

## Draft Genome Sequence of *Streptomyces* sp. Strain PRh5, a Novel Endophytic Actinomycete Isolated from Dongxiang Wild Rice Root

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Here, we report the draft genome sequence of *Streptomyces* sp. strain PRh5 (China Center for Type Culture Collection [CCTCC] number 2013487), which is used to produce nigericin and nocardamine. The genome sequence will allow for the characterization of the molecular mechanisms underlying its beneficial properties.

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igericin is a polyether ionophore antibiotic derived from Streptomyces hygroscopicus. The structure was elucidated by X-ray crystallography in 1968 (1). Nigericin acts as an H<sup>+</sup>/K<sup>+</sup>/ Pb<sup>2+</sup> ionophore to chelate metal ions and transport them across cell membranes. In the past, nigericin was used as an antibiotic active against Gram-positive bacteria (2, 3). Recently, studies have shown that nigericin inhibits the Golgi functions in eukaryotic cells and exhibits anti-HIV activity (4, 5). Nocardamine, a hydroxamate siderophore, was initially isolated as an antibacterial metabolite of a Nocardia strain. Nocardamine exhibits antibacterial activity against mycobacterium species, especially tetracycline-resistant strains (6). Streptomyces sp. strain PRh5, which can produce nigericin and nocardamine, is a novel endophytic actinomycete that we isolated from Dongxiang wild rice root in China. Streptomyces sp. PRh5 was collected in the China Center for Type Culture Collection (CCTCC) with the collection number of CCTCC 2013487. Here, we report the first genome sequence of Streptomyces sp. PRh5, which we determined in an attempt to identify the nigericin and nocardamine biosynthetic gene clusters. The genome information may afford a basis for subsequent research directly connected with the natural product synthesis field.

The genome was sequenced using the Illumina Solexa HiSeq2000 instrument at the Beijing Genomics Institute (BGI) (Shenzhen, China). A library containing 500-bp inserts was constructed. Sequencing was performed with the paired-end strategy of (90, 90)-bp reads to produce 1.0 Gb of filtered sequences, representing 90-fold coverage of the genome. The sequences were assembled into 290 contigs using Velvet software (7).

Genome annotation was performed with the NCBI Prokaryotic Genome Annotation Pipeline 2.0. Open reading frames (ORF) were identified by Glimmer 3.02 (8) and Genemark (9). The resulting translations were used for a BLASTP (10) search against the GenBank NR database, as well as the KEGG (11) and COG (12) databases. tRNA and rRNA genes were identified by tRNAscan-SE (13) and RNAmmer (14), respectively. The PRh5 chromosome is about 11.1 Mbp in length, with an average G+C content of 71.1%. A total of 8,712 protein-coding genes were identified. The genome sequence represents a valuable shortcut for helping scientists find genes. Putative nigericin and nocardamine biosynthetic gene clusters are found in the *Streptomyces* sp. PRh5 genome. Gene expression analysis and bioassays are needed for further investigation into these genes. The genome sequence will accelerate the progress of research involving *Streptomyces* sp. PRh5.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number JABQ00000000. The version described in this paper is version JABQ01000000.

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## **REFERENCES**

- Steinrauf LK, Pinkerton M, Chamberlin JW. 1968. The structure of nigericin. Biochem. Biophys. Res. Commun. 33:29–31. http://dx.doi.org /10.1016/0006-291X(68)90249-0.
- Markin VS, Sokolov VS, Bogulavsky LI, Jaguzhinsky LS. 1975.
  Nigericin-induced charge transfer across membranes. J. Membr. Biol. 25: 23–45.
- Kovács-Hadady K, Kupás K. 1984. Rapid determination of monensin and nigericin in medical premixes and in growth-promoting feed preparations. Acta Vet. Hung. 32:97–101.
- Deng CC, Liang Y, Wu MS, Feng FT, Hu WR, Chen LZ, Feng QS, Bei JX, Zeng YX. 2013. Nigericin selectively targets cancer stem cells in nasopharyngeal carcinoma. Int. J. Biochem. Cell Biol. 45:1997–2006. http://dx.doi.org/10.1016/j.biocel.2013.06.023.
- 5. Sturz GR, Phan TH, Mummalaneni S, Ren Z, DeSimone JA, Lyall V. 2011. The K+–H+ exchanger, nigericin, modulates taste cell pH and

- chorda tympani taste nerve responses to acidic stimuli. Chem. Sens. **36**: 375–388. http://dx.doi.org/10.1093/chemse/bjq146.
- Ueki M, Suzuki R, Takamatsu S, Takagi H, Uramoto M, Ikeda H, Osada H. 2009. Nocardamine production by *Streptomyces avermitilis*. Actinomycetologica 23:34–39. http://dx.doi.org/10.3209/saj.SAJ230203.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res. 18:821–829. http://dx.doi .org/10.1101/gr.074492.107.
- 8. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679. http://dx.doi.org/10.1093/bioinformatics/btm009.
- Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. Nucleic Acids Res. 29:2607–2618. http://dx.doi.org/10.1093/nar/29.12.2607.
- 10. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local

- alignment search tool. J. Mol. Biol. 215:403–410. http://dx.doi.org/10.10 16/S0022-2836(05)80360-2.
- 11. Kanehisa M, Araki M, Goto S, Hattori M, Hirakawa M, Itoh M, Katayama T, Kawashima S, Okuda S, Tokimatsu T, Yamanishi Y. 2008. KEGG for linking genomes to life and the environment. Nucleic Acids Res. 36:D480–D484. http://dx.doi.org/10.1093/nar/gkm882.
- Tatusov RL, Galperin MY, Natale DA, Koonin EV. 2000. The COG database: a tool for genome-scale analysis of protein functions and evolution. Nucleic Acids Res. 28:33–36. http://dx.doi.org/10.1093/nar/28.1.33.
- 13. Lowe TM, Eddy SR. 1997. tRNAscan-SE: A program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25:955–964. http://dx.doi.org/10.1093/nar/25.5.955.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100-3108. http://dx.doi.org/10.1093/nar/gkm160.