



Article Protracted Impairment of Maternal Metabolic Health in Mouse Dams Following Pregnancy Exposure to a Mixture of Low Dose Endocrine-Disrupting Chemicals, a Pilot Study

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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Abstract: Pregnancy, a period of increased metabolic demands coordinated by fluctuating steroid hormones, is an understudied critical window of disease susceptibility for later-life maternal metabolic health. Epidemiological studies have identified associations between exposures to various endocrinedisrupting chemicals (EDCs) with an increased risk for metabolic syndrome, obesity, and diabetes. Whether such adverse outcomes would be heightened by concurrent exposures to multiple EDCs during pregnancy, consistent with the reality that human exposures are to EDC mixtures, was examined in the current pilot study. Mouse dams were orally exposed to relatively low doses of four EDCs: (atrazine (10 mg/kg), bisphenol-A (50 μ g/kg), perfluorooctanoic acid (0.1 mg/kg), 2,3,7,8-tetrachlorodibenzo-p-dioxin (0.036 µg/kg)), or the combination (MIX), from gestational day 7 until birth or for an equivalent 12 days in non-pregnant females. Glucose intolerance, serum lipids, weight, and visceral adiposity were assessed six months later. MIX-exposed dams exhibited hyperglycemia with a persistent elevation in blood glucose two hours after glucose administration in a glucose tolerance test, whereas no such effects were observed in MIX-exposed non-pregnant females. Correspondingly, MIX dams showed elevated serum low-density lipoprotein (LDL). There were no statistically significant differences in weight or visceral adipose; MIX dams showed an average visceral adipose volume to body volume ratio of 0.09, while the vehicle dams had an average ratio of 0.07. Collectively, these findings provide biological plausibility for the epidemiological associations observed between EDC exposures during pregnancy and subsequent maternal metabolic dyshomeostasis, and proof of concept data that highlight the importance of considering complex EDC mixtures based of off common health outcomes, e.g., for increased risk for later-life maternal metabolic effects following pregnancy.

Keywords: type 2 diabetes; maternal health; pregnancy; endocrine-disrupting chemicals

1. Introduction

Historically, toxicological research has studied maternal exposures to understand the health of the fetus but has ignored the long-term health of the mother [1], despite the fact that pregnancy represents a unique critical window for later-life maternal metabolic health due to the unique metabolic demands of supporting fetal development [2,3]. Pregnancy-specific changes in maternal metabolic physiology, i.e., lower fasting glucose, increased insulin secretion, elevated adipose volume and decreased insulin sensitivity, occur synchronously with predictable patterns of endocrine alterations [4,5]. Given that pregnant women are exposed concurrently to multiple endocrine-disrupting chemicals (EDCs) [6–9], pregnancy-specific metabolic demands raise concern that EDC exposures may induce

unique, protracted impairments of maternal metabolic health. Recent epidemiological studies have reported associations between EDC exposures, such as persistent organic pollutants, per and poly-fluoroalkyl substances, and bisphenols, during pregnancy and an increased risk of maternal hyperglycemia and gestational diabetes [10–20]. Following pregnancy, women undergo a metabolic reset to return to the physiological state experienced prior to pregnancy adaptations, the persistence of these pregnancy-specific changes in maternal metabolic physiology increases the risk of later-life maternal metabolic disease [21,22]. Furthermore, epidemiological studies have found per and poly-fluoroalkyl substances and phthalates exposure during pregnancy to be associated with increased postpartum maternal adiposity [23,24]. Pregnancy phthalate exposures have also been found in an epidemiological study to be associated with postpartum maternal elevated plasma glucose, insulin resistance, and adverse lipid changes, i.e., lower HDL and higher triglyceride levels [25]. Studies of non-pregnant women have also shown these same chemicals to be associated with obesity and type 2 diabetes [26–28]. Yet, little remains known about the protracted impact of EDC exposures during pregnancy on later-life maternal metabolic health, including type 2 diabetes risk. Understanding the influence of EDC exposures during pregnancy on later-life maternal metabolic health is critical, as worldwide, approximately 1 in 10 women are living with a form of diabetes [29], a quantity expected to double in the next decade [30].

Complicating the ability to predict metabolic disease risk, pregnant women are exposed to EDC mixtures, not to single EDCs [9,25,31–34]. If combinatorial EDCs confer an enhanced effect on metabolic health, risk assessment based on no adverse effect levels for single EDC exposures could overestimate "safe" exposure levels. Multiple studies focused on reproductive toxicity support this possibility, as the reproductive toxicity of EDC mixtures indicates dose-additive effects of low dose mixtures [35–38], even when single EDC exposures show no phenotype [39–41]. This study hypothesizes that well-studied EDCs known to target metabolic function through different direct molecular targets will produce enhanced metabolic toxicity on a single physiological function, such as glucose metabolism [35,42]. Glucose homeostasis requires the proper functioning of multiple interacting physiological systems [43,44]. Our multiple hits hypothesis [42] predicts that low-dose EDC hits at multiple molecular target sites may overwhelm compensatory mechanisms that normally maintain metabolic homeostasis [28,45], thereby increasing metabolic disease risk. Given the increased metabolic demands of pregnancy and the rapid metabolic changes that occur postpartum, it is essential to study EDC mixtures in pregnant and non-pregnant animals to determine if pregnancy alters sensitivity to EDC exposures with later-life metabolic consequences. This approach is critical to ensure that no adverse effect levels set for single EDC exposures are not underestimated.

Single chemical studies have identified approximately a dozen EDCs that have strong and substantial evidence of metabolic disruption [27,28]. Four well-studied EDC classes herbicides, plasticizers, perfluorinated compounds, and PCB-like dioxin compounds—target glucose tolerance, lipid metabolism, and weight gain, although typically these studies have focused on male rodents [26–28,46,47]. Atrazine, a commonly used herbicide, modifies insulin resistance [48], and alters free fatty acids and hepatic lipids [49]. Bisphenol-A (BPA), a plasticizer, leads to hyperglycemia [27,50–56], increases body and adipose weight [57,58], and alters adipocyte hypertrophy [59] and differentiation [60]. Perfluorooctanoic acid, a perfluorinated compound [61,62], induces hypercholesterolemia, and increases liver weight, bodyweight, adipokines, and lipids [63–66]. 2,3,7,8-tetrachlorodibenzo-p-dioxin [67,68], a PCB-like dioxin compound, has been shown to lower insulin secretion [69–71] and decrease serum lipids [72]. With the exception of BPA [51,52], most animal models have not determined the potential protracted effects of EDC exposures, either singly or in mixtures, on pregnant females on later-life maternal metabolic health.

To address these gaps, the current pilot study assessed the effects of pregnancy exposures to low-dose EDCs either singly or in a mixture on later-life maternal metabolic health. Pregnant mice (hereafter referred to as dams) were exposed to low doses of four EDCs singly shown to have high to moderate evidence for metabolic disruption (atrazine, bisphenol-A, perfluorooctanoic acid, and 2,3,7,8-tetrachlorodibenzo-p-dioxin) or to a mixture (MIX) of these four from gestational day 7 until birth [27]. Additionally, non-pregnant female mice were exposed to MIX for an equivalent 12 days to determine the role of pregnancy. Approximately 6 months after the cessation of exposure, dams and non-pregnant females underwent a glucose tolerance test, and serum lipids, visceral adiposity, and serum adipokines were measured.

2. Materials and Methods

2.1. Animals and Husbandry

To determine whether EDC exposures occurring singly or in a mixture alter later-life maternal metabolic disease risk, adult (postnatal day (PND) 60) male and female C57BL/6J mice obtained from Jackson Laboratories (Bar Harbor, ME, USA) were bred using a monogamous pairing scheme. Male and female mice were paired, and the following morning, female mice were checked for the presence of a mucus plug, which was denoted as gestational day (GD) 0. If a mucus plug was observed, males were removed from the home cage. Pregnant dams remained singly housed throughout weaning. Pregnant dams were exposed orally via EDC-spiked mealworm to either a single EDC at a low dose or the combination of all four EDCs at the same doses, as given in the single exposures: atrazine (ATR-10 mg/kg), bisphenol-A (BPA—50 μg/kg), perfluorooctanoic acid (PFOA—0.1 mg/kg), 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD $-0.036 \,\mu$ g/kg), or their mixture (MIX), from GD 7 until birth (n = 4/group for the weekly weights, glucose tolerance test, serum lipids, serum adipokines, and serum corticosterone, and n = 7/group for the visceral adipose visualization performed sequentially). Single EDC doses were considered relatively low because they were either: at or below levels typically shown to produce effects in animal studies (ATR [73–76], PFOA [65,77,78], and TCDD [69,70,72,79]), at current oral human reference dose levels (BPA) (https://www.epa.gov/iris, accessed on 1 September 2021), or at or below the EPA's NOAEL for animal studies (PFOA) [80] (Table 1. EDC Dosing Method). ATR (>98% pure) and BPA (99% pure) (Sigma-Aldrich, St. Louis, MO, USA) were each separately dissolved in peanut oil, PFOA (96% pure, Sigma-Aldrich, St. Louis, MO, USA) was dissolved in water, and TCDD (98% pure, Cambridge Isotopes, Cambridge, MA, USA) was dissolved in anisole and diluted in peanut oil. Calculated doses were injected into mealworm(s). MIX-mealworms were created by injecting BPA and PFOA and ATR and TCDD into two worms, respectively. Upon injection, the mealworms were immediately flash-frozen to prevent degradation and leakage. Dams were dosed via EDC-spiked mealworm(s) at the same time every day from GD 7 until birth, with vehicle dams given vehicle oil-spiked mealworms [81,82]. This dosing method was selected to eliminate bolus increases in blood sugar from sugar-based treats and to eliminate the influence of stress from gavage administration. Furthermore, we dosed daily to ensure each dam had an equivalent behavioral experience across pregnancy. The limitation of daily dosing is that each EDC has a different half-life influencing bioaccumulation. Pregnancies were monitored daily to evaluate differences in time to parturition, litter size, or sex ratio of the offspring. On average, litters contained 7.19 ± 1.55 pups. Upon weaning at PND 25, dams remained individually housed in a vivarium room maintained at 22 \pm 2 $^\circ$ C with a 12-h light–dark cycle (lights on at 06:00 h) for 18 weeks with weight measured weekly until metabolic reprogramming was assessed. Labdiet Rodent 5001 diet was provided ad libitum throughout the entire experimental duration.

EDC	Dose	Solvent	Dose Reference
Atrazine (ATR)	10 mg/kg	Peanut Oil	Below levels typically shown to cause effects in animal studies [73–76]
Bisphenol-A (BPA)	50 μg/kg	Peanut Oil	Current oral human daily reference dose (https://www.epa.gov/iris; accessed on 1 September 2021)
Perfluorooctanoic acid (PFOA)	0.10 mg/kg	Distilled water	At or below NOAEL referenced in animal studies in the EPA 2016 document 'Health Effects Support Document for Perfluorooctanoic Acid (PFOA)' [80] Below levels typically shown to cause effects in animal studies [65,77]
2,3,7,8-tetrachlorodibenzo-p- dioxin (TCDD)	0.036 μg/kg	Anisole diluted in peanut oil	Below levels typically shown to cause effects in animal studies [69,70,72,79]
Mixture (MIX)	All the above doses added together	All the above	All the above

 Table 1. EDC Dosing Method.

To determine whether any effects of the mixture of EDCs six months postpartum were uniquely produced by pregnancy, sequentially a separate cohort of non-pregnant female mice underwent acute MIX exposure under the same experimental conditions and concurrently with the MIX dams for visceral adipose visualization. Adult (PND60) female C57BL/6J mice were obtained from Jackson Laboratories (Bar Harbor, ME). The female mice were singly housed beginning on day one. Oral exposure to either MIX or a vehicle oil-spiked mealworm started on day seven (n = 5/group). Exposure occurred for approximately twelve days to mimic the average length of exposure pregnant dams received. Following the cessation of exposure, non-pregnant female mice remained individually housed in a vivarium room maintained at 22 ± 2 °C with a 12-h light–dark cycle (lights on at 06:00 h) for 21 weeks with bodyweight measured weekly, starting three weeks after the cessation of exposure to duplicate the weekly bodyweight measurements in pregnant exposed dams until metabolic reprogramming was assessed. Labdiet Rodent 5001 diet was provided ad libitum throughout the entire experimental duration.

All experiments were carried out according to NIH Guidelines and were approved by the University of Rochester Medical School University Committee on Animal Resources (UCAR-2018-025-102139, 19 December 2018), which included a hazard plan with hazard notification signage to ensure proper handling of cages and bedding.

2.2. Glucose Tolerance Test

Six months after the cessation of exposure, an intraperitoneal glucose tolerance test was performed independent of estrous cyclicity [83]. Mice were transferred to cages with food removed while retaining water at 02:00 h. Fasting occurred for 6 h prior to testing at 08:00 h. Weights were obtained at the end of the fasting duration. A tail snip was performed once fasting commenced providing blood for glucose measurements. Baseline glucose was measured immediately following the tail snip utilizing an AlphaTRAK Blood Glucose Monitoring System (Zoetis, Parsippany, NJ, USA) on the setting for dogs. Each mouse was then intraperitoneally injected with a freshly prepared solution of glucose (Sigma-Aldrich, St. Louis, MO, USA) at 2 g/kg, after which blood glucose measurements

were obtained using the AlphaTRAK glucometer at 15, 30, 60, and 120 min. After the final glucose measurement, mice were immediately sacrificed by live decapitation with trunk blood collected.

2.3. Serum Lipids

In pregnant exposed dams, triglycerides were determined in duplicate utilizing fresh serum obtained following the glucose tolerance test using a colorimetric assay kit (Abcam, Cambridge, UK) carried out according to the manufacturer's instructions with a dilution of 1:10. High-density lipoprotein (HDL) and low-density lipoprotein (LDL) were measured with a colorimetric assay kit (Abcam, Cambridge, UK) in duplicate according to the manufacturer's directions, with the HDL run at a dilution of 1:50 and LDL run without dilution. Assays were run on a SynergyH1 Hybrid Reader (BioTek, Winooski, VT, USA) with Gen5 2.01 software. Sample replicates with coefficients of variation (CVs) greater than 15% were excluded from the analysis.

2.4. Visceral Adipose Visualization

Visualization of visceral adipose in pregnant exposed dams was performed six months postpartum on a Scanco Medical VivaCT 40 cone-beam-computed tomography (CT) (Scanco Medical, Brüttisellen, Switzerland). Dams were anesthetized utilizing continuous isoflurane gas during the scan, with one dam from each group initially euthanized by an intraperitoneal injection of Euthasol (Virbac, Fort Worth, TX, USA) to validate and develop the CT parameters. Throughout the scan, respiration rate and movement were monitored to ensure proper isoflurane administration. The scans were run on the following parameters: tube diameter 35.8 mm OD, resolution 35.0-micron isotropic voxels, 1048 samples, 500 projections over 180°, 200 ms integration time, and an energy of 45 kVp with the intensity/current at 177 micro-amps. Analysis was conducted with Scanco evaluation software V6.5 (Scanco Medical, Brüttisellen, Switzerland). Gaussian filters were utilized to reduce beam hardening, Gauss $\sum 0.8$ and Gauss support 1 pixel. In all images, the scaling maximum was set to 10,000 to provide a clearer contrast between the visceral adipose and other soft tissue. The region of interest extended from below the lungs to the proximal end of the tail. Manual contours were drawn to exclude the visible layers of subcutaneous adipose and the femurs, which were tightly pressed against the abdomen, as well as to include the visceral adipose. Renditions were generated on Scanco's Image Processing Language (IPL) and 3-dimensional viewer via a transparent concatenation to merge adipose and bone with the adipose displayed slightly transparent and colored with the ventral side as the front. Volumetric quantifications of the adipose were calculated through the utilization of Scanco's "per mille" value of 40 (μ linear attenuation coefficient = 0.32 cm⁻¹) to remove potential air and an upper threshold of 75 ($\mu = 0.600 \text{ cm}^{-1}$) to remove bone and soft tissue.

2.5. Serum Adipokines

Adipokines were measured in the pregnant exposed dams' serum obtained after the glucose tolerance test. Adiponectin was assessed in duplicate utilizing a Mouse Adiponectin ELISA kit (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's directions and analyzed on a SynergyH1 Hybrid Reader (BioTek, Winooski, VT, USA) with Gen5 2.01 software. Insulin, monocyte chemoattractant protein 1 (MCP-1), plasminogen activator inhibitor-1 (PAI-1 Total), and resistin were measured on a Mouse Adipokine Magnetic Bead Panel (Sigma-Aldrich, St. Louis, MO, USA) according to directions provided by the manufacturer and run on a Bio-Plex 200 (Bio-Rad, Hercules, CA, USA). Sample replicates with CVs higher than 15% were excluded from the analysis.

2.6. Serum Corticosterone

Serum corticosterone was measured in duplicate using commercially available enzyme immunoassay kits (Arbor Assays, Ann Arbor, MI, USA), according to the manufacturer's specifications and run on a SynergyH1 Hybrid Reader (BioTek, Winooski, VT, USA) with Gen5 2.01 software. Sample replicates with CVs higher than 15% were excluded from the analysis. Standard curve CVs fell below 10%.

2.7. Statistical Analyses

Shapiro-Wilk's test and the Bartlett test were used to assess the normality and homogeneity of variances, respectively. For the glucose tolerance test of pregnant dams, repeated-measure ANOVAs were conducted with treatment groups (ATR, BPA, PFOA, TCDD, MIX, and vehicle) with glucose area under the curve and glucose concentrations at the final two-hour draw compared using one-way ANOVA, with subsequent post-hoc tests (Student's *t*-test). The glucose tolerance test of non-pregnant female mice was analyzed separately utilizing a repeated-measures ANOVA, and the glucose area under the curve and final glucose concentration were analyzed with a Student's *t*-test. Serum lipids, adipokines, and corticosterone were analyzed with one-way ANOVA across all treatments (single EDCs, MIX, and control) with subsequent post-hoc tests (Student's *t*-test). Visceral adipose was only measured in the control and MIX exposed dams; as such, only Student's one-way *t*-tests were conducted. The effect of treatment groups on weekly weight gain was assessed utilizing a repeated-measures ANOVA. Outliers were removed following a statistically significant Grubb's test (Graphpad Software Inc., San Diego, CA, USA). Out of 1070 data points, 9 outliers were removed. The outliers removed are as follows: serum lipids, 5 data points out of 96; serum adipokines, 4 data points out of 120. Triglycerides had 4 data points out of 24 removed due to technical error. No outliers were removed from the weekly weights, the glucose tolerance test, or the visceral adipose visualization. Statistical analyses were conducted using JMP Pro 14.0 (SAS Institute Inc., Cary, NC, USA). *p*-values ≤ 0.05 were considered statistically significant.

3. Results

3.1. Glucose Tolerance Test

No significant effects of treatment on bodyweight were observed in pregnant exposed dams (F(5,16) = 0.22, p = 0.95, Figure 1A). Fasting blood glucose levels did not significantly differ between any treatment groups (F(5,18) = 1.0, p = 0.41, Figure 1B). Over the course of the glucose tolerance test, blood glucose concentrations exhibited a marginally significant interaction between time and treatment (F(20,72) = 1.5, p = 0.09), with MIX dams showing a significantly elevated glucose area under the curve compared to vehicle dams (p = 0.033, Cohen's d = 1.63, Figure 1C). There were no significant differences in area under the curve in any other treatment group. Additionally, at the completion of the glucose tolerance test, 120 min following the glucose injection, only MIX dams exhibited significant hyperglycemia compared to vehicle dams (p = 0.001, Cohen's d = 2.75, Figure 1D). Final blood glucose concentrations did not significantly differ in any other treatment group.



Figure 1. Weekly Weights and Glucose Tolerance Tests. (**A**) Dams were weighed weekly following weaning until the glucose tolerance test with no change in weight between groups at any week. Twenty-one weeks following the cessation of EDC exposures a glucose tolerance test was performed. (**B**) A marginally significant interaction was present when performing a repeated-measures ANOVA on the glucose concentrations over time in MIX dams with a significant effect of MIX at 120 min. (**C**) Only in the MIX dams was there a significantly elevated glucose area under the curve. (**D**) MIX dams also had significantly elevated glucose at the completion of the test; full data points displayed. (**E**) Non-pregnant female mice were weighed weekly three weeks after the cessation of exposure to mirror dams with no difference in weight at any week. (**F**) Non-pregnant MIX dams did not have a difference in blood glucose at any time point of the glucose tolerance test. (**G**) The non-pregnant MIX female mice did not have any difference in glucose area under the curve. (**H**) Non-pregnant MIX female mice did not have any difference in glucose area under the curve. (**H**) Non-pregnant MIX female mice did not differ in blood glucose at 120 min after the glucose injection. Data are reported as mean \pm standard error (*n* = 4–5/group). Asterisk indicates *p* ≤ 0.05.

Non-pregnant MIX female mice, likewise, exhibited no bodyweight effects of treatment difference compared to non-pregnant vehicle female mice (F(1,8) = 0.060, p = 0.81, Figure 1E). As with the MIX dams, fasting blood glucose levels were not affected in the non-pregnant MIX female mice (t = -0.48, p = 0.64). Additionally, no difference was found in blood glucose levels in non-pregnant MIX female mice compared to non-pregnant vehicle female mice at any time point following the injection of glucose during the glucose tolerance test (Figure 1F). Nor was a significant interaction between time and treatment present (F(4,5) = 0.43, p = 0.71). Unlike the MIX dams, the non-pregnant MIX female mice did not have a significantly elevated glucose area under the curve (t = -0.62, p = 0.55, Figure 1G), nor did they show significantly elevated glucose at 120 min following the glucose injection (t = -1.44, p = 0.19, Figure 1H).

3.2. Serum Lipids

Compared to vehicle dams, MIX dams showed significant elevations in LDL (p = 0.037, Cohen's d = 1.73, Figure 2B). There were no significant differences in LDL in any other treatment group. No differences in total cholesterol (F(5,17) = 1.02, p = 0.44), the HDL (F(5,15) = 0.98, p = 0.46) or the triglycerides (F(5,14) = 0.78, p = 0.58) were observed in any treatment group (Figure 2A,C,D).



Figure 2. Pregnant Dams Serum Lipids. Following a glucose tolerance test, a cholesterol panel was performed on the serum. LDL (**B**) was significantly elevated only in the MIX dams. Total cholesterol (**A**), HDL (**C**), and triglycerides (**D**) did not differ in any treatment group. Data are presented as mean \pm standard error (n = 3-4/group). Asterisk indicates $p \le 0.05$.

3.3. Visceral Adipose Visualization

No difference was observed in body volume, with a mean body volume of 7055 mm³ in vehicle dams and 7546 mm³ in MIX dams (t = -1.36, p = 0.10). MIX dams had a mean adipose volume of 673 mm³ in MIX dams compared to 511 mm³ in vehicle dams, though this failed to reach statistical significance (t = -1.57, p = 0.07, Cohen's d = 0.83; data not shown). Furthermore, MIX dams had an average visceral adipose volume to body volume ratio of 0.09 while the vehicle dams had an average ratio of 0.07; however, this did not reach statistical significance (t = -1.49, p = 0.08, Cohen's d = 0.79, Figure 3C).



Figure 3. Pregnant Dams Visceral Adipose Visualization. Six months following the cessation of exposure, MIX and vehicle dams underwent a CT scan for visceral adipose assessment. (A) A representative image of the vehicle dams. (B) A representative image of the MIX dams. (C) MIX dams had a non-significant ratio of visceral adipose volume to body volume compared to vehicle dams. Data are shown as mean \pm standard error (n = 7/group, p = 0.08).

3.4. Serum Adipokines

Adiponectin was significantly decreased by both PFOA (p = 0.017, Cohen's d = 2.52) and TCDD (p = 0.023, Cohen's d = 2.67) administered singly (Figure 4A). There were no significant changes in adiponectin in the other single EDC groups or in MIX dams. No differences in insulin (F(5,18) = 0.56, p = 0.73), MCP-1 (F(5,16) = 0.62, p = 0.69), PAI-1 Total (F(5,18) = 0.42, p = 0.83), or resistin (F(5,18) = 1.11, p = 0.39) were observed in any treatment group (Figure 4B–E).



Figure 4. Pregnant Dams Serum Adipokines and Corticosterone. Various adipokines were analyzed following the glucose tolerance test: adiponectin (**A**), insulin (**B**), MCP-1 (**C**), PAI-1 total (**D**), and resistin (**E**). Adiponectin was significantly decreased in the single EDCs, PFOA and TCDD (**A**). No difference was observed between the remaining serum adipokines in any other treatment group. No difference was observed in corticosterone following the glucose tolerance test either (**F**). Data are shown as mean \pm standard error (n = 3-4/group). Asterisk indicates $p \le 0.05$.

3.5. Serum Corticosterone

Corticosterone levels at the completion of the glucose tolerance test did not differ between any treatment group (F(5,18) = 1.2, p = 0.34) (Figure 4F).

4. Discussion

Pregnancy is an understudied window of vulnerability for later-life maternal health [1,84], as the metabolic demands necessary for supporting fetal development may enhance maternal metabolic disease risk [2,3,85,86]. Recent epidemiological studies have shown an increased prevalence of life-long maternal metabolic alterations following gestational EDC exposures [23–25], yet few rodent models have explored the role of pregnancy per se by directly comparing non-pregnant and pregnant female mice, nor have they examined exposure to EDC mixtures, a phenomenon consistent with the human environment. The present pilot study examined the potential for protracted maternal metabolic toxicity following gestational EDC exposures, singly and in the mixture. In this pilot study, MIX exposure, but not single EDCs, produced hyperglycemia in MIX dams but not in non-pregnant MIX female mice. MIX-exposed dams showed significantly elevated blood glucose two hours after glucose administration. Furthermore, in preliminary analyses, MIX dams had elevated serum LDL, suggesting dyslipidemia. Despite higher means, there were no statistically significant increases in weight or visceral adipose. This study was a proof-of-concept experiment focused on a disease-centered approach to mixture studies aimed at investigating how EDCs with diverse molecular targets integrate across multiple signaling pathways in a complex system to evaluate joint effects on later-life maternal metabolic health [87-89]. More work is needed to replicate this study with a larger sample size and to investigate how additional EDC mixtures with diverse molecular modes of action may converge downstream to alter maternal health during gestation and across the lifespan.

Six months following the cessation of exposure, only MIX dams exhibited hyperglycemia during a glucose tolerance test, a phenotype often observed in metabolic syndrome and type 2 diabetes [90]. Furthermore, MIX dams had nearly double the blood glucose levels of vehicle dams at the 120 min time point of the glucose tolerance test. The stark differences in elevated total blood glucose (increased area under the curve values) and final blood glucose concentrations occurred without selection of a genetically predisposed strain and without a high-fat diet [44,48,91–93]. Interestingly, MIX dams did not show significant elevations in bodyweight. Although prior studies have shown that single EDC exposures are associated with hyperglycemia [50-54], such effects were not seen here in response to single EDC exposures. We hypothesize that variations in exposure concentration, exposure window, route of exposure, the time point of testing, and sex differences influence the effect of single EDC exposures on hyperglycemia. Furthermore, dependent on effect sizes, we may have been limited in identifying the effects of single EDCs. In an elegant series of studies, 10 and 100 μ g/kg of BPA exposure via subcutaneous injection during pregnancy were shown to produce hyperglycemia as soon as five and four months postpartum, respectively, in OF-1 dams [51]. In our study, dams were exposed to 50 ug/kg BPA via oral exposure, which may alter toxicokinetic distribution. Alternatively, there may be mouse-strain-specific effects. Interestingly, single EDC exposures did produce multiple biochemical changes. For example, PFOA and TCDD alone resulted in reductions in adiponectin, indicating that these exposure concentrations were not "no adverse effect levels", although these concentrations did not result in an overall increase in hyperglycemia at the completion of the glucose tolerance test. Importantly, MIX effects on blood glucose concentrations were unique to pregnant dams, as females exposed for an equivalent number of days showed no significant elevations in blood glucose six months later.

In addition to hyperglycemia, only MIX dams showed elevated serum LDL. Dyslipidemia is another phenotype commonly observed in individuals with metabolic dysfunction, such as insulin resistance and type 2 diabetes, and has been utilized as a predictor for these diseases [43,94–98]. Previous rodent studies have found that a bolus dose of $30 \mu g/kg$ of TCDD decreased total cholesterol, LDL, and HDL one week post-exposure [99], and that approximately 0.5 mg/kg of PFOA along with a high-fat diet exposure resulted in hypercholesterolemia [65], and approximately 0.26 mg/kg of BPA exposure can also increase total cholesterol [58]. Though the HDL of MIX dams was unchanged, it is unknown how HDL responded at baseline since these data were collected following the glucose tolerance test, and this may reflect acute lipid alterations [100]. Future research is needed to assess lipid profiles in the absence of prior glucose tolerance testing, particularly for HDL and hepatic lipid profiles, to determine the protracted effects of maternal lipid profiles following gestational exposure to EDC mixtures.

There were no significant increases in body volume or visceral adipose volume. MIX dams had an average visceral adipose volume to body volume ratio of 0.09, while vehicle dams had an average bodyweight ratio of 0.07 (p = 0.08). Increased visceral adipose is indicative of metabolic syndrome and an increased risk for type 2 diabetes [90]. Adipocytes have been observed to be impacted by single EDC exposures, with BPA found to increase adipogenesis [101] and maternal adipose weight months postpartum following gestational exposure [51]. TCDD was found to upregulate genes in inflammatory pathways in adipose tissue both in vitro and in vivo [102], and numerous other EDCs have been observed to alter the differentiation of adipocytes and expression of adipokines [24,101–103]. Furthermore, in epidemiological studies, exposures during pregnancy to phthalates and per and polyfluoroalkyl substances, such as PFOA, have been associated with elevated long-term maternal weight gain, despite weight gain in pregnancy [23,24]. While such findings are notable, it is possible that a much greater risk is conferred given that women are exposed to multiple EDCs during pregnancy, and the effect of such mixtures on long-term maternal adiposity remains to be investigated [6-9,31]. Future research should investigate the impact of multiple relevant risk factors, including EDC mixtures, as well as how high-fat diets alter later-life maternal visceral adipose and weight gain in dams exposed during gestation to EDC mixtures and the persistence of such effects.

MIX dams did not show altered serum adipokines: adiponectin [104], insulin [90], monocyte chemoattractant protein-1 (MCP-1) [105], plasminogen activator inhibitor-1 total (PAI-1 Total) [106], or resistin [107], in response to exposures. Fasting and glucose tolerance tests have been shown to alter adipokine levels, which might have led to the lack of adipokine effects seen post-glucose-tolerance test in MIX dams [108,109]. Consequently, future studies are needed to assess adipokine levels at baseline, not after fasting and subsequent glucose-tolerance testing. Though PFOA and TCDD alone resulted in reductions in adiponectin, we did not see a synergistic effect in the MIX. The physiological response from the single EDC exposures might have differed from the MIX exposure, as the MIXexposed dams might be responding to, or perhaps compensating for, effects from the other EDC exposures, e.g., effects simultaneously of TCDD and PFOA that resulted in a unique metabolic shift or triggered compensation. More work on the combinatorial effects of EDCs is needed to assess these changes. Additionally, circulating steroid hormones, for example, high levels of corticosterone can significantly influence metabolism and blood glucose concentrations by inhibiting energy storage and acting in opposition to insulin [110–112]. As such, serum corticosterone levels were assessed following glucose-tolerance testing, but these were not altered by MIX exposure.

It is worth noting, the limitations of the current study include statistical considerations, such as the need for larger sample sizes for identification of effects of single low-dose EDC exposures. If effect sizes of single EDC exposures are small, larger sample sizes are required to confirm effects on these metabolic endpoints. Research comparing low-dose EDC studies should consider the directionality of effects, beta coefficients, and confidence intervals to determine the continuity across replicated studies, even if effects are not statistically significant [113]. Secondly, ANOVA analyses of multiple groups utilize the variance estimates from all groups when determining significance. Therefore, single EDC studies, conducted with just a control and exposed group, would have different variance estimates that may yield different conclusions with respect to statistical significance. Additionally,

the single EDC doses utilized were not chosen based on previous effects in pregnant dams. Selecting single EDC doses based on a dose–response curve would have provided more accurate low-doses for the study. Future mixture experiments should utilize dose–response curves, as the doses selected might produce different effects on later-life maternal health. Finally, we targeted the gestational exposure window only. This experiment tests the sensitivity of this window to later-life maternal metabolic reprogramming. However, this is not the reality of human exposures, as women are exposed pre-pregnancy, during pregnancy, and post-pregnancy. Each of these exposure windows may yield differential effects on later-life maternal metabolic health.

Research on maternal metabolic health following gestational EDC exposures should utilize real-world mixtures, along with integrated metabolic assessments, to determine the influence of complex EDC mixtures on women's health. The current findings indicate that enhanced effects of MIX exposure may result from multiple small hits at different molecular targets that together converge to disrupt the homeostasis of integrated metabolic endpoints, such as serum hyperglycemia and dyslipidemia [87]. Mechanistic studies focused on mixtures should integrate multiple aspects of metabolic physiology, including assessments of the pancreas, liver, and serum lipids, at baseline to understand how such convergence occurs. Future research is needed to advance public health protection of women by assessing the influence of EDC mixtures on later-life maternal metabolic health to ensure "safe" exposure levels are not overestimated.

5. Conclusions

In conclusion, exposure during pregnancy to a curated mixture of EDCs, known to target metabolism but not exposures to single EDCs, altered maternal glucose tolerance, consistent with protracted toxicity and potential reprogramming. Moreover, this protracted alteration of blood glucose levels following MIX was unique to pregnancy as non-pregnant female mice showed no effects. These data indicate pregnancy is a unique window of EDC exposure and support the possibility that mixtures of low-dose EDCs may influence later-life maternal metabolic disease risk. More work is needed with larger sample sizes and more research is needed focused on grouping EDCs for mixture risk assessment based on common adverse health outcomes.

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References

- 1. Catanese, M.C.; Suvorov, A.; Vandenberg, L.N. Beyond a means of exposure: A new view of the mother in toxicology research. *Toxicol. Res.* **2015**, *4*, 592–612. [CrossRef]
- 2. Rahim, M.N.; Williamson, C.; Kametas, N.A.; Heneghan, M.A. Using pregnancy to assess risk and predict women's health. *EClinicalMedicine* **2020**, 20. [CrossRef]
- 3. Bellamy, L.; Casas, J.-P.; Hingorani, A.D.; Williams, D. Type 2 diabetes mellitus after gestational diabetes: A systematic review and meta-analysis. *Lancet* 2009, 373, 1773–1779. [CrossRef]
- 4. Lain, K.Y.; Catalano, P.M. Metabolic changes in pregnancy. Clin. Obstet. Gynecol. 2007, 50, 938–948. [CrossRef]
- 5. Butler, A.; Cao-Minh, L.; Galasso, R.; Rizza, R.; Corradin, A.; Cobelli, C.; Butler, P. Adaptive changes in pancreatic beta cell fractional area and beta cell turnover in human pregnancy. *Diabetologia* **2010**, *53*, 2167–2176. [CrossRef] [PubMed]
- Lee, W.-C.; Fisher, M.; Davis, K.; Arbuckle, T.E.; Sinha, S.K. Identification of chemical mixtures to which Canadian pregnant women are exposed: The MIREC Study. *Environ. Int.* 2017, 99, 321–330. [CrossRef]
- Morello-Frosch, R.; Cushing, L.J.; Jesdale, B.M.; Schwartz, J.M.; Guo, W.; Guo, T.; Wang, M.; Harwani, S.; Petropoulou, S.-S.E.; Duong, W. Environmental chemicals in an urban population of pregnant women and their newborns from San Francisco. *Environ. Sci. Technol.* 2016, 50, 12464–12472. [CrossRef]
- Robinson, O.; Basagaña, X.; Agier, L.; De Castro, M.; Hernandez-Ferrer, C.; Gonzalez, J.R.; Grimalt, J.O.; Nieuwenhuijsen, M.; Sunyer, J.; Slama, R.M. The pregnancy exposome: Multiple environmental exposures in the INMA-Sabadell birth cohort. *Environ. Sci. Technol.* 2015, 49, 10632–10641. [CrossRef] [PubMed]
- 9. Woodruff, T.J.; Zota, A.R.; Schwartz, J.M. Environmental chemicals in pregnant women in the United States: NHANES 2003–2004. *Environ. Health Perspect.* **2011**, *119*, 878–885. [CrossRef] [PubMed]
- Chiu, Y.-H.; Mínguez-Alarcón, L.; Ford, J.B.; Keller, M.; Seely, E.W.; Messerlian, C.; Petrozza, J.; Williams, P.L.; Ye, X.; Calafat, A.M. Trimester-specific urinary bisphenol A concentrations and blood glucose levels among pregnant women from a fertility clinic. *J. Clin. Endocrinol. Metab.* 2017, 102, 1350–1357. [CrossRef]
- Shaffer, R.M.; Ferguson, K.K.; Sheppard, L.; James-Todd, T.; Butts, S.; Chandrasekaran, S.; Swan, S.H.; Barrett, E.S.; Nguyen, R.; Bush, N. Maternal urinary phthalate metabolites in relation to gestational diabetes and glucose intolerance during pregnancy. *Environ. Int.* 2019, 123, 588–596. [CrossRef]
- Vafeiadi, M.; Roumeliotaki, T.; Chalkiadaki, G.; Rantakokko, P.; Kiviranta, H.; Fthenou, E.; Kyrtopoulos, S.A.; Kogevinas, M.; Chatzi, L. Persistent organic pollutants in early pregnancy and risk of gestational diabetes mellitus. *Environ. Int.* 2017, *98*, 89–95. [CrossRef] [PubMed]
- James-Todd, T.M.; Chiu, Y.-H.; Messerlian, C.; Mínguez-Alarcón, L.; Ford, J.B.; Keller, M.; Petrozza, J.; Williams, P.L.; Ye, X.; Calafat, A.M. Trimester-specific phthalate concentrations and glucose levels among women from a fertility clinic. *Environ. Health* 2018, 17, 55. [CrossRef] [PubMed]
- Bellavia, A.; Chiu, Y.-H.; Brown, F.M.; Mínguez-Alarcón, L.; Ford, J.B.; Keller, M.; Petrozza, J.; Williams, P.L.; Ye, X.; Calafat, A.M. Urinary concentrations of parabens mixture and pregnancy glucose levels among women from a fertility clinic. *Environ. Res.* 2019, 168, 389–396. [CrossRef]
- 15. Zhang, W.; Xia, W.; Liu, W.; Li, X.; Hu, J.; Zhang, B.; Xu, S.; Zhou, Y.; Li, J.; Cai, Z. Exposure to bisphenol A substitutes and gestational diabetes mellitus: A prospective cohort study in China. *Front. Endocrinol.* **2019**, *10*, 262. [CrossRef] [PubMed]
- 16. James-Todd, T.M.; Meeker, J.D.; Huang, T.; Hauser, R.; Ferguson, K.K.; Rich-Edwards, J.W.; McElrath, T.F.; Seely, E.W. Pregnancy urinary phthalate metabolite concentrations and gestational diabetes risk factors. *Environ. Int.* **2016**, *96*, 118–126. [CrossRef]
- Rahman, M.L.; Zhang, C.; Smarr, M.M.; Lee, S.; Honda, M.; Kannan, K.; Tekola-Ayele, F.; Louis, G.M.B. Persistent organic pollutants and gestational diabetes: A multi-center prospective cohort study of healthy US women. *Environ. Int.* 2019, 124, 249–258. [CrossRef] [PubMed]
- Saldana, T.M.; Basso, O.; Hoppin, J.A.; Baird, D.D.; Knott, C.; Blair, A.; Alavanja, M.C.; Sandler, D.P. Pesticide exposure and self-reported gestational diabetes mellitus in the Agricultural Health Study. *Diabetes Care* 2007, 30, 529–534. [CrossRef] [PubMed]
- Matilla-Santander, N.; Valvi, D.; Lopez-Espinosa, M.-J.; Manzano-Salgado, C.B.; Ballester, F.; Ibarluzea, J.; Santa-Marina, L.; Schettgen, T.; Guxens, M.; Sunyer, J. Exposure to perfluoroalkyl substances and metabolic outcomes in pregnant women: Evidence from the Spanish INMA birth cohorts. *Environ. Health Perspect.* 2017, 125, 117004. [CrossRef] [PubMed]
- 20. Zhang, C.; Sundaram, R.; Maisog, J.; Calafat, A.M.; Barr, D.B.; Louis, G.M.B. A prospective study of prepregnancy serum concentrations of perfluorochemicals and the risk of gestational diabetes. *Fertil. Steril.* 2015, *103*, 184–189. [CrossRef]
- Groth, S.W.; Fernandez, I.D.; Block, R.C.; Thurston, S.W.; Wong, E.; Brunner, J.; Mayo, N.; Kapula, N.; Yu, Y.; Meng, Y. Biological changes in the pregnancy-postpartum period and subsequent cardiometabolic risk—UPSIDE MOMS: A research protocol. *Res. Nurs. Health* 2021. [CrossRef] [PubMed]
- 22. Barrett, E.S.; Groth, S.W.; Preston, E.V.; Kinkade, C.; James-Todd, T. Endocrine-Disrupting Chemical Exposures in Pregnancy: A Sensitive Window for Later-Life Cardiometabolic Health in Women. *Curr. Epidemiol. Rep.* **2021**, *8*, 130–142. [CrossRef]
- Mitro, S.D.; Sagiv, S.K.; Fleisch, A.F.; Jaacks, L.M.; Williams, P.L.; Rifas-Shiman, S.L.; Calafat, A.M.; Hivert, M.-F.; Oken, E.; James-Todd, T.M. Pregnancy per-and polyfluoroalkyl substance concentrations and postpartum health in Project Viva, a prospective cohort. J. Clin. Endocrinol. Metab. 2020, 105, e3415–e3426. [CrossRef]

- Rodríguez-Carmona, Y.; Cantoral, A.; Trejo-Valdivia, B.; Téllez-Rojo, M.M.; Svensson, K.; Peterson, K.E.; Meeker, J.D.; Schnaas, L.; Solano, M.; Watkins, D.J. Phthalate exposure during pregnancy and long-term weight gain in women. *Environ. Res.* 2019, 169, 26–32. [CrossRef]
- Wu, H.; Just, A.C.; Colicino, E.; Calafat, A.M.; Oken, E.; Braun, J.M.; McRae, N.; Cantoral, A.; Pantic, I.; Pizano-Zárate, M.L. The associations of phthalate biomarkers during pregnancy with later glycemia and lipid profiles. *Environ. Int.* 2021, 155, 106612. [CrossRef] [PubMed]
- Alonso-Magdalena, P.; Quesada, I.; Nadal, A. Endocrine disruptors in the etiology of type 2 diabetes mellitus. *Nat. Rev. Endocrinol.* 2011, 7, 346–353. [CrossRef] [PubMed]
- 27. Heindel, J.J.; Blumberg, B.; Cave, M.; Machtinger, R.; Mantovani, A.; Mendez, M.A.; Nadal, A.; Palanza, P.; Panzica, G.; Sargis, R. Metabolism disrupting chemicals and metabolic disorders. *Reprod. Toxicol.* **2017**, *68*, 3–33. [CrossRef]
- 28. Le Magueresse-Battistoni, B.; Labaronne, E.; Vidal, H.; Naville, D. Endocrine disrupting chemicals in mixture and obesity, diabetes and related metabolic disorders. *World J. Biol. Chem.* 2017, *8*, 108–119. [CrossRef]
- Saeedi, P.; Petersohn, I.; Salpea, P.; Malanda, B.; Karuranga, S.; Unwin, N.; Colagiuri, S.; Guariguata, L.; Motala, A.A.; Ogurtsova, K. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas. *Diabetes Res. Clin. Pract.* 2019, *157*, 107843. [CrossRef]
- 30. Rowley, W.R.; Bezold, C.; Arikan, Y.; Byrne, E.; Krohe, S. Diabetes 2030: Insights from yesterday, today, and future trends. *Popul. Health Manag.* **2017**, *20*, 6–12. [CrossRef] [PubMed]
- Braun, J.M.; Kalkbrenner, A.E.; Just, A.C.; Yolton, K.; Calafat, A.M.; Sjödin, A.; Hauser, R.; Webster, G.M.; Chen, A.; Lanphear, B.P. Gestational exposure to endocrine-disrupting chemicals and reciprocal social, repetitive, and stereotypic behaviors in 4-and 5-year-old children: The HOME study. *Environ. Health Perspect.* 2014, 122, 513–520. [CrossRef] [PubMed]
- Kalloo, G.; Wellenius, G.A.; McCandless, L.; Calafat, A.M.; Sjodin, A.; Karagas, M.; Chen, A.; Yolton, K.; Lanphear, B.P.; Braun, J.M. Profiles and predictors of environmental chemical mixture exposure among pregnant women: The health outcomes and measures of the environment study. *Environ. Sci. Technol.* 2018, 52, 10104–10113. [CrossRef] [PubMed]
- 33. Wolff, M.S.; Engel, S.M.; Berkowitz, G.S.; Ye, X.; Silva, M.J.; Zhu, C.; Wetmur, J.; Calafat, A.M. Prenatal phenol and phthalate exposures and birth outcomes. *Environ. Health Perspect.* **2008**, *116*, 1092–1097. [CrossRef] [PubMed]
- Kelley, A.S.; Banker, M.; Goodrich, J.M.; Dolinoy, D.C.; Burant, C.; Domino, S.E.; Smith, Y.R.; Song, P.X.; Padmanabhan, V. Early pregnancy exposure to endocrine disrupting chemical mixtures are associated with inflammatory changes in maternal and neonatal circulation. *Sci. Rep.* 2019, *9*, 5422. [CrossRef] [PubMed]
- 35. Howdeshell, K.L.; Hotchkiss, A.K.; Gray, L.E., Jr. Cumulative effects of antiandrogenic chemical mixtures and their relevance to human health risk assessment. *Int. J. Hyg. Environ. Health* **2017**, 220, 179–188. [CrossRef]
- Howdeshell, K.L.; Wilson, V.S.; Furr, J.; Lambright, C.R.; Rider, C.V.; Blystone, C.R.; Hotchkiss, A.K.; Gray, L.E., Jr. A mixture of five phthalate esters inhibits fetal testicular testosterone production in the sprague-dawley rat in a cumulative, dose-additive manner. *Toxicol. Sci.* 2008, 105, 153–165. [CrossRef] [PubMed]
- Rider, C.V.; Wilson, V.S.; Howdeshell, K.L.; Hotchkiss, A.K.; Furr, J.R.; Lambright, C.R.; Gray, L.E. Cumulative effects of in utero administration of mixtures of "antiandrogens" on male rat reproductive development. *Toxicol. Pathol.* 2009, 37, 100–113. [CrossRef]
- Conley, J.M.; Lambright, C.S.; Evans, N.; Cardon, M.; Medlock-Kakaley, E.; Wilson, V.S.; Gray, L.E., Jr. A mixture of 15 phthalates and pesticides below individual chemical no observed adverse effect levels (NOAELs) produces reproductive tract malformations in the male rat. *Environ. Int.* 2021, 156, 106615. [CrossRef]
- 39. Rajapakse, N.; Silva, E.; Kortenkamp, A. Combining xenoestrogens at levels below individual no-observed-effect concentrations dramatically enhances steroid hormone action. *Environ. Health Perspect.* **2002**, *110*, 917–921. [CrossRef] [PubMed]
- Kortenkamp, A. Low dose mixture effects of endocrine disrupters: Implications for risk assessment and epidemiology. *Int. J. Androl.* 2008, *31*, 233–240. [CrossRef] [PubMed]
- 41. Kortenkamp, A.; Faust, M. Regulate to reduce chemical mixture risk. Science 2018, 361, 224–226. [CrossRef]
- 42. Cory-Slechta, D.A. Studying toxicants as single chemicals: Does this strategy adequately identify neurotoxic risk? *Neurotoxicology* **2005**, *26*, 491–510. [CrossRef]
- 43. Mooradian, A.D. Dyslipidemia in type 2 diabetes mellitus. Nat. Rev. Endocrinol. 2009, 5, 150–159. [CrossRef]
- 44. Yan, S.; Zhang, H.; Zheng, F.; Sheng, N.; Guo, X.; Dai, J. Perfluorooctanoic acid exposure for 28 days affects glucose homeostasis and induces insulin hypersensitivity in mice. *Sci. Rep.* **2015**, *5*, 11029. [CrossRef] [PubMed]
- 45. Kautzky-Willer, A.; Harreiter, J.; Pacini, G. Sex and gender differences in risk, pathophysiology and complications of type 2 diabetes mellitus. *Endocr. Rev.* 2016, *37*, 278–316. [CrossRef] [PubMed]
- Ehrlich, S.; Lambers, D.; Baccarelli, A.; Khoury, J.; Macaluso, M.; Ho, S.-M. Endocrine disruptors: A potential risk factor for gestational diabetes mellitus. *Am. J. Perinatol.* 2016, 33, 1313–1318.
- 47. Ruiz, D.; Becerra, M.; Jagai, J.S.; Ard, K.; Sargis, R.M. Disparities in environmental exposures to endocrine-disrupting chemicals and diabetes risk in vulnerable populations. *Diabetes Care* **2018**, *41*, 193–205. [CrossRef] [PubMed]
- 48. Lim, S.; Ahn, S.Y.; Song, I.C.; Chung, M.H.; Jang, H.C.; Park, K.S.; Lee, K.-U.; Pak, Y.K.; Lee, H.K. Chronic exposure to the herbicide, atrazine, causes mitochondrial dysfunction and insulin resistance. *PLoS ONE* **2009**, *4*, e5186. [CrossRef]
- 49. Jin, Y.; Lin, X.; Miao, W.; Wu, T.; Shen, H.; Chen, S.; Li, Y.; Pan, Q.; Fu, Z. Chronic exposure of mice to environmental endocrinedisrupting chemicals disturbs their energy metabolism. *Toxicol. Lett.* **2014**, 225, 392–400. [CrossRef]

- 50. Wei, J.; Lin, Y.; Li, Y.; Ying, C.; Chen, J.; Song, L.; Zhou, Z.; Lv, Z.; Xia, W.; Chen, X. Perinatal exposure to bisphenol A at reference dose predisposes offspring to metabolic syndrome in adult rats on a high-fat diet. *Endocrinology* **2011**, *152*, 3049–3061. [CrossRef]
- Alonso-Magdalena, P.; García-Arévalo, M.; Quesada, I.; Nadal, Á. Bisphenol-A Treatment during Pregnancy in Mice: A New Window of Susceptibility for the Development of Diabetes in Mothers Later in Life. *Endocrinology* 2015, 156, 1659–1670. [CrossRef] [PubMed]
- 52. Susiarjo, M.; Xin, F.; Bansal, A.; Stefaniak, M.; Li, C.; Simmons, R.A.; Bartolomei, M.S. Bisphenol a exposure disrupts metabolic health across multiple generations in the mouse. *Endocrinology* **2015**, *156*, 2049–2058. [CrossRef] [PubMed]
- 53. Alonso-Magdalena, P.; Morimoto, S.; Ripoll, C.; Fuentes, E.; Nadal, A. The estrogenic effect of bisphenol A disrupts pancreatic β-cell function in vivo and induces insulin resistance. *Environ. Health Perspect.* **2006**, *114*, 106–112. [CrossRef]
- 54. Moon, M.K.; Jeong, I.-K.; Oh, T.J.; Ahn, H.Y.; Kim, H.H.; Park, Y.J.; Jang, H.C.; Park, K.S. Long-term oral exposure to bisphenol A induces glucose intolerance and insulin resistance. J. Endocrinol. 2015, 226, 35–42. [CrossRef]
- 55. Alonso-Magdalena, P.; Vieira, E.; Soriano, S.; Menes, L.; Burks, D.; Quesada, I.; Nadal, A. Bisphenol A exposure during pregnancy disrupts glucose homeostasis in mothers and adult male offspring. *Environ. Health Perspect.* **2010**, *118*, 1243–1250. [CrossRef]
- 56. Le Magueresse-Battistoni, B.; Multigner, L.; Beausoleil, C.; Rousselle, C. Effects of bisphenol A on metabolism and evidences of a mode of action mediated through endocrine disruption. *Mol. Cell. Endocrinol.* **2018**, 475, 74–91. [CrossRef] [PubMed]
- 57. Angle, B.M.; Do, R.P.; Ponzi, D.; Stahlhut, R.W.; Drury, B.E.; Nagel, S.C.; Welshons, W.V.; Besch-Williford, C.L.; Palanza, P.; Parmigiani, S. Metabolic disruption in male mice due to fetal exposure to low but not high doses of bisphenol A (BPA): Evidence for effects on body weight, food intake, adipocytes, leptin, adiponectin, insulin and glucose regulation. *Reprod. Toxicol.* 2013, 42, 256–268. [CrossRef] [PubMed]
- 58. Miyawaki, J.; Sakayama, K.; Kato, H.; Yamamoto, H.; Masuno, H. Perinatal and postnatal exposure to bisphenol a increases adipose tissue mass and serum cholesterol level in mice. *J. Atheroscler. Thromb.* **2007**, *14*, 245. [CrossRef]
- 59. Yang, M.; Chen, M.; Wang, J.; Xu, M.; Sun, J.; Ding, L.; Lv, X.; Ma, Q.; Bi, Y.; Liu, R. Bisphenol A promotes adiposity and inflammation in a nonmonotonic dose-response way in 5-week-old male and female C57BL/6J mice fed a low-calorie diet. *Endocrinology* **2016**, *157*, 2333–2345. [CrossRef]
- 60. Pereira-Fernandes, A.; Demaegdt, H.; Vandermeiren, K.; Hectors, T.L.; Jorens, P.G.; Blust, R.; Vanparys, C. Evaluation of a screening system for obesogenic compounds: Screening of endocrine disrupting compounds and evaluation of the PPAR dependency of the effect. *PLoS ONE* **2013**, *8*, e77481. [CrossRef] [PubMed]
- 61. Begley, T.; White, K.; Honigfort, P.; Twaroski, M.; Neches, R.; Walker, R. Perfluorochemicals: Potential sources of and migration from food packaging. *Food Addit. Contam.* **2005**, *22*, 1023–1031. [CrossRef]
- 62. Trudel, D.; Horowitz, L.; Wormuth, M.; Scheringer, M.; Cousins, I.T.; Hungerbühler, K. Estimating consumer exposure to PFOS and PFOA. *Risk Anal. Int. J.* 2008, *28*, 251–269. [CrossRef]
- 63. Hines, E.P.; White, S.S.; Stanko, J.P.; Gibbs-Flournoy, E.A.; Lau, C.; Fenton, S.E. Phenotypic dichotomy following developmental exposure to perfluorooctanoic acid (PFOA) in female CD-1 mice: Low doses induce elevated serum leptin and insulin, and overweight in mid-life. *Mol. Cell. Endocrinol.* **2009**, *304*, 97–105. [CrossRef] [PubMed]
- 64. Louisse, J.; Rijkers, D.; Stoopen, G.; Janssen, A.; Staats, M.; Hoogenboom, R.; Kersten, S.; Peijnenburg, A. Perfluorooctanoic acid (PFOA), perfluorooctane sulfonic acid (PFOS), and perfluorononanoic acid (PFNA) increase triglyceride levels and decrease cholesterogenic gene expression in human HepaRG liver cells. *Arch. Toxicol.* **2020**, *94*, 3137–3155. [CrossRef] [PubMed]
- 65. Rebholz, S.L.; Jones, T.; Herrick, R.L.; Xie, C.; Calafat, A.M.; Pinney, S.M.; Woollett, L.A. Hypercholesterolemia with consumption of PFOA-laced Western diets is dependent on strain and sex of mice. *Toxicol. Rep.* **2016**, *3*, 46–54. [CrossRef] [PubMed]
- 66. Marques, E.S.; Agudelo, J.; Kaye, E.M.; Modaresi, S.M.S.; Pfohl, M.; Bečanová, J.; Wei, W.; Polunas, M.; Goedken, M.; Slitt, A.L. The role of maternal high fat diet on mouse pup metabolic endpoints following perinatal PFAS and PFAS mixture exposure. *Toxicology* 2021, 462, 152921. [CrossRef] [PubMed]
- 67. Mastroiacovo, P.; Spagnolo, A.; Marni, E.; Meazza, L.; Bertollini, R.; Segni, G. Birth defects in the Seveso area after TCDD contamination. *JAMA* **1988**, 259, 1668–1672. [CrossRef] [PubMed]
- 68. Lawson, C.C.; Schnorr, T.M.; Whelan, E.A.; Deddens, J.A.; Dankovic, D.A.; Piacitelli, L.A.; Sweeney, M.H.; Connally, L.B. Paternal occupational exposure to 2,3,7,8-tetrachlorodibenzo-p-dioxin and birth outcomes of offspring: Birth weight, preterm delivery, and birth defects. *Environ. Health Perspect.* **2004**, *112*, 1403–1408. [CrossRef] [PubMed]
- 69. Novelli, M.; Piaggi, S.; De Tata, V. 2,3,7,8-Tetrachlorodibenzo-p-dioxin-induced impairment of glucose-stimulated insulin secretion in isolated rat pancreatic islets. *Toxicol. Lett.* **2005**, *156*, 307–314. [CrossRef]
- Kurita, H.; Yoshioka, W.; Nishimura, N.; Kubota, N.; Kadowaki, T.; Tohyama, C. Aryl hydrocarbon receptor-mediated effects of 2,3,7,8-tetrachlorodibenzo-p-dioxin on glucose-stimulated insulin secretion in mice. J. Appl. Toxicol. Int. J. 2009, 29, 689–694. [CrossRef]
- 71. Piaggi, S.; Novelli, M.; Martino, L.; Masini, M.; Raggi, C.; Orciuolo, E.; Masiello, P.; Casini, A.; De Tata, V. Cell death and impairment of glucose-stimulated insulin secretion induced by 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) in the β-cell line INS-1E. *Toxicol. Appl. Pharmacol.* 2007, 220, 333–340. [CrossRef] [PubMed]
- 72. Angrish, M.M.; Dominici, C.Y.; Zacharewski, T.R. TCDD-elicited effects on liver, serum, and adipose lipid composition in C57BL/6 mice. *Toxicol. Sci.* 2013, 131, 108–115. [CrossRef]
- Coban, A.; Filipov, N. Dopaminergic toxicity associated with oral exposure to the herbicide atrazine in juvenile male C57BL/6 mice. J. Neurochem. 2007, 100, 1177–1187. [CrossRef]

- 74. Ross, M.K.; Filipov, N.M. Determination of atrazine and its metabolites in mouse urine and plasma by LC–MS analysis. *Anal. Biochem.* **2006**, *351*, 161–173. [CrossRef] [PubMed]
- 75. Galbiati, V.; Buoso, E.; di Villa Bianca, R.D.; Paola, R.D.; Morroni, F.; Nocentini, G.; Racchi, M.; Viviani, B.; Corsini, E. Immune and Nervous Systems Interaction in Endocrine Disruptors Toxicity: The Case of Atrazine. *Front. Toxicol.* **2021**, *3*, 4. [CrossRef]
- Rowe, A.M.; Brundage, K.M.; Barnett, J.B. Developmental immunotoxicity of atrazine in rodents. *Basic Clin. Pharmacol. Toxicol.* 2008, 102, 139–145. [CrossRef]
- 77. Cope, H.A.; Blake, B.E.; Love, C.; McCord, J.; Elmore, S.A.; Harvey, J.B.; Chappell, V.A.; Fenton, S.E. Latent, sex-specific metabolic health effects in CD-1 mouse offspring exposed to PFOA or HFPO-DA (GenX) during gestation. *Emerg. Contam.* 2021, 7, 219–235. [CrossRef]
- 78. Fenton, S.E.; Reiner, J.L.; Nakayama, S.F.; Delinsky, A.D.; Stanko, J.P.; Hines, E.P.; White, S.S.; Lindstrom, A.B.; Strynar, M.J.; Petropoulou, S.-S.E. Analysis of PFOA in dosed CD-1 mice. Part 2: Disposition of PFOA in tissues and fluids from pregnant and lactating mice and their pups. *Reprod. Toxicol.* 2009, *27*, 365–372. [CrossRef]
- 79. Vorderstrasse, B.A.; Cundiff, J.A.; Paige Lawrence, B. A dose-response study of the effects of prenatal and lactational exposure to TCDD on the immune response to influenza a virus. *J. Toxicol. Environ. Health Part A* **2006**, *69*, 445–463. [CrossRef]
- 80. US Environmental Protection Agency. *Health Effects Support Document for Perfluorooctanoic Acid (PFOA);* US Environmental Protection Agency: Washington, DC, USA, 2016.
- Sobolewski, M.; Conrad, K.; Allen, J.L.; Weston, H.; Martin, K.; Lawrence, B.P.; Cory-Slechta, D.A. Sex-specific enhanced behavioral toxicity induced by maternal exposure to a mixture of low dose endocrine-disrupting chemicals. *Neurotoxicology* 2014, 45, 121–130. [CrossRef]
- 82. Sobolewski, M.; Allen, J.L.; Morris-Schaffer, K.; Klocke, C.; Conrad, K.; Cory-Slechta, D.A. A novel, ecologically relevant, highly preferred, and non-invasive means of oral substance administration for rodents. *Neurotoxicol. Teratol.* **2016**, *56*, 75–80. [CrossRef]
- 83. Fields, A.M.; Welle, K.; Ho, E.S.; Mesaros, C.; Susiarjo, M. Vitamin B6 deficiency disrupts serotonin signaling in pancreatic islets and induces gestational diabetes in mice. *Commun. Biol.* **2021**, *4*, 421. [CrossRef] [PubMed]
- 84. Stillerman, K.P.; Mattison, D.R.; Giudice, L.C.; Woodruff, T.J. Environmental exposures and adverse pregnancy outcomes: A review of the science. *Reprod. Sci.* 2008, *15*, 631–650. [CrossRef]
- Varshavsky, J.; Smith, A.; Wang, A.; Hom, E.; Izano, M.; Huang, H.; Padula, A.; Woodruff, T.J. Heightened Susceptibility: A Review of How Pregnancy and Chemical Exposures Influence Maternal Health. *Reprod. Toxicol.* 2019, 92, 14–56. [CrossRef] [PubMed]
- 86. Reece, E.A. The fetal and maternal consequences of gestational diabetes mellitus. *J. Matern.-Fetal Neonatal Med.* **2010**, *23*, 199–203. [CrossRef] [PubMed]
- Rider, C.V.; McHale, C.M.; Webster, T.F.; Lowe, L.; Goodson, W.H., III; La Merrill, M.A.; Rice, G.; Zeise, L.; Zhang, L.; Smith, M.T. Using the Key Characteristics of Carcinogens to Develop Research on Chemical Mixtures and Cancer. *Environ. Health Perspect.* 2021, 129, 035003. [CrossRef]
- Christiansen, S.; Axelstad, M.; Scholze, M.; Johansson, H.K.; Hass, U.; Mandrup, K.; Frandsen, H.L.; Frederiksen, H.; Isling, L.K.; Boberg, J. Grouping of endocrine disrupting chemicals for mixture risk assessment–Evidence from a rat study. *Environ. Int.* 2020, 142, 105870. [CrossRef]
- 89. Kortenkamp, A. Which chemicals should be grouped together for mixture risk assessments of male reproductive disorders? *Mol. Cell. Endocrinol.* **2020**, *499*, 110581. [CrossRef] [PubMed]
- 90. Eckel, R.H.; Grundy, S.M.; Zimmet, P.Z. The metabolic syndrome. Lancet 2005, 365, 1415–1428. [CrossRef]
- 91. Batista, T.M.; Alonso-Magdalena, P.; Vieira, E.; Amaral, M.E.C.; Cederroth, C.R.; Nef, S.; Quesada, I.; Carneiro, E.M.; Nadal, A. Short-term treatment with bisphenol-A leads to metabolic abnormalities in adult male mice. *PLoS ONE* **2012**, *7*, e33814. [CrossRef]
- Marmugi, A.; Lasserre, F.; Beuzelin, D.; Ducheix, S.; Huc, L.; Polizzi, A.; Chetivaux, M.; Pineau, T.; Martin, P.; Guillou, H. Adverse effects of long-term exposure to bisphenol A during adulthood leading to hyperglycaemia and hypercholesterolemia in mice. *Toxicology* 2014, 325, 133–143. [CrossRef]
- Zheng, F.; Sheng, N.; Zhang, H.; Yan, S.; Zhang, J.; Wang, J. Perfluorooctanoic acid exposure disturbs glucose metabolism in mouse liver. *Toxicol. Appl. Pharmacol.* 2017, 335, 41–48. [CrossRef]
- 94. Reaven, G.M.; Chen, Y.-D.; Jeppesen, J.; Maheux, P.; Krauss, R.M. Insulin resistance and hyperinsulinemia in individuals with small, dense low density lipoprotein particles. *J. Clin. Investig.* **1993**, *92*, 141–146. [CrossRef]
- 95. Kannel, W.B. Lipids, diabetes, and coronary heart disease: Insights from the Framingham Study. *Am. Heart J.* **1985**, *110*, 1100–1107. [CrossRef]
- Lorenzo, C.; Hartnett, S.; Hanley, A.J.; Rewers, M.J.; Wagenknecht, L.E.; Karter, A.J.; Haffner, S.M. Impaired fasting glucose and impaired glucose tolerance have distinct lipoprotein and apolipoprotein changes: The insulin resistance atherosclerosis study. J. Clin. Endocrinol. Metab. 2013, 98, 1622–1630. [CrossRef]
- Festa, A.; Williams, K.; Hanley, A.J.; Otvos, J.D.; Goff, D.C.; Wagenknecht, L.E.; Haffner, S.M. Nuclear magnetic resonance lipoprotein abnormalities in prediabetic subjects in the Insulin Resistance Atherosclerosis Study. *Circulation* 2005, 111, 3465–3472. [CrossRef]
- Garvey, W.T.; Kwon, S.; Zheng, D.; Shaughnessy, S.; Wallace, P.; Hutto, A.; Pugh, K.; Jenkins, A.J.; Klein, R.L.; Liao, Y. Effects of insulin resistance and type 2 diabetes on lipoprotein subclass particle size and concentration determined by nuclear magnetic resonance. *Diabetes* 2003, *52*, 453–462. [CrossRef] [PubMed]

- 99. Brulport, A.; Le Corre, L.; Chagnon, M.-C. Chronic exposure of 2, 3, 7, 8-tetrachlorodibenzo-p-dioxin (TCDD) induces an obesogenic effect in C57BL/6J mice fed a high fat diet. *Toxicology* **2017**, *390*, 43–52. [CrossRef] [PubMed]
- Bahceci, M.; Aydemir, M.; Tuzcu, A. Effects of oral fat and glucose tolerance test on serum lipid profile, apolipoprotein, and CRP concentration, and insulin resistance in patients with polycystic ovary syndrome. *Fertil. Steril.* 2007, *87*, 1363–1368. [CrossRef] [PubMed]
- Somm, E.; Schwitzgebel, V.M.; Toulotte, A.; Cederroth, C.R.; Combescure, C.; Nef, S.; Aubert, M.L.; Hüppi, P.S. Perinatal exposure to bisphenol a alters early adipogenesis in the rat. *Environ. Health Perspect.* 2009, 117, 1549–1555. [CrossRef]
- 102. Kim, M.J.; Pelloux, V.; Guyot, E.; Tordjman, J.; Bui, L.-C.; Chevallier, A.; Forest, C.; Benelli, C.; Clément, K.; Barouki, R. Inflammatory pathway genes belong to major targets of persistent organic pollutants in adipose cells. *Environ. Health Perspect.* 2012, 120, 508–514. [CrossRef] [PubMed]
- Regnier, S.M.; Sargis, R.M. Adipocytes under assault: Environmental disruption of adipose physiology. *Biochim. Biophys. Acta* (BBA)-Mol. Basis Dis. 2014, 1842, 520–533. [CrossRef] [PubMed]
- 104. Yamauchi, T.; Kamon, J.; Waki, H.; Terauchi, Y.; Kubota, N.; Hara, K.; Mori, Y.; Ide, T.; Murakami, K.; Tsuboyama-Kasaoka, N. The fat-derived hormone adiponectin reverses insulin resistance associated with both lipoatrophy and obesity. *Nat. Med.* 2001, 7, 941–946. [CrossRef] [PubMed]
- 105. Panee, J. Monocyte Chemoattractant Protein 1 (MCP-1) in obesity and diabetes. Cytokine 2012, 60, 1–12. [CrossRef] [PubMed]
- 106. Yarmolinsky, J.; Barbieri, N.B.; Weinmann, T.; Ziegelmann, P.K.; Duncan, B.B.; Schmidt, M.I. Plasminogen activator inhibitor-1 and type 2 diabetes: A systematic review and meta-analysis of observational studies. *Sci. Rep.* **2016**, *6*, 17714. [CrossRef]
- 107. Steppan, C.M.; Bailey, S.T.; Bhat, S.; Brown, E.J.; Banerjee, R.R.; Wright, C.M.; Patel, H.R.; Ahima, R.S.; Lazar, M.A. The hormone resistin links obesity to diabetes. *Nature* 2001, 409, 307–312. [CrossRef]
- 108. Gui, Y.; Silha, J.V.; Mishra, S.; Murphy, L.J. Changes in adipokine expression during food deprivation in the mouse and the relationship to fasting-induced insulin resistance. *Can. J. Physiol. Pharmacol.* **2003**, *81*, 979–985. [CrossRef]
- 109. Bertile, F.; Raclot, T. Adipose-derived factors during nutritional transitions. Curr. Nutr. Food Sci. 2006, 2, 127–139. [CrossRef]
- Karatsoreos, I.N.; Bhagat, S.M.; Bowles, N.P.; Weil, Z.M.; Pfaff, D.W.; McEwen, B.S. Endocrine and physiological changes in response to chronic corticosterone: A potential model of the metabolic syndrome in mouse. *Endocrinology* 2010, 151, 2117–2127. [CrossRef] [PubMed]
- 111. Strack, A.M.; Sebastian, R.J.; Schwartz, M.W.; Dallman, M.F. Glucocorticoids and insulin: Reciprocal signals for energy balance. *Am. J. Physiol.-Regul. Integr. Comp. Physiol.* **1995**, *268*, R142–R149. [CrossRef]
- 112. De Boer, S.; Koopmans, S.; Slangen, J.; Van der Gugten, J. Plasma catecholamine, corticosterone and glucose responses to repeated stress in rats: Effect of interstressor interval length. *Physiol. Behav.* **1990**, *47*, 1117–1124. [CrossRef]
- 113. Baker, M. Statisticians issue warning over misuse of *p* values. *Nat. News* **2016**, 531, 151. [CrossRef] [PubMed]