plaques. *Atherosclerosis*, 2015; 241: 443-449
- 103) Dunlay SM, Weston SA, Redfield MM, Killian JM and Roger VL: Tumor necrosis factor-alpha and mortality in heart failure - A community study. *Circulation*, 2008; 118: 625-631
- 104) Cheng JM, Akkerhuis KM, Battes LC, van Vark LC, Hillege HL, Paulus WJ, Boersma E and Kardys I: Biomarkers of heart failure with normal ejection fraction: a systematic review. *Eur J Heart Fail*, 2013; 15: 1350-1362
- 105) Franssen C, Chen S, Unger A, Korkmaz HI, De Keulenaer GW, Tschope C, Leite-Moreira AF, Musters R, Niessen HW, Linke WA, Paulus WJ and Hamdani N: Myocardial Microvascular Inflammatory Endothelial Activation in Heart Failure With Preserved Ejection Fraction. *JACC Heart Fail*, 2016; 4: 312-324
- 106) Bagi Z: Mechanisms of coronary microvascular adaptation to obesity. *Am J Physiol Regul Integr Comp Physiol*, 2009; 297: R556-567
- 107) Vulesevic B, McNeill B, Giacco F, Maeda K, Blackburn NJ, Brownlee M, Milne RW and Suuronen EJ: Methylglyoxal-Induced Endothelial Cell Loss and Inflammation Contribute to the Development of Diabetic Cardiomyopathy. *Diabetes*, 2016; 65: 1699-1713
- 108) Krishnamurthy P, Rajasingh J, Lambers E, Qin G, Losordo DW and Kishore R: IL-10 inhibits inflammation and attenuates left ventricular remodeling after myocardial infarction via activation of STAT3 and suppression of HuR. *Circ Res*, 2009; 104: e9-18
- 109) Gupta R, Liu L, Zhang X, Fan X, Krishnamurthy P, Verma S, Tongers J, Misener S, Ashcherkin N, Sun H, Tian J and Kishore R: IL-10 provides cardioprotection in diabetic myocardial infarction via upregulation of Heme clearance pathways. *JCI Insight*, 2020; 5:
- 110) Banerjee I, Fuseler JW, Intwala AR and Baudino TA: IL-6 loss causes ventricular dysfunction, fibrosis, reduced capillary density, and dramatically alters the cell populations of the developing and adult heart. *Am J Physiol Heart Circ Physiol*, 2009; 296: H1694-1704
- 111) Rienks M, Carai P, Bitsch N, Schellings M, Vanhaverbeke M, Verjans J, Cuijpers I, Heymans S and Papageorgiou A: Sema3A promotes the resolution of cardiac inflammation after myocardial infarction. *Basic Res Cardiol*, 2017; 112: 42
- 112) Freed JK and Guterman DD: Mitochondrial reactive oxygen species and vascular function: less is more. *Arterioscler Thromb Vasc Biol*, 2013; 33: 673-675
- 113) Fukai T and Ushio-Fukai M: Cross-Talk between NADPH Oxidase and Mitochondria: Role in ROS Signaling and Angiogenesis. *Cells*, 2020; 9:
- 114) Oshikawa J, Kim SJ, Furuta E, Caliceti C, Chen GF, McKinney RD, Kuhr F, Levitan I, Fukai T and Ushio-Fukai M: Novel role of p66Shc in ROS-dependent VEGF signaling and angiogenesis in endothelial cells. *Am J Physiol Heart Circ Physiol*, 2012; 302: H724-732
- 115) Speer T, Owala FO, Holy EW, Zewinger S, Frenzel FL, Stahl BE, Razavi M, Triem S, Cvija H, Rohrer L, Seiler S, Heine GH, Jankowski V, Jankowski J, Camici GG, Akhmedov A, Fliser D, Luscher TF and Tanner FC: Carbamylated low-density lipoprotein induces endothelial dysfunction. *Eur Heart J*, 2014; 35: 3021-3032
- 116) Petrosillo G, Matera M, Moro N, Ruggiero FM and Paradies G: Mitochondrial complex I dysfunction in rat heart with aging: critical role of reactive oxygen species and cardiolipin. *Free Radic Biol Med*, 2009; 46: 88-94
- 117) Zhang M, Perino A, Ghigo A, Hirsch E and Shah AM: NADPH oxidases in heart failure: poachers or gamekeepers? *Antioxid Redox Signal*, 2013; 18: 1024-1041
- 118) Fan LM, Geng L, Cahill-Smith S, Liu F, Douglas G, McKenzie CA, Smith C, Brooks G, Channon KM and Li JM: Nox2 contributes to age-related oxidative damage to neurons and the cerebral vasculature. *J Clin Invest*, 2019; 129: 3374-3386
- 119) Tojo T, Ushio-Fukai M, Yamaoka-Tojo M, Ikeda S, Patrushev N and Alexander RW: Role of gp91phox (Nox2)-containing NAD(P)H oxidase in angiogenesis in response to hindlimb ischemia. *Circulation*, 2005; 111: 2347-2355
- 120) Douglas G, Bendall JK, Crabtree MJ, Tatham AL, Carter EE, Hale AB and Channon KM: Endothelial-specific Nox2 overexpression increases vascular superoxide and macrophage recruitment in ApoE(-)/(-) mice. *Cardiovasc Res*, 2012; 94: 20-29
- 121) Murdoch CE, Chaubey S, Zeng L, Yu B, Ivetic A, Walker SJ, Vanhoutte D, Heymans S, Grieve DJ, Cave AC, Brewer AC, Zhang M and Shah AM: Endothelial NADPH oxidase-2 promotes interstitial cardiac fibrosis and diastolic dysfunction through proinflammatory effects and endothelial-mesenchymal transition. *J Am Coll Cardiol*, 2014; 63: 2734-2741
- 122) Zhang M, Brewer AC, Schroder K, Santos CX, Grieve DJ, Wang M, Anilkumar N, Yu B, Dong X, Walker SJ, Brandes RP and Shah AM: NADPH oxidase-4 mediates protection against chronic load-induced stress in mouse hearts by enhancing angiogenesis. *Proc Natl Acad Sci U S A*, 2010; 107: 18121-18126
- 123) Schroder K, Zhang M, Benkhoff S, Mieth A, Pliquett R, Kosowski J, Kruse C, Luedike P, Michaelis UR, Weissmann N, Dimmeler S, Shah AM and Brandes RP: Nox4 is a protective reactive oxygen species generating vascular NADPH oxidase. *Circ Res*, 2012; 110: 1217-1225
- 124) Murphy SP, Ibrahim NE and Januzzi JL, Jr.: Heart Failure With Reduced Ejection Fraction: A Review. *JAMA*, 2020; 324: 488-504
- 125) McMurray JJV and Packer M: How Should We Sequence the Treatments for Heart Failure and a Reduced Ejection Fraction?: A Redefinition of Evidence-Based Medicine. *Circulation*, 2021; 143: 875-877
- 126) McMurray JJ, Ostergren J, Swedberg K, Granger CB, Held P, Michelson EL, Olofsson B, Yusuf S, Pfeffer MA, Investigators C and Committees: Effects of candesartan in patients with chronic heart failure and reduced left-ventricular systolic function taking angiotensin-converting-enzyme inhibitors: the CHARM-Added trial. *Lancet*, 2003; 362: 767-771

- 127) Group CTS: Effects of enalapril on mortality in severe congestive heart failure. Results of the Cooperative North Scandinavian Enalapril Survival Study (CONSENSUS). *N Engl J Med*, 1987; 316: 1429-1435
- 128) Investigators S, Yusuf S, Pitt B, Davis CE, Hood WB and Cohn JN: Effect of enalapril on survival in patients with reduced left ventricular ejection fractions and congestive heart failure. *N Engl J Med*, 1991; 325: 293-302
- 129) Hayashi T, Sohmiya K, Ukimura A, Endoh S, Mori T, Shimomura H, Okabe M, Terasaki F and Kitaura Y: Angiotensin II receptor blockade prevents microangiopathy and preserves diastolic function in the diabetic rat heart. *Heart*, 2003; 89: 1236-1242
- 130) de Boer RA, Pinto YM, Suurmeijer AJ, Pokharel S, Scholtens E, Humler M, Saavedra JM, Boomsma F, van Gilst WH and van Veldhuisen DJ: Increased expression of cardiac angiotensin II type 1 (AT(1)) receptors decreases myocardial microvessel density after experimental myocardial infarction. *Cardiovasc Res*, 2003; 57: 434-442
- 131) Nako H, Kataoka K, Koibuchi N, Dong YF, Toyama K, Yamamoto E, Yasuda O, Ichijo H, Ogawa H and Kim-Mitsuyama S: Novel mechanism of angiotensin II-induced cardiac injury in hypertensive rats: the critical role of ASK1 and VEGF. *Hypertens Res*, 2012; 35: 194-200
- 132) Parenti A, Morbidelli L, Ledda F, Granger HJ and Ziche M: The bradykinin/B1 receptor promotes angiogenesis by up-regulation of endogenous FGF-2 in endothelium via the nitric oxide synthase pathway. *FASEB J*, 2001; 15: 1487-1489
- 133) Silvestre JS, Bergaya S, Tamarat R, Duriez M, Boulanger CM and Levy BI: Proangiogenic effect of angiotensin-converting enzyme inhibition is mediated by the bradykinin B(2) receptor pathway. *Circ Res*, 2001; 89: 678-683
- 134) Tamarat R, Silvestre JS, Kubis N, Benessiano J, Duriez M, deGasparo M, Henrion D and Levy BI: Endothelial nitric oxide synthase lies downstream from angiotensin II-induced angiogenesis in ischemic hindlimb. *Hypertension*, 2002; 39: 830-835
- 135) Imanishi T, Hano T and Nishio I: Angiotensin II potentiates vascular endothelial growth factor-induced proliferation and network formation of endothelial progenitor cells. *Hypertens Res*, 2004; 27: 101-108
- 136) Siddiqui AJ, Mansson-Broberg A, Gustafsson T, Grinnemo KH, Dellgren G, Hao X, Fischer H and Sylven C: Antagonism of the renin-angiotensin system can counteract cardiac angiogenic vascular endothelial growth factor gene therapy and myocardial angiogenesis in the normal heart. *Am J Hypertens*, 2005; 18: 1347-1352
- 137) Jesmin S, Hattori Y, Sakuma I, Mowa CN and Kitabatake A: Role of ANG II in coronary capillary angiogenesis at the insulin-resistant stage of a NIDDM rat model. *Am J Physiol Heart Circ Physiol*, 2002; 283: H1387-1397
- 138) Belabbas H, Zalvidea S, Casellas D, Moles JP, Galbes O, Mercier J and Jover B: Contrasting effect of exercise and angiotensin II hypertension on in vivo and in vitro cardiac angiogenesis in rats. *Am J Physiol Regul Integr Comp Physiol*, 2008; 295: R1512-1518
- 139) Braunwald E: The path to an angiotensin receptor antagonist-neprilysin inhibitor in the treatment of heart failure. *J Am Coll Cardiol*, 2015; 65: 1029-1041
- 140) McMurray JJ, Packer M, Desai AS, Gong J, Lefkowitz MP, Rizkala AR, Rouleau JL, Shi VC, Solomon SD, Swedberg K, Zile MR, Investigators P-H and Committees: Angiotensin-neprilysin inhibition versus enalapril in heart failure. *N Engl J Med*, 2014; 371: 993-1004
- 141) Pfau D, Thorn SL, Zhang J, Mikush N, Renaud JM, Klein R, deKemp RA, Wu X, Hu X, Sinusas AJ, Young LH and Tirziu D: Angiotensin Receptor Neprilysin Inhibitor Attenuates Myocardial Remodeling and Improves Infarct Perfusion in Experimental Heart Failure. *Sci Rep*, 2019; 9: 5791
- 142) Suematsu Y, Tashiro K, Morita H, Ideishi A, Kuwano T and Miura SI: Angiotensin Receptor Blocker and Neprilysin Inhibitor Suppresses Cardiac Dysfunction by Accelerating Myocardial Angiogenesis in Apolipoprotein E-Knockout Mice Fed a High-Fat Diet. *J Renin Angiotensin Aldosterone Syst*, 2021; 2021: 9916789
- 143) Zheng W, Brown MD, Brock TA, Bjercke RJ and Tomanek RJ: Bradycardia-induced coronary angiogenesis is dependent on vascular endothelial growth factor. *Circ Res*, 1999; 85: 192-198
- 144) Lei L, Zhou R, Zheng W, Christensen LP, Weiss RM and Tomanek RJ: Bradycardia induces angiogenesis, increases coronary reserve, and preserves function of the postinfarcted heart. *Circulation*, 2004; 110: 796-802
- 145) Ulu N, Henning RH, Goris M, Schoemaker RG and van Gilst WH: Effects of ivabradine and metoprolol on cardiac angiogenesis and endothelial dysfunction in rats with heart failure. *J Cardiovasc Pharmacol*, 2009; 53: 9-17
- 146) Lamping KG, Zheng W, Xing D, Christensen LP, Martins J and Tomanek RJ: Bradycardia stimulates vascular growth during gradual coronary occlusion. *Arterioscler Thromb Vasc Biol*, 2005; 25: 2122-2127
- 147) Zheng W, Seftor EA, Meininger CJ, Hendrix MJ and Tomanek RJ: Mechanisms of coronary angiogenesis in response to stretch: role of VEGF and TGF-beta. *Am J Physiol Heart Circ Physiol*, 2001; 280: H909-917
- 148) Amann K, Hofstetter J, Campean V, Koch A, Gross ML, Veelken R and Ritz E: Nonhypotensive dose of beta-adrenergic blocker ameliorates capillary deficits in the hearts of rats with moderate renal failure. *Virchows Arch*, 2006; 449: 207-214
- 149) Swedberg K, Komajda M, Bohm M, Borer JS, Ford I, Dubost-Brama A, Lerebours G, Tavazzi L and Investigators S: Ivabradine and outcomes in chronic heart failure (SHIFT): a randomised placebo-controlled study. *Lancet*, 2010; 376: 875-885
- 150) Wu X, You W, Wu Z, Ye F and Chen S: Ivabradine promotes angiogenesis and reduces cardiac hypertrophy in mice with myocardial infarction. *Anatol J Cardiol*, 2018; 20: 266-272
- 151) Neal B, Perkovic V, Mahaffey KW, de Zeeuw D, Fulcher G, Erondou N, Shaw W, Law G, Desai M, Matthews DR and Group CPC: Canagliflozin and Cardiovascular and

- Renal Events in Type 2 Diabetes. *N Engl J Med*, 2017; 377: 644-657
- 152) Perkovic V, Jardine MJ, Neal B, Bompoint S, Heerspink HJL, Charytan DM, Edwards R, Agarwal R, Bakris G, Bull S, Cannon CP, Capuano G, Chu PL, de Zeeuw D, Greene T, Levin A, Pollock C, Wheeler DC, Yavin Y, Zhang H, Zinman B, Meininger G, Brenner BM, Mahaffey KW and Investigators CT: Canagliflozin and Renal Outcomes in Type 2 Diabetes and Nephropathy. *N Engl J Med*, 2019; 380: 2295-2306
 - 153) Zelniker TA, Wiviott SD, Raz I, Im K, Goodrich EL, Bonaca MP, Mosenson O, Kato ET, Cahn A, Furtado RHM, Bhatt DL, Leiter LA, McGuire DK, Wilding JPH and Sabatine MS: SGLT2 inhibitors for primary and secondary prevention of cardiovascular and renal outcomes in type 2 diabetes: a systematic review and meta-analysis of cardiovascular outcome trials. *Lancet*, 2019; 393: 31-39
 - 154) Packer M, Anker SD, Butler J, Filippatos G, Pocock SJ, Carson P, Januzzi J, Verma S, Tsutsui H, Brueckmann M, Jamal W, Kimura K, Schnee J, Zeller C, Cotton D, Bocchi E, Bohm M, Choi DJ, Chopra V, Chuquiure E, Giannetti N, Janssens S, Zhang J, Gonzalez Juanatey JR, Kaul S, Brunner-La Rocca HP, Merkely B, Nicholls SJ, Perrone S, Pina I, Ponikowski P, Sattar N, Senni M, Serrone MF, Spinar J, Squire I, Taddei S, Wanner C, Zannad F and Investigators EM-RT: Cardiovascular and Renal Outcomes with Empagliflozin in Heart Failure. *N Engl J Med*, 2020; 383: 1413-1424
 - 155) Lahnwong S, Chattipakorn SC and Chattipakorn N: Potential mechanisms responsible for cardioprotective effects of sodium-glucose co-transporter 2 inhibitors. *Cardiovasc Diabetol*, 2018; 17: 101
 - 156) Adingupu DD, Gopel SO, Gronros J, Behrendt M, Sotak M, Miliotis T, Dahlqvist U, Gan LM and Jonsson-Rylander AC: SGLT2 inhibition with empagliflozin improves coronary microvascular function and cardiac contractility in prediabetic ob/ob(-/-) mice. *Cardiovasc Diabetol*, 2019; 18: 16
 - 157) Zhou H, Wang S, Zhu P, Hu S, Chen Y and Ren J: Empagliflozin rescues diabetic myocardial microvascular injury via AMPK-mediated inhibition of mitochondrial fission. *Redox Biol*, 2018; 15: 335-346
 - 158) Pitt B, Zannad F, Remme WJ, Cody R, Castaigne A, Perez A, Palensky J and Wittes J: The effect of spironolactone on morbidity and mortality in patients with severe heart failure. Randomized Aldactone Evaluation Study Investigators. *N Engl J Med*, 1999; 341: 709-717
 - 159) Beygui F, Collet JP, Benoliel JJ, Vignolles N, Dumaine R, Barthelemy O and Montalescot G: High plasma aldosterone levels on admission are associated with death in patients presenting with acute ST-elevation myocardial infarction. *Circulation*, 2006; 114: 2604-2610
 - 160) Fuller PJ and Young MJ: Mechanisms of mineralocorticoid action. *Hypertension*, 2005; 46: 1227-1235
 - 161) Fraccarollo D, Berger S, Galuppo P, Kneitz S, Hein L, Schutz G, Frantz S, Ertl G and Bauersachs J: Deletion of cardiomyocyte mineralocorticoid receptor ameliorates adverse remodeling after myocardial infarction. *Circulation*, 2011; 123: 400-408
 - 162) Lother A, Deng L, Huck M, Furst D, Kowalski J, Esser JS, Moser M, Bode C and Hein L: Endothelial cell mineralocorticoid receptors oppose VEGF-induced gene expression and angiogenesis. *J Endocrinol*, 2019; 240: 15-26
 - 163) Kim SK, Biber LA, Moss ME, Man JJ, Aronovitz MJ, Martin GL, Carrillo-Salinas FJ, Salvador AM, Alcaide P and Jaffe IZ: Mineralocorticoid Receptor in Smooth Muscle Contributes to Pressure Overload-Induced Heart Failure. *Circ Heart Fail*, 2021; 14: e007279
 - 164) Salvador AM, Moss ME, Aronovitz M, Mueller KB, Blanton RM, Jaffe IZ and Alcaide P: Endothelial mineralocorticoid receptor contributes to systolic dysfunction induced by pressure overload without modulating cardiac hypertrophy or inflammation. *Physiol Rep*, 2017; 5:
 - 165) Bentley-Lewis R, Adler GK, Perlstein T, Seely EW, Hopkins PN, Williams GH and Garg R: Body mass index predicts aldosterone production in normotensive adults on a high-salt diet. *J Clin Endocrinol Metab*, 2007; 92: 4472-4475
 - 166) Gorini S, Kim SK, Infante M, Mammi C, La Vignera S, Fabbri A, Jaffe IZ and Caprio M: Role of Aldosterone and Mineralocorticoid Receptor in Cardiovascular Aging. *Front Endocrinol (Lausanne)*, 2019; 10: 584
 - 167) Bistola V, Lambadiari V, Dimitriadis G, Ioannidis I, Makrilakis K, Tentolouris N, Tsapas A and Parissis J: Possible mechanisms of direct cardiovascular impact of GLP-1 agonists and DPP4 inhibitors. *Heart Fail Rev*, 2018; 23: 377-388
 - 168) Green JB, Bethel MA, Armstrong PW, Buse JB, Engel SS, Garg J, Josse R, Kaufman KD, Koglin J, Korn S, Lachin JM, McGuire DK, Pencina MJ, Standl E, Stein PP, Suryawanshi S, Van de Werf F, Peterson ED, Holman RR and Group TS: Effect of Sitagliptin on Cardiovascular Outcomes in Type 2 Diabetes. *N Engl J Med*, 2015; 373: 232-242
 - 169) Scirica BM, Bhatt DL, Braunwald E, Steg PG, Davidson J, Hirshberg B, Ohman P, Frederick R, Wiviott SD, Hoffman EB, Cavender MA, Udell JA, Desai NR, Mosenson O, McGuire DK, Ray KK, Leiter LA, Raz I, Committee S-TS and Investigators: Saxagliptin and cardiovascular outcomes in patients with type 2 diabetes mellitus. *N Engl J Med*, 2013; 369: 1317-1326
 - 170) Zannad F, Cannon CP, Cushman WC, Bakris GL, Menon V, Perez AT, Fleck PR, Mehta CR, Kupfer S, Wilson C, Lam H, White WB and Investigators E: Heart failure and mortality outcomes in patients with type 2 diabetes taking alogliptin versus placebo in EXAMINE: a multicentre, randomised, double-blind trial. *Lancet*, 2015; 385: 2067-2076
 - 171) Shigeta T, Aoyama M, Bando YK, Monji A, Mitsui T, Takatsu M, Cheng XW, Okumura T, Hirashiki A, Nagata K and Murohara T: Dipeptidyl peptidase-4 modulates left ventricular dysfunction in chronic heart failure via angiogenesis-dependent and -independent actions. *Circulation*, 2012; 126: 1838-1851
 - 172) Fiordaliso F, Maggioni S, Balconi G, Schiarea S, Corbelli A, De Luigi A, Figliuzzi M, Antoniou X, Chiabrando C,

- Masson S, Cervo L and Latini R: Effects of dipeptidyl peptidase-4 (DPP-4) inhibition on angiogenesis and hypoxic injury in type 2 diabetes. *Life Sci*, 2016; 154: 87-95
- 173) Shimizu I and Minamino T: Cellular senescence in cardiac diseases. *J Cardiol*, 2019; 74: 313-319
- 174) Palmer AK, Tchkonja T, LeBrasseur NK, Chini EN, Xu M and Kirkland JL: Cellular Senescence in Type 2 Diabetes: A Therapeutic Opportunity. *Diabetes*, 2015; 64: 2289-2298
- 175) Baker DJ, Childs BG, Durik M, Wijers ME, Sieben CJ, Zhong J, Saltness RA, Jeganathan KB, Verzosa GC, Pezeshki A, Khazaie K, Miller JD and van Deursen JM: Naturally occurring p16(Ink4a)-positive cells shorten healthy lifespan. *Nature*, 2016; 530: 184-189
- 176) Chang J, Wang Y, Shao L, Laberge RM, Demaria M, Campisi J, Janakiraman K, Sharpless NE, Ding S, Feng W, Luo Y, Wang X, Aykin-Burns N, Krager K, Ponnappan U, Hauer-Jensen M, Meng A and Zhou D: Clearance of senescent cells by ABT263 rejuvenates aged hematopoietic stem cells in mice. *Nat Med*, 2016; 22: 78-83
- 177) Dookun E, Walaszczyk A, Redgrave R, Palmowski P, Tual-Chalot S, Suwana A, Chapman J, Jirkovsky E, Donastorg Sosa L, Gill E, Yausep OE, Santin Y, Miale-Perez J, Andrew Owens W, Grieve D, Spyridopoulos I, Taggart M, Arthur HM, Passos JF and Richardson GD: Clearance of senescent cells during cardiac ischemia-reperfusion injury improves recovery. *Aging Cell*, 2020; 19: e13249
- 178) Dookun E, Passos JF, Arthur HM and Richardson GD: Therapeutic Potential of Senolytics in Cardiovascular Disease. *Cardiovasc Drugs Ther*, 2020;
- 179) Roos CM, Zhang B, Palmer AK, Ogrodnik MB, Pirtskhalava T, Thalji NM, Hagler M, Jurk D, Smith LA, Casaclang-Verzosa G, Zhu Y, Schafer MJ, Tchkonja T, Kirkland JL and Miller JD: Chronic senolytic treatment alleviates established vasomotor dysfunction in aged or atherosclerotic mice. *Aging Cell*, 2016; 15: 973-977
- 180) Walaszczyk A, Dookun E, Redgrave R, Tual-Chalot S, Victorelli S, Spyridopoulos I, Owens A, Arthur HM, Passos JF and Richardson GD: Pharmacological clearance of senescent cells improves survival and recovery in aged mice following acute myocardial infarction. *Aging Cell*, 2019; 18: e12945
- 181) Jia K, Dai Y, Liu A, Li X, Wu L, Lu L, Bao Y and Jin Q: Senolytic Agent Navitoclax Inhibits Angiotensin II-Induced Heart Failure in Mice. *J Cardiovasc Pharmacol*, 2020; 76: 452-460
- 182) Yu S, Kim SR, Jiang K, Ogrodnik M, Zhu XY, Ferguson CM, Tchkonja T, Lerman A, Kirkland JL and Lerman LO: Quercetin Reverses Cardiac Systolic Dysfunction in Mice Fed with a High-Fat Diet: Role of Angiogenesis. *Oxid Med Cell Longev*, 2021; 2021: 8875729
- 183) Justice JN, Nambiar AM, Tchkonja T, LeBrasseur NK, Pascual R, Hashmi SK, Prata L, Masternak MM, Kritchevsky SB, Musi N and Kirkland JL: Senolytics in idiopathic pulmonary fibrosis: Results from a first-in-human, open-label, pilot study. *EBioMedicine*, 2019; 40: 554-563
- 184) Hickson LJ, Langhi Prata LGP, Bobart SA, Evans TK, Giorgadze N, Hashmi SK, Herrmann SM, Jensen MD, Jia Q, Jordan KL, Kellogg TA, Khosla S, Koerber DM, Lagnado AB, Lawson DK, LeBrasseur NK, Lerman LO, McDonald KM, McKenzie TJ, Passos JF, Pignolo RJ, Pirtskhalava T, Saadiq IM, Schaefer KK, Textor SC, Victorelli SG, Volkman TL, Xue A, Wentworth MA, Wissler Gerdes EO, Zhu Y, Tchkonja T and Kirkland JL: Senolytics decrease senescent cells in humans: Preliminary report from a clinical trial of Dasatinib plus Quercetin in individuals with diabetic kidney disease. *EBioMedicine*, 2019; 47: 446-456
- 185) Suda M, Shimizu I, Katsuomi G, Yoshida Y, Matsumoto N, Hayashi Y, Ikegami R, Yoshida Y, Mikawa R, Katayama A, Wada J, Seki M, Suzuki Y, Iwama A, Nakagami H, Nagasawa A, Morishita R, Sugimoto M, Okuda S, Tsuchida M, Ozaki K, Matsui M and Minamino T: Senolytic vaccination improves normal and pathological age-related phenotypes and increases lifespan in progeroid mice. *Nature Aging*, 2021;